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# Molecular speciation controls arsenic and lead bioaccessibility in fugitive dusts from sulfidic mine tailings†

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Communities nearby mine wastes in arid and semi-arid regions are potentially exposed to high concentrations of toxic metal(loid)s from fugitive dusts deriving from impoundments. To assess the relation between potentially lofted particles and human health risk, we studied the relationship between pharmacokinetic bioaccessibility and metal(loid) molecular speciation for mine tailings dust particulate matter (PM), with elevated levels of arsenic and lead (up to 59 and 34 mmol kg<sup>-1</sup>, respectively), by coupling *in vitro* bioassay (IVBA) with X-ray absorption spectroscopy (XAS). Mine tailing efflorescent salts (PM<sub>ES</sub>) and PM from the surface crust (0–1 cm, PM<sub>SC</sub>) and near surface (0–25 cm) were isolated to <10 μm and <150 μm effective spherical diameter (PM<sub>10</sub> and PM<sub>150</sub>) and reacted with synthetic gastric and lung fluid for 30 s to 100 h to investigate toxic metal(loid) release kinetics. Bioaccessible (BAC) fractions of arsenic and lead were about 10 and 100 times greater in gastric than in lung fluid simulant, respectively, and 10–100% of the maximum gastric BAC from PM<sub>10</sub> and PM<sub>150</sub> occurred within 30 s, with parabolic dissolution of fine, highly-reactive particles followed by slower release from less soluble sources. Evaporite salts were almost completely solubilized in gastric-fluid simulants. Arsenate within jarosite and sorbed to ferrihydrite, and lead from anglesite, were identified by XAS as the principal contaminant sources in the near surface tailings. In the synthetic lung fluid, arsenic was released continuously to 100 h, suggesting that residence time *in vivo* must be considered for risk determination. Analysis of pre- and post-IVBA PM indicated the release of arsenic in lung fluid was principally from arsenic-substituted jarosite, whereas in synthetic gastric fluid arsenic complexed on ferrihydrite surfaces was preferentially released and subsequently repartitioned to jarosite-like coordination at extended exposures. Lead dissolved at 30 s was subsequently repartitioned back to the solid phase as pyromorphite in phosphate rich lung fluid. The bioaccessibility of lead in surface tailings PM was limited due to robust sequestration in plumbojarosite. Kinetic release of toxic elements in both synthetic biofluids indicated that a single IVBA interval may not adequately describe release dynamics.

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## Environmental significance

Human health risks from particulate matter (PM) are becoming increasingly clear. Mine tailings serve as PM sources, especially in dry climates. The US Agency for Toxic Substances and Disease Registry shows the top 2 substances of concern are arsenic (#1) and lead (#2), metal(loid)s commonly found in mine tailings. With the associated likelihood of off-site dust transport, inhalation and ingestion of PM can lead to toxic exposures with biological effects controlled by the rate of dissolution *in vivo*. However, the bioavailability, toxicity, and uptake of metal(loid)s also depends on their solid phase speciation, which can be probed with X-ray spectroscopy. This study uniquely combines pharmacokinetic bioassay and solid phase speciation to determine toxic metal(loid) source and kinetic release *in vitro*.

## 1 Introduction

Mine tailings are a significant source of fugitive dust pollution in the form of atmospheric particulate matter (PM). Since fine-grained, unconsolidated, metalliferous particles can be subjected to suspension and transport by wind (and water), potentially resulting in direct human exposure to toxicants, there is a need to know the lability and precise molecular form or speciation of metal(loid)s that may emit from a tailings

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† Electronic supplementary information (ESI) available: The study site, biofluid simulant formulations, details of synchrotron X-ray analysis; XANES fits; XRD; fit spectra for sulfur, iron, arsenic, and lead; tabulated EXAFS fits; zinc IVBA for PM<sub>ES</sub> and PM<sub>SC</sub>; SEM-EDS for PM<sub>ES</sub>, PM<sub>10</sub> and PM<sub>150</sub>, and total elemental analysis of PM<sub>10</sub> and bulk tailings. See <https://doi.org/10.1039/d2em00182a>



Assessment of risk to health from ingested or inhaled contaminants must account for the total toxicant concentration ( $C_i$ ) and its bioactivity ( $a_{i\text{-bio}}$ ), the fraction absorbed at a target organ.<sup>31</sup> Animal model *in vivo* assays have been used to monitor contaminant uptake and elimination to access relative bioavailability (RBA), and *in vitro* bioassays (IVBA) have been used to determine solubilization of particles under lab-controlled biomimicry to access bioaccessibility (BAC), conditionally-defined as the concentration of an analyte of interest (*e.g.*, As, Pb) released to solution in biofluid

Total elemental concentrations were analyzed by inductively coupled plasma mass spectrometry (ICP-MS, PerkinElmer, Elan DRC-II) following microwave assisted dissolution by aqua regia (Arizona Laboratory for Emerging Contaminants, ALEC, University of Arizona) and HF, HNO<sub>3</sub>, HClO<sub>4</sub>, and HCl (Activation Labs, Ontario CA). All chemicals were reagent grade or better and all labware was acid washed and metal-free. Total arsenic was analyzed by instrumental neutron activation analysis (INAA) and total iron was analyzed by ICP-AES following dissolution of the tailings in a lithium metaborate flux-fusion (with LiBO<sub>2</sub> and Li<sub>2</sub>B<sub>4</sub>O<sub>7</sub>) (Activation Labs, Ontario CA).

Certified reference materials were digested and analyzed along with the tailing samples with an acceptance range of  $\pm 10\%$  of the certified value to verify precision and accuracy in sample preparation and analysis. Specific surface area (SSA) was measured by multipoint BET (Brunauer, Emmett, and Teller, Micromeritics Gemini VII 2390 t, Norcross, GA).

## 2.2 Bioaccessibility of metal(loid)s

BAC was used to assess the metal(loid) exposure risk by monitoring the concentration of As, Pb, and host mineral Fe solubilized into IVBA synthetic biofluid and available for biologic absorption.<sup>8,45,46</sup>

$$\text{BAC}_{(c_i)} = \frac{[c_i]_{\text{IVBA}}}{[c_i]_{\text{total}}} \times 100 \quad (1)$$

where the BAC fraction of element  $c_i$  is the concentration dissolved ( $\text{mmol kg}^{-1}$ ) in synthetic lung fluid (SLF) or synthetic gastric fluid (SGF)  $[[c_i]_{\text{IVBA}}]$  normalized to the total solid phase concentration  $[[c_i]_{\text{total}}]$  in the tailings PM sample.

The lung fluid simulant was a modified Gamble's solution, used previously to mimic the fluid released by Type II alveolar cells and estimate the potential *in vivo* dissolution of metal(-loids) following inhalation of dusts.<sup>47–49</sup> The solution contained a mixture of mineral salts, humectants, and an emulsifier, but omitted proteins to eschew putrefaction (Table S1†), and has been found to be analogous to human alveolar fluid in terms of major components.<sup>50,51</sup> This SLF recipe has been used to determine the BAC of various respirable soil particles including, silica,<sup>52</sup> cadmium,<sup>53</sup> uranium,<sup>54</sup> and lead.<sup>55</sup>

It has been established that among the many *in vitro* gastric extractions, a robust predictor of *in vivo* bioavailability, *i.e.*, a consistently good correlation between *in vitro* BAC and *in vivo* RBA with juvenile swine model for gastric exposures, is a low pH (1.5) fluid buffered with hydrochloric acid and glycine.<sup>56,57</sup> Whereas the large swine body size is favorable for repeated blood sampling, this requires surgery, and swine are expensive and difficult to handle in a laboratory setting.<sup>58</sup> For comparing *in vitro* and *in vivo* bioassays, mouse models are favorable due to their smaller size, lower expense, availability, relative ease of use and are already widely applied in medical research.<sup>1,59–62</sup> However, mouse arsenic metabolism differs from that of humans and their small body weight and blood volume is prohibitive for repeat blood sampling.<sup>63,64</sup> Despite these complications, mice and swine are both considered suitable animal bioassays for validation of *in vitro* bioavailability studies. The chosen IVBA method in this study is representative of both USEPA Method 1340 and the Solubility Bioaccessibility Research Consortium (SRC) assay.<sup>39,65</sup> This SGF mimics conditions of a fasting stomach and was adopted by USEPA bioavailability protocol.<sup>66,67</sup> The SGF, prepared in  $\text{N}_2$  sparged deionized water (Milli-Q, 18.2 M $\Omega$ ) adjusted to pH 1.5 with concentrated HCl (ACS grade) and buffered with 0.4 M glycine, has been established to be appropriate for mining wastes<sup>68</sup> and mine impacted soils, and has been validated as a good predictor of arsenic RBA in contaminated soil.<sup>1,57</sup> This single-step gastric fluid approach was used in the current study.<sup>66,69–71</sup>

Kinetic IVBA experiments with SLF and SGF were carried out in triplicate in batch-mode. In each IVBA vessel, a 0.15 g aliquot of tailings PM was reacted with 15.0 g of SLF or SGF in a metal-free polypropylene tube (VWR, PN 89049-172). The IVBA solution was pre-heated to 37.0 °C to simulate internal body temperature and added to the tailings. The vessel was wrapped with aluminum foil to eschew photoreaction, placed on an end over end rotator in a 37.0 °C incubator at 60 rpm for prescribed durations. Reaction times <5 min were agitated by hand in pre-heated (37.0 °C) biofluid. At each interval, samples were removed from the rotator, centrifuged at 23 600 g, and the supernatant solution was aspirated and syringe filtered to 0.45  $\mu\text{m}$  with acid washed hydrophobic polypropylene GHP membranes (acro-disc, Pall Corp. PN 28139). The filtered supernatant was analyzed for pH to assure changes were <0.5 and by ICP-MS for the elemental BAC concentrations. The concentration of the target analyte that migrated from the solid to aqueous phase was operationally defined as the kinetic-step BAC concentration. The residue was washed with DI water and lyophilized at  $-45$  °C and 0.013 kPa prior to solid phase characterization and spectroscopic analysis. Kinetic studies of the tailing samples included five sample times for the  $\text{PM}_{\text{ES}}$  and  $\text{PM}_{\text{SC}}$  from 0.5 h to 48 h, and nine steps from 30 s to 100 h for the  $\text{PM}_{10}$  and  $\text{PM}_{150}$  tailings. Exposure durations were selected to develop (i) relevant particle resident times in lung (days) and gastric (hours) systems and (ii) an understanding of the kinetic release rates of metal(loid)s in these systems at shorter and longer retention times.

## 2.3 X-ray techniques

Bulk PM minerals were identified on pre- and post-IVBA reacted tailings using synchrotron transmission X-ray diffraction (ST-XRD) at 0.965 Å with a CCD detector for collection of Laue images at beamline 11–3 at the Stanford Synchrotron Radiation Lightsource (SSRL, Menlo Park, CA). Diffractograms were converted to conventional Cu K $\alpha$  wavelength and normalized to the  $hkl_{112}$  quartz reflection (1.818 Å) for comparative analysis. Synchrotron X-ray absorption near-edge spectroscopy (XANES) and extended X-ray absorption fine structure (EXAFS) spectra were collected for speciation of arsenic, iron, (K-edge, 11 867 and 7112 eV) and lead (L<sub>III</sub>-edge 13 035 eV) at beamline 11–2 at SSRL. Sulfur (K-edge 2472.04 eV) XANES were collected at beamline 4–3 at SSRL. General procedures for XAS collection and data processing were described previously.<sup>15</sup> The mixed ferric phases in PM were resolved using a non-linear least-squares fit to the Fe EXAFS constrained by a two-mineral model for shell-by-shell fits, simultaneously fitting a jarosite model component and a ferrihydrite model component to the tailing PM samples. Details of sample handling, preparation, data collection and analysis are provided in the (ESI†).

# 3 Results

## 3.1 Characterization of particulate matter

Both arsenic and lead in the tailing samples exceed the regional background levels ( $[\text{As}]_{\text{bkg}} = 0.33 \text{ mmol kg}^{-1}$ ,  $[\text{Pb}]_{\text{bkg}} =$



0.10 mmol kg<sup>-1</sup>),<sup>72</sup> with tailing PM exhibiting multi-metal enrichment.<sup>15</sup> Total metal(loid) concentration increased with decreasing particle-size, consistent with previous studies of mine tailings<sup>8,12</sup> (Table 1). The iron-normalized molar concentrations of arsenic and lead (As : Fe, Pb : Fe) varied by sample type, with values of about 0.02 and 0.01, respectively, for PM<sub>10</sub> and PM<sub>SC</sub>, while PM<sub>150</sub> had lower values of about 0.015 and 0.005, respectively, indicating a relative enrichment in both arsenic and lead in PM<sub>10</sub>. The PM<sub>ES</sub>, which had <63 µm ESD,

were completely soluble in aqua regia digestion, but had only ca. 0.1% of the arsenic and lead content of the other PM. Further, in comparison to the other geodusts, the PM<sub>ES</sub> were depleted in arsenic and lead with As : Fe and Pb : Fe values of 0.0002 and 0.0026. However, PM<sub>ES</sub> had almost 10x elevated Zn ([Zn]<sub>ES</sub> = 496 µmol g<sup>-1</sup>) compared to other PM (e.g. [Zn]<sub>150</sub> = 59, [Zn]<sub>SC</sub> = 46.5 µmol g<sup>-1</sup>) (Fig. S2 and S3†). Despite the greater solubility of efflorescent salts, the arsenic and lead inhalation exposures from PM<sub>ES</sub> were expected to be lower than other the particles as exposure is a function of total concentration and solubility (Table 1).

Major crystalline phases in PM<sub>10</sub>, PM<sub>150</sub>, and PM<sub>SC</sub> determined by synchrotron XRD were quartz [SiO<sub>2</sub>], albite [NaAlSi<sub>3</sub>O<sub>8</sub>], gypsum [CaSO<sub>4</sub>·2H<sub>2</sub>O], pyrite [FeS<sub>2</sub>], jarosite [KFe<sub>3</sub>(SO<sub>4</sub>)<sub>2</sub>(OH)<sub>6</sub>], and chlorite [(Mg,Fe)<sub>5</sub>Al<sub>2</sub>Si<sub>3</sub>O<sub>10</sub>(OH)<sub>8</sub>] (Table 1). Crystalline arsenic or lead phases were not detected by XRD (detection limit ~2 wt%). Efflorescent salts (PM<sub>ES</sub>) comprised gypsum, magnesiocopiapite [MgFe<sup>(III)</sup><sub>4</sub>(SO<sub>4</sub>)<sub>6</sub>(OH)<sub>2</sub>·20H<sub>2</sub>O], goslarite [ZnSO<sub>4</sub>·7H<sub>2</sub>O], copiapite [Fe<sup>(III)</sup>Fe<sup>(III)</sup><sub>4</sub>(SO<sub>4</sub>)<sub>6</sub>(OH)<sub>2</sub>·20H<sub>2</sub>O], hydrated ferric sulfate salts [Fe<sub>2</sub>(SO<sub>4</sub>)<sub>3</sub>·xH<sub>2</sub>O], ferrihydrite [Fe<sub>2</sub>O<sub>3</sub>·5H<sub>2</sub>O], and jarosite. The effective enrichment of metal(loid)s in the finest particulates was due to depletion of coarser primary silicate minerals in the fine size fractions.

### 3.2 Lung fluid IVBA is

**3.2.1 Kinetics.** The fractional releases of arsenic and lead, and the dissolution of host ferric minerals were interrogated with a circumneutral SLF to assess *in vitro* kinetic exposure representing inhalation of PM, where elemental release to the biofluid simulant represented the BAC fraction relative to the total PM concentration. Synthetic lung fluid IVBA showed generally low BAC for metal(loid)s in each PM compared to gastric exposures (about 1–10%) (Table 1), with the greatest relative release occurring at the shortest time-points (e.g., minutes) (Fig. 1). Following this rapid pharmacokinetic release, in the closed IVBA batch reaction, iron and lead were observed to repartition back to the solid phase at longer exposures (>24 h). Arsenic showed release to SLF from PM<sub>10</sub>, PM<sub>150</sub> and PM<sub>SC</sub>, increasing at extended times, whereas PM<sub>ES</sub> at 48 h showed no significant difference from the maximum release at 5 min. The BET SSA for PM<sub>150</sub> increased from 2.3 m<sup>2</sup> g<sup>-1</sup> (unreacted) to 9.6 m<sup>2</sup> g<sup>-1</sup> after 48 h SLF as higher specific surface area particles or mineral coatings precipitated in the lung fluid.

The PM<sub>10</sub>, which showed approximately 2x enrichment of arsenic and lead relative to the PM<sub>150</sub> and PM<sub>SC</sub> and over 2000x enrichment relative to PM<sub>ES</sub>, had a maximum arsenic BAC of 3.6% compared to 2.2% for PM<sub>150</sub>, 0.37% in PM<sub>SC</sub>, and 2.3% in PM<sub>ES</sub> (Table 1). Lead BAC was not significantly different (*p* > 0.05) for PM<sub>10</sub>, PM<sub>150</sub>, and PM<sub>SC</sub> at 0.02%, but 67.5% in PM<sub>ES</sub>.

At 24 h in SLF, the generally used IVBA time point for lung exposure, the arsenic released from the PM<sub>10</sub> was more than 2x PM<sub>150</sub>, and the BAC was significantly larger (*p* < 0.05) than each of the other PM. The lead BAC was low and not significantly different among PM<sub>10</sub>, PM<sub>150</sub> and PM<sub>SC</sub>; the release at 24 h from PM<sub>ES</sub> was the highest BAC (7.8%), and was three orders-of-

Table 1 Bioaccessible iron, arsenic, and lead in Iron King Mine Tailings

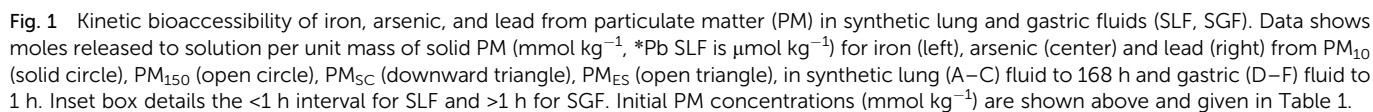
| Sample                                                        | [Fe]                          | [As]                                                | [Pb]               |
|---------------------------------------------------------------|-------------------------------|-----------------------------------------------------|--------------------|
| <b>Total concentration<sup>a</sup> (mmol·kg<sup>-1</sup>)</b> |                               |                                                     |                    |
| PM <sub>10</sub>                                              | 3330                          | 58.7                                                | 34.0               |
| PM <sub>150</sub>                                             | 2220                          | 34.3                                                | 11.4               |
| PM <sub>SC</sub>                                              | 1890                          | 38.3                                                | 19.0               |
| PM <sub>ES</sub>                                              | 93.1                          | 0.0220                                              | 0.0222             |
| SRM 2782                                                      | 4820                          | 2.22                                                | 2.77               |
| Background <sup>b</sup>                                       | na                            | 0.297                                               | 0.101              |
| <b>Molecular speciation<sup>c</sup></b>                       |                               |                                                     |                    |
|                                                               | fh jar pyt <sup>■</sup> /cop* | As <sup>V</sup> -fh As <sup>V</sup> -jar Pb-jar ang |                    |
| PM <sub>10</sub>                                              | 55 41 3 <sup>■</sup>          | 24 76 86 14                                         |                    |
| PM <sub>150</sub>                                             | 30 61 8 <sup>■</sup>          | 38 62 71 29                                         |                    |
| PM <sub>SC</sub>                                              | 39 52 10 <sup>■</sup>         | 46 54 na                                            |                    |
| PM <sub>ES</sub>                                              | 26 39 35*                     | <DL <DL                                             |                    |
| <b>Maximum percent BAC lung<sup>d</sup></b>                   |                               |                                                     |                    |
| PM <sub>10</sub>                                              | 1.55 <sup>1</sup>             | 3.64 <sup>2</sup>                                   | 0.016 <sup>1</sup> |
| PM <sub>150</sub>                                             | 0.15 <sup>3</sup>             | 1.31 <sup>2</sup>                                   | 0.022 <sup>2</sup> |
| PM <sub>SC</sub>                                              | 0.12 <sup>4</sup>             | 0.37 <sup>5</sup>                                   | 0.02 <sup>5</sup>  |
| PM <sub>ES</sub>                                              | 28.3 <sup>4</sup>             | 2.3 <sup>4</sup>                                    | 67.5 <sup>6</sup>  |
| SRM 2782                                                      | na                            | 1.1 <sup>2</sup>                                    | <0.01              |
| <b>Percent BAC lung 24 h</b>                                  |                               |                                                     |                    |
| PM <sub>10</sub>                                              | 0.54 <sup>a</sup>             | 1.16 <sup>d</sup>                                   | 0.003 <sup>g</sup> |
| PM <sub>150</sub>                                             | 0.07 <sup>b</sup>             | 0.74 <sup>c</sup>                                   | 0.003 <sup>g</sup> |
| PM <sub>SC</sub>                                              | 0.07 <sup>b</sup>             | 0.22 <sup>f</sup>                                   | 0.006 <sup>g</sup> |
| PM <sub>ES</sub>                                              | 2.3 <sup>c</sup>              | 0.74 <sup>e</sup>                                   | 7.8 <sup>h</sup>   |
| <b>Maximum percent BAC gastric<sup>d</sup></b>                |                               |                                                     |                    |
| PM <sub>10</sub>                                              | 19.1 <sup>7</sup>             | 11.7 <sup>7</sup>                                   | 3.00 <sup>7</sup>  |
| PM <sub>150</sub>                                             | 32.9 <sup>7</sup>             | 25.3 <sup>7</sup>                                   | 18.2 <sup>7</sup>  |
| PM <sub>SC</sub>                                              | 5.8 <sup>5</sup>              | 13.3 <sup>5</sup>                                   | 0.87 <sup>8</sup>  |
| PM <sub>ES</sub>                                              | 93.3 <sup>5</sup>             | 87.9 <sup>5</sup>                                   | 86.3 <sup>5</sup>  |
| SRM 2782                                                      | na                            | 25.4 <sup>7</sup>                                   | 92.5 <sup>7</sup>  |
| <b>Percent BAC gastric 1 h</b>                                |                               |                                                     |                    |
| PM <sub>10</sub>                                              | 2.9 <sup>i</sup>              | 7.8 <sup>l</sup>                                    | 0.77 <sup>o</sup>  |
| PM <sub>150</sub>                                             | 3.6 <sup>i</sup>              | 9.9 <sup>l</sup>                                    | 0.66 <sup>o</sup>  |
| PM <sub>SC</sub>                                              | 1.1 <sup>j</sup>              | 4.3 <sup>m</sup>                                    | 0.11 <sup>p</sup>  |
| PM <sub>ES</sub>                                              | 76.7 <sup>k</sup>             | 44.9 <sup>n</sup>                                   | 74.5 <sup>q</sup>  |

<sup>a</sup> Total concentration in PM<sub>10</sub>, PM<sub>150</sub>, and PM<sub>SC</sub> determined by Li<sub>2</sub>B<sub>4</sub>O<sub>7</sub>/LiBO<sub>2</sub> fusion with ICP-MS detection, PM<sub>ES</sub> total concentrations determined by aqua regia digest with ICP-MS detection, SRM 2782 is the assigned NIST value. <sup>b</sup> Background is the site-specific level.<sup>73</sup>

<sup>c</sup> Molecular speciation of each element as determined by XAS, <sup>a-q</sup> BAC significantly different at *p* < 0.05 for 24 h SLF and 1 h SGF.

<sup>d</sup> Maximum BAC at specified time points: <sup>1</sup>30 s, <sup>2</sup>168 h (7d), <sup>3</sup>5 min, <sup>4</sup>0.5 h, <sup>5</sup>48 h, <sup>6</sup>1 h, <sup>7</sup>100 h, <sup>8</sup>24 h; fh = ferrihydrite, jar = jarosite, pyt<sup>■</sup> = pyrite (from Hayes *et al.*, 2015), ang = anglesite, cop\* = copiapite, <DL = less than detection limit, na = not applicable, IVBA results reported are average of triplicates.





3.2.2.1 *Iron species.* Evaluation of the PM<sub>10</sub> Fe K $\alpha$  XANES pre-edge (7110–7118 eV), a feature resulting from an allowed transition of the 1s electron to a 3d orbital, showed slight changes in the three assigned peaks (*a-c*) that make up the pre-edge (Fig. 2A). The lowest energy peak (*a*) at 7112 eV was present in jarosite and absent in ferrihydrite. The *a* peak decreased in the 48 h SLF reacted sample compared to the unreacted PM<sub>10</sub> particles. Additionally, the subsequent pre-edge peaks (*b*) at 7113 eV and (*c*) at 7115 eV show a relative increase in *c*, over *b*, post IVBA. The pre-edge in the PM<sub>150</sub> showed slight changes in the three assigned peaks (*a-c*), similar to PM<sub>10</sub> with the (*a*) peak diminished in SLF at 48 h (Fig. S9†). Linear combination fits (LCF) to the PM<sub>10</sub> Fe EXAFS showed that the iron in the unreacted tailings was comprised of jarosite (41%), ferrihydrite (55%) and pyrite (3%), PM<sub>150</sub> was jarosite (61%), ferrihydrite (30%) and pyrite (8%), PM<sub>SC</sub> had, jarosite (52%), ferrihydrite (39%), and pyrite (10%), and unreacted PM<sub>ES</sub> was jarosite (39%), ferrihydrite (26%), and copiapite (35%). Iron EXAFS of unreacted and 48 h post IVBA PM tailings were compared. The 1<sup>st</sup> shell feature in the Fourier transformed Fe K $\alpha$  EXAFS, attributed to the Fe<sup>III</sup>-O<sub>6</sub> backscattering, increased in amplitude for the 48 h SLF-reacted PM<sub>10</sub> relative to the unreacted PM<sub>10</sub> and the Fe-O radial distance increased from 1.94 Å to 1.97

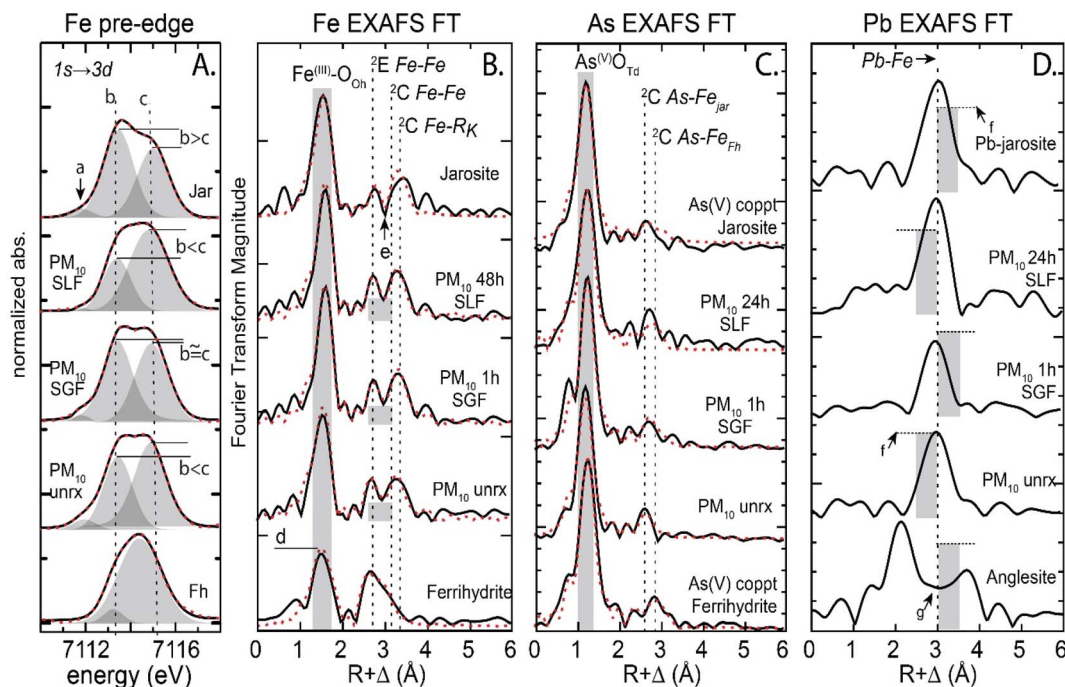


Fig. 2 Iron XANES pre-edge and iron, arsenic, and lead EXAFS Fourier transform of unreacted and IVBA reacted  $PM_{10}$  from Iron King mine tailings. Panel A shows Fe XANES pre-edge, with peaks  $a-c$  characteristic of ferrihydrite v jarosite in oxidized sulfidic tailings. Panel B shows the EXAFS FT for  $PM_{10}$  reacted in lung and gastric fluid *in vitro*,  $d$  indicates Fe–O backscattering and highlights FWHM peak change for ferrihydrite v jarosite and  $e$  indicates separation of edge sharing Fe–S/Fe and corner sharing Fe–Fe in ferrihydrite and jarosite, fits shown in Table S2.† Panel C shows As EXAFS FT with first shell As–O at  $1.68 \pm 0.02$  Å and second shell As–Fe  $\Delta 0.02$  Å. Panel D shows Pb EXAFS FT,  $f$  Pb–Fe FT peak amplitude is greater for  $PM_{10}$  unrx.  $g$  indicates region of FT where Pb–O features would be observed for Pb-sorbed ferrihydrite.

Å (Fig. 2B and S10, Table S2†). Second shell contributions for near neighbor iron backscattering atoms varied with the contribution of jarosite and ferrihydrite, and fits were constrained with a binary structural model for fitting. Backscatterer Fe–Fe distances varied by the major contribution from the identified species, where longer corner sharing Fe–Fe distances at  $3.64 \pm 0.01$  Å and Fe–S at  $3.24 \pm 0.01$  Å were assigned to jarosite and shorter Fe–Fe distances of  $3.07 \pm 0.02$  Å and  $3.36 \pm 0.01$  Å were assigned to ferrihydrite. At 1 h,  $PM_{10}$  showed a decrease by about 50% in coordination of the Fe–Fe at  $3.08 \pm 0.1$  Å and  $3.35 \pm 0.1$  Å from 1.5 and 3.1 to 0.7 and 1.4, respectively (Table S2†). These distances were attributed to the contribution from Fe–Fe backscattering from edge-sharing ( $^2E$ ) and corner-sharing ( $^2C$ )  $Fe^{III}$  octahedra in ferrihydrite, respectively.<sup>74</sup> The Fe second shell contribution showed a decrease in the contribution from ferrihydrite at 1 h and longer, e.g., 48 h (Table S2†). No significant changes were observed in the jarosite associated backscattering near neighbors. The post SLF IVBA  $PM_{150}$  Fe EXAFS showed small decreases in coordination of the Fe–S and Fe–Fe to 1.4 and 1.2, respectively (Fig. S11 and Table S3†). While the minor change noted in the jarosite associated Fe–Fe backscattering near neighbors at 3.63 Å was within the error of fitting coordination numbers in complex matrices, it is nonetheless consistent with a reduction in the jarosite contribution to the EXAFS signal. Fitting the post IVBA  $PM_{ES}$  showed significant alteration of the Fe second shell neighbors with a depletion in Fe–S at 3.27 Å and alteration of the Fe–Fe 3.26 Å

and 3.40 Å distances associated with ferrihydrite (Table S4†). The complex dissolution and reprecipitation in the  $PM_{ES}$  is not resolvable by Fe EXAFS but does indicate a loss of Fe–S second shell ligand, likely attributed to dissolution and loss of soluble ferric sulfate salts and jarosite.

**3.2.2.2 Sulfur species.** The unreacted  $PM_{10}$  sulfur speciation was dominated by gypsum (49.1%), which was depleted by extended incubation *in vitro* (Table S5, Fig S12 and S13†). The relative contribution of pyrite to the sulfur XANES increased with each time step, as sulfate species were solubilized. Other components of the fits increased concomitantly, with the relative contribution of pyrite doubling, and the jarosite signal increasing slightly less than the ferrihydrite-adsorbed sulfate contribution. Sulfur speciation in unreacted  $PM_{150}$  was mostly jarosite (60%), and at long reaction times 7 d in the SLF the contribution from pyrite was unchanged, jarosite increased to over 80% of the S signal while gypsum and adsorbed sulfate were depleted relative to other species.

**3.2.2.3 Arsenic species.** The solid phase arsenic speciation at each kinetic step probed by EXAFS showed  $As^{(V)}-O_4$  in tetrahedral coordination at  $1.68 \pm 0.02$  Å (Table S6 and S7†). The As EXAFS showed changes in second shell ligand distances as a function of the mixing ratio of ferrihydrite and jarosite associated arsenate (Fig. S14†). The spectral contribution from jarosite associated arsenate was greater than arsenic sorbed ferrihydrite in  $PM_{10}$ ,  $PM_{150}$ , and  $PM_{SC}$  and was not detectable in  $PM_{ES}$  (Table 1). In the unreacted  $PM_{10}$ , 76% of the arsenic was





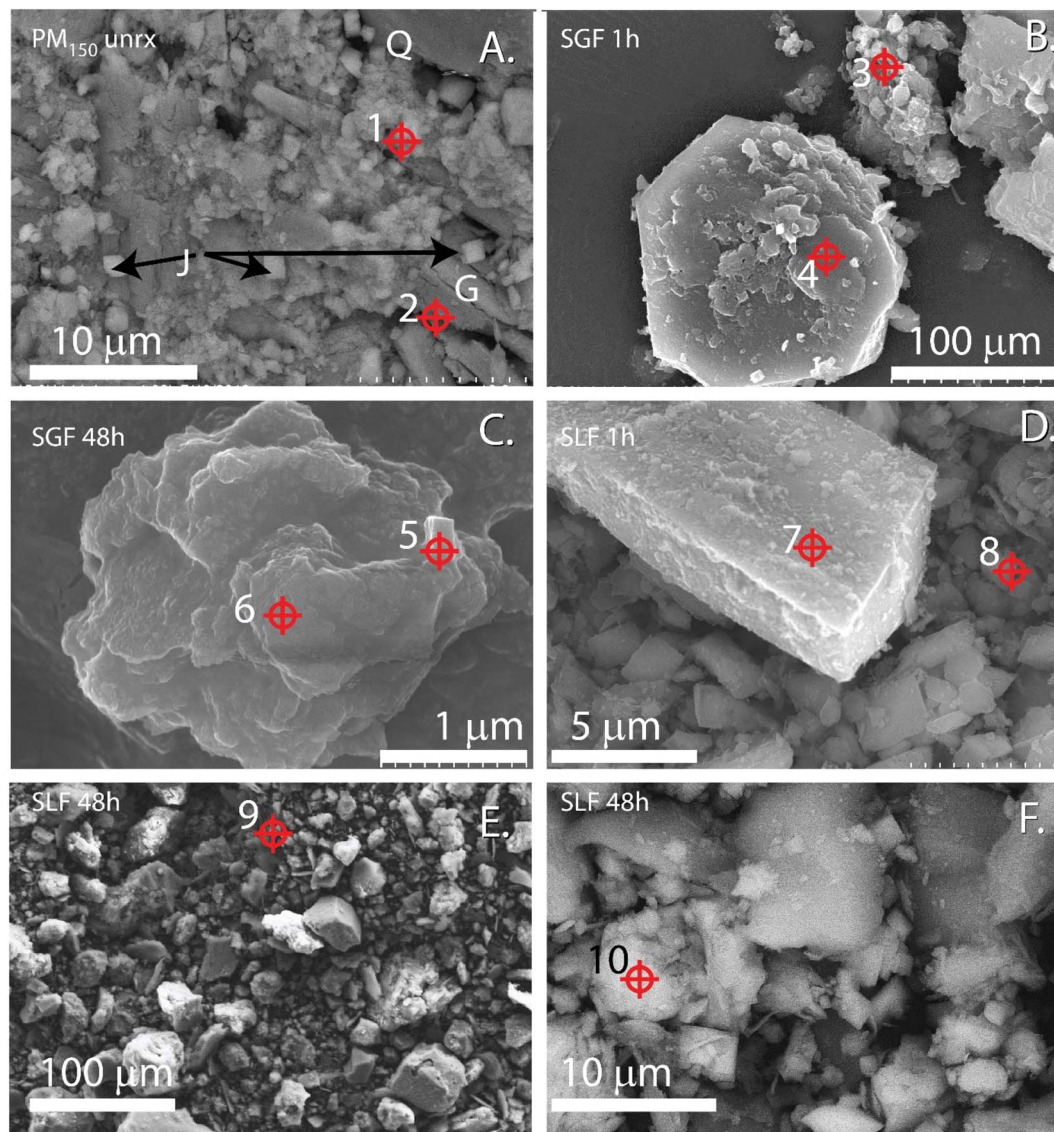


Fig. 3 Electron micrographs of <150  $\mu\text{m}$  effective spherical diameter particulate matter ( $\text{PM}_{150}$ ) isolated from IKMHSS tailings unreacted (A) and after *in vitro* gastric simulant bioassay for 1 hour (B), and 48 hours (C), and lung fluid for 1 h (D) and 48 h (E and F). Crosshairs indicate EDS spots,<sup>1–10</sup> spectra given in Fig. S5.† Panel (A) J = jarosite, Q = quartz, G = gypsum. Scale bar indicates magnification.

attributed to out of phase lead -bearing solids *e.g.* anglesite, consistent with previously reported tailings materials.<sup>20</sup> Reaction in SGF showed a decrease in the contribution of plumbogjarosite for  $\text{PM}_{10}$  and  $\text{PM}_{150}$  after 1 h. (Table S9†).

## 4 Discussion

Fugitive dusts in arid and semi-arid environments exhibit hygroscopic growth *in vivo* under conditions of high humidity that enhance alveolar deposition and inhibit lung clearance.<sup>76</sup> When particles finer than 10  $\mu\text{m}$  effective spherical diameter ( $\text{PM}_{10}$ ) pass the tracheobronchial system into the lungs, the relative solubilities of the remaining compounds will dictate what dissolves into extracellular fluids, while insoluble compounds can accumulate as particles in the lungs.<sup>77</sup> Larger particles (> $\text{PM}_{10}$ ) that do not pass into the respiratory regions of

the lung are trapped in mucus by cilia as part of the body's filtration defense. The muco-ciliary escalator mechanism clears larger particles from the airway toward the pharynx, where they are swallowed and become susceptible to gastric fluid dissolution.<sup>78</sup>

The maximum mass release of toxins in synthetic lung fluid at  $\leq 24$  h (Table 1), and the extrapolated risk to human health from inhalation exposures, from Iron King mine tailings particles for arsenic was  $\text{PM}_{10} > \text{PM}_{150} > \text{PM}_{\text{SC}} > \text{PM}_{\text{ES}}$  and for lead was  $\text{PM}_{\text{ES}} > \text{PM}_{10} > \text{PM}_{\text{SC}} > \text{PM}_{150}$ . The pharmacokinetic release to SLF from  $\text{PM}_{10}$  shows greatest risk at short duration exposures (<0.5 h) for lead and a delayed exposure risk for arsenic, with the greatest arsenic release measured in this study at the longest time step (Fig. 1). Combining sorbate elemental analysis with sorbent multi-element XAS, XRD, SEM and BET at several bio-relevant kinetic steps indicated arsenic-substituted jarosite



was the most bioaccessible species with respect to arsenic, and the toxic metalloid host phase posing the most risk in lung fluid. Precipitation of arsenic or lead bearing minerals in lung fluid can remove a contaminant from the dissolved phase, but it is unclear the extent to which such *in vivo* lung fluids achieve supersaturation and, if so, neoformed particles may also act as irritants and long-term point-sources of chronic exposure at extended residence times (*e.g.*, years). However, in lung tissue, the larger volume of fluid and continuous throughflux of new fluid interacting per unit mass of particle is likely to preclude, to some degree, the back reaction (*i.e.*, secondary precipitation formation).

#### 4.1 Arsenic speciation

In unreacted PM, arsenic was distributed between ferrihydrite and jarosite (Table 1 and Fig. S14†). The surface crust had the highest ferrihydrite associated arsenic, lowest jarosite associated arsenic, and the lowest 24 h BAC in lung fluid, 0.22% (and lowest maximum 0.37% at 48 h) and lowest in gastric fluid, 4.3% at <1 h. With increased content of short-range ordered minerals like ferrihydrite in metalliferous-mine tailings, arsenic shows a corresponding decrease in bioaccessibility for SGF and SLF, attributable to the high activity of high-affinity sites for surface complexation of arsenate.<sup>45</sup>

The reported EXAFS determined As–Fe interatomic distances in jarosites are: 3.25–3.26 Å for arsenic co-precipitated jarosite;<sup>79–81</sup> 3.35 Å for high arsenic jarosite loadings (17 mole%, possible contribution from scorodite);<sup>75,80,82</sup> and 3.22 Å for <sup>2</sup>C coordination of As-adsorbed jarosite.<sup>79</sup> Ferrihydrite with low to moderate arsenic loading (Fe : As > 50) displays As–Fe <sup>2</sup>C coordination at 3.28 Å for bidentate arsenate tetrahedra corner-sharing oxygens with two edge linked iron octahedra, as determined by EXAFS and DFT calculations.<sup>74</sup> These distances are shorter than the longer distances of 3.32 Å (<sup>2</sup>C) reported for colloidal amorphous ferric arsenate.<sup>74,83</sup> Arsenic-rich amorphous particles have been reported to occur as highly-aggregated clusters, 10 to 100 nm in size, composed of corner-linked iron octahedra in bidentate-binuclear complexation with arsenate.<sup>84</sup> While the arsenic in the PM was distributed between surface complexed arsenate on ferrihydrite and substituted for sulfate in jarosite, the long As–Fe distance of 3.30–3.34 Å observed in the unreacted PM<sub>10</sub> indicated arsenic was also associated with colloidal amorphous ferric arsenate in <sup>2</sup>C coordination,<sup>84</sup> whereas the relatively shorter As–Fe distances in PM<sub>150</sub> of 3.27–3.28 Å are in agreement with bidentate binuclear arsenic adsorbed on ferrihydrite reactive surface sites.<sup>74</sup> The PM<sub>10</sub> displayed relatively longer As–Fe bonds (>3.30 Å), consistent with high arsenic loading of ferric hydroxide octahedra. Whether high arsenic ferric solids were linked chains<sup>80</sup> or ferric arsenate clusters,<sup>83</sup> is beyond the scope of this study.<sup>74</sup>

**4.1.1 SGF.** Gastric IVBA carried out to 100 hours approaches an extreme exposure duration. Here, the 1 h and less duration exposures are emphasized as those that are physiologically relevant (Fig. 1). However, long kinetic steps were applied to explore positive and negative interferences in

toxin release from competing reactions and dissolution mechanisms *in vitro* that may not mimic *in vivo* physiology. In contrast to SLF, changes in the post SGF atomic coordination of arsenic showed slight shortening of the As–Fe distance from 3.32 Å (PM<sub>10</sub>) and 3.28 Å (PM<sub>150</sub>) to 3.28 Å and 3.27 Å, respectively, consistent with a reduction in the contribution of arsenate coprecipitated with ferrihydrite 3.28 Å<sup>74</sup> and an increase in the contribution of the shorter distance As–Fe jarosite component of 3.27 Å.<sup>75,80</sup> Arsenic liberated from surface complexation sites on ferrihydrite was the major contributor to the SGF BAC at initial time points, and at extended incubations *in vitro* arsenic may partition back to the solid phase in As-jarosite or Pb–As-jarosite, or as a surface complex on freshly precipitated β-FeOOH (PM<sub>150</sub> log  $\mathcal{Q}_{100h}$  = 2.43). Although neo-precipitate ferric hydroxides were observed to be supersaturated in the closed and extended SGF IVBA, they would not be expected to form in the gastrointestinal tract *in vivo* and would therefore not likely serve as a sink for dissolved arsenic in a physiological system. However, under the acid conditions in gastric fluid, arsenic released from soluble phases like calcium arsenate or arseniosiderite could adsorb to co-occurring, sparingly-soluble ferric hydroxides thus attenuating the measured bioaccessible fraction.<sup>85,86</sup> At the acid pH of SGF (1.5), precipitation of ferrihydrite was not thermodynamically favorable (PM<sub>150</sub> log  $\mathcal{Q}_{100h}$  SGF = −2.2), although there was a thermodynamic drive for akaganeite and plumbojarosite (PM<sub>150</sub> log  $\mathcal{Q}_{100h}$  = 11.3) precipitation. The pre-existence of jarosite surfaces could serve as a template to decrease the activation energy (degree of supersaturation) required for nucleation and crystal growth,<sup>87,88</sup> which could sequester both arsenate and lead during long term low pH SGF IVBA exposure. Because ingestion exposure to PM<sub>ES</sub> exhibited rapid solubilization on the time scale of minutes to hours, this suggests that total concentration was a reasonable indicator of BAC of arsenic and lead in efflorescent salts. However, the total arsenic concentration in the PM<sub>ES</sub> was 3 orders of magnitude lower than the other PM (Table 1). Whereas the 1 h SGF bioaccessible fraction of PM<sub>10</sub>, PM<sub>150</sub> and PM<sub>SC</sub> were all much lower (4.3–9.9%) compared to PM<sub>ES</sub> (44.9%), the total mass of arsenic released per gram of geodust was much lower in the PM<sub>ES</sub> ([As<sub>ES</sub>]<sub>T</sub> = 0.022 μmol g<sup>−1</sup>), and therefore PM<sub>ES</sub> poses a lower IVBA determined risk of arsenic exposure.

**4.1.2 SLF.** About 20% of the total SLF released arsenic occurred at 30 s and aqueous arsenic concentrations increased in all PM at each kinetic measurement step, releasing about 80% of the BAC arsenic after the initial pulse (Fig. 1). Investigation of SLF arsenic bioaccessibility of PM<sub>20</sub> from Au mining wastes from southern Australia, where the principal arsenic bearing phase was scorodite (FeAsO<sub>4</sub>·2H<sub>2</sub>O), showed a rapid release followed by a modest increase in arsenic dissolution to 24 h, but in that study total BAC arsenic was <0.1%.<sup>89</sup> Here, BAC values were more than an order of magnitude higher, indicating that the mixture of arsenate-rich jarosite and ferrihydrite pose a much greater inhalation risk than scorodite dominated tailings. Additionally, studies have shown soil arsenic BAC varies with speciation; BAC showed jarosite > ferrihydrite > scorodite > arsenopyrite.<sup>90</sup>



At extended exposures ( $>48$  h),  $\text{PM}_{150}$  in SLF resulted in a lengthening of the As-Fe distance consistent with  $^2\text{C}$  coordination of amorphous ferric arsenate. *In silico* calculations indicate atomic coordination similar to the substitution of tetrahedral sulfate by arsenate in the  $\text{T-Oh}_3$  jarosite structure in tridentate penta- and hexa-nuclear complexes.<sup>91</sup> The substitution of an arsenate tetrahedron ( $\text{As-O}_4$  1.69 Å) into a sulfate tetrahedron site ( $\text{S-O}_4$  1.54 Å) in the jarosite crystal system creates torsional and steric strain in the iron octahedral kagome lattice, decreasing the activation energy for breaking the As-O-Fe bonds, which may make arsenic from highly substituted jarosite more susceptible to release *in vivo* than from bidentate binuclear surface complexes at ferric (hydr)oxide surface sites.

Here, Fe XANES pre-edge combined with iron and arsenic EXAFS indicated that jarosite was the species most susceptible to SLF extraction. The lengthening of the 3.32 Å As-Fe bond in post SLF solids at 48 h further indicates a relative loss of the shorter As-Fe distance from As-loaded jarosite and/or accretion of arsenic complexed with ferric hydroxides. The affinity of arsenate for adsorption to ferric (hydr)oxide surfaces generates a surface complex that is relatively resistant to SLF IVBA. The high activity of phosphate in the SLF and sulfate from the dissolution of soluble salts (*e.g.*, gypsum) contributed competing ions that inhibited arsenic re-sorption, evidenced by S XANES showing increased loading of sulfate on ferrihydrite as the SLF reaction progressed, from 13% to 22% (Table S5†). The release of arsenic by SLF IVBA was parabolic, but not asymptotic, over the duration examined for lung fluid extractions in the  $\text{PM}_{10}$  and  $\text{PM}_{150}$  size range (Fig. 1), indicating more than one kinetic reaction rate, with an initial release dominated by dissolution of strained arsenic-substituted jarosite and secondary competing processes of solid phase repartitioning or precipitation under saturated conditions. It should be considered that the observed parabolic release may be due to accumulation of solutes in the closed IVBA batch system that would not develop in an open biosystem with continuously exchanging fresh biofluid. The release of arsenic from PM in SLF was not stoichiometric with iron. The ratio of Fe : As in unreacted  $\text{PM}_{10}$  and  $\text{PM}_{150}$  solids was 56.7 and 64.7, respectively. At the first SLF extraction step, the  $\text{PM}_{10}$  release was dominated by iron with respect to arsenic, 30 s Fe : As = 124, while  $\text{PM}_{150}$  showed diminished iron release relative to arsenic, Fe : As = 18.2 (Fig. 4). Both particle types showed an enrichment of arsenic release relative to iron release with increased reaction times, and the release of arsenic in both  $\text{PM}_{10}$  and  $\text{PM}_{150}$  to SLF approached unity ( $\text{PM}_{10}$  Fe : As<sub>100h</sub> = 2.13,  $\text{PM}_{150}$  Fe : As<sub>100h</sub> = 2.27) with extended incubations (Fig. 4). The Fe : As ratio in the solubilized (bioaccessible) pool decreased at each interval in  $\text{PM}_{10}$  and  $\text{PM}_{150}$  and fit to non-linear natural log of time curve ( $\text{PM}_{10}$   $R^2$  = 0.946,  $\text{PM}_{150}$   $R^2$  = 0.829) (Fig. 4). Where the molar ratio of Fe : As released to SLF was lower than the bulk ratio, *i.e.* after 15 minutes, for  $\text{PM}_{10}$  and over the duration of the IVBA experiment for  $\text{PM}_{150}$ , indicated that (i) dissolution of an arsenic enriched species, (ii) preferential arsenic release, or (iii) reprecipitation of iron were responsible for the solubilized species; and where the arsenic release exceeded iron it was seen as evidence that iron was precipitating as ferric (hydr)oxide without concurrent arsenic sorption.

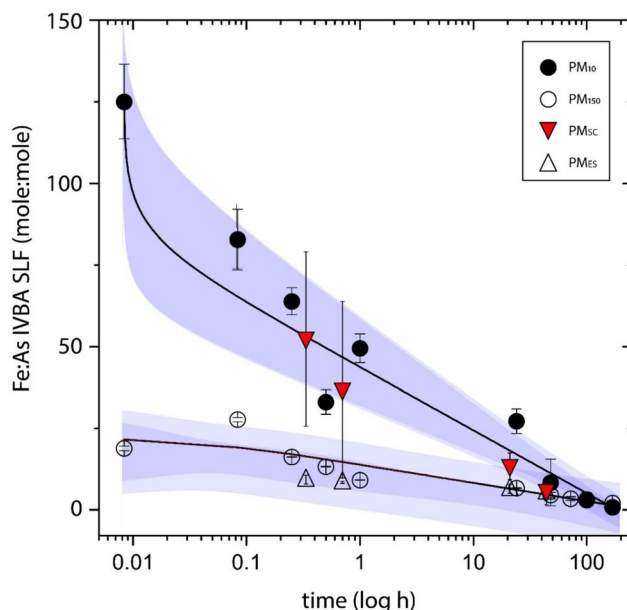


Fig. 4 Mole ratio of released iron and arsenic in SLF. Solid circles are  $\text{PM}_{10}$ , open circles are  $\text{PM}_{150}$ , closed downward red triangles are  $\text{PM}_{\text{SC}}$ , open triangles  $\text{PM}_{\text{ES}}$ , lines represent the best fit ( $y = a - b \cdot \ln(x + c)$ ) to the data,  $\text{PM}_{10}$   $R^2$  = 0.946,  $\text{PM}_{150}$   $R^2$  = 0.829, fits to SC and ES not shown. Dark shaded bands represent the 95% confidence interval, light shading represents the 95% prediction band.

## 4.2 Lead speciation

**4.2.1 SGF.** The release of lead from  $\text{PM}_{10}$  to SGF increased with time to 100 h, and BAc reached a maximum of 18.2% (Fig. 1). The iron gastric IVBA followed a release curve very similar to lead (and arsenic), whereby it increased at each step to 100 h. The initial, a rapid, release of lead from the tailings was attributed to lead sequestered in acid soluble anglesite, which was released early in the reaction. Equilibrium modeling of anglesite solubility at the low pH and geochemical conditions of the SGF show a drive toward instability ( $\log Q_{100\text{h-SGF}} = -17.1$ ), which would release lead to gastric fluid.<sup>92</sup> Whereas equilibrium modeling showed the potential for precipitation of akaganeite, at the low pH of SGF  $\text{Pb}^{2+}$  sorption would be minimized and would not serve as a sink for lead. Further, because of the low phosphate activity and low pH, precipitation of pyromorphite as a sink for lead was not expected in SGF.<sup>92,93</sup> Lead released to the aqueous phase at extended reaction times could be limited by the stability of plumbojarosite ( $\text{PM}_{10}$ - $\log Q_{100\text{h-SGF}} = 10.9$ ), for which the 100 h solution was supersaturated and could promote sequestration of lead (and iron) into the solid phase. The low pH SGF solution was not supersaturated with respect to any other lead minerals. Therefore, it is not expected that any neo-phase provided a sink for  $\text{Pb}^{2+}$  at extended exposures.

**4.2.2 SLF.** The maximum lead released in  $\text{PM}_{10}$  SLF occurred very quickly (less than 30 s), and the release at very short intervals suggests dissolution of a sulfate salt like anglesite ( $\text{PbSO}_4$ ). The removal of lead from the aqueous phase after the initial pulse to SLF was attributed to supersaturation with respect to lead bearing apatite group minerals, *e.g.*







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