

RSC Sustainability

Accepted Manuscript

This article can be cited before page numbers have been issued, to do this please use: P. S. Mateos, S. Sampaolesi, M. V. Toledo and L. E. Briand, *RSC Sustain.*, 2025, DOI: 10.1039/D4SU00745J.



This is an Accepted Manuscript, which has been through the Royal Society of Chemistry peer review process and has been accepted for publication.

Accepted Manuscripts are published online shortly after acceptance, before technical editing, formatting and proof reading. Using this free service, authors can make their results available to the community, in citable form, before we publish the edited article. We will replace this Accepted Manuscript with the edited and formatted Advance Article as soon as it is available.

You can find more information about Accepted Manuscripts in the [Information for Authors](#).

Please note that technical editing may introduce minor changes to the text and/or graphics, which may alter content. The journal's standard [Terms & Conditions](#) and the [Ethical guidelines](#) still apply. In no event shall the Royal Society of Chemistry be held responsible for any errors or omissions in this Accepted Manuscript or any consequences arising from the use of any information it contains.

Sustainability spotlight

View Article Online
DOI: 10.1039/D4SU00745J

The depletion of fossil feedstocks and the CO₂ emissions is turning the governments and the research community towards the upgrading the industrial processes to more eco-friendly chemo-biological based technologies. The use of plastic and biomass wastes as biorefinery feedstocks represents an unlimited and ubiquitous alternative that can be adapted to each country, region and climate's availability of renewable resources. In particular, the biobased valorization of mixed wastes such as, plastic and textiles, plastic and biomass, and food wastes and plastics, are emerging research fields that needs further development for industrial application.



Thermochemical and Chemo-biological Molecular Recycling of Plastic Waste and Plastic-Biomass Waste Mixtures: An Updated Revision

View Article Online

DOI: 10.1039/D4SU00145J

Plastic Waste and Plastic-Biomass Waste Mixtures: An Updated Revision

Paula S. Mateos, Sofía Sampaolesi, María Victoria Toledo and Laura E. Briand*

Centro de Investigación y Desarrollo en Ciencias Aplicadas “Dr. Jorge J. Ronco” CINDECA, CCT La Plata-CONICET, UNLP, CICpBA, Calle 47 N° 257, B1900AJK La Plata, Buenos Aires, Argentina. *E-mail: briand@quimica.unlp.edu.ar*

Massive amounts of plastic and biomass waste is mismanaged worldwide, causing detrimental consequences to the human health and the environment. In fact, the disposal of residues through landfill without further processing and burning for household heating and cooking are common practices. Thermochemical processing such as pyrolysis, chemical depolymerization and bioprocessing, prove feasible for recovering valuable building block molecules from plastic residues. The main goal of pyrolysis is to obtain aliphatic hydrocarbons useful as fuel, while chemical processing conducts to constitutive molecules of the plastic (i.e., monomers and polyols) able to be repolymerized and reinserted in the market. On the other hand, the bioprocessing of plastic waste requires prior chemical depolymerization in order to unleash the building blocks. Chemo-enzymatic treatment of waste plastic-biomass mixtures is an open challenge due to the diverse composition of the residues, along with the presence of additives and contaminants. The few reports found in the literature regarding the bioprocessing of plastic residues with lignocellulosic biomass and paper, indicate that a chemical pretreatment cannot be avoided and that some substances present in the residues can act as fermentation inhibitors that affect waste bioprocessing.

11

12

13

14

15

16

17

18

19

20

21

22

23

24

25

26

27

28

29

30

31

32

33

34

35

RSC Sustainability Accepted Manuscript



Sustainability spotlight

The depletion of fossil feedstocks and CO₂ emissions is turning the governments and the research community towards upgrading the industrial processes to more eco-friendly chemo-biological based technologies. The use of plastic and biomass wastes as biorefinery feedstocks represents an unlimited and ubiquitous alternative that can be adapted to each country, region and climate's availability of renewable resources. In particular, the biobased valorization of mixed wastes, such as plastic combined with textiles residues, biomass or food waste, are emerging research fields that needs further development for industrial application.

1. Motivation and outline of the review

The development of novel technology towards the conversion of wastes in valuable substances is at the cutting edge of the scientific community's interest. Nevertheless, attempts to process complex mixtures of wastes have been assessed only recently. The most recent advances in the valorization of residues will be addressed in this review with special attention on the chemical, catalytic and biological treatment of mixtures of various types of plastics and mixtures of plastic and biomass wastes. This overview of cutting-edge processes for mixed waste valorization provides niches and opportunities for further advancement of the research devoted to solve the billions of tons of wastes that are dumped around the world.

Before discussing the more recent investigations concerning the valorization of plastics and plastic-biomass waste mixtures it is important to present fundamental aspects regarding the magnitude of the problem in terms of quantity of plastic waste, the end-of-life management and the emission of greenhouse gases (GHG) as equivalent carbon dioxide (CO₂e).

28

29

30

31

32

33

34

35

36

37

38

39

40

41

42

43

44

45



2. Global plastic residues generation, disposal and environmental impact

View Article Online
DOI: 10.1039/D4SU00745J

47

According to the reports of the International Energy Agency and the World Bank, 44% of the worldwide waste is composed of biodegradable type of residues, such as food leftovers, food industry residues (i.e., potato peel, waste cooking oil, etc.) and green waste that includes tree pruning, grass clippings, branches, wood chips, bark, wood, palm trees and branches, and weeds^{1,2}. Then, 17% of the waste is paper and cardboard and 12% is of plastic origin. Nowadays, plastic residues get worldwide attention due to the debris found in the ocean along with the detection of microplastics in water streams. The Organization for Economic Cooperation and Development (OECD) estimated that this year around 23.5 million tons of macro and microplastic waste leaked to the environment around the world³.

48

49

50

51

52

53

54

55

56

Recently, Cottom *et al.* published a global macroplastic pollution emissions data analysis⁴. Interestingly, the authors defined the term “*pollution emission*” as materials that have moved from the managed or mismanaged system (controlled or contained state) to the uncontrolled or uncontained state, that is released towards the environment. Somehow, this is important in the sense that the analysis is focused in the plastic waste management rather than the amount of plastic produced. The findings of Cottom *et al.* demonstrated that 52.1 million metric tons per year (Mt year⁻¹) of plastic debris are not adequately managed worldwide, that in turn would have an impact in the carbon and environmental footprints. India generates the largest amount of plastic pollution, accounting for 9.3 Mt year⁻¹, followed by Nigeria (3.5 Mt year⁻¹), Indonesia (3.4 Mt year⁻¹) and China (2.8 Mt year⁻¹).

57

58

59

60

61

62

63

64

65

66

Going deeply in the relevancy of proper management of plastic waste, Figure 1 shows the amount of plastic produced per capita and recycled in various countries; the percentage of recycled plastic based on the total amount of plastic wastes is indicated above each column⁵. In this context, South Korea possesses the highest percentage of plastic waste recycling (60%) followed by various European countries, such as Germany, Denmark, Belgium and Norway (48-35%). United States, the larger waste producer per capita (811 kg), recycles only 23%.

67

68

69

70

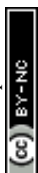
71

72

Landfilling, even though leads to long-term environmental contamination, is the end disposal of 40% of the global plastic waste, as depicted in Figure 2. In addition, 32% goes

73

74



directly to open non-regulated dump sites and only 8% is disposed of in sanitary landfills with gas collection systems^{2,6}. In fact, plastic management is directly related to the socio-economic profile, since low-income countries dump 93% of their plastic (solid) waste. The non-regulated dumping causes the pollution of waterways, that in turn generates marine litter as microplastics, accounting for 11.6-21.1 Mt in the Atlantic Ocean in 2020⁶. In addition, non-regulated dumping goes along with open burning of plastics within all type of solid garbage and the emission of harmful gases and ashes⁷. For instance, bottles made of polyethylene terephthalate (PET) release CO₂, methane, formaldehyde and polycyclic aromatic hydrocarbons; grocery bags, made of high-density polyethylene (HDPE), produce olefins, aldehydes, CO and aromatic compounds; foam cups of polystyrene (PS) generate styrene gas, acrolein, hydrogen cyanide; and curtains, made of polyurethane (PU), release phosgene, among others⁷.

The waste-to-energy incineration (WtE) is the end disposal of 14% of plastic waste. This method involves CO₂ emissions unless a technology for carbon capture and storage (CCS) or carbon capture and utilization (CCU) is applied downwards. Nowadays, only the Netherlands has three operational CCU facilities, one of large scale and two pilot plants⁸. The former has the capability of processing 360.635 ton/year of waste with 60 kt/year of CO₂ capture through absorption. Norway and Japan have also operational pilot WtE-CCU plants.

Rubio-Domingo and Halevi gathered and analyzed various reports of the GHG emissions generated by the plastic end-of-life management option⁹. The authors concluded that landfilling and mechanical recycling have the lowest GHG among the disposal methods. On the other hand, incineration possesses the highest emissions with 1-2.5 kg CO₂e/kg for WtE and 1.8-2.0 kg CO₂e/kg (per kg of plastic) for incineration without an energy conversion associated process. The investigation of Rubio-Domingo and Halevi also considered that the gasification (0.2-1.8 kg CO₂e/kg) and pyrolysis (almost zero emission) are low emission methods. This last method will be further discussed in the following sections due to its low environmental impact and high potential to generate valuable substances.



The term “CO₂e” means CO₂ equivalent, and is used to compare the emissions from various greenhouse gases (GHG) on the basis of their global-warming potential (GWP), by converting amounts of other gases to the equivalent amount of carbon dioxide with the same global warming potential. This concept is related with the carbon and environmental footprints of a product. The former is the total amount of GHG generated along the life cycle of a product. Moreover, the environmental footprint (called also Life Cycle Assessment) involves, not only the GHG emissions, but also the environmental impact caused by: particulate matter emission, human toxicity, ozone depletion, eutrophication, land use, resource depletion, among others.

Zheng and Suh¹⁰ calculated an emission of 1.8 Gt CO₂e of fossil fuel-based plastics along their life cycle in 2015 (see Fig. 2). By 2020, that number raised to 2.2 Gt CO₂e and is projected to grow up 31% by the year 2030, unless mitigation actions took place¹¹. The major contributors to GHG emissions at the resin production stage (most polluting one) are: polypropylene (PP), polyurethane (PU), low-density polyethylene (LDPE), high-density polyethylene (HDPE) and polyethylene terephthalate (PET)¹⁰. In this context, the various strategies of valorization of actual residues based on those type of plastics will be discussed in the following sections.

Carbon footprint is calculated considering that the life cycle of a plastic involves coal combustion for the resin-production stage, which includes all activities from cradle to polymer-production factory gate, accounting for 61% of the total emission. In addition, the conversion stage, which covers the manufacturing processes that turn polymers into final plastic products (30% of the global emission) and the end-of-life stage, which refers to the treatment and disposal processes of plastic waste with 9% of CO₂e emission. Zheng and Suh pointed out that further efforts towards bio-based plastics and renewable energy (wind power and biogas) sources, lowering the demand, and recycling, are the key to diminish the carbon footprint of plastics.

Currently, only 9% of all the plastic waste is recycled. This observation is a driving force towards the development of valorization processes applicable to large amount of plastic



waste. In this context, the following sections discuss the various investigations of thermo-chemical-biological strategies of plastic waste recycling towards valuable platform molecules. 130-131

3. Strategies of valorization of plastic residues towards valuable products 132-133

Nowadays, the mechanical processing is the main route for recycling waste plastics of various sources^{1,12}. This methodology involves the classification of the collected waste according to the polymers' nature and color. Then, it is washed and mechanically ground into a secondary raw material in the form of plastic flakes¹². Then, the flakes are melted (extrusion stage) and filtered to remove impurities. This recyclate that is ready to be reuse in new plastic products is generally of a lower quality than starting virgin-grade plastics mainly due to the changes in the polymer structure during the melting process. 134-141

In contrast, the chemical and biological recycling pursues the breaking down of the polymer into valuable molecules suitable to be converted into new materials. The so-called tertiary recycling of plastics, comprises the pyrolysis and hydrolysis of the wastes^{13,14}. Those processes often involve a sequence of procedures that might begin with the mechanical treatment, followed by a chemical (catalytic or not) process and further biotransformation of the obtained molecules. 142-147

The biological upgrading of that building block molecules uses biocatalysts based on enzymes or microorganisms. Microbial bioprocessing of plastics involves assimilation and mineralization of the carbon degradation products to build more complex molecules. In contrast, the enzymatic treatment produces substances that can be further valorized into second generation products. 148-151

An efficient recycling and valorization of plastic waste is a challenge since there is a large number of different plastics, many of them consisting of a combination of different polymers as well as the presence of additives, such as plasticizers, fillers and reinforcements, thermal stabilizers and antioxidants, colorants, metals, among others. The variety and complexity of their composition is a drawback that traditionally involves multiple processing steps to be overcome. 152-158

159



3.1. Thermochemical (chemical and pyrolytic) based treatments: An overview

Tables 1 to 4 shows a pick of the latest reports on thermochemical and chemical methods that use plastic residues as feedstocks. In particular, the nature of the process, operative conditions, yield, recovery, purification and valorization, have been addressed on the treatment of polyethylene terephthalate¹⁴⁻²², polyethylene²³⁻³⁰, polypropylene³¹⁻³⁸ and polyurethane³⁹⁻⁴⁴ based wastes.

In general, the *pyrolysis* is a thermochemical process carried under an inert gaseous environment (non-oxidative atmosphere) provided by argon or nitrogen. More recently, the use of carbon dioxide has also been investigated as will be discuss later on. The process involves the decomposition of a substrate through heating carried out either in a slow or fast mode, with or without a catalyst, in a batch, fluidized or spouted bed reactor.

The pyrolysis generates a liquid fraction called oil or biooil (if comes from biomass), composed of organic molecules that are lighter than the ones of the feedstock; a non-condensable gaseous fraction and a solid phase composed of a carbonaceous material (char or biochar). It is somehow obvious that the nature of the products of pyrolysis is related primarily to the starving oxygen environment that suppresses the gasification and combustion, preserving the integrity of the organic molecules and avoiding the generation of non-condensable gases. Secondly, the abundance of the liquid, gaseous and solid fractions and their composition depends on the heating rate and the temperature. A slow heating (slow pyrolysis), high residence time of the inert gas with the substrate (5-60 min) and temperatures from 300 °C to 650 °C, conduct to a higher proportion of the solid fraction. Fast heating (fast and flash pyrolysis), low residence time (0.5-1.0 s) and temperatures in the range of 450-600 °C, improve the yield of oil³³.

In turn, the pyrolysis might use a catalytic material in order to direct the decomposition towards targeted reactions and products. For instance, the non-catalyzed pyrolysis of PET



typically yields terephthalic acid, benzoic acid vinyl ester and acetophenone. However, the
pyrolysis of PET waste carpet catalyzed with a basic material such as CaO with steam co-
feeding, conducts to the deoxygenation of PET's oligomers towards benzene¹⁴ (see Table 1,
row 1).

The pyrolysis of low-density polyethylene (LDPE) waste in the presence of a HZSM-11
zeolite yields light aliphatic hydrocarbons in the C₈-C₂₁ range rather than the C₁₈-C₃₅ obtained
in the non-catalyzed process²³. The acidic material catalyzes not only the cracking of
hydrocarbons, but also the dehydrogenation of propane towards propylene (see Table 2, row
1).

In this context, the typical heterogeneous catalysts used in pyrolysis are aluminosilicate
materials such as, zeolites (ZSM-5, ZSM-11, Y, HY)^{14, 23,25-27,33,34} and the commercial catalyst
used in the fluid catalytic cracking (FCC) process of petroleum refinery^{25,27,32,33} (see Table 2,
rows 3 and 5; and Table 3, rows 2, 3 and 4).

Recently, Kanattukara *et al.* published an extensive investigation of the influence of
various catalysts, such as ZSM5, activated alumina, the FCC catalyst and a halloysite nanotube
clay, in the pyrolysis of wastes containing polyethylene (HDPE and LDPE) and polypropylene
(PP)³⁴ (see Table 3, row 4). The catalysts allowed diminishing the temperature of the pyrolysis
from 470 °C to 450 °C, reaction time from 5h to 4h, and improved the yield of the oil fraction
compared with the non-catalyzed pyrolysis. These observations were attributed to the acid
property of the materials that catalyze the cracking of fragments initially produced in the
pyrolysis to even lower molecular weight hydrocarbons. In general, 70-80% of the oil was
composed of olefins, followed by 20% of paraffins and cycloalkanes and a minor content of
aromatic compounds.

Overall, the catalytic pyrolysis is intended to improve the amount of the liquid fraction
towards suitable hydrocarbons to be applied as fuels. More precisely, C₅-C₁₅ hydrocarbons
containing olefins and aromatic compounds, are key pyrolytic products. This goal is achieved
primarily in the catalytic pyrolysis of polyethylene wastes, as shown in Table 2²³⁻²⁷. In some
cases, a tandem pyrolysis, that is two successive pyrolysis, and tandem pyrolysis-steam



cracking was applied in order to further tuning the desired products^{25,27} (Table 2, rows 3 and 5). 218

Typically, pyrolysis is carried out through conventional conduction heating, that is the feedstock is heating up at the surface and then the energy is conducted towards the inner part of the particles. Microwave assisted pyrolysis uses radiation that directly penetrates plastic material (of insulating nature) without absorption or is absorbed by dielectric materials (i.e., biomass) that are heated from the inside out. In this context, Table 2 (see row 4) shows that microwave pyrolysis of polyethylene (LDPE and HDPE) generates a high proportion of oil enriched in hydrocarbons that can be used as fuels²⁶. 219
220
221
222
223
224
225

Another non-conventional heating is plasma pyrolysis. This one provides extreme heat and high temperature (around 1200 °C) in a short period of time conducting to the generation of gases (i.e., CO, H₂ and hydrocarbons) and a low proportion of residues. This process is particularly indicated for the treatment of infectious medical plastic waste³¹ (see Table 3, row 1). 226
227
228
229
230

The pyrolysis of polyurethane (PU) gives rise to harmful aromatic compounds, such as benzene, toluene, aniline, styrene, p-xylene, methylenedianiline (MDA), among others (Table 4, rows 1 and 2). Jung *et al.* reported the pyrolysis of waste PU catalyzed with 5 wt.% Ni/SiO₂ in a N₂/CO₂ environment to convert those chemicals in H₂ and CO (this gas mixture is known as syngas)³⁹. The process was performed in a tandem mode through a pyrolytic reactor followed by a second one containing the catalyst. The slow pyrolysis was carried out between 100 °C to 700 °C at 10 °C/min, while the second reactor was set at 600 °C. 231
232
233
234
235
236
237

A similar approach was used to obtain syngas from the pyrolysis of disposable facemasks, composed of polypropylene, polyethylene and nylon³¹ (see Table 3, row 1). 238
239

The non-catalyzed co-pyrolysis of PU with an epoxy resin also suppresses the decomposition of the methylene diphenyl diisocyanate monomer of PU towards HCN and aromatics (Table 4, row 2). In fact, the investigation of Wu *et al.* suggested that the acid sites of the epoxy resin catalyze the secondary cracking of those substances towards aliphatic hydrocarbons, alcohols, ethers and epoxides at temperatures above 500 °C⁴⁰. 240
241
242
243
244



As observed in the Tables 1 to 4, the pyrolytic process is a commonality in plastic waste treatment since it is suitable to be applied regardless of the nature of the polymeric matrix. Nevertheless, less harsh processes for plastic waste depolymerization, such as hydrolysis, glycolysis, methanolysis, aminolysis, hydrothermal degradation, hydrogenolysis and hydrocracking, have also been investigated.

The *hydrolysis* is the reaction with water at high temperature that is performed under acidic, alkaline or neutral conditions, with or without a catalyst¹⁴. The hydrolysis depolymerizes the plastic waste towards the terephthalic acid (TPA) monomer of PET and polyols in the case of PU. Table 1 (rows 2, 3 and 4) shows that the hydrolysis of waste bottles of PET towards the monomer is highly effective to recover up to 100% of TPA¹⁴⁻¹⁶. Similarly, the acidolysis of PU with dicarboxylic acids produces polyol, amine and esters (see Table 4, row 4)⁴².

Polyethylene is based on the polymerization of ethylene C₂H₂; therefore, the linear alkyl chains of the polymer (C₂H₄)_n do not have polar functions. In turn, PE is rather inert and not suitable for hydrolysis. In the case of polypropylene, the hydrothermal degradation with water in sub and supercritical conditions, at about 450 °C, in an inert gas at high pressure, proved effective to degrade the PP waste towards an oil containing alkanes, alkenes, alcohols, among others^{36,37} (see Table 3, rows 6 and 7). The hydrolysis of ice cream packaging under oxidative conditions, provided by hydrogen peroxide and CO₂, yields mostly acetic acid at a lower temperature (150 °C vs 450 °C) than the process described before³⁵ (Table 3, row 5).

The *glycolysis* comprises the cleavage of the ester bonds of PET with ethylene glycol to release oligomers, dimers and finally the 2-hydroxyethyl terephthalate (BHET)^{20,21} (see Table 1, rows 8 and 9). Typically, heterogeneous catalysts, such as metal (Zn, Mn, Co and Pb) salts, sulfated niobia, ZnMn₂O₄/g-Fe₂O₃, zeolites and silica nanoparticles, are used^{13,20}. More recently, deep eutectic solvents catalysts have been successfully used in the glycolysis of PET waste with an important reduction of the temperature of reaction compared with the heterogeneous catalyzed process²¹ (Table 1, row 9). PU also undergoes glycolysis through the reaction of the urethane group NHCOO with diethylene glycol, releasing a polyol and



carbamate compounds, $R_1NHCOOR_2$. The nature of these last ones depends on the isocyanate that was originally used for synthesizing the polymer. Most frequently, the isocyanate is diphenylmethane-4,4'-diisocyanate, which provides the carcinogenic amine 4,4'-methylenedianiline (MDA) upon PU glycolysis^{41,43,44}. In this context, Donadini *et al.* studied the reaction of MDA with 2-ethylhexylglycidyl ether, acetic anhydride and ethylene carbonate in order to diminish its concentration in the reaction media⁴¹. The deaminated solution was then successfully used to synthesize new rigid PU foam (see Table 4, row 3). Microwave assisted glycolysis of rigid foams made of PU catalyzed with potassium acetate and stannous octoate proved far less time consuming and energy saving than the conventional heating (Table 4, row 5). The combination of the catalysts and diethylene glycol provided a dielectric media for an efficient heating that conducted to PU depolymerization towards polyols along with a low content of the harmful MDA⁴³.

The depolymerization of PET waste through *methanolysis* and *ethanolysis* implies the transesterification with methanol or ethanol at 70 °C to 200 °C, catalyzed with zinc acetate, potassium carbonate or biomass ashes^{14,17-19}. The reaction yields dimethyl terephthalate and diethyl terephthalate, between other substituted monomers containing the terephthalate backbone (see Table 1, rows 5 and 6).

The *aminolysis* involves the reaction with an aqueous solution of a primary amine at 20-200 °C, under an inert environment with the presence of a catalyst or not^{13,14}. This process was investigated in the depolymerization of PET^{13,14,19} and PU⁴². The aminolysis of PET might be carried out with ethanolamine, ethylene diamine, allyl amine, hydrazine hydrate, hydroxylamine hydrochloride or alkyl amine, yielding bis(-2-hydroxyethyl) terephthalamide, bis(2-aminoethyl) terephthalamide; N,N'-diallyl terephthalamide, terephthalic dihydrazide, terephthalohydroxamic acid and N,N'-dialkyl terephthalamide, respectively¹³. In particular, the non-catalyzed solvent-free aminolysis of PET bottles waste, with 1,2-diaminopropane at 130 °C, provided a water-soluble amide of the monomer. This product is suitable to be condensed with salicylaldehyde towards a Schiff base¹⁹ (see Table 1, row 7).

Table 4 (row 4) shows that the aminolysis of PU generates a biphasic system containing an upper phase with a polyol and a bottom one with disubstituted ureas. In this context, the polyols are easily recovered and reuse to synthesize new PU. In addition, the ureas might be further valorize through decomposition in amines and CO_2 ⁴².

View Article Online
DOI: 10.1039/D4SU00743J

275

276

277

278

279

280

281

282

283

284

285

286

287

288

289

290

291

292

293

294

295

296

297

298

299

300

301

302

303

304



The *hydrogenolysis* is the selective scission of C-C and C-O bonds through reaction with hydrogen. Typically, metal particles in acid or basic media are used as catalysts⁴⁵. Depolymerization of plastic through hydrogenolysis has gained attention since numerous studies demonstrated that polyolefins are successfully converted to short-chain hydrocarbons⁴⁶. At 200-250 °C, liquid products are obtained from polyolefins and the selective alkoxy C-O bond hydrogenolysis of polyesters drives to high yields of the terephthalic acid (see Table 1, row 10).

Table 4 (row 6) shows that the hydrogenation of upholstery PU foam catalyzed with a Mn-complex and t-BuOK, generates methylenedianiline, the corresponding formate and the polyol⁴⁴.

Hydrocracking is a process that converts heavy plastic molecules into lighter molecules by breaking the long polymer chains in the presence of hydrogen with a bifunctional metal/acid catalyst^{46,47}. Zeolites are often used to catalyze these reactions due to their strong acidity, high thermal and hydro-thermal stability and regeneration capacity. Furthermore, the porosity of zeolites allows accessibility of certain reactants to the reaction sites, leading to high selectivity⁴⁸. Efficient hydrogen transfer from the hydrogen donors to the polyolefins is crucial to this process. Typically, the reaction temperature is 150-450 °C and the hydrogen pressure is between 20 and 100 bar. Hydrocracking of PE or PP with a bifunctional metal/acid catalyst can achieve a liquid yield over 60% and the distribution of hydrocarbons in liquid fuels will depend on the metal and acid sites of the catalyst^{49,50} (see Table 2, rows 7 and 8 and Table 3, rows 8 and 9).

3.2. Chemo-biological valorization of plastics residues

The section above presented the investigations regarding the chemical and thermochemical methods for the valorization of plastic wastes that were reported in the past 5 years. In comparison, biodegradation is a more environmentally friendly approach, since bioprocesses use milder reaction conditions, such as room temperatures and mid-range pH. More interesting, they avoid or minimize the use of biologically incompatible -toxic- reagents. Interaction of bacteria and fungi with plastic waste has been successfully applied to degrade the polymers into shorter low-molecular weight chains, either under aerobic or anaerobic conditions^{51,52}. Till today, three different strategies have been explored, including fungal,



bacterial (isolates and consortia) and enzymatic biodegradation (native and bio-engineered isolated enzymes). Each of them is best suited to depolymerize different post-consumer plastics, as we will review in this section.

Combined strategies that valorize plastic waste through both chemical and biotechnological methods were designed to overcome the challenges of deconstructing highly crystalline polymers, such as PET or PU. Table 5 gathers reported tandem processes involved in the treatment of plastic wastes to recover valuable building block molecules. In addition, the biological and enzymatic based technologies developed for further valorization of the recovered substances are summarized and illustrated in the Figures 3A and 3B. Chemo-enzymatic treatment of textile waste based on polyesters was investigated by Quartinello *et al.*⁵³. As a first step, the plastic waste was hydrolyzed in an aqueous environment at 250 °C and 40 bar (neutral hydrolysis), which depolymerized 85% of the PET fibers into terephthalic acid and oligomers (see Table 5, row 1, Fig. 3A). This energy-consuming pre-treatment was necessary to degrade highly crystalline fractions of PET into oligomers that enzymes can catabolize. These last ones were further hydrolyzed with 1-2 mg mL⁻¹ of *Humicola insolens* cutinase, an enzyme from the α/β hydrolase family, at pH 7, 50 °C, for 6 h. The overall process provides 97% of terephthalic acid (TPA).

Going even further in the biorefinery concept, Shingwekar *et al.* developed a two-step process for the depolymerization of post-consumer PET bottles to 92 wt% of bis(2-hydroxyethyl) terephthalate (BHET)⁵⁴. The glycolysis performed at 190 °C for 1.5 h was catalyzed with ashes obtained from orange peel (Table 5, row 2, Fig. 3A), yielding a biocompatible mixture rich in BHET suitable for biological degradation, since the crystalline fraction of PET was eliminated. A native consortium of five bacteria strains, investigated previously by León-Zayas *et al.*, was isolated from soils polluted with petroleum products, taking advantage of the rapid adaptation and evolution capacities of bacteria⁵⁵. Composed of *Bacillus thuringiensis* C15, *Bacillus albus*, *Pseudomonas* sp. B10, *Pseudomonas* sp. SW136 and *Pseudomonas* sp. PFYNo1, the consortium uses more readily BHET than PET, synergistically degrading the glycolysis products by 62.63% in 2 weeks towards TPA and ethylene glycol (EG). The overall 2-step process stands as a sustainable methodology to valorize plastic waste without using toxic or costly reagents.



In a similar approach, Kim *et al.* investigated the chemical glycolysis and enzymatic hydrolysis of PET cup waste followed by the biological transformation of TPA towards catechol⁵⁶. Altogether, the cascade-type process comprises the glycolysis of the polymer to BHET and mono(2-hydroxyethyl) terephthalate (MHET) with potassium carbonate K₂CO₃ catalyst, followed by the enzymatic degradation of BHET, MHET and oligomers to TPA (Table 5, row 3, Fig. 3A). Four commercial esterases from *Bacillus subtilis*, *Paenibacillus barcinonensis* and *Rhizopus oryzae*, *Methylobacterium populi* were assayed. This last one catalyzed a complete depolymerization of BHET at a loading of 41.8 µg mL⁻¹, 30 °C in 10 h. Finally, the biotransformation of terephthalic acid to catechol was achieved with a bioengineered strain of *Escherichia coli* at 30 °C for 20 h. Addressing the complete biorefinery concept, the authors proposed a PET upcycling using the catechol as a coating agent directly from the previous step, by simply removing *E. coli* cells without further purification. The catechol coating provided the scaffolding to further functionalization of different materials, with a broad range of applications.

Previous work of Yoshida *et al.*, reviewed by Blank *et al.*, supply the bases for more recent studies that engineer bacteria with a set of genes codifying PET-degrading enzymes⁵⁷. The authors isolated the bacteria *Ideonella sakaiensis* 201-F6, capable to degrade PET and assimilate its monomers, and characterized the specific enzymes involved with that unusual feature, namely PETase and MHETase. The enzymes have the potential to be improved by genetic manipulation and to achieve PET depolymerization at milder temperatures and biologically relevant conditions.

A catalyzed glycolysis of model PET (not from a waste source) coupled with biological upgrading of BHET towards β-ketoadipic acid (β-KA, monomer of nylon 66) was reported by Werner *et al.*⁵⁸. In this case, degrading enzymes from *I. sakaiensis* were used to transform *Pseudomonas putida* KT2440, obtaining a strain capable of harnessing EG by constitutive expression of native genes and BHET by heterologous expression of PETase, MHETase, TPA transporters and enzymes for TPA conversion to protocatechuate (PCA). The strain further converts BHET into the β-ketoadipic acid with a molar yield of 76% at 30 °C, pH 7 in 96 h of fermentation. This scheme of chemical depolymerization coupled with biodegradation proved to be yield-efficient, but improvement of biocompatibility of the glycolysis products is necessary to achieve process fluency and scaling. Valorizing the EG obtained as a by-product remains a future challenge, since β-KA represses EG utilization by bacteria⁵⁸.



The performance of *P. putida* KT2440 and engineered derivatives on post-consumer PET feedstocks is reviewed next. In this context, Sullivan *et al.* reported the chemical and biological processing of a mixture of high-density polyethylene, polystyrene, and poly(ethylene terephthalate) that are regular components of post-consumer plastics waste⁵⁹. The authors used expanded polystyrene cups, milk containers made of polyethylene, and single-use beverage bottles (Table 5, row 4, Fig. 3A). At a first step, the mixture was subjected to auto-oxidization and depolymerization through a catalyzed process which conducts to a random type of chain scissoring. The process, carried out typically with manganese/copper containing catalysts and N-hydroxyphthalimide as oxidation promoter, was performed at 180-200 °C for 5.5 h. As a result, benzoic acid, dicarboxylic acids and terephthalic acids were produced, with substantial energy consumption and need for wastewater treatment. An important observation highlighted by the authors was the fact that the catalytic treatment in an oxygen atmosphere, unlike a typical pyrolysis, generates a mixture of products with enhanced water solubility, suitable for biological fermentation. In this context, the authors used *P. putida* genetically engineered for the bioconversion of acetate, C4 to C17 dicarboxylates, benzoate, and terephthalate to polyhydroxyalkanoate, that is a natural polyester. In addition, these remarkable strains were designed to use acetate and dicarboxylates as a carbon source for cellular growth, while converting benzoate and terephthalate to β -ketoacid, a monomer of polymers. The authors called that downstream valorization a “biological funneling”, since the various molecules produced in the chemical treatment were bio-transformed to only two building block molecules. This approach enables the treatment of blends of plastic residues without previous sorting, which makes it cost-effective.

In the case of polyethylene (PE), it is interesting to discuss the investigation of Li *et al.* who isolated the marine bacteria *Microbulbifer hydrolyticus* IRE-31, capable to degrade the recalcitrant low-density polyethylene (LDPE)⁶⁰. The bacteria, found in the wastewater of a lignin-rich pulp mill, were able to oxidize the surface of linear LDPE, monitored by scanning electron microscopy after 30 days of incubation. FTIR analysis brought insights of the unknown metabolic pathways of biodegradation of PE, revealing the formation of additional hydroxyl and carbonyl functional groups at the polymer surface, meaning that oxidative reactions may be the initial step for depolymerization. Nevertheless, the products of degradation were not informed.



A study by Shao *et al.* reports the biodegradation of un-treated PE with the native strain *Streptomyces albogriseolus* LBX-2 isolated from soil⁶¹. The authors highlight the importance of using microorganisms that can degrade virgin PE, avoiding the photo- and thermal pre-treatments. It has been proposed that the alkane hydrolase system, particularly alkane monooxygenase, is involved in PE degradation, supported by the similarities in the chemical structures of these substances. Arguing this hypothesis, the authors found 21 monooxygenase genes in the genome of *S. albogriseolus* LBX-2, while other bacterial genomes commonly harbor a few. Further genomic, transcriptomic and metabolic studies of the novel strain are needed to design rational biodegradation processes.

Gregory *et al.* reported the catalytic hydrogenolysis of waste polyolefins followed by biotransformation towards an ester wax and alcohols⁶² (see Table 5, row 5, Fig. 3A). Interestingly, the authors isolated a consortium of bacteria (composed of two *Rhodococcus aetherivorans* strains) from LDPE debris found in the soil of a plastic recycling plant. Those bacteria were cultivated in a C₁₀-C₄₀ alkane mixture as a sole carbon source in order to enhance the bioconversion of the PE deconstruction mixture.

The investigations discussed above evidences that most of the research efforts are devoted to the treatment and valorization of PET based wastes. Nevertheless, this kind of plastics represents 10.2% of the global plastic production. But, what about the enzymatic treatment of more recalcitrant and by far less recycled plastics, such as polyurethane (PU)?

The diverse composition and variety of monomers of PU hinders the implementation of chemical depolymerization processes and requires the design of new recycling strategies. As described by Rossignolo *et al.*⁶³, degradation of PU takes place in three steps; namely, breaking of the polymer chains into oligomers; deeper depolymerization towards low-molecular weight species; and conversion to carbon dioxide and water (aerobic conditions) or, alternatively, to methane (anaerobic conditions).

In this context, an early investigation by Schmidt *et al.* demonstrated that enzymatic hydrolysis of model PU and thermoplastic polyester PU (TPU) with various strains of cutinases was achievable⁶⁴. The authors cloned synthetic gene constructs corresponding to polyester hydrolases LC cutinase (LCC), TfCut2, Tcur1278 and Tcur0390 in *E. coli*, expressed and purified the recombinant enzymes that allowed the hydrolysis of solid polyurethane plastic. The



degradation, performed under incubation at 70 °C for 200 h, was concluded from surface depletions and weight loss of PU, but the products of degradation were not reported.

Biological funneling for the upcycling of PU hydrolysates was proposed by Catur Utomo *et al.*⁶⁵. The researchers applied a defined microbial mixed culture composed of microorganisms trained to use specific PU monomers and genetically engineered to yield rhamnolipids. The advantage of this strategy relays in saving time and effort by developing various strains with different metabolic capacities rather than a single strain with multiple biotransformation events or multiple targeted features achieved by directed evolution. While three *P. putida* KT2440 derivatives harboring different genetic optimizations enable the utilization of adipic acid, 1,4-butanediol and EG, the addition of a fourth *Pseudomonas* sp. strain that degrades 2,4-toluene diamine (TDA) was not enough to reduce the inhibition caused by this isocyanate by-product. To overcome the drawback, the authors proposed a chemical removal of TDA from the PU hydrolysate prior to incubation with the microbial consortium. The scheme allows the recovery of valuable TDA and the full utilization of other PU monomers originated from a variety of PU wastes, but some issues regarding biocompatibility of the extractants remains.

The review of Magnin *et al.* pointed out that there is only one investigation about the use of oxidoreductases, such as laccase and a horseradish peroxidase, in the enzymatic degradation of PU⁶⁶. That review dates from the year 2021 and to our knowledge, there is not additional publications on that matter. Magnin *et al.* discovered that the combination of an amidase (E4143) and an esterase (E3576) was capable of hydrolyze model PU films towards 6-hydroxycaproic acid (HCA) and 4,4'-methylene dianiline (MDA)⁶⁷. The researchers developed an interesting strategy, using specifically designed TPU that allows to further understand the molecular mechanisms of enzymatic catalysis. It was postulated a synergistic two-discrete step degradation, by which esterase first attacks the polymer and releases water soluble oligomers containing urethane bond, allowing amidase to better access and exert its urethanase (hydrolase) activity. Finally, the efficient enzymatic system yielded 1 g/L of 6-hydroxycaproic acid and 0.3 to 3 mg/L of MDA and MDA linked to caprolactone. As a limitation, this time-consuming process requires boosting/replacement of the lost enzymatic activity every two to three days during 51 days of incubation.

More recently, the same research group demonstrated that a commercial laccase from *Trametes versicolor* fungi was active in the degradation of model foams, thermoplastic,

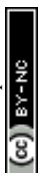


polyester and polyether-based PU incubated in 1-hydroxybenzotriazole at 37 °C for 18 days⁶⁸. Further investigation is needed to test if the combination of the prior studied esterase and amidase with the novel laccase could create an efficient enzymatic system, suitable for different kinds of PU. Moreover, the development of coupled schemes with physical (grinding towards PU powder) or chemical (glycolysis) pre-treatments could enable full degradation of the plastic.

It is worth noticing that the investigations discussed above used model materials which, somehow, denotes the difficulty of the biological recycling of the actual PU waste towards substances suitable for valorization, excepting for those motivated by basic research. In fact, a very recent review by Rossignolo *et al.* pointed out that biodegradation is limited by the number of microorganisms and enzymes able to degrade polyurethanes. In addition, the various structures (polyester PU, polyether PU, among others) and forms (flexible and rigid foams, elastomers, thermoplastic, etc.) would demand a previous PU waste separation to enable an effective upcycling⁶³. In this context, more research is needed to obtain engineered microorganisms harboring the gene constructs necessary to overproduce genetically optimized enzymes. The use of the metagenomic approach to find new enzymatic activities among microorganisms belonging to degrading communities of PU and petroleum derivatives seems to be the next step to overcome these issues.

Such strategy was used by Branson *et al.* in the only investigation that, to our knowledge, reports the chemo-enzymatic recycling of an actual PU waste⁶⁹ (see Table 5, row 6, Fig. 3B). The authors isolated DNA from soil largely exposed to PU residues and developed a metagenome library. The screening for urethanase activity lead to the discovery of three new enzymes, identified as: UMG-SP-1 to UMG-SP-3 with GenBank accession codes: OP972509, OP972510, and OP972511. These enzymes converted 65% of the dicarbamates generated in the glycolysis of polyether-polyurethane waste foam towards aromatic diamines under mild conditions and at room temperature.

The development of tandem processes for plastic recycling is an ongoing effort that requires a critical analysis of the technical feasibility, economic and environmental factors before gone towards a scaling stage. In this sense, the need for waste sorting before treatment is a bottle neck. Even though some bio-funneling strategies has been proposed to overcome this drawback, the development of more integral waste processing approach to



achieve circular economy goals is still a challenge. The few operative technologies that reached that stage of maturity will be further addressed in the section 4 of this review.

3.3. Valorization of plastic and biomass waste mixtures: a synergic effect exists?

The previous sections described the valorization of plastics waste of various sort. This section dives into the chemo-biological treatment of mixtures of plastic and biomass wastes and the effect of combining those major streams of residues. A detailed analysis has been published by Seath *et al.* regarding the synergistic effect of the co-pyrolysis of biomass and plastics to improve the yield and quality of biofuels⁷⁰. In this context, the up-to-date reports (published in the past 5 years) on to the treatment of mixtures composed of plastic waste with textile fabrics (cotton based and synthetic)⁷¹⁻⁷⁶, waste food⁷⁷⁻⁷⁹ and paper wastes⁸⁰ are discussed.

Table 6 summarizes the feedstocks, characteristics of the chemical pretreatment and the bioprocessing of the waste mixture, products obtained and further bioprocessing of them, if applied.

In the particular case of textiles containing PET (with the exception of those containing wool), grinding and a chemical pretreatment (i.e., alkaline hydrolysis) are required to increase the available sites for the biocatalytic saccharification with cellulases. Table 6 (see rows 1 and 2) shows that saccharification of cotton-based wastes is performed through a commercial cocktail of cellulases. This biocatalyst contains endoglucanases that randomly cut cotton cellulose chains; exoglucanases, that act at the ends of the cellulose chain yielding cellobiose; and β -glucosidases, that degrades cellobiose towards glucose^{71,72}. Only one research presents the simultaneous depolymerization of PET and cotton biocatalyzed with cutinases and cellulases, giving rise to terephthalic acid and glucose⁷³ (Table 6, row 1). Unlike other methods, the work reported by Kabel *et al.* uses the minimum amount of liquid just to provide enough moisture for the bioprocessing, but no pre-treatment is performed. Bioethanol is also produced through fermentation with *Saccharomyces cerevisiae* of carbohydrates obtained from textiles^{72,76} (Table 6, rows 1 and 2).

Various types of plastics have also been recently addressed. These include PET microplastics; polyethylene, polypropylene and polystyrene found in films; plastics from disposable bags and food containers and, also, polylactic acid from biodegradable bags.

View Article Online
DOI: 10.3390/S1007431

526
527

528

529

530

531

532

533

534

535

536

537

538

539

540

541

542

543

544

545

546

547

548

549

550

551

552

553

554

555

556

557

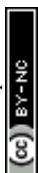


Particularly, complex mixtures of plastics with waste food and sewage sludge, have been treated through anaerobic co-digestion both in mesophilic and thermophilic fashions with bacteria and archaea microorganisms⁷⁷⁻⁷⁹. In this process, organic matter is degraded to form biogas by the action of anaerobic bacteria at temperatures of 30 to 50 °C. The first stage involves acid-forming bacteria that use carbohydrates as raw material. A second step implicates the generation of acetic acid. The last stage comprises the bioconversion of acetic acid, carbon monoxide, and hydrogen in biogas composed mainly of methane.

The mixture of wastes is frequently pretreated before the anaerobic digestion. In this context, Farghali *et al.* reported the pretreatment of 2 m³ of a mixture of plastic films, waste food and rice husks under subcritical water in a large-scale reactor at high temperature and pressure⁷⁷ (see Table 6, row 3). A pretreatment was required to reduce the wastes to liquid and solid fractions containing solubilized lignocellulose. In fact, volatile fatty acids released from biomass through the hydrothermal process served as carbon-source for bacteria, improving the biogas production.

In contrast with the positive effect of biomass-plastic mixture described above, the investigations of Wang *et al.* regarding the co-digestion of PET microplastics with sewage sludge and waste food was not synergistic⁷⁸. In this case, the decomposition of PET into diisobutyl phthalate and dibutyl phthalate resulted harmful to the microbial community of the anaerobic digester (see Table 6, row 4). Indeed, those substances caused a decrease in the amount of hydrolytic bacteria *Bacteroides vadin HA17* and the acidification performed by *Clostridium* and *Sphaerochaeta*.

A similar outcome was obtained by Yu *et al.* in the co-digestion of food waste and biodegradable plastic bags made of polybutylene adipate terephthalate (PBAT)/polylactic acid (PLA)/starch⁷⁹. The authors detected a decline in the biogas production at a bioplastic/waste food ratio above 30% due to a detrimental effect on the bacterial community diversity (see Table 6, row 5). The less effective anaerobic digestion under mesophilic conditions and high plastic loading was attributed to the decrease of the *Synergistota* phylum type of bacteria. Similarly, a decrease in the abundance of bacteria related to the hydrolysis and acidification of organic substances (i.e., *Firmicutes* and *Bacteroidota*) was observed under thermophilic conditions and high plastic loading. On the other hand, the composition of the archaeal community (i.e., *Methanosarcina*, *Methanospirillum*, *Methanothermobacter* and



unclassified_k_norank_d_Archaea) was not influenced by the proportion of bioplastics mixed with food waste.

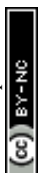
Finally, Brown *et al.* investigated the co-fermentation of various plastics (PET, polypropylene, low-density and high-density polyethylene, polystyrene, and polyvinyl chloride) and paper waste through enzymatic hydrolysis with the commercial cellulase CTec2[®] 80. The authors demonstrated that the presence of acetic acid in the copy paper acts as an enzymatic inhibitor of cellulases (Table 6, row 6). In this context, an alkaline pre-treatment proved to be effective in removing contaminants and increasing the surface area of the cellulose fibers that in turn, enhances the substrate-enzyme interaction. However, the presence of plastic, ink, and stickies within the mixed paper stream, did not have an impact on the enzymatic hydrolysis of cellulose towards glucose and xylose.

Figure 4 summarizes the synergistic and non-synergistic effect of mixing plastic and biomass wastes. PET is one of the most common plastic wastes polluting the planet, and it is also the most deeply studied in this section. Better yields in glucose and bioethanol production were achieved employing cotton or wool combined with PET as biomass feedstock. Nevertheless, the PET presence was detrimental when methane production from sewage sludge-waste food or glucose generation from paper waste was studied. With regard to polyethylene, polypropylene and polystyrene, there were analyzed combined with paper and waste food, giving only synergistic effect in the anaerobic digestion of rice husks. Finally, plastic residues from biodegradable bags did not present beneficial effect in the anaerobic digestion studied with waste food as raw material.

4. From lab bench to industry: actual high scale processes

Previous sections discussed a variety of investigations dealing with thermochemical and chemo-biological methods to treat plastic waste and mixtures of plastic-biomass waste, in order to obtain valuable substances. This section presents the technologically mature and cost-effective processes that scaled up towards industrial application and are effectively applied to this day.

The report published by the capital investment firm called Closed Loop Partners and our own search in the world wide web performed in January 2024, show that pyrolysis is the first



choice for plastic recycling with 36 companies all over the world. Chemical depolymerization possesses the second place with 19 companies⁸¹. There is only one company called Carbios (located in France) that applies chemo-biological methods. Carbios depolymerizes PET and other polyesters from urban plastic and textile wastes through enzymatic hydrolysis. The process uses an engineered cutinase where a divalent-metal-binding (formed by the side chains of three acidic amino acid residues) site was replaced by a disulfide bridge to increase the thermal stability of the hydrolase. The obtained TPA monomer is recycled to produce new plastic⁸¹⁻⁸³.

In addition, it is worth noticing that the startups: Scindo (London, UK), the University of Portsmouth (USA) and Xampla (Cambridge, UK), developed biological based recycling technologies that are currently at lab scale and moving towards large scale application⁸¹.

5. Conclusions and future perspectives

This review provides insights in the up-to-date research regarding the valorization of the most common plastic type residues and plastic-biomass mixed wastes that account for the major quantity of residues generated worldwide.

The valorization of waste-derived platform molecules for the production of refined chemicals and commodities has been extensively explored in the last decades. Nowadays, most of these compounds are originated from petroleum-based raw materials. However, the depletion of fossil feedstocks and the GHG emissions associated to those energy sources is turning the governments and research community into the obtention of bio-based building blocks, upgrading the industrial processes to more eco-friendly ways.

The use of wastes as biorefinery feedstocks represents an unlimited and ubiquitous alternative that can be adapted to each country, region and climate's availability of renewable resources. Nevertheless, molecular recycling of mixtures of plastic and biomass residues is not an easy task due to the variety of compositions and properties of those wastes, which in turn might be either an advantage, as in the case of the pyrolysis of mixed plastic-biomass residues, or a problem to overcome with an appropriate pretreatment. A remarkable fact is that always a chemical or physical, or both treatments, are required for further application of



either an enzymatic or biological process in order to obtain valuable substances out of the waste. In fact, multiple approaches of pretreatment have been investigated and novel, less harsh ones are ongoing research.

The use of microorganisms as microscopic bioreactors for the production of commodities has been largely exploited by humanity since early times. They have the complex metabolic pathways and biologic machinery to synthesize all kind of biomolecules, including biopolymers and its constituent building blocks. By this approach, with the appropriate selection of the bacteria or fungi to be cultured, bulk mixtures enriched in target biomolecules can be obtained.

This review clearly shows that there is a great deal of research to be done in order to develop reliable chemo-enzymatic bioprocesses to treat and valorize mixtures of wastes. In particular, the biobased valorization of mixed wastes, such as plastic and textiles, plastic and biomass, and food wastes and plastics, are emerging research fields that needs further development for industrial application. In fact, the cutting-edge investigations outlined in this contribution shows a variety of vacancies and therefore, opportunities for the advancement in the field.

Author Contributions

The manuscript was written through contributions of all authors.

Data availability

No primary research results, software or code have been included and no new data were generated or analyzed as part of this review.

Funding

This research was funded by Consejo Nacional de Investigaciones Científicas y Técnicas CONICET of Argentina (project PIP 11220200102016CO and project PIBAA 2872021010 0040CO) and Universidad Nacional de La Plata (project 11X-898).

Conflicts of interest

The authors do not declare conflicts of interest.

View Article Online
DOI: 10.1039/D4SU00743J

651

652

653

654

655

656

657

658

659

660

661

662

663

664

665

666

667

668

669

670

671

672

673

674

675

676

677

678

679

680

681



References

1. I. Johansson, M.E. Giménez, D. Roberts, B. Hoffman, M. Becidan, G. Ciceri, F. Murphy, C. Trois, T. P. Curran, D. Stapf. Material and energy valorization of waste as part of a circular model. Bioenergy Annual Report, Technology Collaboration Programme [internet]. International Energy Agency IEA; 2023. <https://www.ieabioenergy.com/wp-content/uploads/2023/05/Colour-Feature-Article-Annual-Report-2022.pdf> (accessed September 2024).
2. Worldbank.org. What a waste 2.0. A global snapshot of solid waste management to 2050. Trends in solid waste management. Washington (DC): World Bank; 2023. <https://datatopics.worldbank.org/what-a-waste/trends-in-solid-waste-management.html> (accessed September 2024).
3. Organization for Economic Co-operation and Development [internet]. Plastic leakage to the environment. OECD.Stat: OECD environment statistics; 2023. https://stats.oecd.org/viewhtml.aspx?datasetcode=PLASTIC_LEAKAGE_V2_3&lang=en# (accessed September 2024).
4. J. W. Cottom, E. Cook, C. A. Velis, *Nature*, 2024, **633**, 101-108.
5. Global waste index 2022: These are biggest waste producers in the world [internet]. <https://sensoneo.com/global-waste-index/> (accessed September 2024).
6. A. F. J. Tang, S. Yu, Ch. Wang, G. H. Yeoh, W. Y. Teoh, A. C. K. Yip, *npj Mater. Sustain.*, 2024, **3**, article number 2.
7. G. Pathak, M. Nichter, A. Hardon, E. Moyer, A. Latkar, J. Simbaya, D. Pakasi, E. Taqueban, J. Love, *Global Environ. Change*, 2023, **80**, 102648.
8. M. Bertone, L. Stabile, G. Buonanno, *Sustainability*, 2024, **16**, 4117.
9. G. Rubio-Domingo, A. Halevi, "Making plastics emissions transparent", Coalition on Materials Emissions Transparency eds., published in February 2022. <https://ccsi.columbia.edu/sites/default/files/content/COMET-making-plastics-emissions-transparent.pdf> (accessed September 2024).
10. J. Zheng, S. Suh, *Nat. Clim. Change*, 2019, **9**, 374-378.
11. L. Cabernard, S. Pfister, Ch. Oberschelp, S. Hellweg, *Nature*, 2022, **5**, 139-148.
12. H. Jeswani, Ch. Krüger, M. Russ, M. Horlacher, F. Antony, S. Hann, A. Azapagic, *Sc. Total Environ.*, 2021, **769**, 144483.
13. K. Ghosal and Ch. Nayak, *Mat. Adv.*, 2022, **3**, 1974-1992.



14. T. Thiounn and R. C. Smith, *J. Polym. Sci.*, 2020, **58**, 1347-1364. View Article Online
DOI: 10.1039/D4SU00743J
15. H. Chen and H. Hu, *I&EC Res.*, 2023, **62**, 12925-12934. 715
16. P. Pereira, P. E. Savage, ChW. Pester, *ACS Sustain. Chem. Eng.*, 2023, **11**, 7203-7209. 716
17. F. Tollini, L. Brivio, P. Innocenti, M. Sponchioni, D. Moscatelli, *Chem. Eng. Sci.*, 2022, **260**, 117875. 717
718
18. Z. T. Laldinpuii, V. Khiangte, S. Lalhmangaihzuala, Ch. Lalmuanpuia, Z. Pachuau, C. Lalhriatpuia, K. Vanlaldinpuia, *J. Polym. Environ.*, 2022, **30**, 1600-1614. 719
720
19. A. A. Al Otaibi, A. K. D. Alsukaibi, M. A. Rahman, M. Mushtaque, A. Hauqe, *Polymers*, 2022, **14**, 1861. 721
722
20. S. Shirazimoghaddam, I. Amin, J. A. Faria Albanese, N. R. Shiju, *ACS Eng. Au*, 2023, **23**, 37-44. 723
724
21. J. Huang, D. Yan, Q. Zhu, X. Cheng, J. Tang, X. Lu, J. Xin, *Polym. Degrad. Stabil.*, 2023, **208**, 110245. 725
726
22. A. A. Y. Kratish, T. J. Marks, *Angew. Chem. Int.*, 2022, **61**, e202112576. 727
23. N. Lee, J. Joo, K. A. Lin, J. Lee, *Polymers*, 2021, **13**, 1198. 728
24. K. P. Rajan, I. Mustafa, A. Gopanna, S. P. Thomas, *Recycling*, 2023, **8**, 63. 729
25. A. Eschenbacher, R. J. Varghese, M. S. Abbas-Abadi, K. M. Van Geem, *Chem. Eng. J.*, 2022, **428**, 132087. 730
731
26. X. Hu, D. Ma, G. Zhang, M. Ling, Q. Hu, K. Liang, J. Lu, Y. Zheng, *Carbon Res. Conv.*, 2023, **6**, 215-228. 732
733
27. S. W. Kim, Y. T. Kim, Y. F. Tsang, J. Lee, *Sci. Total Environ.*, 2023, **903**, 166789. 734
28. X. Xue, J. Mi, F. Huang, J. Wang, L. Chen, J. Liang, *J. Anal. App. Pyrol.*, 2023, **170**, 105909. 735
29. Z. Chen, B. J. Erwin, L. Che, *J. Clean. Prod.*, 2023, **424**, 138861. 736
30. S. Liu, P. A. Kots, B. C. Vance, A. Danielson, D. G. Vlachos, *Sci. Adv.*, 2021, **7**, eabf8283. 737
31. M. M. Harussani, S. M. Sapuan, U. Rashid, A. Khalina, R. A. Ilyas, *Sci. Total Environ.*, 2022, **803**, 149911. 738
739
32. P. Palmay, C. Medina, C. Donoso, D. Barzallo, J. C. Bruno, *Clean Technol. Envir.*, 2023, **25**, 1539-1549. 740
741
33. A. Zabaniotou and I. Vaskalis, *Energies*, 2023, **16**, 593. 742
34. B. V. Kanattukara, J. Singh, P. Sarkar, A. Chopra, D. Singh, S. Mondal, G. Singh Kapur, S. S. V. Ramakumar, *Environ. Sci. Pollut R.*, 2023, **30**, 64994-65010. 743
744



35. V. V. Zefirov, I. V. Elmanovich, A. I. Stakhanov, A. A. Pavlov, S. V. Stakhanova, E. P. Kharitonova, M. O. Gallyamov, *Polymers*, 2022, **14**, 744. View Article Online
DOI: 10.20597/D43U00743J 745
36. M. Čolnik, P. Kotnik, Ž. Knez, M. Škerget, *Polymers*, 2022, **20**, 4415. 746
37. Z. Fu, Y. S. Zhang, G. Ji, A. Li, *Chemosphere*, 2024, **350**, 141045. 747
38. M. S. Al-Iessa, B. Y. Al-Zaidi, R. S. Almukhtar, Z. M. Shakor, I. Hamawand, *Energies*, 2023, **16**, 4871. 748
39. J. M. Jung, T. Lee, S. Jung, Y. F. Tsang, A. Bhatnagar, S. S. Lee, H. Song, W-K. Park, E. E. Kwon, *Chem. Eng. J.*, 2022, **442**, 136358. 749
40. Z. Wu, C. Li, R. Shan, J. Zhang, *Waste Biomass Valori.*, 2024, **15**, 1603-1614. 750
41. R. Donadini, R. Boaretti, L. Scopel, A. Lorenzetti, M. Modesti, *Chem. Eur. J.*, 2023, e202301919. 751
42. B. Liu, Z. Westman, K. Richardson, D. Lim, A. L. Stottleyer, T. Farmer, P. Gillis, V. Vlcek, P. Christopher, M. M. Abu-Omar, *ACS Sustain. Chem. Eng.*, 2023, **11**, 6114-6128. 752
43. R. Donadini, R. Boaretti, A. Lorenzetti, M. Roso, D. Penzo, E. Dal Lago, M. Modesti, *ACS Omega*, 2023, **8**, 4655-4666. 753
44. V. Zubar, A. T. Haedler, M. Shütte, A. S. K. Hashmi, T. Schaub, *Chem. Sus. Chem.*, 2022, **15**, e202101606. 754
45. C.M. Osmundsen, K. Egeblad, E. Taarning, *in New and Future Developments in Catalysis*, ed. Steven L. Suib, Amsterdam, Elsevier, 2013, **4**, 73-89. 755
46. X.B. Wu, K-H. Lin, W-T. Lee, R.C. Turnell-Ritson, P.J. Dyson, P.C.L. Delannoi, *Nat. Commun.*, 2023, **14**, 6524. 756
47. D. Munir, M.F. Irfan, M.F. Usman, *Renew. Sustain. Energy Rev.*, 2018, **90**, 490-515. 757
48. R. Saab, K. Polychronopoulou, L. Zheng, S. Kumar, A. Schiffer, *J. Ind. Eng. Chem.*, 2020, **89**, 83-103. 758
49. P. A. Kolts, B. C. Vance, D. G. Vlachos, *React. Chem. Eng.*, 2022, **7**, 41-54. 759
50. M. S. Al-Iessa, B. Y. Al-Zaidi, R. S. Almukhtar, Z. M. Shakor, I. Hamawand, *Energies*, 2023, **16**, 4871. 760
51. M. Farghali, A. Shimahata, I. M. A. Mohamed, M. Iwasaki, J. Lu, I. Ihara, K. Umetsu, *Biochem. Eng. J.*, 2022, **186**, 108546. 761
52. L.M. Blank, T. Narancic, J. Mampel, T. Tiso, K. O'Connor, *Current Opinion in Biotechnol.*, 2020, **62**, 212-219. 762
53. F. Quartinello, S. Vajnhandl, J. Volmajer Valh, T. J. Farmer, B. Vončina, A. Lobnik, E. H. Acero, A. Pellis, G. M. Guebitz, *Microb Biotechnol.*, 2017, **10**, 1376-1383. 763
54. D. Shingwekar, H. Laster, H. Kemp, J. L. Mellies, *Bioengineering*, 2023, **10**, 1253. 764



55. R. León-Zayas, C. Roberts, M. Vague, J. L. Mellies, *Microbiol. Resour. Announce*, 2019, **8**(25), e00237-19. 779
DOI: 10.1039/C9RU00743J
56. H. T. Kim, M. H. Ryu, Y. J. Jung, S. Lim, H. M. Song, J. Park, S. Y. Hwang, H-S. Lee, Y. J. Yeon, B. H. Sung, U. T. Bornscheuer, S. J. Park, J. Chan Joo, D. X. Oh, *ChemSusChem*, 2021, **14**, 4251-4259. 781
782
783
57. L. M. Blank, T. Narancic, J. Mampel, T. Tiso, K. O'Connor, *Current Opinion in Biotechnol.*, 2020, **62**, 212–219. 784
785
58. A. Z. Werner, R. Clare, T. D. Mand, I. Pardo, K. J. Ramirez, S. J. Hauge, F. Bratti, G. N. Dexter, J. R. Elmore, J. D. Huenemann, V. G. L. Peabody, C. W. Johnson, N. A. Rorrer, D. Salvachúa, A. M. Guss, G. T. Beckman, *Metab. Eng.*, 2021, **67**, 250-261. 786
787
788
59. K. P. Sullivan, A. Z. Werner, K. J. Ramirez, L. D. Ellis, J. R. Bussard, B. A. Black, D. G. Brandner, F. Bratti, B. L. Buss, X. Dong, S. J. Haugen, M. A. Ingraham, M. O. Konev, W. E. Michener, J. Miscall, I. Pardo, S. P. Woodworth, A. M. Guss, Y. Román-Leshkov, S. S. Stahl, T. Gregg, G. T. Beckham, *Science*, 2022, **378**, 207-211. 789
790
791
792
60. Z. Li, R. Wei, M. Gao, Y. Ren, B. Yu, K. Nie, H. Xu, L. Liu, *J. Environ. Manage.*, 2020, **263**, 110402. 793
794
61. H.H. Shao, M.J. Chen, X.T. Fei, R.L. Zhang, Y. Zhong, W.M. Ni, X. Tao, X.Y. He, E.L. Zhang, B. Yong, X.M. Tan, *Microorganisms.*, 2019, **7**, 1–13. 795
796
62. G. J. Gregory, C. Wang, S. Sadula, S. Koval, R. F. Lobo, D. G. Vlachos, E. T. Papoutsakis, *ACS Sustain. Chem. Eng.*, 2023, **11**, 3494-3505. 797
798
63. G. Rossignolo, G. Malucelli, A. Lorenzetti, *Green Chem.*, 2024, **26**, 1132-1152. 799
64. J. Schmidt, W. Ren, T. Oeser, L. A. D. e Silva, D. Breite, A. Schulze, W. Zimmermann, *Polymers*, 2017, **9**, 65. 800
801
65. R. N. Catur Utomo, W.-J. Li, T. Tiso, C. Eberlein, M. Doeker, H.J. Heipieper, A. Jupke, N. Wierckx, L. M. Blank, *ACS Sustain. Chem. Eng.*, 2020, **8** (47), 17466-17474. 802
803
66. A. Magnin, E. Pollet, L. Avérous, *Methods Enzymol.*, 2021, **648**, 317-336. 804
67. A. Magnin, E. Pollet, R. Perrin, C. Ullman, C. Persillon, V. Phalip, L. Avérous, *Waste Manag.*, 2019, **85**, 141-150. 805
806
68. A. Magnin, L. Entzmann, E. Pollet, L. Avérous, *Waste Manag.*, 2021, **132**, 23-30. 807
69. Y. Branson, S. Sörtl, C. Buchmann, R. Wei, L. Schaffert, C. P. S. Badenhorst, L. Reisky, G. Jäger, U. T. Bornscheuer, *Angew. Chem. Int. Ed.*, 2023, **62**, e202216220. 808
809
70. C. C. Seah, C. H. Tan, N. A. Arifin, R. S. R. M. Hafriz, A. Salmiaton, S. Nomanbhay, A. H. Shamsuddin, *Results Eng.*, 2023, **17**, 100989. 810
811



71. Piribauer, A. Bartl, W. Ipsmiller, *Waste Manage. Res.*, 2021, **39**, 1277-1290. View Article Online
DOI: 10.1039/D4SU00745J 812
72. S. M. Gritsch, S. Mihalyi, A. Bartl, W. Ipsmiller, U. Jenull-Halver, R. F. Putz, F. Quartinello, 813
G. M. Guebitz, *Resour. Conserv. Recycl.*, 2023, **188**, 106701. 814
73. S. Kaabel, J. Arciszewski, T. H. Borchers, J. P. D. Therien, T. Friščić, K. Auclair, *ChemSusChem*, 815
2023, **16**, e202201613. 816
74. A. Boondaeng, J. Keabpimai, P. Srichola, P. Vaithanomsat, C. Trakunjae, N. Niyomvong, 817
Polymers, 2023, **15**, 1964. 818
75. J. Egan, S. Wang, J. Shen, O. Baars, G. Moxley, S. Salmon, *Resour. Environ. Sustainability*, 819
2023, **13**, 100118. 820
76. F. Quartinello, S. Vecchiato, S. Weinberger, K. Kremenser, L. Skopek, A. Pellis, G. M. 821
Guebitz, *Polymers*, 2018, **10**, 1107. 822
77. M. Farghali, A. Shimahata, I. M. A. Mohamed, M. Iwasaki, J. Lu, I. Ihara, K. Umetsu, 823
Biochem. Eng. J., 2022, **186**, 108546. 824
78. P. Wang, Y. Guo, M. Yu, S. Riya, Y. Zheng, L. Ren, *Biochem. Eng. J.*, 2023, **198**, 109012. 825
79. C. Yu, B. Dongsu, Z. Tao, K. Zhe, J. Xintong, W. Siqi, C. Ming, S. Zheng, Z. Yalei, *Biochem.* 826
Eng. J., 2023, **199**, 109072. 827
80. R. M. Brown, A. N. Hoover, J. L. Klinger, B. D. Wahlen, D. Hartley, H. Lee, V. S. Thompson, 828
Front Energy Res., 2022, **10**, art. 834832. 829
81. Closed Loop Partners- Investors in the Circular Economy [internet]. B. Miñana and E. Mills. 830
Global Directory of Molecular Recycling Technologies. Supplemental Resource from 831
Transitioning to a Circular System for Plastics: Assessing Molecular Recycling Technologies 832
in the United States and Canada. New York (US): Closed Loop partners; cited 2021 Nov 13. 833
[https://www.closedlooppartners.com/wp-content/uploads/2021/11/CLP_Molecular- 834](https://www.closedlooppartners.com/wp-content/uploads/2021/11/CLP_Molecular-Recycling-Directory-2021.pdf)
[Recycling-Directory-2021.pdf](https://www.closedlooppartners.com/wp-content/uploads/2021/11/CLP_Molecular-Recycling-Directory-2021.pdf) (assessed October, 2024). 835
82. V. Tournier, C. M. Topham, A. Gilles, B. David, C. Folgoas, A. Moya-Leclair, E. Kamionka, 836
M-L. Desrousseaux, H. Texier, S. Gavalda, M. Cot, E. Guémard, M. Dalibey, J. Nomme, G. 837
Cioci, S. Barbe, M. Chateau, I. André, S. Duquesne, A. Marty, *Nature*, 2020, **580**, 216-219. 838
83. Carbios website [internet]. Carbios produces first clear plastic bottles from enzymatically 839
recycled textile waste. Clermont-Ferrand (France): Carbios industries; 2020 (press release 840
2020 Nov 19). <https://www.carbios.com/en/enzymatic-recycling/>(assessed November, 841
2024). 842



No primary research results, software or code have been included and no new data were generated or analysed as part of this review.

