REVIEW

Check for updates

Corey–Fuchs reaction enabled synthesis of natural products: a review

Rabia Ashraf,[a] Ameer Fawad Zahoor, [ID] *[a] Samreen Gul Khan,[a] Muhammad Jawwad Saif, [ID] [b] Syed Makhdoom Hussain,[c] Syed Ali Raza Naqvi,[a] Bushra Parveen,[a] Kulsoom Ghulam Ali,[a] Asim Mansha[a] and Ahmad Irfan[d]

Natural products can be derived from a vast array of animals, plants and microorganisms and are generally characterized by a wide spectrum of bioactive properties such as anti-viral, anti-cancer, anti-inflammatory and anti-bacterial properties. Synthesis of natural products is of paramount importance in various fields including medicine, biotechnology and agriculture. The Corey–Fuchs reaction, also known as the Ramirez Corey–Fuchs reaction, is a pivotal organic transformation and plays a significant role in the synthesis of intricate natural products and their analogues. This review article highlights the development of the Corey–Fuchs reaction in recent years towards the synthesis of complex natural products including polyketides, alkaloids, terpenoids and peptides.

1. Introduction

The Corey–Fuchs reaction was established by Nobel laureate E. J. Corey and his colleague P. L. Fuchs in 1972.[1] It is a pioneering transformation in organic synthesis involving the construction of a terminal alkyne *via* one carbon homologation of aldehyde. The reaction protocol includes a two steps methodology entailing the formation of α,α-dibromoalkene from an aldehyde followed by its conversion to a terminal alkyne under the assistance of a strong base (Scheme 1). The general reaction mechanism involves the formation of phosphorus ylide **8** from the reaction of triphenylphosphine and carbon tetrabromide, succeeded by its treatment with aldehyde **1** to furnish dibromoolefin **2**. In the next step, the lithium halogen exchange process and elimination lead to the synthesis of desired alkyne **3** (Scheme 2).[2]

Terminal alkynes constitute a primary class of organic substances that have gained considerable acclaim since the turn of the century due to their diverse utilization and applications in multiple fields. Over 400 naturally occurring substances possessing terminal alkyne as a building block, have been characterized by a range of biological active properties including anti-viral, anti-cancer, anti-microbial, anti-malarial and anti-inflammatory[3] properties. Moreover, alkynes

synthesized through Corey–Fuchs reaction, act as key intermediates in the assembly of various other natural products, such as macrolides,[4] δ-lactone,[5] alkaloids,[6] unsaturated fatty acids,[7] polyketides[8] and terpenoids[9] *etc.*

A well-utilized strategy for the construction of alkynes is the one-carbon homologation of carbonyl compounds. Building on Corey's pioneering studies in this field,[10] numerous methodologies including Ramirez Corey–Fuchs reaction, Colvin rearrangement,[11] Ohira–Bestmann reaction[12] and Seyferth–Gilbert homologation[11] have been come forth in the literature. Corey–Fuchs reaction is often found as an optimal method for the alkyne synthesis owing to mild reaction conditions, wide substrate scope and high yields over other phosphorous-catalyst based reactions.[13] Ohira–Bestman reagent (dimethyl 1-diazo-2-oxopropylphosphonate) (termed as Seyferth–Gilbert homologation) was widely employed for the synthesis of terminal alkynes, however, these types of reagents are relatively expensive, precarious and also have limited applications for the preparation of acetylenic group starting from aldehydes.[14] Moreover, Corey–Fuchs procedure allows for the preparation of enantiomerically pure products in contrast to Ohira–Bestmann reaction protocol as it is incompatible due to the racemization of the product.[15]

[a]Department of Chemistry, Government College University Faisalabad, Faisalabad, 38000, Pakistan. E-mail: fawad.zahoor@gcuf.edu.pk

[b]Department of Applied Chemistry, Government College University Faisalabad, Faisalabad, 38000, Pakistan
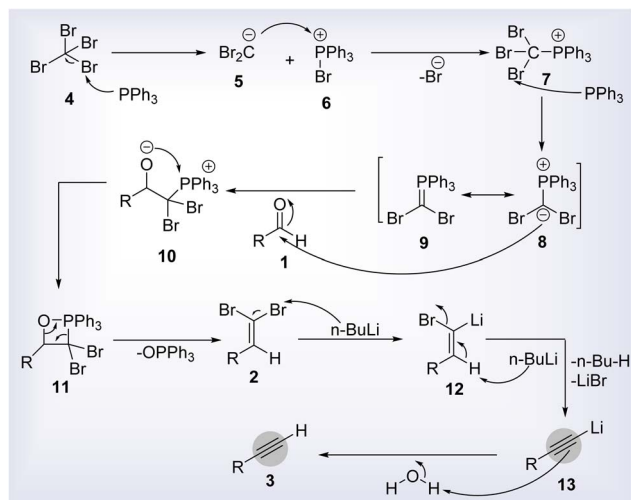
[c]Department of Zoology, Government College University Faisalabad, Faisalabad, 38000, Pakistan

[d]Department of Chemistry, College of Science, King Khalid University, P. O. Box 9004, Abha, 61413, Saudi Arabia
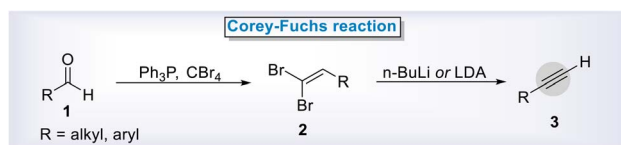
Scheme 1 General reaction of Corey–Fuchs protocol.

Scheme 2 General mechanism for Corey–Fuchs reaction involving the synthesis of terminal alkyne.

Various organic reactions are employed to carry out the total synthesis of naturally occurring compounds and other intricate biologically active heterocyclic functionalities.[16–19] Surplus literature regarding the construction of several sophisticated natural products and other intricate organic compounds involving the Corey–Fuchs reaction as a main step, has been reported yet.[20–23] Natural products have been the backbone of various biological active and potential lead compounds and their total synthesis has witnessed a pronounced advancement in recent years.[24] In this perspective, Lee *et al.*, in 2014, demonstrated the total synthesis of (+)-Violapyrone C employing Corey–Fuchs reaction. (+)-Violapyrone C **14** is featured with high cytotoxic activity against six human tumor cell lines.[25] Similarly, in the same year, Sengoku and coworkers described the total synthesis of Kendomycin **15** utilizing the Corey–Fuchs reaction conditions.[26] This natural product not only exhibit strong anti-tumor activity against human call lines but also show excellent anti-bacterial properties against Gram positive and negative bacteria. In 2015, Huang *et al.* devised the total synthesis of 6-*epi*-Aculeatin D **16** *via* Corey–Fuchs alkyne synthesis, which showcases strong anti-malarial activity with IC$_{50}$ value of 0.18–3 μM against *P. falciparum* (Fig. 1).[27]

The Corey–Fuchs reaction has endured notable breakthrough in the recent past and the need for a thorough and

state-of-the-art review on this topic has gained great prominence. The most recent comprehensive review on this topic dates back to 2015, emphasizing the need for a contemporary assessment of its scope and applications.[28] This manuscript intends to provide a latest (2016 to date) update on the cutting-edge development, showcasing the versatility and potential of this invaluable reaction in natural product synthesis.

2. Review of literature

2.1. Synthesis of polyketide-based natural products

2.1.1. Chatenaytrienin-4 synthesis. Acetogenin membranacin is a naturally occurring polyketide which belongs to a large group of fatty acid type compounds called annonaceous acetogenins. These natural products are characterized by a number of biological active properties such as pesticidal, cytotoxic, anti-malarial, *anti*-feedant, immunosuppressive and most likely, anti-tumor properties.[29] Adrian and Stark[30] in 2016, described modified strategy for the efficient stereodivergent construction of straight chain 1,5,9, n-polyenes and employed this for the first total synthesis of chatenaytrienin-4 **24**, which is the hypothetical biosynthetic forerunner of membranacin **25**. The total synthesis commenced from the unexpensive 1-dodecyne **17** which was subjected to subsequent allylation to yield **18** followed by hydroboration to form bis-homopropargylic alcohol **19** in 70% yield. Next, the alcohol **19** was subjected to PCC/silica mediated oxidation in DCM to furnish aldehyde compound **20** (86%), followed by Corey–Fuchs alkyne synthesis reaction under the presence of CBr$_3$ and PPh$_3$ to furnish a dibromo compound **21** in conjugation with its lithiation and allylation in the presence of TBAI to furnish enediyne **22** in 80% yield. Further, enediyne **22** was subjected to chemoselective hydroboration in the presence of 9-BBN-dimer to deliver alcohol **23** in 63% yield. Carboxylic compound **24** was synthesized from **23** in a few steps followed by its treatment with hydroxy butanolide **25** in the presence of DMAP, ensued by Fries rearrangement and successive selective hydrogenation to deliver compound **26** (90%). Further, the hydroxyl group was protected under the presence of Tf$_2$O followed by Lindlar's catalyst mediated reduction and then Pd-mediated reduction to finally achieve chatenaytrienin-4 **27** (51% over 2 steps) in overall 6% yield in 15 linear steps (Scheme 3).

2.1.2. Tetrangulol synthesis. Tetrangulol **35** is a naturally occurring quinone, which belongs to the group of angucyclines. This group of compounds exhibit a wide range of biological properties such as anti-cancer and anti-bacterial activities.[31] Tetrangulol **35** was synthesized for the first time by Brown & Thomson in 1976.[32] Since then, a number of synthetic methods have been reported on the synthesis of biological active tetrangulol.[33] Owing to high medicinal properties, Ngwira[34] *et al.*, in 2016 reported the synthesis of tetrangulol by employing Suzuki–Miyaura coupling reaction & Corey–Fuchs alkyne synthesis as key synthetic steps. The total synthesis initiated *via* generation of substituted benzaldehyde **30** which is a key precursor, from commercially available 1,5-dihydroxynaphthalene **29** over few steps. Next, the compound **31** was subjected to reductive methylation followed by its reaction with *n*-BuLi
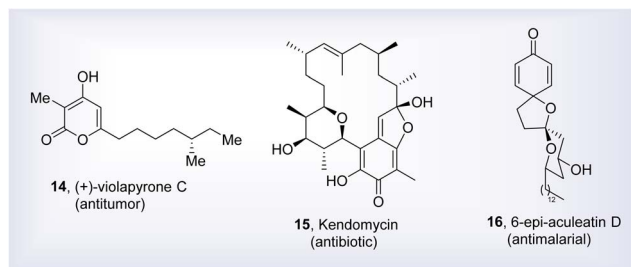

Fig. 1 Structure of some biological important natural products having Corey–Fuchs reaction as main step in total synthesis.

Scheme 3 Total synthesis of chatenaytrienin-4 **27** by Adrian and Stark.

and triisopropylborate to furnish boronic acid **32** (96%). The synthesized acid **32** underwent Suzuki–Miyaura coupling reaction in the presence of Pd(PPh$_3$)$_4$ to access 3-methoxy-5-methyl-2-(1,4,5-trimethoxynaphthalen-2-yl)benzaldehyde **33** in 80% yield. Compound **33** was then subjected to Corey–Fuchs reaction under the presence of CBr$_4$ and PPh$_3$ to furnish dibromoalkene **34** in conjugation with n-BuLi to form desired alkyne compound **35** in 88% yield. In the next step, formation of a mixture of tetracyclic product **36** (61%) and **37** (23%) took

place when compound **35** was reacted in the presence of PtCl$_2$ and PhMe. Both the products were separated *via* chromatography and compound **36** were further subjected to CAN oxidation followed by its treatment with boron tribromide to afford desired tetrangulol **38** in 97% yield (Scheme 4).

2.1.3. Synthesis of C16–C28 fragment of hemicalide. Hemicalide is a naturally occurring compound, fits into the class of polyketides.[35] It is extracted from marine sponge *Hemimycale* sp. in the south pacific and featured with potent
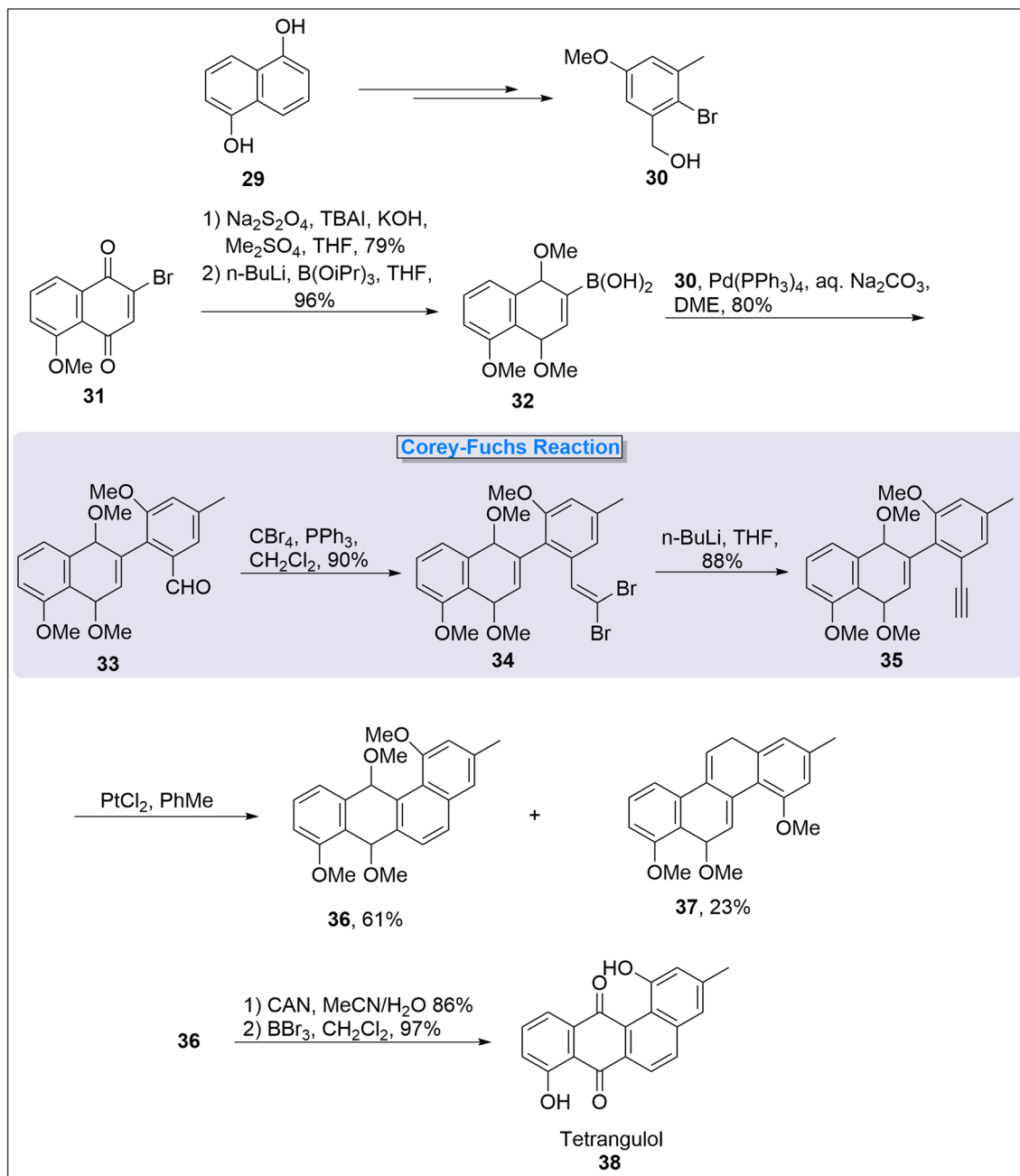
Scheme 4 Total synthesis of tetrangulol **38** by Ngwira and coworkers.

cytotoxic properties. Hemicalide exhibit excellent growth inhibitory potential against human cancer cell lines with IC_{50} values in picomolar, processing *via* tubulin targeting antimitotic mechanism.[36,37] In this regard, MacGregor[36] *et al.*, in 2016 synthesized the advanced C16–C28 fragment **47** of the hemicalide **48** with restructured 18,19-*syn* geometry. The synthesis commenced with the formation of alkyne compound **40** (94%) under the Corey–Fuchs reaction followed by hydrozirconation and iodination in conjugation with subsequent 2,3-dichloro-5,6-dicyano-1,4-benzoquinone (DDQ) induced ether cleavage and oxidation to furnish (*Z*)-vinyl iodide **41** (80%). In the next step, vinyl iodide **41** were subjected to aldol addition

reaction with PMB ether **42** under the conditions of ((−)-Ipc₂BCl, Et₃N) to generate a *syn* adduct **43** in 70% yield with exclusive diastereoselectivity. Next, subsequent formation of TBS ether and enolate addition in the presence of EtOAc and LDA led to the formation of ester **44** (97%), which was further subjected to TBS ether cleavage and induced lactonization in conjugation with elimination to furnish α-lactone **45** in 83% yield. Next, aldehyde compound **46** was achieved in 4 steps including *syn*-dihydroxylation and TES ether generation in the presence of TESOTf and 2,6-lutidine followed by PMB ether cleavage in DDQ and Dess–Martin periodinane (DMP) mediated oxidation. In the next step, Horner–Worsworth Emmons olefination in the

Scheme 5 Total synthesis of C16–C28 fragment of hemicalide **48** by MacGregor and coworkers.

presence (MeO)$_2$P(O)CH$_2$COMe & Ba(OH)$_2$ and Stryker reduction under the conditions of [Ph$_3$PCuH]$_6$ and PhMe subsequently led towards the synthesis of completed C16–C28 fragments **47** in 67% yield (Scheme 5).

2.1.4. Actinoallolide synthesis. Actinoallolides belong to the class of 12 or 14-membered macrolides, derived from culture medium of *Actinoallomurus fulvus* MK10-036.[38] These compounds showcase a number of activities, such as *in vitro* anti trypanosomal potential against *Trypanosoma brucei brucei* GUTat 3.1 strain which is a bioweapon of Nagana disease in animals, *Trypanosoma brucei rhodesiense* which is a bioweapon of African *Trypanosomiasis* and *Trypanosoma cruzi Tulahun* strain which cause Chagas disease.[38] Actinoallolide A **61** possessed most of the potent activities such as *anti*-trypanosomal activity against *T. b. rhodesiense* STIB900 strain and also exhibit cytotoxic properties against MRC-5 cell lines and MIC; >10 µg per paper disk against Gram-negative and Gram-positive

bacteria.[38] Since macrolides depict a variety of biological activities, actinoallolides are first naturally occurring macrolides possessing *anti*-trypanosomal properties. In 2016, Ohista[39] *et al.*, reported the construction of the key intermediate (+)-**60** towards the convergent total synthesis of **61**. The synthesis commenced from the synthesis of right part (–)-**54** of the compound (+)-**60**. For this purpose, the compound **49** was transformed into asymmetric alcohol **50** (78%) *via* protection of hydroxyl group in the presence of MsCl, DMAP and pyridine followed by its treatment with cesium acetate under the presence of 18-crown-6, in conjugation with DIBAL-H mediated reduction. Next, alcohol **50** was subjected to acetalization followed by ozonolysis to furnish aldehyde compound **51** (79%) which was further reacted under the Corey–Fuchs reaction conditions to furnish alkyne **52**. Next, PMP acetal ring opening furnished ketone **53** in 4 steps. The hydrozirconation of compound **53** followed by TIPS protection of hydroxyl group

finally furnished first precursor (−)-**54** (97%) for the synthesis of intermediate (+)-**60**. Next, the other coupling partners *i.e.*, *E*-vinyl stannane (+)-**56** and alkyne compound (+)-**58** were also synthesized from commercially available mono-methyl hydroquinone **55** and methyl-3-hydroxy methyl propanoate **57** respectively over a few steps. Next, the construction of main

carbon skeleton of the key intermediate (+)-**60** was obtained from above synthesized precursors *i.e.*, (−)-**54** and (+)-**58**. As cross coupling reactions facilitate the robust synthesis of C–C and C–heteroatom bonds, accelerating the synthesis of coupled products.[40,41] For this purpose, compound (+)-**58** was subjected to Negishi cross coupling with (−)-**54** to generate coupling
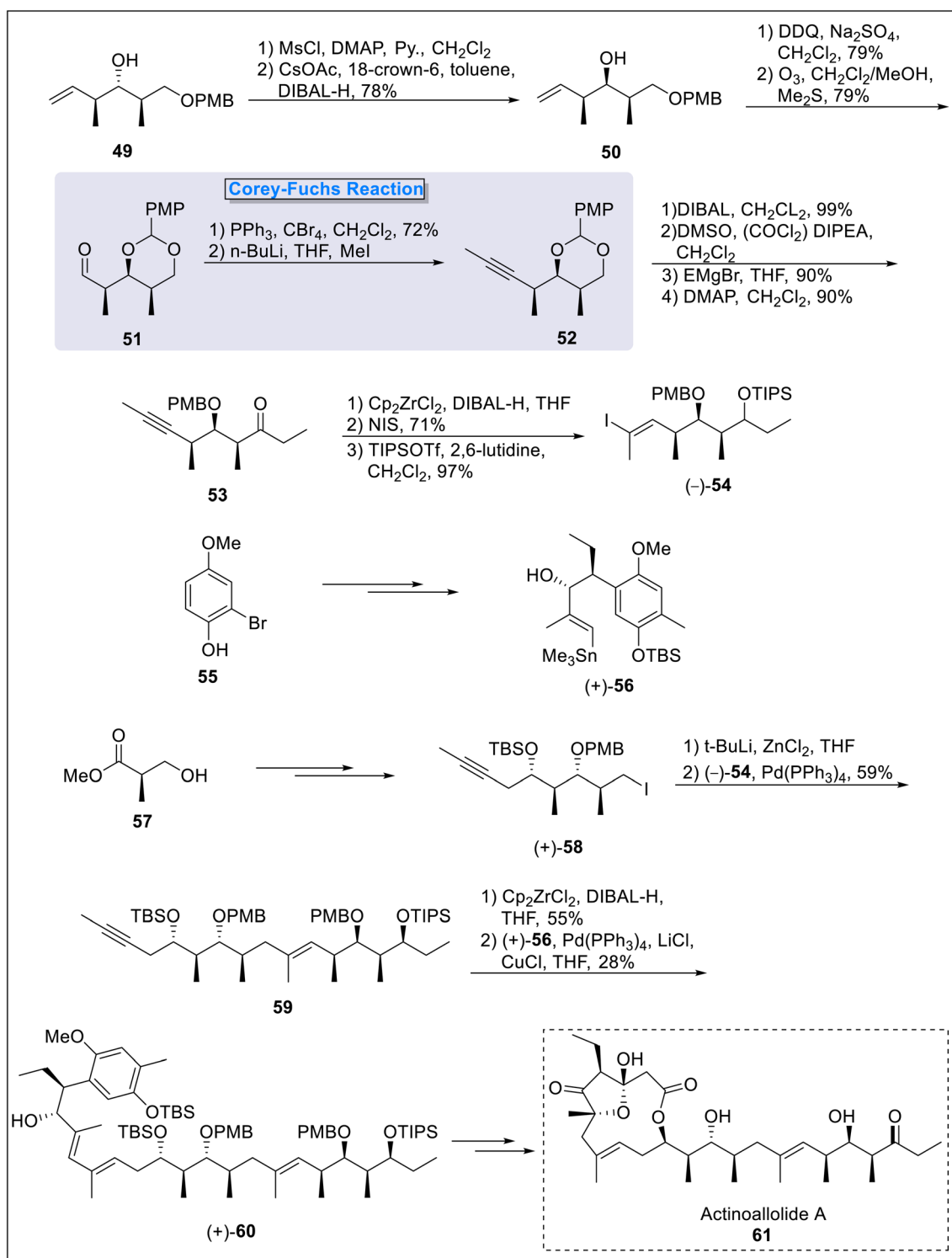
Scheme 6 Total synthesis of key intermediate of the actinoallolide A (+)-**60** by Ohista and coworkers.

product **59** (59%). In the next step, the compound **59** underwent hydrozirconation followed by Stille coupling with (+)-**56** to finally furnish (+)-**60** in 28% yield (Scheme 6).

2.1.5. Orevactaene and epipyrone synthesis. Orevactaene **74** and epipyrone A **76** are significant naturally occurring scaffolds, comprised of 4-hydroxy-2-pyrone core structure.[42] Both compounds were extracted from *Epicoccum purpurascens* strains and featured with a number of biological actives properties such as orevactaene depicts HIV-1 inhibitory and telomerase inhibitory properties.[42] Whereas epipyrone A showcase potential against the influenza A virus and also act as a strong fungicide.[43] Owing to the unique structural and biological properties, Preindl[44] *et al.*, in 2017 devised total synthesis of these intriguing moieties *via* a number of key steps such as Corey–Fuchs reaction, two alkyne cycloisomerization reaction and Stille coupling. The total synthesis began with the generation of first precursor **63** from commercially available D-arabinose **62** over a few steps. In the next step, alcohol **64** was transformed into propargyl alcohol **65** (51% over 3 steps) *via* subsequent oxidation and Corey–Fuchs reaction in conjugation with the treatment of resulting alkyne with paraformaldehyde. Compound **65** was then reacted with ethyl trifluoropyruvate **66** followed by hydroboration in the presence of the oxyboran succeeded by subsequent oxidation in the presence of trimethylamine *N*-oxide and Suzuki-type cross-coupling with 1-iodopropyne under the conditions of Pd catalysis furnished compound **67** (57% over 3 steps). Further, compound **67** was reacted with PhMe$_2$SiLi to form silyl compound **68** (90%) which was converted to compound **69** over 4 steps yielding 89%. In the next step, commercially available epichlorohydrin **70** was subjected to alkylation under the presence of sodium acetylide in basic conditions followed by stannylcupration in conjugation with oxidation to yield **71**. Next, Wittig olefination, reduction, oxidation and bora-Wittig reaction subsequently delivered heterodimetalated tetraene **73** (84%). Next, compound **73** was subjected to Stille coupling with compound **69** in the presence of Pd catalysis followed by Suzuki coupling reaction with compound **63** that resulted in formation of core structure of orevactaene. Next, deprotection using TSAF ultimately furnished desired **74**. Apipyrone A **76** was also synthesized in the same manner from galactopyranosyl fluoride **75** over a few steps (Scheme 7).

2.1.6. (−)-Dactylolide synthesis. (−)-Dactylolide **85** is a naturally occurring compound fits into the class of cytotoxic macrolides, derived from *Dactylospongia* sp. of Vanuatu sponge.[45] In 2018, Tanaka[46] *et al.*, published the convergent total synthesis of (−)-dectylolide **85**, centered around the inter- and intramolecular allylation strategies. The synthesis commenced with the generation of key reactant **80** from known alcohol **77** *via* protection in the presence of MOMCl/iPr$_2$NEt/DMAP succeeded by the DIBAL-H mediated reduction. Next, treated it with CBr$_4$ and PPh$_3$ to furnish dibromoalkene **78** *via* Corey–Fuchs reaction in conjugation with its reaction in the presence of *n*-BuLi and then ethyl chloroformate to generate ester compound **79** (88%). Precursor **80** was afforded from compound **79** over a few steps. Next, the other reacting partner **82** was also generated over a few steps from a known alcohol **81**. Both the reacting partners **80** & **82** in hand, the total synthesis of

(−)-dactylolide **85** was performed under various reaction conditions. First, both the reactants *i.e.*, **80** & **82** were reacted *via* Mukaiyama aldol condensation in the presence of CMPI followed by its reduction with DIBAL-H in conjugation with treatment with (CH$_2$ClCO)$_2$O, pyridine and DMAP to afford α-acetoxy ether **83**. Next, compound **83** was subjected to react with ZnBr$_2$.OEt$_2$ and MS5A succeeded by its reaction with TMSBr/TBAI in conjugation with MnO$_2$ *via* oxidation (73% over 2 steps) and then Pinnick oxidation to access seco-acid **84** (99%). The compound **84** was subjected to react with MNBA and DMAP succeeded by its deprotection using TBAF in conjugation with Dess–Martin oxidation to synthesize (−)-dactylolide **85** in 88% yield (Scheme 8).

2.1.7. Synthesis of polyketide fragment of seragamide. Seragamides are naturally occurring compounds featured with excellent cytotoxic properties.[47] These compounds were isolated from marine sponge *Suberites japonicus*.[48] Lang and Lindel[49] in 2019, described the total synthesis of the polyketide fragment of the Seragamide employing Corey–Fuchs reaction as a main step. The synthesis involved nine overall steps with 21% yield. The total synthesis commenced from the preparation of organozinc homoenolate **87** which was utilized as Negishi coupling partner, from the treatment of β-bromopropionic acid **86** in the presence of manganese bromide, copper chloride and di ethyl zinc. Next, *tert*-butyl ester **89** was synthesized from *tert*-butyl propionate **88** over a few steps. Compound **89** was then reduced in the presence of DIBAL followed by DMAP-catalyzed oxidation to obtain aldehyde **90**. In the next step, aldehyde **90** was subjected to Corey–Fuchs reaction under the presence of carbon tetrabromide and triphenyl phosphine to form dibromoalkene **91** succeeded by its reaction with LDA and then methyl iodide to form internal alkyne **92** in 91% yield (over 2 steps). Next, compound **92** underwent subsequent hydrozirconation and iodination under the presence of Schwartz reagent and iodine to form (*E*)-olefin **93** in 84% yield (over 2 steps) with exclusive regio- and stereo-selectivity. Compound **93** underwent Negishi coupling with homoenolate **87** under the presence of palladium catalysis in DCM to furnish protected nonenoic acid **94** in 85% yield. In the final step, PMP deprotection of **94** in the presence of CAN afforded fragment **95** in 82% yield. Next, tripeptide **97** was subjected to BEP-catalyzed reaction with compound **96** (synthesized from polyketide section **95** *via* saponification), to form peptide-polyketide open chain compound **98** in 60% yield (over 2 steps) with 9 : 1 ratio of diastereoselectivity. This open chain peptide-polyketide compound **98** was further transformed into natural product seragamide A **99** over few steps (Scheme 9).

2.1.8. (−)-Enigmazole A and (−)-15-*O*-methylenigmazole synthesis. (−)-Enigmazole A **110** and its natural chemical analog (−)-15-*O*-methylenigmazole **114** fits into the family of phosphomacrolides derived from marine sponge *Cinachyrella enigmatica* of New Guinean Papua.[50] These naturally occurring compounds are featured by 18-membered macro-lactone possessing biological active properties. (−)-Enigmazole A **110** exhibit cytotoxic activity against the NCl 60 cell line, with a GI$_{50}$ value of 1.7 μM.[51] In 2020, Sakurai[52] *et al.*, reported the total synthesis of (−)-enigmazole A **110** and (−)-15-*O*-methyl-enigmazole **114** by employing a number of key reaction
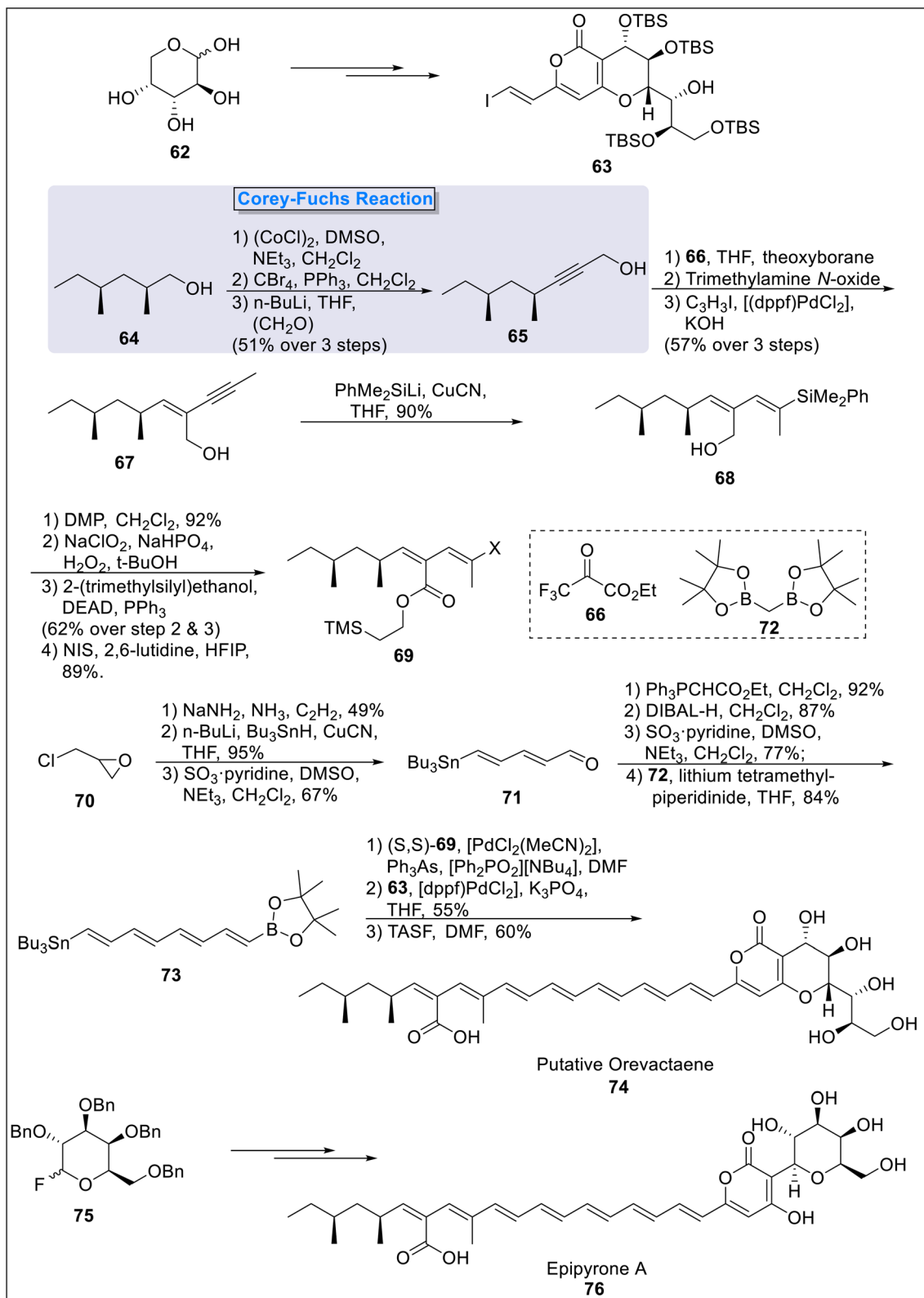
Scheme 7 Total synthesis of putative orevactaene **74** & epipyron A **76** by Preindl and coworkers.

procedures and Corey–Fuchs alkyne synthesis reaction is one of them. The total synthesis commenced with protection of homoallylic alcohol **100** to generate TBS protected ether **101**

(97%), which on oxidative cleavage, furnished aldehyde compound **102**. The aldehyde **102** underwent two steps Corey–Fuchs reaction to access alkyne **104** in 94% yield in the presence
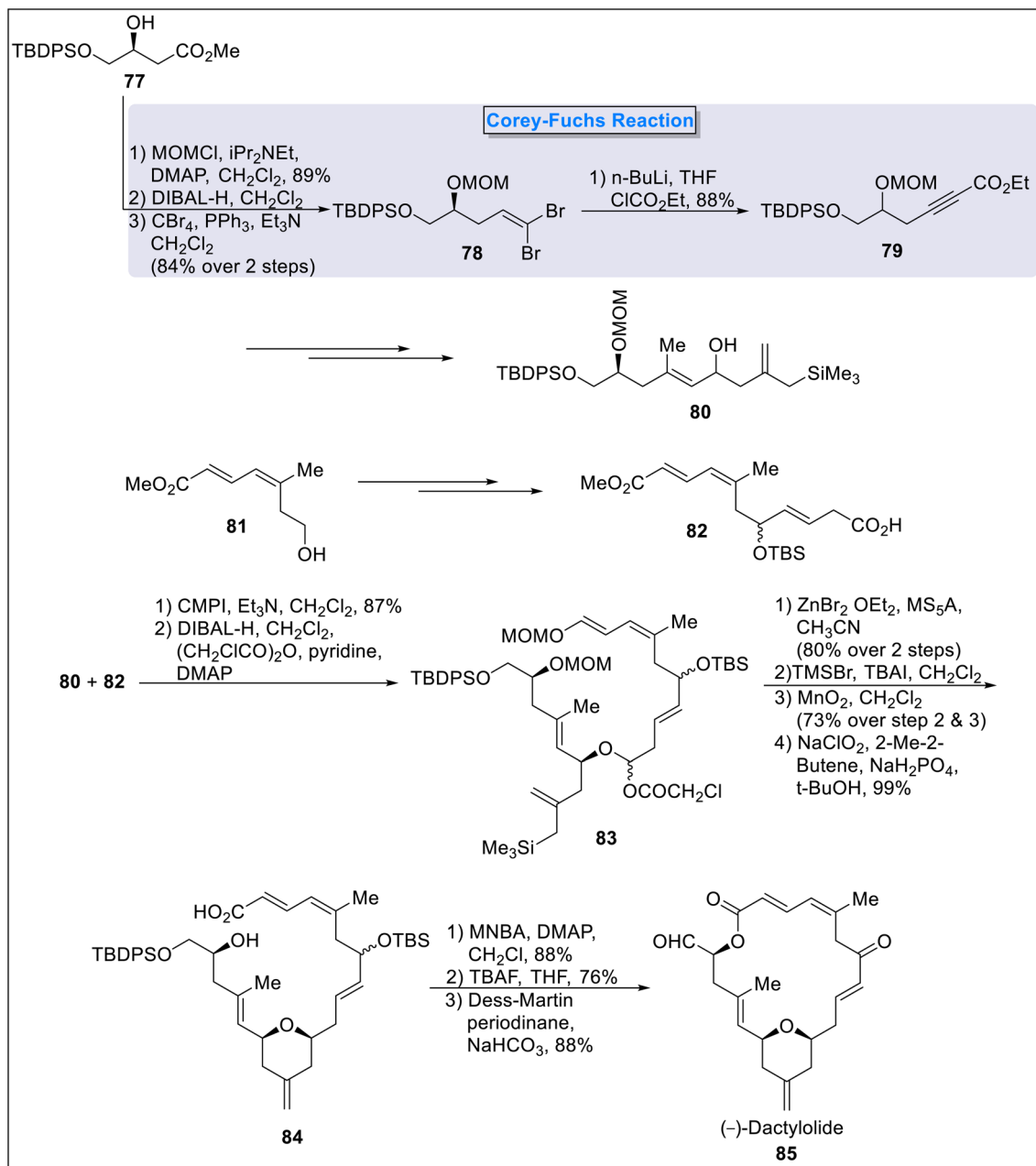
Scheme 8 Total synthesis of (−)-dactylolide **85** by Tanaka and coworkers.

of standard Corey–Fuchs conditions. Next, alkyne **104** was deprotected in the presence of n-BuLi followed by acetylide trapping with acetaldehyde succeeded by benzoyl chloride/pyridine mediated acylation and then TBAF catalyzed deprotection generated compound **105** (92% over 2 steps). In the next step, compound **105** was made to react with the synthesized carboxylic acid **106** in the presence of 3,4,6-Cl$_3$C$_6$H$_2$COCl, Et$_3$N and DMAP (Yamaguchi conditions) to furnish ester compound **107**. Finally, (−)-enigmazole A **108** was synthesized from ester **107** over a few steps. The total synthesis of (−)-15-O-methyl-enigmazole **109** was also accessible from ester compound **107**, which was transformed into natural product **109** over a few synthetic steps (Scheme 10).

2.1.9. Metacridamide B synthesis. Metacridamide B **120** fits into the class of macrolides, extracted for the first time from conidia of the entomopathogenic fungus *M. acridum* By Krasnoff *et al.*, in 2012.[53] It is a 17 membered macrocyclic secondary metabolite featuring biological active property *i.e.*, anti-cytotoxicity with IC$_{50}$ of 18.2 mM against HepG2/C3A.[53] Due to its unique structural architecture with remarkable biological potential, researchers explore various synthetic pathways towards its total synthesis.[54] In this regard, Sharma and coworkers[55] in 2021 documented the total synthesis of meta-cridamide B in overall 27 steps. The key methodological steps involved Yamaguchi macrolactonization, Corey–Fuchs alkyne synthesis, Ru-catalyzed alkyne functionalization, Stille coupling
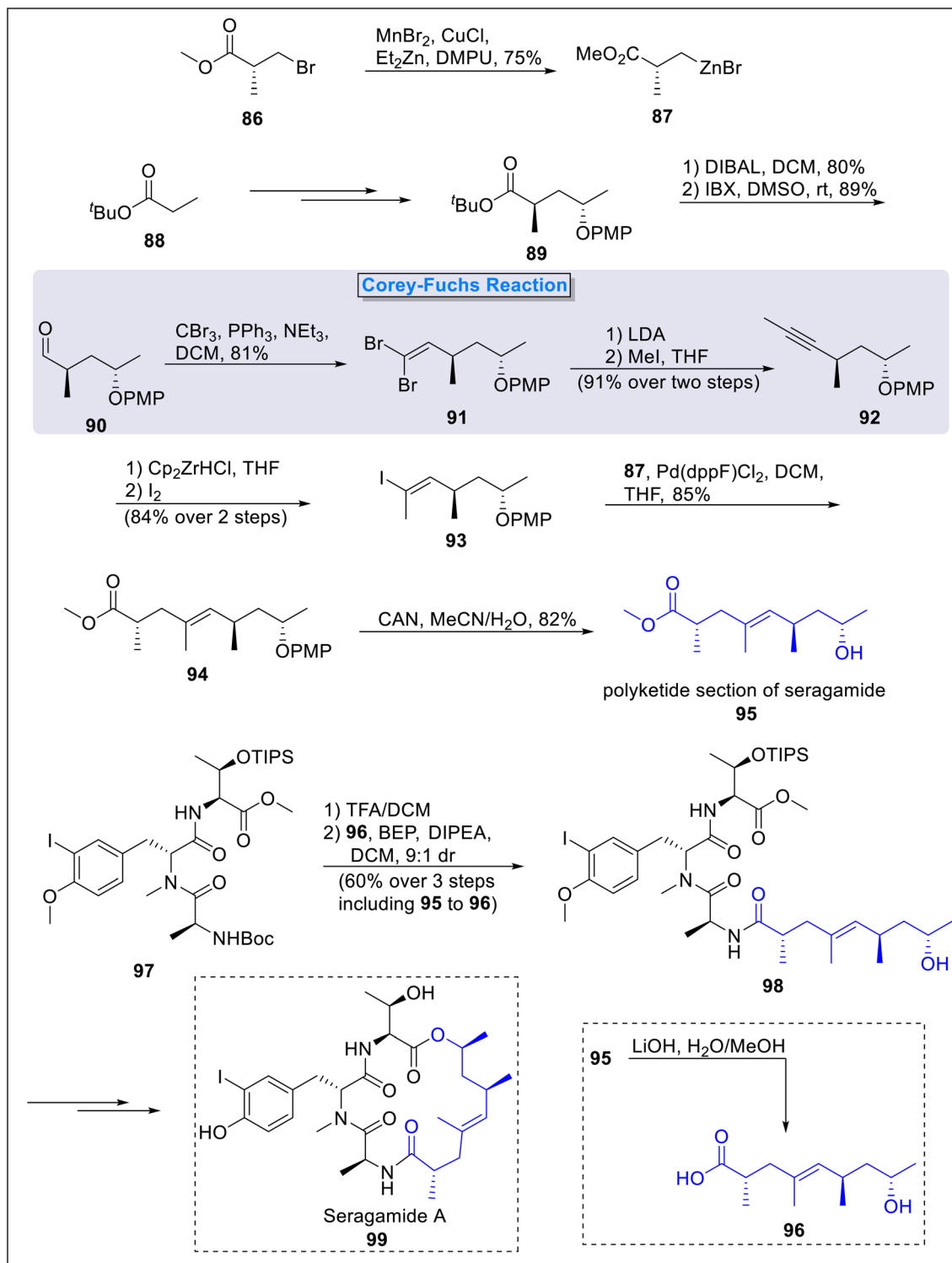
Scheme 9 Total synthesis of polyketide fragment of the natural product seragamide A **99** by Lang and Lindel.

and Micalizio coupling. The total synthesis was proceeded with the synthesis of a key intermediate **111**. The alkyne moiety **111** (79%) was accomplished from primary alcohol **110** in two reaction steps including DMP oxidation and Corey–Fuchs alkyne synthesis reaction in the presence of TPP, CBr$_4$ in conjugation with the use of *n*-BuLi. The synthesis of the second reactant

aldehyde compound **114** was accomplished over few steps starting from an alkyne **112** and aldehyde compound **113**. Next, both reacting entities in hands, coupling of **111** & **114** took place to yield enantiopure secondary alcohol **115** (80%) in three steps involving reaction with *n*-BuLi, DMP oxidation and asymmetric reduction in the presence of (*S*)-CBS catalysis. Secondary alcohol
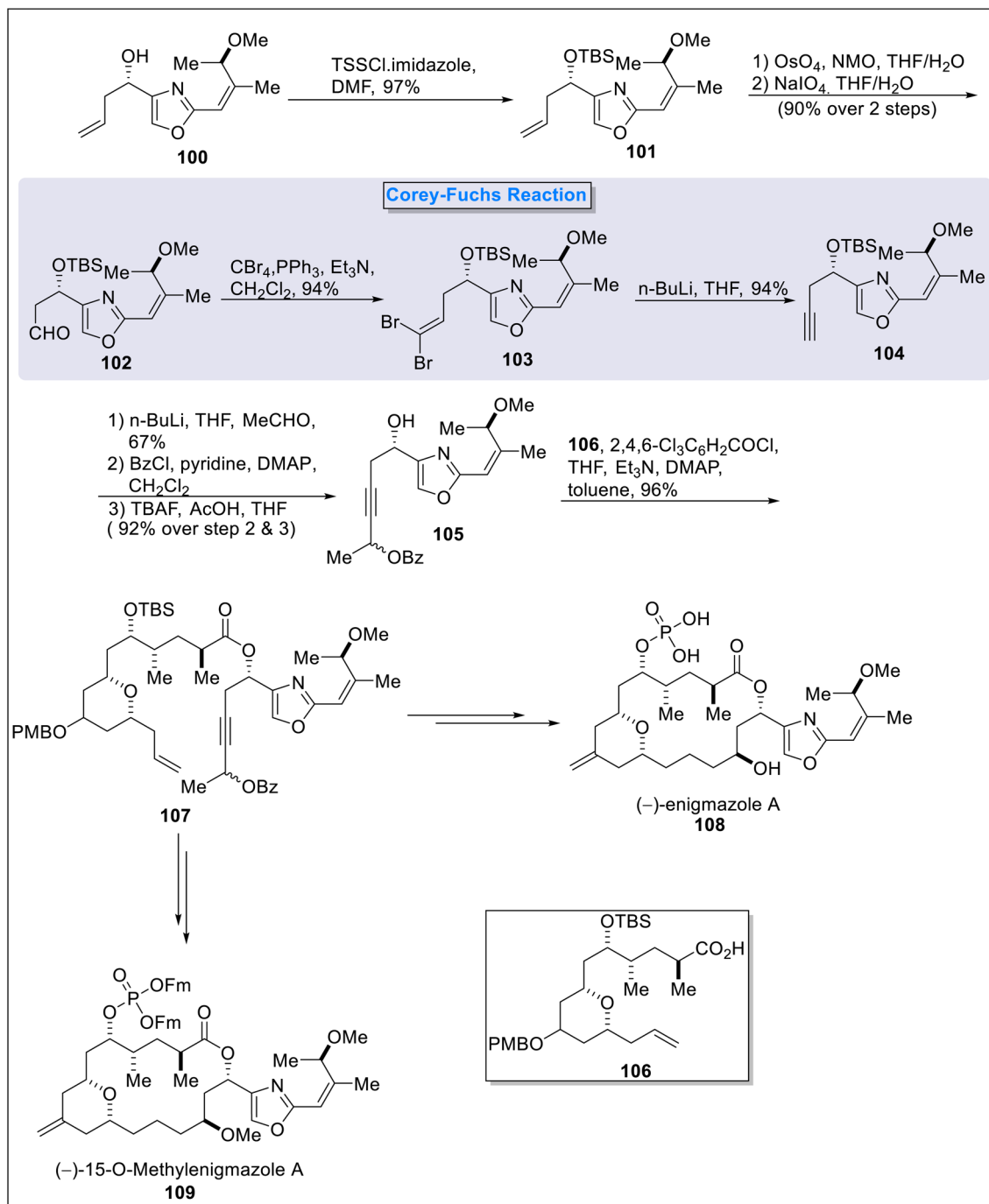
Scheme 10 Total synthesis of (−)-enigmazole A **108** & (−)-15-*O*-methylenigmazole **109** by Sakurai and coworkers.

115 was subjected to regioselective functionalization under the presence of Bu₃SnH and ruthenium catalyst followed by its treatment with NIS in conjugation with Stille coupling in the presence of CuTC, Pd[PPh₃]₄ and [Ph₂PO₂][NBu₄] to afford compound **116** in 84% yield. Further, protection of free hydroxyl with TBS group took place followed by selective deprotection in the presence of CSA (catalyst). Next, DMP oxidation was followed by Pinnick oxidation to furnish intermediate **117** in 77% yield over 2 steps. Intermediate **117** was further treated with L-

phenylalanine methyl ester **118** under the presence of 2-(7-aza-1*H*-benzotriazole-1-yl)-1,1,3,3-tetramethyluronium hexa-fluorophosphate (HATU) followed by PMB deprotection with DDQ to form alcohol compound **119** in 94% yield. In the final step, compound **119** was subjected to ester hydrolysis in the presence of Me₃SnOH succeeded by Yamaguchi macro-lactonization to generate TBS protected metacridamide B which on HF-pyridine catalyzed silyl deprotection gave metacridamide B **120** in 50% yield over 2 steps (Scheme 11).
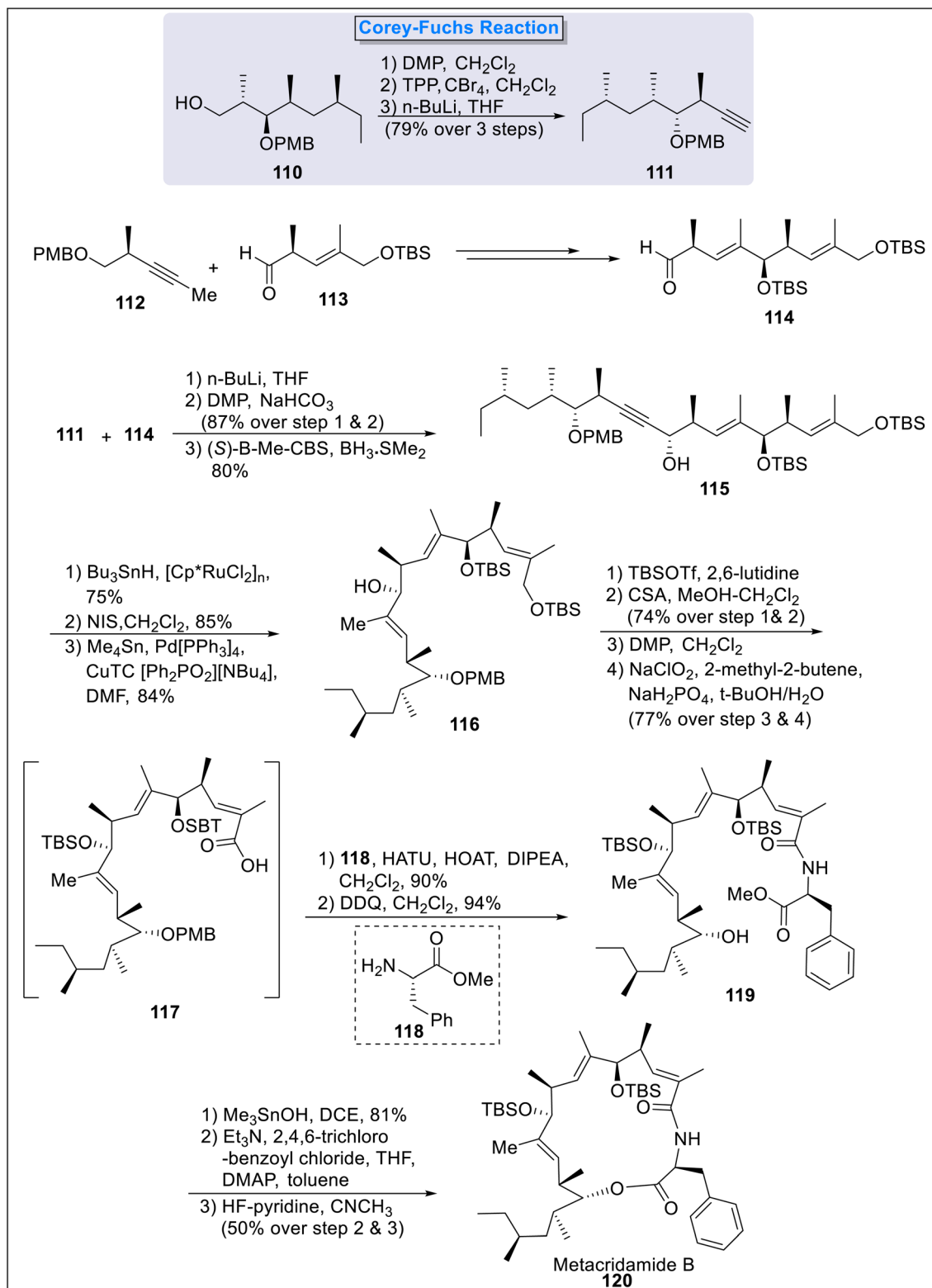
Scheme 11 Total synthesis of metacridamide B **120** by Sharma and coworkers.

2.1.10. Veramycin A, NFAT-133 & TM-123 synthesis. Veramycins along with their chemical analogues NFAT-133 and TM-123 are well known polyketides, were derived from culture broth of *Streptomyces* sp. from Sanofi microbial strain. These natural products showcased an enhanced baseline deoxy glucose uptake in a tailored L6 rat skeletal muscle cell line.[56] In 2022, Dardić[57] *et al.*, reported a detailed total synthesis of α-pyrone featured veramycin A **135** and its congeners NFAT-133 **126** and

TM-123 **132**. The synthesis was commenced with the formation of Weinreb amide intermediate **123** (95%) from (S)-aldehyde **121** in two steps involving the aldol addition with benzyl-3-propionyloxazolidinone **122** in the presence of Bu_2BOTf and NEt_3 followed by oxazolidinone cleavage. Next, compound **123** was coupled with pinacol boronate **124** via Suzuki coupling in the presence of XPhos Pd G4 (as catalyst) to afford **125** in 96% yield. Compound **125** was efficiently converted to NFAT-133 **126** in two steps entailing the formation of methyl ketone in the presence of Grignard reagent in conjugation with deprotection of TBS group in 25% overall yield. Next, to accomplish the synthesis of veramycin A **135** and TM-123 **132**, the intermediate **125** was converted into aldehyde **127** (97% yield) via TBS protection of hydroxyl group, succeeded by reduction in the presence of DIBAL-H. The prepared aldehyde **127** underwent two step Corey–Fuchs reaction in the presence of CBr_4, PPh_3, and NEt_3 to yield dibromoalkene **128** followed by its treatment with butyl lithium and methyl chloroformate to furnish propiolic acid methyl ester **129** in 76% yield. Next, the ester **129** was reacted with t-butyl ester **88** & TMSE protected ester **132** to furnish cyclized intermediates **130** (84% yield) and **133** (94% yield) correspondingly under the Claisen reaction conditions. These cyclized precursors were then transformed into their corresponding pyrones i.e., veramycin A **134** and TM-123 **131** in the presence of $(SPhos)AuNTf_2$ (gold catalyst) and $MeNO_2/AcOH$ followed by deprotection of alcohols (in 16% and 12% overall yield respectively) (Scheme 12).

2.1.11. (±)-Parvistilbine B & (±)-stemenone B synthesis. (±)-Parvistilbine B **149** and (±)-stemenone B **148** are naturally occurring compounds that fit into the class of polyketides and extracted from the *Stemona parviflora* and *Stemona tuberosa* respectively, exhibiting profound historical roots in Chinese phytomedicines.[58] Parvistilbine B was extracted from the same room which exhibit excellent nematocidal activity while (±)-stemenone B has anti-inflammatory potential with IC_{50} value of 2.97 μM.[59] In 2022, Kozlowski[60] et al., first proposed a new dearomative oxidation strategy for the construction of para-quinol moiety which is the core structure of both abovementioned natural compounds, then performed the total synthesis of these compounds. The main steps involving the total synthesis were Corey–Fuchs alkyne synthesis and Sonogashira-hydrogenation. The total synthesis began with the synthesis of alkyne precursors **137** and **140** from commercially available o-anisaldehyde **135** and p-hydroxy-2-anisaldehyde **136** respectively via Corey–Fuchs reaction. Next, the second coupling partner **144** was synthesized from dimethyl resorcinol **141** which was subjected to debromination to synthesize **142** (99%) succeeded by subsequent methylation and debromination to achieve **143** in 99% yield. Next, the compound **143** was subjected to Li–halogen exchange reaction and selective deprotection to furnish compound **144** with 55% yield. All the coupling partners **137**, **140** & **144** in hand, Sonogashira coupling of **144** with **137** and **140** led to the formation of compounds **145** (51%) and **146** (45%) respectively, followed by hydrogenation–debenzylation and photo-catalytic dearomatization strategy to construct (±)-stemenone B **147** (86% over 2 steps) and (±)-parvistilbine B **148** (90% over 2 steps) respectively (Scheme 13).

2.1.12. Aigialomycin D synthesis. Aigialomycin D **160** is a fungal polyketide, derived from the marine mangrove fungus *Aigialus parvus* BCC 5311. This is a 14 membered resorcyclic acid lactone, featuring a number of bioactive properties including anti-malarial activity (IC_{50}: 6.6 μg mL^{-1} for *P. falciparum*) and cytotoxicity in human cell lines (IC_{50}: 3.0 & 18 μg mL^{-1} in KB and BC-1 cells respectively).[61] Total synthesis of aigialomycin D was disclosed for the first time in 2004 by Danishefsky and coworkers in 18 steps and with 8% overall yield.[62] In continuation of exploring the new ways towards the total synthesis of bioactive natural products, Sudhakar[63] et al., in 2024 published the total synthesis of aigialomycin D **160**. The key reactions in synthetic approach entailed Corey–Fuchs alkyne synthesis, Yamaguchi esterification and ring closing metathesis. The synthesis was initiated with epoxide **149**, which was subjected to ring opening reaction with homoallyl magnesium bromides under the conditions of dry ether to generate an alcohol.[64] The synthesized alcohol on reaction with BnBr and NaH furnished benzyl ether **150** in 86% yield. Next, benzyl ether **150** was transformed into diol under the acidic medium (70% aq. acetic acid) which on reaction with BzCl and Et_3N got converted to monobezoate **151** in 81% yield. On treatment with TBSCl and imidazole, a TBS protected product was furnished which subjected debenzoylation in the presence of K_2CO_3 followed by Swern oxidation to form respective aldehyde **152**. Aldehyde **152** underwent Corey–Fuchs reaction in the presence of CBr_4 and PPh_3 to access corresponding alkyne **153** in 77% yield. On treatment with n-BuLi, resulting acetylenic anion reacted with (S)-propylene oxide followed by its reaction with red-Al to give intermediate **154** in 74% yield. In the next step, the synthesis of another key precursor **157** took place from 2,4-dimethoxybenzoic acid **155**, which was converted into aldehyde compound **156** in 2 steps including the synthesis of amide in the presence of thionyl chloride and its conversion to aldehyde **156** in the presence of n-BuLi and DMF. Next, Wittig olefination of aldehyde **156** consequently led to the formation of styrene derivative which on hydrolysis gave **157** in 82% yield. The prepared intermediate **157** subjected to Yamaguchi reaction to form an anhydride which on further condensation with above synthesized intermediate **154** in the presence of DMAP delivered ester **159** in 72% yield. Finally, the ester **159** transformed into aigialomycin D **160** (74% & 2.67% overall yield) in two steps entailing the reaction of **159** with Grubb's 2nd generation catalyst in conjugation with deprotection step in the presence of AlI_3 in benzene (Scheme 14).

2.2. Synthesis of alkaloid-based natural products

2.2.1. Synthesis of C22–C40 fragment of azaspiracids. Alkaloids are a diverse class of naturally products, comprise of millions of biologically active compounds.[65] Azaspiracids are naturally occurring compounds fits into the category of lipophilic algal toxins (originally alkaloids), extracted from the dinoflagellates *A. spinosum* and *A. poporum*.[66] These azaspiracids are responsible for various biological responses in higher animals as its consumption by human through shellfish may cause diarrhetic shellfish poisoning.[67] Having intricate
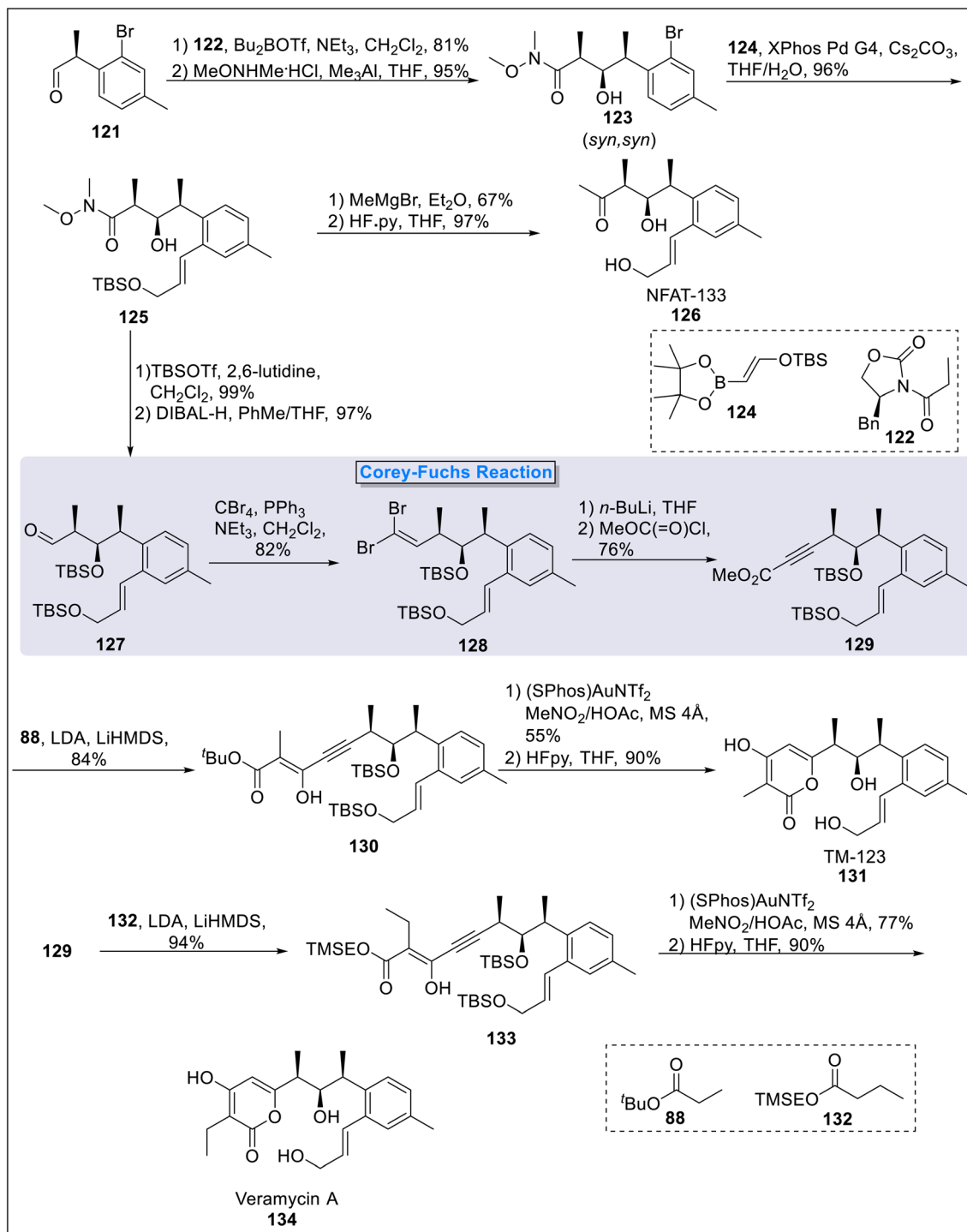
Scheme 12 Total synthesis of veramycin A **134**, NFAT-133 **126** & TM-123 **131** by Dardić and coworkers.

structural features along with bioactive properties, synthesis of these compounds has gained much interest. In this regard, Zhang[68] *et al.* in 2016 demonstrated the total synthesis of C22–C40 region **173** of the azaspiracids utilizing Corey–Fuchs reaction, Nozaki–Hiyama–Kishi coupling and intramolecular-hetero-Michael addition reaction as main steps. The total synthesis commenced from the synthesis of TMS alkyne fragment **166** involving the anti-aldol reaction of (*S*)-lactate ketone

161 with easily available aldehyde **162** in the presence of c-Hex$_2$BCl to access β-hydroxy ketone **163**. Next, compound **163** was first protected and then reduction of keto-ester took place with the assistance of lithium borohydride to furnish **164** (78%) followed by sodium periodate mediated oxidative cleavage to furnish aldehyde compound **165**. Next, the synthesis of TMS protected alkyne **166** (81%) was took place utilizing the Corey–Fuchs reaction followed by the silyl protection. Next,
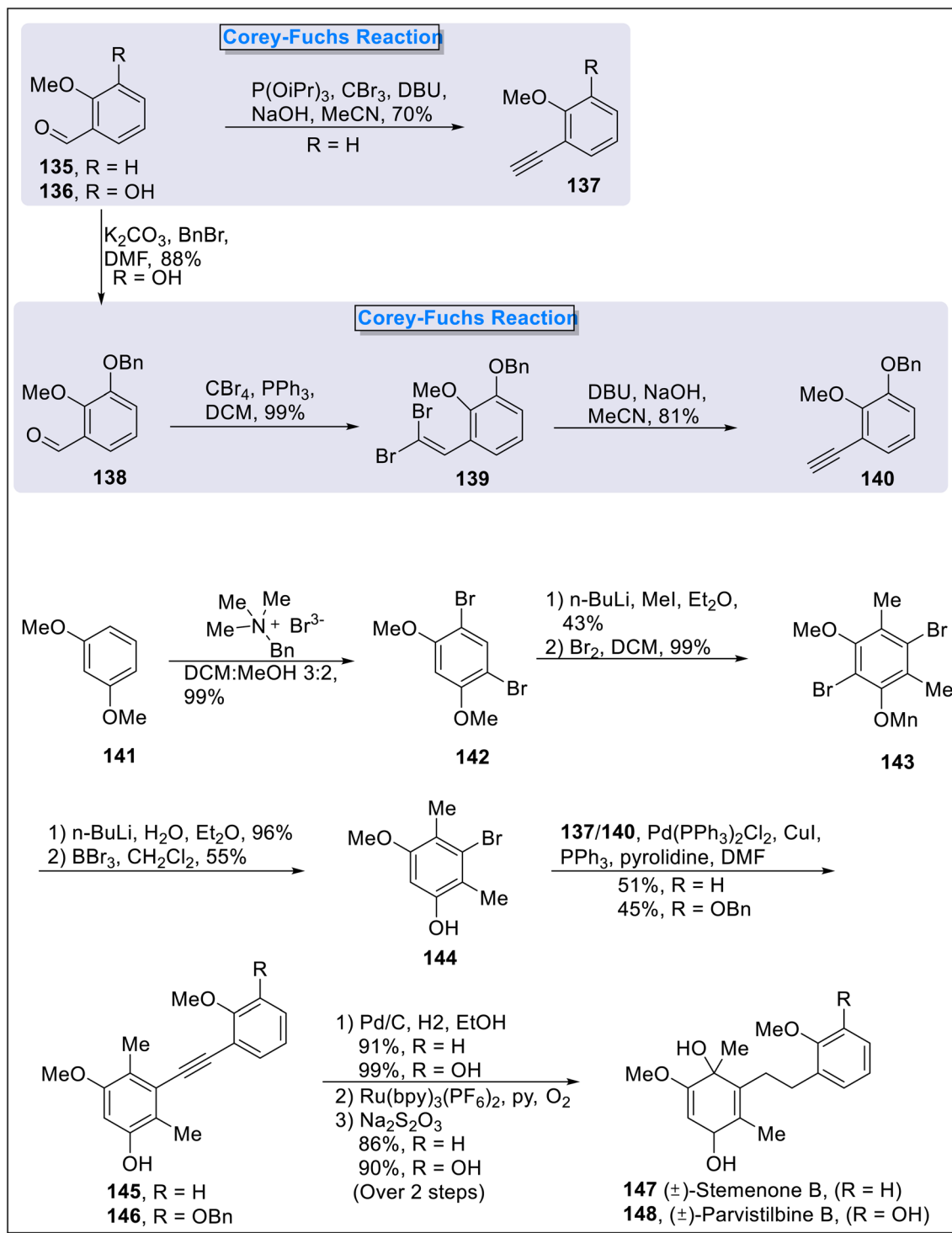
Scheme 13 Total synthesis of (±)-parvistilbine B **148** & (±)-stemenone B **147** by Kozlowski and coworkers.

deprotection of the TBS group led the synthesis of compound **167** (90%), followed by its conversion to epimeric acetal **168** in the presence of DDQ in DCM. Epimeric acetal **168** underwent DIBAL-H mediated reduction followed by the Parikh–Doering oxidation to achieve compound **169** in 95% yield. Next, compound **170** was treated in the presence of NIS and AgOTf in DMF to prepare iodo-alkyne followed by Nozaki–Hiyama–Kishi

coupling with **169** to furnish propargylic alcohol **171** in 95% yield. Next, acyclic intermediate **172** was generated over a few steps. The acyclic intermediate **172** underwent ring closing reaction over a few steps to finally generate C22–C40 fragment **173** of the naturally occurring azaspiracids (Scheme 15).

2.2.2. Tetradecapentaenoic acid synthesis. Tetradeca-pentaenoic acids, a class of poly-unsaturated fatty acid amides
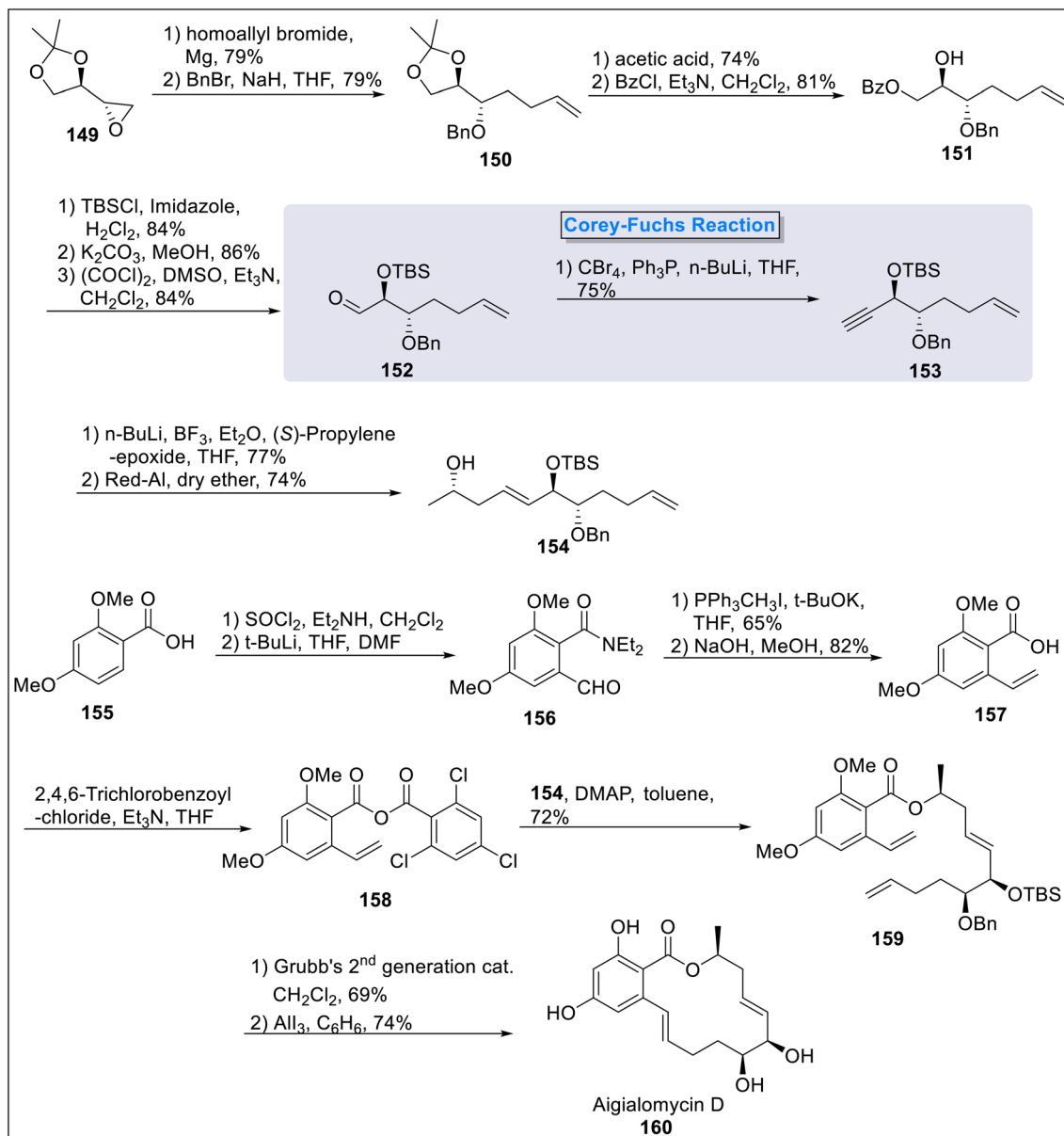
Scheme 14   Total synthesis of aigialomycin D **160** by Sudhakar and coworkers.

present in many tropical plant species such as *Z. bungeanum*, *Z. piperitum* and *Z. ailanthoi*.[69] Until now, more than 50 compounds of poly-unsaturated fatty acids have been extracted from Zanthoxylum plant species displaying remarkable bioactivities. Amides of these fatty acids depict *anti*-helmintic, antioxidant, apoptotic and anti-proliferative potential. Tetradecapentaenoic acid amide derivative **178** also termed as sanshool derivative, extracted from *Zanthoxylum bungeanum*.[70] Sanshool derivatives exhibit promising biological active properties as they act as power activator for cell apoptosis and also depict anticancer properties[71] as cancer continues to pose as a causative agent for death worldwide.[72] In 2019, Kolodiazhna & Kolodiazhnyi[73] demonstrated the total synthesis of sanshool derivative **178** *via* employing Corey–Fuchs reaction as the key step. The total synthesis commenced with the reaction of known

unsaturated aldehyde **174** in the presence of dibromomethylene phosphorane (generated *via* reaction of CBr$_4$ and PPh$_3$) to furnish dibromoalkene **175**. Next, dibromoalkene **175** was treated with *n*-BuLi to form terminal alkyne **176** followed by its reaction with oxalyl dichloride in conjugation with triphenyl phosphine to form unsaturated ester compound **177**. In the final step, desired tetradecapentaenoic acid amide **178** was obtained, upon treatment of ester compound **177** with *n*-BuLi (Scheme 16).

Same group of scientists[74] also represented the stereoselective synthesis of tetradecapentaenoic acid derivatives *i.e.*, **178** & **186**. The key steps in the synthesis involved Wittig reaction, Ramirez–Corey–Fuchs reaction and Trost–Kazmaier rearrangement. The synthesis began with the synthesis of sorbaldehyde **180** from hexadienol **179** *via* Parikh–Doering
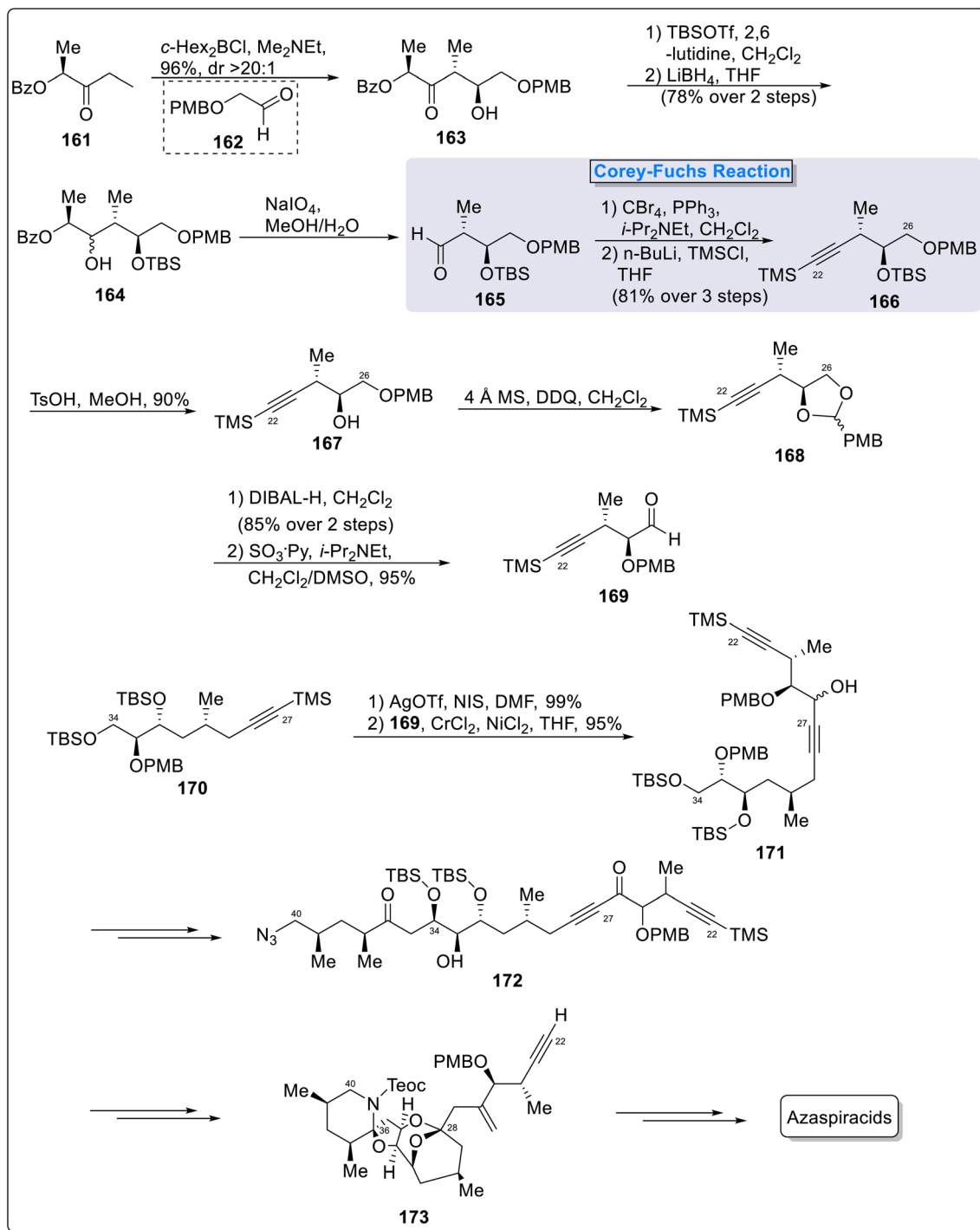
Scheme 15 Total synthesis of C22–C40 fragment **173** of azaspiracids by Zhang and coworkers.

oxidation in the presence of pyridine sulfur trioxide complex. Next step involved the synthesis of second reactant *i.e.*, phosphonium salt **182** which was generated from aldehyde compound **181** in two steps. The above synthesized compound **180** underwent Wittig olefination reaction with phosphonium compound followed by lithium aluminum hydride mediated reduction to generate alcohol **184**. Next, Parikh–Doering oxidation of alcohol **184** generated aldehyde compound **174**

which was subjected to two steps Corey–Fuchs reaction in the presence of CBr₄, PPh₃ to form dibromoalkene **175** and then *n*-BuLi in conjugation with methyl chloroformate to furnish alkyne compound **185**. Next, amides **178** and **186** were synthesized from compound **185** over a few steps (Scheme 17).

2.2.3. (−)-Chelonin A synthesis. (−)-Chelonin A **196** having 2,6-disubstituted morpholine core structure, was first extracted from the marine sponge *Chelonaplysilla* sp. in 1991.[75] This
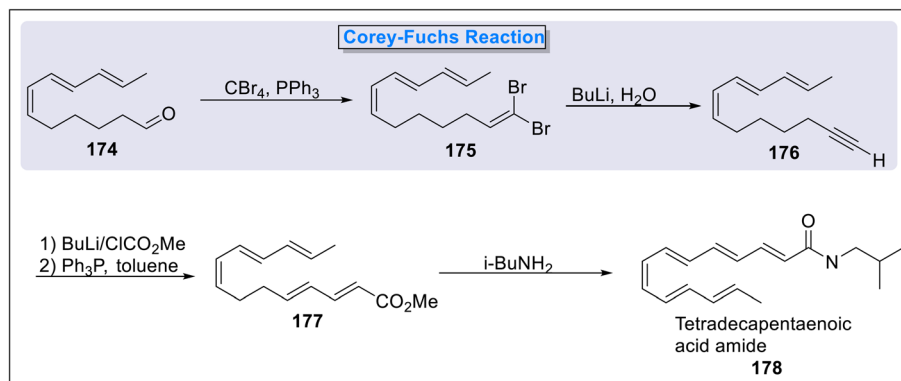
Scheme 16 Total synthesis of tetradecapentaenoic acid amide **178** by Kolodiazhna & Kolodiazhnyi.
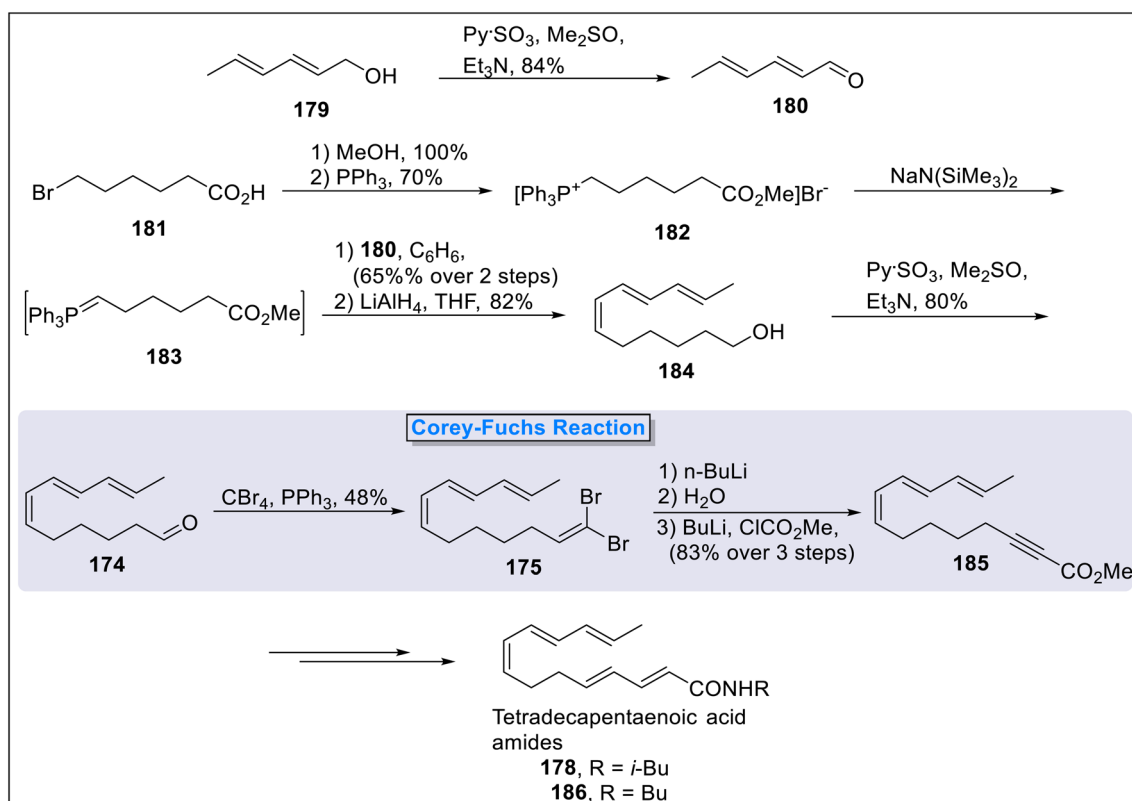
Scheme 17 Total synthesis of tetradecapentaenoic acid derivatives **178** & **186** by Kolodiazhna & Kolodiazhnyi.

compound showcased good anti-microbial potential against *B. subtilis* and also depicted 60% inhibition of phorbol myristate acetate triggered inflammation in the host animals.[75] Hitherto, four total syntheses of chelonin A and an asymmetric synthesis of (+)-chelonin A (unnatural enantiomer) have been reported to date.[76–79] Thus, Gunawana[80] *et al.*, in 2023 described the asymmetric total synthesis of (−)-chelonin A by employing various reaction processes *i.e.*, Corey–Fuchs, 1,3-addition of rhodium carbenoid, and annulation reaction. The first step for the total synthesis of (−)-chelonin included the synthesis of two starting substrates *i.e.*, bromohydrin (−)-**188** and triazole precursor **194**. In this regard, α-bromoketone **187** was converted to

enantioenriched bromohydrin (−)-**188** in the presence of Noyori catalyst **189** and HCO_2H/HCO_2K to furnish target product (−)-**188** in 87% yield with exceptional enantiomeric excess (>80% ee). In the next step, indole-3-carbaldehyde **190** was subjected to *N*-tosylation, followed by Corey–Fuchs reaction to synthesize terminal alkyne **193** in 62% yield. For this purpose, the protected aldehyde **191** (synthesized *via N*-tosylation) was treated under the conditions of CBr_4 and PPh_3 to generate gem-dibromo alkene **193** followed by its reaction with *n*-BuLi to form alkyne **193**. In the next step, alkyne **193** was then transformed into triazole **195** (75%) when treated with TsN_3 under the catalytic conditions of Cu thiophene carboxylate (CuTc). The

synthesized substrates *i.e.*, triazole **194** and (−)-bromohydrin **188** were both subjected to the Rh₂OcT₄ catalyzed reaction followed by cyclization reaction in the presence of CH₃CN & Cs₂CO₃ succeeded by its treatment with a mixture of trifluoroacetic acid and triethylsilane to generate the diastereoselective *syn*-compound **195** in 77% yield. In the last step, deprotection of protected morpholine **195** took place in the presence of Na naphthalenide to finally deliver enantioselective (−)-chelonin A **196** in 52% yield (Scheme 18).

2.2.4. Schwarzinicines A–D synthesis. Schwarzinicines A–D, chemically referred as 1,4-diarylbutanoidphenethylamine derivatives are naturally occurring alkaloids, isolated from *Ficus schwarzii*.⁸¹ These natural products are attributed to vasorelaxant, anti-diarrheal, anti-ulcer and anti-inflammatory

properties.⁸² Annapurna⁸³ *et al.*, in 2024 accomplished the total synthesis of these Schwarzinicines A–D compounds by employing Corey–Fuchs, Henry and Eschweiler–Clarke reactions. The synthesis began with the formation of 2-arylacetaldehyde **198** from veratraldehyde **197** in two steps *via* Wittig reaction and ether cleavage protocol in the presence of MeOCH₂PPh₃Cl, ᵗBuOK and HCl. Alongside, phenyl acetylene **200** (90%) was synthesized *via* Corey–Fuchs reaction from veratraldehyde **197** in two steps under the conditions of CBr₄, PPh₃, *n*-BuLi. Next, phenyl acetylene **200** and 2-arylacetaldehyde **198** underwent coupling reaction in the presence of *n*-BuLi to furnish alkynol **201** (85%) followed by the reduction of triple bond and subsequent oxidation in the presence of Dess–Martin periodinane to access diaryl ketone **202** in 90% yield. In the next
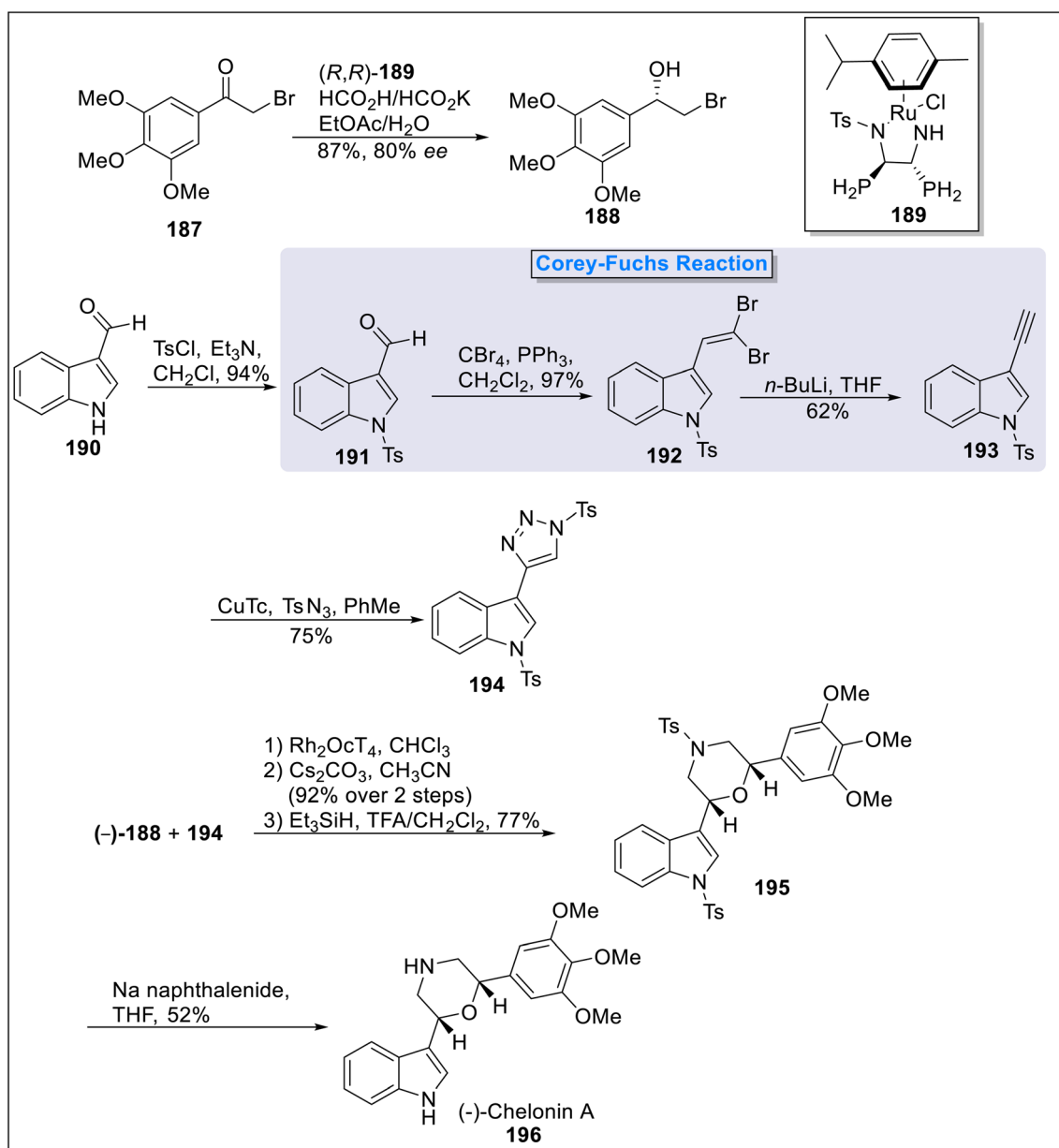
Scheme 18 Total synthesis of (−)-chelonin A **196** by Gunawana and coworkers.
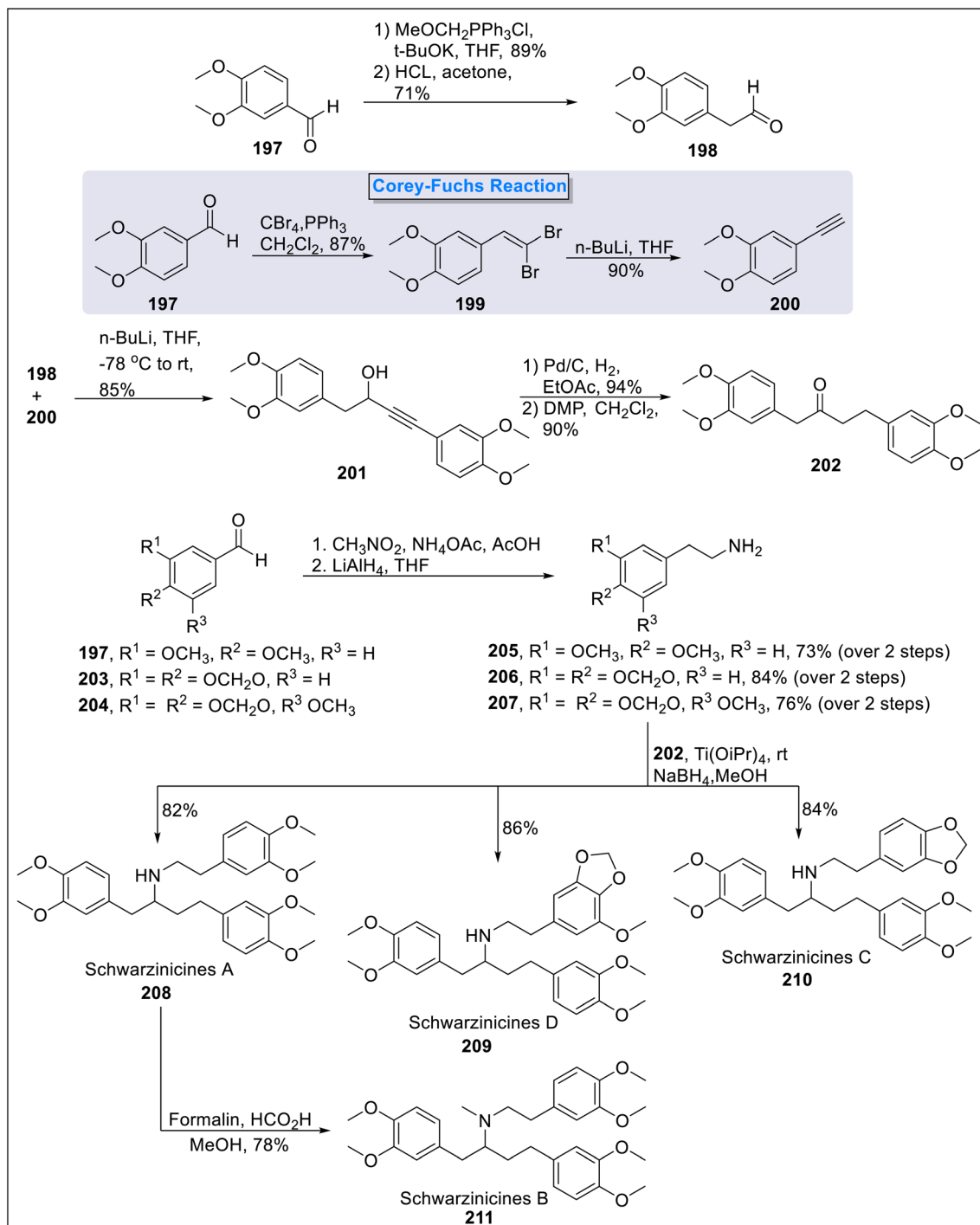
Scheme 19 Total synthesis of schwarzinicines A–D **208–211** by Annapurna and coworkers.

step, 2-arylethanamines **205**, **206**, **207** were synthesized from veratraldehyde **197**, piperonal **203** and 3-methoxy-4,5-methylenedioxybenzaldehyde **204** respectively in the presence of CH_3NO_2, NH_4OAc, AcOH and $LiAlH_4$ in two steps. The synthesized 2-arylethanamines *i.e.*, **205**, **206** & **207** were reacted with diaryl ketone **202** in the presence of titanium(IV)isopropoxide and $NaBH_4$ to furnish schwarzinicines A **208** (82%), schwarzinicines C **210** (84%) and Schwarzinicines D **209** (86%) respectively. Finally, schwarzinicines A **208** was converted into

schwarzinicines B **211** *via* Eschweiler–Clarke methylation providing 78% yield (Scheme 19).

2.3. Synthesis of polyunsaturated lipid-based natural products

2.3.1. Xestospongenyne synthesis. Xestospongenyne is a marine brominated naturally occurring compound which belongs to the class of polyunsaturated lipids.[84] It is considered as strong anti-obesity agent with exclusive pancreatic lipase
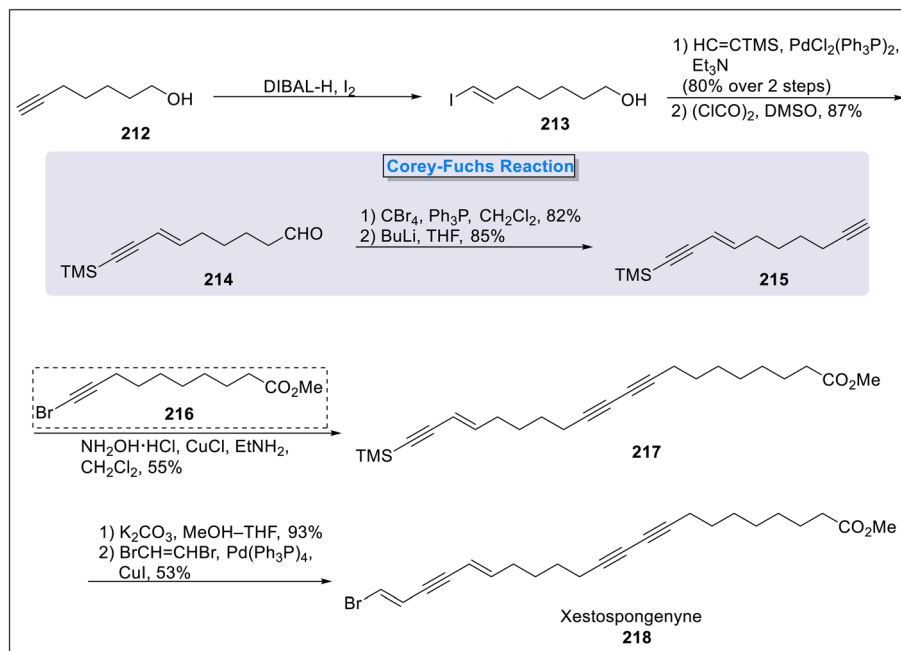
Scheme 20 Total synthesis of xestospongenyne **218** by Gong and coworkers.

inhibition potential with $IC_{50} = 0.61$ μM. Owing to the significant importance as potent pancreatic lipase inhibitor, the total synthesis of xestospongenyne attain much interest in organic synthesis.[85] In the regard, Gong and coworkers[86] in 2016, represented first total synthesis of xestospongenyne **218** in 13% overall yield. There were three key steps involved in the total synthesis of xestospongenyne **218** including Corey–Fuchs reaction, Cadiot–Choskiewicz cross-coupling and Sonogashira coupling. The total synthesis initiated with the synthesis of vinyl iodide **213** by the treatment of commercially available compound hept-6-yn-1-ol **212** with DIBAL-H in the presence of iodine. Further, the compound **213** was reacted in the presence of ethynyl trimethyl silane under the catalytic dichlorobis(triphenylphosphine)palladium in Et₃N followed by Swern's oxidation to furnish enynal compound **214** (87%). In the next step, compound **214** was transformed into silylated enyne **215** (85%) *via* Corey–Fuchs reaction under the conditions of tetra-bromomethane and triphenylphosphine in conjugation with butyl lithium in THF. Next, the alkyne compound **215** was subjected to Cadiot–

Choskiewicz cross-coupling with methyl 10-bromodec-9-ynoate **216** in the presence of hydroxylamine hydrochloride and copper chloride to furnish enyne ester **217** (55%). In the final step, subsequent potassium carbonate-mediated deprotection and Sonogashira coupling with dibromoethane under the presence of tetrakis(triphenylphosphine)palladium consequently gave xestospongenyne **218** in 53% yield (Scheme 20).

2.3.2. (5Z,9Z)-Eicosa-5,9-dienoic acid synthesis. (5Z,9Z)-Eicosa-5,9-dienoic acid **220** falls under the category of (5Z,9Z)-dienoic fatty acids which belongs to the class of non-methylene interrupted, very long chain fatty acids (VLCFA), mostly found in microorganisms, marine invertebrates and in some higher plants.[87] As these fatty acids are amidst the primitive and simplest I and IIα inhibitors known so far and hence considered for the development of new antitumor agents.[88,89] Owing to these properties, researchers are eager to develop new strategies towards the total synthesis of these structurally important natural products. In this prospective, Adrian and Stark[90] in 2016, demonstrated the total synthesis of topoisomerase
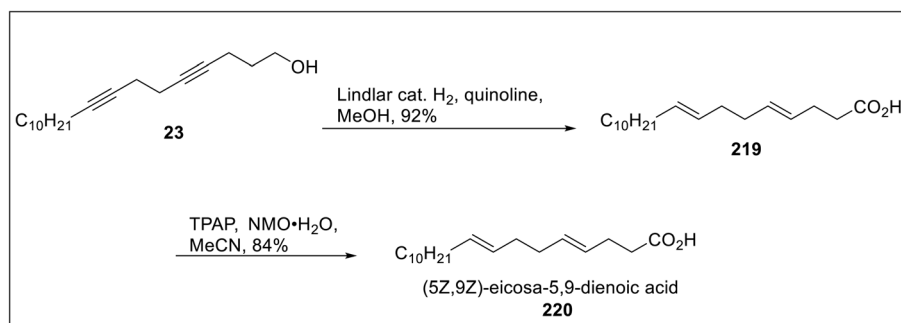
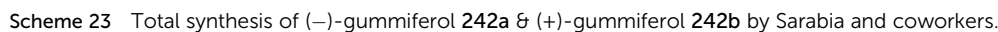**Scheme 21** Total synthesis of (5Z,9Z)-eicosa-5,9-dienoic acid **220** by Adrian and Stark.

inhibitor, (5Z,9Z)-eicosa-5,9-dienoic acid **220**, which is a potent human antitumor agent. The main steps towards the total synthesis involved Corey–Fuchs reaction, Lindlar reduction, TPAP-mediated oxidation of alcohol, and Arndt Eistert homologation. The synthesis commenced with precursor **17**, which was converted to compound **23** (provided in Scheme 3). Further,

Lindlar catalyst-mediated reduction of alkyne **23** resulted in (Z,Z)-alcohol **219** in 92% yield followed by TPAP/NMO·H₂O catalyzed direct oxidation that led towards the efficient total synthesis of natural (5Z,9Z)-eicosa-5,9-dienoic acid **220**. The straightforward total synthesis of compound **220** was achieved in 10 steps and with 20% overall yield (Scheme 21).
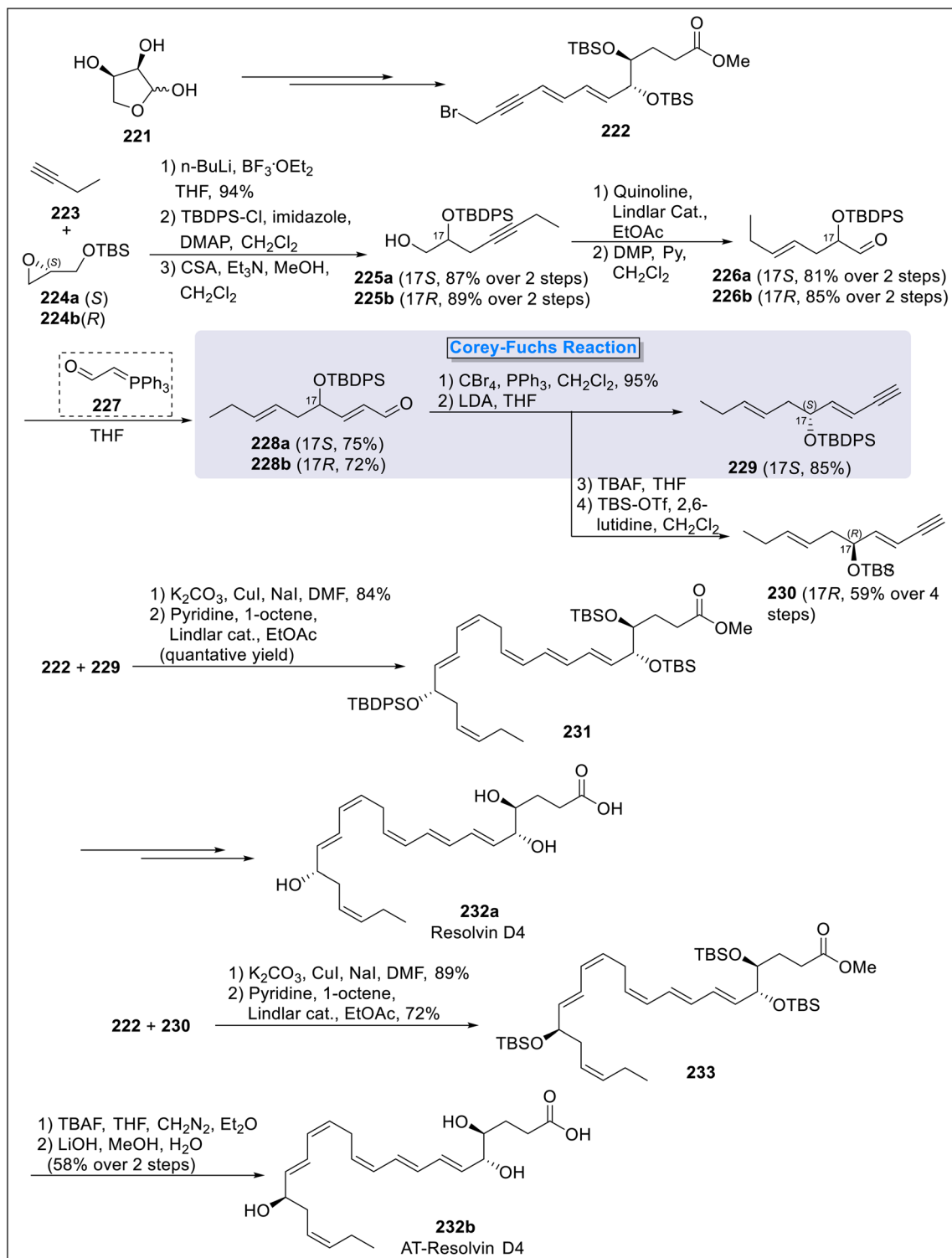
Scheme 22 Total synthesis of resolvin D4 **232a** and its 17(R)-hydroxy-epimer **232b** by Nshimiyimana and coworkers.

Scheme 23 Total synthesis of (−)-gummiferol **242a** & (+)-gummiferol **242b** by Sarabia and coworkers.

**Scheme 24** Total synthesis of atractylodinol **248** by Kraus and coworkers.

2.3.3. Resolvin D4 and its 17(R)-hydroxy-epimer synthesis. Resolvin D4 and its 17(R)-hydroxy-epimer are lipid based biosynthetic natural compounds derived from docosahexaenoic acid. These compounds have many biological active properties such as anti-inflammatory and pro resolution property which enhanced the activity of human macrophages.[91] In this context, Nshimiyimana[92] et al., in 2023 disclosed a convergent, stereocontrolled total synthesis of resolvin D4 **232a** and its epimer 17(R)-resolvin D4 **232b**. The synthesis was initiated by the generation of propargyl bromide **222** from D-erythrose **221** in multiple steps. The other key intermediates i.e., **229** & **230** was synthesized via a number of reaction protocols. In this regard, 1-butyne **223** was reacted with chiral glycidol **224** in the presence of boron trifluoride etherate followed by silylation in conjugation with mild deprotection to generate isomers of intermediate **225** (87–89%) in weak acidic medium. Next, Lindlar hydrogenation of **225** succeeded by Dess–Martin oxidation generated aldehyde **226** which on Wittig olefination with (triphenylphosphoranylidene)acetaldehyde **227** delivered elongated enol **228** (**228a** in 75% & **228b** in 72% yield). This enol was converted to terminal alkyne **229** via two steps Corey–Fuchs alkyne synthesis reaction in the presence of CBr$_4$ and PPh$_3$ in CH$_2$Cl$_2$ in conjugation with dehydrohalogenation using LDA in tetrahydrofuran (THF). To generate second isomer i.e., **230**, protection of **229** with less bulky silyl group was achieved which assisted in uniform desilylation in final step of the synthesis of AT-resolvin

D4 **232b**. In the next step, the reactions between **222** & **229** and **222** & **230** were took place via copper-catalyzed coupling to form bis-acetylenic moieties followed by Lindlar catalyzed selective reduction to generate silyl ethers **231** & **233** respectively. Next, desilylation in the presence of TBAF succeeded by its reaction with diazomethane and alkaline hydrolysis in the presence of LiOH furnished resolvin D4 **232a** and its 17(R)-hydroxy-epimer **232b** (Scheme 22).

2.4. Synthesis of polyacetylene-based natural products

2.4.1. (−)-Gummiferol synthesis. (−)-Gummiferol **242a** is a naturally occurring polyacetylene compound extracted from *Adenia gummifera* by Wall et al., in 1995.[93]. (−)-Gummiferol **242a** is characterized by excellent cytotoxic activities as it shows potential against P388 murine leukemia with ED$_{50}$ values of 0.03 μg mL^{-1} and against U373 human glioma cell lines with the ED$_{50}$ value 0.05 μg mL^{-1}.[94] Regarding the promising anticancer properties, the synthesis of different enantiomeric gummiferol is crucial in organic synthesis. In 2016, Sarabia and coworkers[95] designed the total synthesis of natural and non-natural gummiferol utilizing the novel asymmetric approach of epoxide construction. The two epoxide rings, installed on natural product were formed via novel asymmetric sulfonium salts. The total synthesis commenced with the oxidation of commercially available alcohol **234** in the presence of PCC and
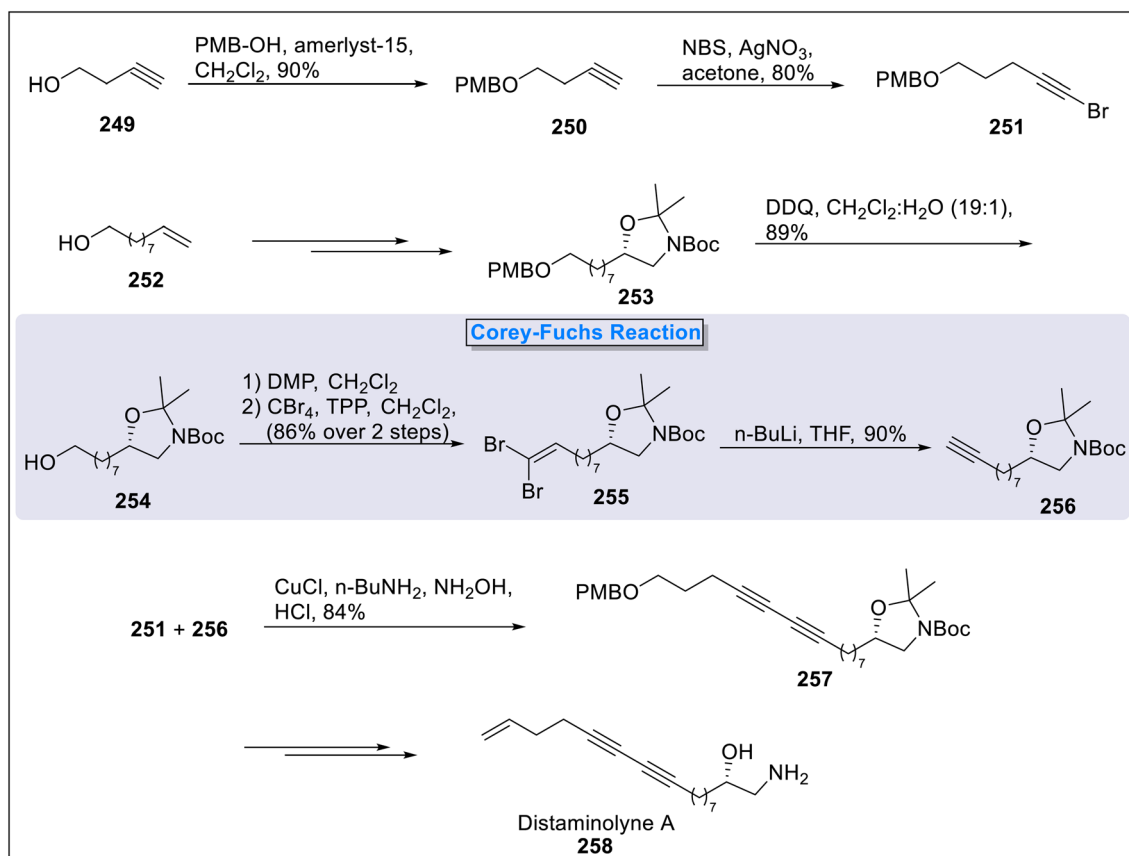

Scheme 25 Total synthesis of distaminolyne A **258** by Dumpala and coworkers.

sodium acetoacetate to form an aldehyde compound **235** followed by its reaction under the influence of sulfonium salt **236a** in basic conditions to access epoxy amide **237** (54%). Compound **237** were efficiently converted to diepoxy compound **238** (95%) *via* Red-Al induced reduction followed by its reaction with novel sulfonium salt **236a**. Subsequent reduction and Corey–Fuchs reaction delivered dibromo compound **239** in 40% yield. Next, the treatment of **239** with TBAF and coupling with dialkyne compound **240** in the presence of NH$_2$OH·HCl and CuCl followed by acetylation that led to the synthesis of compound **241** (51%). Finally, compound **241** was subjected to silyl deprotection in the presence of HF·Py to furnish natural (−)-gummiferol **242a** in overall 10% yield with 10 steps. Unnatural (+)-gummiferol **242b** was also obtained *via* same

synthetic protocol with the assistance of sulfonium salt **236b** (Scheme 23).

2.4.2. Atractylodinol synthesis. Atractylodinol **248** is an unsaturated natural product extracted from the rhizomes of *Atractylodes lancea. A. lancea* and known for its activity against porcine reproductive and respiratory syndrome virus (*anti*-PRRSV) potency with IC$_{50}$ values of 7.9 μmol L^{-1}.[96] It was utilized as traditional Chinese and Japanese drug against various diseases including night blindness, influenza, rheumatic and digestive disorders.[97] Owing to various biological activities, Kraus[98] *et al.*, in 2016, reported the total synthesis of atractylodinol **248** *via* utilizing a number of key synthetic procedure *i.e.*, Corey–Fuchs alkyne synthesis, Dess–Martin oxidation of alcohol and Wittig reaction. The total synthesis
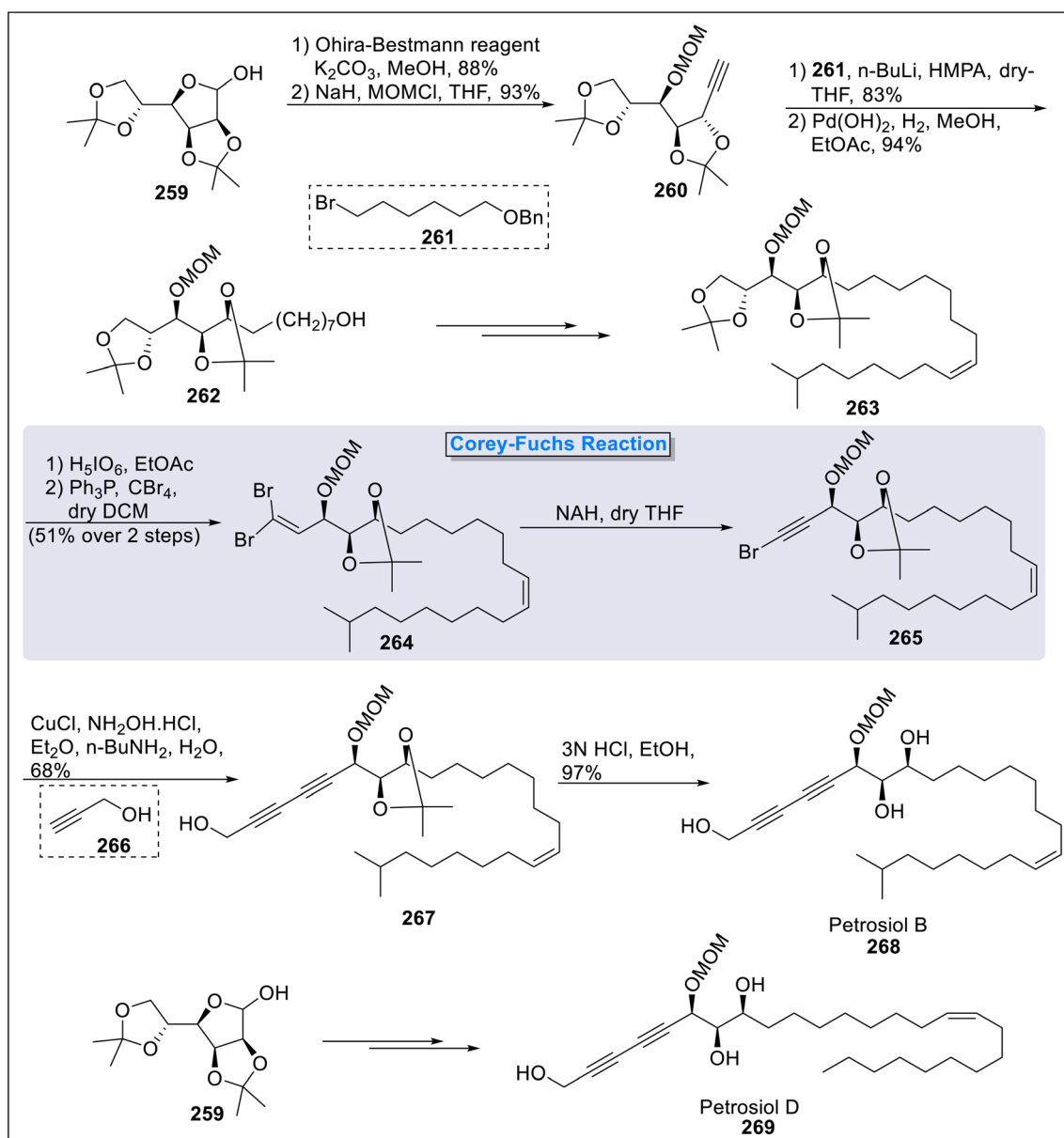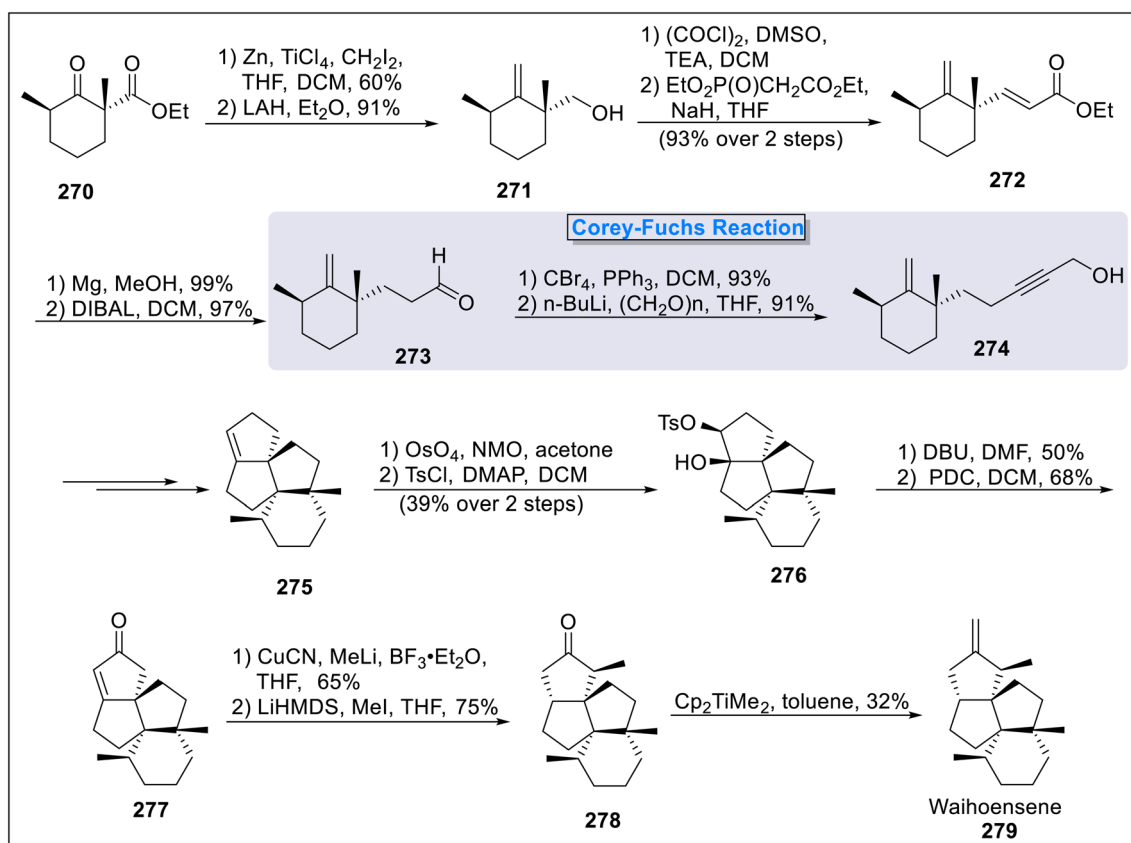

Scheme 26 Total synthesis of petrosiol B **268** & petrosiol D **269** by Geng and coworkers.

began with the known two step Corey–Fuchs reaction in which 3-(2-furyl)acrolein **243** was transformed into terminal alkyne **244** (86% overall yield) under the presence of carbon tetrabromide and triphenylphosphine in DCM. Next, terminal alkyne **244** was coupled with 3-iodopropargyl alcohol **245** in the presence of copper chloride, sodium hydroxide in butyl amine to furnish compound **246**, which was subjected to Dess–Martin oxidation in conjugation with Wittig reaction in the presence of phosphorane to form ester **247**. In the final step, reduction using DIBAL led to the synthesis of final natural product atractylodinol **248** in 88% yield (Scheme 24).

2.4.3. Distaminolyne A synthesis. Distaminolyne A **258** is a first naturally occurring compound, featuring diacetylene 1-amino 2-alcohol core structure. It was extracted from the New Zealand ascidian *Pseudodistoma opacum* exhibiting antimicrobial activity against several bacterial species such as *E. coli*, *S. aureus* and *M. tuberculosis*.[99] As *M. tuberculosis* remains a global health threat with unprecedented levels, spurring the need for effective inhibitors.[100,101] Owing to potential biological property, Dumpala[102] *et al.*, in 2017 proposed the total synthesis of distaminolyne A **258** utilizing a number of key synthetic steps. These steps including Corey–Fuchs reaction, Cadiot–Chodkiewicz coupling and Witting olefination reaction. The total synthesis started with the formation of bromoalkynes analogue **251** in two steps involving the reaction of commercially available 4-pentyn-1-ol **249** under the presence of PMB-OH

and amberlyst-15 to form PMB ether **250** (90%) followed by its conversion to bromoalkyne **251** (80%) in the presence of *N*-bromosuccinimide (NBS) and AgNO$_3$ (as catalyst). Next, the second coupling partner **256** of Cadiot–Chodkiewicz reaction was synthesized. For this purpose, 9-dacene-1-ol **252** was transformed into compound **253** over few steps. Next, compound **253** was subjected to *p*-anisyl group deprotection in the presence of DDQ to furnish alcohol **254** in 89% yield followed by DMP catalyzed oxidation to form Corey–Fuchs reaction precursor. Subsequent reaction with CBr$_4$ and triphenyl phosphine (TPP) yielded dibromide compound **255** (86%) succeeded by its treatment with *n*-BuLi in THF to afford alkyne **256** in 90% yield. Having the both coupling partners in hand, Cadiot–Chodkiewicz coupling of **251** & **256** led to the synthesis of diacetylene-1-amino-2-alcohol **257** (84%) under the influence of copper catalysis. Next, the desired natural product **258** was synthesized from compound **257** over few steps (Scheme 25).

2.4.4. Petrosiol B & D synthesis. Polyacetylenes are a large group of naturally occurring compounds featured with multitudinous double and triple bonds.[84] Petrosiol B & D fits into the class of these polyacetylene natural products extracted from marine sponge named *Petrosia strongylata*. These types of compounds are emphasized by various medicinal properties including neuritogenic, anti-inflammatory, antiviral, antitumor and anti-microbial.[103] Taking into account the significant potentials of these type of natural products Geng[104] *et al.*, in
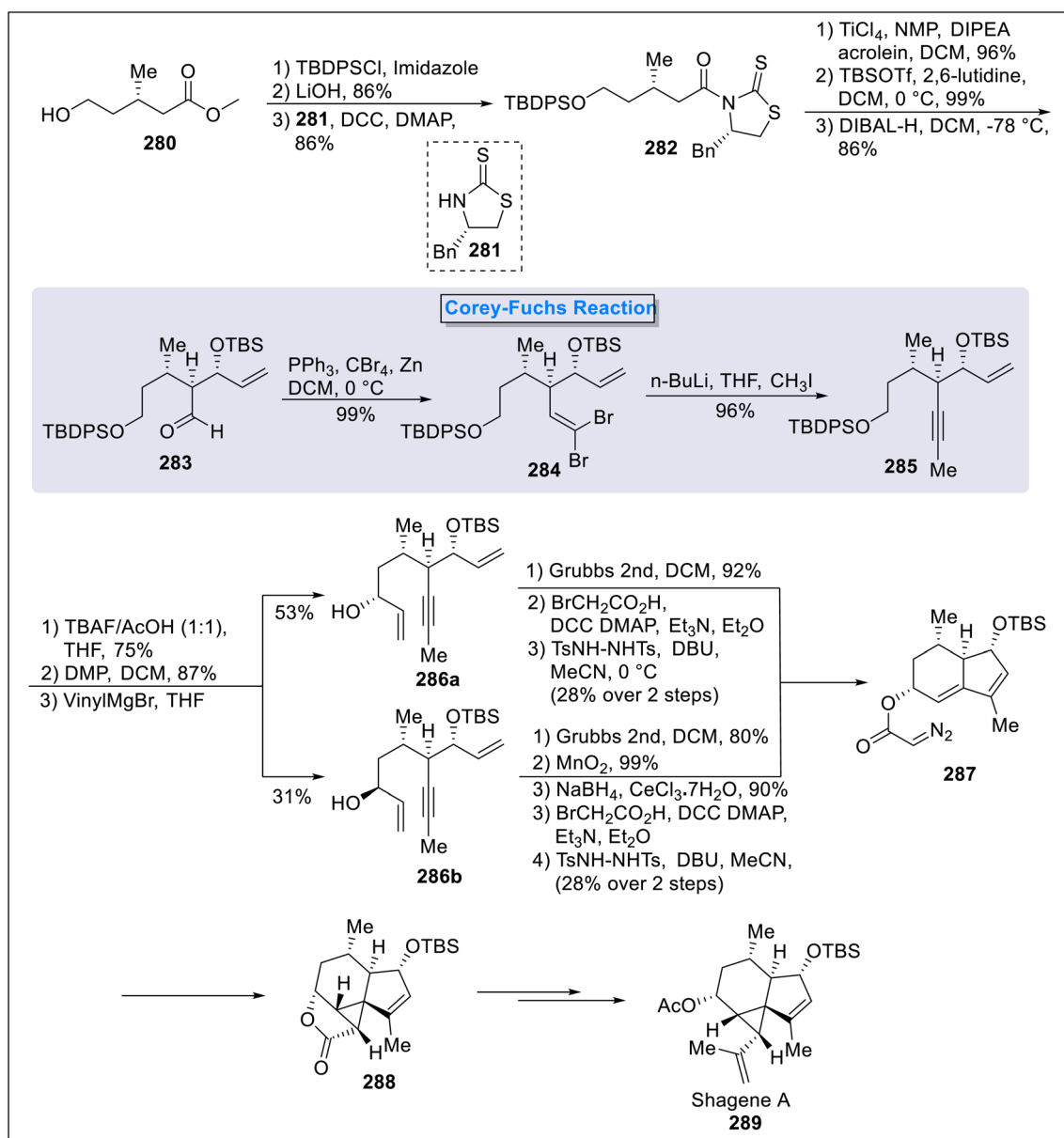

Scheme 27 Total synthesis of Waihoensene **279** by Lee and coworkers.

2019 devised the total synthesis of petrosiol B **268** & D **269**, reporting for the first time the total synthesis of petrosiol B **268** with 13 steps in 10% overall yield. The trajectory of the total synthesis involved Ohira–Bestmann homologation, Corey–Fuchs alkynes synthesis and Cadiot–Chodkiewicz reaction as key steps. The total synthesis commenced from unexpensive isopropylidene-α-D-mannofuranose **259** which was subjected to Ohira–Bestmann homologation in conjugation with its treatment with K$_2$CO$_3$ and MOM protection to furnish terminal alkyne **260** (93%). The synthesized alkyne **260** was then subjected to alkylation with bromide compound **261** under the influence of HMPA and butyl lithium followed by its hydrogenation in the presence of ethyl acetate and methanol to furnish compound **262** in 94% yield. Next, *cis*-olefin **263** was obtained from compound **263** over few steps. The regioselective cleavage

of isopropylidene region of the compound **263** under the presence of periodic acid in conjugation with Corey–Fuchs reaction in assistance of CBr$_4$ and PPh$_3$ led to the formation of dibromoalkene **264** in 51% yield. The synthesized dibromoalkene **264** underwent sodium hydride catalyzed dehydrobromination to furnish mono-bromoalkyne compound **265**. In the next step, compound **265** was subjected to Cadiot–Chodkiewicz coupling reaction with propargyl alcohol **266** in the presence of copper chloride, hydroxylamine, butylamine and diethyl ether to access diynes compound **267** in 68% yield. In the last step, the deprotection of terminal isopropylidene group of **267** in the presence of 3 N HCl led to the synthesis of natural product petrosiol B **268** in 97% yield. The other natural product *i.e.*, petrosiol D **269** (98%) was also synthesized from the same precursor **259** over a few steps (Scheme 26).


Scheme 28   Route leading towards the total synthesis of shagene A **289** by Bai and coworkers.

## 2.5. Synthesis of terpene-based natural products

2.5.1. Waihoensene synthesis.
Waihoensene **279** is a naturally occurring tetracyclic diterpene, derived from New Zealand podocarp, *Podocarpus totara* var. *waihoensis* by Weavers *et al.*, in 1997.[105] Waihoensene has intriguing architectural features as it is decorated with four contiguous quaternary carbons and thus also recognized as structurally challenging compound for total synthesis. In 2017, Lee[106] and coworkers reported the total of Waihoensene for the first time from commercially available reagents. The total synthesis commenced from a racemic keto ester **270** which was subjected to Lombardo–Takai olefination in the presence of Zn, TiCl$_4$ and CH$_2$I$_2$ followed by lithium aluminum hydride (LAH) mediated reduction to furnish alcohol **271** (91%). Next, Swern's oxidation of compound **271** in conjugation with Horner Wadsworth–Emmons reaction yielded α,β-unsaturated ester **272** (93%). Selective reduction of compound **272** employing magnesium and then partial reduction using DIBAL generated aldehyde **273** in 97% yield. Aldehyde **273** was
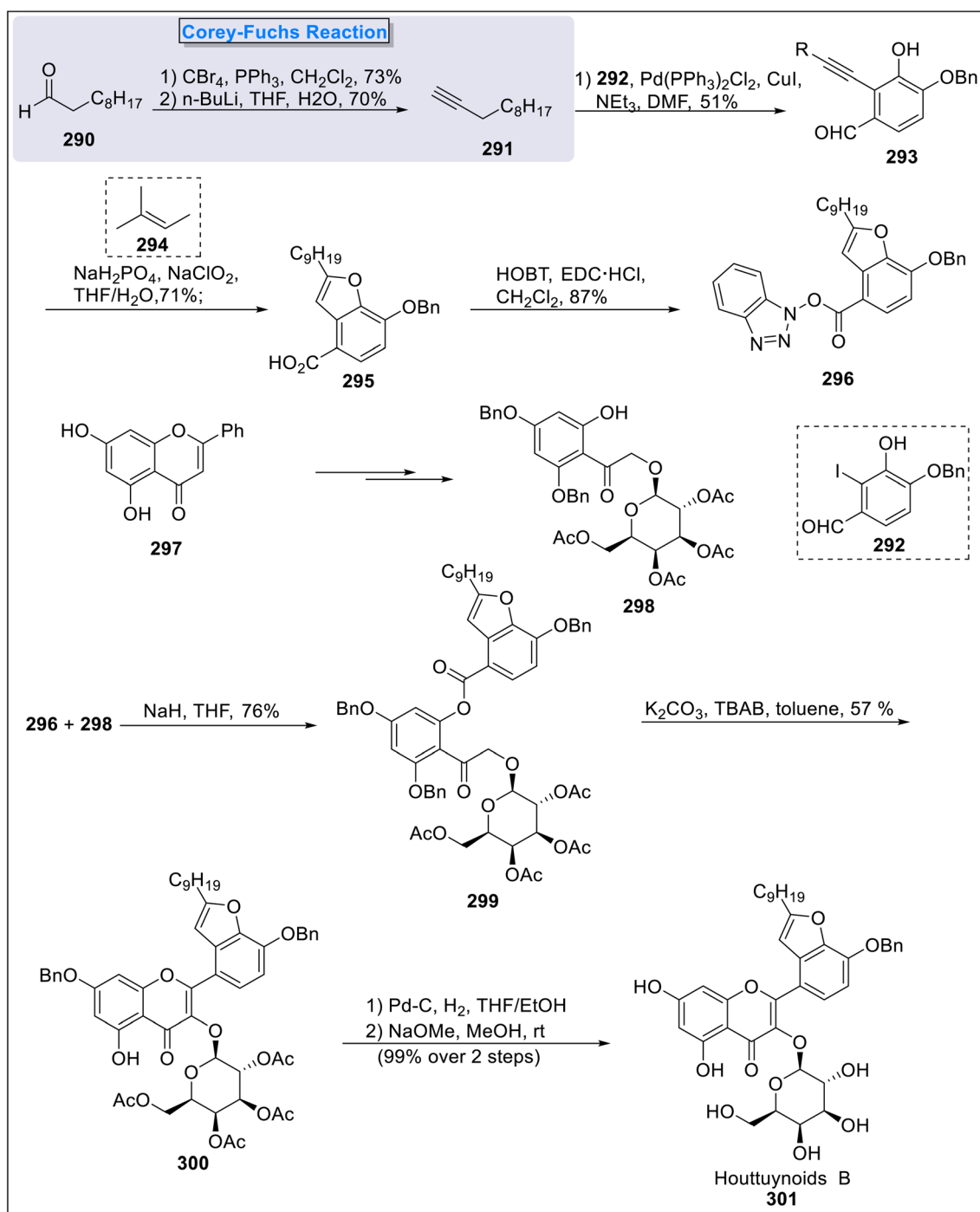
Scheme 29 Total synthesis of houttuynoid B **301** by Kerl and coworkers.
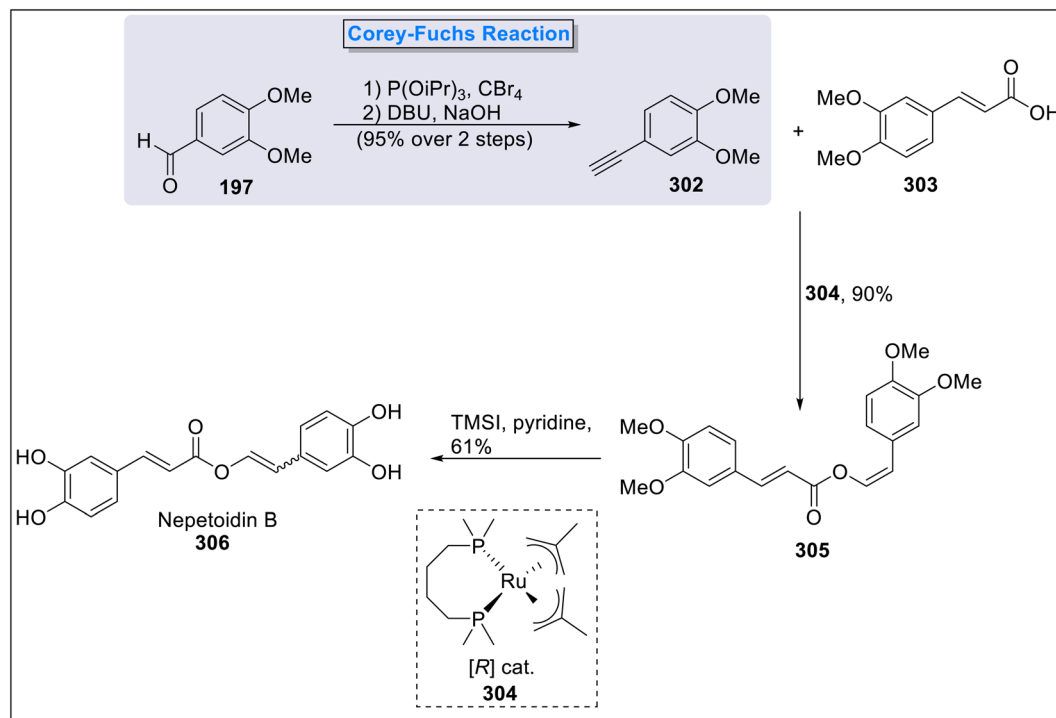
Scheme 30 Total synthesis of nepetoidin B **306** by Yao and coworkers.

transformed into propargylic alcohol **274** (91%) utilizing Corey–Fuchs reaction conditions in conjugation with hydroxymethylation. Next, tetracyclic product **275** was accessed from compound **274** over a few steps. Compound **275** was further subjected to dihydroxylation in the presence of OsO$_4$ succeeded by selective tosylation using TsCl to furnish compound **276** (39%). In the next step, the formation of enone **277** was achieved *via* elimination reaction in the presence of DBU followed by PDC mediated oxidation. Reaction of enone compound **277** with cyanocopper and boron trifluoride etherate in conjugation with LiHMDS and methyl iodide furnished methyl substituted compound **278** in 75% yield. In the final step, employing Petasis reagent (Cp$_2$TiMe$_2$) in toluene, natural product Waihoensene **279** was achieved in 32% yield (Scheme 27).

2.5.2. Synthesis of key intermediate of shagene A. Shagene A **289** is a tricyclic sesquiterpenoid, extracted for the first time from soft corals of Scotia Sea by baker group in 2014.[107] Shagene A showcase high toxicity against parasite *Leishmania donovani* (IC$_{50}$ = 5 μM), which is a causative agent for serious visceral leishmaniasis.[108] In 2024, Bai[9] *et al.*, demonstrated an efficient route leading towards the total synthesis of Shagene A **289** by employing ring closing cascade reaction of dienyne **286** constructed *via* Corey–Fuchs reaction, into 5/6 membered bicyclic compound. The ring closing metathesis was succeeded by the formation of diazoacetate **287** which was considered hypothetical key intermediate for the total synthesis of shagene A **289**. The total synthesis was initiated with hydroxyl group protection of ester **280** in conjunction with hydrolysis followed by installation of chiral auxiliary **281** in the presence of DCC and DMAP to furnish amide **282** in

86% yield. Next, **282** was subjected to the subsequent aldol reaction by reacting it with acrolein followed by hydroxyl group protection with TBS and reductive cleavage of previously installed chiral auxiliaries in the presence of DIBAL-H to access compound **283** (in 86% yield). The conversion of aldehyde **283** to alkyne **285** took place by Corey–Fuchs reaction utilizing PPh$_3$ and CBr$_4$ and then *n*-BuLi and alkyl iodide over 2 steps reaction (resulting in 96% yield). The internal alkyne **285** was then subjected to selective deprotection of TBDPS followed by oxidation in the presence of DMP to form an aldehyde in 87% yield. Further, next step proceeded with the treatment of aldehyde with Grignard reagent *via* nucleophilic addition to generate two diastereomers **286a** (53%) and **286b** (31%). Both the isomers were transformed to the cyclic compound **287** over a few steps. The compound **286a** was subjected to ring closing cascade reaction to deliver cyclic compound **287** (in 28% yield) catalyzed by Grubbs 2nd catalyst, 2-bromoacetic acid, DCC DMAP and *N*,*N*′-ditosylhydrazine (for the installation of diazo group) over three steps. Compound **286b** was also transformed into compound **287** over several reaction steps. It was first reacted with Grubb's 2nd catalyst followed by its treatment with MnO$_2$ and then sodium borohydride and CeCl$_3$·7H$_2$O. Next, its treatment with bromoacetic acid, DCC DMAP and *N*,*N*′-ditosylhydrazine led to the synthesis of **287** in 28%. The target compound **289** was assumed to achieve over few steps from compound **288**. Several reaction conditions were applied for the efficient conversion of the intermediate **287** to 3-membered ring compound **288** *via* cyclopropanation reaction but the starting material was decomposed (Scheme 28).

2.6. Synthesis of flavonoid-based natural products

2.6.1. Houttuynoid synthesis. Houttuynoid **301** is a naturally occurring flavonoid glycoside that belongs to the family of *Sauruaceae* and extracted from Chinese *Houttuynia cordata* by Yao *et al.*, in 2012.[109] It is considered as strong antiviral flavonoid having various viral inhibition properties such as inhibition of herpes simplex virus (with IC_{50} values of 57.7 μM).[109] Owing to the importance of development of new antiviral agents, Kerl and coworkers[110] in 2016 reported for the first the total synthesis of houttuynoid B **301** in overall 9 linear steps with 11% yield. The key steps involving the total synthesis include Corey–Fuchs alkyne synthesis reaction, Sonogashira coupling and Baker–Venkataraman rearrangement. The

synthesis commenced from the unexpensive decanal **290** which was subjected to Corey–Fuchs reaction under the presence of carbon tetrabromide and triphenyl phosphine in conjugation with *n*-butyl lithium to furnish terminal alkyne **291** (70%). Next, alkyne **291** underwent Sonogashira coupling reaction with iodophenol **292** in the presence of $Pd(PPh_3)_2Cl_2$, CuI and triethylamine to furnish **293** (51%) which was then rearranged to a benzofuran moiety. In the next step, the aldehydic functionality on **293** underwent Pinnick oxidation in the presence of NaH_2PO_4, $NaClO_2$ and 2-methyl-2-butene2 **294** to furnish carboxylic framework **295** in 71% yield. Further, to achieve HOBT activated ester **296** (87%), the carboxylic compound **295** was reacted with 1-hydroxybenzotriazole (HOBT) and ethyl-3-(3-dimethylaminopropyl)-carbodiimide in CH_2Cl_2. On the other
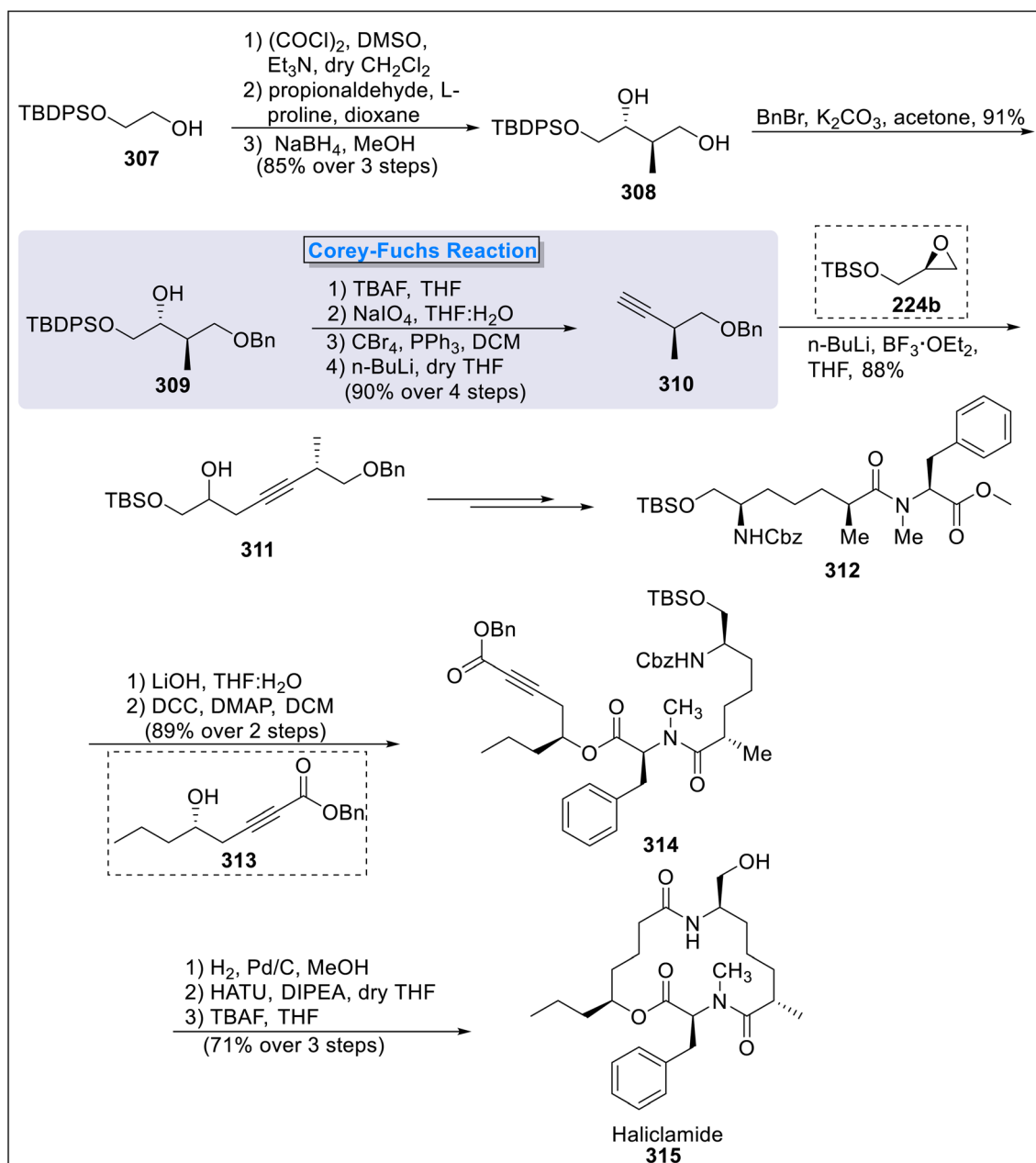
Scheme 31 Total synthesis of haliclamide **315** by Gahalawat and coworkers.

side, the second precursor **298** was synthesized from commercially available chrysin **297** over a few steps. Having both the reaction partners *i.e.*, **296** & **298** in hand, HOBT activated ester **296** was made to react with **298** in the presence of sodium hydride in THF to form arylester **299** (76%). Next, chromenone **300** (57%) was accessed from arylester **299** *via* Baker–Venkataraman rearrangement under the presence of K_2CO_3 and TBAB in toluene. Finally, the desired natural product houttuynoid B **301** was synthesized by deprotection of chromenone **300** under the presence of sodium methoxide in toluene with 99% yield (Scheme 29).

2.7. Synthesis of polyphenol-based natural products

2.7.1. Nepetoidin B synthesis. Nepetoidin B **306** belongs to the class of phenolic compounds existing in two isomeric forms *i.e.*, (*Z*, *E*)- and (*E*, *E*). It is extracted from *P. frutescens*,[111] *P.*

forsteri[112] and *S. miltiorrhiza*[113] and depicting remarkable bioactive properties including anti-viral, anti-fungal and anti-oxidant properties.[114] It is also used for the production of nitric oxide and the inhibition of xanthine oxidase. Nepetoidin B was synthesized for the first time in 2018 by Timokhin and co-workers in 17% overall yield.[115] Due to the ongoing research on the synthesis of bioactive phenolic compounds, Yao[116] *et al.*, in 2020 developed a three steps synthesis of nepetoidin B with an overall 52% yield. The key steps involved Corey–Fuchs alkyne synthesis reaction, Ru-catalyzed *anti*-Markovnikov addition and demethylation. The synthesis commenced from the commercially available 3,4-dimethoxybenzaldehyde **197** which underwent Corey–Fuchs reaction to form respective terminal alkyne **302** (95%). Next, the alkyne **302** was subjected to *anti*-Markovnikov addition with 3,4-dimethoxycinnamic acid **303** under the presence of Ru-catalyst **304** to afford tetramethylated
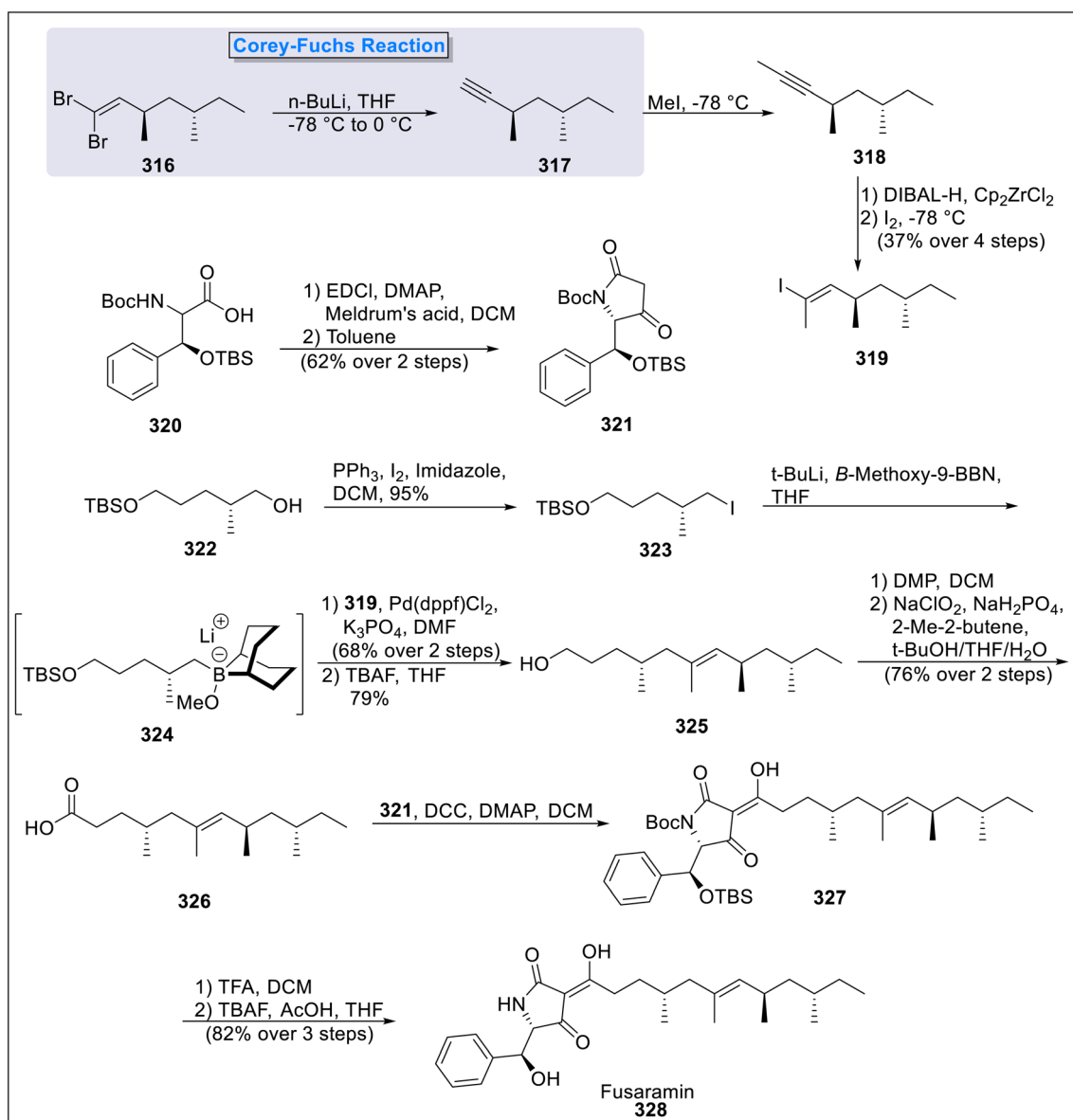
Scheme 32 Total synthesis of fusaramin **328** by Kimishima and coworkers.

nepetoidin B **305** in 90% yield. Subsequent demethylation of compound **305** in the presence of Me₃SiI and pyridine afforded the mixture of (Z, E)/(E, E) isomers of nepetoidin B **306** with 2; 8 : 1 ratio and 61% yield (Scheme 30).

2.8. Synthesis of peptide-based natural products

2.8.1. Haliclamide synthesis. Haliclamide **315** is a naturally occurring compound, extracted from the marine sponge *Haliclona* sp. of Vanuatu island by Randazzo *et al.*, in 2001.[117] Haliclamide exhibited excellent *in vitro* anti-tumor properties against human lung carcinoma cell line. The first total synthesis of haliclamide was also done by the same group at the time of its extraction. Due to its significant anti-tumor properties, the total synthesis of haliclamide has gained much interest of synthetic chemists. In this perspective, Gahalawat[118] *et al.*, in 2016 published the total synthesis of haliclamide **315** employing MacMillan cross aldol reaction, Corey–Fuchs alkyne synthesis, Yamaguchi–Hirao Reaction, Steglich esterification reaction and macro-lactamization reaction as key steps. The total synthesis began with the commercially available ethylene glycol **307**, which was subjected to MacMillan cross aldol strategy in the presence of L-proline followed by NaBH₄-mediated reduction to furnish 1,3-diol **308** (85%). Next, compound **308** was easily transformed into monobenzylated derivative **309** (91%) *via* treating it with K₂CO₃ and BnBr followed by subsequent silyl deprotection, NaIO₄ catalyzed oxidation and Corey–Fuchs alkyne synthesis reaction under the presence of carbon tetrabromide and phosphine triphenyl to furnish acetylene moiety **310** in 90% yield. Next, compound **310** was subjected to Yamaguchi–Hirao alkylation protocol with glycidol silyl ether **229b** under the presence of boron trifluoride diethyl etherate to access homopropargylic alcohol **311** in 88% yield. Compound **311** was then transformed into compound **312** over a few steps. Next, LiOH mediated saponification of compound **312** took place succeeded by Steglich esterification reaction with alcohol **313** under the presence of DCC and DMAP to form compound **314** (89%). Next, subsequent Cbz deprotection, hydrogenation and HATU mediated macro-lactamization finally furnished haliclamide **315** in 71% yield (Scheme 31).

2.9. Miscellaneous natural products

2.9.1. Fusaramin synthesis. Fusaramin **328** is a naturally occurring potent antimitochondrial compound obtained from agar broth of *Fusarium* sp. FKI-7550.[119] This compound shows excellent anti-fungal activities against plant disease-causing fungi. Thus, in 2024, Kimishima[120] *et al.*, envisioned the total synthesis of Fusaramin **328** and enabled the elucidation of its stereochemical structure. The total synthesis commenced with the formation of vinyl iodide **319** (37% yield) from gem-dibromo alkene **316** *via* sequential Corey–Fuchs, methylation and Schwartz hydrozirconation reaction in the presence of *n*-BuLi, MeI, DIBAL-H, Cp₂ZrCl₂ and I₂. Next, phenylalanine derivative **320** was condensed with Meldrum's acid followed by the ketene formation and attack of the Boc-amine group to access tetramic acid fragment **321** with 62% yield. The next step involved treatment of asymmetric silyl protected alcohol **322** under the

conditions of PPh₃, I₂ and imidazole (Appel conditions) in DCM to afford alkyl iodo compound **323**. This reaction was followed by the generation of an intermediate **324** *via* lithium–halogen exchange in the presence of *n*-BuLi which was then quenched with *B*-methoxy-9-BBN. The intermediate **324** underwent Suzuki–Miyaura coupling reaction with **319**, followed by silyl deprotection in the presence of TBAF to deliver chiral alcohol **325** in 79% yield. Further, two steps oxidation of the compound **325** took place to form carboxylic acid compound **326** (76%) which was reacted with **321** to form Boc protected fusaramin **327**. Finally, the deprotection of **327** in the presence of TFA, TBAF and AcOH led to the synthesis of desired product **328** with 82% yield (Scheme 32).

3. Conclusion

In conclusion, this manuscript summarizes the significance of Corey–Fuchs reaction in the total synthesis of various biologically important natural products as a key step. The protocol facilitates the direct and efficient pathway to alkynes from aldehydes and has been the area of extensive research and development. This review covers the total synthesis of various natural products *i.e.*, polyketides, alkaloids, peptides and polyacetylene-based natural products utilizing the Corey–Fuchs reaction as one of the significant steps. Current breakthroughs and modifications of the reaction have further broadened its viability and practicality, and is expected to remain a cornerstone in the synthesis of complex compounds for the foreseeable future.

Data availability

No primary research results, software or code have been included and no new data were generated or analyzed as part of this review.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

References

1 N. Desai, N. McKelvie and F. Ramirez, *J. Am. Chem. Soc.*, 1962, **84**, 1745–1747.

2 F. Pandolfi, I. Chiarotto and M. Feroci, *Beilstein J. Org. Chem.*, 2018, **14**, 891–899.

3 H. Okumura, N. I. Prakoso, T. Morozumi and T. Umezawa, *Org. Lett.*, 2024, **26**, 9817–9821.

4 A. K. Ghosh and Y. Wang, *Tetrahedron Lett.*, 2000, **41**, 4705–4708.

5 S. Das, S. Paul, K. K. Mandal, A. Anoop and S. Nanda, *J. Org. Chem.*, 2024, **89**, 15764–15776.

6 Y. Garg, S. Gahalawat and S. K. Pandey, *RSC Adv.*, 2015, **5**, 38846–38850.

7 M. G. Jakobsen, A. Vik and T. V. Hansen, *Tetrahedron Lett.*, 2014, **55**, 2842–2844.

8 H. F. Sneddon, M. J. Gaunt and S. V. Ley, *Org. Lett.*, 2003, **5**, 1147–1150.

9 M. Bai, X. Wang and Q. Li, *Tetrahedron*, 2024, **154**, 133846.

10 E. Corey and P. Fuchs, *Tetrahedron Lett.*, 1972, **13**, 3769–3772.

11 D. Habrant, V. Rauhala and A. M. Koskinen, *Chem. Soc. Rev.*, 2010, **39**, 2007–2017.

12 M. Dhameja and J. Pandey, *Asian J. Org. Chem.*, 2018, **7**, 1502–1523.

13 F. Eymery, B. Iorga and P. Savignac, *Synthesis*, 2000, **2000**, 185–213.

14 Y. Thummala, G. V. Karunakar and V. R. Doddi, *Adv. Synth. Catal.*, 2019, **361**, 611–616.

15 W. Doherty, N. Adler, A. Knox, D. Nolan, J. McGouran, A. P. Nikalje, D. Lokwani, A. Sarkate and P. Evans, *Eur. J. Org Chem.*, 2017, **2017**, 175–185.

16 S. Kumar, M.-T. Ho and Y.-T. Tao, *Org. Lett.*, 2016, **18**, 200–203.

17 A. Irfan, S. Faisal, S. Ahmad, S. A. Al-Hussain, S. Javed, A. F. Zahoor, B. Parveen and M. E. Zaki, *pharmaceuticals*, 2023, **16**, 344.

18 S. Faiz, A. F. Zahoor, N. Rasool, M. Yousaf, A. Mansha, M. Zia-Ul-Haq and H. Z. Jaafar, *Molecules*, 2015, **20**, 14699–14745.

19 P. J. Parsons, C. S. Penkett and A. J. Shell, *Chem. Rev.*, 1996, **96**, 195–206.

20 Y. Bai, X. Shen, Y. Li and M. Dai, *J. Am. Chem. Soc.*, 2016, **138**, 10838–10841.

21 D. L. Boger, S. Ichikawa and W. Zhong, *J. Am. Chem. Soc.*, 2001, **123**, 4161–4167.

22 H. Johansson and D. S. Pedersen, *Eur. J. Org Chem.*, 2012, **2012**, 4267–4281.

23 M. Yao, W. Yang, J. Li, C. Huang, J. Fang, S. Qin, S. Liu and X. Yang, *Chin. J. Chem.*, 2024, **42**, 1509–1514.

24 A. Mushtaq, A. F. Zahoor, M. Bilal, S. M. Hussain, M. Irfan, R. Akhtar, A. Irfan, K. Kotwica-Mojzych and M. Mojzych, *Molecules*, 2023, **28**, 2722.

25 J. S. Lee, J. Shin, H. J. Shin, H. S. Lee, Y. J. Lee, H. S. Lee and H. Won, *Eur. J. Org Chem.*, 2014, **2014**, 4472–4476.

26 T. Sengoku, S. Xu, K. Ogura, Y. Emori, K. Kitada, D. Uemura and H. Arimoto, *Angew. Chem.*, 2014, **126**, 4297–4300.

27 S. Huang, S. Chen, G. Wang, J. Zhang, L. Tang, G. Du and X. Wang, *Synthesis*, 2015, **47**, 1303–1308.

28 M. M Heravi, S. Asadi, N. Nazari and B. Malekzadeh Lashkariani, *Curr. Org. Chem.*, 2015, **19**, 2196–2219.

29 N. Li, Z. Shi, Y. Tang, J. Chen and X. Li, *Beilstein J. Org. Chem.*, 2008, **4**, 48.

30 J. Adrian and C. B. Stark, *J. Org. Chem.*, 2016, **81**, 8175–8186.

31 K. Krohn and J. Rohr, *Bioorganic Chemistry Deoxysugars, Polyketides and Related Classes: Synthesis, Biosynthesis, Enzymes*, 2008, pp. 127–195.

32 P. M. Brown and R. H. Thomson, *J. Chem. Soc., Perkin Trans. 1*, 1976, 997–1000.

33 D.-S. Hsu and J.-Y. Huang, *J. Org. Chem.*, 2012, **77**, 2659–2666.

34 K. J. Ngwira, A. L. Rousseau, M. M. Johnson and C. B. de Koning, *Eur. J. Org Chem.*, 2017, **2017**, 1479–1488.

35 S. M. Dalby and I. Paterson, *Curr. Opin. Drug Discovery Dev.*, 2010, **13**, 777–794.

36 C. I. MacGregor, B. Y. Han, J. M. Goodman and I. Paterson, *Chem. Commun.*, 2016, **52**, 4632–4635.

37 K.-H. Altmann and J. Gertsch, *Nat. Prod. Rep.*, 2007, **24**, 327–357.

38 Y. Inahashi, M. Iwatsuki, A. Ishiyama, A. Matsumoto, T. Hirose, J. Oshita, T. Sunazuka, W. Panbangred, Y. Takahashi and M. Kaiser, *Org. Lett.*, 2015, **17**, 864–867.

39 J. Oshita, Y. Noguchi, A. Watanabe, G. Sennari, S. Sato, T. Hirose, D. Oikawa, Y. Inahashi, M. Iwatsuki and A. Ishiyama, *Tetrahedron Lett.*, 2016, **57**, 357–360.

40 F. Doraghi, L. Rezainia, M. H. Morshedsolouk, H. Navid, B. Larijani and M. Mahdavi, *RSC Adv.*, 2025, **15**, 1134–1151.

41 R. Akhtar, A. F. Zahoor, M. Irfan, T. H. Bokhari and A. ul Haq, *Chem. Pap.*, 2022, **76**, 7275–7293.

42 Y.-Z. Shu, Q. Ye, H. Li, K. F. Kadow, R. A. Hussain, S. Huang, D. R. Gustavson, S. E. Lowe, L.-P. Chang and D. M. Pirnik, *Bioorg. Med. Chem. Lett.*, 1997, **7**, 2295–2298.

43 J. Peng, J. Jiao, J. Li, W. Wang, Q. Gu, T. Zhu and D. Li, *Bioorg. Med. Chem. Lett.*, 2012, **22**, 3188–3190.

44 J. Preindl, S. Schulthoff, C. Wirtz, J. Lingnau and A. Fürstner, *Angew. Chem., Int. Ed.*, 2017, **56**, 7525–7530.

45 A. Cutignano, I. Bruno, G. Bifulco, A. Casapullo, C. Debitus, L. Gomez-Paloma and R. Riccio, *Eur. J. Org Chem.*, 2001, **2001**, 775–778.

46 T. Tanaka, Y. Murai, T. Kishi, H. Takamura and I. Kadota, *Tetrahedron Lett.*, 2018, **59**, 763–766.

47 C. Tanaka, J. Tanaka, R. F. Bolland, G. Marriott and T. Higa, *Tetrahedron*, 2006, **62**, 3536–3542.

48 H. D. Arndt, S. Rizzo, C. Nöcker, V. N. Wakchaure, L. G. Milroy, V. Bieker, A. Calderon, T. T. Tran, S. Brand and L. Dehmelt, *Chem.–Eur. J.*, 2015, **21**, 5311–5316.

49 J. H. Lang and T. Lindel, *Beilstein J. Org. Chem.*, 2019, **15**, 577–583.

50 N. Oku, K. Takada, R. W. Fuller, J. A. Wilson, M. L. Peach, L. K. Pannell, J. B. McMahon and K. R. Gustafson, *J. Am. Chem. Soc.*, 2010, **132**, 10278–10285.

51 C. K. Skepper, T. Quach and T. F. Molinski, *J. Am. Chem. Soc.*, 2010, **132**, 10286–10292.

52 K. Sakurai, K. Sakamoto, M. Sasaki and H. Fuwa, *Chem.–Asian J.*, 2020, **15**, 3494–3502.

53 S. B. Krasnoff, U. Englich, P. G. Miller, M. L. Shuler, R. P. Glahn, B. G. Donzelli and D. M. Gibson, *J. Nat. Prod.*, 2012, **75**, 175–180.

54 M. Yoshida, Y. Okoshi and H. Kigoshi, *Chem. Commun.*, 2023, **59**, 9880–9883.

55 A. Sharma, S. Athe, P. Ramesh, K. Vishali and S. Ghosh, *Tetrahedron Lett.*, 2021, **82**, 153374.

56 W. Zhou, H. A. Alharbi, E. Hummingbird, A. T. Keatinge-Clay and T. Mahmud, *ACS Chem. Biol.*, 2022, **17**, 2039–2045.

57 D. Dardić, N. Böhringer, A. Plaza, F. Zubeil, J. Pohl, S. Sommer, L. Padva, J. Becker, M. A. Patras and M.-K. Bill, *Org. Chem. Front.*, 2022, **9**, 1604–1615.

58 S.-Z. Huang, F.-D. Kong, G. Chen, X.-H. Cai, L.-M. Zhou, Q.-Y. Ma, Q. Wang, W.-L. Mei, H.-F. Dai and Y.-X. Zhao, *Phytochemistry*, 2019, **159**, 208–215.

59 L. Fang, X.-Q. Song, T.-T. He, K.-K. Zhu, J.-H. Yu, J.-T. Song, J. Zhou and H. Zhang, *Fitoterapia*, 2018, **129**, 150–153.

60 M. C. Carson, B. J. Orzolek and M. C. Kozlowski, *Org. Lett.*, 2022, **24**, 7250–7254.

61 S. DGS, V. R. R. Ch, T. Syed, G. Sridhar and S. R. Alapati, *Synth. Commun.*, 2024, 1–7.

62 X. Geng and S. J. Danishefsky, *Org. Lett.*, 2004, **6**, 413–416.

63 D. Sudhakar, V. R. R. Ch, T. Syed, G. Sridhar and S. R. Alapati, *Synth. Commun.*, 2024, **54**, 992–998.

64 S. Faiz and A. F. Zahoor, *Mol. Diversity*, 2016, **20**, 969–987.

65 S. Munawar, A. F. Zahoor, S. Ali, S. Javed, M. Irfan, A. Irfan, K. Kotwica-Mojzych and M. Mojzych, *Molecules*, 2022, **27**, 6953.

66 U. Tillmann, M. Elbrächter, B. Krock, U. John and A. Cembella, *Eur. J. Phycol.*, 2009, **44**, 63–79.

67 E. Ito, M. Satake, K. Ofuji, N. Kurita, T. McMahon, K. James and T. Yasumoto, *Toxicon*, 2000, **38**, 917–930.

68 Z. Zhang, Y. Chen, D. Adu-Ampratwum, A. A. Okumu, N. T. Kenton and C. J. Forsyth, *Org. Lett.*, 2016, **18**, 1824–1827.

69 M. Bader, T. D. Stark, C. Dawid, S. Lösch and T. Hofmann, *J. Agric. Food Chem.*, 2014, **62**, 2479–2488.

70 K. Aoki, Y. Igarashi, H. Nishimura, I. Morishita and K. Usui, *Tetrahedron Lett.*, 2012, **53**, 6000–6003.

71 C. Mugnaini and F. Corelli, *Synthesis*, 2016, **48**, 2085–2092.

72 A. Irfan, S. Faiz, A. Rasul, R. Zafar, A. F. Zahoor, K. Kotwica-Mojzych and M. Mojzych, *Molecules*, 2022, **27**, 1023.

73 A. Kolodiazhna and O. Kolodiazhnyi, *Phosphorus, Sulfur Silicon Relat. Elem.*, 2019, **194**, 275–276.

74 A. Kolodyazhnaya and O. Kolodyazhny, *Russ. J. Gen. Chem.*, 2019, **89**, 1998–2004.

75 S. C. Bobzin and D. J. Faulkner, *J. Org. Chem.*, 1991, **56**, 4403–4407.

76 K. Nakagawa, *Heterocycles*, 1995, **41**, 5.

77 S. J. Gharpure, D. Anuradha, J. V. Prasad and P. Srinivasa Rao, *Eur. J. Org Chem.*, 2015, **2015**, 86–90.

78 M. Borah, U. Borthakur and A. K. Saikia, *J. Org. Chem.*, 2017, **82**, 1330–1339.

79 Y. Wang, W.-Y. Zhang and S.-L. You, *J. Am. Chem. Soc.*, 2019, **141**, 2228–2232.

80 N. Gunawan, M. J. Nutt, A. C. Bissember, J. A. Smith and S. G. Stewart, *Synlett*, 2023, **34**, 1524–1528.

81 P. Krishnan, F.-K. Lee, V. A. Yap, Y.-Y. Low, T.-S. Kam, K.-T. Yong, K.-N. Ting and K.-H. Lim, *J. Nat. Prod.*, 2020, **83**, 152–158.

82 Y. Y. Mak, B. J. Loong, P. Millns, C. C. Bauer, R. S. Bon, Y. Mbaki, F. K. Lee, K. H. Lim, C. Kong and S. M. Then, *Phytother. Res.*, 2022, **36**, 2952–2963.

83 K. Annapurna and A. V. Narsaiah, *Tetrahedron*, 2024, **155**, 133909.

84 Z.-F. Zhou, M. Menna, Y.-S. Cai and Y.-W. Guo, *Chem. Rev.*, 2015, **115**, 1543–1596.

85 M. Smietana, E. Benedetti, C. Bressy and S. Arseniyadis, *Efficiency in Natural Product Total Synthesis*, 2018, 319–344.

86 J.-X. Gong, H.-Y. Wang, L.-G. Yao, X.-W. Li and Y.-W. Guo, *Synlett*, 2016, **27**, 391–394.

87 C. Djerassi and W. K. Lam, *Acc. Chem. Res.*, 1991, **24**, 69–75.

88 K. Suzuki, F. Shono, H. Kai, T. Uno and M. Uyeda, *J. Enzyme Inhib.*, 2000, **15**, 357–366.

89 N. M. Carballeira, N. Montano, R. Alvarez-Velilla, C. F. Prada, R. M. Reguera and R. Balaña-Fouce, *Mar. Drugs*, 2013, **11**, 3661–3675.

90 J. Adrian and C. B. Stark, *Eur. J. Org Chem.*, 2016, **2016**, 4607–4610.

91 R. Nshimiyimana, T. F. Lam, S. Aggarwal, C. N. Serhan and N. A. Petasis, *RSC Adv.*, 2022, **12**, 11613–11618.

92 R. Nshimiyimana, S. J. Glynn, C. N. Serhan and N. A. Petasis, *Org. Biomol. Chem.*, 2023, **21**, 1667–1673.

93 F. Fullas, D. M. Brown, M. C. Wani, M. R. Wall, T. E. Chagwedera, N. R. Farnsworth, J. M. Pezzuto and A. D. Kinghorn, *J. Nat. Prod.*, 1995, **58**, 1625–1628.

94 H. Takamura, H. Wada, N. Lu and I. Kadota, *Org. Lett.*, 2011, **13**, 3644–3647.

95 C. Garcia-Ruiz, I. Cheng-Sanchez and F. Sarabia, *Synthesis*, 2016, **48**, 1655–1662.

96 W.-g. Li, F.-y. Dai, Y.-x. Cheng, G.-f. Yin, J.-l. Bi and D.-p. Li, *Chem. Res. Chin. Univ.*, 2013, **29**, 290–293.

97 Y. Nishikawa, I. Yasuda, Y. Watanabe and T. Seto, *Yakugaku Zasshi*, 1976, **96**, 1322–1326.

98 G. A. Kraus, P. Dong, Y. Qu, A. Evans and S. Carpenter, *Tetrahedron Lett.*, 2016, **57**, 5185–5187.

99 J. Wang, A. N. Pearce, S. T. Chan, R. B. Taylor, M. J. Page, A. Valentin, M.-L. Bourguet-Kondracki, J. P. Dalton, S. Wiles and B. R. Copp, *J. Nat. Prod.*, 2016, **79**, 607–610.

100 A. Irfan, S. Faisal, A. F. Zahoor, R. Noreen, S. A. Al-Hussain, B. Tuzun, R. Javaid, A. A. Elhenawy, M. E. Zaki and S. Ahmad, *Pharmaceuticals*, 2023, **16**, 829.

101 A. F. Zahoor, M. Yousaf, R. Siddique, S. Ahmad, S. A. R. Naqvi and S. M. A. Rizvi, *Synth. Commun.*, 2017, **47**, 1021–1039.

102 M. Dumpala, S. Theegala and R. K. Palakodety, *Tetrahedron Lett.*, 2017, **58**, 1273–1275.

103 R. Negri, *Fitoterapia*, 2015, **106**, 92–109.

104 J. Geng, Q. Ren, C. Chang, X. Xie, J. Liu and Y. Du, *RSC Adv.*, 2019, **9**, 10253–10263.

105 D. B. Clarke, S. F. Hinkley and R. T. Weavers, *Tetrahedron Lett.*, 1997, **38**, 4297–4300.

106 H. Lee, T. Kang and H. Y. Lee, *Angew. Chem.*, 2017, **129**, 8366–8369.

107 J. L. von Salm, N. G. Wilson, B. A. Vesely, D. E. Kyle, J. Cuce and B. J. Baker, *Org. Lett.*, 2014, **16**, 2630–2633.

108 World Health Organization, *Fourth WHO report on neglected tropical diseases*, World Health Organization, Geneva, 2017.

109 S.-D. Chen, H. Gao, Q.-C. Zhu, Y.-Q. Wang, T. Li, Z.-Q. Mu, H.-L. Wu, T. Peng and X.-S. Yao, *Org. Lett.*, 2012, **14**, 1772–1775.

110 T. Kerl, F. Berger and H. G. Schmalz, *Chem.–Eur. J.*, 2016, **22**, 2935–2938.

111 T. Nakanishi, M. Nishi, A. Inada, H. Obata, N. Tanabe, S. Abe and M. Wakashiro, *Chem. Pharm. Bull.*, 1990, **38**, 1772–1774.

112 R. Kubínová, E. Švajdlenka, K. Schneiderová, Z. Hanáková, S. Dall'Acqua and O. Farsa, *Biochem. Syst. Ecol.*, 2013, **49**, 39–42.

113 X. Wu, H. Gao, W. Sun, J. Yu, H. Hu, Q. Xu and X. Chen, *Phytother. Res.*, 2017, **31**, 1072–1077.

114 R. J. Grayer, M. R. Eckert, N. C. Veitch, G. C. Kite, P. D. Marin, T. Kokubun, M. S. Simmonds and A. J. Paton, *Phytochemistry*, 2003, **64**, 519–528.

115 V. Timokhin, M. Regner, Y. Tsuji, J. Grabber and J. Ralph, *Synlett*, 2018, **29**, 1229–1231.

116 M. Yao, J. Zhang, S. Yang and H. Xiong, *ARKIVOC*, 2020, **2020**, 52–59.

117 A. Randazzo, C. Debitus and L. Gomez-Paloma, *Tetrahedron*, 2001, **57**, 4443–4446.

118 S. Gahalawat and S. K. Pandey, *Org. Biomol. Chem.*, 2016, **14**, 9287–9293.

119 K. Sakai, Y. Unten, M. Iwatsuki, H. Matsuo, W. Fukasawa, T. Hirose, T. Chinen, K. Nonaka, T. Nakashima and T. Sunazuka, *J. Antibiot.*, 2019, **72**, 645–652.

120 A. Kimishima, D. Hagimoto, M. Honsho, K. Sakai, S. Honma, S.-i. Fuji, M. Iwatsuki, T. Tokiwa, K. Nonaka and T. Chinen, *Org. Lett.*, 2024, **26**, 597–601.