

A water soluble fluorescent polymer as a dual colour sensor for temperature and a specific protein†

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We present two thermoresponsive water soluble copolymers prepared *via* free radical statistical copolymerization of *N*-isopropylacrylamide (NIPAm) and of oligo(ethylene glycol) methacrylates (OEGMAs), respectively, with a solvatochromic 7-(diethylamino)-3-carboxy-coumarin (DEAC)-functionalized monomer. In aqueous solutions, the NIPAm-based copolymer exhibits characteristic changes in its fluorescence profile in response to a change in solution temperature as well as to the presence of a specific protein, namely an anti-DEAC antibody. This polymer emits only weakly at low temperatures, but exhibits a marked fluorescence enhancement accompanied by a change in its emission colour when heated above its cloud point. Such drastic changes in the fluorescence and absorbance spectra are observed also upon injection of the anti-DEAC antibody, attributed to the specific binding of the antibody to DEAC moieties. Importantly, protein binding occurs exclusively when the polymer is in the well hydrated state below the cloud point, enabling a temperature control on the molecular recognition event. On the other hand, heating of the polymer–antibody complexes releases a fraction of the bound antibody. In the presence of the DEAC-functionalized monomer in this mixture, the released antibody competitively binds to the monomer and the antibody-free chains of the polymer undergo a more effective collapse and inter-aggregation. In contrast, the emission properties of the OEGMA-based analogous copolymer are rather insensitive to the thermally induced phase transition or to antibody binding. These opposite behaviours underline the need for a carefully tailored molecular design of responsive polymers aimed at specific applications, such as biosensing.

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Introduction

The need to continuously monitor and quantify an analyte or an environmental parameter in the fastest and cheapest way has been the driving force of sensor research for several decades. Given its facile operation, remote monitoring, easy read-out and high sensitivity, fluorescence is by far the most applied

detection method.^{1,2} The output signal comes in a variety of ways namely the lifetime, anisotropy or intensity of the emission of the fluorescent probe. A well-exploited sensing strategy *via* fluorescence methods employs *solvatochromic* (environmentally sensitive) dyes which change their emission and/or absorbance properties with respect to physical or chemical changes in their surroundings.³ Such environmental changes, *e.g.*, polarity or viscosity, can be induced by direct or indirect interactions of the fluorophore with target molecules whose presence and concentration can be correlated with the extent of alterations of the photophysical spectra.^{4–6} However, many of the conventional small molecule-based probes suffer from major limitations such as poor water solubility, low quantum yield, and low structural stability.⁷ Moreover, due to the presence of different media and an uncountable variety of targets to detect, not only new sensing schemes but also new materials are mandatory.¹

In this context, synthesis and characterization of a large variety of thermoresponsive polymers bearing covalently attached fluorophores has been reported.^{7–9} Typically, induced by an increase in solution temperature above their lower critical solution temperature (LCST), these polymers undergo a solubility transition which involves drastic, yet reversible changes in

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† Electronic supplementary information (ESI) available: Photophysical data of the monomer (2), temperature dependent absorbance and time-resolved PL spectra of **P1**, steady-state PL spectra of the monomer (2) at 20 and 50 °C, concentration dependent PL spectra of **P1**, decay curves of **P1** in the presence of AB, evolution of absorbance, steady state and time resolved PL spectra of aqueous **P1** solution upon heating and AB injection, evolution of absorbance and emission of the heated **P1** solution upon monomer addition, decay curves of **P2** below/above its LCST, ELISA tests on all polymers and the monomer at 4 °C, and the influence of AB on the turbidity profile of **P2**. See DOI: 10.1039/c3tb21245a

ethylmethacrylate (MEO₂MA) and oligo(ethylene glycol) methacrylate (OEGMA) monomers. The resulting NIPAm-based copolymer reports the LCST-type phase transition *via* a remarkable fluorescence output. Such a change in fluorescence characteristics is also induced as a monoclonal anti-DEAC antibody is injected into the solution. This biological sensing can be (de)activated simply by tuning the conformation of polymer chains with temperature. We suggest that the correlation of fluorescence enhancement with the solution temperature and antibody concentration should pave the way for new strategies of the detection and quantification of multiple stimuli using responsive polymers.

The NIPAm-based platform is furthermore intriguing as a biological assay. Above the LCST of the polymer, a fraction of the bound antibody is released from the polymer-attached DEAC sites out to the solution. In the presence of the DEAC-functionalized monomer in this mixture, the released antibody binds to the monomer enabling rather effective inter-chain aggregation/collapse. This *competitive binding* mechanism can be directly monitored from the polymer's temperature-turbidity profile. Remarkably, when the same dye is attached to the polymethacrylate bearing OEG side chains, we find that the fluorescence properties are almost insensitive to the polymer phase transition as well as to the presence of the antibody. This contrasting behaviour stresses the importance of structural considerations for the design of such molecular sensors.

Synthetic procedures

Synthesis of 7-(diethyl amino)-2-oxo-2H-chromene-3-carboxylic acid (7-(diethylamino) 3-carboxy coumarin) (1)

Motivated by these previously reported designs, we envisaged to construct a proof of concept, multifunctional water soluble material which can report changes in solution temperature as well as in analyte concentration through its emission profile. For this purpose, we combined the fundamental ability of the responsive polymer to react on temperature changes with the sensitivity of the probe's fluorescence to a specific, targeted biological macromolecule. Instead of utilizing an additional moiety for molecular recognition, here, the fluorophore itself acts as the recognition unit. We incorporated a solvatochromic fluorophore, 7-(diethylamino)-coumarin-3-carboxylic acid (DEAC) *via* statistical copolymerisation into two well-established types of thermoresponsive polymers based on either NIPAm or on a mixture of 2-(2'-methoxyethoxy)-

titration of hydrochloric acid while cooling the mixture in an ice bath. The orange precipitate formed was filtered off and crystallized in ethanol. Yield: 2.89 g (11.06 mmol, 55%); mp. 230 °C. Mass spectrum (ESI) signal at 262.1079 $[M + H]^+$, 284.0899 $[M + Na]^+$ found: 262.1092 $[M + H]^+$, 284.0924 $[M + Na]^+$. Elemental analysis ($C_{14}H_{15}NO_4$): calcd: C = 64.36%, H = 5.79%, N = 5.36%, found: C = 64.03%, H = 5.60%, N = 5.79%; FT-IR (cm^{-1}) selected bands: 2982 $\nu(CH_2)$, 2935 $\nu(CH_2)$, 1737 $\nu(C=O)$, 1669 $\nu(C=O)$, 1611 $\nu(C=C)$, 1582 $\nu(C=C)$; 1H NMR (300 MHz, $CDCl_3$): δ = 12.35 (s, 1H, OH), 8.65 (s, 1H, C=CH), 7.45 (d, J = 9.0 Hz, 1H, $CH_{aromatic}$), 6.71 (dd, J^1 = 9.0 Hz, J^2 = 2.4 Hz, 1H, $CH_{aromatic}$), 6.53 (d, J = 2.2 Hz, 1H, $CH_{aromatic}$), 3.50 (q, J = 7.1 Hz, 4H, CH_2), 1.27 (t, J = 7.1 Hz, 6H, CH_3), ^{13}C NMR (75 MHz, $CDCl_3$): δ = 165.6, 164.5, 158.2, 153.9, 150.4, 132.0, 111.0, 108.7, 105.8, 97.0, 45.4, 12.5.

Synthesis of 2-(7-(diethylamino)-2-oxo-2H-chromene-3-carboxylic acid amido) ethyl-methacrylate (2)

Thionyl chloride (0.33 mL, ρ = 1.64 g mL^{-1} , 4.59 mmol) was carefully added to the mixture of 7-(diethylamino)-2-oxo-2H-chromene-3-carboxylic acid (1, 1.00 g, 3.83 mmol) in pyridine. The colour of the mixture changed from green to red and then finally to black. Upon stirring this mixture for 1 h at room temperature, 2-aminoethylmethacrylate hydrochloride (755 mg, 4.67 mmol) was added to the mixture. After stirring overnight at room temperature, the solvent was evaporated, the residue was dissolved in a small amount of acetone, and this solution was poured into water. The yellow precipitate was collected *via* filtration. Yield: 1.28 g (3.31 mmol, 86%); mp. 103 °C. Mass spectrum (ESI) signal at 373.1763 $[M + H]^+$, found: 373.1735 $[M + H]^+$. Elemental analysis: calcd: C = 64.5%, H = 6.5%, N = 7.52%, found: C = 62.9%, H = 6.25%, N = 7.55%; FT-IR (cm^{-1}) selected bands: 3334 $\nu(NH)$, 2975 $\nu(CH_2)$, 2926 $\nu(CH_2)$, 1698 $\nu(C=O)$, 1650 $\delta(NH)$, 1616 $\nu(C=C)$, 1583 $\nu(C=C)$; 1H NMR (300 MHz, $CDCl_3$): δ = 9.08 (s, 1H, CONH), 8.70 (s, 1H, C=CH), 7.43 (d, J = 9.0 Hz, 1H, $CH_{aromatic}$), 6.65 (d, J = 9.0 Hz, 1H, $CH_{aromatic}$), 6.50 (s, 1H, $CH_{aromatic}$), 6.20 (s, 1H, C=CH_{2, methacrylate}), 5.60 (s, 1H, C=CH_{2, methacrylate}), 4.32 (t, J = 5.5 Hz, 2H, CH_2), 3.76 (q, J = 5.6 Hz, 2H, CH_2), 3.46 (q, J = 7.1, 4H, CH_2), 1.98 (s, 3H, $CH_3, methacrylate$), 1.24 (t, J = 7.1, 6H, CH_3) ^{13}C NMR (75 MHz, $CDCl_3$) δ = 167.4, 163.5, 162.9, 157.9, 152.8, 148.4, 131.3, 126.2, 110.2, 108.6, 96.8, 63.6, 45.3, 38.6, 18.5, 12.6.

Synthesis of the copolymers

NIPAm-based copolymer (P1). DEAC-functionalized monomer (2) and NIPAm with ca. 1 mol% of AIBN were dissolved in ethanol. The flask was sealed with a rubber septum. The mixture was purged with dry nitrogen for 30 min, and heated to 65 °C for 17 h. The solution was cooled, the solvent evaporated, and the residue was dissolved in a small amount of acetone. The solution was precipitated into diethyl ether. The precipitate was redissolved in acetone, and precipitated into diethyl ether, at least thrice, until the supernatant liquid became colourless. The polymer was dried to give a yellowish powder (labelled pNIPAm: 1.0 g NIPAm, 14 mg AIBN, 34 mg (2), 10 mL ethanol, yield: 860 mg (81%) of P1).

OEGMA-based copolymer (P2). DEAC-functionalized monomer (2) and a mixture of OEGMA (475 g mol^{-1}) and MEO₂MA with ca. 1 mol% of AIBN were dissolved in DMF (10 wt%). The flask was sealed with a rubber septum; the mixture was purged with dry nitrogen for 30 min, and heated in an oil bath to 65 °C for 17 h. The mixture was then diluted with de-ionized water, and dialyzed against de-ionised water (Roth, ZelluTrans membrane, molecular weight cut off 4000–6000). The purified polymer was isolated by freeze drying, to yield a yellow gluey mass (labelled p(MEO₂MA-co-OEGMA): 875 mg MEO₂MA, 555 mg OEGMA, 13 mg AIBN, 24 mg (2), 10 mL ethanol, yield: 1.33 g (90%) of P2).

Preparation of the monoclonal anti-DEAC antibody

The monoclonal antibody G100-HB9 was generated by standard hybridoma technology using electrofusion.²⁶ In brief, spleen cells of Balb/c mice immunized with a DEAC-KLH-conjugate (keyhole limpet hemocyanin, emp biotech, Berlin, Germany) were fused with the myeloma cell line P3 \times 63Ag8.653 (ATCC CRL-1580). Positive clones were screened for binding DEAC-ovalbumin (emp biotech) in an indirect ELISA and verified in a competitive ELISA using free DEAC. Clone G100-HB9 was mass-produced in serum free medium and the antibody was purified on protein A columns to avoid contamination with SLPI.²⁷

Photophysical characterization

Absorption spectra of solutions were recorded with a Varian Cary 5000 spectrometer. Turbidity measurements were performed on a Varian Cary 50 UV-VIS photometer. The transmittance of polymer solutions at 700 nm was monitored as a function of temperature (with a heating-cooling cycle at rates of 0.1 °C min^{-1} , cell path length of 12 mm). Steady-state fluorescence spectra were recorded with a HORIBA Jobin Yvon Fluorolog-3. The samples in PMMA cells of 1 cm path length were excited at the wavelength of their maximum absorption and the emission was detected at the front face. PL quantum efficiencies were determined with a Hamamatsu C9920 set-up, including an integrating sphere combined with a photonic multi-channel analyzer. The time-resolved fluorescence measurements were performed by using a single photon counting setup (TCSPC) with a Becker&Hickl PML-spectrometer (modified Oriel MS-125). As the excitation source, a picosecond diode laser at 405 nm was operated at a repetition rate of 20 MHz. The detector comprises a Becker&Hickl PML-16-C-1 (modified Hamamatsu) multi-alkaline photomultiplier. Considering that

$$I_{PL}(t) = \sum_{i=1}^{i=n} a_i e^{-t/\tau_i}, \quad (1)$$

where τ_i is the lifetime and a_i is the amplitude of the i^{th} component, the intensity-averaged fluorescence lifetime was calculated as

$$\langle \tau \rangle = \frac{\sum_{i=1}^{i=n} a_i \tau_i^2}{\sum_{i=1}^{i=n} a_i \tau_i} \quad (2)$$

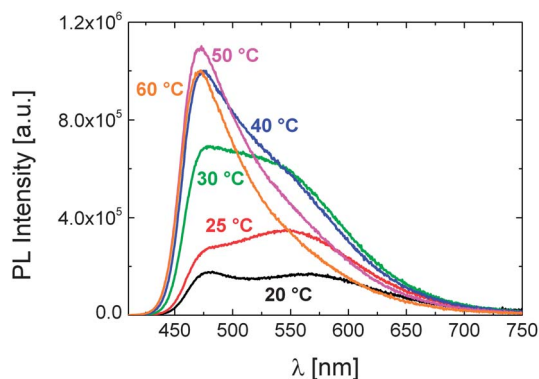


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Table 1 Molecular characterisation and thermoresponsive properties of the copolymers

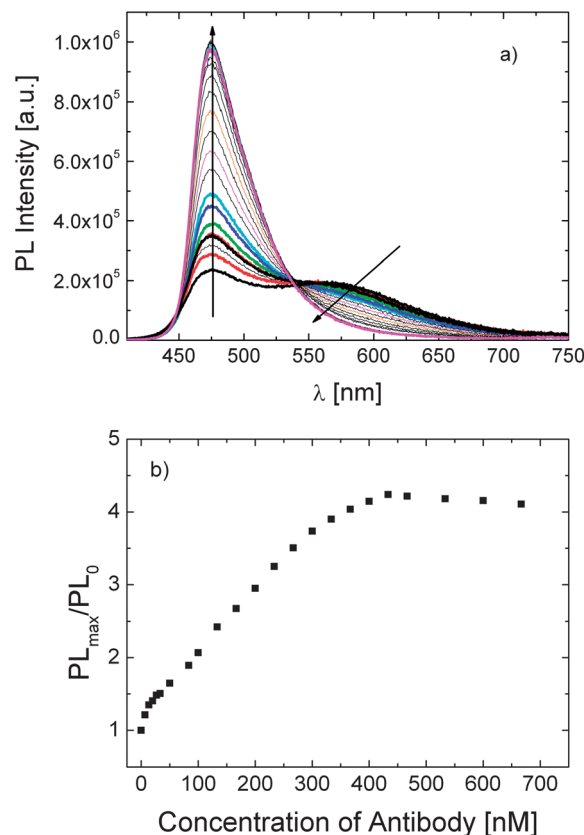
Polymer	M_n^{appa} [kg mol ⁻¹]	M_w^{app}/M_n^{appa}	DP ^b	Dye content ^c [% mol]	NC ^d	Cloud point ^e [°C]	T (50%) ^f [°C]
P1	23	1.9	196	1.1	2.2	20	23
P2	44	2.5	177	0.9	1.6	41	44

^a Determined by size exclusion chromatography in DMF, using polystyrene standards. ^b Nominal number average of monomer repeat units per polymer chain. ^c Calculated from the UV-VIS spectra of the monomer and the copolymer solutions in CHCl₃ according to Beer-Lambert's law (molar extinction coefficient (ϵ): 35,000 L mol⁻¹ cm⁻¹). ^d Average number of fluorophores per polymer chain. ^e Onset of the decrease of transmission in PBS solution (2 g L⁻¹). ^f Temperature at which the transmission is reduced by 50% in PBS solution (2 g L⁻¹).

**Fig. 2** The evolution of the PL spectra of **P1** in PBS (0.15 g L⁻¹) with temperature, recorded at an excitation wavelength of 400 nm.

transition, which sets in as the solution temperature is raised above 20 °C (cf. Table 1). Notably, the emission spectra of the monomer and of the polymer solution in PBS do not resemble each other, *i.e.* the monomer spectrum involves only the peak centred at about 475 nm. For the polymer, the species responsible for the emission at longer wavelengths also exhibit a ground-state absorption (Fig. S1†). With its relatively long emission lifetime (>10 ns), this feature is only encountered in aqueous solutions of **P1** below its LCST and disappears when the **P1** concentration is lower than 10⁻⁴ g L⁻¹ (Fig. S4†). These findings evidence the presence of DEAC aggregates, promoted presumably by interchain associations. Although it is generally favoured that the photophysical properties of the label are preserved upon incorporation into the polymer structure, the possibility to modulate the relative emission intensity of the two emitting species, *i.e.*, the single dye and its aggregates, by a stimulus, such as the temperature here, makes this polymer very promising and advantageous to construct a dual-colour sensor.

Apart from the distinguished fluorescence response to variations in solution temperature, **P1** alerts the presence of a monoclonal antibody through its optical spectra. When the anti-DEAC antibody is introduced into the aqueous **P1** solution, remarkable changes in the PL characteristics are induced (Fig. 3). Despite the similarity to those triggered by increasing temperature, these changes are more drastic: while the aggregate band is suppressed, the emission from the isolated dye moieties at shorter wavelengths builds up. The slight blue shift in λ_{max}^{PL} points to a more hydrophobic environment around the dye. As more protein is present in the solution, we measure a longer lifetime of the emission at 475 nm (Fig. S5†). For a

**Fig. 3** (a) Fluorescence spectra of **P1** solution in PBS (0.005 g L⁻¹) at 15 °C containing varying amounts of the antibody [from 6.7 nM to 660 nM]. The direction of the arrows shows the increase in antibody concentration. The excitation wavelength is 400 nm. (b) The relative change of the maximum emission intensity as a function of antibody concentration. Data are taken from Fig. 3a.

polymer concentration of 0.005 g L⁻¹, the presence of the antibody in the solution at any concentration between about 6 and 300 nM invokes correspondingly different emission intensity values, evident from the linear region shown in Fig. 3b. Also, as scattering does not interfere, the antibody-induced changes in the absorbance spectrum are clearly visible, and qualitatively correlated with its concentration (Fig. 4).

In Fig. 3 and 4, almost complete disappearance of the spectroscopic features attributed to DEAC aggregates evidences their disruption which might indicate changes in chain conformation/association. In parallel to these, the presence of antibody in the solution shifts the LCST of **P1** to somewhat higher temperatures accompanied by lowered turbidity values



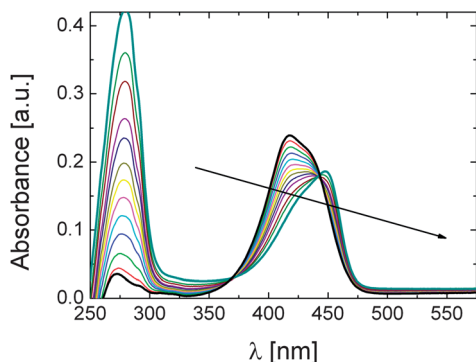


Fig. 4 Absorbance spectra of **P1** solution in PBS (0.005 g L^{-1}) at 15°C containing varying amounts of the antibody [from 6.7 nM to 660 nM]. The direction of the arrow shows the increase in antibody concentration.

(Fig. 5a). Here, in addition to the increased hydrophilicity that the polymer chains attain upon complexation with the antibody, steric hindrance might play a role in retarding and/or hindering the interchain aggregation.

Nevertheless, the effect of the interaction of a thermoresponsive polymer with another macromolecule is *a priori* not obvious. For instance, Rimmer *et al.* reported that binding of highly branched pNIPAm to a Gram negative bacterium decreased the LCST of the polymer by more than 20°C .³⁶ On the other hand, the phase transition temperature of a biotinylated OEGMA-based copolymer shifted to higher temperatures upon

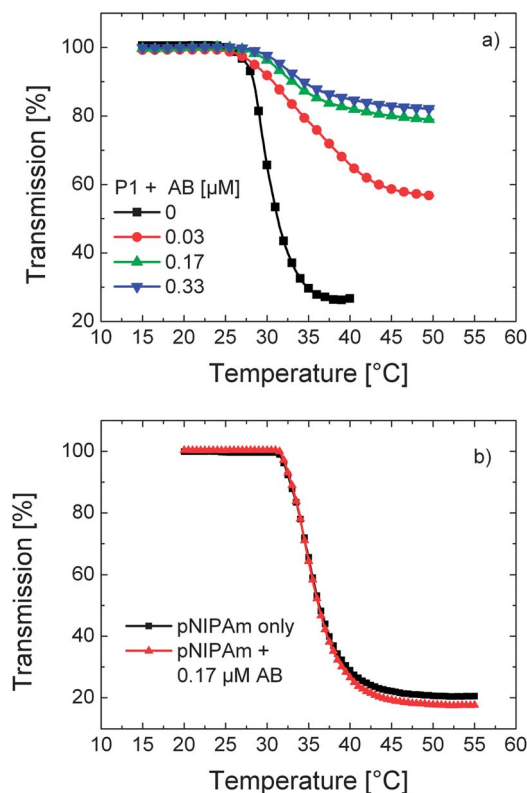


Fig. 5 Temperature-transmission profiles of (a) **P1** solution in PBS (0.1 g L^{-1}) containing varying amounts of the antibody (AB) (0 , 0.03 , 0.17 , and $0.33 \mu\text{M}$) and (b) a solution of unlabelled pNIPAm in PBS (0.01 g L^{-1}) in the presence (red triangles) and absence of the antibody ($0.17 \mu\text{M}$) (black squares).

specific binding of Avidin to the biotin moieties.³⁷ The direction of the LCST shift depends thus on the type of the analyte, on the recognition unit, and on the nature of interactions between these groups and water. It must be emphasised that pNIPAm synthesised analogously to **P1** but devoid of DEAC moieties does not exhibit changes in its turbidity profile in the presence of the antibody (Fig. 5b). This reveals that the LCST shift of **P1** is caused by the *specific binding* of the antibody to DEAC, rather than a complexation with NIPAm units. Apparently, when the antibody captures a dye, the co-aggregated chains are separated and further aggregation is inhibited. Within the binding pocket of the antibody, the dye has a rigid, water-free, and relatively nonpolar environment. This situation may be comparable to the enhanced fluorescence of DEAC dyes upon their inclusion in the hydrophobic nanocavities of cyclodextrins.³⁴ The increase in emission intensity was attributed to the inhibition of the TICT state in such an environment.

Injection of the antibody into the solution of **P1** at 40°C , *i.e.* well above its LCST, does affect neither the shape nor the intensity of the PL spectrum. However, when this mixture is cooled down to 15°C , we once again detect marked photo-physical changes indicative of the binding event (for the evolution of PL and absorbance spectra with the relevant heating and injection steps, see Fig. S6†). Regulating the access of a recognition unit linked to a thermoresponsive polymer by the particular temperature sensitivity of the chains has been reported before.^{38–40} Apparently, DEAC is accessible to the antibody only when **P1** chains adopt a well-extended conformation, whereas their collapse blocks the binding sites. Therefore, by exploiting the intrinsic response of pNIPAm to temperature, *i.e.*, changing the conformational state of the chains, we can switch the protein sensing on and off.

The control of biological sensing with temperature led us to the question whether the antibody bound to **P1** is available for *competitive binding*. Fig. 6 shows that after treatment with a 20 fold excess of DEAC molecules (the functional monomer **2**), the mixture of **P1** (0.01 g L^{-1}) with $0.17 \mu\text{M}$ of antibody exhibits a “re-clouding”. This ternary mixture was prepared by injecting the DEAC labelled monomer into the solution, comprising of **P1** and the antibody, either at 15°C followed by heating to 40°C or directly at 40°C (lines c and d, respectively). Scattering is clearly identified by the appearance of an absorption tail which extends towards the NIR region. Note that the addition of the fluorescent monomer to pure **P1** solution does not introduce such a scattering contribution (Fig. S7†). In all experiments, the heating times and rates were identical, excluding the effect of annealing on the size/density of globules formed, therefore on the scattering intensity.

The increase in turbidity (compared to the binary mixture of **P1** with the antibody) in the presence of the DEAC-labelled monomer indicates that a fraction of the antibody is released from the polymer-attached DEAC sites at 40°C . Apparently, the detached antibody forms new complexes with the monomer available in the solution so that **P1** chains can more efficiently collapse and inter-aggregate.

The question then remains whether this competitive binding occurs already at 15°C or whether the process necessitates



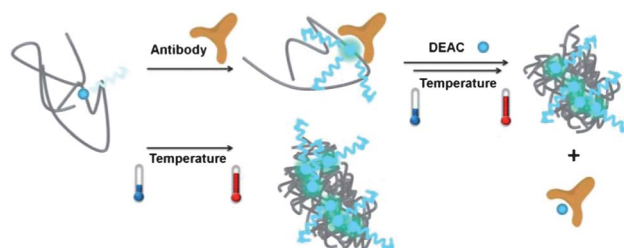
elevated temperatures. One way to address this is to examine the effect of monomer addition at 15 and at 40 °C on the fluorescence spectrum of the binary mixture at the respective temperature. As pointed out above (cf. Fig. 2), the emission spectrum of the **P1** solution below its LCST includes an aggregate-assigned band at *ca.* 575 nm. While this feature is still visible in the spectrum of the **P1** solution at 40 °C, it becomes fully suppressed upon addition of a sufficient amount of the antibody at 15 °C, below the LCST (cf. Fig. 3a). Here, we detect that the aggregate emission reappears as the monomer is injected into the mixture at 40 °C (Fig. S8a†), while no such effect is present for the low temperature spectrum of the ternary mixture prepared at 15 °C (Fig. S8b†). This shows that below the LCST, the antibody is not released from the polymer. The driving force of the observed antibody detachment at 40 °C is yet not clear, but it might be related to a temperature-activated process, e.g., increased mobility/diffusion of the sensing components or the dehydration of the chains. There have been examples in which the latter was assigned as the trigger of similar kinds of dissociation mechanisms. For example, Stayton and co-workers reported that the biotin bound-streptavidin functionalized-NIPAm chains released a significant amount of the biotin upon heating the mixture above its LCST.^{40,41} Importantly, at this temperature, the released antibody cannot re-bind to the DEAC sites of **P1** as chain collapse and aggregation partly block the accessibility of the polymer-bound dye. Instead, it remains stably bound to the low molar mass dye present excessively in the solution. The relatively firm nature of the new antibody–DEAC complexes is evidenced by the temperature–transmission profile of the ternary mixture that was cooled down to room temperature and reheated, shown in Fig. 7. The ternary mixture displays a situation that is intermediate between the presence of only pure **P1** (no interference to inter-chain aggregation) and the case where **P1** solution is saturated with the antibody (full suppression of inter-aggregation).

Temperature [°C]	P1 Transmission [%]	P1 + AB Transmission [%]	P1 + AB + DEAC Transmission [%]
20	100	100	100
25	100	100	100
30	98	100	100
35	88	100	98
40	82	100	92
45	81	100	90
50	81	100	90
55	81	100	89
60	81	100	89

sensor assays utilizing thermoresponsive polymers. Scheme 1 depicts the proposed mechanism of sensing with **P1**.

In comparison, we also examined the effect of temperature on photophysical properties of the OEGMA-based copolymer **P2**. To our surprise, no change in emission properties could be associated with its phase transition. Fig. 8 demonstrates the temperature independent shape of the PL spectrum. As the fluorescence intensity decreases gradually with the increase in temperature, the average fluorescence lifetime also shortens (Fig. S9[†]). These observations are attributed to the heat-facilitated non-radiative deactivation processes.^{22,42}

Although copolymers like **P2** exhibit an LCST-transition,^{29,37,43} the constant $\lambda_{\text{max}}^{\text{PL}}$ below and above the cloud point suggests that the dye's environment is not affected by phase transition. This is quite impressive considering the distinct response of DEAC to pNIPAm phase transition. This dependence of the dye's fluorescence response to the chemical structure of the polymer was observed for another solvatochromic fluorophore, a naphthalimide derivative, incorporated into NIPAm- or OEGMA-based copolymers analogous to **P1** and **P2**, respectively.²¹ Certain peculiarities about the



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We, lastly, addressed the effect of antibody binding on the optical spectra of **P2**. It was proven by ELISA tests that the antibody also binds to the DEAC moieties of **P2**, upon which the LCST shifts to slightly higher temperatures (Fig. S11 and S12†). However as opposed to **P1**, the emission peak displays a bathochromic shift of *ca.* 8 nm with a slight decrease in its intensity while there is no significant change in the absorbance spectrum (Fig. 9). With respect to PL spectra, similar observations were reported for a chromophore-modified cyclodextrin upon its interaction with guest molecules.⁴⁴ Bound to the guest molecule, the chromophore inherently located in the interior of the hydrophobic cavity of dextrin was excluded into the polar aqueous solution. Although the dyes which are bound to the antibody have now less segmental mobility, i.e. an inhibited

Fig. 9 Normalised absorbance (solid lines) and PL (dashed lines) spectra of the aqueous solution of **P2** (0.01 g L^{-1}) in the presence and absence of $0.17 \text{ }\mu\text{M}$ of antibody at $20 \text{ }^{\circ}\text{C}$. The absolute PL spectra are given in Fig. S10.†

TICT state, this new environment might be less hydrophobic than that provided by the polymer chains, leading to the slight red-shift in the emission spectrum. On the other hand, the antibody complexion with a certain fraction of the dyes might enhance the water exposure of some others as a result of the expansion of the chains. A detailed analysis on the processes responsible for such spectral changes is on the way.

Conclusions

We demonstrate the synthesis and photophysical characterisation of two thermoresponsive polymers bearing a covalently attached TICT-type DEAC dye, promising for optical sensing applications. In aqueous solutions of the OEGMA-based polymer, the quantum yield of DEAC is relatively high. The dye is mostly insensitive to the polymer phase transition, and to the anti-DEAC antibody. In contrast, aqueous solutions of the NIPAm-based polymer exhibit a very weak emission at low temperatures or in the absence of antibody, which increases dramatically above the LCST or upon binding of the antibody. Heating of the solution of antibody-bound polymer above its cloud point releases a fraction of the immobilized antibody. In the presence of the DEAC-labelled monomer in this mixture, the detached antibody binds competitively to the low molar mass monomer allowing the free polymer chains to aggregate. Consequently, on one hand, this work presents a water soluble polymer with a distinct fluorescence response towards temperature and a biological macromolecule. On the other hand, by comparing two analogous DEAC-labelled polymers, the need to design a specific polymer structure is stressed, *e.g.*, composed of alternative thermo-sensitive blocks, one of which only produces a marked fluorescence signal of the thermal transition, or of the binding event.

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