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Self-Amplified Depolymerization of Oligo(thiourethanes) for the Release of COS/H₂S

Received 00th January 20xx, Accepted 00th January 20xx Chadwick R. Powell, Jeffrey C. Foster, Sarah N. Swilley, Kuljeet Kaur, Samantha J. Scannelli, Diego Troya and John B. Matson^{*}

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Herein we report the self-amplified depolymerization of an aryl oligo(thiourethane) (OTU) for the release of COS/H_2S . The OTU was synthesized via polyaddition of 4-isothiocyanatobenzyl alcohol and end-capped with an aryl azide. The aryl azide chain-end was reduced by tris(2-carboxyethyl)phosphine or H_2S to the corresponding aniline, resulting in depolymerization (i.e., self-immolation) and the release of COS/H_2S . Depolymerization was monitored by ¹H NMR and UV-Vis spectroscopy, and the released COS was converted into H_2S by the ubiquitous enzyme carbonic anhydrase in aqueous media.

Depolymerizable or degradable polymers (i.e., selfimmolative polymers) are a class of materials which depolymerize in the presence of a specific stimulus, typically resulting in the release of small molecules.¹ These stimuliresponsive depolymerizable polymers are comprised of three discrete portions: a triggering moiety, a spacer, and an output.² The triggering moiety is a functional group that responds to a specific stimulus such as light,³ redox reactions,⁴ or a small molecule.^{5, 6} Application of the stimulus results in the formation of an unstable intermediate, typically on the polymer chainend, which causes the depolymerization of a single monomer unit and the subsequent regeneration of the unstable intermediate (Scheme 1A). This process repeats until each monomer unit in the polymer chain has depolymerized. The utility of depolymerizable polymers derives from the release of the output molecule, which occurs concurrently with depolymerization. Output molecules are often quantifiable (i.e., fluorescent small molecules), making depolymerizable polymers intriguing motifs for signal detection and amplification.7, 8 Despite this progress in depolymerizable for detection of biological events, polymers few depolymerizable polymers have been developed that release biologically active output molecules.^{1, 9} Here we envisioned developing an depolymerizable polymer capable of releasing hydrogen sulfide (H₂S), a biological signalling gas.

Department of Chemistry

A) Self-propagating depolymerizable poly(urethane)



B) Benzyl thiocarbamate small molecule H₂S donors



C) Self-propagating depolymerizable poly(thiourethane) - This work



Scheme 1. A) Depolymerizable aryl poly(urethanes). B) Small molecule benzyl thiocarbamate COS/H₂S donors developed by Pluth and coworkers. C) Proposed COS/H₂S releasing depolymerizable polyurethane.

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In 1996 H₂S was established as a critical signalling molecule in mammals.¹⁰ As such, alterations in H₂S production in the body have systemic consequences and have been linked to a variety of disease states including cardiovascular disease,¹¹ cystic fibrosis,¹² and diabetes,¹³ among others.¹⁴ A commonality in these disease states is a decrease in endogenous H_2S production.¹⁵ As a result, exogenous delivery of H₂S through inorganic donor salts (NaSH, Na2S) or synthetic donor compounds may mitigate disease symptoms and improve healing.¹⁶⁻¹⁹ To aid in understanding H₂S physiology and investigate possible benefits of H₂S therapy, several types of small molecule and polymeric H₂S donors have been developed over the past few years.²⁰⁻²⁴ However, many classes of synthetic donors release only one equivalent of H₂S per equivalent of consumed trigger, which is often a redox-active thiol. This net neutral redox balance may ultimately limit the long-term efficacy of common of synthetic donors and complicate in vivo analysis.^{25, 26} Thus, H₂S donors that release multiple equivalents of H₂S per triggering event may be critical in furthering the therapeutic benefit of exogenous H₂S delivery.

In an effort to develop a depolymerizable polymer that can release multiple equivalents of H₂S in response to low concentrations of H₂S itself, we were inspired by Pluth and coworkers' 2016 report that introduced benzyl thiocarbamates as a class of small molecule, dual carbonyl sulfide (COS)/H₂S donors based on the benzyl elimination reaction (Scheme 1B).²⁷ These benzyl thiocarbamates released COS via a 1,6-elimination reaction triggered by a reducing stimulus, such as H₂S itself. The elimination reaction led to release of the desired COS payload via an unstable thiocarbamic acid intermediate (Scheme 1B). The released COS was then rapidly hydrolyzed to H₂S via the action of the ubiquitous enzyme carbonic anhydrase (CA). Since this seminal work on small molecule thiocarbamates as dual COS/H₂S donors, Pluth and coworkers have demonstrated the ability to trigger COS/H₂S release in the presence of other stimuli including hydrogen peroxide,²⁸ cysteine,²⁹ light,³⁰ and others.^{29, 31} We envisioned that leveraging benzyl thiocarbamates as the repeat unit of a depolymerizable polymer would provide an exciting opportunity for a platform from which endogenous H₂S production may be amplified, creating a self-amplified depolymerizable polymer (SADP) (Scheme 1C).

A) NaBH₄ Pd/C Azide end-capper (EC1) H₂O 1) CS₂, TEA, EtOH N₂ HO 2) Boc₂O, EtOH 3) DMAP SADM1 B) 1) DBTDL NCS DMF, 60 °C, 7.5 h 2) EC1 DMF, 60 °C, 16 h

Scheme 2. A) Synthesis of SADM1. B) Synthesis of aryl azide end-capped SADP1.

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In order to synthesize a COS/H₂S-donating SADP, we first set out to prepare monomer that could undergo a step-growth polyaddition to make the desired poly(thiourethane) (PTU). Typically, PTUs are prepared as S-alkyl thiocarbamates through the reaction of thiols and isocyanates mediated by the soft (DBTDL).32-34 Lewis-acid catalyst dibutyltin dilaurate Unfortunately, the S-alkyl thiocarbamate isomer is a less efficient COS donor than the O-alkyl isomer, likely stemming from an unfavorable Gibb's free energy (ΔG) for the COSreleasing reaction.³⁵ In contrast, the O-alkyl thiourethane isomer readily decomposes to form COS via the 1,6-benzyl elimination. Accordingly, synthesis of a depolymerizable O-alkyl thiourethane repeating unit would require a monomer containing both an aryl isothiocyanate (Ar-NCS) and a benzyl alcohol to facilitate efficient COS release.

To meet this challenge, we designed and synthesized a bifunctional monomer containing the desired aryl isothiocyanate and benzyl alcohol functional groups (SADM1, Scheme 2). Starting from 4-nitrobenzaldehyde, a one-pot reduction of both the nitro and aldehyde was accomplished by addition of sodium borohydride (NaBH₄) and palladium on carbon (Pd/C, 5 mol %) in water to give 4-aminobenzyl alcohol (4-AB). To access the aryl isothiocyanate, a method developed by Boas and coworkers³⁶ was employed wherein the aniline of 4-AB was converted into the corresponding dithiocarbamate salt by reaction with carbon disulfide in the presence of triethylamine, followed by addition of Boc anhydride, which led to the spontaneous evolution of COS gas and tert-butyl alcohol and the formation of the desired aryl isothiocyanate (SADM1).

With the desired AB monomer in hand, we envisioned that it would undergo polyaddition in the presence of DBTDL, as this catalyst has been used successfully in analogous, non-sulfurcontaining systems. Thus, the polymerization of SADM1 was carried out in dry DMF (1 M) at 60 °C under N₂ in the presence of DBTDL (5 mol %). Under these conditions we observed a plateau in monomer conversion (p) after 7.5 h at approximately 85 % (Figure S11). At this stage in the polymerization, 1 equiv of 4-azidobenzylalcohol (EC1) was added as an end-capping reagent, and the reaction mixture was allowed to stir overnight. The azide end-capped SADP (SADP1) was then isolated as a yellow powder after precipitating from Et₂O. The presence of the aryl azide on the oligomer chain end was confirmed by FTIR spectroscopy (Figure S9), and the oligomer M_n was measured to be 1.6 kg/mol by ¹H NMR end-group analysis (degree of polymerization (X_n) of ~7), which is consistent with the expected $M_{\rm n}$ for 85% conversion. Size exclusion chromatography (SEC) with light scattering detection was attempted, but the low molecular weight of the oligomer coupled with its low dn/dc value in the elution solvent (THF) prevented accurate analysis.

To explain the limited conversion of monomer under these conditions, the reaction Gibbs energy for a model small molecule reaction between benzyl alcohol and phenyl isothiocyanate was calculated using density functional theory. The 60 °C reaction Gibbs energy using the M06-2X functional and aug-cc-pVDZ basis with implicit DMF solvation is -2.28 kcal/mol. Combining Carother's equation for step-growth polymerizations with the reaction Gibbs energy relationship to



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the equilibrium constant (*K*) (equation 1) we calculated p to be 0.85 for this system under ideal conditions. These calculations indicate a maximum degree of polymerization (X_n) of 6.7. The polymerization of **SADM1** routinely yielded oligomers with $X_n \sim 7$, in good agreement with the predictions. Therefore, the limited conversion observed experimentally appears to be due to the small polymerization excergicity under the experimental conditions.

$$p_{eq} = \frac{\sqrt{e^{\frac{-\Delta G}{RT}}}}{1 + \sqrt{e^{\frac{-\Delta G}{RT}}}} \quad (Equation 1)$$

We next investigated the depolymerization of azideterminated SADP1 by ¹H NMR spectroscopy. For these experiments, tris(2-carboxyethyl)phosphine (TCEP, 1.7 equiv with respect to azide chain ends) was employed as an organosoluble reducing agent to facilitate the reduction of the chainend aryl azide, leading to depolymerization and ultimately COS release. In order to follow the reaction, changes in the peaks attributed to SADP1 were monitored as well as the generation of 4-AB, an expected major byproduct of SADP1 depolymerization. Due to the low water solubility of SADP1 at concentrations required for NMR spectroscopy, ¹H NMR analysis was performed in DMSO- d_6 , which dramatically decreases reaction rates for 1,6-elimination reactions relative to water.³⁷ However, despite the slow reaction rate, ¹H NMR analysis revealed a decrease in the broad heteroatomic peak attributed to the thiocarbamate repeating unit proton (Ar-NHC(S)O) as well as the appearance of well resolved aromatic doublets consistent with 4-AB. Under the same conditions, depolymerization of a benzyl alcohol end-capped control SADP (Ctrl-SADP) occurred more slowly in the presence of TCEP (Figures S12 and S13), indicating that reduction of the chain-end azide is critical for initiating depolymerization.

Monitoring depolymerization by UV-Vis spectroscopy allowed for use of aqueous media because lower concentrations of **SADP1** could be employed than in the ¹H NMR spectroscopy experiments. For these experiments, **SADP1** was dissolved in PBS buffer (pH 7.4) containing DMSO (2 % v/v) and cetrimonium bromide (CTAB) to aid in solubility. Prior to adding a reducing agent, a broad absorbance for **SADP1** was observed at 284 nm (Figure 1A). Upon addition of a reducing agent (TCEP or Na₂S), the absorbance maximum began to gradually shift to lower wavelength over the course of 2 h. We attribute the shift in λ_{max} for the oligomer to depolymerization and the generation of 4-AB. Isosbestic points at 240 and 271 nm were observed, although they shifted slightly during the course of the analysis, which we attribute to low MW SADP species and oxidized byproducts of 4-AB.

In order to better evaluate the depolymerization kinetics of **SADP1**, a series of UV-Vis experiments were conducted using varying amounts of TCEP, Na₂S, and no trigger (hydrolysis) in the presence of CA (which converts COS into H₂S). By plotting the change in absorbance at 284 nm (λ_{max} of **SADP1**) over time, a

clear trend in reaction kinetics was observed. Addition of 1 equiv TCEP relative to aryl azide **SADP1** chain-end gave the greatest decrease in λ_{max} for **SADP1** over the course of 2 h. When the amount of TCEP was reduced to 0.1 equiv, a concomitant decrease in rate was observed. Addition of Na₂S (0.1 equiv) as an alternative means of reducing the **SADP1** chain-end gave data similar to that for the addition of 0.1 equiv TCEP, indicating that TCEP and Na₂S are roughly equal in reduction capacity under these conditions and that both generate multiple equivalents of H₂S per equivalent of added trigger. Additionally, the same shift in λ_{max} was observed without the addition of any reducing agent, albeit at a much slower rate, indicating that hydrolysis, likely of the thiourethane units, contributes to the depolymerization of **SADP1**.



Figure 1. A) Representative UV-Vis spectra of **SADP1** (10 μ M) prior to the addition of reducing agent (λ_{max} = 284 nm) and 2 h after the addition of reducing agent (λ_{max} = 254 nm). B) Change in absorbance of **SADP1** at 284 nm over time in the presence and absence of reducing agents. Data are normalized to the absorbance at t₀, prior to the addition of reducing agent. All depolymerization experiments were run in PBS buffer (pH 7.4) with 2% DMSO, 1 mM CTAB, and 300 nM CA.

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In light of the data indicating that hydrolysis is a viable mechanism for depolymerization of **SADP1**, UV-Vis experiments of **Ctrl-SADP** were conducted to investigate whether the reducing agent (Na₂S) or the H₂S released in the depolymerization reaction might also contribute to degradation of the benzyl thiocarbamate moiety. The spectral profile of **Ctrl-SADP** is very similar to **SADP1**, with a broad absorbance centred at 284 nm (Figure S14). However, there was no significant change the λ_{max} of **Ctrl-SADP** in the presence or absence of Na₂S, indicating that sulfide does not degrade the benzyl thiocarbamate moiety. Therefore, we conclude that the observed enhanced rate of **SADP1** depolymerization in the presence of Na₂S compared with water is due to the reduction of the chain-end azide and subsequent depolymerization.

Lastly, analysis of the H₂S release profile for SADP1 was performed using an H₂S-selective electrochemical probe. H₂S release experiments were performed in PBS buffer (pH 7.4) at 100 μ M **SADP1** with the addition of CTAB, similar to the UV-Vis experiments. Addition of Na₂S (0.1 equiv) as the reducing agent to a solution of SADP1 containing CA generated an initial spike in H₂S due to the presence of Na₂S, followed by a rapid decrease in H₂S concentration, followed by steady generation of H₂S, ultimately reaching a peak concentration after 220 min (Figure 2, purple curve). In contrast, addition of Na₂S (0.1 equiv) to a solution of SADP1 in the absence of CA generated a spike in H₂S concentration followed by a rapid return to baseline, similar to probe response when only Na₂S is added (orange curve). This result indicates that the released COS from SADP1 was not converted into H_2S , as expected when CA is not present. The apparently low peak H_2S concentration (0.9 μ M) is due to the long peaking time of SADP1, where low peak concentration is a result of slow H₂S generation combined with COS/H₂S volatilization and H₂S oxidation. Taken together, results from these H₂S release experiments demonstrate that SADP1 successfully generates H₂S in the presence of submolar quantities of a Na₂S trigger, acting in an autoinductive selfpropagating amplification reaction $^{\rm 38}$ where $\rm H_2S$ derived from hydrolysis of COS generates increasing amounts of H₂S.

Conclusions

The first COS/H₂S-releasing SADP is reported. The depolymerizable oligo(thiourethane) was synthesized in a polyaddition reaction from a bifunctional monomer containing an aryl isothiocyanate and a benzyl alcohol. The oligomer structure was confirmed by ¹H NMR and FTIR spectroscopy. Aryl azide-terminated **SADP1** underwent depolymerization in the presence of reducing agents, with a greater concentration of reducing agent resulting in an enhanced reaction rate. Additionally, upon addition of submolar concentrations of reducing agents, including Na₂S, **SADP1** demonstrated COS release, which was converted to H₂S in the presence of CA, generating multiple equivalents of H₂S per triggering event in a manner consistent with signal amplification.



Figure 2. A) Scheme depicting H_2S release from SADP1. B) H_2S release data from SADP1 (100 μ M) in the presence of Na₂S (0.1 equiv) with 300 nM CA (purple curve) and without CA (orange curve).

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Conflicts of interest

There are no conflicts to declare.

Notes and references

- 1 G. I. Peterson, M. B. Larsen and A. J. Boydston,
- Macromolecules, 2012, **45**, 7317-7328.
- 2 P. L. Carl, P. K. Chakravarty and J. A. Katzenellenbogen, *J. Med. Chem.*, 1981, **24**, 479-480.
- F. M. H. de Groot, C. Albrecht, R. Koekkoek, P. H. Beusker, H.
 W. Scheeren, R. J. Amir, N. Pessah, M. Shamis and D. Shabat, Angew. Chem. Int. Ed., 2003, 42, 4411-4411.
- 4 R. Weinstain, P. S. Baran and D. Shabat, *Bioconjugate Chem.*, 2009, **20**, 1783-1791.
- 5 W. Seo and S. T. Phillips, *J. Am. Chem. Soc.*, 2010, **132**, 9234-9235.
- 6 H. Zhang, K. Yeung, J. S. Robbins, R. A. Pavlick, M. Wu, R. Liu, A. Sen and S. T. Phillips, *Angew. Chem. Int. Ed.*, 2012, **51**, 2400-2404.
- 7 A. Sagi, R. Weinstain, N. Karton and D. Shabat, J. Am. Chem. Soc., 2008, **130**, 5434-5435.
- 8 O. Redy, E. Kisin-Finfer, E. Sella and D. Shabat, *Org. Biomol. Chem.*, 2012, **10**, 710-715.

Journal Name

Journal Name

- 9 D. Shabat, J. Polym. Sci., Part A: Polym. Chem., 2006, 44, 1569-1578.
- 10 K. Abe and H. Kimura, J. Neurosci., 1996, 16, 1066-1071.
- D. J. Lefer, Proc. Natl. Acad. Sci. U. S. A., 2007, 104, 17907.
 L. Fang, H. Li, C. Tang, B. Geng, Y. Qi and X. Liu, Can. J. Physiol. Pharmacol., 2009, 87, 531-538.
- 13 C. Szabo, Antioxidants & redox signaling, 2012, **17**, 68-80.
- 14 C. R. Powell, K. M. Dillon and J. B. Matson, *Biochem. Pharmacol.*, 2018, **149**, 110-123.
- 15 R. Wang, FASEB J., 2002, 16, 1792-1798.
- 16 W. Zhao, J. Zhang, Y. Lu and R. Wang, *The EMBO Journal*, 2001, **20**, 6008-6016.
- 17 M. Whiteman, N. S. Cheung, Y. Z. Zhu, S. H. Chu, J. L. Siau, B. S. Wong, J. S. Armstrong and P. K. Moore, *Biochem. Biophys. Res. Commun.*, 2005, **326**, 794-798.
- 18 Y. Zhao, C. Yang, C. Organ, Z. Li, S. Bhushan, H. Otsuka, A. Pacheco, J. Kang, H. C. Aguilar, D. J. Lefer and M. Xian, *J. Med. Chem.*, 2015, **58**, 7501-7511.
- 19 Z. Li, C. L. Organ, Y. Zheng, B. Wang and D. J. Lefer, *Circulation*, 2016, **134**, A17903-A17903.
- 20 J. C. Foster, C. R. Powell, S. C. Radzinski and J. B. Matson, Org. Lett., 2014, 16, 1558-1561.
- 21 C. R. Powell, J. C. Foster, B. Okyere, M. H. Theus and J. B. Matson, *J. Am. Chem. Soc.*, 2016, **138**, 13477-13480.
- 22 Z. Xiao, T. Bonnard, A. Shakouri-Motlagh, R. A. L. Wylie, J. Collins, J. White, D. E. Heath, C. E. Hagemeyer and L. A. Connal, *Chem. Eur. J.*, 2017, **23**, 11294-11300.
- 23 F. Ercole, F. M. Mansfeld, M. Kavallaris, M. R. Whittaker, J. F. Quinn, M. L. Halls and T. P. Davis, *Biomacromolecules*, 2016, 17, 371-383.
- 24 Y. Zheng, B. Yu, K. Ji, Z. Pan, V. Chittavong and B. Wang, Angew. Chem. Int. Ed. Engl., 2016, **55**, 4514-4518.
- 25 Y. Zhao, H. Wang and M. Xian, J. Am. Chem. Soc., 2011, **133**, 15-17.
- 26 A. Martelli, L. Testai, V. Citi, A. Marino, I. Pugliesi, E. Barresi, G. Nesi, S. Rapposelli, S. Taliani, F. Da Settimo, M. C. Breschi and V. Calderone, ACS Med. Chem. Lett., 2013, 4, 904-908.
- 27 A. K. Steiger, S. Pardue, C. G. Kevil and M. D. Pluth, *J. Am. Chem. Soc.*, 2016, **138**, 7256-7259.
- 28 Y. Zhao and M. D. Pluth, Angew. Chem. Int. Ed., 2016, 55, 14638-14642.
- 29 Y. Zhao, A. K. Steiger and M. D. Pluth, *Chem. Commun.*, 2018, 54, 4951-4954.
- 30 Y. Zhao, S. G. Bolton and M. D. Pluth, *Org. Lett.*, 2017, **19**, 2278-2281.
- 31 A. K. Steiger, Y. Yang, M. Royzen and M. D. Pluth, Chem. Commun., 2017, 53, 1378-1380.
- 32 A. Kultys, M. Rogulska and S. Pikus, J. Polym. Sci., Part A: Polym. Chem., 2008, **46**, 1770-1782.
- 33 D. Eglin, S. Griffon and M. Alini, *Journal of Biomaterials* Science -- Polymer Edition, 2010, **21**, 477-491.
- 34 M. D. Campiñez, C. Ferris, M. V. de Paz, A. Aguilar-de-Leyva, J. Galbis and I. Caraballo, Int. J. Pharm., 2015, 480, 63-72.
- 35 Y. Zhao, H. A. Henthorn and M. D. Pluth, J. Am. Chem. Soc., 2017, 139, 16365-16376.
- 36 H. Munch, J. S. Hansen, M. Pittelkow, J. B. Christensen and U. Boas, *Tetrahedron Lett.*, 2008, **49**, 3117-3119.
- K. M. Schmid, L. Jensen and S. T. Phillips, J. Org. Chem., 2012, 77, 4363-4374.
- 38 X. Sun, D. Shabat, S. T. Phillips and E. V. Anslyn, J. Phys. Org. Chem., 2018, **31**, e3827.



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