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Lipid-based nanoparticles for psoriasis treatment: a review on conventional treatments, recent works, and future prospects

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Psoriasis is a lingering inflammatory skin disease that attacks the immune system. The abnormal interactions between T cells, immune cells, and inflammatory cytokines causing the epidermal thickening. International guidelines have recommended topical treatments for mild to moderate psoriasis whilst systemic and phototherapy treatments for moderate to severe psoriasis. However, current therapeutic approaches have a wider extent to treat moderate to severe type of psoriasis especially since the emergence of diverse biologic agents. In the meantime, topical delivery of conventional treatments has prompted many unsatisfactory effects to penetrate through the skin (stratum corneum). By understanding the physiology of stratum corneum barrier functions, scientists have developed different types of lipid-based nanoparticles like solid lipid nanoparticles, nanostructured lipid carriers, nanovesicles, and nanoemulsions. These novel drug delivery systems help the poorly solubilised active pharmaceutical ingredient reaches the targeted site seamlessly because of the bioavailability feature of the nanosized molecules. Lipid-based nanoparticles for psoriasis treatments create a paradigm for topical drug delivery due to their lipids' amphiphilic feature to efficiently encapsulate both lipophilic and hydrophilic drugs. This review highlights different types of lipid-based nanoparticles and their recent works of nano formulated psoriasis treatments. The encapsulation of psoriasis drugs through lipid nanocarriers unfold numerous research opportunities in pharmaceutical applications but also draw challenges for the future development of nano drugs.

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1 Introduction

Psoriasis is an immune-mediated inflammatory dermatological disease that is also widely referred to as a systemic inflammatory disorder.^{1,2} Psoriasis can be indicated by a rapid build-up of cells on the skin's surface, forming erythematous (redness) papules and plaques with silver scales.³ The red, scaly plaques on the skin are itchy and sometimes painful.⁴ The psoriasis lesions typically formed on joints such as elbows and knees and may develop on any part of the skin such as hands, feet, scalp, neck, and face.⁵ The inflammation comes from the aberrant interactions between innate immune cells, T cells, and keratinocytes.⁶ However, psoriasis is non-contagious since it inherently involves innate and adaptive immune systems.⁷

The prevalence rate differs by region,⁸ which is epidemiologically attributable to racial factors, genetic background, lifestyle, and diet.^{9,10} Psoriasis prevalence amongst adults and children is reported to be ranging from 0.91% to 8.5% and 0 to 2.1%, respectively.^{11,12} Generally, it depicts a lower occurrence in Asian and some African populations but has a higher occurrence amongst Caucasian and Scandinavian people, notably in high-income countries.^{2,8} Nevertheless, the actual data amongst worldwide occurrence is not precise as Parisi *et al.* reported that the majority of low-income countries did not report the epidemiology of psoriasis in their countries.²

The disease may be triggered by several known factors or may also develop over the course of time. Infections from bacteria and viruses are reportedly exacerbated psoriasis conditions.¹³ Besides, certain medications (such as beta-blockers, lithium antimalarials, nonsteroidal anti-inflammatory medications, and tetracyclines) for adult psoriasis patients could also trigger the disease.^{14,15} They are also associated with numerous psoriasis comorbidities, including psoriatic arthritis, heart disease, hypertension, dyslipidaemia, obesity, type II diabetes, metabolic syndrome, inflammatory

bowel disease, anxiety, and depression.^{14,16} From psychosocial impacts, psoriasis has been an intimidating disease as some studies show that the risk for anxiety and depression has become dominant comorbidities in psoriasis pathophysiology that influences patients' physical appearance and self-esteem.^{17,18}

In determining the type of psoriasis (mild to moderate or moderate to severe), Psoriasis Area and Severity Index (PASI) score acts as the gold standard of severity lesion measurement, with PASI ≥ 10 indicating moderate to severe psoriasis.^{19,20} For mild to moderate psoriasis conditions, topical therapies are generally recommended over systemic therapies.³ The latter is preferable in cases involving moderate to severe disease conditions.^{3,21} Conventional topical delivery of antipsoriatic drugs typically utilises dosage methods at high concentrated therapeutics for a certain period.²² Nonetheless, this treatment possesses several limitations, including inadequate and unpredictable delivery to the afflicted skin as well as local toxicity due to skin irritation.^{22,23}

Biologic drugs (biologics or biologic agents) are the recent treatment strategy with superior efficacy to other treatments for moderate to severe type of psoriasis.^{3,24} This treatment employs recombinant proteins to inhibit inflammatory cytokines at the target site while functions systemically.²⁵ The advent of biologic agents has transformed the psoriasis treatment landscape after the first of its kind, etanercept, was approved by the Food and Drug Administration (FDA) in 2004.²⁶

Currently, the breakthrough of psoriasis treatments emerges from the utilisation of novel drug carrier systems or nanotechnology-based approaches leading to greater drug efficacy and long-term effects.^{22,27,28} Poorly soluble active pharmaceutical ingredients (API) can be improved through the use of nanotechnology whilst increasing their bioavailability and physicochemical stability.^{28–32} It is presumed that ineffective APIs have low pharmacokinetic profiles, lack of stability and



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the micelle structures. Other than that, she also works on the use of ultrasound on other subjects such as to synthesize gold nanoparticles, on natural rubber, sonoencapsulation, sonoextraction, sonopolymerization and sonodegradation such as on food wastes and organic pollutants.



solubility, or ones with limited dose toxicity.³³ Therefore, by employing novel drug delivery systems, these drawbacks can be overcome and thus improve the therapeutic actions.^{33–35} Nanoparticles have been proven in preclinical and clinical studies to reduce the drug's side effects at highly controlled drug delivery with relevant therapeutic concentrations.³⁶ Nanoparticles through the application of nanomedicine are designed to improve the drug's half-life, thus facilitating the API delivery to its targeted action site through nanocarriers.³³ Nano-based drug delivery or known as nanocarriers are nanosized particles that enhance the API's therapeutic effectiveness and reduce adverse effects. This is mainly because of their large surface area to carry the API.³⁷ Nanocarriers are drug delivery systems employed to regulate the pharmacokinetics and pharmacodynamic properties of the drug of interest.³⁸ Widespread attention to utilising topical nanocarriers for skin diseases has multiplied lately because they provide better skin permeation and controlled release at a reduced dosage regimen.²¹

Different interests in various types of nanoparticles have been widely investigated for topical psoriasis treatments. Polymeric nanoparticles, namely nanospheres³⁹ and nanocapsules⁴⁰ as well as polymer-based nanoparticles such as dendrimers,⁴¹ and polymeric micelles⁴² are a growing interest in these applications. Generally, many types of polymers are attractive biocompatible materials with proven favourable skin penetration and deposition effects.^{43–45} Despite so, their constituents are inclined to cytotoxicity while the cost of production is expensive for a scale-up and requires high-end instruments.^{45,46}

Hydrogels, water-insoluble cross-linked hydrophilic polymers, have become a popular trend in psoriasis with/without nanoformulations lately, as they can improve anti-psoriatic activity after drug encapsulation. The gels can solely act as drug vehicles,⁴⁷ or formulated into nanostructured supramolecules,⁴⁸ or integrated as excipients in various types of nanocarriers, such as graphene oxide,⁴⁹ polymeric nanoparticles,⁴³ nanosponge,⁵⁰ and niosomes.⁵¹ With the growing interest and research in hydrogels, it is expected that they would become a significant component of nano-based drug delivery systems in the future to further improve the therapeutic profiles.⁵²

At a more advanced approach than conventional phototherapy treatments, photothermal therapy (PTT) is a new technology combined with nano-agents to kill cancer cells by localised thermal damage,⁵³ which can also be utilised for psoriasis treatments. PTT results in the apoptosis of keratinocytes upon photo absorption of nano-agents that is activated by light.⁵⁴ Nirmal *et al.* developed gold nanorods and isatin (an anti-inflammatory agent) loaded into poly(lactic-co-glycolic acid) (PLGA) to form nanocomplexes for PTT, which was found to be able to reduce hyperthermia as a result of near-infrared (NIR) irradiation and also improve psoriasiform hyperproliferation.⁵⁴ Nevertheless, the use of gold as a nano-agent is associated with low photostability, which is liable to structural deformation.⁵⁵ Photodynamic therapy (PDT) is also a form of phototherapy, which is illuminated by a laser light beam of a specific wavelength, matching the absorption band of the topical administrated photosensitizer drug in psoriasis

treatment. For example, a study reported by Liu *et al.*⁵⁶ showed that zinc phthalocyanine (ZnPc) *via* PDT significantly inhibited the hyperproliferation of mouse vaginal epithelium induced by diethylstilbestrol and improved propranolol- and imiquimod-induced psoriasis-like symptoms. However, the PDT may perform damage to the plasma membrane or DNA of harmful cells, which may lead to cell death due to the highly reactive ¹O₂ generated *in situ*. Thus, nanocarriers based on carboxymethyl chitosan loaded with ZnPc has been reported.⁵⁷

Lipid-based nanoparticles (also known as lipid-based nanocarriers or nano-delivery systems) have been futuristic for topical nanocarriers in the treatments of wound-healing activity of skin diseases such as those found from psoriasis.^{58–60} Such interest in lipid-based nanoparticles is the result of their low toxicity potential and much easier sample preparation with low-production cost allowing mass industrial production, and better thermal stability when compared to other types of nanoparticles.^{61,62} On top of that, their constituents are also more biodegradable and biocompatible relative to synthetic polymer constituents like in other types of nanocarriers.^{58,63}

In this review, the pathophysiology of psoriasis is briefly explained such as the interaction of the inflammatory infiltrating cells with specific cytokines and alleles in the dermal layers. Multiple diagnosis strategies and the conventional treatments as recommended by dermatologists depending on the types of psoriasis are presented to provide information on different existing therapies that are regulated worldwide. As psoriasis is a skin disease, the skin penetration barrier of the stratum corneum and the challenges in delivering current therapies are further explained. This includes the discussion on unsatisfactory results from existing approaches. Subsequently, the emerging roles of lipid-based nanocarriers including their pros and cons are to be associated with the conventional strategies of psoriasis treatments, as well as their recent works. Lastly, current opportunities and challenges that arise from this novel therapeutic in treating psoriasis are also discussed comprehensively.

2 Pathogenesis of psoriasis

Psoriasis is a T cell-mediated inflammatory disease originating from the autoimmune system of the body. It is also construed to stem from a combination of genetic, epigenetic, and environmental factors,⁶⁴ and also attributable to endocrine hormones and immunological factors.¹⁰ Human leukocyte antigen (HLA) alleles (also known as major histocompatibility complex (MHC) molecules) that reside in both innate and adaptive immune systems and other susceptibility loci that bear genetic factors lead to the risk of developing psoriasis.^{65–67} It remains vague whether the inciting antigens are self-antigens, environmental antigens, or some combination of the two to a certain extent.⁶⁷ This autoimmune expedites the life cycle of skin cells and often be characterised by an excessively rare hyperproliferation of keratinocytes,⁶⁴ caused by inflammatory cell infiltration that generates into the epidermis and dermis skin layers.¹¹ Inflammatory cells are from many types of leukocyte population including lymphocytes, monocytes, and neutrophils that are



accelerated by tumour necrosis factor (TNF)- α , a type of inflammatory cytokine aided by dendritic cell activation.^{10,68} Hence, the feedback loop between adaptive immune cells (T cells), innate immune cells (*e.g.*, dendritic cells, Langerhans cells, natural killer cells, macrophages, granulocytes), and resident skin cells (*e.g.*, keratinocytes, endothelial cells) will increase the inflammatory response, thus, cause cell hyperproliferation.⁶⁹

TNF- α has the highest concentration of cytokine in psoriatic lesions since it is the target of immune response that is produced by various skin cells such as activated T cells, natural killer cells, keratinocytes, and Langerhans cells.⁷⁰ Psoriatic lesions are also indicated by the rich in mature CD4⁺ helper T (T_H1 and T_H17) and CD8⁺ cytotoxic T cells as well as the increase in both T_H1 and T_H17-related cytokines.^{67,70} The major pathway of antigens and cytokines in the psoriasis pathophysiology is illustrated in Fig. 1 and summarised below.

(1) Antigen-presenting cells (APCs), such as Langerhans cells (the specialised form of dendritic cells), act as T-cell recognition. They mediate specific HLA antigens that carry psoriasis alleles to migrate from the epidermis to the lymph nodes *via* afferent lymphatic vessels. The migration process is promoted by inflammatory cytokines such as TNF- α and interleukin (IL)-1 β . The APCs present their antigens (a process called antigen presentation) to naïve T cells, which results in the activation of T-cell proliferation in the dermis to regulate the immune response.^{70–72}

(2) Dermal dendritic cells activate T_H1 and T_H17 infiltrate by secreting pro-inflammatory cytokines IL-12 and IL-23, respectively. Besides, other APCs (inflammatory dendritic cells) help secrete IL-18, IL-23, and TNF- α cytokines. This process is also accompanied by the change of phenotype of the naïve T cells, from CD45RA that are often expressed as cutaneous lymphocyte-associated antigens (CLA) on their surface (known as CD45RA⁺ CLA⁺), to the effector memory T cells, CD45RO⁺ CLA[−].^{70,72–74}

(3) The memory CD45RO⁺ CLA[−] T cells are accompanied by the T_H1 and T_H17 cells to be carried out through the blood

vessels from the dermis to the epidermis as the result of the intradermal proliferation of T-cells.⁷⁴

(4) In the epidermis, the natural killer T cells are responsible for direct antigen presentation due to their proximity to the keratinocytes. The activated T_H1 cells are stimulated by IL-18 and IL-23 to secrete interferon-gamma (IFN- γ) cytokine. Meanwhile, T_H17 cells are stimulated by IL-23 to secrete IL-17 and IL-22 cytokines.⁷⁰ Then, the IL-23/IL-17 immune axis induces the infiltration of skin resident innate lymphocytes.⁷⁵ IFN- γ enhances IL-22 to initiate keratinocyte activation causing epidermal acanthosis (skin thickening) such as found in plaque psoriasis,^{72,76} thus the activation also induces IL-17 family to promote keratinocyte amplification and further chemokine secretion.⁷⁷

3 Treatment options and management of psoriasis

Psoriasis is an acute recurrent disease that requires long-term therapy, therefore tailored treatment is normally recommended based on the drug's compatibility with the patient.⁷⁸ Another pertinent factor is the patient's background in terms of gender, age, and previous medical history since individuals respond to drugs is somehow unique. The quality of life and the effects of the disease on the patient's responsiveness and tolerability are also essential psychosocial assessments to be evaluated by dermatologists before prescriptions.⁷⁹

In general, there are three different treatment options according to the type of psoriasis, which are topical, systemic, and phototherapy. Milder topical treatments can be recommended for patients with mild to moderate psoriasis. On the other hand, systemic treatments that include oral and biologic drugs are suitable for patients with moderate to severe type of

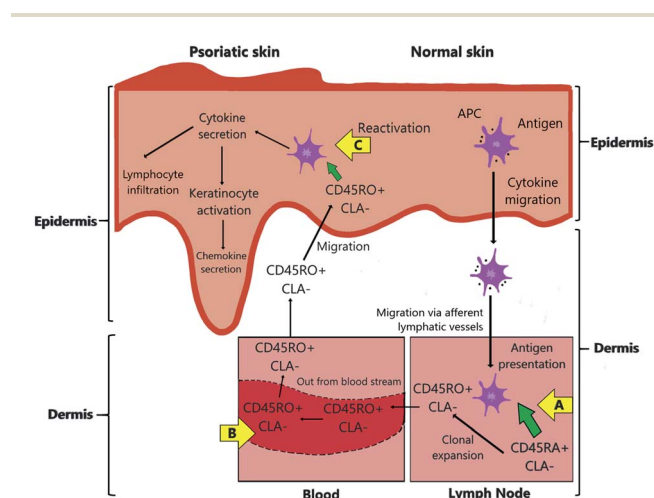


Fig. 1 A schematic pathway diagram of psoriasis pathology.

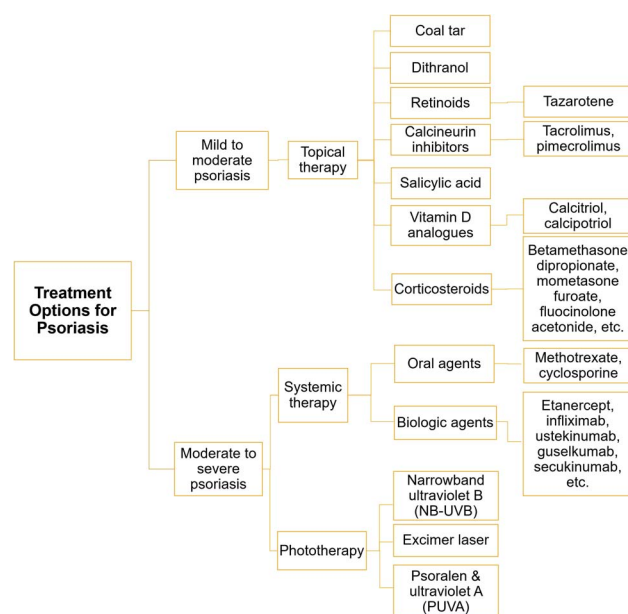


Fig. 2 Currently available treatment options by type of psoriasis.



psoriasis. Phototherapy treatment is commonly suggested for patients with moderate to severe type of psoriasis in the event of unsuccessful topical and systemic treatments. A list of treatment options under topical, systemic, and phototherapy are depicted in Fig. 2.^{21,27}

3.1 Topical treatments

The recommended first line of defence is topical treatment, in which the therapeutic is applied onto the afflicted skin. Such topical formulations are typically rendered in cream, solution, ointment, and shampoo (for scalp treatment). A wide range of topical treatments is available to treat psoriasis with different mechanisms of action. Topical drugs are expedient and accessible but usually deliver better efficacy in combination with other topical agents. In this section, the type of topical agents is sorted by older to the latest treatments relevant to the medications.

3.1.1 Coal tar. Since ancient times, coal tar has been part of psoriasis treatment, especially for scalp psoriasis but shows declining popularity in the past decades when compared to other topicals.^{80,81} This is probably because of its cosmetically unpleasant nature such as malodorous smell and staining of cloth, especially in patients with non-black hair.^{80,82} Out of thousand compounds constituted in the tar, polycyclic aryl hydrocarbon is the most analysed compound as it is believed to generate an effect in psoriasis treatment.⁸³ Contrariwise, carbazole of aryl hydrocarbons is suggested to exhibit a potent effect in the coal tar.⁸³ Even so, coal tar has raised concerns among clinicians and patients due to its thousands of unidentified components.⁸⁴ It is also banned in Canada and the European Union for cosmetic use because of carcinogenic concerns.⁸⁵

3.1.2 Dithranol. Dithranol (also known as anthralin) is one of the oldest treatments to alleviate plaque psoriasis⁸⁶ through inhibiting proliferation and inducing keratinocyte apoptosis, as demonstrated through human keratinocyte (HaCaT) cells.^{87,88} In essence, the direct or indirect effects of dithranol on the inflammatory response of psoriasis are not entirely comprehended, although they have been investigated by multifaceted studies.^{89,90} Notwithstanding the statement, a recent study showed that topical dithranol leads to a rapid reduction in the PASI score, resultant in a decrease in epidermal hyperproliferation and delayed reduction of inflammatory penetration in psoriatic skin.⁸⁹ The study also suggested that psoriatic activity can be suppressed by targeting the keratinocytes (residing in the epidermis) rather than the immune cells. Despite the benefits, dithranol may not be powerful enough for skin permeation as it is associated with low solubility and stability, toxicity, staining, and skin irritation.⁹¹

3.1.3 Retinoids. Retinoids are the derivatives or vitamers of Vitamin A. The sole FDA-approved retinoids for psoriasis treatments are tazarotene and acitretin.⁹² Tazarotene is a topical drug, whilst acitretin is an oral agent. The retinoids work on retinoic acid receptors and retinoid-X-receptors, initiating gene expression's modification of inflammatory cytokines and thus, inhibits the keratinocyte proliferation. However, topical

tazarotene can cause localised toxicity such as skin irritation and photosensitivity.^{93,94} Tazarotene's efficacy has been widely studied through combination with topical corticosteroids as they optimally enhance the post-treatment effects of psoriasis, in which tazarotene reduces the irritation whilst corticosteroids provide anti-inflammatory effects.^{95,96}

3.1.4 Calcineurin inhibitors. Tacrolimus and pimecrolimus are two different types of topical calcineurin inhibitors (immunosuppressants) that act as the first-line therapeutic options for inverse psoriasis (body fold area) for long-time use.^{97,98} They are functional in anti-inflammatory and immunomodulatory activities to inhibit calcineurin phosphatase; hence, suppressing T-cell activation and cytokine production.⁹⁹ Tacrolimus is also used to treat genital psoriasis, is well-tolerated amongst adults, and is suitable for patients who are sensitive to steroids but with certain side effects like mild pruritus and burning sensation onset the treatment.^{100–102} Similarly, pimecrolimus has also been used for such psoriasis conditions and facial psoriasis with equivalent potency in children but has less therapeutic efficacy than tacrolimus in adults.^{98,101,102}

3.1.5 Salicylic acid. Salicylic acid is a potent keratolytic agent, often used in combination with other topical agents such as corticosteroids and calcineurin inhibitors. Its combination with calcineurin inhibitors is believed to help in the excellent drug absorption into psoriasis plaques.¹⁰³ The use of salicylic acid alone for scalp psoriasis may result in temporary shedding of telogen hair, but the cessation of use may resolve the issue.¹⁰⁴ Besides, it is not recommended to use salicylic acid with a concentration greater than 10% on large body surface areas. This is mainly to avoid possible side effects such as oral mucosal burn, frontal headache, and nausea. Some previous research has shown the potent effects of salicylic acid as a penetration enhancer through combination with a calcineurin inhibitor, vitamin D analogue, and corticosteroid for better skin permeation and sustained release profile.^{105–107}

3.1.6 Vitamin D analogues. Calcipotriol and calcitriol are two types of vitamin D analogues derived from sunlight. Presumably, vitamin D deficiency is associated with psoriasis as its presence at an adequate amount helps suppress the production of inflammation's potent cytokine mediators, namely IL-2, IL-6, and IFN- γ ; thus, keep the body healthy. Moreover, there is a correlation between T_H17 cells (that work for a healthy immune system) and vitamin D in psoriasis patients.¹⁰⁸ Vitamin D evidently stimulates suppressor T cells that regulate other cells in the immune system and inhibits cytotoxicity and, thus, is a natural killer of cell hyperproliferation. Nevertheless, their association remains debatable, albeit well-documented,¹⁰⁹ and vitamin D analogues are often recommended with other topical agents. Calcipotriol and betamethasone dipropionate (a type of corticosteroid) has been studied widely to treat trunk and limb psoriasis as they both work synergistically to reduce inflammation. The combination therapy of the two helps control the imbalance between regulatory CD8⁺ or CD4⁺ T cells and pro-inflammatory CCR6⁺ gamma delta ($\gamma\delta$) T cells for antipsoriatic activity.¹¹⁰



3.1.7 Corticosteroids. Corticosteroids are the mainstay for topical therapies of psoriasis. They work by inhibiting the release of phospholipase A2 and act directly on deoxyribonucleic acid (DNA) and inflammatory cytokines.¹¹¹ The use of corticosteroids alone acts as first-line topical therapy by reducing inflammation and slowing down cell hyperproliferation, especially in genital psoriasis.¹¹² As discussed earlier, the effects of corticosteroids are more powerful through combination with other topical agents, namely retinoids, salicylic acid, and vitamin D analogues, as evidenced by multiple studies. However, local side effects like perioral dermatitis, striae, hypertrichosis, and infections are reportedly developed from the long-time use of topical corticosteroids.¹¹³ On a side note, studies showed that there is a moderately high perception of steroid phobia among our current generation, especially in women.^{114,115} In general, the studies indicate that the fear of using steroid-related drugs are due to skin deterioration conditions, weight gain, asthma, and stunted growth after obtaining sources from dermatologists and media.

3.2 Systemic treatments

The systemic treatment utilises drug that circulates through the whole body. It can be divided into two types: (1) oral agent; and (2) biologic agent that is administered into the system by injection or intravenous (IV) infusion. International guidelines for psoriasis treatments have recommended that if topical and phototherapy treatments are deemed unsuccessful, systemic nonbiologic (oral agent) and biologic therapies may be pursued to achieve skin clearance. Since monotherapy of topical treatments to moderate to severe psoriasis causes many dissatisfactions, patients can also benefit from systemic therapies that provide better efficiency and favourable tolerability to reduce clinical symptoms, accomplish treatment efficacy targets, and improve their quality of life.¹¹⁶

3.2.1 Oral drugs. Systemic nonbiologic therapies work by suppressing inflammatory responses in the form of oral consumption. Methotrexate and cyclosporine are the two famous oral, systemic therapies used for decades to treat psoriasis, often recommended as monotherapy or other systemic therapies. Although biologics have widespread attention in psoriasis treatments recently, systemic nonbiologic therapies are with many advantages. Oral systemic generally curb pervasive psoriasis, is lower cost than older medications and biologics, and has the convenience of use and availability.¹¹⁷ Besides can be administered orally, the active ingredients are also loaded in nanoformulations to treat psoriasis topically.

3.2.1.1 Methotrexate. Methotrexate is a comprehensive treatment for all types of psoriasis conditions. It normalises psoriasis conditions by inciting the apoptosis process of keratinocyte proliferation.¹¹⁸ It is usually administered weekly at a low single dose to reduce side effects before the dose can be gradually increased to achieve its optimal efficacy.¹¹⁹ However, the continuous use of methotrexate would cause liver damage and pulmonary toxicity (lung damage). Besides, myelosuppression (low blood cell production) and immunosuppression (weakened immune system) are other problems associated with

methotrexate's long-term use. It is also reported that methotrexate-receiving patients have a higher cardiovascular event risk than TNF inhibitor users.¹²⁰

3.2.1.2 Cyclosporine. Cyclosporine (sometimes spelt as cyclosporin and known as cyclosporine A) is another type of calcineurin inhibitor that has been one of the most effective systemic psoriasis treatments. Both cyclosporine and methotrexate's rotational use are found to help reduce long-term kidney toxicity (nephrotoxicity) whilst minimising the adverse impacts of plaque psoriasis.¹²¹ The use of cyclosporine drug is reportedly equally safe and effective for severe paediatric and adult plaque psoriasis patients¹²² but with mild side effects such as pain abdomen and an increase in serum creatinine (a chemical waste product in the blood).^{123,124} However, liver toxicity (hepatotoxicity) has been subject to the most affected organ for prolonged use of cyclosporine.¹²⁵

3.2.2 Biologic drugs. After the successful case of treating a patient that suffered from Crohn's disease and moderate to severe psoriasis with a TNF- α blocker in 2000,¹²⁶ the last decade witnessed the discovery of IL-17 and IL-23 pathways responsible for the pathogenesis of psoriasis.^{127,128} Biologics have pragmatic strength of recommendations for psoriasis treatments, typically recommended based on the patient's case evidence and the drug's performance. The American Academy of Dermatology (AAD) and the National Psoriasis Foundation define biologic agents as "engineered monoclonal antibodies and fusion proteins that exert their therapeutic actions by blocking specific cytokines or cytokine receptors critical to psoriatic inflammation".²⁶ Presently, there are 11 types of FDA-approved biologic agents for adult psoriasis treatments, classified by their cytokine classes (TNF, IL-12, IL-23, IL-17A) (Fig. 3). Secukinumab, as the first-line systemic biological treatment¹²⁹ is discussed to represent the overview of this type of advanced drug.

Secukinumab (Cosentyx®) inhibits IL-17A from binding to its cytokine receptor for inflammation.¹³⁰ It is highly favourable in safety profile as its efficacy demonstrates comparable with the commonly used psoriasis treatments (e.g., methotrexate and TNF- α blockers).¹³¹ Its efficacy is widely reported in treating moderate to severe plaque psoriasis¹³² and psoriatic arthritis.¹³³ Other than that, those with a past treatment failure, difficult-to-treat patterns of psoriasis (nail, scalp, and palmoplantar psoriasis),¹³⁴ and have clear skin desire¹³⁵ can also be benefited

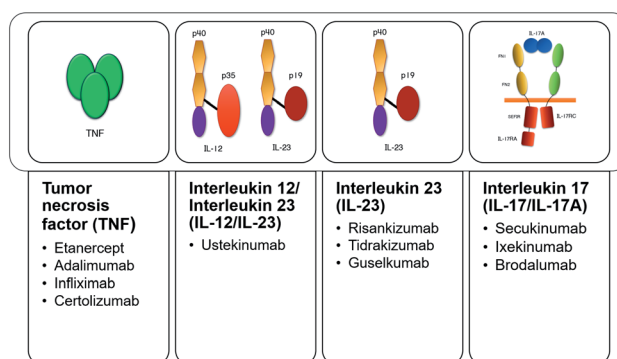


Fig. 3 FDA-approved biologic agents by their classes.



from this biologics. Having said that, secukinumab is also perceived to have the most frequent side effects, mainly infections, compared with other biologic agents.¹³⁶ It has been reported that nasopharyngitis, upper respiratory tract infections, and headaches are the expected side effects of secukinumab.¹³³

Overall, biologic agents are considered expensive or as high-investment treatment based on the type of biologics and individual's PASI score.¹³⁷ In the wake of the globally impacted pandemic era of COVID-19 (novel coronavirus disease) that severely affects the human respiratory system, the continuation of biologic treatments is controversial. This is due to the high respiratory rate before the pandemic emerged.¹³⁸ That raises concern if it could increase the consumers' susceptibility to the virus. On the contrary, treatment discontinuation would cause loss of response when treatments are reintroduced or further exacerbated by antibody formation to the biologics.¹³⁸

3.3 Phototherapy treatments

Photo-based therapy has been developed for psoriasis treatments since the 1920s, and presently serves as a mainstay for the moderate to severe type of psoriasis.¹³⁹ In general, phototherapy is deemed an effective and comparatively affordable treatment. For instance, it is estimated that a 3 year phototherapy treatment would cost about \$5000 as compared to the 3 year duration cost of secukinumab at about \$182 718.¹⁴⁰ Phototherapy treatments also have relatively rare side effects, particularly immunosuppression as compared to systemic therapies.^{141,142} Presently, narrowband ultraviolet B (NB-UVB), excimer laser/lamp (targeted phototherapy), and psoralen plus ultraviolet A (PUVA) are three different types of phototherapy treatments to treat psoriasis as recommended by international guidelines.

Nevertheless, phototherapy might be unpopular among psoriasis patients due to concerns about its carcinogenicity and is therefore not recommended for long-term treatment.¹⁴¹ The declining usage of phototherapy is also contributed after the advent of biological agents.^{3,143} Moreover, NB-UVB is also not suitable for psoriasis patients with photosensitive disorders (such as those suffering from xeroderma pigmentosa).^{141,143} Other than that, the literature reveals that PUVA undergoes a higher adverse risk than UVB and is not recommended for maintenance therapy due to cutaneous malignancy risks.¹⁴⁴

3.3.1 Narrowband ultraviolet B (NB-UVB) and excimer laser. NB-UVB light (311 nm) and excimer lamp/laser (308 nm) are two effective phototherapies that currently act as the first-line treatment for stable plaque psoriasis.¹⁴⁵ Both therapies are equally useful for psoriasis' skin clearance.^{141,145} The choice of NB-UVB is usually made after the failure of a series of topical agents.¹⁴⁶ Phototherapy requires repetitive skin exposure to UV light. The generation of phototherapy works where the UV radiation from the treatments induce apoptosis in T lymphocytes and keratinocytes in the epidermis to control the psoriatic lesions.¹⁴⁷ The trend of the highly cost-effective UVB treatment is the exclusive option for a particular group of patients, such as human immunodeficiency virus (HIV), internal malignancy, and pregnancy, where contraindication of their systemic

immune reaction might happen (if modified while using other psoriasis treatments).¹⁴⁸

The excimer laser is remarked to be more efficacious than the excimer lamp and is recommended by the AAD for PASI < 10.¹⁴¹ Whilst other parts of the body can be induced by the NB-UVB phototherapy, the nose, ear, and palpebral region represent difficult-to-treat areas of psoriasis. Therefore, the excimer laser is a better phototherapy alternative to target those mentioned psoriasis areas.¹⁴⁹ Besides being a powerful and effective treatment, it is preferable amongst psoriasis patients as the laser offers long-term remission of psoriasis.¹⁵⁰

3.3.2 Psoralen plus ultraviolet A (PUVA). PUVA is a photochemotherapy treatment considered another effective treatment of chronic plaque psoriasis.¹⁵¹ The combination of psoralens and UVA has been shown to reduce PASI by 75% to 80% amongst the users, making this phototherapy useful as a second-line treatment after unresponsive topical and NB-UVB treatments.¹⁵² Instances of specific failure treatment indications of NB-UVB relapse are when the patients develop pustular psoriasis and pityriasis rubra pilaris conditions. Consequently, to introduce PUVA phototherapy to the patients after such psoriasis deterioration, psoralen needs to be sensitised upon them as a form to enhance the therapeutic procedure.¹⁴⁶ Psoralen is a group of light-sensitive compounds derived from natural plants, commonly found in *Psoralea corylifolia* (Babchi) that exhibit therapeutic properties.¹⁵³ They are likely to instigate skin-related conditions namely burning, itching, and pigmentation, and also nausea after this therapeutic action.¹⁵⁴

4 Overcoming skin barrier challenges for nano-based drug delivery

A thorough analysis and understanding of the various skin's structures and physiology is required to design drugs with improved permeability and skin penetration. Skin is the largest organ in the human body and is also responsible for protecting one's system against exogenous pathogens and environmental factors and preventing water and electrolyte loss from the internal.^{155,156} As a result, the stratum corneum prevents the penetration of larger molecules (>500 Da)¹⁵⁷ across the outermost layer of the skin epidermis. The barrier is referred to as a 'brick-and-mortar system' and comprises approximately 15 layers of flattened corneocytes (anucleated keratinocytes) within a multilamellar, lipid-rich extracellular matrix.¹⁵⁸ The three essential components of stratum corneum are: (1) natural moisturising factor (NMF)-laden and lipid-bound corneocytes

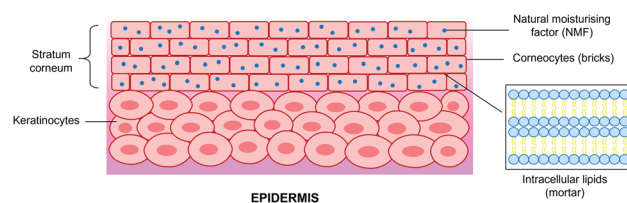


Fig. 4 The 'brick-and-mortar' system of the stratum corneum within the epidermis.



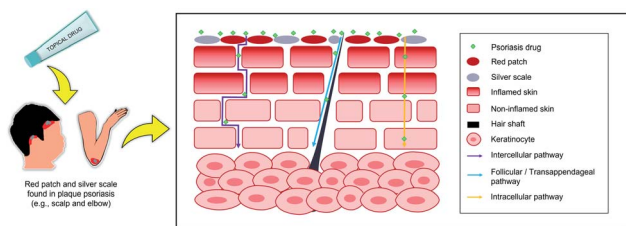


Fig. 5 The pathways of topical drug delivery for psoriasis treatments.

(terminally differentiated keratinocytes); (2) corneodesmosomes (proteinaceous rivets holding corneocytes together); and (3) lipids as shown in Fig. 4.

Optimal skin barrier function can be achieved by the correct stratum corneum lipid lamellar structure with well-structured lipid bilayers, a massive amount of phytosphingosine-containing ceramides, and a high stratum corneum lipid/protein ratio.¹⁵⁹ Other than that, the lipid matrix is also vital in the skin barrier function. Ceramides, fatty acids, and cholesterol are the principal components of the skin barrier lipids present in an equimolar ratio.¹⁶⁰ Headgroup chemistry (hydroxyl group's number and position) and tail structure (unsaturation of sphingoid moiety) play parts in the ceramide bilayers' structural and transport properties.¹⁶¹

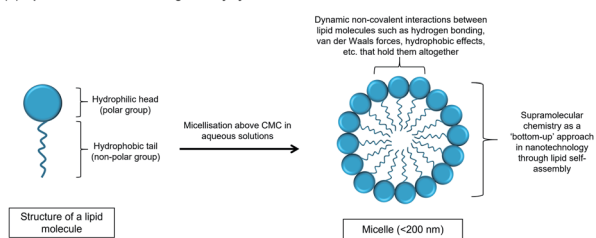
As a result of the complicated skin barrier lipids, nanoparticles may act as a tool to overcome the complexity of the stratum corneum in which chemical enhancers are unable to. In this way, nanoparticles stimulate drugs' residence time by channelling them to the target site while improving their penetration.¹⁶² As conventional topical treatments possess low drug permeation across the scaly psoriasis lesions, nanoparticles' microscopic morphology ensures that drugs can penetrate through the skin layers effectively.

It is widely regarded that topical drug delivery is the ideal route for skin diseases owing to its ability to deliver a high concentration of drugs whilst reducing systemic toxicity,¹⁶³ as illustrated in Fig. 5. Its mechanism allows the API or drug to pass through the skin to the local therapy or across the skin by systemic treatment. Topical drug formulations like cream, gel, and ointment have been studied for local effect rather than systemic action. Thus, chemical enhancers like surfactants, alcohols, and amines are employed to deliver active therapeutics by overcoming the skin barrier (topical delivery).¹⁶⁴ Besides topical, transdermal drug delivery can also target local skin disease and reach the bloodstream (systemic treatment). In transdermal delivery, the drug's systemic circulation (distribution process) is deemed successful upon skin absorption to the body's tissues *via* capillary action at a specific rate.¹⁶⁵ In a common occurrence, the release of drugs are influenced by environmental parameters such as pH, redox reaction, and enzymes.³⁷

5 Types and properties of lipid-based nanoparticles for psoriasis treatments

Lipids (Fig. 6) are amphipathic since they possess both the hydrophilic (polar head group) and the hydrophobic (non-polar tail group) properties.¹⁶⁶ They self-assemble in aqueous

(A) Lipid as a surfactant for drug delivery systems



(B) Influence of critical packing parameter (Cpp) on the geometrical shape of lipid-based nanoparticles

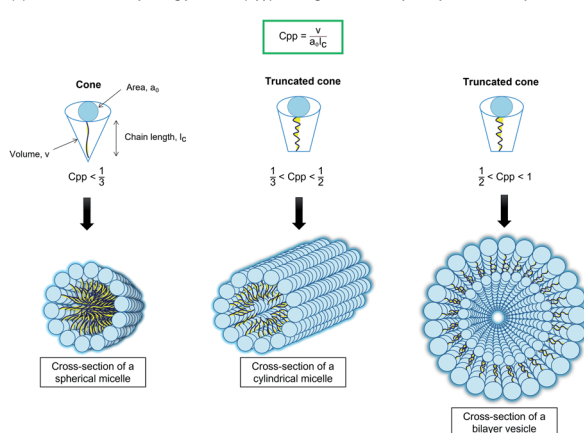


Fig. 6 Properties of lipids as nanocarriers.

solutions to minimise contact of their non-polar part with the water molecules. This characteristic allows them to spontaneously organise themselves into ordered patterns without human interference.¹⁶⁷ This usually results in a micelle formation or micellization process when the surfactant concentration is above the critical micelle concentration (CMC).^{167,168} Such a phenomenon is one of the most important physicochemical properties since many types of lipid-based are micelles and act as surface-active agents (surfactants). Micelles act as carriers for the majority of hydrophobic (lipophilic) drugs upon their incorporation at nanoscale size whereby their architecture has a resemblance to lipid-based nanoparticles.¹⁶⁹ The micellar resemblance of drug carriers has benefited nearly all possible drug administrations to increase bioavailability whilst reducing adverse effects.¹⁷⁰

In addition to CMC, the critical packing parameter, Cpp of the lipid surfactants also governs the geometrical shape of the aggregates,^{171–173} as shown in Fig. 6. Their self-assembly morphology in an aqueous medium generally can be predicted to be either spherical micelles, cylindrical micelles or vesicles, as proposed by Israelachvili.^{171–173} For example, spherical ($Cpp < 1/3$), cylindrical, ($1/3 < Cpp < 1/2$), and vesicles ($1/2 < Cpp < 1$). Traditionally, the self-assembly properties and supramolecular chemistry are important for drug delivery systems through the bottom-up approach in nanotechnology.^{174,175} Supramolecular chemistry is generally referred to as the soft and dynamic non-covalent interactions of the assemblies, such as hydrogen bonding and van der Waals forces, but the interactions are capable of holding the amphiphiles together.^{174,176,177} This bottom-up approach has been an



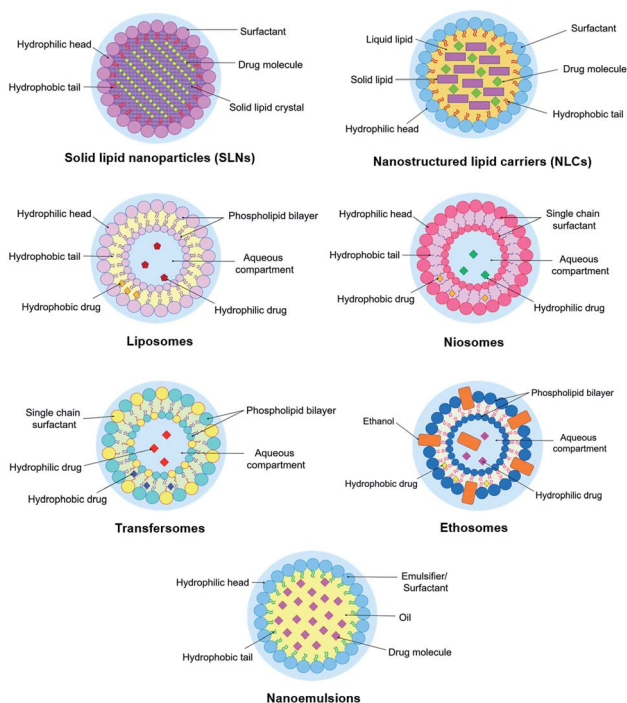


Fig. 7 Structural diagrams of lipid-based nanoparticles.

exciting route before the aggregates are formed into their shapes that are mainly influenced by their Cpp prediction.^{171,174} Therefore, the stable colloidal lipid systems in nanoscale supramolecular structures help overcome skin physiological barriers for optimised targeting and controlled release of drug delivery.^{168,178,179}

Solid lipid nanoparticles (SLNs), nanostructured lipid carriers (NLCs), liposomes, niosomes, transfersomes, ethosomes, and nanoemulsions are classified as lipid-based nanoparticles because of their lipid arrangements. This section focuses on these different types of lipid-based nanoparticles because of their significant applications in psoriasis treatments lately. Their structural representations are illustrated in Fig. 7. This section also describes their overall concept, roles, components, preparation methods, advantages, and disadvantages for topical and transdermal psoriasis delivery. Recent up-to-date works involving psoriasis drugs loaded in the lipid-based nano delivery systems (nanoformulations) are compiled in Table 1.

5.1 Solid lipid nanoparticles (SLNs)

SLNs is the first generation of lipid nanocarrier systems introduced in the 1990s. It is an advanced drug delivery carrier where the submicron particles (40 to 1000 nm) are lipids that are dispersed in an aqueous surfactant solution.¹⁸⁰ The surfactant used for SLNs acts as an emulsifier while the lipids (*e.g.*, fatty acids, steroids, glycerides) are solid at the body and ambient temperature.¹⁸¹ SLNs are commonly formulated by a cold or hot homogenisation method, depending on the drug's thermal stability.^{182,183} SLNs can also be formulated by high-pressure homogenisation and ultrasonication methods.^{182,184} SLNs are

site-specific, developed to overcome the drawbacks of polymeric nanoparticles and liposomes, such as polymer degradation and cytotoxicity, and drug leakage, and also liposome's phospholipid degradation.¹⁸⁵ Their dynamic system also allows modification in formulating the components compared to the liposomes, such as the surfactant can either be ionic or nonionic and is often used with a co-surfactant to reduce the particle size.¹⁸⁶ Nevertheless, established studies show that there is a particular type of surfactant and lipid components in SLN formulation which might have influenced their high cytotoxicity level and thus, have less biocompatibility feature such as sodium dodecyl sulphate (surfactant) and stearic acid (lipid).^{187,188}

Within SLN systems, there exists a perfect crystal structure that resembles 'a wall of brick' in which drug molecules are encompassed between similar lipid components.¹⁸⁹ For this reason, the nanosized particles remain in the solid-state even after administration, which offers a controlled release profile.¹⁹⁰ Because of the same reason, SLNs can provide drug protection from external factors by their rigid core matrix.¹⁸⁶ They are also possible to be produced on large scale by the high-pressure homogenisation method.⁶² Despite the basis of SLNs' core matrix that is better suited for hydrophobic-loaded drugs, research has reported for SLNs' capability to entrap and release hydrophilic drugs.^{191,192} However, since the solid core encapsulated in the lipid system has a perfect crystalline structure, this would lead to inefficient drug loading and possible drug repulsion due to the crystallisation process. Other inevitable factors may also affect SLNs' drug encapsulation in the lipid matrix, such as the melting of lipids, and the miscibility or dispersion of the drug with the lipids.⁶²

5.2 Nanostructured lipid carriers (NLCs)

About two decades ago, researchers observed some limitations in SLNs, such as drug leakage through the matrix during storage and the less efficient drug loading as mentioned previously.⁶² The limitations led to the invention of NLCs, second-generation lipid carrier systems that are capable of better drug loading efficiency. Within NLCs' system, the drug is encapsulated in the mixture of unsaturated, amorphous, or liquid lipids (oils) to the solid lipids.¹⁹³ The premise of NLCs' design was also improved to obtain a solidified core with a little-to-no crystalline matrix.¹⁸³

NLCs also have similar preparation methods to SLNs, namely cold homogenisation, hot homogenisation, and hot emulsification-ultrasonication.¹⁸³ However, unlike SLNs, the oils in NLCs formulation ensure the expulsion of the loaded drug is minimised during the storage period as they obstruct the recrystallisation process.¹⁹⁰ The incorporation of drugs is also improved due to lipid imperfections within their matrix systems.¹⁹⁴ Thus, higher drug solubility and a controlled release profile can be achieved by NLCs.⁶² Currently, NLCs are regarded as a more superior drug carrier than SLNs due to their more excellent formulation and biocompatibility properties. NLCs are also site-specific, and their topical applications improve skin irritation and occlusive properties, as well as skin permeation and retention.^{195–197}



Table 1 Recent developments of lipid-based nanoparticles for psoriasis treatments (2016–2020)

No.	Lipid carrier	Antipsoriatic agent	Route of drug administration	Method	Remark	Reference
1	Solid lipid nanocarriers (SLNs)	Methotrexate and etanercept	Topical	Hot ultrasonication	Prolonged drug release <i>in vitro</i> and nontoxic towards human keratinocytes and fibroblasts	272
		Cyclosporine	Topical	Microemulsion	Exhibited systemic absorption and reduced side effects whilst increased its concentration in the skin layers <i>in vitro</i>	273
		Thymoquinone (<i>Nigella sativa</i> extract)	Topical	Melt-emulsification and ultrasonication	Low skin irritation score and reduced symptoms in erythema, oedema, and thickening in PASI score	274
		Tacrolimus	Topical	Emulsification and low-temperature solidification	High <i>ex vivo</i> skin penetration value and deeper <i>in vivo</i> skin layer penetration	275
2	Nanostructured lipid carriers (NLCs)	Methotrexate	Topical	Solvent diffusion	Significant entrapment efficiency and drug deposition whilst no erythema was indicated at primary skin irritation index	276
		Mometasone furoate	Topical	Microemulsion	High skin deposition and low primary skin irritation index with complete clearance of parakeratosis <i>in vivo</i>	277
		Dithranol	Topical	Hot melt homogenisation	Reduced the symptoms by PASI score and enzyme-linked immunosorbent assay with depletion in disease severity and cytokines like ILs-17, 22, 23, and TNF- α	278
		Methotrexate	Topical	Solvent diffusion	Improved therapeutic response and reduced local side effects <i>in vivo</i> with a decline of PASI score, oxidative stress, inflammatory cytokines like TNF- α , IL-1 β , and IL-6, and IMQ-induced histopathological alterations in mouse ear models	279
3	Liposomes	Cyclosporine	Topical	Thin-film hydration	Showed acceptable safety profile in the remedy of chronic plaque psoriasis	280
		Psoralen	Topical PUVA	Cationic liposomes by thin-film hydration method	Multiple fold increase in skin permeation study with reduced psoriasis plaque symptoms and levels of psoriatic cytokines (TNF- α , IL-17, and IL-22)	281
		Cyclosporine	Topical	Cationic liposomes by ethanol injection method	Had shear-thinning behaviour and reduced the symptoms of psoriasis plaque and levels of psoriatic cytokines (TNF- α , IL-17, and IL-22)	282



Table 1 (Contd.)

No.	Lipid carrier	Antipsoriatic agent	Route of drug administration	Method	Remark	Reference
4	Niosomes	Zedoary turmeric oil and tretinoin	Topical	Ethanol injection	High drug penetration and drug retention <i>in vitro</i> and significant psoriasis drug-dependent <i>in vivo</i>	283
		<i>trans</i> -Retinoic acid and betamethasone	Topical	Thin-film hydration	Nontoxic and time-dependent cellular uptake on HaCaT cells whilst reducing the epidermal thickness and cytokine level (TNF- α and IL-6) <i>in vivo</i>	284
		Liposomal spherical nucleic acids (L-SNA)	Topical	IL-17A receptor targeting	Reduced significantly in the PASI score and epidermal thickness on imiquimod (IMQ)-remedied mouse skin, and in the expressions of IL17RA, IL-17C, defensin, beta 4 (DEFB4), TNF- α , and phosphoinositide 3-kinase (PI3K) on psoriatic 3D rafts	285
		Bexarotene (retinoid X receptor)	Topical	Thin-film hydration	A significant reversal of psoriasis (reduced scaling, inflammation without any toxicity) on IMQ-induced psoriatic plaque model mice whilst reducing the expression of cytokines (IL-17, IL-23, and IL-2)	286
		Diacerin	Topical	Thin-film hydration	Prominent skin penetration in epidermal and dermal layers <i>in vitro</i> and found stable at low temperature	287
5	Transfersomes	Acitretin	Topical	Thin-film hydration	Improved <i>ex vivo</i> permeability assay and drug deposition in HaCaT cells	288
		Celastrol	Topical	Thin-film hydration	Alleviated erythema and scaling psoriasis symptoms <i>in vivo</i>	51
		Tacrolimus	Transdermal	Rotary evaporation-sonication	Higher mean residence time and skin permeation <i>in vivo</i> compared to that of liposomal formulation	289
6	Ethosomes	Methotrexate	Topical	The fusion method	Reduced the thickness score in dose-dependent IMQ-induced psoriasis mouse model with no organ toxicity observed	290
		Methotrexate and salicylic acid (MTX-SA)	Topical	The cold method	Decreased in PASI score whilst MTX-SA had more prolonged release than MTX alone	291
		Anthralin (dithranol)	Topical	Thin-film hydration	High permeation rate <i>ex vivo</i> with minimised adverse effects after treatment in clinical studies	292
7	Nanoemulsions	Cyclosporine	Topical	Phase inversion composition	High efficacy rate <i>in vitro</i> and improvement in skin hydration <i>in vivo</i>	293



Table 1 (Contd.)

No.	Lipid carrier	Antipsoriatic agent	Route of drug administration	Method	Remark	Reference
		Tacrolimus	Topical	Spontaneous emulsification	Prolonged-release pattern and dermal bioavailability improvement <i>in vitro</i> whilst showed depletion in serum cytokines and improvement in psoriasis symptoms <i>in vivo</i>	294
		Methotrexate	Topical	Low energy emulsification	Improved skin permeation and retention in deep skin layers <i>ex vivo</i> with an increase in antipsoriatic efficacy, effective skin retention, and lesser serum and tissue accumulation <i>in vivo</i>	295
		Curcumin (natural compound)	Topical	Low-energy emulsification	Multiple fold increase in skin permeation and fast healing in psoriatic activity	296
		Imiquimod and curcumin	Topical	Low energy emulsification	Prevented the appearance of psoriasis-like symptoms and slowing down the psoriatic activity	297

5.3 Nanovesicular systems

Nanovesicular systems (also known as nanovesicles) can typically be distinguished by their unilamellar structures, which have become up-and-coming drug delivery and targeting systems.¹⁹⁸ The original vesicles are conventional liposomes before they are adapted into niosomes, transfersomes, and ethosomes, named after their characteristics and compositions.¹⁹⁹ They can be assembled per need of their membrane components, and the type of the drug (hydrophobic or hydrophilic),¹⁹⁹ which are useful for topical delivery of psoriasis.²⁷

5.3.1 Liposomes. The liposomes are nano-, and micro-sized colloidal multilayer vesicles (50–1000 nm in diameter) consist of an aqueous compartment enclosed by natural or synthetic lipid bilayers.^{37,200} Liposomes are permeable to water and osmotically sensitive.²⁰¹ Liposomes may present in multilamellar, small unilamellar, large unilamellar, or multivesicular shapes. Regardless of that, liposomes' size and homogeneity are more essential for drug encapsulation compared to the lamellar numbers.^{202,203} Liposomes can be prepared conventionally by thin-film hydration, reverse-phase evaporation, detergent dialysis, or solvent injection method.^{199,204} Further formation steps namely sonication, high-pressure homogenisation or extrusion technique is required to obtain smaller unilamellar vesicles.^{199,204}

Several drawbacks associated with phospholipid bilayers of conventional liposomes are revealed in literature such as high-cost production, short half-life, and low solubility.^{199,200} Besides, the encapsulated drug molecules within liposomes may leak out and fuse because of their less stable surfactants (phospholipids).²⁰⁰ Phospholipids that make up the membrane components of liposomes have a negatively charged phosphate

hydrophilic head group with two adjacent fatty acid chains on their tail. Phospholipids are disposed to undergo hydrolysis-oxidation reactions due to several factors: (1) small liposome size and low concentration; (2) the presence of oxidant agents (chemical oxidants and enzymatic oxidants); and (3) modification on fatty acyl chain. The literature also stated that liposome size is an essential parameter for a stable liposome, whilst phospholipids' starting material could cause free radical oxidation.²⁰⁵

Accordingly, some characteristics are modified to improve liposomal formulations for gene delivery application and are pertinent to antipsoriatic activity. The recent shift towards cationic liposomes by adding cationic lipids on the surface of liposomes to form stable complexes has gained a lot of interest for select nucleic acid delivery.^{206,207} In this regard, liposomes provide some protection to the genes from degradation reactions. The cationic liposomal structures will get entrapped to the large negatively charged DNA fragments, possibly about chromosome size, and, most importantly, can target specific cells or tissues.²⁰⁰

5.3.2 Niosomes. Niosomes are the best alternative to liposomes. They are developed to overcome the low physical stability and expensive production of liposomes.^{208,209} Niosomes obtained the name because they consist of nonionic surfactants, thus are less toxic to the human's body since they carry no charge.^{208,210} However, the earliest reported use of niosomes (known as synthetic liposomes) was in cosmetical application, coined by cosmetic giant L'Oréal in the 1970s and 1980s,^{211,212} before being explored in pharmaceutical applications. Since they are made up of nonionic surfactants, they are less haemolytic and disrupt the cellular surfaces, thus, exhibiting



biocompatible properties.^{213–215} The formation of niosomes is usually with or without cholesterol, in which the latter affords the membrane rigidity and stability due to its charge presence.^{216,217}

Different preparation methods of niosomes have been reported including ether injection method, thin-film hydration method, emulsion method, reverse phase evaporation method, and micro fluidisation method, which each could yield different vesicles sizes.^{209,218,219} It is signified that a favourable niosome of amphiphilic bilayers for high entrapment efficiency can be produced by taking into consideration of the hydrophilic-lipophilic balance (HLB) value of nonionic surfactants between 4 and 8.^{218,220} The choice of nonionic surfactants that falls within the range is typically amongst Tween, Span, and Brij. Besides, a stable niosome formation for better drug loading can be achieved had long hydrocarbon chains used without a double bond.²²⁰ It is also ubiquitous for nonionic surfactants with a high HLB value, notably Tween 20, Tween 60, and Tween 80 to integrate cholesterol for stabilisation due to their imbalance hefty hydrophilic heads and small hydrophobic tails.²²⁰

In general, niosomes exhibit similar physicochemical properties, drug delivery studies, and *in vivo* behaviour with liposomes.^{210,221} Even so, they have different structural composition bilayer as niosomes exhibit higher chemical, physical, and biological stabilities of the vesicles.²²¹ As a result, niosomal formulations may offer several advantages over liposomes in respect of higher drug entrapment and cheaper industrial manufacturing.^{209,210,221} However, this interesting nanovesicle is relatively new and young in pharmaceutical applications compared to liposomes.²²¹ The lack of generally recognised as safe (GRAS) components in niosomes (like phospholipids in liposomes) might be one of their drawbacks,²¹² whilst the aforementioned commercial surfactants (Tween and Span) reportedly exhibit polydispersity feature, which is held as another shortcoming from niosomes.²¹⁰

5.3.3 Transfersomes. Transfersomes (also known as ultra-deformable liposomes or elastic liposomes),^{222,223} was designed by Cevc and Blume in 1992.^{224,225} The main components of transfersomes are phospholipids (similar to liposomes) and edge activators (EA), a single chain surfactant that is also a membrane-softening agent (*e.g.*, Tween 80, Span 80, and sodium cholate).^{224,226} Novel approaches have been introduced to provide better deformability, but there is no general recipe in formulating the vesicles since their medium suspending composition must be tailored individually to each type of drug and the specific payload.^{222,224} Besides, the high bilayer adaptability and vesicle stability are the two vital factors in the process of forming transfersomes.²²⁴

In essence, transfersomes are more deformable and elastic to efficiently penetrate through the small channels in the skin compared to conventional liposomes.^{226,227} As a result of their self-optimising deformability, the membrane changing flexibility upon reaching and passing through the skin pores can be attained spontaneously as they open up extracellular pathways of the cells.²²⁷ EA acts as a permeation enhancer to enable transdermal penetration by destabilising lipid bilayers of the vesicles while enhancing the fluidity of stratum corneum

lipid.^{163,225} Thus, transdermal delivery can be enhanced by avoiding the first-pass effect and be controlled and prolonged the duration of the drug activity. These pharmacokinetic effects allow the efficiency of short half-life drugs and increase the physiological and pharmacological response.²²⁸ Amongst the common preparation methods of transfersomes are either thin-film hydration, reverse-phase evaporation, ethanol injection, or freeze-thaw method.^{226,229} It is generally regarded that transfersomes are superior to liposomes in terms of skin penetration^{226,227,230} but relatively inferior to ethosomes in terms of deformability.²³¹ However, transfersomes and ethosomes have equivalent skin deposition ability had ethosomes not incorporated another type of alcohol as will be discussed in the next subsection.²³²

5.3.4 Ethosomes. Ethosomes were developed by Tuitou *et al.* in 1996, acting as another enhanced delivery of active agents.^{233–235} Ethosomes are composed of phospholipids (similar to liposomes) and ethanol, in which ethanol plays an essential role in efficient topical drug delivery.^{236,237} Ethosomes have a better penetration rate through the skin, probably due to their high ethanolic content (20–50%). The soft and malleable vesicles are obtained as a result of the coexistence of both phospholipids and ethanol as proved by Tuitou *et al.*^{238,239} Similar to transfersomes, therapeutic delivery can be administered *via* transdermal route by ethosomal systems.²⁴⁰ Thus, the vesicles have a higher therapeutic efficacy over liposomes in terms of the greater permeation of the drug's ability as they can squeeze themselves through the skin pores as established by several studies.^{241–243} A study by Yang *et al.* showed that ethosomal formulation had a higher quantity and depth of lipophilic drugs to be delivered across the skin (transdermal delivery) for a longer period when compared to liposomes due to their interdependent composition.²⁴⁴

The proposed mechanism of ethosome transportation across the skin is well-described by the literature; however, the exact method of how they act as vehicles is not fully understood.²³⁷ The underlying mechanism is upon ethanol's polar head group interaction with the lipid regions of the skin to allow the increase of cell membrane lipid fluidity, which results in improved skin penetrability of the ethosomes.²³⁷ These ethosomes then permeate the skin, merge with cell membrane lipids, and release the system's drug.²⁴⁵ A recent investigation by Niu and co-workers reported that the ethanol solution influenced the swelling of the stratum corneum and the increase in the size of keratinocytes, except that the ethosomes were not responsible for the results. The findings also concluded that the ethosomes had better transdermal permeation effects since some of the ethosomes fused to the upper stratum corneum whilst some others travelled and released the drug to the intercellular space during their penetration into the deeper skin layers.²⁴⁶

Amongst popular ethosomal formulations are the classical cold method, the hot method, ethanol injection-sonication, thin-film hydration, and reverse phase-evaporation methods.²³³ Ethosomal systems can be further classified into their composition as (1) classical ethosomes; (2) binary ethosomes (added with another type of alcohol like isopropyl



alcohol); and (3) transethosomes (added with a chemical surfactant or edge activator).²³³ Several studies published comparative studies between these different types of ethosomes and transfersomes. For instance, upon the comparison between classical ethosomes, binary ethosomes, and transfersomes, it was reported that binary ethosomes have the highest penetration depth and fluorescence intensity but both types of ethosomes have lesser skin deposition than transfersomes.²³¹ Similarly, Zhang *et al.* concluded that the significant amount of ethanol had influenced the transdermal drug delivery rather than the skin deposition (topical delivery) of ethosomes. Hence, the ethosomal formulations could be adjusted with propylene glycol to prolong the movement of the vesicles and accumulate in the skin.²⁴¹ On top of that, transethosomes showed the highest drug's incorporation efficiency as opposed to ethosomes and transfersomes, and as such, were more deformable than the two because of the presence of both ethanol and surfactant compositions.²³²

5.4 Nanoemulsions

Nanoemulsions are isotropic, heterogeneous system, consisting of two immiscible liquids of a sufficient drug's dispersion in nanodroplet sizes.²⁴⁷ The two liquids usually are either water in oil (W/O) or oil in water (O/W) or double emulsion (W/O/W and O/W/O)²⁴⁸ stabilised by amphiphilic surfactants, with mean droplets of <200 nm depending on the phase media.²⁴⁹ The systems have been proven to virtually encapsulate lipophilic active compounds for enhanced skin delivery topically.²⁵⁰ Besides, nanoemulsions are also excellent vehicles to encapsulate essential oils and natural bioactive compounds as stated in the literature and proven in recent studies.^{251–254} The emulsion technique is versatile, and as such, it can be rendered in spray, gel, cream, and aerosol. Several other reasons also lead to the overwhelming number of research that utilises nanoemulsion methods as drug delivery, such as they are feasible, non-invasive, and cost-effective, especially for high metabolic active drugs.^{255,256}

The choice of surfactants is vital in reducing the surface and interfacial tension between oil and water, which can either be ionic or nonionic.²⁴⁹ Amongst common surfactants or emulsifying agents are Tween® 80, amphiphilic proteins (caseinate), phospholipids (soy lecithin), polysaccharides, and polymers (PEG).²⁵⁷ Generally, nonionic surfactants are regarded as safer than anionic surfactants because they have less local irritation, which is suitable for pharmaceutical applications.²⁵⁸ In addition, they also have a lower CMC value than their ionic counterparts of the same alkyl chain length for a more stable drug delivery system.^{259,260} Concurrently, a co-surfactant is often needed whereby alcohol groups with C3–C8 chain length like ethylene glycol, propylene glycol, ethanol, and propanol are added to enhance the entropy and stabilise the colloidal system.²⁴⁷ It is interesting to note that, nanoemulsion structure has the closest resemblance of a micelle, out of other lipid nanocarriers, except that the innermost part of nanoemulsions is the oil phase that solubilises hydrophobic drugs. In choosing the oils, different types of triglycerides from long-chain

triglycerides (LCT), medium-chain triglycerides (MCT), and short-chain triglycerides (SCT) with potent therapeutic benefits have been utilised as excipients and act as determinants in the bioavailability of the encapsulated API.²⁶¹ Nevertheless, it is assumed that a more stable O/W formulation could be achieved through a less polar oil base.²⁶²

The preparation methods of nanoemulsions can generally be divided into high energy and low energy methods.²⁶³ The high-energy method, such as high-pressure homogenisation, produces a robust way of synthesising nanoemulsions by applying excess shear or mechanical device, to break large droplets into nanosized particles.^{263,264} However, the high-energy method merely produces 1% nanoemulsion whilst 99% of its heat is dissipated off, thus, a repeated series of cycles is required, which is more applicable in large scale production.²⁶⁵ On the other hand, two different types of low-energy methods, namely phase inversion composition (PIC) and phase inversion temperature (PIT) are known to be energy-saving since nanoemulsions are produced by exploiting the system's low interfacial property.^{264,266,267} The PIC and PIT methods have gained special attention lately and become more favourable methods in which temperature and composition are kept constant, respectively.²⁶⁶ Given the requirements of the low-energy methods, composition variables act as determinants in producing nanodroplets rather than the preparation variables as in the high-energy method.²⁶⁷

Nanoemulsions are non-equilibrium systems with thermodynamically unstable and kinetically stable systems that are sensitive to environmental parameters like pH and temperature.^{268,269} Consequently, they have a propensity to undergo flocculation, coalescence, and/or Ostwald ripening phenomena that are associated with unstable emulsions.^{270,271} However, with appropriate optimisation in the selection of their components as well as during formulating preparation, a stable nanoemulsion system can be achieved for drug delivery.

6 Future prospects of lipid-based nanoparticles in pharmaceutical applications

In line with nanotechnology evolution, we witness an overwhelming number of nanotechnology uses in pharmaceutical applications. As a result, the utilisation of nanomaterials into nanotechnology-based drug delivery systems is futuristic, and the market value is projected to skyrocket six-times fold for approximately US\$334 billion by 2025 compared to that of over a decade ago.²⁹⁸ Nanomedicine will witness rigorous research progress within the next 20 years as more focus on materials design is driven by clinical needs.²⁹⁹ Therefore, a new paradigm of psoriasis treatments is moving towards this novel therapeutic platform.^{300,301}

Interestingly, lipid-based nanoparticles have garnered special attention in psoriasis treatments because of their adaptability and feasibility approach. Each different type of lipid-based nanoparticle is distinctive since their lipid components as surfactants can be tailored per suitability. This leads to



a further novel approach as researchers can broaden their horizon in selecting the lipid components as the starting materials for more natural and greener products, such as cardanol, sugar-based surfactants, glycolipids, and bio-surfactants.^{302–306} However, the Cpp of the lipid surfactants must be acknowledged since the morphology prediction would determine the success of preparing any lipid-based nanoparticles. For example, non-ionic lipid surfactants that have $(1/2 < \text{Cpp} < 1)$ would form vesicles (niosomes) in an aqueous medium but other types of non-ionic lipid surfactants with $(1/3 < \text{Cpp} < 1/2)$ would form cylindrical micelles (nano-emulsions),^{171–173} as discussed in Section 5.0.

Moreover, the option of encapsulating the API does not limit to a certain type of lipid-based nanoparticles based on the compilation in Table 1. In this regard, we can see the developments of the same API *via* different nanocarriers such as methotrexate was successfully encapsulated into SLNs,²⁷² NLCs,^{276,279} transfersomes,²⁹⁰ ethosomes,²⁹¹ and nano-emulsions,²⁹⁵ with significant therapeutic profiles. Furthermore, these different types of lipid-based nanoparticles have successfully encapsulated several different antipsoriatic agents or API. For example, liposomes have encapsulated cyclosporine,^{280,282} psoralens,²⁸¹ zedoary turmeric oil and tretinoin,²⁸³ *trans*-retinoic acid and betamethasone,²⁸⁴ L-SNA,²⁸⁵ and bexarotene.²⁸⁶ Therefore, these discriminations demonstrate that lipid-based nanoparticles are versatile and highly promising drug carriers for antipsoriatic activity. They also have a high potential for clinical studies and thus, to be developed in pharmaceuticals.

An online platform, <http://www.ClinicalTrials.gov>, serves as the clinical trials database that gathers worldwide research and any clinical studies should be reported there to avoid evidence distortion for clinical practice.^{307,308} However, based on the database, there is no psoriasis nano-drug yet to be approved for use after clinical studies. The database also shows that there is only one psoriasis nano-drug that is undergoing clinical trial. The study (Identifier: NCT03348462) successfully developed two different anthralin nanoformulations encapsulated into liposomes and ethosomes, which has already completed phase 4 clinical trial and showed higher potent effects through ethosomal systems.²⁹²

In the pharmaceutical industry, it was reported that commercialised nano drugs are currently at the first generation of phase II clinical trials carried out by small and medium-sized enterprises (SMEs) before entering the market.²⁹⁸ The tedious procedures of clinical trials and financial constraints have hindered the current progress whilst big pharmaceutical companies are only interested in acquiring these SMEs once these emerging drugs get approved by the FDA.²⁹⁸ Some other challenges to realise in pharmaceuticals include that there is a need for further nanoparticle characterisation profile for safety and toxicity-related issues, as well as a lack of nanotechnology-regulated guidelines for the industry.^{309,310} It is also noteworthy to mention that current nanomedicine investigation on nano drugs is limited to the evaluation of *in vitro* and *in vivo* studies.³¹¹ Lastly, the interactions between the tissues and lipid-based nanoparticles should also be deeply

explored since the drug delivery systems may exhibit intricate behaviour in the human's body.³¹² For example, nanoparticles may be deposited in the internal organs and thus interact with other immune cells, which could develop other immune reactions.³¹ However, the bottom-up approach through the self-assembled lipid nanoparticles could improve the safety measures.³¹ Besides, the application of state-of-the-art technologies namely artificial intelligence (AI) and machine learning could facilitate the nanotherapeutic delivery *via* stimulation and modelling processes by understanding both biochemical and physicochemical properties of the nano drugs.³¹³ Therefore, profound knowledge on the cytotoxicity and biological activities of nano-drugs are vital to be further investigated inclusively to providing coherent knowledge on their short-term and long-term effects before being pursued in clinical studies.^{31,311,312}

7 Conclusion

Psoriasis is a skin disease that is affected by several known factors including environmental and genetic, causing cell hyperproliferation. Psoriasis treatments provide a wide array of treatment options, namely topical, systemic nonbiologic, systemic biologics, and phototherapy. Despite the many antipsoriatic drugs with different mechanisms of action, topical treatment serves as the most convenient way to transport the therapeutics across the skin barrier. Lipid-based nanoparticles are promising novel nano delivery systems to improve the API delivery to the target site. Recent works show that different APIs can be encapsulated into different lipid nanocarriers but liposomes and nanoemulsions have the highest number of successful nano-based drugs formulated for psoriasis treatments. Enhanced skin permeation and alleviated psoriasis symptoms are the improvements of antipsoriatic agents in tandem with lipid-based nanoparticles. Besides, the therapeutics also show highly favourable outcomes in alleviating psoriasis lesions as they enhance skin penetration, retention, and prolonged release due to their large surface area at nanoscale size. However, further research and opportunities with other types of lipid-based nanocarriers like niosomes, transfersomes, and ethosomes are needed to explore their tremendous potential in the antipsoriatic application's topical delivery. In general, the production cost of nano drugs is expensive in the pharmaceutical industry. The development of nano drugs is also associated with cytotoxicity and safety issues but could be overcome by an in-depth understanding of their interactions in the human's body. Based on the compilation of the recent works of the API with lipid-based nanocarriers, it is also observed that they exhibit higher drug penetration and bioavailability compared to that of the control variable (conventional treatments without nanocarrier). Therefore, the production of nano drugs is guaranteed success if optimised strategies are taken in nanomedicine applications.

Author contributions

Conceptualization, U. U. M. N., N. A., N. S. and N. S. M. Y.; methodology, U. U. M. N.; writing – original draft preparation,



Review

U. U. M. N.; writing – review & editing, N. A., N. S. and N. S. M. Y.; visualization, U. U. M. N.; supervision, N. A. and N. S.; project administration, N. A. and N. S.; funding acquisition, N. A.; all authors have read and agreed to the published version of the manuscript.

Conflicts of interest

There are no conflicts to declare.

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References

- J. Takeshita, S. Grewal, S. M. Langan, N. N. Mehta, A. Ogdie, A. S. Van Voorhees and J. M. Gelfand, *J. Am. Acad. Dermatol.*, 2017, **76**, 377–390.
- R. Parisi, I. Y. Iskandar, E. Kontopantelis, M. Augustin, C. E. Griffiths and D. M. Ashcroft, *Br. Med. J.*, 2020, **369**, m1590.
- A. W. Armstrong and C. Read, *J. Am. Med. Assoc.*, 2020, **323**, 1945–1960.
- B. Elewski, A. F. Alexis, M. Lebwohl, L. S. Gold, D. Pariser, J. D. Rosso and G. Yosipovitch, *J. Eur. Acad. Dermatol. Venereol.*, 2019, **33**, 1465–1476.
- Z. Yiu, R. Warren and A. Menter, in *Handbook of Psoriasis and Psoriatic Arthritis*, ed. R. Warren and A. Menter, Adis, Cham, 2016.
- S. K. Ippagunta, R. Gangwar, D. Finkelstein, P. Vogel, S. Pelletier, S. Gingras, V. Redecke and H. Häcker, *Proc. Natl. Acad. Sci. U.S.A.*, 2016, **113**, E6162–E6171.
- W. Masson, M. Lobo and G. Molinero, *Adv. Ther.*, 2020, **37**, 2017.
- A. Rendon and K. Schäkel, *Int. J. Mol. Sci.*, 2019, **20**, 1475.
- J. M. Fernández-Armenteros, X. Gómez-Arbonés, M. Buti-Solé, A. Betriu-Bars, V. Sanmartín-Novell, M. Ortega-Bravo, M. Martínez-Alonso and J. M. Casanova-Seuma, *Actas Dermo-Sifiliográficas*, 2019, **110**, 385–392.
- E. Ogawa, Y. Sato, A. Minagawa and R. Okuyama, *J. Dermatol.*, 2018, **45**, 264–272.
- C.-H. Lin, S.-H. Chang and J.-Y. Fang, *Curr. Drug Ther.*, 2016, **11**, 21–40.
- R. Parisi, D. P. Symmons, C. E. Griffiths and D. M. Ashcroft, *J. Invest. Dermatol.*, 2013, **133**, 377–385.
- M. Rademaker, K. Agnew, N. Anagnostou, M. Andrews, K. Armour, C. Baker, P. Foley, K. Gebauer, M. Gupta and G. Marshman, *Australas. J. Dermatol.*, 2019, **60**, 91–98.
- A. Menter, K. M. Cordoro, D. M. R. Davis, D. Kroshinsky, A. S. Paller, A. W. Armstrong, C. Connor, B. E. Elewski, J. M. Gelfand, K. B. Gordon, A. B. Gottlieb, D. H. Kaplan, A. Kavanaugh, M. Kiselica, D. Kivelevitch, N. J. Korman, M. Lebwohl, C. L. Leonardi, J. Lichten, H. W. Lim, N. N. Mehta, S. L. Parra, A. L. Pathy, E. A. Farley Prater, R. N. Rupani, M. Siegel, B. Stoff, B. E. Strober, E. B. Wong, J. J. Wu, V. Hariharan and C. A. Elmets, *J. Am. Acad. Dermatol.*, 2020, **82**, 161–201.
- D. M. Balak and E. Hajdarbegovic, *Psoriasis*, 2017, **7**, 87–94.
- N. J. Korman, *Br. J. Dermatol.*, 2020, **182**, 840–848.
- S. González-Parra and E. Daudén, *Actas Dermo-Sifiliográficas*, 2019, **110**, 12–19.
- F. M. Sahi, A. Masood, N. A. Danawar, A. Mekaiel and B. H. Malik, *Cureus*, 2020, **12**(8), e9708.
- C. Fink, C. Alt, L. Uhlmann, C. Klose, A. Enk and H. A. Haenssle, *Br. J. Dermatol.*, 2019, **180**, 390–396.
- D. Hägg, A. Sundström, M. Eriksson and M. Schmitt-Egenolf, *Am. J. Clin. Dermatol.*, 2017, **18**, 583–590.
- V. K. Rapalli, G. Singhvi, S. K. Dubey, G. Gupta, D. K. Chellappan and K. Dua, *Biomed. Pharmacother.*, 2018, **106**, 707–713.
- G. Singhvi, S. Hejmady, V. K. Rapalli, S. K. Dubey and S. Dubey, in *Delivery of Drugs*, Elsevier, 2020, pp. 75–96.
- M. Bhat, S. Pukale, S. Singh, A. Mittal and D. Chitkara, *J. Drug Deliv. Sci. Technol.*, 2021, **62**, 102328.
- A. W. Armstrong, L. Puig, A. Joshi, M. Skup, D. Williams, J. Li, K. A. Betts and M. Augustin, *JAMA Dermatol.*, 2020, **156**, 258–269.
- P. Rider, Y. Carmi and I. Cohen, *Int. J. Cell Biol.*, 2016, **2016**, 9259646.
- A. Menter, B. E. Strober, D. H. Kaplan, D. Kivelevitch, E. F. Prater, B. Stoff, A. W. Armstrong, C. Connor, K. M. Cordoro, D. M. R. Davis, B. E. Elewski, J. M. Gelfand, K. B. Gordon, A. B. Gottlieb, A. Kavanaugh, M. Kiselica, N. J. Korman, D. Kroshinsky, M. Lebwohl, C. L. Leonardi, J. Lichten, H. W. Lim, N. N. Mehta, A. S. Paller, S. L. Parra, A. L. Pathy, R. N. Rupani, M. Siegel, E. B. Wong, J. J. Wu, V. Hariharan and C. A. Elmets, *J. Am. Acad. Dermatol.*, 2019, **80**, 1029–1072.
- M. Pradhan, A. Alexander, M. R. Singh, D. Singh, S. Saraf and S. Saraf, *Biomed. Pharmacother.*, 2018, **107**, 447–463.
- A. R. Fernandes, C. Martins-Gomes, A. Santini, A. M. Silva and E. B. Souto, in *Design of Nanostructures for Versatile Therapeutic Applications*, ed. A. M. Grumezescu, William Andrew Publishing, 2018, pp. 379–406.
- M. R. Gigliobianco, C. Casadidio, R. Censi and P. Di Martino, *Pharmaceutics*, 2018, **10**, 134.
- W. Kaialy and M. Al Shafiee, *Adv. Colloid Interface Sci.*, 2016, **228**, 71–91.
- W. Lu, J. Yao, X. Zhu and Y. Qi, *Biomed. Pharmacother.*, 2021, **134**, 111103.
- X. Zhang, Z. Li, J. Gao, Z. Wang, X. Gao, N. Liu, M. Li, H. Zhang and A. Zheng, *Molecules*, 2020, **25**, 1080.
- M. Swierczewska, R. M. Crist and S. E. McNeil, in *Characterization of Nanoparticles Intended for Drug Delivery*, Humana Press, New York, NY, 2018, pp. 3–16.
- S. L. Jyothi, K. L. Krishna, V. K. Ameena Shirin, R. Sankar, K. Pramod and H. V. Gangadharappa, *J. Drug Deliv. Sci. Technol.*, 2021, **62**, 102364.
- N. Vincent, D. D. Ramya and H. B. Vedha, *Dermatol. Rep.*, 2014, **6**, 5451.



- 36 T. W. Prow, J. E. Grice, L. L. Lin, R. Faye, M. Butler, W. Becker, E. M. T. Wurm, C. Yoong, T. A. Robertson, H. P. Soyer and M. S. Roberts, *Adv. Drug Deliv. Rev.*, 2011, **63**, 470–491.
- 37 D. Medina-Cruz, B. Saleh, A. Vernet-Crua, A. Ajo, A. K. Roy and T. J. Webster, in *Wound Healing, Tissue Repair, and Regeneration in Diabetes*, ed. D. Bagchi, A. Das and S. Roy, Academic Press, 2020, pp. 439–488.
- 38 C. Su, Y. Liu, R. Li, W. Wu, J. P. Fawcett and J. Gu, *Adv. Drug Deliv. Rev.*, 2019, **143**, 97–114.
- 39 T. Ramezanli, B. E. Kilfoyle, Z. Zhang and B. B. Michniak-Kohn, *Int. J. Pharm.*, 2017, **516**, 196–203.
- 40 A. L. Savian, D. Rodrigues, J. Weber, R. F. Ribeiro, M. H. Motta, S. R. Schaffazick, A. I. H. Adams, D. F. de Andrade, R. C. R. Beck and C. B. da Silva, *Mater. Sci. Eng. C*, 2015, **46**, 69–76.
- 41 P. K. Tripathi, B. Gorain, H. Choudhury, A. Srivastava and P. Kesharwani, *Heliyon*, 2019, **5**, e01343.
- 42 M. Lapteva, V. Santer, K. Mondon, I. Patmanidis, G. Chiriano, L. Scapozza, R. Gurny, M. Möller and Y. N. Kalia, *J. Control. Release*, 2014, **196**, 9–18.
- 43 K.-L. Mao, Z.-L. Fan, J.-D. Yuan, P.-P. Chen, J.-J. Yang, J. Xu, D.-L. ZhuGe, B.-H. Jin, Q.-Y. Zhu, B.-X. Shen, Y. Sohawon, Y.-Z. Zhao and H.-L. Xu, *Colloids Surf. B Biointerfaces*, 2017, **160**, 704–714.
- 44 S. A. Fereig, G. M. El-Zaafarany, M. G. Arafa and M. M. A. Abdel-Mottaleb, *Carbohydr. Polym.*, 2021, **268**, 118238.
- 45 K. Yadav, A. Soni, D. Singh and M. R. Singh, *Prog. Biomater.*, 2021, **10**, 1–17.
- 46 S. Sur, A. Rathore, V. Dave, K. R. Reddy, R. S. Chouhan and V. Sadhu, *Nano-Struct. Nano-Objects*, 2019, **20**, 100397.
- 47 M. T. C. P. Bernardes, S. B. N. Agostini, G. R. Pereira, L. P. da Silva, J. B. da Silva, M. L. Bruschi, R. D. Novaes and F. C. Carvalho, *Eur. J. Pharm. Sci.*, 2021, **165**, 105956.
- 48 D. Limón, K. Talló Domínguez, M. L. Garduño-Ramírez, B. Andrade, A. C. Calpena and L. Pérez-García, *Colloids Surf. B Biointerfaces*, 2019, **181**, 657–670.
- 49 Q. Li, F. Li, X. Qi, F. Wei, H. Chen and T. Wang, *Colloids Surf. B Biointerfaces*, 2020, **195**, 111246.
- 50 S. Kumar, M. Prasad and R. Rao, *Mater. Sci. Eng. C*, 2021, **119**, 111605.
- 51 S. Meng, L. Sun, L. Wang, Z. Lin, Z. Liu, L. Xi, Z. Wang and Y. Zheng, *Colloids Surf. B Biointerfaces*, 2019, **182**, 110352.
- 52 S. Jacob, A. B. Nair, J. Shah, N. Sreeharsha, S. Gupta and P. Shinu, *Pharmaceutics*, 2021, **13**, 357.
- 53 X. Li, J. F. Lovell, J. Yoon and X. Chen, *Nat. Rev. Clin. Oncol.*, 2020, **17**, 657–674.
- 54 G. R. Nirmal, Z.-C. Lin, M.-J. Tsai, S.-C. Yang, A. Alalaiwe and J.-Y. Fang, *J. Controlled Release*, 2021, **333**, 487–499.
- 55 J. Chen, Z. Zeng, L. Huang, S. Luo, J. Dong, F. H. Zhou, K. Zhou, L. Wang and L. Kang, *Am. J. Transl. Res.*, 2020, **12**, 3089–3115.
- 56 H.-Q. Liu, Y.-M. Wang, W.-F. Li, C. Li, Z.-H. Jiang, J. Bao, J.-F. Wei, H.-T. Jin and A.-P. Wang, *Cell. Physiol. Biochem.*, 2017, **44**, 200–214.
- 57 P. R. Ebani, L. Stefanello, B. Luiza Kuhn, C. Piccinin Frizzo, T. Augusto Lima Burgo, C. Luisa Kloster and M. Antonio Villetti, *J. Mol. Liq.*, 2021, **336**, 116874.
- 58 R. Kumar, in *Nanocarriers for Drug Delivery*, ed. S. S. Mohapatra, S. Ranjan, N. Dasgupta, R. K. Mishra and S. Thomas, Elsevier, 2019, pp. 249–284.
- 59 M. Pradhan, D. Singh and M. R. Singh, *J. Controlled Release*, 2013, **170**, 380–395.
- 60 M. Gupta, U. Agrawal and S. P. Vyas, *Expet Opin. Drug Deliv.*, 2012, **9**, 783–804.
- 61 B. García-Pinel, C. Porras-Alcalá, A. Ortega-Rodríguez, F. Sarabia, J. Prados, C. Melguizo and J. M. López-Romero, *Nanomaterials*, 2019, **9**, 638.
- 62 P. Ghasemiyeh and S. Mohammadi-Samani, *Res. Pharm. Sci.*, 2018, **13**, 288–303.
- 63 S.-Y. Chuang, C.-H. Lin, T.-H. Huang and J.-Y. Fang, *Nanomaterials*, 2018, **8**, 42.
- 64 C. Albanesi, in *Clinical Immunology*, Elsevier, 2019, pp. 871–878. e1.
- 65 B. Lukasch, H. Westerdahl, M. Strandh, H. Winkler, Y. Moodley, F. Knauer and H. Hoi, *PeerJ*, 2017, **5**, e3679.
- 66 M. Wiczorek, E. T. Abualrous, J. Sticht, M. Álvaro-Benito, S. Stolzenberg, F. Noé and C. Freund, *Front. Immunol.*, 2017, **8**, 292.
- 67 V. Kumar, A. K. Abbas and J. C. Aster, *Robbins Basic Pathology*, Elsevier, Philadelphia, PA, 10th edn, 2018.
- 68 J. B. Zabriskie, *Essential clinical immunology*, Cambridge University Press, 2009.
- 69 M. P. Schön, *Front. Immunol.*, 2019, **10**, 1764.
- 70 J. M. Hugh and J. M. Weinberg, *Cutis*, 2018, **102**, 6–12.
- 71 H. R. Hampton and T. Chtanova, *Front. Immunol.*, 2019, **10**, 1168.
- 72 A. Wang and Y. Bai, *J. Dermatol.*, 2020, **47**, 104–113.
- 73 C. de Jesús-Gil, L. Sans-de SanNicolàs, I. García-Jiménez, M. Ferran, A. Celada, A. Chiriac, R. M. Pujol and L. F. Santamaria-Babi, *Front. Immunol.*, 2021, **12**, 524.
- 74 V. R. Khairutdinov, A. F. Mikhailichenko, I. E. Belousova, E. Sh. Kuligina, A. V. Samtsov and E. N. Imyanitov, *An. Bras. Dermatol.*, 2017, **92**, 41–44.
- 75 B. Polese, H. Zhang, B. Thuraiajah and I. L. King, *Front. Immunol.*, 2020, **11**, 242.
- 76 B. Wawrzycki, A. Pietrzak, E. Grywalska, D. Krasowska, G. Chodorowska and J. Roliński, *Arch. Immunol. Ther. Exp.*, 2019, **67**, 103–108.
- 77 N. C. Brembilla, L. Senra and W.-H. Boehncke, *Front. Immunol.*, 2018, **9**, 1682.
- 78 M. Tampa, M.-I. Sarbu, M.-I. Mitran, C.-I. Mitran, C. Matei and S.-R. Georgescu, *Dis. Markers*, 2018, **2018**, 5823684.
- 79 A. M. Martins, A. Ascenso, H. M. Ribeiro and J. Marto, *Mol. Neurobiol.*, 2020, **57**, 2391–2419.
- 80 G. P. Thami and R. Sarkar, *Clin. Exp. Dermatol.*, 2002, **27**, 99–103.
- 81 J. A. Zeichner, *J. Clin. Aesthetic Dermatol.*, 2010, **3**, 37–40.
- 82 A. Bhatia, B. Singh, B. Amarji, P. Negi, A. Shukla and O. P. Katara, *Int. J. Dermatol.*, 2011, **50**, 1246–1248.



- 83 S. Sekhon, C. Jeon, M. Nakamura, L. Afifi, D. Yan, J. J. Wu, W. Liao and T. Bhutani, *J. Dermatol. Treat.*, 2018, **29**, 230–232.
- 84 W. R. Heymann, *J. Am. Acad. Dermatol.*, 2019, **80**, 56–57.
- 85 K. Blakely and M. Gooderham, *Psoriasis Auckl. NZ*, 2016, **6**, 33–40.
- 86 H. Myśliwiec, P. Myśliwiec, A. Baran and I. Flisiak, *Adv. Med. Sci.*, 2016, **61**, 207–211.
- 87 K. A. Hollywood, C. L. Winder, W. B. Dunn, Y. Xu, D. Broadhurst, C. E. M. Griffiths and R. Goodacre, *Mol. Biosyst.*, 2015, **11**, 2198–2209.
- 88 C. Mo, D. Shetti and K. Wei, *Molecules*, 2019, **24**, 2727.
- 89 T. Benezeder, C. Painsi, V. Patra, S. Dey, M. Holcman, B. Lange-Asschenfeldt, M. Sibilia and P. Wolf, *eLife*, 2020, **9**, e56991.
- 90 J. Holstein, B. Fehrenbacher, J. Brück, E. Müller-Hermelink, I. Schäfer, M. Carevic, B. Schitteck, M. Schaller, K. Ghoreschi and F. C. Eberle, *J. Dermatol. Sci.*, 2017, **87**, 236–245.
- 91 V. Kadian, S. Kumar, K. Saini, V. Kakkar and R. Rao, *Curr. Drug Res. Rev.*, 2020, **12**, 82–96.
- 92 M. S. Heath, D. R. Sahni, Z. A. Curry and S. R. Feldman, *Expert Opin. Drug Metab. Toxicol.*, 2018, **14**, 919–927.
- 93 İ. Erol, N. Ü. Okur, D. Orak, H. Sipahi, A. Aydın and Ö. Özer, *Pharm. Dev. Technol.*, 2020, **25**, 909–918.
- 94 E. L. Roux and H. Frow, *Prescriber*, 2020, **31**, 9–17.
- 95 E. Tanghetti, M. Lebwohl and L. Stein Gold, *J. Drugs Dermatol. JDD*, 2018, **17**, 1280–1287.
- 96 L. S. Gold, M. G. Lebwohl, J. L. Sugarman, D. M. Pariser, T. Lin, G. Martin, R. Pillai, R. Israel and T. Ramakrishna, *J. Am. Acad. Dermatol.*, 2018, **79**, 287–293.
- 97 G. Micali, A. E. Verzi, G. Giuffrida, E. Panebianco, M. L. Musumeci and F. Lacarrubba, *Clin. Cosmet. Investig. Dermatol.*, 2019, **12**, 953–959.
- 98 J. F. Merola, A. Qureshi and M. E. Husni, *Dermatol. Ther.*, 2018, **31**, e12589.
- 99 L. Guenther, C. Lynde and Y. Poulin, *J. Cutan. Med. Surg.*, 2019, **23**, 27S–34S.
- 100 S. Jindal, R. Awasthi, D. Singhare and G. T. Kulkarni, *Med. Hypotheses*, 2020, **142**, 109838.
- 101 Z. Yin, J. Xu and D. Luo, *J. Biomed. Res.*, 2011, **25**, 385–391.
- 102 X. Huang and B. Xu, *Dermatology*, 2015, **231**, 41–49.
- 103 R. Torsekar and M. M. Gautam, *Indian Dermatol. Online J.*, 2017, **8**, 235.
- 104 C. Stull, S. Grossman and G. Yosipovitch, *Am. J. Clin. Dermatol.*, 2016, **17**, 617–624.
- 105 S. Baboota, M. S. Alam, S. Sharma, J. K. Sahni, A. Kumar and J. Ali, *Int. J. Pharm. Investig.*, 2011, **1**, 139–147.
- 106 C. L. Carroll, J. Clarke, F. Camacho, R. Balkrishnan and S. R. Feldman, *Arch. Dermatol.*, 2005, **141**, 43–46.
- 107 A. Tosti, B. M. Piraccini, N. Cameli, F. Kokely, C. Plozzer, G. E. Cannata and C. Benelli, *Br. J. Dermatol.*, 1998, **139**, 655–659.
- 108 M. S. Al-Dhubaibi, *Int. J. Health Sci.*, 2018, **12**, 33.
- 109 A. Filoni, M. Vestita, M. Congedo, G. Giudice, S. Tafuri and D. Bonamonte, *Medicine*, 2018, **97**, e11185.
- 110 K. Satake, T. Amano and T. Okamoto, *Sci. Rep.*, 2019, **9**, 16322.
- 111 S. Gabros, T. A. Nessel and P. M. Zito, in *StatPearls*, StatPearls Publishing, Treasure Island (FL), 2021.
- 112 E. C. Kley, E. Morsman, L. Griffin, J. J. Wu, P. Cm van de Kerkhof, W. Gulliver, J. M. van der Walt and L. Iversen, *J. Dermatol. Treat.*, 2019, **30**, 311–319.
- 113 T. Frantz, E. G. Wright, E. A. Balogh, A. Cline, A. L. Adler-Neal and S. R. Feldman, *Children*, 2019, **6**, 125.
- 114 E. Choi, N. S. Chandran and C. Tan, *Singapore Med. J.*, 2020, **61**(3), 149.
- 115 S. Y. Song, S.-Y. Jung and E. Kim, *J. Dermatol. Treat.*, 2019, **30**, 245–250.
- 116 G. Martin, M. Young and L. Aldredge, *J. Clin. Aesthetic Dermatol.*, 2019, **12**, 13–26.
- 117 A. Menter, J. M. Gelfand, C. Connor, A. W. Armstrong, K. M. Cordoro, D. M. R. Davis, B. E. Elewski, K. B. Gordon, A. B. Gottlieb, D. H. Kaplan, A. Kavanaugh, M. Kiselica, D. Kivelevitch, N. J. Korman, D. Kroshinsky, M. Lebwohl, C. L. Leonardi, J. Lichten, H. W. Lim, N. N. Mehta, A. S. Paller, S. L. Parra, A. L. Pathy, E. F. Prater, R. S. Rahimi, R. N. Rupani, M. Siegel, B. Stoff, B. E. Strober, E. B. Tapper, E. B. Wong, J. J. Wu, V. Hariharan and C. A. Elmets, *J. Am. Acad. Dermatol.*, 2020, **82**, 1445–1486.
- 118 T. Elango, A. Thirupathi, S. Subramanian, P. Ethiraj, H. Dayalan and P. Gnanaraj, *Clin. Exp. Med.*, 2017, **17**, 371–381.
- 119 C. A. da Silva, K. Von Kossel, M. Leszczynski, T. Melnik and R. Riera, *Cochrane Database Syst. Rev.*, 2019, **2019**, CD010498.
- 120 J. J. Wu, A. Guérin, M. Sundaram, K. Dea, M. Cloutier and P. Mulani, *J. Am. Acad. Dermatol.*, 2017, **76**, 81–90.
- 121 C. W. Choi, B. R. Kim, J. Ohn and S. W. Youn, *Ann. Dermatol.*, 2017, **29**, 55–60.
- 122 E. Baskan, S. Yazici, S. Tunali and H. Saricaoglu, *J. Dermatol. Treat.*, 2016, **27**, 328–331.
- 123 V. Di Lernia, L. Stingeni, V. Boccaletti, P. G. Calzavara Pinton, C. Guarneri, A. Belloni Fortina, M. Panzone, M. Corazza, I. Neri, S. Cambiaghi, C. Lasagni, E. Ficarelli and P. Gisondi, *J. Dermatol. Treat.*, 2016, **27**, 395–398.
- 124 S. Dogra, R. Mahajan, T. Narang and S. Handa, *J. Dermatol. Treat.*, 2017, **28**, 18–20.
- 125 B. R. Kim, S. Yang, E. J. Doh, C. W. Choi and S. W. Youn, *Ann. Dermatol.*, 2018, **30**, 143–149.
- 126 C. J. Oh, K. M. Das and A. B. Gottlieb, *J. Am. Acad. Dermatol.*, 2000, **42**, 829–830.
- 127 M. Frleta, S. Siebert and I. B. McInnes, *Curr. Rheumatol. Rep.*, 2014, **16**, 414.
- 128 J. E. Hawkes, B. Y. Yan, T. C. Chan and J. G. Krueger, *J. Immunol.*, 2018, **201**, 1605–1613.
- 129 K. Godse, *Indian J. Dermatol.*, 2017, **62**, 195–199.
- 130 M. Shirley and L. J. Scott, *Drugs*, 2016, **76**, 1135–1145.
- 131 A. Blauvelt, *Expert Opin. Drug Saf.*, 2016, **15**, 1413–1420.
- 132 R. Reszke and J. C. Szepletowski, *Immunotherapy*, 2017, **9**, 229–238.
- 133 P. Mease and I. B. McInnes, *Rheumatol. Ther.*, 2016, **3**, 5–29.



- 134 A. Gottlieb, J. Sullivan, M. van Doorn, A. Kubanov, R. You, A. Parneix, S. Hugot and M. Milutinovic, *J. Am. Acad. Dermatol.*, 2017, **76**, 70–80.
- 135 A. Johar, S. Thevarajah, A. Heng, L. C. Chan, C. C. Ch'ng, N. A. Mohd Safdar, P. Muniandy, T. Taib, W. C. Tan and K. E. Tey, *Dermatol. Res. Pract.*, 2019, **2019**, 8923168.
- 136 A. Egeberg, J. P. Thyssen, J. J. Wu and L. Skov, *Br. J. Dermatol.*, 2019, **180**, 116–121.
- 137 J. J. Wu, S. R. Feldman, S. Rastogi, B. Menges, M. Lingohr-Smith and J. Lin, *J. Dermatol. Treat.*, 2018, **29**, 769–774.
- 138 M. Lebwohl, R. Rivera-Oyola and D. F. Murrell, *J. Am. Acad. Dermatol.*, 2020, **82**, 1217–1218.
- 139 P. C. M. van de Kerkhof and F. R. de Gruijl, *J. Eur. Acad. Dermatol. Venereol.*, 2020, **34**, 926–931.
- 140 K. Hyde, L. A. Cardwell, R. Stotts and S. R. Feldman, *Am. J. Pharm. Benefits*, 2018, **10**, 18–21.
- 141 C. A. Elmets, H. W. Lim, B. Stoff, C. Connor, K. M. Cordoro, M. Lebwohl, A. W. Armstrong, D. M. R. Davis, B. E. Elewski, J. M. Gelfand, K. B. Gordon, A. B. Gottlieb, D. H. Kaplan, A. Kavanaugh, M. Kiselica, D. Kivelevitch, N. J. Korman, D. Kroshinsky, C. L. Leonardi, J. Lichten, N. N. Mehta, A. S. Paller, S. L. Parra, A. L. Pathy, E. A. Farley Prater, R. N. Rupani, M. Siegel, B. E. Strober, E. B. Wong, J. J. Wu, V. Hariharan and A. Menter, *J. Am. Acad. Dermatol.*, 2019, **81**, 775–804.
- 142 J. M. Vassantachart, T. Soleymani and J. J. Wu, *J. Drugs Dermatol. JDD*, 2016, **15**, 995–1000.
- 143 V. T. Ma, C. S. Katzman, P. L. Palmbos, R. M. Patel, J. E. Gudjonsson and A. S. Alva, *Curr. Probl. Cancer Case Rep.*, 2020, **1**, 100004.
- 144 A. Ighani, A. C. R. Partridge, N. H. Shear, C. Lynde, W. P. Gulliver, C. Sibbald and P. Fleming, *J. Cutan. Med. Surg.*, 2019, **23**, 204–221.
- 145 P. Zhang and M. X. Wu, *Lasers Med. Sci.*, 2018, **33**, 173–180.
- 146 S. H. Ibbotson, *Front. Med.*, 2018, **5**, 184.
- 147 A. Morita, *J. Dermatol.*, 2018, **45**, 287–292.
- 148 E. Rácz and E. P. Prens, in *Ultraviolet Light in Human Health, Diseases and Environment. Advances in Experimental Medicine and Biology*, ed. S. Ahmad, Springer, Cham, 2017, pp. 287–294.
- 149 V. Ardeleanu, D. S. Radaschin and A. L. Tatu, *Exp. Ther. Med.*, 2020, **20**, 52–55.
- 150 M. Nakamura, B. Farahnik and T. Bhutani, *F1000Res.*, 2016, **5**, 1684.
- 151 S. Das, *Int. J. Res. Rev.*, 2019, **6**, 258–263.
- 152 B. Farahnik, M. Nakamura, R. K. Singh, M. Abrouk, T. H. Zhu, K. M. Lee, M. V. Jose, R. DaLovichio, J. Koo, T. Bhutani and W. Liao, *Dermatol. Ther.*, 2016, **6**, 315–324.
- 153 Y. Ren, X. Song, L. Tan, C. Guo, M. Wang, H. Liu, Z. Cao, Y. Li and C. Peng, *Front. Pharmacol.*, 2020, **11**, 571535.
- 154 J. Koo and M. Nakamura, in *Clinical Cases in Phototherapy. Clinical Cases in Dermatology*, Springer, Cham, 2017, pp. 13–31.
- 155 E. S. Chambers and M. Vukmanovic-Stejic, *Immunology*, 2020, **160**, 116–125.
- 156 S. J. van Rensburg, A. Franken and J. L. D. Plessis, *Skin Res. Technol.*, 2019, **25**, 595–605.
- 157 J. D. Bos and M. M. H. M. Meinardi, *Exp. Dermatol.*, 2000, **9**, 165–169.
- 158 M. B. Murphrey, J. H. Miao and P. M. Zito, in *StatPearls*, StatPearls Publishing, Treasure Island (FL), 2020.
- 159 D. J. Moore and A. V. Rawlings, *Int. J. Cosmet. Sci.*, 2017, **39**, 366–372.
- 160 K. Vávrová, A. Kováčik and L. Opálka, *Eur. Pharm. J.*, 2017, **64**, 28–35.
- 161 Y. Badhe, R. Gupta and B. Rai, *J. Mol. Model.*, 2019, **25**, 140.
- 162 F. Arriagada and J. Morales, *Curr. Pharm. Des.*, 2019, **25**, 455–466.
- 163 T. Jiang, T. Wang, T. Li, Y. Ma, S. Shen, B. He and R. Mo, *ACS Nano*, 2018, **12**, 9693–9701.
- 164 N. Tiwari, A. S. Sonzogni and M. Calderón, *Nanomed*, 2019, **14**, 2891–2895.
- 165 X. Zhou, Y. Hao, L. Yuan, S. Pradhan, K. Shrestha, O. Pradhan, H. Liu and W. Li, *Chin. Chem. Lett.*, 2018, **29**, 1713–1724.
- 166 T. Sych, Y. Mély and W. Römer, *Philos. Trans. R. Soc. Lond. B Biol. Sci.*, 2018, **373**, 20170117.
- 167 W. Lu, in *Encyclopedia of Nanotechnology*, ed. B. Bhushan, Springer Netherlands, Dordrecht, 2012, pp. 2371–2382.
- 168 H. Su, F. Wang, W. Ran, W. Zhang, W. Dai, H. Wang, C. F. Anderson, Z. Wang, C. Zheng, P. Zhang, Y. Li and H. Cui, *Proc. Natl. Acad. Sci. U.S.A.*, 2020, **117**, 4518–4526.
- 169 S. H. Crayton, A. K. Chen, J. F. Liu, E. M. Higbee-Dempsey, C.-H. Huang, A. Tsourkas and Z. Cheng, in *Comprehensive Biomaterials II*, ed. P. Ducheyne, Elsevier, Oxford, 2017, pp. 424–466.
- 170 S. Rana, J. Bhattacharjee, K. C. Barick, G. Verma, P. A. Hassan and J. V. Yakhmi, in *Nanostructures for Cancer Therapy*, ed. A. Ficaí and A. M. Grumezescu, Elsevier, 2017, pp. 177–209.
- 171 R. A. Khalil and A. A. Zarari, *Appl. Surf. Sci.*, 2014, **318**, 85–89.
- 172 D. Lombardo, M. A. Kiselev, S. Magazù and P. Calandra, *Adv. Condens. Matter Phys.*, 2015, **2015**, e151683.
- 173 J. N. Israelachvili, D. J. Mitchell and B. W. Ninham, *J. Chem. Soc., Faraday Trans. 2*, 1976, **72**, 1525–1568.
- 174 D. Lombardo, P. Calandra, L. Pasqua and S. Magazù, *Materials*, 2020, **13**, 1048.
- 175 P. Iqbal, J. A. Preece and P. M. Mendes, in *Supramolecular Chemistry*, American Cancer Society, 2012.
- 176 Z. Huang, W. Song and X. Chen, *Front. Chem.*, 2020, **8**, 380.
- 177 A. Sikder and S. Ghosh, *Mater. Chem. Front.*, 2019, **3**, 2602–2616.
- 178 L. T. Varma, N. Singh, B. Gorain, H. Choudhury, M. M. Tambuwala, P. Kesharwani and R. Shukla, *Curr. Drug Deliv.*, 2020, **17**, 279–291.
- 179 A. Walduck, P. Sangwan, Q. Anh Vo, J. Ratcliffe, J. White, B. W. Muir and N. Tran, *RSC Adv.*, 2020, **10**, 33608–33619.
- 180 R. H. Müller, K. Mäder and S. Gohla, *Eur. J. Pharm. Biopharm.*, 2000, **50**, 161–177.
- 181 N. Naseri, H. Valizadeh and P. Zakeri-Milani, *Adv. Pharm. Bull.*, 2015, **5**, 305–313.
- 182 W. Mehnert and K. Mäder, *Adv. Drug Deliv. Rev.*, 2012, **64**, 83–101.



- 183 V. R. Salvi and P. Pawar, *J. Drug Deliv. Sci. Technol.*, 2019, **51**, 255–267.
- 184 A. C. Silva, E. González-Mira, M. L. García, M. A. Egea, J. Fonseca, R. Silva, D. Santos, E. B. Souto and D. Ferreira, *Colloids Surf. B Biointerfaces*, 2011, **86**, 158–165.
- 185 V. Mishra, K. K. Bansal, A. Verma, N. Yadav, S. Thakur, K. Sudhakar and J. M. Rosenholm, *Pharmaceutics*, 2018, **10**, 191.
- 186 Y. Duan, A. Dhar, C. Patel, M. Khimani, S. Neogi, P. Sharma, N. S. Kumar and R. L. Vekariya, *RSC Adv.*, 2020, **10**, 26777–26791.
- 187 C. D. Pizzol, F. B. Filippin-Monteiro, J. A. S. Restrepo, F. Pittella, A. H. Silva, P. Alves de Souza, A. Machado de Campos and T. B. Creczynski-Pasa, *Int. J. Environ. Res. Public Health*, 2014, **11**, 8581–8596.
- 188 A. H. Silva, F. B. Filippin-Monteiro, B. Mattei, B. G. Zanetti-Ramos and T. B. Creczynski-Pasa, *Sci. Total Environ.*, 2012, **432**, 382–388.
- 189 V.-A. Duong, T.-T.-L. Nguyen and H.-J. Maeng, *Molecules*, 2020, **25**, 4781.
- 190 E. B. Souto, I. Baldim, W. P. Oliveira, R. Rao, N. Yadav, F. M. Gama and S. Mahant, *Expet Opin. Drug Deliv.*, 2020, **17**, 357–377.
- 191 G. Amasya, G. Sandri, A. Onay-Besikci, U. Badilli, C. Caramella, M. C. Bonferoni and N. Tarimci, *Curr. Drug Deliv.*, 2016, **13**, 1100–1110.
- 192 R. A. Khallaf, H. F. Salem and A. Abdelbary, *Drug Deliv.*, 2016, **23**, 3452–3460.
- 193 G. M. Gelfuso, M. S. Cunha-Filho and T. Gratieri, *Ther. Deliv.*, 2016, **7**, 735–737.
- 194 B. Gaba, M. Fazil, S. Khan, A. Ali, S. Baboota and J. Ali, *Bull. Fac. Pharm. Cairo Univ.*, 2015, **53**, 147–159.
- 195 A. Khosa, S. Reddi and R. N. Saha, *Biomed. Pharmacother.*, 2018, **103**, 598–613.
- 196 V. K. Rapalli, V. Kaul, T. Waghule, S. Gorantla, S. Sharma, A. Roy, S. K. Dubey and G. Singhvi, *Eur. J. Pharm. Sci.*, 2020, **152**, 105438.
- 197 S. Vaz, R. Silva, M. H. Amaral, E. Martins, J. M. Sousa Lobo and A. C. Silva, *Colloids Surf. B Biointerfaces*, 2019, **179**, 242–249.
- 198 S. Wadhwa, V. Garg, M. Gulati, B. Kapoor, S. K. Singh and N. Mittal, in *Pharmaceutical Nanotechnology: Basic Protocols*, ed. V. Weissig and T. Elbayoumi, Springer, New York, NY, 2019, pp. 1–17.
- 199 N. Grimaldi, F. Andrade, N. Segovia, L. Ferrer-Tasies, S. Sala, J. Veciana and N. Ventosa, *Chem. Soc. Rev.*, 2016, **45**, 6520–6545.
- 200 H. Daraee, A. Etemadi, M. Kouhi, S. Alimirzalu and A. Akbarzadeh, *Artif. Cells Nanomed. Biotechnol.*, 2016, **44**, 381–391.
- 201 N. Filipczak, J. Pan, S. S. K. Yalamarty and V. P. Torchilin, *Adv. Drug Deliv. Rev.*, 2020, **156**, 4–22.
- 202 E. Elizondo, E. Moreno, I. Cabrera, A. Córdoba, S. Sala, J. Veciana and N. Ventosa, in *Progress in Molecular Biology and Translational Science*, ed. A. Villaverde, Academic Press, 2011, vol. 104, pp. 1–52.
- 203 L. Maja, K. Željko and P. Mateja, *J. Supercrit. Fluids*, 2020, **165**, 104984.
- 204 K. S. Ahmed, S. A. Hussein, A. H. Ali, S. A. Korma, Q. Lipeng and C. Jinghua, *J. Drug Target.*, 2019, **27**, 742–761.
- 205 A. Reis and C. M. Spickett, *Biochim. Biophys. Acta Biomembr.*, 2012, **1818**, 2374–2387.
- 206 M. A. Obeid, R. J. Tate, A. B. Mullen and V. A. Ferro, in *Lipid Nanocarriers for Drug Targeting*, ed. A. M. Grumezescu, William Andrew Publishing, 2018, pp. 313–359.
- 207 K. Yadav, D. Singh, M. R. Singh and M. Pradhan, *Med. Hypotheses*, 2020, **145**, 110322.
- 208 P. Bhardwaj, P. Tripathi, R. Gupta and S. Pandey, *J. Drug Deliv. Sci. Technol.*, 2020, **56**, 101581.
- 209 S. Chen, S. Hanning, J. Falconer, M. Locke and J. Wen, *Eur. J. Pharm. Biopharm.*, 2019, **144**, 18–39.
- 210 R. Bartelds, M. H. Nematollahi, T. Pols, M. C. A. Stuart, A. Pardakhty, G. Asadikaram and B. Poolman, *PloS One*, 2018, **13**, e0194179.
- 211 G. M. El Maghraby, M. F. Arafa and E. A. Essa, in *Applications of Nanocomposite Materials in Drug Delivery*, ed. E. M. Inamuddin, A. M. Asiri and A. Mohammad, Woodhead Publishing, 2018, pp. 469–490.
- 212 T. G. Singh and N. Sharma, in *Nanobiomaterials in Galenic Formulations and Cosmetics*, ed. A. M. Grumezescu, William Andrew Publishing, 2016, pp. 149–174.
- 213 I. Ali, M. R. Shah, S. Yousuf, S. Ahmed, K. Shah and I. Javed, *Toxicol. Res.*, 2018, **7**, 771–778.
- 214 G. P. Kumar and P. Rajeshwarrao, *Acta Pharm. Sin. B*, 2011, **1**, 208–219.
- 215 Y. Zhang, K. Zhang, Z. Wu, T. Guo, B. Ye, M. Lu, J. Zhao, C. Zhu and N. Feng, *Int. J. Pharm.*, 2015, **478**, 138–146.
- 216 R. Liang, L. Chen, W. Yokoyama, P. A. Williams and F. Zhong, *J. Agric. Food Chem.*, 2016, **64**, 9180–9188.
- 217 M. H. Nematollahi, A. Pardakhty, M. Torkzadeh-Mahanai, M. Mehrabani and G. Asadikaram, *RSC Adv.*, 2017, **7**, 49463–49472.
- 218 R. Rajera, K. Nagpal, S. K. Singh and D. N. Mishra, *Biol. Pharm. Bull.*, 2011, **34**, 945–953.
- 219 P. L. Yeo, C. L. Lim, S. M. Chye, A. P. K. Ling and R. Y. Koh, *Asian Biomed.*, 2018, **11**, 301–314.
- 220 B. Lu, Y. Huang, Z. Chen, J. Ye, H. Xu, W. Chen and X. Long, *Molecules*, 2019, **24**, 2322.
- 221 R. Muzzalupo and E. Mazzotta, *Expet Opin. Drug Deliv.*, 2019, **16**, 1145–1147.
- 222 J. Chen, W.-L. Lu, W. Gu, S.-S. Lu, Z.-P. Chen and B.-C. Cai, *Expet Opin. Drug Deliv.*, 2013, **10**, 845–856.
- 223 B. Piumitali, U. Neeraj and J. Jyotivardhan, *Int. J. Nanosci.*, 2020, **19**, 1950033.
- 224 G. Cevc and A. Chopra, in *Percutaneous penetration enhancers chemical methods in penetration enhancement*, Springer, 2016, pp. 39–59.
- 225 P.-S. Wu, Y.-S. Li, Y.-C. Kuo, S.-J. J. Tsai and C.-C. Lin, *Molecules*, 2019, **24**, 600.
- 226 S. A. T. Opatha, V. Titapiwatanakun and R. Chutoprapat, *Pharmaceutics*, 2020, **12**, 855.
- 227 S. Rai, V. Pandey and G. Rai, *Nano Rev. Exp.*, 2017, **8**, 1325708.



- 228 P. Chaurasiya, E. Ganju, N. Upmanyu, S. K. Ray and P. Jain, *J. Drug Deliv. Ther.*, 2019, **9**, 279–285.
- 229 P. Chauhan and B. K. Tyagi, *J. Drug Deliv. Ther.*, 2018, **8**, 162–168.
- 230 S. Jain, P. Jain, R. B. Umamaheshwari and N. K. Jain, *Drug Dev. Ind. Pharm.*, 2003, **29**, 1013–1026.
- 231 J.-P. Zhang, Y.-H. Wei, Y. Zhou, Y.-Q. Li and X.-A. Wu, *Arch. Pharm. Res.*, 2012, **35**, 109–117.
- 232 A. Ascenso, S. Raposo, C. Batista, P. Cardoso, T. Mendes, F. G. Praça, M. V. L. B. Bentley and S. Simões, *Int. J. Nanomedicine*, 2015, **10**, 5837–5851.
- 233 I. M. Abdulbaqi, Y. Darwis, N. A. K. Khan, R. A. Assi and A. A. Khan, *Int. J. Nanomedicine*, 2016, **11**, 2279–2304.
- 234 M. M. A. Elsayed, O. Y. Abdallah, V. F. Naggar and N. M. Khalafallah, *Int. J. Pharm.*, 2006, **322**, 60–66.
- 235 E. Touitou, B. Godin and C. Weiss, *Drug Dev. Res.*, 2000, **50**, 406–415.
- 236 D. Aggarwal and U. Nautiyal, *Int. J. Pharm. Med. Res.*, 2016, **4**, 354–363.
- 237 P. Verma and K. Pathak, *J. Adv. Pharm. Technol. Res.*, 2010, **1**, 274–282.
- 238 D. Mishra, N. Balekar, V. Dhote and P. Mishra, 2017.
- 239 E. Touitou, N. Dayan, L. Bergelson, B. Godin and M. Eliaz, *J. Control. Release*, 2000, **65**, 403–418.
- 240 B. Godin and E. Touitou, *Crit. Rev. Ther. Drug Carr. Syst.*, 2003, **20**, 63–102.
- 241 L.-N. Shen, Y.-T. Zhang, Q. Wang, L. Xu and N.-P. Feng, *Int. J. Pharm.*, 2014, **460**, 280–288.
- 242 Y.-T. Zhang, L.-N. Shen, Z.-H. Wu, J.-H. Zhao and N.-P. Feng, *Int. J. Pharm.*, 2014, **471**, 449–452.
- 243 C. Fan, X. Li, Y. Zhou, Y. Zhao, S. Ma, W. Li, Y. Liu and G. Li, *BioMed Res. Int.*, 2013, 161943.
- 244 L. Yang, L. Wu, D. Wu, D. Shi, T. Wang and X. Zhu, *Int. J. Nanomedicine*, 2017, **12**, 3357–3364.
- 245 P. K. Jaiswal, S. Kesharwani, R. Kesharwani and D. K. Patel, *J. Drug Deliv. Ther.*, 2016, **6**, 7–17.
- 246 X.-Q. Niu, D.-P. Zhang, Q. Bian, X.-F. Feng, H. Li, Y.-F. Rao, Y.-M. Shen, F.-N. Geng, A.-R. Yuan, X.-Y. Ying and J.-Q. Gao, *Int. J. Pharm. X*, 2019, **1**, 100027.
- 247 V. K. Rai, N. Mishra, K. S. Yadav and N. P. Yadav, *J. Controlled Release*, 2018, **270**, 203–225.
- 248 H. H. Tayeb and F. Sainsbury, *Nanomed*, 2018, **13**, 2507–2525.
- 249 N. Salim, N. Ahmad, S. H. Musa, R. Hashim, T. F. Tadros and M. Basri, *RSC Adv.*, 2016, **6**, 6234–6250.
- 250 S. A. Yousef, Y. H. Mohammed, S. Namjoshi, J. E. Grice, H. A. E. Benson, W. Sakran and M. S. Roberts, *Pharmaceutics*, 2019, **11**, 639.
- 251 O. Bayraktar, İ. Erdoğan, M. D. Köse and G. Kalmaz, in *Nanostructures for Antimicrobial Therapy*, Elsevier, 2017, pp. 395–412.
- 252 R. K. Harwansh, R. Deshmukh and M. A. Rahman, *J. Drug Deliv. Sci. Technol.*, 2019, **51**, 224–233.
- 253 A. de J. Cenobio-Galindo, J. Ocampo-López, A. Reyes-Munguía, M. L. Carrillo-Inungaray, M. Cawood, G. Medina-Pérez, F. Fernández-Luqueño and R. G. Campos-Montiel, *Antioxidants*, 2019, **8**, 500.
- 254 Q. Liu, H. Huang, H. Chen, J. Lin and Q. Wang, *Molecules*, 2019, **24**, 4242.
- 255 L. Pavoni, D. R. Perinelli, G. Bonacucina, M. Cespi and G. F. Palmieri, *Nanomaterials*, 2020, **10**, 135.
- 256 D. S. Shaker, R. A. H. Ishak, A. Ghoneim and M. A. Elhuoni, *Sci. Pharm.*, 2019, **87**, 17.
- 257 E. Sánchez-López, M. Guerra, J. Dias-Ferreira, A. Lopez-Machado, M. Ettcheto, A. Cano, M. Espina, A. Camins, M. L. Garcia and E. B. Souto, *Nanomaterials*, 2019, **9**, 821.
- 258 C. Pulce and J. Descotes, in *Human Toxicology*, ed. J. Descotes, Elsevier Science B.V., Amsterdam, 1996, pp. 683–702.
- 259 Y. Lu, E. Zhang, J. Yang and Z. Cao, *Nano Res.*, 2018, **11**, 4985–4998.
- 260 T. Tadros, in *Encyclopedia of Colloid and Interface Science*, ed. T. Tadros, Springer, Berlin, Heidelberg, 2013, pp. 209–210.
- 261 Y. Singh, J. G. Meher, K. Raval, F. A. Khan, M. Chaurasia, N. K. Jain and M. K. Chourasia, *J. Controlled Release*, 2017, **252**, 28–49.
- 262 M. Miastkowska, E. Sikora and J. Ogonowski, *Period. Polytech. Chem. Eng.*, 2014, **58**, 43–48.
- 263 M. Kumar, R. S. Bishnoi, A. K. Shukla and C. P. Jain, *Prev. Nutr. Food Sci.*, 2019, **24**, 225–234.
- 264 A. Gupta, A. Z. M. Badruddoza and P. S. Doyle, *Langmuir*, 2017, **33**, 7118–7123.
- 265 O. Alliod, J.-P. Valour, S. Urbaniak, H. Fessi, D. Dupin and C. Charcosset, *Colloids Surf., A*, 2018, **557**, 76–84.
- 266 L. Wang, X. Guan, C. Zheng, N. Wang, H. Lu and Z. Huang, *Langmuir*, 2020, **36**, 10082–10090.
- 267 M. N. Yukuyama, D. D. M. Ghisleni, T. J. A. Pinto and N. A. Bou-Chacra, *Int. J. Cosmet. Sci.*, 2016, **38**, 13–24.
- 268 S. Manickam, K. Sivakumar and C. H. Pang, *Ultrason. Sonochem.*, 2020, **69**, 105258.
- 269 L. A. Pascual-Pineda, E. Flores-Andrade, M. Jiménez-Fernández and C. I. Beristain, *Int. J. Food Sci. Technol.*, 2015, **50**, 1174–1181.
- 270 A. Simonazzi, A. G. Cid, M. Villegas, A. I. Romero, S. D. Palma and J. M. Bermúdez, in *Drug Targeting and Stimuli Sensitive Drug Delivery Systems*, ed. A. M. Grumezescu, William Andrew Publishing, 2018, pp. 81–116.
- 271 C. Solans and I. Solé, *Curr. Opin. Colloid Interface Sci.*, 2012, **17**, 246–254.
- 272 M. Ferreira, L. Barreiros, M. A. Segundo, T. Torres, M. Selores, S. A. Costa Lima and S. Reis, *Colloids Surf. B Biointerfaces*, 2017, **159**, 23–29.
- 273 S. Trombino, R. Russo, S. Mellace, G. P. Varano, A. S. Laganà, F. Marcucci and R. Cassano, *J. Drug Deliv. Sci. Technol.*, 2019, **49**, 563–569.
- 274 A. Ali, S. Ali, M. Aqil, S. S. Imam, A. Ahad and A. Qadir, *J. Drug Deliv. Sci. Technol.*, 2019, **52**, 713–721.
- 275 J.-H. Kang, J. Chon, Y.-I. Kim, H.-J. Lee, D.-W. Oh, H.-G. Lee, C.-S. Han, D.-W. Kim and C.-W. Park, *Int. J. Nanomedicine*, 2019, **14**, 5381–5396.
- 276 P. Tripathi, A. Kumar, P. K. Jain and J. R. Patel, *Int. J. Biol. Macromol.*, 2018, **120**, 1322–1334.



- 277 N. Kaur, K. Sharma and N. Bedi, *Pharm. Nanotechnol.*, 2018, **6**, 133–143.
- 278 P. Sathe, R. Saka, N. Kommineni, K. Raza and W. Khan, *Drug Dev. Ind. Pharm.*, 2019, **45**, 826–838.
- 279 Y. O. Agrawal, U. B. Mahajan, H. S. Mahajan and S. Ojha, *Int. J. Nanomedicine*, 2020, **15**, 4763–4778.
- 280 R. Kumar, S. Dogra, B. Amarji, B. Singh, S. Kumar, K. Vinay, R. Mahajan and O. Katore, *JAMA Dermatol.*, 2016, **152**, 807–815.
- 281 S. Doppalapudi, A. Jain, D. K. Chopra and W. Khan, *Eur. J. Pharm. Sci.*, 2017, **96**, 515–529.
- 282 M. Walunj, S. Doppalapudi, U. Bulbake and W. Khan, *J. Liposome Res.*, 2020, **30**, 68–79.
- 283 J. Chen, Y. Ma, Y. Tao, X. Zhao, Y. Xiong, Z. Chen and Y. Tian, *J. Liposome Res.*, 2020, 1–15.
- 284 W. Wang, G. Shu, K. Lu, X. Xu, M. Sun, J. Qi, Q. Huang, W. Tan and Y. Du, *J. Nanobiotechnol.*, 2020, **18**, 80.
- 285 H. Liu, R. S. Kang, K. Bagnowski, J. M. Yu, S. Radecki, W. L. Daniel, B. R. Anderson, S. Nallagatla, A. Schook, R. Agarwal, D. A. Giljohann and A. S. Paller, *J. Invest. Dermatol.*, 2020, **140**, 435–444.
- 286 R. Saka, H. Jain, N. Kommineni, N. Chella and W. Khan, *J. Drug Deliv. Sci. Technol.*, 2020, **58**, 101691.
- 287 S. R. M. Moghddam, A. Ahad, M. Aqil, S. S. Imam and Y. Sultana, *Mater. Sci. Eng. C*, 2016, **69**, 789–797.
- 288 I. I. Abu Hashim, N. F. Abo El-Magd, A. R. El-Sheakh, M. F. Hamed and A. E.-G. H. Abd El-Gawad, *Int. J. Nanomedicine*, 2018, **13**, 1059–1079.
- 289 V. Parkash, S. Maan, V. Chaudhary, V. Jogpal, G. Mittal and V. Jain, *J. Bioequivalence Bioavailab.*, 2018, **10**, 99–106.
- 290 M. Bahramizadeh, M. Bahramizadeh, B. Kiafar, A. H. Jafarian, A. R. Nikpoor, M. Hatamipour, H. Esmaily, Z. Rezaeemehr, S. Golmohammadzadeh, S. A. Moosavian and M. R. Jafari, *Int. J. Pharm.*, 2019, **569**, 118623.
- 291 A. Chandra, G. Aggarwal, S. Manchanda and A. Narula, *Pharm. Nanotechnol.*, 2019, **7**, 362–374.
- 292 D. Fathalla, E. M. K. Youssef and G. M. Soliman, *Pharmaceutics*, 2020, **12**, 446.
- 293 S. H. Musa, M. Basri, H. R. Fard Masoumi, N. Shamsudin and N. Salim, *Int. J. Nanomedicine*, 2017, **12**, 2427–2441.
- 294 S. Sahu, S. S. Katiyar, V. Kushwah and S. Jain, *Nanomed*, 2018, **13**, 1985–1998.
- 295 P. Rajitha, P. Shammika, S. Aiswarya, A. Gopikrishnan, R. Jayakumar and M. Sabitha, *J. Drug Deliv. Sci. Technol.*, 2019, **49**, 463–476.
- 296 M. S. Algahtani, M. Z. Ahmad and J. Ahmad, *J. Drug Deliv. Sci. Technol.*, 2020, **59**, 101847.
- 297 M. S. Algahtani, M. Z. Ahmad, I. H. Nouredin and J. Ahmad, *Biomolecules*, 2020, **10**, 968.
- 298 R. Bosetti and S. L. Jones, *Nanomed*, 2019, **14**, 1367–1370.
- 299 J. J. Richardson and F. Caruso, *Nano Lett.*, 2020, **20**, 1481–1482.
- 300 E. C. Murphy, S. W. Schaffter and A. J. Friedman, *Curr. Dermatol. Rep.*, 2019, **8**, 14–25.
- 301 S. Saleem, M. K. Iqbal, S. Garg, J. Ali and S. Baboota, *Expet Opin. Drug Deliv.*, 2020, **17**, 817–838.
- 302 N. Ahmad, R. Ramsch, M. Llinàs, C. Solans, R. Hashim and H. A. Tajuddin, *Colloids Surf. B Biointerfaces*, 2014, **115**, 267–274.
- 303 L. Bai and D. J. McClements, *J. Colloid Interface Sci.*, 2016, **479**, 71–79.
- 304 M. S. Behalo, E. Bloise, L. Carbone, R. D. Sole, D. Lomonaco, S. E. Mazzetto, G. Mele and L. Mergola, *J. Exp. Nanosci.*, 2016, **11**, 1274–1284.
- 305 O. W. Mak, N. Seman, N. H. Harun, R. S. D. Hussien, N. M. Rodzi and T. Heidelberg, *J. Surfactants Deterg.*, 2015, **18**, 973–980.
- 306 A. Sedaghat Doost, J. Van Camp, K. Dewettinck and P. Van der Meeren, *Food Chem.*, 2019, **293**, 134–143.
- 307 N. J. DeVito, S. Bacon and B. Goldacre, *Lancet*, 2020, **395**, 361–369.
- 308 M. Zwierzyna, M. Davies, A. D. Hingorani and J. Hunter, *Br. Med. J.*, 2018, **361**, k2130.
- 309 J. K. Patra, G. Das, L. F. Fraceto, E. V. R. Campos, M. del Pilar Rodriguez-Torres, L. S. Acosta-Torres, L. A. Diaz-Torres, R. Grillo, M. K. Swamy and S. Sharma, *J. Nanobiotechnol.*, 2018, **16**, 71.
- 310 C. L. Ventola, *Pharm. Ther.*, 2017, **42**, 742–755.
- 311 R. Kumar, S. V. Dalvi and P. F. Siril, *ACS Appl. Nano Mater.*, 2020, **3**, 4944–4961.
- 312 D. Lombardo, M. A. Kiselev and M. T. Caccamo, *J. Nanomater.*, 2019, **2019**, 3702518.
- 313 A. V. Singh, M. H. D. Ansari, D. Rosenkranz, R. S. Maharjan, F. L. Kriegel, K. Gandhi, A. Kanase, R. Singh, P. Laux and A. Luch, *Adv. Healthc. Mater.*, 2020, **9**, 1901862.

