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The role of spin states in the catalytic mechanism of the intra- and extradiol cleavage of catechols by O₂[†]

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Iron-dependent enzymes and biomimetic iron complexes can catalyze the ring cleavage of very inert, aromatic compounds. The mechanisms of these transformations and the factors that lead either to extradiol cleavage or intradiol cleavage have not been fully understood. By using density functional theory we have elucidated the mechanism of the catalytic cycle for two biomimetic complexes, and explained the difference in the experimentally obtained products.

Introduction

The microbial aerobic degradation of aromatic pollutants, which are widespread contaminants in soils and groundwaters, obviously represents an intense research area in the scientific community.^{1,2} A key step in the biodegradation of aromatic compounds is the oxidative cleavage of catechol derivatives and is facilitated by a class of enzymes known as catechol dioxygenases.^{3,4} Two different groups of enzymes play an essential role in the ring cleavage of catechol derivatives: extradiol- and intradiol-cleaving dioxygenases.⁵ These two enzyme groups have different active sites, and thus catalyse different metabolic transformations.⁶ In the case of extradiol enzymes, one of the oxygens from O₂ is incorporated next to one of the hydroxyl groups (see Fig. 1). On the other hand, in the intradiol dioxygenases, the insertion of the oxygen takes place between the two hydroxyl groups of catechol.

The most remarkable feature of these two enzymes is the use of different oxidation states of iron in them. Extradiol-cleaving dioxygenases use Fe^{II} in their active site, coordinated by a 2-His-1-carboxylate facial triad in an octahedral environment. The vacant sites are occupied by solvent molecules that can be easily replaced by the substrate and oxygen in the catalytic process, see Fig. 1. In the subsequent step the formation

of an alkyl peroxide is followed by a homolytic cleavage of the O–O bond and the insertion of an oxygen atom into the catechol ring occurs. On the other hand, the active site of intradiol dioxygenases consists of Fe^{III} coordinated by two histidines and two tyrosines, and one hydroxide anion.

One of the proposed mechanistic pathways suggested that for an intradiol enzyme, first, Fe^{III} has to oxidize the aromatic ring to a catechol radical, thereby enabling the coordination of oxygen and the subsequent formation of an alkyl peroxide. It is believed that the incorporation of an oxygen atom into the catechol ring proceeds *via* a concerted process, the so-called Criegee rearrangement.^{6,9,10}

Despite many studies on intra/extradiol enzymes, many questions still remain concerning the details of the reaction mechanisms of these enzymes along with the factors that determine the intra- *versus* extradiol specificity.⁶ This is in particular true because of the finding that biomimetic complexes based on Fe^{III} showed both intra- and extradiol activities.¹¹ Previous experimental investigations on iron complexes with rigid L-N₂H₂ (2,11-diaza[3.3](2,6)pyridinophane) and L-N₂Me₂ (*N,N'*-dimethyl-2,11-diaza[3.3](2,6)pyridinophane) ligands (see Fig. 2) have shown that the complexation with L-N₂Me₂ mainly follows the intradiol pathway (as expected for a Fe^{III} complex),¹² while for the complex with L-N₂H₂ both pathways are being followed.¹¹ Previous experimental and theoretical DFT studies indicated that the formation of an alkyl peroxide is the rate determining step in both cases.¹⁰ The selection of one or the other pathway might involve the inclusion of different spin states during the reaction, and therefore, we have studied in detail the influence of the spin-state on the energetics and the mechanism of these reactions.

The relatively small size of biomimetic complexes compared to the enzymes enables both a facile and well-controlled adaptation of the ligand structure, and an in-depth inspection by

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Fig. 1 Proposed catalytic cycle in the active site for (a) extradiol and (b) intradiol cleaving catechol dioxygenases.^{7,8}



Fig. 2 Schematic representation of ligands used (left) to model the reactivity of extradiol and intradiol catechol dioxygenases, L-N₂H₂ (2,11-diaza[3.3](2,6)pyridinophane) and L-N₂Me₂ (N,N'-dimethyl-2,11-diaza[3.3](2,6)pyridinophane), and the structure of the [Fe^{III}(L-N₂H₂)Cl₃] complex (right).

means of quantum mechanical (QM) modelling; this provides meaningful guidelines for the investigation of the functioning of complicated enzymatic systems.^{6,13–15} In theory, there is a

wide variety of QM levels that can be applied to biomimetic complexes, ranging from semi-empirical methods to high level *ab initio* (post-Hartree Fock) approaches,^{16,17} and including a plethora of Density Functional Approximations (DFAs).^{18,19} The DFAs offer a fast and easy way to calculate the properties of medium-to-large sized molecules, considering their good compromise between the accuracy and the computational efficiency. However, even though DFA methods are in general very accurate for structures and energies, the reliable prediction of spin state splitting in transition metal compounds becomes a difficult challenge.²⁰ There are a large number of studies^{21–29} devoted to the validation of Density Functional Approximation (DFA) for accurate description of relative spin state energetics, although the prediction of the ground state in complicated systems requires special caution.^{20,23,30–34} LDA and early GGA DFAs show intrinsic preferences for the low spin (LS) state, while hybrid DFAs systematically favour high spin (HS) states, which is a consequence³⁵ of the inclusion of HF exchange that does not address the correlation between electrons with different spins in a proper way.³⁶ There has not yet been a generally accepted level of theory for the diversity of complicated transition metal systems, although there is a clear preference for DFAs that are spin-state consistent in their design.¹⁹ However, so far, almost all studies on enzyme active sites and their small molecule mimics have used hybrid DFAs.^{9,10,37–42} As mentioned before, because of the inclusion of the HF exchange in the hybrid DFAs, a tendency to overestimate the stability of high spin states is present in them.^{23,32} Recently, new spin-state consistent DFAs (SSB-D,⁴³ S12g⁴⁴ and S12h⁴⁴) have been designed, showing good performance for spin state energetics and reaction barriers.¹⁹ Furthermore, Perdew has recently reported a non-empirical meta-GGA 'Made Very Simple' (MGGA-MVS),⁴⁵ which could also perform well.⁴⁶

Computational details

All DFT calculations were performed with the Amsterdam Density Functional (ADF) program suite.^{47,48} MOs were expanded in an uncontracted set of Slater type orbitals (STOs) of triple- ζ quality containing diffuse functions (TZ2P)⁴⁹ and two sets of polarization functions. Core electrons (1s for the 2nd period, 1s2s2p for 3rd–4th periods) were not treated explicitly during the geometry optimization (frozen core approximation), as it was shown to have a negligible effect on the obtained geometries.⁵⁰ An auxiliary set of s, p, d, f, and g STOs was used to fit the molecular density and represent the Coulomb and exchange potentials accurately for each SCF cycle. Electronic energies were used without any thermochemistry correction throughout the paper, with Gibbs free energies available in the ESI.†

The geometries were optimized using the QUILD program⁵¹ (as implemented in the ADF2016 suite) using adapted delocalized coordinates⁵² until the maximum gradient component was less than 10^{-4} a.u. The energies and gradients were calculated by the S12g⁴⁴ DFA, using a Becke grid^{53,54} with a numeri-



cal accuracy of very good quality. The COSMO^{55–57} dielectric continuum model was used for implicit treatment of the environment (with methanol as a solvent).^{58,59} Scalar relativistic corrections have been included self-consistently by using the zeroth-order regular approximation (ZORA).^{60–62} The nature of the stationary points is confirmed by calculating analytical Hessians. The subsequent single point calculations that use all-electron basis sets have been performed on all optimized geometries with the S12g, SSB-D,⁴³ M06-L,^{13,63} MVS⁴⁵ and S12h⁴⁴ levels of theory (see the ESI†).

Results and discussion

General mechanism

The proposed mechanism for both the extradiol and intradiol cleavages of an iron catechol complex is shown in Fig. 3 (with the N₂H₂ ligand, it is assumed that the same pathways are being followed using the N₂Me₂ ligand). This mechanism represents stepwise progression *via* multiple local minima. In the examined biomimetic complexes, iron is coordinated by four nitrogen atoms from the chelate ligand, having two adjacent sides accessible for the coordination of the catechol which donates two oxygen atoms to the iron in the resting state (**0**), see Fig. 3.

After binding of O₂, in the first step a weakly-bound van der Waals complex **1** is obtained. Subsequent electron transfer from the catechol, *via* iron(III) to oxygen, and coordination of the latter, occur *via* a concerted process,⁹ forming intermediate **2** (see Fig. 3). In the next step, the coordinated superoxide

attacks the catechol, which results in a bridging alkyl peroxide **3**. The O₂ bond is cleaved homolytically, leading to intermediate **4**, which immediately decays in one of the two possible routes: intradiol or extradiol. This intermediate **4** is therefore the branching point where the selectivity for one of the two routes is decided.

The intradiol pathway leads to the open chain radical intermediate **5i**, *via* breaking of the C–C bond and subsequently, the insertion of the oxygen to leads to the cyclization product **6i** (see Fig. 3). The extradiol pathway involves the formation of cyclic epoxide **5e**, and then breaking of the C–C bond, and the insertion of the oxygen leads to structure **6e**. It should be noted that **6i** and **6e** dissociate forming two isomers, a muconic anhydride and a seven-membered ring lactone, respectively.

Iron(III) complexes have three possible spin-states: high-spin (HS) with five unpaired electrons on the metal; intermediate-spin (IS) with three unpaired electrons on the metal; and low-spin (LS) with one unpaired electron on the metal. In order to fully understand the proposed mechanism, the spin-state splittings of all intermediates and transition states involved in the reaction mechanisms shown in Fig. 3 were investigated. The spin-state splittings of **0**, **6i** and **6e** at S12g/TZ2P are given in Table 1, while those with SSB-D, S12h, M06-L and MVS⁴⁵ can be found in Tables S1–S8.†

There are a number of experimentally available crystal structures of the complexes that are related to the investigated catalytic cycle.^{11,12,64,65} They are either precursors of complex **0** with chloride ions instead of catechol with L-N₂H₂ and L-N₂Me₂ ligands, or **0** itself (only characterized structurally with L-N₂Me₂). The calculated bond distances for all com-

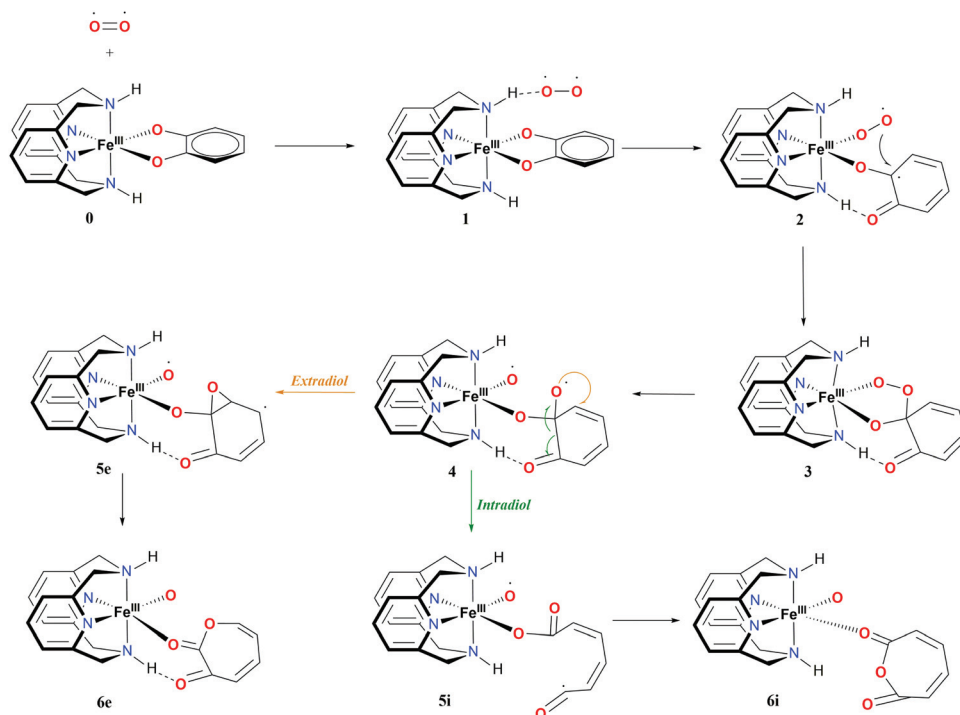


Fig. 3 Proposed mechanism and nomenclature for intra- and extradiol catechol cleavages catalyzed by the L-N₂H₂ iron(III) complex.



Table 1 Electronic energies (kcal mol⁻¹) for the resting state of **0** and final products **6i** and **6e**, relative to reactants, at S12g/TZ2P

Ligand molecule	L-N ₂ H ₂			L-N ₂ Me ₂		
	0	6i	6e	0	6i	6e
LS	0	-28.3	-15.0	0	-25.8	-12.6
IS	5.0	-28.7	-15.5	4.3	-26.5	-13.3
HS	4.5	-30.0	-16.8	4.1	-27.4	-21.7

plexes show reasonable agreement with the experimentally determined bond lengths (see Table S9†). Experimental studies have concluded (based on the bond lengths of similar systems) that all of these complexes are in the HS ground state.^{9,10} The calculated electronic energies show that the chloride precursors are in the HS ground state, which corroborates the experimental conclusions. However, for the complex with catechol, the LS state is obtained as the ground state. Similar results are obtained previously at the B3LYP* level of theory.¹⁰ This indicates that the spin ground-state for this complex is not determined by the electronic energies, but instead a significant effect is to be expected from enthalpy and entropy corrections that systematically favour the HS state. Irrespective of the coordinated ligand (L-N₂H₂ or L-N₂Me₂), the HS intradiol product is thermodynamically favoured, see Table 1.

Reaction path with the L-N₂H₂ ligand

The first part of the energy landscape for the catalytic cycle of the complex with the L-N₂H₂ ligand (before branching into intradiol and extradiol pathways) is represented in Fig. 4. The splitting of the pathways after branching point 4 is shown in Fig. 5. For the sake of clarity, only the S12g results are given. Some of the specific details are different using SSB-D, M06-L,

S12h and MVS, but the overall conclusions regarding the spin state that governs the mechanism and product distribution are the same (see Tables S1–S8†).

Following the schematic representation shown in Fig. 1, the complete catalytic dioxygenation reaction for all the available spin states is calculated, starting from a [Fe(LN₄H₂)(Cat)]⁺ complex **0**, Fig. 4. As mentioned earlier, the initial structure **0** is suggested to be in the HS state. Close lying LS and the HS states are obtained using S12g, with the LS slightly lower in energy. The reaction proceeds with the initial formation of the van der Waals complex **1** that has a negligible effect on the relative spin state energies. Since complex **1** is coordinatively saturated, the subsequent coordination of molecular oxygen needs to be accompanied by decooordination of one part of catechol, in an endothermic process, leading to structure **2**. The oxygen that moved away is now able to form an H-bond with the ligand, thereby fixating its position during the reaction. This step is a complex set of events that include catechol oxidation coupled with the iron mediated electron transfer to the O₂ molecule, accompanied by the large structural changes. As a result, the spin state ordering in intermediate **2** has changed, with the sextet state now being the lowest in energy. This result is in accordance with experimental results for the enzymatic reaction⁶⁶ and similar iron superoxo intermediates.⁶⁷

In the next step, after the catechol is attacked by the superoxo moiety, an intermediate with a five-membered ring (**3**) is formed. This corroborates the scattered experimental results for the enzymatic process.⁶⁸ During the formation of the peroxo intermediate, the quartet state becomes the lowest in energy, and remains so until the final point where the muconic anhydride and seven-membered ring lactone dissociate from the iron centre. After the formation of intermediate **3**, the O–O bond weakens, and subsequently it breaks, leading the reaction profile to the branching point region in

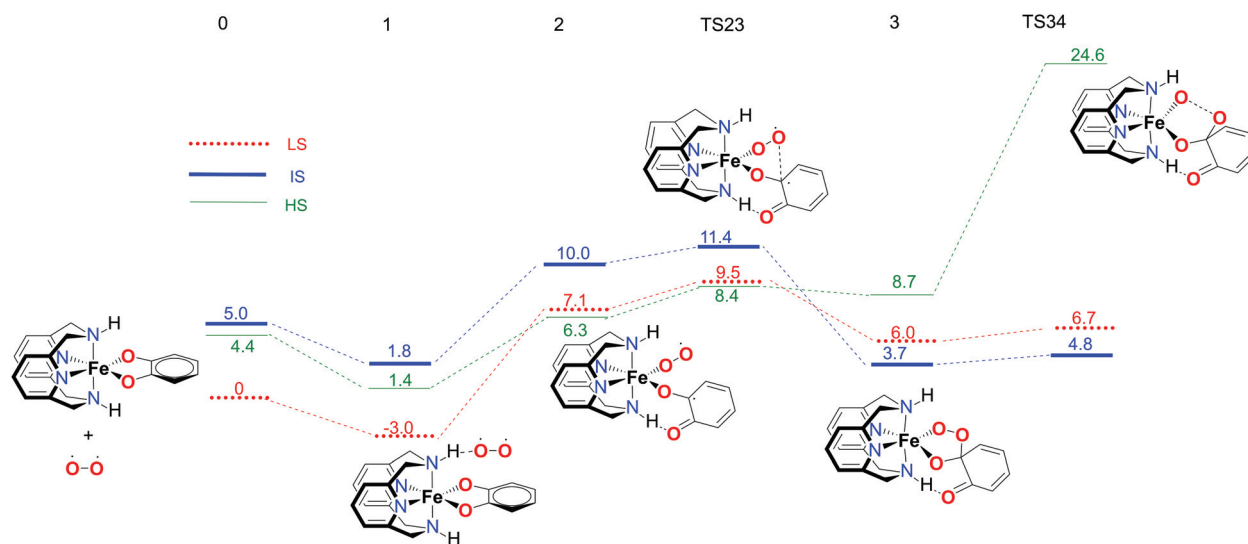
**Fig. 4** Calculated reaction profile for the first part of the catechol dioxygenation mechanism, starting from the [Fe(LN₄H₂)(Cat)]⁺ complex **0**. Electronic energies (kcal mol⁻¹), obtained using the S12g/TZ2P level of theory, are given relative to reactants.



Fig. 5 Calculated reaction profile for the second part of the catalytic mechanism with the LN_4H_2 ligand, branching to the extradiol (left) and intradiol (right) pathways. Electronic energies (kcal mol^{-1}), obtained using the S12g/TZ2P level of theory, are given relative to reactants.

which it can continue either along the intradiol or the extradiol pathway. The reaction profile that follows the cleavage of the O–O bond in intermediate 3 is depicted in Fig. 5. The intradiol pathway is presented on the right, while the extradiol pathway is shown in the same figure on the left. These two mechanisms are characterized by distinct structural transformations, reaction courses and final products.

Our calculations indicate that the intradiol mechanism starts with the breaking of the C–C bond, which is then followed by a rearrangement and cyclization, creating the final product, a seven-membered ring lactone. The extradiol pathway is initiated by the formation of the cyclic epoxide, followed by the breaking of a different C–C bond to give the extradiol product, muconic anhydride (see Fig. 5). However, in most of the previous studies concerning the mechanism of intradiol catechol dioxygenase⁶ and its biomimetics,^{9,10} a concerted mechanism was found for the formation of seven-membered lactone, whereas in this study the C–C bond breaking occurs first.

The ratio of the products is determined by the barriers for the two pathways, *i.e.* the relative energies of two transition states $\text{TS}_{45\text{i}}$ and $\text{TS}_{45\text{e}}$. In the IS state, the energy difference between $\text{TS}_{45\text{i}}$ and $\text{TS}_{45\text{e}}$ is $1.9 \text{ kcal mol}^{-1}$, favouring the intradiol pathway. However, much smaller barriers are found in the sextet state, with a difference between $\text{TS}_{45\text{i}}$ and $\text{TS}_{45\text{e}}$ of only $0.4 \text{ kcal mol}^{-1}$. Since a spin-state switch from IS to HS itself also involves a small barrier (through a so-called minimum-energy crossing point, MECP),^{69,70} no attempt was made here

to localize this MECP because of the small barriers involved. It indicates that both products should in principle be formed, which is in excellent agreement with the experiment.¹¹ The inspection of the reaction landscape shows that the quartet and sextet states are almost equi-energetic in many cases, see Fig. 5, and hence two-state reactivity^{71–73} seems to play a role here to govern the reaction until the dissociation of the seven-membered ring products and the formation of a five-coordinate iron-oxo complex occur. This is in good agreement with previous studies⁹ which concluded that iron-oxo complexes tend to be in close lying spin states, with the dominant HS state in a penta-coordinated environment, and IS when hexa-coordinated.

Reaction path with the $\text{L-N}_2\text{Me}_2$ ligand

In order to fully understand the behaviour and intrinsic properties of the spin state governed reactions, calculations have also been carried out for the Fe^{III} complex with the $\text{L-N}_2\text{Me}_2$ ligand. The observed reaction profile with S12g is shown in Fig. 6 and 7 (SSB-D, M06-L, S12h and MVS results are given in Tables S1–S8†).

The overall reaction pathway is very similar to the catalytic cycle of the complex with the $\text{L-N}_2\text{H}_2$ ligand. The sextet state is the ground state for the superoxo intermediate 2, as well as for the final product, a five-coordinate iron-oxo complex. The quartet state governs the reaction mechanism and product distribution due to the fact that the barrier for the O–O bond dissociation TS_{34} is very high in energy on a sextet PES.





Fig. 6 Calculated reaction profile for the first part of the catechol dioxygenation mechanism, starting from the $[\text{Fe}(\text{LN}_4\text{Me}_2)(\text{Cat})]^+$ complex 0. Electronic energies (kcal mol⁻¹), obtained using the S12g/TZ2P level of theory, are given relative to reactants.

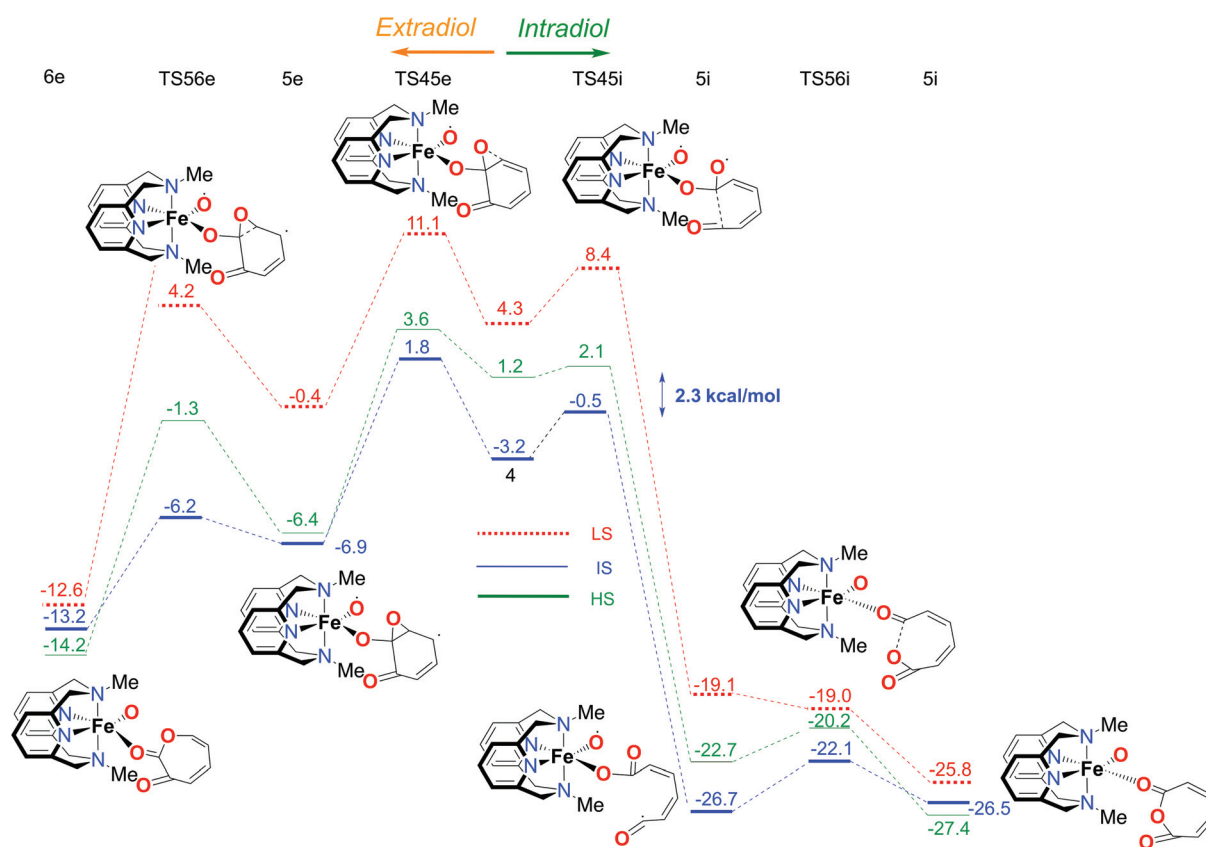


Fig. 7 Calculated reaction profile for the second part of the catalytic mechanism with the LN_4Me_2 ligand, branching to the extradiol (left) and intradiol (right) pathways. Electronic energies (kcal mol⁻¹), obtained using the S12g/TZ2P level of theory, are given relative to reactants.



However, there are two notable differences. First, the step for the coordination of O₂, 1 → 2, is more endothermic for the L-N₂Me₂ ligand. The obvious reason for this is easily found to be the absence of a H-bond donor here (see Fig. 7), to which the decoordinating catechol oxygen can bind (unlike the situation for the L-N₂H₂ ligand, see Fig. 4).

Secondly and most importantly, the differences in the reaction barriers (TS_{45i} vs. TS_{45e}) after the branching point 4 were twice as large (2.8 kcal mol⁻¹) for the L-N₂Me₂ ligand compared to those for the L-N₂H₂ ligand (1.4 kcal mol⁻¹). This corroborates the experimental findings that the iron(III)-complex with the L-N₂Me₂ ligand selectively cleaves the catechol in an intradiol pathway.¹²

The difference in the product selectivity with Fe^{III} complexes from L-N₂H₂ and L-N₂Me₂ ligands

The overall selectivity of the catalytic cycle is determined by the differences in the energies of the transition states for the two reaction pathways, TS_{45i} for the intradiol and TS_{45e} for the extradiol routes. The difference in the intradiol/extradiol barriers can be explained in terms of the relative orientation of the substrate toward the bridging peroxide.¹⁰ The extradiol pathway has a higher barrier, probably due to the necessity of reorientation of the O_{2p} orbital originating from the O–O cleavage.¹⁰ The reason for the difference in the product selectivity of the two complexes, *i.e.* barrier heights at the branching point, can be found in the inspection of H-bonding towards the catechol substrate with the L-N₂H₂ ligand. During the intradiol pathway, the H-bond with the L-N₂H₂ ligand is lost in the 4 → 5i step, as a consequence of the breaking of the C–C bond. This does not occur during the extradiol reaction path, explaining why the intradiol pathway is higher in energy and closer to the extradiol alternative with the L-N₂H₂ ligand. Similarly, the larger reaction rate for the L-N₂H₂ ligand compared to L-N₂Me₂ results from the smaller endothermicity for binding of O₂ with L-N₂H₂ because of the capability to form H-bonds (see Fig. 4). In addition, it seems that the methyl groups on the ligand tend to further stabilize the intermediate spin if we compare intermediate 4 with the two ligands. Although on an absolute scale the barrier of the extradiol pathway remains almost identical (*ca.* 5 kcal mol⁻¹) the barrier for the intradiol path is *ca.* 1 kcal mol⁻¹ lower for the L-N₂Me₂ ligand. The origin of this change in the barrier height can be found in L-N₂Me₂ 4 where the IS state shows an increased bond distance between iron and the catechol oxygen. At the same time a higher Mulliken spin-density charge is observed on the oxygen and carbon atoms that are involved in the cleavage. The Fe core also seems to have a higher spin density in the L-N₂Me₂ ligand than the L-N₂H₂ one, and therefore, the geometry and the spin densities of the L-N₂Me₂ complex in the IS state are more similar to a HS electronic configuration, while in the case of 4 with the L-N₂H₂ ligand the geometry and electronic configuration are closer to a low spin electronic configuration.

Conclusions

Density functional theory calculations have been performed for the reaction mechanism of the catechol cleaving dioxygenase biomimetic iron complexes with L-N₂H₂ and L-N₂Me₂ ligands. The complete details of the catalytic cycle, with all accessible spin states and two possible pathways, intradiol and extradiol, were determined. Our calculations show that the formation of both intradiol and extradiol products is substantially exothermic, but the intradiol product formation is thermodynamically more favourable. For the L-N₂H₂ ligand, H-bonding plays an important role, both in stabilizing the binding of O₂, and in making the reaction barriers after the branching point 4 more equal (and hence reducing the preference for an intradiol pathway). Several spin states are involved throughout the reaction mechanism, where again for the L-N₂H₂ ligand in the critical steps a two-state reactivity seems to be present. On the other hand, for the L-N₂Me₂ ligand only the quartet state is shown to be involved in an energetically plausible mechanism, completely determining the product distribution. Finally, the difference in the intradiol/extradiol barriers is explained with the change in the H-bonding of the catechol substrate with the L-N₂H₂ ligand during the intradiol pathway. The obtained results and rationalizations can assist in the design of more selective catechol dioxygenation mimics.

Conflicts of interest

There are no conflicts of interest to declare.

Acknowledgements

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References

- 1 M. Costas, M. P. Mehn, M. P. Jensen and L. Que, *Chem. Rev.*, 2004, **104**, 939–986.
- 2 C. K. Brown, M. W. Vetting, C. A. Earhart and D. H. Ohlendorf, *Annu. Rev. Microbiol.*, 2004, **58**, 555–585.
- 3 S. J. Lippard and J. M. Berg, *Principles of bioinorganic chemistry*, University Science Books, 1994.



- 4 J. J. R. Fraústo da Silva and R. J. P. Williams, *The Biological Chemistry of the Elements*, Oxford University Press, 2001.
- 5 K. Ray, F. F. Pfaff, B. Wang and W. Nam, *J. Am. Chem. Soc.*, 2014, **136**, 13942–13958.
- 6 A. Company, J. Lloret-Fillol and M. Costas, in *Comprehensive Inorganic Chemistry II*, ed. J. R. Poeppelmeier, Elsevier, Amsterdam, 2nd edn, 2013, pp. 487–564. DOI: 10.1016/B978-0-08-097774-4.00323-5.
- 7 E. G. Kovaleva, M. B. Neibergall, S. Chakrabarty and J. D. Lipscomb, *Acc. Chem. Res.*, 2007, **40**, 475–483.
- 8 T. Borowski and P. E. M. Siegbahn, *J. Am. Chem. Soc.*, 2006, **128**, 12941–12953.
- 9 R. Jastrzebski, M. G. Quesne, B. M. Weckhuysen, S. P. de Visser and P. C. A. Bruijninx, *Chem. – Eur. J.*, 2014, **20**, 15686–15691.
- 10 V. Georgiev, H. Noack, T. Borowski, M. R. A. Blomberg and P. E. M. Siegbahn, *J. Phys. Chem. B*, 2010, **114**, 5878–5885.
- 11 N. Raffard, R. Carina, A. J. Simaan, J. Sainton, E. Rivière, L. Tchertanov, S. Bourcier, G. Bouchoux, M. Delroisse, F. Banse and J.-J. Girerd, *Eur. J. Inorg. Chem.*, 2001, **2001**, 2249–2254.
- 12 W. O. Koch and H.-J. Krüger, *Angew. Chem., Int. Ed. Engl.*, 1996, **34**, 2671–2674.
- 13 Y. Zhao and D. G. Truhlar, *Theor. Chem. Acc.*, 2008, **120**, 215–241.
- 14 M. K. Panda, A. John, M. M. Shaikh and P. Ghosh, *Inorg. Chem.*, 2008, **47**, 11847–11856.
- 15 S. Chatterjee, D. Sheet and T. K. Paine, *Chem. Commun.*, 2013, **49**, 10251–10253.
- 16 C. Daul, M. Zlatar, M. Gruden-Pavlovic and M. Swart, in *Spin States in Biochemistry and Inorganic Chemistry: Influence on Structure and Reactivity*, ed. M. Swart and M. Costas, Wiley, Oxford, 2015, pp. 7–34. DOI: 10.1002/9781118898277.ch2.
- 17 C. Sousa and C. de Graaf, in *Spin States in Biochemistry and Inorganic Chemistry: Influence on Structure and Reactivity*, ed. M. Swart and M. Costas, Wiley, Oxford, 2015, ch. 3, pp. 35–57. DOI: 10.1002/9781118898277.ch3.
- 18 A. D. Becke, *J. Chem. Phys.*, 2014, **140**, 18A301.
- 19 M. Swart and M. Gruden, *Acc. Chem. Res.*, 2016, **49**, 2690–2697.
- 20 M. Swart, *Int. J. Quantum Chem.*, 2013, **113**, 2–7.
- 21 H. Paulsen, L. Duelund, H. Winkler, H. Toftlund and A. X. Trautwein, *Inorg. Chem.*, 2001, **40**, 2201–2203.
- 22 M. Reiher, *Inorg. Chem.*, 2002, **41**, 6928–6935.
- 23 M. Swart, A. R. Groenhof, A. W. Ehlers and K. Lammertsma, *J. Phys. Chem. A*, 2004, **108**, 5479–5483.
- 24 J. Conradie and A. Ghosh, *J. Phys. Chem. B*, 2007, **111**, 12621–12624.
- 25 K. P. Jensen, *Inorg. Chem.*, 2008, **47**, 10357–10365.
- 26 L. M. Lawson Daku, A. Vargas, A. Hauser, A. Fouqueau and M. E. Casida, *ChemPhysChem*, 2005, **6**, 1393–1410.
- 27 B. J. Houghton and R. J. Deeth, *Eur. J. Inorg. Chem.*, 2014, **2014**, 4573–4580.
- 28 W. C. Isley III, S. Zarra, R. K. Carlson, R. A. Bilbeisi, T. K. Ronson, J. R. Nitschke, L. Gagliardi and C. J. Cramer, *Phys. Chem. Chem. Phys.*, 2014, **16**, 10620–10628.
- 29 P. Verma, Z. Varga, J. E. M. N. Klein, C. J. Cramer, L. Que Jr. and D. G. Truhlar, *Phys. Chem. Chem. Phys.*, 2017, **19**, 13049–13069.
- 30 S. Ye and F. Neese, *Inorg. Chem.*, 2010, **49**, 772–774.
- 31 M. Swart, M. Güell and M. Solà, in *Quantum Biochemistry*, Wiley-VCH, 2010, pp. 551–583. DOI: 10.1002/9783527629213.ch19.
- 32 M. Swart, *J. Chem. Theory Comput.*, 2008, **4**, 2057–2066.
- 33 M. Gruden-Pavlovic, S. Stepanovic, M. Peric, M. Güell and M. Swart, *Phys. Chem. Chem. Phys.*, 2014, **16**, 14514–14522.
- 34 S. Stepanovic, L. Andjelkovic, M. Zlatar, K. Andjelkovic, M. Gruden-Pavlovic and M. Swart, *Inorg. Chem.*, 2013, **52**, 13415–13423.
- 35 M. Swart, *Inorg. Chim. Acta*, 2007, **360**, 179–189.
- 36 J. N. Harvey, *Struct. Bonding*, 2004, **112**, 151–183.
- 37 A. T. P. Carvalho, A. Barrozo, D. Doron, A. V. Kilshtain, D. T. Major and S. C. L. Kamerlin, *J. Mol. Graphics Modell.*, 2014, **54**, 62–79.
- 38 M. R. A. Blomberg, T. Borowski, F. Himo, R.-Z. Liao and P. E. M. Siegbahn, *Chem. Rev.*, 2014, **114**, 3601–3658.
- 39 S. P. de Visser, in *Comprehensive Inorganic Chemistry II*, ed. J. R. Poeppelmeier, Elsevier, Amsterdam, 2nd edn, 2013, pp. 619–634. DOI: 10.1016/B978-0-08-097774-4.00928-1.
- 40 D. Li, Y. Wang and K. Han, *Coord. Chem. Rev.*, 2012, **256**, 1137–1150.
- 41 S. Shaik, S. Cohen, Y. Wang, H. Chen, D. Kumar and W. Thiel, *Chem. Rev.*, 2009, **110**, 949–1017.
- 42 S. Shaik, D. Kumar, S. P. de Visser, A. Altun and W. Thiel, *Chem. Rev.*, 2005, **105**, 2279–2328.
- 43 M. Swart, M. Solà and F. M. Bickelhaupt, *J. Chem. Phys.*, 2009, **131**, 094103.
- 44 M. Swart, *Chem. Phys. Lett.*, 2013, **580**, 166–171.
- 45 J. Sun, J. P. Perdew and A. Ruzsinszky, *Proc. Natl. Acad. Sci. U. S. A.*, 2015, **112**, 685–689.
- 46 F. Feixas and M. Swart, unpublished data, 2017.
- 47 G. te Velde, F. M. Bickelhaupt, E. J. Baerends, C. Fonseca Guerra, S. J. A. van Gisbergen, J. G. Snijders and T. Ziegler, *J. Comput. Chem.*, 2001, **22**, 931–967.
- 48 E. J. Baerends, T. Ziegler, A. J. Atkins, J. Autschbach, D. Bashford, A. Bérce, F. M. Bickelhaupt, C. Bo, P. M. Boerrigter, L. Cavallo, D. P. Chong, D. V. Chulhai, L. Deng, R. M. Dickson, J. M. Dieterich, D. E. Ellis, M. V. Faassen, L. Fan, T. H. Fischer, C. Fonseca Guerra, M. Franchini, A. Ghysels, A. Giammona, S. J. A. van Gisbergen, A. W. Götz, J. A. Groeneveld, O. V. Gritsenko, M. Grüning, S. Gusarov, F. E. Harris, T. Heine, P. van den Hoek, C. R. Jacob, H. Jacobsen, L. Jensen, J. W. Kaminski, G. van Kessel, F. Kootstra, A. Kovalenko, M. V. Krykunov, E. van Lenthe, D. A. McCormack, A. Michalak, M. Mitoraj, S. M. Morton, J. Neugebauer, V. P. Nicu, L. Noodleman, V. P. Osinga, S. Patchkovskii, M. Pavanello, C. A. Peeples, P. H. T. Philipsen, D. Post, C. C. Pye, W. Ravenek, J. I. Rodríguez, P. Ros, R. Rüger, P. R. T. Schipper, H. van Schoot, G. Schreckenbach, J. S. Seldenthuis, M. Seth, J. G. Snijders, M. Solà, M. Swart, D. Swerhone, G. te Velde, P. Vernooijs, L. Versluis, L. Visscher, O. Visser, F. Wang,



- T. A. Wesolowski, E. M. van Wezenbeek, G. Wiesenekker, S. K. Wolff, T. K. Woo and A. L. Yakovlev, *ADF2016, SCM, Theoretical Chemistry*, Vrije Universiteit, Amsterdam, 2016.
- 49 E. van Lenthe and E. J. Baerends, *J. Comput. Chem.*, 2003, **24**, 1142–1156.
- 50 M. Swart and J. G. Snijders, *Theor. Chem. Acc.*, 2003, **110**, 34–41. Erratum: M. Swart and J. G. Snijders, *Theor. Chem. Acc.*, 2003, **111**, 156.
- 51 M. Swart and F. M. Bickelhaupt, *J. Comput. Chem.*, 2008, **29**, 724–734.
- 52 M. Swart and F. M. Bickelhaupt, *Int. J. Quantum Chem.*, 2006, **106**, 2536–2544.
- 53 M. Franchini, P. H. T. Philipsen and L. Visscher, *J. Comput. Chem.*, 2013, **34**, 1819–1827.
- 54 A. D. Becke, *J. Chem. Phys.*, 1988, **88**, 2547–2553.
- 55 A. Klamt and V. Jonas, *J. Chem. Phys.*, 1996, **105**, 9972–9981.
- 56 A. Klamt, *J. Phys. Chem.*, 1995, **99**, 2224–2235.
- 57 A. Klamt and G. Schuurmann, *J. Chem. Soc., Perkin Trans. 2*, 1993, 799–805.
- 58 M. Swart, E. Rösler and F. M. Bickelhaupt, *Eur. J. Inorg. Chem.*, 2007, **2007**, 3646–3654.
- 59 C. C. Pye and T. Ziegler, *Theor. Chem. Acc.*, 1999, **101**, 396–408.
- 60 E. van Lenthe, A. Ehlers and E.-J. Baerends, *J. Chem. Phys.*, 1999, **110**, 8943–8953.
- 61 E. van Lenthe, E. J. Baerends and J. G. Snijders, *J. Chem. Phys.*, 1994, **101**, 9783–9792.
- 62 E. v. Lenthe, E. J. Baerends and J. G. Snijders, *J. Chem. Phys.*, 1993, **99**, 4597–4610.
- 63 Y. Zhao and D. G. Truhlar, *J. Chem. Phys.*, 2006, **125**, 194101.
- 64 W. O. Koch, V. Schünemann, M. Gerdan, A. X. Trautwein and H.-J. Krüger, *Chem. – Eur. J.*, 1998, **4**, 686–691.
- 65 W. O. Koch, V. Schünemann, M. Gerdan, A. X. Trautwein and H.-J. Krüger, *Chem. – Eur. J.*, 1998, **4**, 1255–1265.
- 66 M. M. Mbughuni, M. Chakrabarti, J. A. Hayden, E. L. Bominaar, M. P. Hendrich, E. Münck and J. D. Lipscomb, *Proc. Natl. Acad. Sci. U. S. A.*, 2010, **107**, 16788–16793.
- 67 C.-W. Chiang, S. T. Kleespies, H. D. Stout, K. K. Meier, P.-Y. Li, E. L. Bominaar, L. Que, E. Münck and W.-Z. Lee, *J. Am. Chem. Soc.*, 2014, **136**, 10846–10849.
- 68 C. J. Knoot, V. M. Purpero and J. D. Lipscomb, *Proc. Natl. Acad. Sci. U. S. A.*, 2015, **112**, 388–393.
- 69 J. N. Harvey, M. Aschi, H. Schwarz and W. Koch, *Theor. Chem. Acc.*, 1998, **99**, 95–99.
- 70 J. N. Harvey, *Wiley Interdiscip. Rev.: Comput. Mol. Sci.*, 2014, **4**, 1–14.
- 71 S. Shaik, S. P. de Visser, F. Ogliaro, H. Schwarz and D. Schroder, *Curr. Opin. Chem. Biol.*, 2002, **6**, 556–567.
- 72 S. Shaik, H. Chen and D. Janardanan, *Nat. Chem.*, 2011, **3**, 19–27.
- 73 D. Usharani, B. Wang, D. A. Sharon and S. Shaik, in *Spin States in Biochemistry and Inorganic Chemistry: Influence on Structure and Reactivity*, ed. M. Swart and M. Costas, Wiley, Oxford, 2015, pp. 131–156. DOI: 10.1002/9781118898277.ch7.

