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Chiral and non-conjugated fluorescent salen ligands: AIE, anion probes, chiral recognition of unprotected amino acids, and cell imaging applications†

Guangyu Shen, Fei Gou, Jinghui Cheng, Xiaohong Zhang, Xiangge Zhou  and Haifeng Xiang *

Natural products are usually non-conjugated and chiral, but organic luminescent materials are commonly polycyclic aromatic molecules with extended π -conjugation. In the present work, we combine with the advantages of non-conjugation and chirality to prepare a series of novel and simple salen ligands (41 samples), which have a non-conjugated and chiral (*S,S*) and (*R,R*) cyclohexane or 1,2-diphenylethane bridge but display strong blue, green, and red aggregation-induced emission (AIE) with large Stokes shifts (up to 186 nm) and high fluorescence quantum yields (up to 0.35). Through hydrogen and halogen bonds, these flexible salen ligands can be used as universal anion probes and chiral receptors of unprotected amino acids (enantiomeric selectivity up to 0.11) with fluorescence quantum yields up to 0.29 and 0.27, respectively. Moreover, the effects of different chiral bridges on the molecule arrangement, AIE, and anion and chiral recognition properties are also explored, which provide unequivocal insights for the design of non-conjugated chiral and soft fluorescent materials.

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Introduction

Chirality is a universal phenomenon throughout nature.¹ Not only many biologically active molecules, such as the naturally occurring amino acids and sugars, are chiral, but also many key chemical and biological processes are profoundly influenced by chiral structures. Therefore, chiral recognition² through fluorescence method have attracted increasing attention due to its unique advantages, including high sensitivity, good selectivity, short response time, real-time and parallel monitoring, cell imaging, and *in situ* and non-invasive measurement. Up to date, a lot of different fluorescent chiral receptors have been designed and prepared for enantioselective recognition of chiral amines, acids, or modified amino acids. In the literature, however, there are only a few 1,1'-bi-2-naphthol³ (BINOL)- and tetraphenylethylene⁴ (TPE)-based fluorescent receptors for chiral recognition of unprotected amino acids, because zwitterionic amino acids have not only a poor solubility in organic solvents but also little interaction with organic receptors in water.⁵

Organic luminescent (fluorescent or phosphorescent) materials⁶ are commonly polycyclic aromatic molecules with extended π -electron conjugation, but these rigid materials might suffer some disadvantages, such as the synthesis difficulty, poor solubility, and fluorescence aggregation-caused quenching⁷ (ACQ). On the other hand, non-conjugated soft materials that might have a better solubility, lower cost, higher flexibility, lower cytotoxicity, better biocompatibility, and naturally occurring,⁸ are usually taken for granted as non-emissive materials. Until recently, some non-conjugated materials, such as 1,1,2,2-tetraphenylethane,⁹ poly[(maleic anhydride)-*alt*-(vinyl acetate)],¹⁰ polyisobutene succinic anhydrides and imides,¹¹ and steroidal saponin digitonin,¹² were reported to show amazing ultraviolet (UV) or blue aggregation-induced emission (AIE).^{9,13} Our research work continuously focus on the synthesis, optical properties, and sensing applications of *N,N'*-bis(salicylidene)ethylenediamine (salen), a particular class of salicylaldehyde-based bis-Schiff base (Fig. 1), owing to its facile preparation, good stability, and rich optical properties.^{7b,14} Our recently work demonstrated that step-like salen ligands^{14g} (*R-C_n*) and tripod-like tri-Schiff bases¹⁵ (TSBs, Fig. 1a) linking with non-conjugated long alkyl bridges display strong red-green-blue (RGB) AIE. If chiral (*S,S*) and (*R,R*) cyclohexane or 1,2-diphenylethane bridges are employed, it is easy to prepare chiral salen ligands (Fig. 1b). These non-conjugated chiral ligands are widely used as asymmetric catalysts,¹⁶ and they are deemed to be non-emissive and used as turn-on fluorescence

College of Chemistry, Sichuan University, Chengdu, 610041, China. E-mail: xianghaifeng@scu.edu.cn; Fax: +86-28-8541-2291

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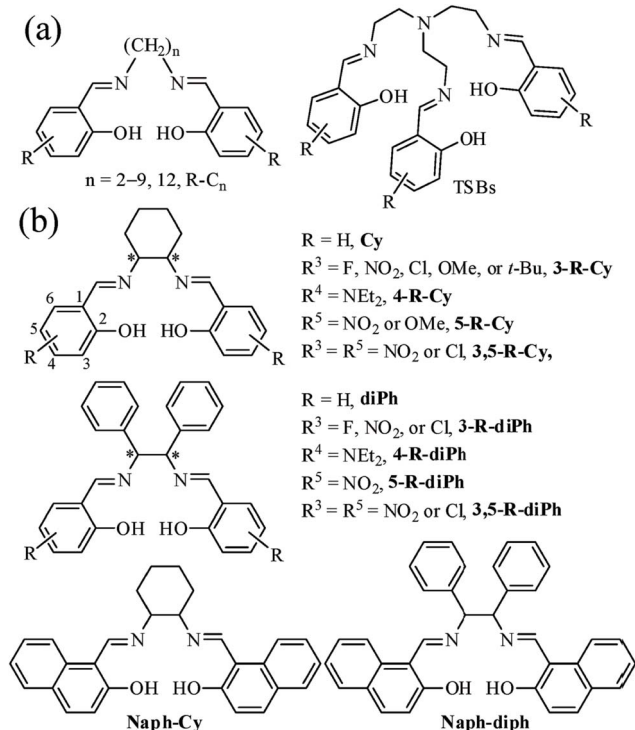


Fig. 1 Non-conjugated fluorescent salicylaldehyde-based Schiff bases: (a) step-like $R-C_n$ and tripod-like TSBs in our previous works; (b) chiral V-type **Cys** and propeller-type **diPhs** in this work.

probes for detecting metal ions.¹⁷ Moreover, non-conjugated soft materials might be used as anion probes, because they are flexible and might have multiple and strong intermolecular interactions with anions through hydrogen bonds.¹⁵ Particularly, the effects of different chiral bridges on the molecule arrangement, AIE, and chiral recognition properties are still unexplored. Herein, we demonstrate a novel and simple class of colorful chiral salen ligands (41 samples, Fig. 1b) that are linked by a non-conjugated cyclohexane (**Cy**) or 1,2-diphenylethane (**diPh**) bridge but have strong AIE and potential applications in cell imaging and recognition of anion and chiral unprotected amino acids.

Results and discussion

Synthesis and characterization

All salen ligands were reasonably easy to synthesize by the condensation of primary diamine with 2 equivalents of salicylaldehyde precursor in ethanol under refluxing condition.¹⁴ Most salen ligands have a bad solubility in petroleum ether, hexane, and water but a good solubility in CH_3CN , CH_2Cl_2 , DMF, and DMSO. All salen ligands either in solution or in solid state are stable within several months under air. For most salen ligands, good-quality single crystals can be obtained by the method of slow solvent diffusion/evaporation (CH_2Cl_2 /hexane). For the purpose of comparison, the reference substance **C₂** (ref. 14g) (Fig. 1a) was prepared as well.

Photophysical and AIE properties

The UV/visible absorption and fluorescence data of all synthesized **Cys** and **diPhs** at room-temperature are listed Tables S1, S2, Fig. S1, and S2 (in ESI†). Since the photophysical and AIE properties of (*S,S*), (*R,R*), and racemic salen ligands are similar, only racemic salen ligands are discussed in this section. As we expected, all salen ligands exhibit weak fluorescence in pure organic solvent of MeCN or DMSO (Tables S1–S3†). For example, racemic **3,5-Cl-Cy** shows very weak blue fluorescence ($\Phi = 0.010$) in the dilute MeCN, because the dynamic intramolecular rotations (IRs) of central cyclohexane bridge in **3,5-Cl-Cy** molecules provide a possible way to non-radiatively annihilate its excited states and result in the absence of fluorescence consequently. However, if water is added into MeCN, the green fluorescence of **3,5-Cl-Cy** is strengthened (Fig. 2), because **3,5-Cl-Cy** molecules cannot be dissolved in water, which causes **3,5-Cl-Cy** molecules to precipitate and aggregate. At the same time, adding water would red shift its absorption spectrum (Fig. S3†), which further confirms this aggregation. Its green fluorescence reaches the maximum ($\lambda_{em} = 511 \text{ nm}$, $\Phi = 0.064$) with a large Stokes shift of 117 nm, when the volume fraction (*f*) of water is increased to 90%. Furthermore, solid **3,5-Cl-Cy** displays strong green-yellow fluorescence ($\lambda_{em} = 532 \text{ nm}$, $\Phi = 0.32$) under 360 nm UV lamp (Fig. 2 and 3). All these findings reveal the fact of its AIE nature.

All the salen ligands have stronger fluorescence in solid state or water than in organic solvents (Tables S1 and S2†). Most solid samples, such as crystals, powders, and casting films (Fig. S4†), exhibit not only red-shifted but also much stronger fluorescence compared with water samples (Fig. 3, Tables S1, and S2†), and thus we will focus on their solid AIE characteristics. As shown in Fig. 4, Tables S1, and S2,† for solid salen ligands, different substituents have different effects on the emission peaks (λ_{em} : 474–565 nm) and colors (blue, green, and red). The presence of a π -extended system (**Naph-Cy**, $\lambda_{em} = 486 \text{ nm}$) to the simplest **Cy** ($\lambda_{em} = 502 \text{ nm}$) leads to a blue shift in fluorescence spectra. On the other hand, electron-donating $-OMe$ (**3-OMe-Cy**, $\lambda_{em} = 523 \text{ nm}$; **5-OMe-Cy**, $\lambda_{em} = 548 \text{ nm}$) or electron-accepting $-F$ (**3-F-Cy**, $\lambda_{em} = 509$), $-Cl$ (**3-Cl-Cy**, $\lambda_{em} = 520 \text{ nm}$; **3,5-Cl-Cy**, $\lambda_{em} = 532 \text{ nm}$), or $-NO_2$ (**3-NO₂-Cy**, $\lambda_{em} = 565 \text{ nm}$; **3,5-NO₂**, $\lambda_{em} = 550 \text{ nm}$) substituents result in red-shifted emission (Fig. 4

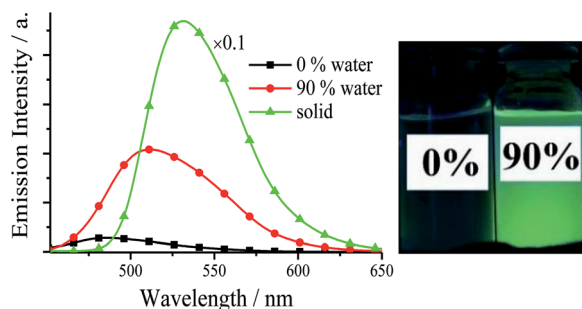


Fig. 2 Emission spectra and photographs (under 360 nm UV light) of **3,5-Cl-Cy** in MeCN–H₂O with different *f* values ($1.0 \times 10^{-5} \text{ mol dm}^{-3}$) and solid.



Fig. 3 Photographs (top: under room light, bottom: under 360 nm UV light) of Cys (a) and diPhs (b) 1 = diPh, 2 = 3-F-(*R,R*)-diPh, 3 = 3-F-(*S,S*)-diPh, 4 = 3-Cl-diPh, 5 = 3-Cl-(*R,R*)-diPh, 6 = 3-Cl-(*S,S*)-diPh, 7 = 3,5-Cl-diPh, 8 = 3,5-Cl-(*R,R*)-diPh, 9 = 3,5-Cl-(*S,S*)-diPh, 10 = 4-NEt₂-diPh; 11 = Naph-diPh, 12 = 3-NO₂-diPh, 13 = 5-NO₂-diPh, 14 = 3,5-NO₂-diPh) powders.

and Table S1†). A similar substituent effect was observed for solid diPhs (Fig. 4 and Table S2†), *R*-C_n^{14g} and TSBs¹⁵ (Fig. 1a) as well, indicating that a substituent has a very obvious effect on λ_{em} of these non-conjugated AIE-active materials.

All fluorescence quantum yields (Φ) (Tables S1 and S2†) of the aggregated salen ligands in water and solid state were

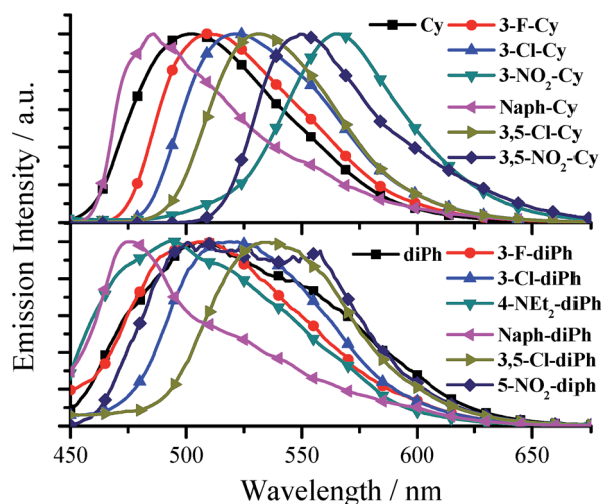


Fig. 4 Normalized emission spectra of some Cys and diPhs solids.

measured by the optical dilute method of Demas and Crosby¹⁸ with a standard of quinine sulfate ($\Phi_r = 0.55$, quinine in 0.05 mol dm⁻³ sulfuric acid) and an integrating sphere, respectively. Although the salen ligands with a non-conjugated linker have a small π -conjugated system, the fluorescence quantum yields of some salen ligands are unexpected high (up to 0.35 and 0.22 for Cys and diPhs, respectively, Tables S1 and S2†). In general, the introduction of -F (3-F-Cy, $\Phi = 0.060$; 3-F-diPh, $\Phi = 0.12$), -Cl (3-Cl-Cy, $\Phi = 0.14$; 3,5-Cl-Cy, $\Phi = 0.32$; 3-Cl-diPh, $\Phi = 0.16$; 3,5-Cl-diPh, $\Phi = 0.20$) substituents to Cy ($\Phi = 0.018$) or diPh ($\Phi = 0.10$) would improve its fluorescence quantum yield, but the presence of other substituents would bring positive or negative effects on fluorescence quantum yield. Combining with the previous results,^{14g,15} we can draw a conclusion that chlorination or fluorination is an much more efficient way to improve the fluorescence quantum yields of non-conjugated materials than those of π -conjugated materials.

Mechanism of AIE

The molecular structures and arrangements play a key role in AIE. For most AIE-active materials, they are π -conjugated molecules and stack closely with not only a short interplanar distance (d , for plane molecules^{14f}) or intermolecular aromatic H_{Ar}... π hydrogen bonds (for non-plane molecules, such as silole¹⁹ or TPE²⁰) but also weak intermolecular face-to-face π - π interactions. The former can ensure to eliminate the molecular rotation, the later would prevent the formation of excimer. For non-conjugated AIE-active *R*-C_n (ref. 14g) and TSBs,¹⁵ however, the restriction of C-C single bond rotations in the central alkyl chain bridges through some other non-covalent intermolecular interactions, such as N...H, O...H, C...H, H...H, F...H, Cl...H, and Cl...Cl must be considered as well.

In order to investigate the effect of molecule arrangements, the X-ray single crystal structures of Cy,²¹ 3-F-Cy (CCDC 1551151), 3-F-(*S,S*)-Cy (CCDC 1496280), 3-F-(*R,R*)-Cy (CCDC 1496279), 3,5-Cl-Cy,²² 3-NO₂-(*R,R*)-Cy,²³ 3-F-diPh (CCDC 1551150), and 3-Cl-diPh (CCDC 1551147) are depicted in Fig. 5–7 and

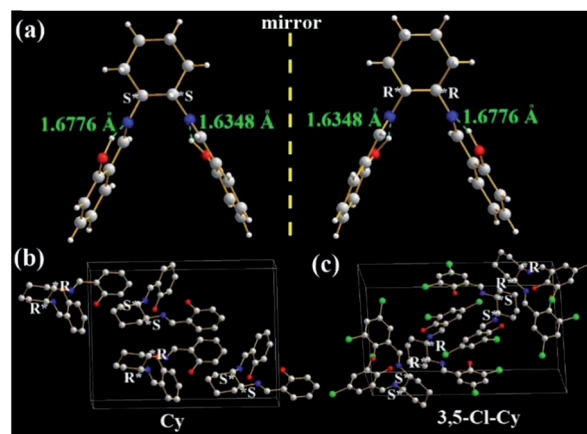


Fig. 5 X-ray single crystal structures and packing of Cy and 3,5-Cl-Cy molecules: (a) (*S,S*) and (*R,R*) enantiomers of Cy; (b) and (c) packing in a unit cell (H atoms are omitted).



Fig. 6 X-ray single crystal structures and packing of 3-F-Cy, 3-F-(*S,S*)Cy, and 3-F-(*R,R*)Cy molecules: (a) (*S,S*) and (*R,R*) enantiomers; (b) and (c) packing in a unit cell (H atoms are omitted); (d) intermolecular interactions of the two closest molecules (H atoms are omitted).

S5–S18.[†] Unlike step-like *R-C_n* (ref. 14g) and tripod-like TSBs¹⁵ molecules, these **Cys** and **diPhs** molecules are V- and propeller-type, respectively.

As expected, two intramolecular N $\cdots$ H hydrogen bonds (1.6776 and 1.6348 Å) between H (–OH) and N (C=N) atoms are found in **Cy** (Fig. 5a), which would eliminate free rotations of phenol rings. The dihedral angles between two panels of π -conjugated phenol rings are about 56.6°, resulting in its V-type molecular configuration and no intramolecular face-to-face π – π interactions in one **Cy** molecule. In dilute organic solution, it is obvious that the rotatable C–N and C–C single bonds in the central cyclohexane bridge afford a possible way to non-

radiatively annihilate its excited states and quench the fluorescence consequently. In the solid state, however, **Cy** molecules exhibit a head (cyclohexane)-to-tail (phenol ring) orientation (Fig. 5b). No intermolecular face-to-face π – π interactions (Fig. 5a, b, and S5[†]) are found between any neighboring **Cy** molecules. On the contrary, many other strong intermolecular interactions including H $\cdots$ H (2.4578, 2.5302, 2.6280, 2.7014, 2.7544, 2.7770, 2.7896, 2.8042, 2.8886, 2.9240, and 2.9792 Å), C $\cdots$ H (2.8077 and 2.8296 Å), and O $\cdots$ H (2.8111 Å) interactions are found in the two adjacent **Cy** molecules (Fig. S5[†]). These above intramolecular and intermolecular interactions would help to restrain the IRs and lead to the presence of AIE consequently. Owing to the existence of chiral carbon atoms, two (*S,S*) and (*R,R*) enantiomers (1 : 1) are found in the single crystals of racemic **Cy** (Fig. 5a and b). The *R/S*-chirality-induced interactions between two enantiomers (Fig. S5[†]) would help **Cy** molecules arrange in a tight head-to-tail orientation packing.

Similar two (*S,S*) and (*R,R*) enantiomers (1 : 1) are also found in the single crystals of racemic 3,5-Cl-**Cy** (Fig. 5c, S6, and S7[†]), but 3,5-Cl-**Cy** molecules exhibit a different tail-to-tail orientation from **Cy** molecules (Fig. 5c). The *R/S*-chirality of two enantiomers induce their phenol rings to pack together with a *d* of 3.46 Å (Fig. S6a[†]) and little intermolecular face-to-face π – π interactions (overlaps in two benzene rings). At the same time, many other strong intermolecular interactions including H $\cdots$ H (2.6687, 2.6899, 2.8377, 2.8587, and 2.8978 Å), O $\cdots$ H (2.4244, 2.6160, 2.6374, 2.6849 Å), and Cl $\cdots$ H (2.8689 and 2.8843 Å) interactions are found in the two adjacent 3,5-Cl-**Cy** molecules (Fig. S6c, d, and S7[†]). Moreover, strong Cl $\cdots$ Cl interactions (3.2848 Å, Fig. S7c[†]) are observed, because two benzene rings are packed together. The F and Cl atoms would help to induce intermolecular F $\cdots$ H, F $\cdots$ F, Cl $\cdots$ O, Cl $\cdots$ H, or Cl $\cdots$ Cl interactions (halogen bond²⁴) that is one possible reason why chlorination or fluorination can improve fluorescence quantum yield (see later discussion). All these data are consistent with the fact that solid 3,5-Cl-**Cy** and its enantiomers have the highest Φ values (0.32–0.35).

In order to evaluate the effects of different chiral (*R,R*) and (*S,S*) bridges on the molecule arrangement, molecular packing of 3-F-**Cy**, 3-F-(*S,S*)**Cy**, and 3-F-(*R,R*)**Cy** single crystal structures are shown in Fig. 6 and S8–S15.[†] Like 3,5-Cl-**Cy** molecules, racemic 3-F-**Cy** molecules exhibit a tight *R/S*-chirality-induced tail-to-tail orientation packing with little intermolecular face-to-face π – π interactions (*d* = 3.54 Å, Fig. 6a, b, S8, and S9[†]). Many other strong intermolecular interactions including H $\cdots$ H (2.6159, 2.6679, 2.6804, 2.7083, 2.8033, 2.8395, 2.8640, 2.8837, and 2.9128 Å), C $\cdots$ H (2.7592, 2.8238, and 2.9546 Å), O $\cdots$ H (2.4532, 2.4701, 2.6238, 2.6691, 2.7395, and 2.8609 Å), and F $\cdots$ H (2.7008, 2.7167, 2.7771, 2.8422, and 2.8584 Å) interactions are found in the two adjacent 3-F-**Cy** molecules. The photophysical and AIE properties of 3-F-**Cy**, 3-F-(*S,S*)**Cy**, and 3-F-(*R,R*)**Cy** are similar, but 3-F-**Cy** has a totally different molecule arrangement from 3-F-(*S,S*)**Cy** and 3-F-(*R,R*)**Cy** (Fig. 6b and c). The molecule arrangements of 3-F-(*S,S*)**Cy** and 3-F-(*R,R*)**Cy** single crystals overlap to form a triplex tail-to-tail, head-to-tail, and head-to-tail arrangement. No intermolecular π – π

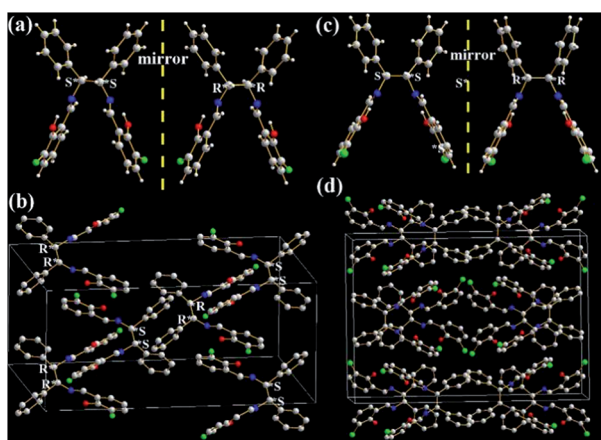


Fig. 7 X-ray single crystal structures and packing of 3-F-diPh (a, b) and 3-Cl-diPh (c, d) molecules: (a) and (c) (*S,S*) and (*R,R*) enantiomers; (b) and (d) packing in a unit cell (H atoms are omitted).



interactions but many similar strong intermolecular interactions ($\text{H}\cdots\text{H}$: 2.4496–2.9179 Å, $\text{C}\cdots\text{H}$: 2.8408–2.9800 Å, $\text{O}\cdots\text{H}$: 2.3803–2.8672 Å, $\text{H}_{\text{Ar}}\cdots\pi$: 2.54–2.57 Å, and $\text{F}\cdots\text{H}$: 2.4464–2.9903 Å) are found in the two adjacent 3-F-(*S,S*)Cy or 3-F-(*R,R*)Cy molecules (Fig. S10–S15†). However, there is no intermolecular $\text{F}\cdots\text{F}$ interactions in 3-F-Cy, 3-F-(*S,S*)Cy, and 3-F-(*R,R*)Cy, which would be contributed to the fact that their Φ values (0.059–0.062) are lower than that of 3,5-Cl-Cy (0.32).

Our previous work demonstrated that the introduction of strong electron-accepting $-\text{NO}_2$ substituents would be an efficient way to red shift fluorescence bands of non-conjugated AIE-active materials.^{14g,15} In this work, 3- NO_2 -Cy also shows a red-shifted fluorescence band up to 565 nm (Fig. 4). We failed to grow good-quality single crystals for X-ray analysis in the previous and present work. It is fortunate that the X-ray single crystal structure of 3- NO_2 -(*R,R*)Cy is obtained in the literature.²³ Unlike the above results, there are strong intermolecular face-to-face π - π interactions in 3- NO_2 -(*R,R*)Cy ($d = 3.31$ Å, Fig. S16†), according with the fact that of 3- NO_2 -Cy has a red-shifted fluorescence band with a not-so-high Φ value (0.051).

As shown in Fig. 7, two phenol rings and two benzene rings in 3-F-diPh and 3-Cl-diPh molecules are *cis* form rather than *trans* form, due to the existence of two chiral (*R,R*) or (*S,S*) carbon atoms. Two phenol rings and two benzene rings are separated each other to form their propeller structures, just like another well-known AIE-active material of TPE.^{9,13,20} Therefore, 3-F-diPh (2.4888–2.8510 Å) and 3-Cl-diPh (2.3507–2.8279 Å) molecules can tightly packed without any face-to-face π - π interactions in their single crystals (Fig. S17 and S18†). It should be noted that strong intermolecular $\text{F}\cdots\text{F}$ (3.2922 Å) and $\text{F}\cdots\text{O}$ (3.1760 Å) interactions are found in 3-F-diPh and 3-Cl-diPh molecules, respectively. All these factors enable solid 3-F-diPh and 3-Cl-diPh have a high Φ value of 0.12 and 0.16, respectively.

Having small π -conjugated systems, these pure organic salen ligands emit strong solid-state emission along with large Stokes shifts (up to 186 nm). This would enable us to doubt that their emission originates from phosphorescence²⁵ rather than fluorescence. Time-resolved emission decay spectra (Fig. S19†) reveal that the emission decay lifetime of 3-F-Cy, 3-F-(*S,S*)Cy, 3-F-(*R,R*)Cy, 3-F-diPh, 3-F-(*S,S*)diPh, and 3-F-(*R,R*)diPh powders is 0.70, 0.66, 0.66, 1.32, 1.72, and 0.93 ns, respectively. These data are in the range of ns, indicating that their emission is not phosphorescence but fluorescence. The large Stokes shifts might be contributed to the intramolecular hydrogen bonds which would lead to keto and enol tautomers through an ultrafast excited state intramolecular-proton transfer (ESIPT) process.²⁶ With the same *R* substituent, *R*-Cy and *R*-diPh have the similar fluorescence properties, because the low-energy transition is mainly be assigned to the $\pi \rightarrow \pi^*$ transition involving molecular orbitals essentially localized on the iminomethylphenol units with little contribution from the non-conjugated cyclohexane and 1,2-diphenylethane bridges (see the later discussion). Moreover, our previous work revealed that both the multi-Schiff base structure and $-\text{OH}$ groups are of the greatest importance for AIE.^{14g}

Anion probe properties

As shown in our previous work,¹⁵ TSBs (Fig. 1a) are tripod-like side-single-opening cages with three flexible arms of phenol ring and can be used as anion hosts through hydrogen bond. In this work, the two phenol rings of these salen ligands are *cis* form with a small dihedral angles, and thus they acting as clamps might react with anion through hydrogen bonds and detect anion consequently.

In general, (*S,S*), (*R,R*), and racemic salen ligands have similar anion responses, and thus racemic salen ligands are discussed in this section. The anion probe properties of Cy, 3-F-Cy, 3-Cl-Cy, 3,5-Cl-Cy, diPh, 3-F-diPh, 3-Cl-diPh, and 3,5-Cl-diPh are demonstrated in Table S3, Fig. 8, and S20–S28.† As example, upon adding many anions, including OH^- , F^- , Cl^- , Br^- , I^- , NO_3^- , CO_3^{2-} , HCO_3^- , SO_4^{2-} , S^{2-} , SO_3^{2-} , $\text{P}_2\text{O}_7^{4-}$, PO_4^{3-} , and HPO_4^{2-} , to dilute 3,5-Cl-diPh DMSO solution would turn on the blue fluorescence (up to 41.1-fold enhancement) of 3,5-Cl-diPh

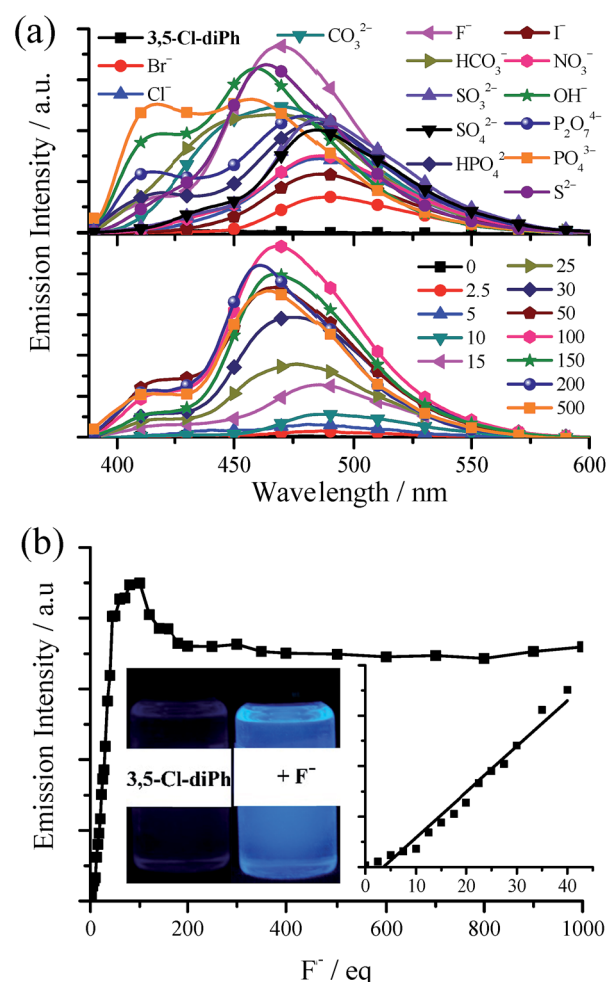


Fig. 8 (a) Top: emission spectra of 3,5-Cl-diPh upon the addition of 100 equivalent of different anions. Bottom: emission spectra of 3,5-Cl-diPh upon the addition of different equivalent of F^- . (b) Plot of emission intensity of 3,5-Cl-diPh at 468 nm as a function of F^- concentration. Inset: photographs of 3,5-Cl-diPh, 3,5-Cl-diPh + 100 equivalent of F^- under 360 nm UV light. In all cases, the probe is 1.0×10^{-5} mol dm^{-3} in DMSO and is excited at 380 nm.



(Fig. 8a). Alkaline OH^- (Fig. S20 and S21†) and neutral F^- (Fig. 8) were used to investigate the concentration effect. The fluorescence of **3,5-Cl-diPh** exhibits a linear enhancement (correlation coefficient $R^2 = 0.970$, $n = 15$) upon the addition of 0–40 equivalent of F^- and then reaches maximum upon the addition of 100 equivalent of F^- ($\Phi = 0.24$). If adding F^- further, the fluorescence would reduce and finally reach saturation ($\Phi = 0.21$). A similar fluorescence enhancement is observed upon adding OH^- to dilute **3,5-Cl-diPh** solution (Φ up to 0.29). The probe performance would be much worse if other solvents, such as MeCN, EtOH, and MeOH, are used.¹⁵

Cy, **3-F-Cy**, **3-Cl-Cy**, **3,5-Cl-Cy**, **diPh**, **3-F-diPh**, and **3-Cl-diPh** have similar anion effects with **3,5-Cl-diPh** that they are very sensitive to F^- , OH^- , S^{2-} and PO_4^{3-} (Table S3†). On the contrary, step-like C_4 (ref. 15) and C_2 have much less anion effect on fluorescence (Fig. S29†). Our previous work¹⁵ revealed that the reaction between TSBs and X^- is not a normal chemical reaction but intermolecular hydrogen bonding ($-\text{O}-\text{H}\cdots\text{X}^-$). The two phenol rings of **Cy**, **3-F-Cy**, **3,5-Cl-Cy**, **3-F-diPh**, and **3-Cl-diPh** are *cis* and diagonal form with a small dihedral angle of 56.6, 51.5, 53.8, 54.6, and 47.5°, respectively. The intramolecular distance $\text{O}\cdots\text{O}$ of **Cy**, **3-F-Cy**, **3,5-Cl-Cy**, **3-F-diPh**, and **3-Cl-diPh** is 6.081, 6.494, 6.352, 5.342, and 5.602 Å, respectively. However, C_4 and C_2 are step-like and their two phenol rings are parallelly separated with much longer intramolecular distance $\text{O}\cdots\text{O}$ of 8.314 and 10.52 Å, respectively.^{14g} It means that, like TSBs,¹⁵ **Cy**, **3-F-Cy**, **3,5-Cl-Cy**, **3-F-diPh**, and **3-Cl-diPh** are easier to act as tetradentate chelating agents (see the later discussion) and consequently react with an anion more tightly than C_4 and C_2 (Fig. S30†).

Chirality and chiral recognition

The chirality properties of **3-F-Cy**, **3-F-(S,S)Cy**, **3-F-(R,R)Cy**, **3-F-diPh**, **3-F-(S,S)diPh**, and **3-F-(R,R)diPh** are examined by circular dichroism (CD) spectra. As depicted in Fig. 9, **3-F-(S,S)diPh** shows a positive cotton effect at 326 nm and further strong peaks at 270, and 249 nm. The CD spectrum of **3-F-(R,R)diPh**

exactly mirrors that of **3-F-(S,S)diPh**, indicating that the two compounds are a pair of enantiomers. Moreover, racemic **3-F-diPh** has little CD signals. Similar phenomena are observed for **3-F-Cy**, **3-F-(S,S)Cy**, and **3-F-(R,R)Cy** as well. Moreover, **3-F-(S,S)Cy** and **3-F-(S,S)diPh** (or **3-F-(R,R)Cy** and **3-F-(R,R)diPh**) exhibit similar CD spectra, revealing that they have the same chiral configuration and energy transition.

The fluorescence responses towards various L- or D-amino acids were investigated under the similar experimental conditions with anion detection (Tables S4–S10, Fig. 10, and S31–S63†). Considering the problem of solubility in both water and DMSO, we chose ten amino acids including alanine (Ala), valine (Val), leucine (Leu), serine (Ser), proline (Pro), histidine (His), tryptophan (Trp), arginine (Arg), glutamate (Glu), and glutamine (Gln). Upon adding amino acids, the fluorescence enhancements of **Cy** and **diPh** (Table S4 and Fig. S31†) are lower than those of **3-F-Cy** (Table S5 and Fig. S32–S37†), **3-Cl-Cy** (Table S6 and Fig. S38–S43†), **3,5-Cl-Cy** (Table S7 and Fig. S44–S49†), **3-F-diPh** (Table S8 and Fig. S50–S55†), **3-Cl-diPh** (Table S9 and Fig. S56–S61†), and **3,5-Cl-diPh** (Table S10, Fig. 10, S62, and S63†). Since all nature amino acids are L-type ones, firstly we used **3,5-Cl-(S,S)diPh** and **3,5-Cl-(R,R)diPh** to probe different L-amino acids. As shown in Fig. 10a and Table S10,† like adding anion, adding different L-amino acids to **3,5-Cl-(S,S)diPh** or **3,5-Cl-(R,R)diPh** DMSO solutions would turn on their fluorescence (Φ up to 0.26), indicating that they might be used as universal amino acid probes (Fig. 10c, S62, and S63†). On the whole, **3,5-Cl-(S,S)diPh** and **3,5-Cl-(R,R)diPh** have a similar response that L-Arg, L-Pro, and L-Ala would induce stronger fluorescence enhancements than other amino acids. Moreover, chiral **3,5-Cl-(S,S)diPh** and **3,5-Cl-(R,R)diPh** could also discriminate the enantiomers of some amino acids (Fig. 10b and Table S10†). For example, adding D- or L-Gln to **3,5-Cl-(S,S)diPh** solution, the fluorescence of **3,5-Cl-(S,S)diPh** would turn on with a 36.2- and 5.82-fold enhancement, respectively. The selective recognition of chiral molecular enantiomers is related to the enantiomeric fluorescence difference ratio, ef, according to $\text{ef} = (I_D - I_0)/(I_L - I_0)$, in which I_0 represents the fluorescence emission intensity of receptor in the absence of a chiral substrate and I_D and I_L are the fluorescence intensities in the presence of D- and L-substrates, respectively. **3,5-Cl-(S,S)diPh** shows a good ef value of 6.22 for enantioselective recognition of D- and L-Gln. For other amino acids, His (ef = 2.71), Arg (ef = 2.27), and Trp (ef = 2.13) enantiomers could be discriminated by the **3,5-Cl-(S,S)diPh**. **3,5-Cl-(R,R)diPh** can identify Gln (ef = 2.93) and His (ef = 2.46) enantiomers.

In addition, **3-F-(S,S)diPh** has an excellent ef value of 0.11 for enantioselective recognition of D- and L-Val (Table S8 and Fig. S54†). **3-F-(R,R)diPh** can discriminate D- and L-Arg (ef = 4.89) (Table S9†). **3-Cl-(S,S)diPh** can efficiently discriminate Arg (ef = 5.48) and Gln (ef = 0.21) enantiomers, on the other hand, Arg (ef = 4.43) and Trp (ef = 0.18) enantiomers can be distinguished by **3-Cl-(R,R)diPh** (Table S9†). **3-F-Cy** (Table S5†), **3-Cl-Cy** (Table S6†), and **3,5-Cl-Cy** (Table S7†) generally exhibit worse ef values than **3-F-diPh** (Table S8†), **3-Cl-diPh** (Table S9†), and **3,5-Cl-diPh** (Table S10†). However, since it is too difficult to grow good-quality single crystals from the mixture of the dye

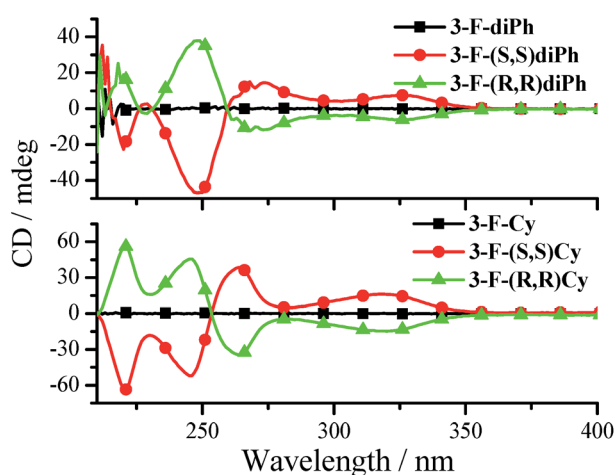


Fig. 9 CD spectra of **3-F-Cy**, **3-F-(R,R)Cy**, **3-F-(S,S)Cy**, **3-F-diPh**, **3-F-(R,R)diPh**, and **3-F-(S,S)diPh** in MeCN (3.0×10^{-5} mol dm^{-3}).





Fig. 10 (a) Emission spectra of 3,5-Cl-(S,S)diPh and 3,5-Cl-(R,R)diPh upon the addition of 100 equivalent of different L-amino acids. (b) Emission spectra of 3,5-Cl-(S,S)diPh and 3,5-Cl-(R,R)diPh upon the addition of 100 equivalent of different L- and D-amino acids. (c) Plot of emission intensity of 3,5-Cl-(R,R)diPh at 440 nm as a function of L-Ala concentration. Inset: emission spectra of 3,5-Cl-(R,R)diPh upon the addition of different equivalent of L-Ala. In all cases, the probe is 1.0×10^{-5} mol dm $^{-3}$ in DMSO and is excited at 380 nm.

and amino acid in DMSO solution, we failed to get a clear insight into the chiral discrimination ability (see the later discussion). We just guess that diPhs have a 1,2-diphenylethane

bridge but not a circular bridge of cyclohexane, hence, compared with Cys, they would be more flexible and consequently might have a better ability to form a cavity^{4a} for the recognition of a tiny chiral structure difference between D- and L-amino acid.

Mechanism of anion and amino acid probes

In order to investigate the possible interaction mechanism between the dye and anion/amino acid, ^1H nuclear magnetic resonance (^1H NMR) analysis was done. As shown in Fig. 11a, the ^1H NMR signal of H^1 atoms in 3,5-Cl-diPh appears a small broad peak at downfield ($\delta = 14.5$ ppm) due to the existence of the intramolecular hydrogen bonds ($-\text{OH}\cdots\text{N}$, Fig. 5).^{15,27} If adding 100 equivalent of OH^- (Fig. 11b), F^- (Fig. 11c), or L-Arg (Fig. 11d) (in D_2O), this downfield peak would disappear and reform a new small sharp peak at upfield ($\delta = 10.1$ – 10.2 ppm), which might be assigned to the ArOH without intramolecular hydrogen bonds.^{27,28} This upfield shift indicates that the strong intramolecular hydrogen bonds are destroyed by adding anion/amino acid. At the same time, the reduction (about 1/2–2/3) of peak area in new peak reveals the formation of new intermolecular hydrogen bonds. These intermolecular hydrogen bonds would increase the molecular electron density through-bond effect and lead to upfield shifts of other H atoms consequently.²⁹ Moreover, the ^1H NMR signal of H^3 at chiral carbon atoms also shows an obvious upfield shift from $\delta = 5.2$ ppm into $\delta = 4.7$, (5.0 and 4.2), and 3.9 ppm upon adding OH^- , F^- , and L-Arg, respectively, which might indicate that the neighboring nitrogen atoms have strong intermolecular interactions with anion and amino acid through hydrogen and halogen bonds.

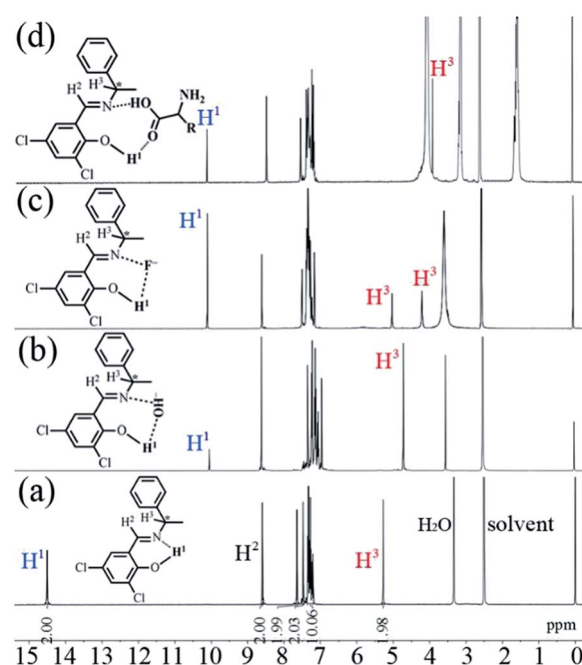


Fig. 11 ^1H NMR spectra of 3,5-Cl-diPh (a), 3,5-Cl-diPh + 100 equivalent of OH^- (b), 3,5-Cl-diPh + 100 equivalent of F^- (c), and 3,5-Cl-(R,R)diPh + 100 equivalent of L-Arg (d) in DMSO- d_6 .

Furthermore, the blank experiments (Fig. S64 and S65†) revealed that the ^1H NMR peak at 10.1–10.2 was not caused by an impurity.

The mechanisms of the aggregation- and anion/amino acid-induced fluorescence enhancement are totally different. For example, the green AIE of **3-F-Cy** is originated from the aggregated state (in water or solid) but its blue anion/amino acid-induced fluorescence is originated from the monomer ($1.0 \times 10^{-5} \text{ mol dm}^{-3}$ in DMSO). To gain insight into the nature of the excited states and transitions, gas-phase density functional theory (DFT) and time-dependent-DFT (TD-DFT) were also carried out for **3-F-Cy** with the Gaussian 09 program package (B3LYP 6-31G(d,p)). The energy absorption bands of **3-F-Cy** ($\lambda_{\text{abs}} = 316, 253, \text{ and } 211 \text{ nm}$ in MeCN) are reproduced well by the computation, which predict three absorption peaks at 318, 248, and 213 nm (Fig. 12). The lower energy absorption is mainly contributed to the highest occupied molecular orbital (HOMO) $\rightarrow$ lowest unoccupied molecular orbital (LUMO) (318 nm, oscillator strength, $f_{\text{osc}} = 0.0814$). The energy level and orbital isosurfaces diagrams of **3-F-Cy** (Fig. 12) reveal that its HOMO and LUMO are mainly made up of the π -functions of iminomethylphenol units rather than the cyclohexane bridge. Therefore, the lower energy absorption can mainly be assigned to the $\pi \rightarrow \pi^*$ transition involving molecular orbitals essentially localized on the iminomethylphenol units with little contribution from the non-conjugated cyclohexane bridge. As mentioned above, F^- , OH^- , or L-Arg would lead to changing $-\text{OH}$ into O^- , and thus **3-F-Cy** in form of O^- is used to evaluate the nature of the anion/amino acid-induced excited states and transitions by calculation. For optimized structure, **3-F-Cy** (in form of O^- , Fig. 12) has a much bigger dihedral angle (79.0°) between two phenol rings than **3-F-Cy** (in form of OH , 51.5°). The lower energy absorption band of **3-F-Cy** (in form of O^- , $\lambda_{\text{abs}} = 365$ and 350 nm for experiment and computation, respectively) are mainly composed of three transitions, including HOMO $\rightarrow$ LUMO (360 nm , $f_{\text{osc}} = 0.150$), HOMO-1 $\rightarrow$ LUMO+1 (344 nm , $f_{\text{osc}} = 0.175$), and HOMO $\rightarrow$ LUMO+1

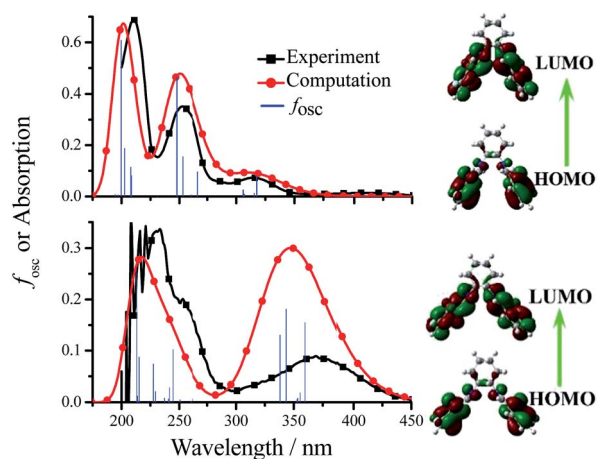


Fig. 12 Computational (gas-phase) and experimental absorption spectra and Frontier molecular orbitals of **3-F-Cy** (top, in form of OH) and **3-F-Cy** + 100 equivalent of OH^- (bottom, in form of O^-) MeCN solution.

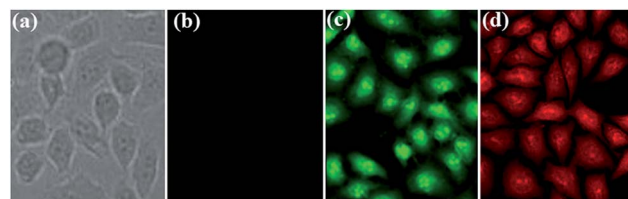


Fig. 13 Brightfield (a) incubated without dye and fluorescence images (b) incubated without dye; (c) incubated with **3,5-Cl-Cy**; (d) incubated with **3-NO₂-Cy** of HeLa cells.

(338 nm, $f_{\text{osc}} = 0.127$). The π -functions of HOMO, HOMO-1, LUMO and LUMO+1 of **3-F-Cy** in form of OH and O^- are similar, which reveals that anion/amino acid would not induce to change of the way of transition ($\pi \rightarrow \pi^*$ transition). This is much different from our previous work¹⁵ that anion would induce to change of the way of transition from $n \rightarrow \pi^*$ into $\pi \rightarrow \pi^*$ for TSBs. Moreover, anion/amino acid would red shift the lower energy absorption band with an increment of molar extinction co-efficient (Fig. 12 and S66†). This phenomenon can be reasonably explained by the fact that electron-rich ArO^- anions have resonance structures of benzoquinone.²⁸ These resonance structures lead the π -function Frontier molecular orbitals to a uniform distribution in the molecule skeleton, and consequently **3-F-Cy** (in form of O^-) has not only a smaller energy gap of transition but also an increment of molar extinction co-efficient and oscillator strength. This also might be contributed to its high fluorescence quantum yield in dilute DMSO solution. It is strange that dilute **3-F-Cy** (in form of O^-) DMSO solution having a red-shifted absorption spectrum shows a blue-shifted fluorescence spectrum, but AIE of **3-F-Cy** (in form of OH , in solid or water) is green-yellow. There is no anymore ESIPT for **3-F-Cy** (in form of O^-), which might be contributed to its blue fluorescence.

Living cell imaging

These non-conjugated AIE-active materials might have a lower cytotoxicity and better biocompatibility, and thus green-light-emitting **3,5-Cl-Cy** and red-light-emitting **3-NO₂-Cy** were used in cell imaging applications. The HeLa cells were imaged by the dye using a standard cell-staining protocol. As can be seen from Fig. 13, incubated with **3,5-Cl-Cy** or **3-NO₂-Cy**, the HeLa cells grow similar as in the control experiments in the absence of the dyes, indicating that both dyes have little toxicity on the living cells. HeLa cells show negligible background fluorescence. However, intense intracellular green and red AIE is observed after HeLa cells are incubated with **3,5-Cl-Cy** or **3-NO₂-Cy**, respectively. Therefore, these AIE-active materials may have potentially applications in probing or monitoring biologically important processes *in vitro* as well as *in vivo*.

Conclusion

In the present work, we demonstrate the unique AIE, chirality, anion/amino acid probe, and cell imaging properties a series of non-conjugated and cyclohexane/1,2-diphenylethane-linked



salen ligands. These V- or propeller-type salen ligands with small π -conjugated systems emit strong blue, green, and red AIE through little π - π interactions and strong non-covalent intermolecular interactions, such as O $\cdots$ H, H $\cdots$ H, C $\cdots$ H, F $\cdots$ H, F $\cdots$ F, Cl $\cdots$ H, Cl $\cdots$ O, and Cl $\cdots$ Cl. Moreover, combine with the advantages of non-conjugation and chirality, these soft materials have multiple and strong interactions with anions and amino acids by hydrogen and halogen bonds, and thus they might be used as universal anion probes and chiral receptors of unprotected amino acids with a good enantiomeric selectivity. Therefore, we believe that these simple salen ligands provide a new paradigm in the design of chiral and non-conjugated fluorescent materials for developing advanced organic optoelectronic devices, fluorescent bio-probes, and cell imaging, and so on. Further studies on circularly polarized luminescence³⁰ of these salen ligands are currently underway in our laboratory.

Conflicts of interest

There are no conflicts to declare.

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Notes and references

- (a) H. C. Aspinall, *Chem. Rev.*, 2002, **102**, 1807; (b) J. Crassous, *Chem. Soc. Rev.*, 2009, **38**, 830; (c) M. Liu, L. Zhang and T. Wang, *Chem. Rev.*, 2015, **115**, 7304.
- (a) L. Pu, *Chem. Rev.*, 2004, **104**, 1687; (b) G. A. Hembury, V. V. Borovkov and Y. Inoue, *Chem. Rev.*, 2008, **108**, 1; (c) A. Zehnacker and M. A. Suhm, *Angew. Chem., Int. Ed.*, 2008, **47**, 6970; (d) A. Accetta, R. Corradini and R. Marchelli, *Top. Curr. Chem.*, 2011, **300**, 175; (e) L. Pu, *Acc. Chem. Res.*, 2012, **45**, 150; (f) D. Leung, S. O. Kang and E. V. Anslyn, *Chem. Soc. Rev.*, 2012, **41**, 448; (g) X. Zhang, J. Yin and J. Yoon, *Chem. Rev.*, 2014, **114**, 4918; (h) L. You, D. Zha and E. V. Anslyn, *Chem. Rev.*, 2015, **115**, 7840.
- (a) X. Yang, X. Liu, K. Shen, C. Zhu and Y. Cheng, *Org. Lett.*, 2011, **13**, 3510; (b) F. Li, L. Li, W. Yang, L. S. Zheng, Z. J. Zheng, K. Jiang, Y. Lu and L. W. Xu, *Tetrahedron Lett.*, 2013, **54**, 1584; (c) Z. Huang, S. Yu, K. Wen, X. Yu and L. Pu, *Chem. Sci.*, 2014, **5**, 3457.
- (a) H. T. Feng, X. Zhang and Y. S. Zheng, *J. Org. Chem.*, 2015, **80**, 8096; (b) J. B. Xiong, W. Z. Xie, J. P. Sun, J. H. Wang, Z. H. Zhu, H. T. Feng, D. Guo, H. Zhang and Y. S. Zheng, *J. Org. Chem.*, 2016, **81**, 3720.
- (a) H. L. Liu, H. P. Zhu, X. L. Hou and L. Pu, *Org. Lett.*, 2010, **12**, 4172; (b) H. Jintoku, M. Takafuji, R. Odac and H. Ihara, *Chem. Commun.*, 2012, **48**, 4881; (c) T. Liu, Y. Su, H. Song and Y. Lv, *Analyst*, 2013, **138**, 6558; (d) B. Aswathy and G. Sony, *J. Lumin.*, 2014, **154**, 541.
- (a) U. Mitschke and P. Bauerle, *J. Mater. Chem.*, 2000, **10**, 1471; (b) L. S. Hung and C. H. Chen, *Mater. Sci. Eng., A*, 2002, **39**, 143; (c) J. D. Slinker, J. Rivnay, J. S. Moskowicz, J. B. Parker, S. Bernhard, H. D. Abruoac and G. Malliaras, *J. Mater. Chem.*, 2007, **17**, 2976; (d) Q. Zhao, C. H. Huang and F. Y. Li, *Chem. Soc. Rev.*, 2011, **40**, 2508; (e) J. F. Zhang, Y. Zhou, J. Yoon and J. S. Kim, *Chem. Soc. Rev.*, 2011, **40**, 3416; (f) Y. Feng, J. H. Cheng, L. Zhou, X. G. Zhou and H. F. Xiang, *Analyst*, 2012, **137**, 4885; (g) H. F. Xiang, J. H. Cheng, X. F. Ma, X. G. Zhou and J. J. Chruma, *Chem. Soc. Rev.*, 2013, **42**, 6128; (h) Y. M. Yang, Q. Zhao, W. Feng and F. Y. Li, *Chem. Rev.*, 2013, **113**, 192; (i) J. H. Cheng, X. G. Zhou and H. F. Xiang, *Analyst*, 2015, **140**, 7082; (j) J. Liang, B. Z. Tang and B. Liu, *Chem. Soc. Rev.*, 2015, **44**, 2798.
- (a) J. B. Birks, *Photophysics of Aromatic Molecules*, Wiley, New York, 1970; (b) X. F. Ma, R. Sun, J. H. Cheng, J. Y. Liu, F. Gou, H. F. Xiang and X. G. Zhou, *J. Chem. Educ.*, 2016, **93**, 345.
- B. Z. Tang, New Structural Motifs for Solid Light Emitters, *Cutting-Edge Chemistry*, ACS, 2016, https://www.acs.org/content/acs/en/pressroom/cutting-edge-chemistry/new-structural-motifs-for-solid-light-emitters.html?cid=home_promo.
- Y. Hong, J. W. Y. Lam and B. Z. Tang, *Chem. Soc. Rev.*, 2011, **40**, 5361.
- C. M. Xing, J. W. Y. Lam, A. Qin, Y. Dong, M. Haussler, W. T. Yang and B. Z. Tang, *Polym. Mater. Sci. Eng.*, 2007, **96**, 418.
- A. Pucci, R. Rausa and F. Ciardelli, *Macromol. Chem. Phys.*, 2008, **209**, 900.
- M. S. Mathew, K. Sreenivasan and K. Joseph, *RSC Adv.*, 2015, **5**, 100176.
- (a) J. Luo, Z. Xie, J. W. Y. Lam, L. Cheng, H. Chen, C. Qiu, H. S. Kwok, X. Zhan, Y. Liu, D. Zhu and B. Z. Tang, *Chem. Commun.*, 2001, 1740; (b) J. Mei, N. L. C. Leung, R. T. K. Kwok, J. W. Y. Lam and B. Z. Tang, *Chem. Rev.*, 2015, **115**, 11718.
- (a) L. Zhou, P. Y. Cai, Y. Feng, J. H. Cheng, H. F. Xiang, J. Liu, D. Wu and X. G. Zhou, *Anal. Chim. Acta*, 2012, **735**, 96; (b) L. Zhou, Y. Feng, J. H. Cheng, N. Sun, X. G. Zhou and H. F. Xiang, *RSC Adv.*, 2012, **2**, 10529; (c) J. H. Cheng, K. Y. Wei, X. F. Ma, X. G. Zhou and H. F. Xiang, *J. Phys. Chem. C*, 2013, **117**, 16552; (d) J. H. Cheng, Y. H. Zhang, X. F. Ma, X. G. Zhou and H. F. Xiang, *Chem. Commun.*, 2013, **49**, 11791; (e) J. H. Cheng, X. F. Ma, Y. H. Zhang, J. Y. Liu, X. G. Zhou and H. F. Xiang, *Inorg. Chem.*, 2014, **53**, 3210; (f) X. F. Ma, J. H. Cheng, J. Y. Liu, X. G. Zhou and H. F. Xiang, *New J. Chem.*, 2015, **39**, 492; (g) J. H. Cheng, Y. X. Li, R. Sun, J. Y. Liu, F. Gou, X. G. Zhou, H. F. Xiang and J. Liu, *J. Mater. Chem. C*, 2015, **3**, 11099.
- X. H. Zhang, G. Y. Shen, F. Gou, J. H. Cheng, X. G. Zhou and H. F. Xiang, *Mater. Chem. Front.*, 2017, **1**, 1041.
- (a) L. Canali and D. C. Sherrington, *Chem. Soc. Rev.*, 1999, **28**, 85; (b) P. G. Cozzi, *Chem. Soc. Rev.*, 2004, **33**, 410; (c) C. Baleizao and H. Garcia, *Chem. Rev.*, 2006, **106**, 3987.
- P. S. Hariharan and S. P. Anthony, *Spectrochim. Acta, Part A*, 2015, **136**, 1658.



- 18 J. N. Demas and G. A. Crosby, *J. Phys. Chem.*, 1971, **75**, 991.
- 19 (a) Y. Liu, Y. Tang, N. N. Barashkov, I. S. Irgibaeva, J. W. Y. Lam, R. Hu, D. Birimzhanova, Y. Yu and B. Z. Tang, *J. Am. Chem. Soc.*, 2010, **132**, 13951; (b) Y. Yuan, R. T. K. Kwok, B. Z. Tang and B. Liu, *J. Am. Chem. Soc.*, 2014, **136**, 2546.
- 20 (a) J. Yang, L. Li, Y. Yu, Z. C. Ren, Q. Peng, S. H. Ye, Q. Q. Li and Z. Li, *Mater. Chem. Front.*, 2017, **1**, 91; (b) C. W. T. Leung, Y. Hong, S. Chen, E. Zhao, J. W. Y. Lam and B. Z. Tang, *J. Am. Chem. Soc.*, 2013, **135**, 62; (c) S. Chen, Y. Hong, Y. Liu, J. Liu, C. W. T. Leung, M. Li, R. T. K. Kwok, E. Zhao, J. W. Y. Lam, Y. Yu and B. Z. Tang, *J. Am. Chem. Soc.*, 2013, **135**, 4926; (d) L. Chen, G. W. Lin, H. R. Peng, S. Y. Ding, W. W. Luo, R. R. Hu, S. M. Chen, F. Huang, A. J. Qin, Z. J. Zhao and B. Z. Tang, *Mater. Chem. Front.*, 2017, **1**, 176; (e) J. Yang, J. Huang, Q. Q. Li and Z. Li, *J. Mater. Chem. C*, 2016, **4**, 2663.
- 21 J. C. Cannadine, J. P. Corden, W. Errington, P. Moore and M. G. H. Wallbridge, *Acta Crystallogr., Sect. C: Cryst. Struct. Commun.*, 1996, **52**, 1014.
- 22 E. Hadjoudis, A. Rontoyianni, K. Ambroziak, T. Dziembowska and I. M. Mavridis, *J. Photochem. Photobiol., A*, 2004, **162**, 521.
- 23 E. Rafii, M. Giorgi, N. Vanthuyne and C. Roussel, *ARKIVOC*, 2005, **10**, 86.
- 24 (a) P. Metrangolo, H. Neukirch, T. Pilati and G. Resnati, *Acc. Chem. Res.*, 2005, **38**, 386; (b) G. Cavallo, P. Metrangolo, T. Pilati, G. Resnati, M. Sansotera and G. Terraneo, *Chem. Soc. Rev.*, 2010, **39**, 3772.
- 25 S. Mukherjee and P. Thilagar, *Chem. Commun.*, 2015, **51**, 10988.
- 26 V. S. Padalkar and S. Seki, *Chem. Soc. Rev.*, 2016, **45**, 169.
- 27 J. H. Cheng, F. Gou, X. H. Zhang, G. Y. Shen, X. G. Zhou and H. F. Xiang, *Inorg. Chem.*, 2016, **55**, 9221.
- 28 J. Y. Liu, J. H. Cheng, X. F. Ma, X. G. Zhou and H. F. Xiang, *Res. Chem. Intermed.*, 2016, **42**, 5027.
- 29 (a) M. Boiocchi, L. Del Boca, D. E. Gomez, L. Fabbrizzi, M. Licchelli and E. Monzani, *J. Am. Chem. Soc.*, 2004, **126**, 16507; (b) Q. Li, Y. Guo, J. Xu and S. J. Shao, *Sens. Actuators, B*, 2011, **158**, 427.
- 30 (a) E. M. Sanchez-Carnerero, A. R. Agarrabeitia, F. Moreno, B. L. Maroto, G. Muller, M. J. Ortiz and S. de la Moya, *Chem.-Eur. J.*, 2015, **21**, 13488; (b) J. Kumar, T. Nakashima and T. Kawai, *J. Phys. Chem. Lett.*, 2015, **6**, 3445.

