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Reactivity variance between stereoisomers of saturated N-heterocyclic carbenes on gold surfaces†

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Controlling the chirality of molecule-surface systems is essential for applications ranging from heterogeneous catalysis to biosensing. N-heterocyclic carbenes (NHCs) are quickly becoming a dominant ligand for noble metal surface passivation, but the potential of chiral NHC scaffolds remains untapped. In this work, two stereoisomers, one C_2 symmetric and one C_s symmetric, of saturated N-heterocyclic carbenes (NHCs) and a structurally related unsaturated NHC (C_{2v} symmetric) were synthesized as NHC- CO_2 adducts. These CO_2 -protected NHCs were deposited on gold films and their presence on the films was confirmed by laser desorption ionization mass spectrometry (LDI-MS) and surface enhanced Raman spectroscopy (SERS). Surprisingly, the C_s NHC, but not the chiral C_2 NHC, partially degrades to the independently synthesized unsaturated NHC upon binding to the gold. Theoretical calculations assist in explaining this phenomenon by showing that the NHCs primarily lie flat on the gold surfaces, which exposes the backbone protons on the C_s -symmetric NHC to a formal elimination of H_2 , while the C_2 -symmetric NHC remains protected from this elimination reaction. These results raise critical questions as to how the structure of NHC ligands may be tuned to influence binding and reactivity on gold surfaces.

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Introduction

Self-assembled monolayers (SAMs) on gold surfaces are critical for many applications in catalysis, sensing, and medicine.^{1–7} Chiral thiols, in particular, are known to form SAMs on gold that deliver single enantiomer selectivity^{8–12} and L-cysteine-based chiral ligands are widely employed as fabricating agents for surfaces with enantiomeric recognition (Fig. 1A).^{13,14} Nevertheless, thiol-based SAMs degrade over time, especially when exposed to oxygen or native thiols, which limits their application in long term biomedical studies.^{15–17}

N-heterocyclic carbenes (NHCs) are rapidly displacing thiols across myriad applications that employ SAMs due to their strong binding affinity to gold.^{18–20} CO_2 -adducts or bicar-

bonate salts of NHCs easily transfer the NHC ligands to a gold surface such as gold mirror or gold film-over nanospheres (AuFONs).^{19,21–23} However, regular NHCs, which have imidazole or benzimidazole rings, are poorly suited for developing chiral NHCs, except with chiral wingtips (Fig. 1B).^{24–29} Previous studies mostly focus on the organic synthesis^{24,26,29} of NHCs with chiral wingtips and their implementation in homogeneous catalysis.^{25,27,28,30} Chiral NHCs have very rarely been deposited on gold surfaces³¹ and even in these limited cases the chirality does not point away from the surface, which would be most valuable for sensing and medical applications.

Non-standard NHCs, specifically saturated NHCs, offer an alternative that would allow for chiral moieties at the backbone position as opposed to the wingtips (Fig. 1C). However, these saturated NHCs cannot be synthesized by simple addition of electrophilic wingtips to imidazoles or benzimidazoles.^{32,33} As such, this category of chiral saturated NHC has mostly been limited to a few examples for alkene metathesis catalysis.^{34–39}

In this manuscript, we synthesized two stereoisomers, one chiral (C_2) and one *meso* (C_s), of saturated N-heterocyclic carbenes (NHCs) and a structurally related unsaturated NHC (C_{2v}) as their CO_2 adducts (Fig. 1D). The NHC precursors were characterized by multi-nuclear NMR as well as single crystal X-ray which confirmed their absolute stereochemistry. The

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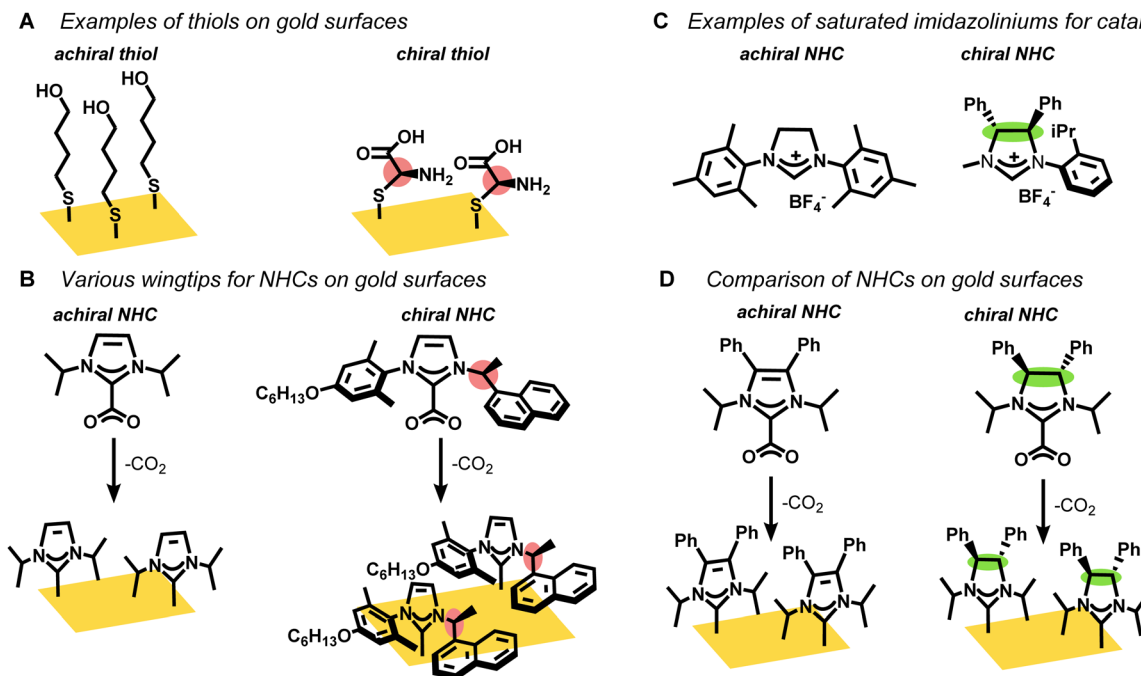


Fig. 1 An overview of achiral and chiral monolayers on surfaces, where red and green represents chiral wingtips and chiral backbone respectively. (A) Thiol SAMs on gold surfaces. (B) NHCs with varying wingtip groups on gold surfaces. (C) Saturated imidazolium studied for homogeneous catalysis. (D) This work that includes saturated NHCs with chiral backbones on gold surfaces.

CO₂-protected NHCs were deposited on gold films and their chemisorption was confirmed by surface enhanced Raman spectroscopy (SERS) and laser desorption ionization mass

spectrometry (LDI-MS). Remarkably, we find that the C_s NHC, but not the chiral C₂ NHC, partially degrades to the unsaturated NHC upon binding to the gold. This surprising result



Scheme 1 Synthesis of C₂, C_s, and C_{2v} symmetric NHC-CO₂ adducts, 6-CO₂, 7-CO₂, and 8-CO₂, respectively.



arises from the surface geometry, which exposes the backbone protons on only the C_s -symmetric NHC, but not the C_2 -symmetric NHC.

Results and discussion

NHC-CO₂ adducts or their related bicarbonate salts are an effective method of NHC transfer to gold films.^{21–23,40} However, for synthesizing saturated NHCs (imidazolinium ring), the wingtips for the NHC must be appended followed by a ring closing.^{23,35} Thus, we followed this strategy for preparing compounds **1** and **2** as the precursors for the chiral and *meso* NHCs, **6** and **7** (Scheme 1).^{41,42} (4*S*,5*S*)-1,3-diisopropyl-4,5-diphenylethanediamine (**1**) was synthesized by addition of excess 2-iodopropane to (1*S*,2*S*)-(-)-1,2-diphenylethylenediamine under basic conditions in a heated pressurized glass vessel in 49% yield (see Experimental section for

additional details†). *meso*-1,3-Diisopropyl-4,5-diphenylethanediamine (**2**) was prepared in a nearly identical fashion in 20% yield but required an additional purification step *via* column chromatography with amine-treated silica gel.

Refluxing **1** and ammonium chloride with excess triethyl orthoformate under acidic conditions yielded (4*S*,5*S*)-1,3-diisopropyl-4,5-diphenylimidazolium chloride (**3a**) in 69% yield. *meso*-1,3-Diisopropyl-4,5-diphenylimidazolium chloride (**4a**) was synthesized under analogous conditions in 58% yield. Since the imidazolium salts are quite stable and the deprotonation occurs at the 2-position to form the NHC-CO₂ adducts, we wanted to confirm the absolute stereochemistry for these compounds *via* single crystal X-ray diffraction. Despite multiple attempts, both **3a** and **4a** had significant disorder in their unit cells due to the chloride counter anion.

To solve the crystallographic disorder due to the counter anion, we switched to hexafluorophosphate by replacing ammonium chloride with ammonium hexafluorophosphate in



Fig. 2 Left: ¹H NMR spectra in CD₃CN from 3–9 ppm of NHC hexafluorophosphate salts (A) **3b**, (B) **4b**, and (C) **5**. Right: X-ray crystal structures of (A) **3b**, (B) **4b**, and (C) **5** shown with thermal ellipsoids at 50% probability. Grey, blue, black, green, and pink ellipsoids represent C, N, H, F, and P atoms, respectively. Hydrogens are omitted except for ones on stereogenic carbons.



the synthesis of **3** and **4**. This switch led to the formation of (4*S*,5*S*)-1,3-diisopropyl-4,5-diphenylimidazolium hexafluorophosphate (**3b**) and *meso*-1,3-diisopropyl-4,5-diphenylimidazolium hexafluorophosphate (**4b**) in 66% and 76% yield, respectively. A comparison of the ¹H NMR spectra for **3b** and **4b** shows mostly similarities but one key distinction due to their difference in stereochemistry (Fig. 2, left). The imidazolium protons (labelled “a” in Fig. 2, left) are close to each other at 8.24 and 8.30 ppm for **3b** and **4b**, respectively, which shows that their relative acidity is similar. However, backbone protons (labelled “c” in Fig. 2, left) are at 5.11 and 5.75 ppm for **3b** and **4b**, respectively. The significant upfield shift for **3b** versus **4b** is due to the proximity to the aryl ring and has been observed by Grisi for similar imidazoliums.⁴³

A typical reaction to append the wingtips was successfully carried out to synthesize 1,3-diisopropyl-4,5-diphenylimidazolium hexafluorophosphate (**5**). Excess 2-iodopropane was refluxed with 4,5-diphenylimidazole under basic conditions followed by an anion exchange with KPF₆ which gave **5** in 86% yield as a pale-yellow powder (Scheme 1).⁴⁴ Notably, there is no peak (labelled “c” in Fig. 2, left) between 5 and 6 ppm in **5** due to the absence of backbone protons in the ¹H NMR that is observed in **3b** and **4b**.

Single crystal X-ray diffraction was necessary to deduce the absolute stereochemistry of **3b** and **4b**. Crystals for **3b** cracked upon cooling, so this structure was determined at room temperature. Fig. 2A (right) shows the structure of **3b** with the view from the back of the molecule, which clearly demonstrates its C₂ symmetry. Likewise, **4b** displays C_s symmetry as the protons are on the same side of the ring (Fig. 2B, right). **3b** crystallizes in the chiral space group P2₁ while **4b** crystallizes in P $\bar{1}$. Finally, the crystal structure for compound **5** shows the expected C_{2v} symmetry and no protons on the back of the ring (Fig. 2C, right).

The NHC-CO₂ adducts for all three compounds, **3b**, **4b**, and **5**, were prepared by deprotonation with potassium bis(trimethylsilyl)amide in THF followed by addition of excess CO₂ gas (Scheme 1).^{45–47} After the completion of reaction, the THF was removed and the solids were extracted with pentane. The pentane was removed under vacuum to yield white solids. **6**-CO₂ was synthesized in 41% yield and the downfield imidazolium proton for **3b** is not observed in the ¹H NMR (see ESI†). Instead, an additional resonance at 164.89 ppm is observed in the ¹³C NMR which is due to the bound CO₂.^{47,48} Similar results were obtained for the synthesis of **7**-CO₂ and **8**-CO₂. The formation of these NHC-CO₂ adducts was further validated by the presence of CO₂ stretches in the FTIR spectra at 1672, 1672, and 1668 cm⁻¹ for **6**-CO₂, **7**-CO₂, and **8**-CO₂, respectively (see ESI†).²²

The deposition of the NHCs on gold mirrors followed a protocol that we previously developed for gold film-over-nanospheres (AuFONs).²³ The gold mirror was placed in a vacuum oven with the solid powders placed directly on top. The vacuum oven was heated to at least 105 °C and placed under reduced pressure (<5 Torr) for 12 minutes. The gold mirror was removed from the oven and then rinsed with alcohol, water and then dried with dinitrogen gas. SERS spectra were

collected for gold mirrors functionalized with **6**, **7**, and **8** (Fig. S51†). The resulting spectra illustrate the adsorption of each of these ligands on the gold surface and little to no spectroscopic differences were observed, which is distinct from our previous reports with benzimidazole-based NHCs.^{21,22} However, the relative differences in SERS intensities may arise from several different variables, such as the molecule orientation relative to the gold surface, number of molecules in the probe volume, and the Raman cross section of each NHC ligand.^{22,49,50}

LDI-MS is a highly efficient method for detecting NHCs on gold and for tracking subsequent chemical reactions.^{51–53} The reason that this approach is so successful is that the NHCs do not fragment like thiols, and each gold ion binds two NHCs to form a cationic bis(NHC)gold(I) complex.⁵¹ LDI-MS analysis was carried for **6**, **7**, and **8** on gold mirror. LDI-MS for **6** shows the expected [(**6**)₂Au]⁺ ion at 809.37 *m/z* (Fig. 3, bottom). However, the corresponding measurement for **7** was quite surprising. In this case, three separate ions were observed in this region at 805.35, 807.36, and 809.38 *m/z* (Fig. 3, middle). One possibility is that NHC **7** is losing backbone protons and becomes unsaturated, which would transform **7** into **8**. A comparison to the LDI-MS of **8** supports this supposition since only one ion is observed at 805.34 *m/z* (Fig. 3, top). A pair of LDI-MS experiments with **7**-Au at 0 ns or 110 ns delay time confirm that the dehydrogenation is not a result of the LDI-MS measurement (Fig. S52†). Since very few studies of NHCs on gold have been performed with saturated NHCs, it is difficult to say how this reaction takes place.^{54,55} Nevertheless, an additional piece of evidence is the small feature found at 1290 cm⁻¹ on the SERS spectrum for **7** (Fig. S48†). This band may be due to the formation of **8** on the surface but since **8**

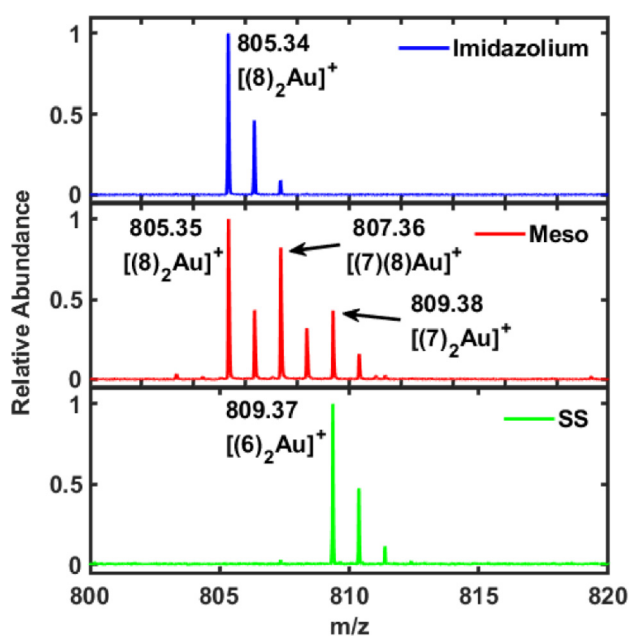


Fig. 3 LDI-MS for **6** (SS, bottom), **7** (*meso*, middle), and **8** (Imidazolium, top) on gold mirrors. Surprisingly, NHC **7** appears to transform into **8**, *i.e.*, become unsaturated, once bound to the gold.



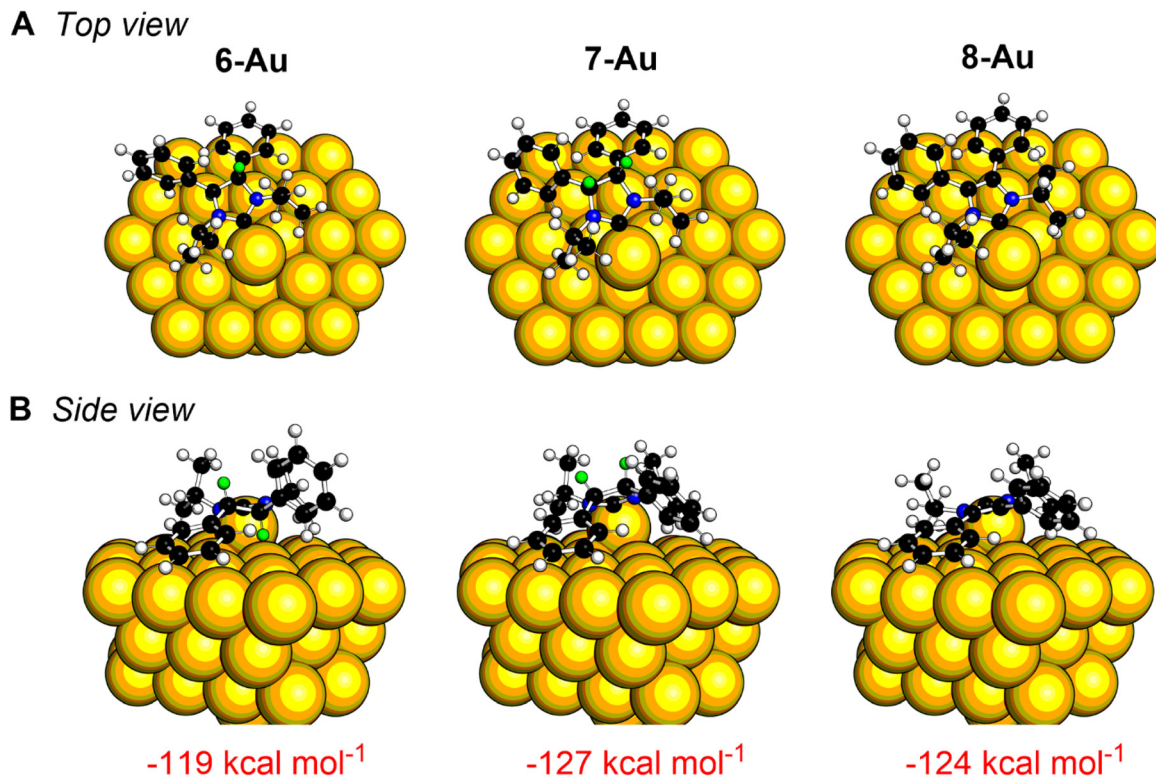


Fig. 4 Binding energies (BE) of NHCs in their energetically favored flat-lying geometry on gold clusters calculated with DFT. (A) represents top view of the NHCs (B) represents the side-view. Black, blue, white, and gold, spheres represent C, N, H, and Au atoms, respectively. The green spheres represent hydrogens binds to chiral carbons.

has much weaker SERS intensity, even if most of 7 is transformed it gives a small signal.

Theoretical calculations can assist in understanding the different reactivity and binding on the gold surface between 6 and 7. Binding energies were calculated for each NHC in both vertical and flat motifs on 58-atom gold clusters.^{52,56} As shown in Fig. 4, the flat binding mode is significantly more energetically favorable for each NHC (with the caveat that only one NHC is modelled on the surface, which precludes NHC–NHC interactions). Of particular note, the flat binding geometry for NHC 7 with the backbone protons pointing away from the gold surface is more favorable by 17 kcal mol⁻¹ energy than the one with protons aimed towards the gold surface. In contrast, for NHC 6, one proton must be pointed towards the gold surface in the flat position since the NHC is *C*₂ symmetric. While future experiments are needed to unravel the mechanism of this dehydrogenation reaction, the variance in reactivity can clearly be accounted for by the differences in stereochemical relationship of the NHCs to the gold surface.

Conclusions

In conclusion, we have synthesized three NHCs which have three different symmetries (*C*₂, *C*_s, and *C*_{2v}) and bound them to gold mirrors *via* their CO₂ adducts. The different stereo-

chemistry led to key differences in the ¹H NMR and their absolute configuration was determined by single crystal X-ray diffraction. This study is the first example where NHCs with a chiral backbone have been affixed to a gold surface. Once bound to the gold, all three NHCs were evaluated with SERS and LDI-MS. Notably, the *C*_s NHC (7), but not the chiral *C*₂ NHC (6), partially degrades to the independently synthesized unsaturated NHC (8) upon binding to the gold. Theoretical calculations assist in explaining this phenomenon by showing that all three NHCs primarily lie flat on the gold surfaces, which exposes the backbone protons on 7 but not on 6.

Experimental

Materials and general considerations

All the reactions and workups for synthesizing organic compounds were done under air unless otherwise stated. All the reactions and workups to synthesize NHC–CO₂ adducts were conducted under dry N₂ atmosphere using standard Schlenk techniques or a glovebox. All the glassware for air and water-sensitive reactions were dried in oven at 170 °C overnight prior to their use in glovebox or on Schlenk line. The solvents used under N₂ were dried on an Innovative Technologies (Newburyport, MA) Pure Solv MD-7 solvent purification system, degassed by three freeze–pump–thaw cycles on a Schlenk line



to remove dioxygen, and stored over activated 4 Å molecular sieves prior to use. Celite was dried at 240 °C under vacuum overnight and stored in the glove box. Compounds **1**, **2**, **3a**, **3b**, **4a**, **4b**, **5**, **6-CO₂**, **7-CO₂**, and **8-CO₂** were synthesized by employing modifications from previously published procedures that reported synthesis of similar compounds.^{41,42,44–47} (1*S*,2*S*)-(–)-1,2-Diphenylethylenediamine and *meso*-1,2-diphenylethylenediamine were purchased from Combi-Blocks and Ambeed, respectively, at highest available purity and used as such without further purification. All other reagents were purchased from typical commercial vendors and used without any further purification.

General instrumentation

Solution ¹H NMR and ¹³C{¹H} NMR spectroscopies were performed on Varian VNMRS 500 MHz and Bruker AVANCE NEO 500 MHz. The ¹H NMR spectral peaks were referenced to the residual protonated solvents. The ¹³C NMR spectral peaks were referenced to the solvents. Mass spectrometry analyses of NHC compounds were conducted at the Biological and Small Molecule Mass Spectrometry Center located in the Department of Chemistry at the University of Tennessee. DART analyses were performed using a JEOL AccuTOF-D time-of-flight (TOF) mass spectrometer with a DART (direct analysis in real time) ionization source from JEOL USA, Inc. (Peabody, MA). Matrix-assisted laser desorption/ionization (MALDI) mass analyses were performed using Waters Synapt G2-Si quadrupole time-of-flight mass spectrometer. Infrared spectra were collected on a Thermo Scientific Nicolet iS10 with a Smart iTR accessory for attenuated total reflectance (ATR) using pure samples of each complex.

Crystal structure determination

Single crystal X-ray diffraction (SCXRD) data of all compounds were collected with a Bruker D8 Venture diffractometer with Mo K α radiation ($\lambda = 0.71073$ Å) and PHOTON II detector. The crystal of compound **3b** was mounted on the capillary glass (Hampton Research) with epoxy glue and cooled at 275 K, while the crystals of **4b** and **5** were mounted on loops (MitiGen) with Paratone-N (Hampton Research) in 100 K cold stream provided by an Oxford Cryostream 800 system. The diffraction data were obtained, integrated, and calculated with Bruker APEX4 with SAINT and SADABS, SHELXL, SHELXT, and OLEX2 software. The structural figures were used by DIAMOND software for visualization.

Surface-enhanced Raman spectroscopy (SERS) measurements

SERS spectra were collected on a home-made Raman microscope described previously.^{57–59} Briefly, one 633 nm HeNe laser (Thorlabs) was guided into an inverted microscope (Nikon Ti-U) and then focused onto the mirror-nanoparticle substrate with an objective lens (20 \times , N.A. = 0.5). Scattered light was collected through the same objective, filtered through a Rayleigh rejection filter (Semrock), and fed into a spectrometer (Princeton Instruments Acton SP2300, $f =$

0.3 mm, 1200 g mm⁻¹). The resulting spectra were background subtracted in Matlab and plotted using Igor software.

Laser desorption/ionization mass spectrometry (LDI-MS) measurements

A Bruker UltrafleXtreme MALDI-TOF-TOF mass spectrometer in positive ion, reflection mode with ultra-beam parameters and a 355 nm frequency tripled ND: YAG laser was employed for LDI-MS measurements. The laser power was set to 100% of the total available power with a global attenuator offset of 20%. Red phosphorous clusters were used to calibrate the mass spectrometer with a cubic enhanced fit.

The gold substrates were mounted onto a metal target plate slide adapter (Bruker) and grounded using conductive copper tape. In general, three microliters of red phosphorous suspended in acetonitrile (1 mg mL⁻¹) was deposited directly onto the gold mirrors and used to calibrate the mass spectrometer.⁶⁰

Computational details

All calculations were carried out with Amsterdam Density Functional (ADF) engine^{61,62} from AMS2021⁶³ software package. The Becke-Perdew (BP86) XC functional^{64,65} with dispersion correction Grimme3 BJDAMP⁶⁶ was used. The triple-zeta polarized Slater type (TZP) basis set with large frozen cores from ADF basis set library⁶⁷ was used and numerical quality is set to normal. The scalar relativistic effects were accounted for by the zeroth-order regular approximation (ZORA).^{68,69} Geometry optimization was performed where only carbene molecule and topmost Au atom were relaxed. The structure and normal modes diagrams were plotted using PyMOL.⁷⁰

Synthetic details

Synthesis of (4*S*,5*S*)-1,3-diisopropyl-4,5-diphenylethanediamine (1). (1*S*,2*S*)-(–)-1,2-Diphenylethylenediamine (1.013 g, 4.769 mmol, 1 eq.) was added to a 35 mL thick-walled glass pressure tube. Sodium carbonate (1.264 g, 11.93 mmol, 2.5 eq.), 2-iodopropane (1.19 mL, 11.9 mmol, 2.5 eq.), and acetonitrile (15 mL) were then added. The pressure tube was sealed with Teflon screw cap and stirred at 82 °C for 72 hours. The glass tube was then cooled to room temperature and the solvent was removed using a rotary evaporator. The solid was then extracted using 3 \times 10 mL dichloromethane and then filtered over Celite. The filtrate was concentrated to dryness under vacuum to yield a pale yellow waxy solid. The solid was used without any further purification (0.689 g, 49.1% yield).

¹H NMR (CDCl₃, 500 MHz): δ 7.27 (m, 6H), 7.13 (m, 4H), 3.98 (s, 2H), 2.53 (m, $J = 6.7$ Hz, 2H), 0.91 (d, $J = 6.3$ Hz, 6H), 0.89 (d, $J = 6.3$ Hz, 6H). ¹³C NMR (CDCl₃, 126 MHz): δ 128.39, 128.22, 127.42, 65.29, 45.78, 24.21, 21.84. IR: 3309, 3023, 2958, 1602, 1492, 1465, 1451, 1435, 1379, 1363, 1337, 1277, 1269, 1238, 1170, 1142, 1123, 1069, 1026, 913, 864, 835, 812, 779, 756, 724, 695 cm⁻¹. DART HRMS (m/z): [M + H]⁺: (C₂₀H₂₉N₂)⁺: 297.2328 (found), (C₂₀H₂₉N₂)⁺: 297.2331 (calculated).



Synthesis of meso-1,3-diisopropyl-4,5-diphenylethanediamine (2). meso-1,2-Diphenylethylenediamine (1.00 g, 4.70 mmol, 1 eq.) was added to a 35 mL thick-walled glass pressure tube. Sodium carbonate (1.247 g, 11.77 mmol, 2.5 eq.), 2-iodopropane (1.18 mL, 11.8 mmol, 2.5 eq.), and acetonitrile (15 mL) were then added. The pressure tube was sealed with Teflon screw cap and stirred at 82 °C for 72 hours. The glass tube was then cooled to room temperature and the solvent was removed using a rotary evaporator. The pale-yellow solid was then extracted using 3 × 10 mL dichloromethane and then filtered over Celite. The filtrate was concentrated to dryness under vacuum to get a pale-yellow solid. This crude product was purified *via* column chromatography on silica-gel that had been treated with 1% triethylamine with an eluant of 1 : 1 ethyl acetate and hexanes. Removing the solvent *via* rotary evaporation yielded the pure product as an off-white powder. (0.342 g, 20.2% yield).

¹H NMR (CD₂Cl₂, 500 MHz): δ 7.24 (m, 6H), 7.14 (m, 4H), 3.87 (s, 2H), 2.45 (sept, *J* = 6.4 Hz, 2H), 0.85 (dd, *J* = 7.4, 6.2 Hz, 12H). ¹³C NMR (CD₂Cl₂, 126 MHz): δ 142.49, 128.89, 128.41, 127.47, 66.00, 46.04, 24.52, 22.23. IR: 2964, 1455, 1385, 1282, 1237, 1223, 1162, 799, 758, 703 cm⁻¹. DART HR MS (*m/z*): [M + H]⁺: (C₂₀H₂₉N₂)⁺: 297.2318 (found), (C₂₀H₂₉N₂)⁺: 297.2331 (calculated).

Synthesis of (4S,5S)-1,3-diisopropyl-4,5-diphenylimidazolium chloride (3a). Compound 1 (3.94 g, 13.4 mmol, 1 eq.) was added along with ammonium chloride (NH₄Cl) (0.716 g, 13.4 mmol, 1 eq.) in a 250 mL round bottom flask. Excess triethyl orthoformate (52.3 mL, 314.9 mmol, 23.5 eq.) was added and the mixture was stirred vigorously. 2-3 drops of formic acid were then added. The solution was refluxed overnight at 142 °C. Upon cooling the solution to room temperature, diethyl ether (~50 mL) was added causing a white solid to precipitate. The solid was collected on a medium 30 mL porosity frit and washed with additional diethyl ether (3 × 10 mL) to obtain the pure product as a white powder (3.17 g, 69.0% yield).

¹H NMR (CD₂Cl₂, 500 MHz): δ 10.01 (s, 1H), 7.49 (m, 6H), 7.29 (m, 4H), 4.92 (s, 2H), 3.93 (sept, *J* = 6.8 Hz, 2H), 1.58 (d, *J* = 6.7 Hz, 6H), 1.24 (d, *J* = 6.6 Hz, 6H). ¹³C NMR (CD₂Cl₂, 126 MHz): δ 156.30, 137.21, 130.61, 130.47, 127.50, 73.10, 51.19, 22.03, 21.84. IR: 2975, 2876, 1627, 1495, 1455, 1391, 1370, 1343, 1269, 1228, 1211, 1177, 1118, 1080, 1045, 931, 641, 615, 755, 665 cm⁻¹. MALDI HR MS (C₂₁H₂₇N₂)⁺: 307.2178 (found), (C₂₁H₂₇N₂)⁺: 307.2169 (calculated).

Synthesis of (4S,5S)-1,3-diisopropyl-4,5-diphenylimidazolium hexafluorophosphate (3b). Compound 1 (10.99 g, 37.35 mmol, 1 eq.) was added along with ammonium hexafluorophosphate (NH₄PF₆) (6.088 g, 37.35 mmol, 1 eq.) in a 500 mL round bottom flask. Excess triethyl orthoformate (142.9 mL, 877.7 mmol, 23.5 eq.) was added and the mixture was stirred vigorously. 2-3 drops of formic acid were then added. The solution was refluxed overnight at 142 °C. Upon cooling the solution to room temperature, diethyl ether (150 mL) was added in excess causing pale-yellow solid to precipitate. The solid was collected on a medium 60 mL porosity

frit and washed with additional diethyl ether (3 × 10 mL) to obtain the pure product as a pale yellow solid. (11.18 g, 65.61% yield). Crystals suitable for single crystal XRD were grown by vapor diffusion from THF : Et₂O.

¹H NMR (CD₃CN, 500 MHz): δ 8.24 (s, 1H), 7.50 (m, 6H), 7.34 (m, 4H), 5.11 (s, 2H), 3.54 (sept, *J* = 6.6 Hz, 2H), 1.37 (d, *J* = 6.8 Hz, 6H), 1.20 (d, *J* = 6.6 Hz, 6H). ¹³C NMR (CD₃CN, 126 MHz): δ 154.86, 137.36, 130.90, 130.62, 128.42, 73.13, 50.93, 21.55, 21.09. ¹⁹F NMR (CD₃CN, 470 MHz): δ -73.00 (d, *J* = 706 Hz). ³¹P NMR (CD₃CN, 202 MHz): δ -144.62 (pent, *J* = 706 Hz). IR: 2985, 1725, 1632, 1497, 1457, 1397, 1379, 1270, 1222, 1179, 1120, 755.97, 700 cm⁻¹. MALDI HR MS (C₂₁H₂₇N₂)⁺: 307.2174 (found), (C₂₁H₂₇N₂)⁺: 307.2169 (calculated).

Synthesis of meso-1,3-diisopropyl-4,5-diphenylimidazolium chloride (4a). Compound 2 (2.36 g, 8.02 mmol, 1 eq.) was added to a 250 mL round bottom flask along with ammonium chloride (0.43 g, 8.02 mmol, 1 eq.). Triethyl orthoformate (31.3 mL, 23.5 eq.) was added in excess and the mixture was stirred vigorously. 2-3 drops of formic acid were added, and the mixture was refluxed overnight at 142 °C. 60 mL of diethyl ether was added upon cooling the reaction mixture to room temperature resulting in the formation of white precipitates. The solid was collected over medium 60 mL porosity frit and washed with 3 × 10 mL diethyl ether to obtain pure product (1.59 g, 58.0% yield).

¹H NMR (CD₂Cl₂, 500 MHz): δ 9.42 (s, 1H), 7.14 (m, 6H), 7.00 (m, 4H), 5.89 (s, 2H), 3.76 (sept, *J* = 6.7 Hz, 2H), 1.71 (d, *J* = 6.8 Hz, 6H), 1.37 (d, *J* = 6.6 Hz, 6H). ¹³C NMR (CD₂Cl₂, 126 MHz): δ 156.58, 131.93, 129.43, 129.05, 128.95, 69.30, 50.85, 21.95, 21.85. IR: 2972, 1630, 1583, 1494, 1455, 1393, 1374, 1341, 1268, 1230, 1177, 1129, 1079, 1050, 928, 856, 772, 749, 702 cm⁻¹. MALDI HR MS (C₂₁H₂₇N₂)⁺: 307.2172 (found), (C₂₁H₂₇N₂)⁺: 307.2169 (calculated).

Synthesis of meso-1,3-diisopropyl-4,5-diphenylimidazolium hexafluorophosphate (4b). Compound 2 (1.00 g, 3.39 mmol, 1 eq.) was added to a 100 mL round bottom flask along with ammonium hexafluorophosphate (0.55 g, 3.39 mmol, 1 eq.) and excess triethyl orthoformate (15 mL, 23.5 eq.). 2-3 drops of formic acid were then added. The reaction was refluxed overnight at 142 °C while continuously stirring. Addition of diethyl ether (50 mL) resulted in formation of white precipitates. The white solid was filtered over medium 30 mL porosity frit and was washed (3 × 10 mL) with diethyl ether to obtain pure product (1.16 g, 75.8% yield).

¹H NMR (CD₃CN, 500 MHz): δ 8.30 (s, 1H), 7.15 (m, 6H), 6.99 (m, 4H), 5.75 (s, 2H), 3.49 (sept, *J* = 6.7 Hz, 2H), 1.49 (d, *J* = 6.7 Hz, 6H), 1.23 (d, *J* = 6.5 Hz, 6H). ¹³C NMR (CD₃CN, 126 MHz): δ 156.15, 132.84, 129.74, 129.57, 129.36, 69.35, 50.68, 21.48, 20.96. ¹⁹F NMR (CD₃CN, 470 MHz): δ -73.00 (d, *J* = 706 Hz). ³¹P NMR (CD₃CN, 202 MHz): δ -144.63 (sept, *J* = 706 Hz). IR: 2972, 1632, 1584, 1495, 1455, 1375, 1266, 1178, 1129, 774, 702 cm⁻¹. MALDI HR MS (C₂₁H₂₇N₂)⁺: 307.2169 (found), (C₂₁H₂₇N₂)⁺: 307.2169 (calculated).

Synthesis of 1,3-diisopropyl-4,5-diphenylimidazolium hexafluorophosphate (5). 4,5-Diphenylimidazole (1.00 g,



4.419 mmol, 1 eq.) was added to a 35 mL thick-walled pressurized tube. 2-Iodopropane (8.6 mL, 88.38 mmol, 20 eq.) was then added in excess followed by potassium carbonate (0.916 g, 6.63 mmol, 1.5 eq.) and acetonitrile (5 mL). The pressure tube was sealed with a Teflon cap and was stirred at 120 °C for 72 hours. The glass tube was then cooled to room temperature and the solvent was removed using a rotary evaporator. The brown solid was then extracted using 3 × 30 mL dichloromethane and then filtered over Celite. The filtrate was evaporated to dryness and the brown powder was dissolved in minimum amount of acetonitrile (5 mL). The product was triturated by adding diethyl ether (50 mL) yielding a pale-yellow powder of 1,3-diisopropyl-4,5-diphenylimidazolium iodide (1.64 g, 85.6% yield).

The above yellow powder (0.80 g, 1.85 mmol, 1 eq.) was dissolved in 25 mL of dimethyl sulfoxide in a 1 L Erlenmeyer flask and a stir bar was added to it. Potassium hexafluorophosphate (5.436 g, 29.53 mmol, 16 eq.), DI water (300 mL), and a stir bar was added to another 500 mL beaker. After both the solutions had stirred for 5 min, the KPF₆ solution was added to the solution in first beaker resulting in the formation of white solid. The white solid was additionally washed with 50 mL of DI water and 100 mL of diethyl ether. The product was further dried under vacuum until all the volatiles were removed to obtain pure compound 5 (0.799 g, 96% yield).

¹H NMR (CD₃CN, 500 MHz): δ 8.71 (s, 1H), 7.45 (m, 6H), 7.37 (m, 4H), 4.40 (sept, *J* = 6.7 Hz, 2H), 1.48 (d, *J* = 6.7 Hz, 12H). ¹³C NMR (CD₃CN, 126 MHz): δ 132.90, 132.07, 131.34, 131.24, 129.98, 126.63, 52.14, 23.16. ¹⁹F NMR (CD₃CN, 470 MHz): δ -73.00 (d, *J* = 706 Hz). ³¹P NMR (CD₃CN, 202 MHz): δ -144.61 (pent, *J* = 706 Hz) IR: 3151, 1550, 1466, 1446, 1398, 1381, 1363, 1340, 1254, 1191, 1169, 1077, 1013, 942, 794, 761, 750, 736, 699, 664. MALDI HR MS (C₂₁H₂₅N₂)⁺: 305.2011 (found), (C₂₁H₂₇N₂)⁺: 305.2018 (calculated).

Synthesis of 6-CO₂. Compound **3b** (0.200 g, 0.442 mmol, 1 eq.) was dissolved in a 20 mL vial in 10 mL of THF and solid potassium bis(trimethylsilyl)amide (KHMDs) (0.0881 g, 0.442 mmol, 1 eq.) was added to the vial. The reaction mixture was stirred overnight at room temperature in a glove box. The solvent was then removed under vacuum. The residue was extracted with 15 mL of pentane and filtered using 15 mL medium porosity frit over a short Celite plug into a 50 mL Schlenk flask. The Schlenk flask containing the free carbene solution in pentane was removed from glove box and excess CO₂ gas was added for 15 min under constant stirring. Upon addition of CO₂, the solution turned cloudy, and the white solid formed shortly thereafter. The Schlenk flask was returned to the glovebox and the solid was filtered over fine 15 mL frit and rinsed with pentane. The white solid was dried under vacuum for 5 minutes to yield the product (0.063 g, 41% yield).

¹H NMR (CD₂Cl₂, 500 MHz): δ 7.46 (m, 6H), 7.31 (m, 4H), 4.67 (s, 2H), 4.35 (sept, *J* = 6.8 Hz, 2H), 1.31 (d, *J* = 6.7 Hz, 6H), 0.89 (d, *J* = 6.9 Hz, 6H). ¹³C NMR (CD₂Cl₂, 126 MHz): δ 164.89, 156.93, 139.91, 130.12, 129.85, 126.45, 70.14, 50.77, 22.20,

20.92. IR: 2975, 1672, 1586, 1547, 1517, 1496, 1454, 1392, 1370, 1336, 1291, 1273, 1242, 1169, 1132, 1079, 1030, 1018, 976, 932, 913, 867, 805, 788, 761, 706, 696, 664, 635, 621, 611, 595, 576, 557 cm⁻¹.

Synthesis of 7-CO₂. Compound **4a** (0.200 g, 0.5765 mmol, 1 eq.) was dissolved in 10 mL of THF and KHMDs (0.1150 g, 0.5765 mmol) was added to the reaction vial. The mixture was stirred in the glove overnight at room temperature. The reaction mixture was concentrated under vacuum and the residue was then extracted with 15 mL of pentane. The pentane solution was filtered over a Celite plug into 50 mL Schlenk flask. The flask was brought out of the box and CO₂ gas was passed constantly over 15 minutes under continuous stirring. The solution turned cloudy, and the white precipitates formed soon after. The flask was returned to the glove box and the white precipitates were filtered over fine 15 mL porosity frit. The white solid was dried for a very short time under vacuum to obtain the product (0.093 g, 48.27% yield).

¹H NMR (CD₂Cl₂, 500 MHz): δ 7.14 (m, 6H), 6.95 (m, 4H), 5.33 (s, 2H), 4.10 (sept, *J* = 6.8 Hz, 2H), 1.41 (d, *J* = 6.8 Hz, 6H), 1.13 (d, *J* = 6.8 Hz, 6H). ¹³C NMR (CD₂Cl₂, 126 MHz): δ 167.90, 157.50, 133.37, 129.11, 128.96, 128.64, 67.99, 51.04, 21.73, 20.80. IR: 2981, 2171, 1672, 1588, 1550, 1495, 1456, 1394, 1373, 1326, 1283, 1212, 1163, 1130, 1025, 985, 927, 858, 796, 777, 759, 707, 649 cm⁻¹.

Synthesis of 8-CO₂. Compound **5** (0.200 g, 0.444 mmol, 1 eq.) was suspended in a 20 mL vial in 10 mL of THF and solid KHMDs (0.0886 g, 0.444 mmol, 1 eq.) was added. The reaction mixture was stirred overnight at room temperature in a glove box. The reaction mixture was then filtered using 15 mL medium porosity frit over Celite into a 50 mL Schlenk flask. The Schlenk flask containing the free carbene solution in pentane was removed from glove box and excess CO₂ gas was added for 30 min under constant stirring. Upon addition of CO₂, the solution turned cloudy. The Schlenk flask was returned to the glovebox and the white solid was filtered over fine 15 mL frit. The product was additionally washed with diethyl ether (3 × 10 mL) to obtain a white powder (0.075 g, 48% yield).

¹H NMR (CDCl₃, 500 MHz): δ 7.41 (m, 6H), 7.25 (d, 4H), 4.45 (sept, *J* = 6.8 Hz, 2H), 1.69 (d, *J* = 6.8 Hz, 12H). ¹³C NMR (CDCl₃, 126 MHz): δ 131.63, 130.84, 130.50, 129.35, 125.43, 52.11, 23.27. IR: 2979, 2937, 1668, 1623, 1548, 1463, 1443, 1370, 1298, 1262, 1203, 1120, 1075, 1010, 979, 936, 835, 796, 768, 737, 722, 704, 686, 669 cm⁻¹.

Preparation of gold mirrors

Gold mirrors were prepared in house using a physical vapor deposition system equipped with a quartz crystal microbalance (Nano36, Kurt J. Lesker). Glass substrates were cleaned with piranha acid (caution, extremely hazardous) made by mixing concentrated sulfuric acid (VWR) and 30% hydrogen peroxide (VWR) at a 4 : 1 ratio. Slides were rinsed with copious amounts of water and mounted on the deposition platform. 5 nm of chromium (Kurt J. Lesker) and then 38 nm of gold (Kurt J. Lesker) were deposited *via* thermal evaporation at a base



pressure of $<10^{-5}$ Torr. Gold coated substrates were rinsed with reagent alcohol, then water, and finally blown dry with nitrogen prior to use.

Citrate-capped gold nanoparticle synthesis

60 nm quasi-spherical AuNPs were synthesized according to a modified Frens method.^{71,72} 180 mL of 0.01% by weight tetrachloroaurate trihydrate salt (Sigma Aldrich) was brought to a boil. Then 120 μ L of 1% by weight trisodium citrate (Sigma Aldrich) was added under vigorous stirring. The solution was kept at a boil for 30 minutes. The suspension was then allowed to cool to room temperature slowly. Ultrapure water (>18 M Ω) was generated using a Thermo Fisher Barnstead System in house and used for nanoparticle synthesis. All Glassware used for nanoparticle synthesis was cleaned using Aqua Regia (caution, extremely dangerous) and rinsed with ultrapure water prior to use.

Deposition of NHC-CO₂ adducts onto gold mirrors

NHC-CO₂ adducts were deposited onto gold mirrors in a vacuum oven (National Appliance) at between 105 and 120 °C. Gold mirrors were cut into 12.5 \times 12.5 mm squares using a glass cutting knife and cleaned with 0.5 M sulfuric acid prior to use. Approximately, 1 to 2 mg of the solid powder was placed directly onto the mirror and heated under vacuum (<5 Torr) for at least 12 minutes. The substrates were rinsed with reagent alcohol, then water, and finally blown dry with nitrogen. For SERS analysis, 100 μ L of 60 nm, citrate-capped gold nanoparticles (AuNPs) were deposited onto the gold mirror and allowed to dry overnight. Finally, the substrates were rinsed again with reagent alcohol, then water, and dried with nitrogen. As a control, gold nanoparticles were drop-cast onto a gold mirror and allowed to dry.

Author contributions

G. K. performed ligand synthesis for all NHCs along with their characterization. N. L. D. appended NHCs to gold films and performed LDI-MS measurements. G. H. performed theoretical calculations. P. N. performed single crystal X-ray diffraction analysis. R. L. T. appended NHCs to gold films and performed SERS measurements. S. L. S. performed initial synthesis for chiral compounds. L. J., J. P. C., and D. M. J. designed and supervised the project. Original draft of the manuscript was prepared by G. K. and D. M. J. Review and editing for draft of the manuscript was conducted by G. K., N. L. D., G. H., L. J., J. P. C., and D. M. J.

Conflicts of interest

There are no conflicts to declare.

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References

- 1 S. Rana, A. Bajaj, R. Mout and V. M. Rotello, Monolayer coated gold nanoparticles for delivery applications, *Adv. Drug Delivery Rev.*, 2012, **64**, 200–216.
- 2 R. B. K. C., B. Thapa and N. Bhattarai, Gold nanoparticle-based gene delivery: promises and challenges, *Nanotechnol. Rev.*, 2014, **3**, 269–280.
- 3 M. Falahati, F. Attar, M. Sharifi, A. A. Saboury, A. Salihi, F. M. Aziz, I. Kostova, C. Burda, P. Priece, J. A. Lopez-Sanchez, S. Laurent, N. Hooshmand and M. A. El-Sayed, Gold nanomaterials as key suppliers in biological and chemical sensing, catalysis, and medicine, *Biochim. Biophys. Acta, Gen. Subj.*, 2020, **1864**, 129435.
- 4 K. Nejati, M. Dadashpour, T. Gharibi, H. Mellatyar and A. Akbarzadeh, Biomedical Applications of Functionalized Gold Nanoparticles: A Review, *J. Cluster Sci.*, 2022, **33**, 1–16.
- 5 P. M. Tiwari, K. Vig, V. A. Dennis and S. R. Singh, Functionalized Gold Nanoparticles and Their Biomedical Applications, *Nanomaterials*, 2011, **1**, 31–63.
- 6 M. Koy, P. Bellotti, M. Das and F. Glorius, N-Heterocyclic carbenes as tunable ligands for catalytic metal surfaces, *Nat. Catal.*, 2021, **4**, 352–363.
- 7 G. Pieters and L. J. Prins, Catalytic self-assembled monolayers on gold nanoparticles, *New J. Chem.*, 2012, **36**, 1931–1939.
- 8 A. Gogoi, N. Mazumder, S. Konwer, H. Ranawat, N.-T. Chen and G.-Y. Zhuo, Enantiomeric Recognition and Separation by Chiral Nanoparticles, *Molecules*, 2019, **24**, 1007.
- 9 H. Li, X. Gao, C. Zhang, Y. Ji, Z. Hu and X. Wu, Gold-Nanoparticle-Based Chiral Plasmonic Nanostructures and Their Biomedical Applications, *Biosensors*, 2022, **12**, 957.
- 10 C. Gautier and T. Bürgi, Chiral Gold Nanoparticles, *ChemPhysChem*, 2009, **10**, 483–492.
- 11 Y. Wang, X. Zhao, Z. Yu, Z. Xu, B. Zhao and Y. Ozaki, A Chiral-Label-Free SERS Strategy for the Synchronous Chiral Discrimination and Identification of Small Aromatic Molecules, *Angew. Chem., Int. Ed.*, 2020, **59**, 19079–19086.



- 12 I. I. S. Lim, D. Mott, M. H. Engelhard, Y. Pan, S. Kamodia, J. Luo, P. N. Njoki, S. Zhou, L. Wang and C. J. Zhong, Interparticle Chiral Recognition of Enantiomers: A Nanoparticle-Based Regulation Strategy, *Anal. Chem.*, 2009, **81**, 689–698.
- 13 I. A. Stoian, B.-C. Iacob, J. P. P. Ramalho, I. O. Marian, V. Chiş, E. Bodoki and R. Oprean, A chiral electrochemical system based on l-cysteine modified gold nanoparticles for propranolol enantiodiscrimination: Electroanalysis and computational modelling, *Electrochim. Acta*, 2019, **326**, 134961.
- 14 M. Matsunaga, T. Nakanishi, T. Asahi and T. Osaka, Effect of surface coverage of gold(111) electrode with cysteine on the chiral discrimination of DOPA, *Chirality*, 2007, **19**, 295–299.
- 15 A. V. Zhukhovitskiy, M. J. MacLeod and J. A. Johnson, Carbene Ligands in Surface Chemistry: From Stabilization of Discrete Elemental Allotropes to Modification of Nanoscale and Bulk Substrates, *Chem. Rev.*, 2015, **115**, 11503–11532.
- 16 C. Vericat, M. E. Vela, G. Benitez, P. Carro and R. C. Salvarezza, Self-assembled monolayers of thiols and dithiols on gold: new challenges for a well-known system, *Chem. Soc. Rev.*, 2010, **39**, 1805–1834.
- 17 L. Srisombat, A. C. Jamison and T. R. Lee, Stability: A key issue for self-assembled monolayers on gold as thin-film coatings and nanoparticle protectants, *Colloids Surf., A*, 2011, **390**, 1–19.
- 18 C. A. Smith, M. R. Narouz, P. A. Lummis, I. Singh, A. Nazemi, C.-H. Li and C. M. Crudden, N-Heterocyclic Carbenes in Materials Chemistry, *Chem. Rev.*, 2019, **119**, 4986–5056.
- 19 G. Kaur, R. L. Thimes, J. P. Camden and D. M. Jenkins, Fundamentals and applications of N-heterocyclic carbene functionalized gold surfaces and nanoparticles, *Chem. Commun.*, 2022, **58**, 13188–13197.
- 20 S. Engel, E.-C. Fritz and B. J. Ravoo, New trends in the functionalization of metallic gold: from organosulfur ligands to N-heterocyclic carbenes, *Chem. Soc. Rev.*, 2017, **46**, 2057–2075.
- 21 J. F. DeJesus, M. J. Trujillo, J. P. Camden and D. M. Jenkins, N-Heterocyclic Carbenes as a Robust Platform for Surface-Enhanced Raman Spectroscopy, *J. Am. Chem. Soc.*, 2018, **140**, 1247–1250.
- 22 M. J. Trujillo, S. L. Strausser, J. C. Becca, J. F. DeJesus, L. Jensen, D. M. Jenkins and J. P. Camden, Using SERS To Understand the Binding of N-Heterocyclic Carbenes to Gold Surfaces, *J. Phys. Chem. Lett.*, 2018, **9**, 6779–6785.
- 23 L. M. Sherman, S. L. Strausser, R. K. Borsari, D. M. Jenkins and J. P. Camden, Imidazolium N-Heterocyclic Carbene Ligands for Enhanced Stability on Gold Surfaces, *Langmuir*, 2021, **37**, 5864–5871.
- 24 B. W. Gung, L. N. Bailey, D. T. Craft, C. L. Barnes and K. Kirschbaum, Preparation and Characterization of Two New N-Heterocyclic Carbene Gold(I) Complexes and Comparison of Their Catalytic Activity to Au(IPr)Cl, *Organometallics*, 2010, **29**, 3450–3456.
- 25 B. W. Gung, M. R. Holmes, C. A. Jones, R. Ma and C. L. Barnes, Structure–enantioselectivity correlation in NHC–Au(I) catalysis for 1,6-enynecyclizations, *Tetrahedron Lett.*, 2016, **57**, 3912–3915.
- 26 M. R. Holmes, J. F. Manganaro, C. L. Barnes and B. W. Gung, Synthesis and characterization of novel chiral [(NHC)Au(I)Cl] complexes: Featuring ortho-biphenyl substituents, *J. Organomet. Chem.*, 2015, **795**, 18–24.
- 27 D. Banerjee, A. K. Buzas, C. Besnard and E. P. Kündig, Chiral N-Heterocyclic Carbene Gold Complexes: Synthesis, Properties, and Application in Asymmetric Catalysis, *Organometallics*, 2012, **31**, 8348–8354.
- 28 E. P. Kündig, T. M. Seidel, Y.-x. Jia and G. Bernardinelli, Bulky Chiral Carbene Ligands and Their Application in the Palladium-Catalyzed Asymmetric Intramolecular α -Arylation of Amides, *Angew. Chem., Int. Ed.*, 2007, **46**, 8484–8487.
- 29 L. Benhamou, C. Besnard and E. P. Kündig, Chiral PEPPSI Complexes: Synthesis, Characterization, and Application in Asymmetric Suzuki–Miyaura Coupling Reactions, *Organometallics*, 2014, **33**, 260–266.
- 30 M. Michalak and W. Kośnik, Chiral N-heterocyclic Carbene Gold Complexes: Synthesis and Applications in Catalysis, *Catalysts*, 2019, **9**, 890.
- 31 J. Ren, M. Freitag, C. Schwermann, A. Bakker, S. Amirjalayer, A. Rühling, H.-Y. Gao, N. L. Doltsinis, F. Glorius and H. Fuchs, A Unidirectional Surface-Anchored N-Heterocyclic Carbene Rotor, *Nano Lett.*, 2020, **20**, 5922–5928.
- 32 P. J. Czerwiński and M. Michalak, Synthetic Approaches to Chiral Non-C₂-symmetric N-Heterocyclic Carbene Precursors, *Synthesis*, 2019, **51**, 1689–1714.
- 33 L. Benhamou, E. Chardon, G. Lavigne, S. Bellemin-Laponnaz and V. César, Synthetic Routes to N-Heterocyclic Carbene Precursors, *Chem. Rev.*, 2011, **111**, 2705–2733.
- 34 F. Wang, L.-j. Liu, W. Wang, S. Li and M. Shi, Chiral NHC–metal-based asymmetric catalysis, *Coord. Chem. Rev.*, 2012, **256**, 804–853.
- 35 V. César, S. Bellemin-Laponnaz and L. H. Gade, Chiral N-heterocyclic carbenes as stereodirecting ligands in asymmetric catalysis, *Chem. Soc. Rev.*, 2004, **33**, 619–636.
- 36 D. Janssen-Müller, C. Schleppehorst and F. Glorius, Privileged chiral N-heterocyclic carbene ligands for asymmetric transition-metal catalysis, *Chem. Soc. Rev.*, 2017, **46**, 4845–4854.
- 37 V. Paradiso, V. Bertolasi, C. Costabile and F. Grisi, Ruthenium olefin metathesis catalysts featuring unsymmetrical N-heterocyclic carbenes, *Dalton Trans.*, 2016, **45**, 561–571.
- 38 V. Paradiso, C. Costabile and F. Grisi, NHC Backbone Configuration in Ruthenium-Catalyzed Olefin Metathesis, *Molecules*, 2016, **21**, 117.
- 39 M. Scholl, S. Ding, C. W. Lee and R. H. Grubbs, Synthesis and Activity of a New Generation of Ruthenium-Based



- Olefin Metathesis Catalysts Coordinated with 1,3-Dimesityl-4,5-dihydroimidazol-2-ylidene Ligands, *Org. Lett.*, 1999, **1**, 953–956.
- 40 G. Lovat, E. A. Doud, D. Lu, G. Kladnik, M. S. Inkpen, M. L. Steigerwald, D. Cvetko, M. S. Hybertsen, A. Morgante and X. Roy, Determination of the structure and geometry of N-heterocyclic carbenes on Au (111) using high-resolution spectroscopy, *Chem. Sci.*, 2019, **10**, 930–935.
- 41 J. F. DeJesus, L. M. Sherman, D. J. Yohannan, J. C. Becca, S. L. Strausser, L. F. P. Karger, L. Jensen, D. M. Jenkins and J. P. Camden, A Benchtop Method for Appending Protic Functional Groups to N-Heterocyclic Carbene Protected Gold Nanoparticles, *Angew. Chem., Int. Ed.*, 2020, **59**, 7585–7590.
- 42 F. Guillen, C. L. Winn and A. Alexakis, Enantioselective copper-catalyzed conjugate addition using chiral diamino-carbene ligands, *Tetrahedron: Asymmetry*, 2001, **12**, 2083–2086.
- 43 V. Paradiso, V. Bertolasi and F. Grisi, Novel Olefin Metathesis Ruthenium Catalysts Bearing Backbone-Substituted Unsymmetrical NHC Ligands, *Organometallics*, 2014, **33**, 5932–5935.
- 44 C. O’Beirne, H. T. Althani, O. Dada, J. Cassidy, K. Kavanagh, H. Müller-Bunz, Y. Ortin, X. Zhu and M. Tacke, Novel derivatives of the antibiotic NHC–Ag(I) drug candidate SBC3: Synthesis, biological evaluation and ¹⁰⁹Ag NMR studies, *Polyhedron*, 2018, **149**, 95–103.
- 45 H. Zhou, W.-Z. Zhang, C.-H. Liu, J.-P. Qu and X.-B. Lu, CO₂ Adducts of N-Heterocyclic Carbenes: Thermal Stability and Catalytic Activity toward the Coupling of CO₂ with Epoxides, *J. Org. Chem.*, 2008, **73**, 8039–8044.
- 46 B. R. Van Ausdall, J. L. Glass, K. M. Wiggins, A. M. Aarif and J. Louie, A Systematic Investigation of Factors Influencing the Decarboxylation of Imidazolium Carboxylates, *J. Org. Chem.*, 2009, **74**, 7935–7942.
- 47 S. Naumann, F. G. Schmidt, R. Schowner, W. Frey and M. R. Buchmeiser, Polymerization of methyl methacrylate by latent pre-catalysts based on CO₂-protected N-heterocyclic carbenes, *Polym. Chem.*, 2013, **4**, 2731–2740.
- 48 Z. Kelemen, B. Péter-Szabó, E. Székely, O. Hollóczki, D. S. Firaha, B. Kirchner, J. Nagy and L. Nyulászi, An Abnormal N-Heterocyclic Carbene–Carbon Dioxide Adduct from Imidazolium Acetate Ionic Liquids: The Importance of Basicity, *Chem. – Eur. J.*, 2014, **20**, 13002–13008.
- 49 E. C. Le Ru, E. Blackie, M. Meyer and P. G. Etchegoin, Surface Enhanced Raman Scattering Enhancement Factors: A Comprehensive Study, *J. Phys. Chem. C*, 2007, **111**, 13794–13803.
- 50 P. L. Stiles, J. A. Dieringer, N. C. Shah and R. P. Van Duyne, Surface-Enhanced Raman Spectroscopy, *Annu. Rev. Anal. Chem.*, 2008, **1**, 601–626.
- 51 N. L. Dominique, S. L. Strausser, J. E. Olson, W. C. Boggess, D. M. Jenkins and J. P. Camden, Probing N-Heterocyclic Carbene Surfaces with Laser Desorption Ionization Mass Spectrometry, *Anal. Chem.*, 2021, **93**, 13534–13538.
- 52 N. L. Dominique, R. Chen, A. V. B. Santos, S. L. Strausser, T. Rauch, C. Q. Kotseos, W. C. Boggess, L. Jensen, D. M. Jenkins and J. P. Camden, Ad aurum: tunable transfer of N-heterocyclic carbene complexes to gold surfaces, *Inorg. Chem. Front.*, 2022, **9**, 6279–6287.
- 53 N. L. Dominique, I. M. Jensen, G. Kaur, C. Q. Kotseos, W. C. Boggess, D. M. Jenkins and J. P. Camden, Giving Gold Wings: Ultrabright and Fragmentation Free Mass Spectrometry Reporters for Barcoding, Bioconjugation Monitoring, and Data Storage, *Angew. Chem., Int. Ed.*, 2023, **62**, e202219182.
- 54 O. Schuster, L. Yang, H. G. Raubenheimer and M. Albrecht, Beyond Conventional N-Heterocyclic Carbenes: Abnormal, Remote, and Other Classes of NHC Ligands with Reduced Heteroatom Stabilization, *Chem. Rev.*, 2009, **109**, 3445–3478.
- 55 J. Li, J. Jiao, C. Zhang, M. Shi and J. Zhang, Facile Syntheses of N-Heterocyclic Carbene Precursors through I₂- or NIS-Promoted Amidiniumation of N-Alkenyl Formamidines, *Chem. – Asian J.*, 2016, **11**, 1361–1365.
- 56 R. L. Thimes, A. V. B. Santos, R. Chen, G. Kaur, L. Jensen, D. M. Jenkins and J. P. Camden, Using Surface-Enhanced Raman Spectroscopy to Unravel the Wingtip-Dependent Orientation of N-Heterocyclic Carbenes on Gold Nanoparticles, *J. Phys. Chem. Lett.*, 2023, **14**, 4219–4224.
- 57 T. B. Demille, R. A. Hughes, N. Dominique, J. E. Olson, S. Rouvimov, J. P. Camden and S. Neretina, Large-area periodic arrays of gold nanostars derived from HEPES-, DMF-, and ascorbic-acid-driven syntheses, *Nanoscale*, 2020, **12**, 16489–16500.
- 58 A. S. Preston, R. A. Hughes, N. L. Dominique, J. P. Camden and S. Neretina, Stabilization of Plasmonic Silver Nanostructures with Ultrathin Oxide Coatings Formed Using Atomic Layer Deposition, *J. Phys. Chem. C*, 2021, **125**, 17212–17220.
- 59 Z. R. Lawson, A. S. Preston, M. T. Korsa, N. L. Dominique, W. J. Tuff, E. Sutter, J. P. Camden, J. Adam, R. A. Hughes and S. Neretina, Plasmonic Gold Trimers and Dimers with Air-Filled Nanogaps, *ACS Appl. Mater. Interfaces*, 2022, **14**, 28186–28198.
- 60 L. Kolářová, L. Prokeš, L. Kučera, A. Hampl, E. Peñaméndez, P. Vaňhara and J. Havel, Clusters of Monoisotopic Elements for Calibration in (TOF) Mass Spectrometry, *J. Am. Soc. Mass Spectrom.*, 2017, **28**, 419–427.
- 61 S. J. A. van Gisbergen, J. G. Snijders and E. J. Baerends, A density functional theory study of frequency-dependent polarizabilities and van der Waals dispersion coefficients for polyatomic molecules, *J. Chem. Phys.*, 1995, **103**, 9347–9354.
- 62 E. J. Baerends, T. Ziegler, A. J. Atkins, J. Autschbach, O. Baseggio, D. Bashford, *et al.*, *ADF 2021.204, SCM, Theoretical Chemistry*, 2021, Vrije Universiteit, Amsterdam, The Netherlands, <http://www.scm.com>.
- 63 R. Rüger, M. Franchini, T. Trnka, A. Yakovlev, E. v. Lenthe, P. Philipsen, T. v. Vuren, B. Klumpers and T. Soini, *AMS 2021.204, SCM, Theoretical Chemistry*, 2021, Vrije Universiteit, Amsterdam, The Netherlands, <https://www.scm.com>.



- 64 J. P. Perdew, Density-functional approximation for the correlation energy of the inhomogeneous electron gas, *Phys. Rev. B: Condens. Matter Mater. Phys.*, 1986, **33**(12), 8822–8824.
- 65 A. D. Becke, Density-functional exchange-energy approximation with correct asymptotic behavior, *Phys. Rev. A*, 1988, **38**, 3098–3100.
- 66 S. Grimme, S. Ehrlich and L. Goerigk, Effect of the damping function in dispersion corrected density functional theory, *J. Comput. Chem.*, 2011, **32**, 1456–1465.
- 67 E. Van Lenthe and E. J. Baerends, Optimized Slater-type basis sets for the elements 1–118, *J. Comput. Chem.*, 2003, **24**, 1142–1156.
- 68 E. van Lenthe, J. G. Snijders and E. J. Baerends, The zero-order regular approximation for relativistic effects: The effect of spin-orbit coupling in closed shell molecules, *J. Chem. Phys.*, 1996, **105**, 6505–6516.
- 69 E. V. Lenthe, E. J. Baerends and J. G. Snijders, Relativistic regular two-component Hamiltonians, *J. Chem. Phys.*, 1993, **99**, 4597–4610.
- 70 Schrödinger, LLC, *The PyMOL molecular graphics system, version 2.2.0 2018*.
- 71 G. Frens, Controlled Nucleation for the Regulation of the Particle Size in Monodisperse Gold Suspensions, *Nat. Phys. Sci.*, 1973, **241**, 20–22.
- 72 A. S. Moody, T. D. Payne, B. A. Barth and B. Sharma, Surface-enhanced spatially-offset Raman spectroscopy (SESORS) for detection of neurochemicals through the skull at physiologically relevant concentrations, *Analyst*, 2020, **145**, 1885–1893.

