Chemical Science

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Cite this: Chem. Sci., 2023, 14, 13842

All publication charges for this article have been paid for by the Royal Society of Chemistry

Received 8th August 2023 Accepted 23rd October 2023

DOI: 10.1039/d3sc04123a

rsc.li/chemical-science

Introduction

Triplet dynamic nuclear polarization of pyruvate *via* supramolecular chemistry†

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Dynamic nuclear polarization (DNP) significantly improves the sensitivity of magnetic resonance imaging, and its most important medical application is cancer diagnosis *via* hyperpolarized ¹³C-labeled pyruvate. Unlike cryogenic DNP, triplet-DNP uses photoexcited triplet electrons under mild conditions. However, triplet-DNP of pyruvate has not been observed because of incompatibility of the hydrophobic polarizing agent with hydrophilic pyruvate. This work demonstrates that supramolecular complexation with β -cyclodextrin can disperse 4,4'-(pentacene-6,13-diyl)dibenzoate (NaPDBA), a pentacene derivative with hydrophilic substituents, even in the presence of high sodium pyruvate concentrations. The polarization of photoexcited triplet electron spins in NaPDBA was transferred to the ¹³C spins of sodium pyruvate *via* triplet-DNP of ¹H spins in water and ¹H-to-¹³C cross-polarization. This provides an important step toward the widespread use of ultra-sensitive MRI for cancer diagnosis.

Nuclear magnetic resonance (NMR) is an important nondestructive technique for analyzing chemical structures, and magnetic resonance imaging (MRI) is an essential medical procedure. However, both methods are inherently insensitive because the sensitivity is proportional to spin polarization, which is 0.004% for ¹H spins and 0.001% for ¹³C spins under a 6 T magnetic field at room temperature. Hence, MRI is essentially limited to abundant water molecules.

Dynamic nuclear polarization (DNP) improves the sensitivity of NMR¹⁻¹⁰ because it creates a hyperpolarized nuclear spin state by transferring the high polarization of unpaired electron spins.

^eDepartment of Data Science, School of Frontier Engineering, Kitasato University, Sagamihara, Kanagawa, 252-0373, Japan Dissolution-DNP¹¹⁻¹³ rapidly dissolves hyperpolarized solid samples and has been used for various NMR analyses, such as highly sensitive protein NMR14,15 and in vivo metabolite imaging.¹⁶⁻²⁰ In particular, [1-¹³C] pyruvate is the most important probe for high-sensitivity MRI because it is at the center of metabolism and its metabolic kinetics are used in the diagnosis of various diseases, including cancer.^{16,17,21-24} Dissolution-DNP has achieved ¹³C-NMR signal enhancement of more than 10 000-fold;²⁵ however, it requires severe conditions, such as a high magnetic field (~7 T) and cryogenic temperatures near 1 K, to use the near-unity electron spin polarization. Alternatively, Overhauser-DNP is a powerful technique that polarizes solutions even at room temperature,26-29 but the maximum enhancement factor depends on the difference in the gyromagnetic ratio between nuclear spins and electron spins (γ_e/γ_H \sim 660 and $\gamma_{\rm e}/\gamma_{\rm C} \sim$ 2600).

In contrast, DNP *via* photoexcited triplet electron spins (triplet-DNP) can produce hyperpolarization under milder conditions.³⁰⁻³⁴ Photoexcited triplets have large non-equilibrium spin polarizations (\sim 70% for pentacene³⁵) that are independent of temperature, which enables DNP at higher temperatures and lower magnetic fields. Fig. 1A depicts triplet-DNP. After photoexcitation of a polarizing agent, a temporary spin-polarized state is generated by spin-selective intersystem crossing. After the polarization is transferred from the electron spin to ¹H spin, it is propagated throughout the solid *via* ¹H spin diffusion. Various host molecules and materials have been examined to hyperpolarize drugs,^{36–38} water,^{39–42} and biologically relevant molecules.^{43,44} Dissolution triplet-DNP has been demonstrated.^{45,46} However, spin-polarization transfer to lower

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[†] Electronic supplementary information (ESI) available. See DOI: https://doi.org/10.1039/d3sc04123a

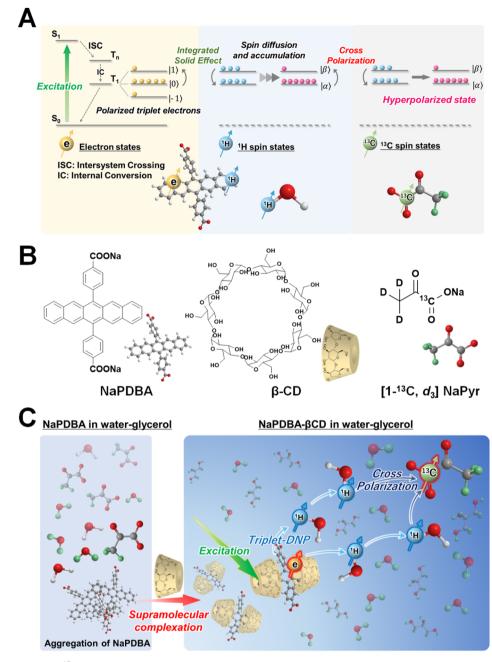


Fig. 1 Hyperpolarization of $[1^{-13}C, d_3]$ sodium pyruvate (NaPyr) by triplet dynamic nuclear polarization (DNP) and cross polarization (CP). (A) Scheme of triplet-DNP and CP. (B) Molecular structures of NaPDBA, β CD, and $[1^{-13}C, d_3]$ NaPyr. (C) NaPDBA aggregate in DNP juice in the absence of β -cyclodextrin (β CD), but the dispersibility is significantly increased by supramolecular complexation with β CD. Polarization transfer from photoexcited triplet electron spins to ¹H spins of water and then to ¹³C spins of $[1^{-13}C, d_3]$ NaPyr.

gyromagnetic ratio spins such as in ¹⁹F and ¹³C *via* cross polarization (CP) has been limited to aromatic molecules,^{33,36,37,47} and triplet-DNP of $[1-^{13}C]$ pyruvate has never been demonstrated because of poor miscibility between the hydrophobic polarizing agent and hydrophilic pyruvate.

Here, we report triplet-DNP of $[1^{-13}C, d_3]$ sodium pyruvate (NaPyr) at ~100 K and 0.64 T. Higher concentrations of NaPyr are desired for dissolution-DNP application because the polarized spins are diluted after dissolution.²⁰ Water-soluble polarizing agents were developed and used for triplet-DNP in

aqueous matrices,^{41,43,48} but hydrophilic polarizing agents easily aggregate with high concentrations of polar pyruvate. To solve this problem, we used supramolecular chemistry to increase the dispersion of the polarizing agent (Fig. 1B). Cyclodextrin can encapsulate hydrophobic dyes in water,^{49,50} and the triplet-DNP of water was observed by using 4,4'-(pentacene-6,13-diyl) dibenzoate (NaPDBA) as a guest in cyclodextrin.⁴⁸ NaPDBA aggregation was prevented by supramolecular complexation with β-cyclodextrin (βCD) with a saturated concentration of 1.5 M NaPyr in DNP juice (H₂O/D₂O/glycerol-d₈ = 1/3/6, v/v/v),

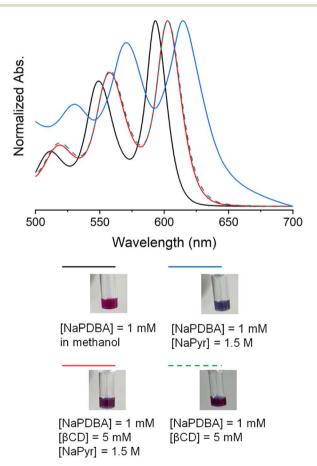
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a general glass forming solvent.²⁰ The well-dispersed NaPDBA- β CD complex generated polarization of triplet electron spins, which was transferred to the ¹H spins of water in DNP juice by triplet-DNP and then to the ¹³C spins of [1-¹³C, *d*₃] NaPyr by CP (Fig. 1C).

Results and discussion

Evaluation of dispersibility of polarizing agents

The dispersed state of NaPDBA in DNP juice was evaluated *via* absorption spectra (Fig. 2) since random aggregation of polarizing agents could induce rapid relaxation of photoexcited triplet electron-spin polarization, significantly decreasing triplet-DNP efficiency. The π - π * absorption peak of 1 mM dispersed NaPDBA in methanol was observed at 593 nm (Fig. S5†). A red-shift to 604 nm was observed in DNP juice, indicating NaPDBA aggregation (Fig. S6†).⁴⁸ When 1.5 M of NaPyr was dissolved in DNP juice, the NaPDBA peak exhibited a large red shift to 614.5 nm, turning the solution blue (Fig. 2). This indicated that the NaPyr salt disrupted NaPDBA hydration and increased its aggregation.⁵¹ The absorption peak was blue-



shifted to 603 nm by adding 5 mM of β CD in DNP juice, in the presence of NaPyr (Fig. 2 and S7†). Previously, the 1 : 2 inclusion complex of NaPDBA- β CD was formed in a water–glycerol mixture [glycerol/H₂O (5/5, v/v)],⁴⁸ and the absorption peak was also blue-shifted by adding 5 mM of β CD in DNP juice [glycerol/H₂O (6/4, v/v)] in the absence of NaPyr (Fig. S8†). No significant change in the absorption spectra of NaPDBA– β CD in DNP juice was observed with or without NaPyr (Fig. 2). Here, the NaPDBA– β CD inclusion complex was intact at high NaPyr concentrations because the addition of salt increased water structuring and shifted the equilibrium toward the bound state.^{52,53}

Evaluation of supramolecular structures

The structure of the NaPDBA-BCD inclusion complex was investigated with NMR and molecular dynamics (MD) simulations. The room-temperature ¹H NMR spectra of NaPDBA shifted upfield when adding β CD in glycerol- d_8/D_2 O (6/4, v/v) that contained NaPyr, which indicated formation of the inclusion complex (Fig. S9[†]).^{54,55} Nuclear-Overhauser-effect roomtemperature NMR of NaPDBA and β CD in D₂O with NaPyr revealed cross peaks between the NaPDBA pentacene skeleton and the β CD inner region (Fig. S10[†]). This indicated that NaPDBA was incorporated into the BCD hydrophobic cavity. NMR spectra of NaPDBA and BCD with different concentrations in D2O which contained NaPyr indicated a 1:2 molar ratio of NaPDBA to β CD in the inclusion complex (Fig. S11[†]). In the MD simulations, the 1:2 inclusion complex was stable in glycerol/ H_2O (6/4, v/v) (Fig. 3A and B), and remained stable in the presence of 1.5 M NaPyr (Fig. 3C and D). These results were

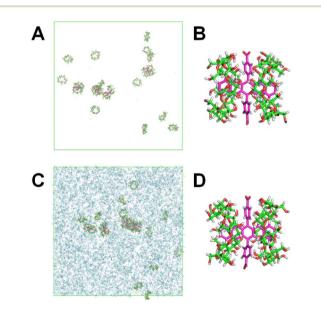


Fig. 2 Dispersion of NaPDBA in the presence of NaPyr. Absorption spectra of NaPDBA in methanol (black line), NaPDBA in DNP juice containing NaPyr (blue line), the NaPDBA- β CD complex in DNP juice (dashed green line) and the NaPDBA- β CD complex in DNP juice containing NaPyr (red line) at room temperature. The concentrations of NaPDBA, NaPyr, and β CD were 1 mM, 1.5 M and 5 mM, respectively. Photographs of each solution are shown.

Fig. 3 Molecular dynamics (MD) simulations of NaPDBA- β CD supramolecular complexes. (A and B) MD simulation snapshots of the NaPDBA- β CD complex ([NaPDBA] = 1 mM and [β CD] = 5 mM) in glycerol/H₂O (6/4, v/v) at 300 K. Glycerol and H₂O were omitted for clarity. (C and D) MD simulation snapshots of the NaPDBA- β CD complex with NaPyr ([NaPDBA] = 1 mM [β CD] = 5 mM and [NaPyr] = 1.5 M) in glycerol/H₂O (6/4, v/v) at 300 K. Glycerol and H₂O are omitted for clarity.

consistent with the dispersibility of the NaPDBA- βCD complex with and without NaPyr.

Time-resolved ESR measurements

The polarization of photoexcited triplet electron spins in NaPDBA in DNP juice was evaluated by time-resolved X-band

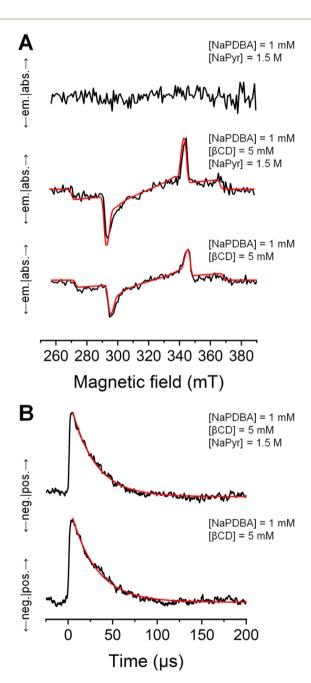


Fig. 4 Time-resolved electron spin resonance (ESR) spectra of NaPDBA and peak signal decays in DNP juice (glycerol/H₂O (6/4, v/v)). (A) ESR spectra of NaPDBA containing NaPyr (top), the NaPDBA– β CD complex containing NaPyr (middle), and the NaPDBA– β CD complex (bottom) after 527 nm photoexcitation at 140 K. The concentrations of NaPDBA, NaPyr and β CD were 1 mM, 1.5 M and 5 mM, respectively. Spectra were fitted with the EasySpin toolbox in MATLAB (red lines). (B) Decays of peak ESR signals (black lines). Single-exponential fits are also shown (red lines).

electron spin resonance (ESR) at 140 K (Fig. 4) after 527 nm pulsed-laser excitation. Almost no ESR signal was observed for NaPDBA in DNP juice containing NaPyr in the absence of BCD (Fig. 4A). This was because of random NaPDBA aggregation, which induced rapid relaxation of the triplet electron polarization. When a chromophore aggregate is in a random orientation, the photoexcited triplet electron spins hop between chromophores with different orientations, causing the electron spins subject to magnetic field fluctuations and inducing relaxation of electron spin polarization.56,57 In contrast, the NaPDBA-BCD complex exhibited a clear ESR signal in DNP juice, even with NaPyr, reflecting well-dispersed NaPDBA via supramolecular complexation. The ESR spectra were fitted with the EasySpin toolbox in MATLAB.58 The zero-field splitting parameters and relative populations of the NaPDBA-BCD complex were almost the same with and without NaPyr, and were almost the same as those for dispersed pentacene and its derivatives (Table S1[†]).^{59,60} This confirmed that the supramolecular structure was intact in the presence of NaPyr, consistent with the absorption measurements. Fig. 4B shows the signal decay of the ESR absorption peak. The NaPDBA-BCD complex had 27 µs and 29 µs polarization lifetimes in the presence and absence of NaPyr, respectively. These lifetimes were long enough to use the triplet-DNP sequence.

Evaluation of the spin-lattice relaxation time

Because the accumulation of spin polarization *via* triplet-DNP competes with ¹H spin relaxation, a solid sample must have a sufficiently long spin–lattice relaxation time (T_1) for efficient triplet-DNP. The ¹H T_1 of glassy DNP juice with NaPDBA and β CD was 46 s at 100 K and 0.64 T, while the addition of NaPyr significantly decreased it to 2 s (Fig. S12†). This was because of increased ¹H spin relaxation *via* methyl group rotation in NaPyr.⁶¹ The use of deuterated sodium pyruvate (d_3 -NaPyr) recovered the ¹H T_1 (26 s), and the difference from ¹H T_1 without NaPyr (46 s) may have been because of remaining ¹H spins in the methyl groups of [1-¹³C, d_3] NaPyr.

DNP measurements

Polarization of the photoexcited NaPDBA triplet electron spins was transferred to the ¹H spins of water in DNP juice [glycerol $d_8/D_2O/H_2O$, 60/30/10, (v/v/v)] by the ISE sequence at 100 K and 0.64 T (see the ESI for details, Fig. S13[†]).^{31,34} NaPDBA was photoexcited with a 527 nm pulsed laser to produce the polarized electron spins. Then, 17.3 GHz microwaves were irradiated to transfer the polarization from electron spins to ¹H spins by matching the frequency of the electron spin in the effective magnetic field in the rotating frame with the Larmor frequency of the ¹H spins in the laboratory frame. The magnetic field was swept during microwave irradiation to use more electron spin packets, since the resonant field of the triplet electron spins is broadened by the random orientation of the polarizing agents and hyperfine interactions. By repeating the ISE sequence at 500 Hz, the ¹H spins were polarized throughout the glassy DNP juice until the accumulation of ¹H spin polarization reaches equilibrium with the spin-lattice relaxation. After the triplet-

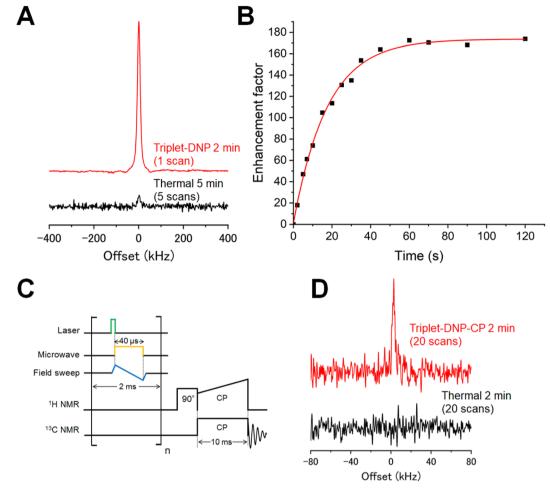


Fig. 5 (A) ¹H NMR signals of water in DNP juice (glycerol- $d_8/D_2O/H_2O$, 60/30/10, v/v/v) containing NaPDBA, β CD and [1-¹³C, d_3] NaPyr under thermal conditions (five scans every 5 min) and after triplet-DNP (integrated solid effect sequence for 2 min and 1 scan) at 100 K. (B) ¹H polarization buildup curve of DNP juice containing NaPDBA, β CD and [1-¹³C, d_3] NaPyr at 100 K. The enhancement factors were calculated by comparing the peak areas after triplet-DNP with that of thermal equilibrium. The enhancement factor relative to thermal equilibrium at room temperature is shown. (C) Sequence of triplet-DNP and ramped-amplitude cross-polarization (RAMP-CP). (D) 6.864 MHz ¹³C NMR spectra of [1-¹³C, d_3] NaPyr. The red line shows the spectra after triplet-DNP and RAMP-CP (integrated solid effect sequence for 2 min, followed by RAMP-CP, 20 scans) and the black line shows the spectra after RAMP-CP with thermal ¹H spins (20 scans).

DNP process in a microwave resonator, the sample was shuttled within 1 s to a coil *via* a motor to acquire ¹H NMR. Fig. 5A shows ¹H NMR of DNP juice containing 1 mM NaPDBA, 5 mM β CD, and 1.5 M [1-¹³C, *d*₃] NaPyr obtained at the thermal state after 2 min of triplet-DNP, in which the accumulation of spin polarization and spin relaxation reached equilibrium. Enhancement factors of 58 and 174 were obtained relative to thermal equilibria at 100 K and room temperature, respectively (Fig. 5B).

The ¹H spin polarization in DNP juice was transferred intermolecularly to ¹³C spins in [1-¹³C, d_3] NaPyr with a rampedamplitude cross-polarization (RAMP-CP) sequence (Fig. 5C).^{62,63} After triplet-DNP at 100 K and 0.64 T, the sample was shuttled into a double-resonance coil with 27.30 MHz and 6.864 MHz resonance frequencies for ¹H and ¹³C spins, respectively. Polarization transfer from ¹H spins to ¹³C spins was conducted using a 10 ms contact. The irradiated field for the ¹³C spins was fixed at 20 kHz, while that for the ¹H spins was swept over the range of 15–25 kHz. After triplet-DNP and RAMP-CP, enhanced ¹³C NMR of $[1^{-13}C, d_3]$ NaPyr was observed (Fig. 5D). The enhancement was clear from the fact that no ¹³C NMR peak was observed via RAMP-CP when using ¹H spins at thermal equilibrium. ¹³C-methanol at room temperature was used as a reference, and an enhancement factor of 122 was estimated (Fig. S14[†]). ¹³C spins in [1-¹³C, d_3] NaPyr accounted for ~85% of the total ¹³C spins in the entire solid sample; thus, the polarization enhancement factor of the ¹³C spins in $[1^{-13}C, d_3]$ NaPyr should have been close to 122. Because the theoretical ¹³C NMR enhancement was ~4 times $(\gamma_{\rm H}/\gamma_{\rm C})$ that of ¹H NMR with ideal CP, the CP efficiency was 18%. The low efficiency could be attributed to the absence of ¹H spins in $[1-^{13}C, d_3]$ NaPyr, because 40-60% efficiencies were observed when ¹H spins were present in the target molecule.64 Furthermore, ideal CP requires that the radiofrequency pulse amplitude be stronger than the ¹H NMR linewidth (~50 kHz).⁶⁵ A higher CP efficiency could be obtained by using a stronger radiofrequency pulse.

Conclusions

In conclusion, triplet-DNP of $[1^{-13}C, d_3]$ NaPyr, the most important biomolecular probe in MRI applications, was observed by increasing the polarizing agent dispersion via supramolecular complexation. It is important to perform DNP with highly concentrated NaPyr, but the method to disperse the polarizing agent in such a highly polar medium had not been clear. Here, the polarizing agent could be modified with ionic carboxylate moieties and then complexed in supramolecular cyclodextrin, which enabled adequate dispersion. This enabled hyperpolarization of the $[1^{-13}C, d_3]$ NaPyr ¹³C spins *via* triplet-DNP-CP under a low magnetic field (~0.64 T) and above liquid nitrogen temperatures (~100 K). Since the final polarization obtained by triplet-DNP is determined by the build up time constant, T_b, and T₁, higher ¹H and ¹³C spin polarizations require a shorter $T_{\rm b}$ and longer $T_{\rm 1}$. Recently, novel pentacene derivatives showing sharper ESR lines exhibits a shorter T_b than pentacene and the ¹H spin polarization is four times higher than that of pentacene.66 In addition, the polarization increased 10-fold as T_1 increased from 1 to 3 min, and a spin polarization of 8% was achieved in a model amorphous matrix.66 Higher ¹H and ¹³C spin polarization can be obtained by using such a new polarizing agent instead of NaPDBA to obtain shorter $T_{\rm b}$ and by diluting the ¹H spins or using other matrices with longer T_1 . In addition, the implementation of more advanced polarization transfer methods with field/frequency modulation would improve the final spin polarization.67,68 Hyperpolarized MRI with triplet-DNP will be possible by combining the optimized molecular design of polarizing agents with the key finding of the present study that supramolecular complexation is useful to hyperpolarize NaPyr at high concentrations.

Data availability

All experimental data are available in the article and ESI.†

Author contributions

T. H., Y. K., H. K. and N. Y. designed the research. T. H. and S. K. prepared and characterized the materials. K. T. and T. U. built time-resolved ESR and DNP setups. T. H. and S. K. conducted time-resolved ESR measurements. T. H. and K. N. conducted DNP measurements. S. S. and G. W. carried out MD simulation. T. H., K. N. and N. Y. wrote the manuscript with contributions from all authors.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

This work was partially supported by the JST-PRESTO program on "Creation of Life Science Basis by Using Quantum Technology" (JPMJPR18GB), JST-FOREST Program (JPMJFR201Y), JSPS KAKENHI (JP20H02713, JP22K19051, JP21J13049, and JP22J21293), the Shinnihon Foundation of Advanced Medical Treatment Research, RIKEN-Kyushu Univ. Science and Technology Hub Collaborative Research Program, and the RIKEN Cluster for Science, Technology and Innovation Hub (RCSTI).

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