

CrossMark  
click for updatesCite this: *Chem. Sci.*, 2016, 7, 5568

## Tandem reactions in self-sorted catalytic molecular hydrogels†

Nishant Singh,<sup>a</sup> Kai Zhang,<sup>b</sup> César A. Angulo-Pachón,<sup>a</sup> Eduardo Mendes,<sup>b</sup> Jan H. van Esch<sup>\*b</sup> and Beatriu Escuder<sup>\*a</sup>

By equipping mutually incompatible carboxylic acid and proline catalytic groups with different self-assembling motives we have achieved self-sorting of the resulting catalytic gelators, namely **SucVal8** and **ProValDoc**, into different supramolecular fibers, thus preventing the acidic and basic catalytic groups from interfering with each other. The resulting spatial separation of the incompatible catalytic functions is found to be essential to achieve one-pot deacetalization–aldol tandem reactions with up to 85% efficiency and 90% enantioselectivity. On the contrary, when **SucVal8** was co-assembled with a structurally similar catalytically active hydrogelator (**ProVal8**), self-sorting was precluded and no tandem catalysis was observed.

Received 21st March 2016  
Accepted 6th May 2016

DOI: 10.1039/c6sc01268j

www.rsc.org/chemicalscience

### Introduction

Tandem catalysis has received increasing attention in recent years due to the process simplicity, economy of time and energy and waste reduction, among other practical advantages, associated with one-pot procedures. In particular, orthogonal tandem catalysis is a one-pot reaction in which sequential catalytic processes occur through two or more functionally distinct and non-interfering catalytic cycles.<sup>1</sup> Excellent examples are found in multi-enzymatic complexes present in biosynthetic pathways, in which a substrate is sequentially transformed towards a product in an effective and selective manner.<sup>2</sup> Multifunctional catalysts have been prepared based on the spatial separation of catalytic groups in nanostructured materials such as star polymers, silicas, zeolites and MOFs, among others, quite often requiring tedious preparation procedures.<sup>3</sup> Some of these systems are based on the controlled organization of incompatible acidic and basic catalytic groups into non-interacting locations. In this context, self-assembly of discrete functional components into multifunctional self-organized materials by bottom-up non-covalent synthesis could represent a powerful strategy incorporating both functional diversity as well as synthetic economy. Catalytic supramolecular nanomaterials can be prepared by self-assembly of tailor-made low molecular weight functional building blocks.<sup>4</sup> In this respect, we and others have reported several examples of

catalytic molecular gels in which catalytic activity has emerged as a consequence of the supramolecular organization of functional groups.<sup>5</sup>

These examples were based on a single catalytic component. However, supramolecular tandem catalysis requires the spatial separation of two incompatible catalytic moieties. One way to achieve spatial separation by self-assembly is self-sorting, a relevant phenomenon in complex supramolecular systems that can be defined as a high fidelity recognition between molecules based on non-covalent interactions and structural complementarity.<sup>6</sup> Seminal work in this field was reported by Menger *et al.* showing self-sorting of fibers and microcrystallites based upon side chain differences of related compounds, as well as by van Esch *et al.* who reported orthogonal self-assembly of surfactants and hydrogelators into co-existing fibers and vesicles.<sup>7</sup> Since then, several groups have studied this phenomenon in depth in order to prepare spatially and temporally resolved multicomponent nanoscale materials.<sup>8,9</sup>

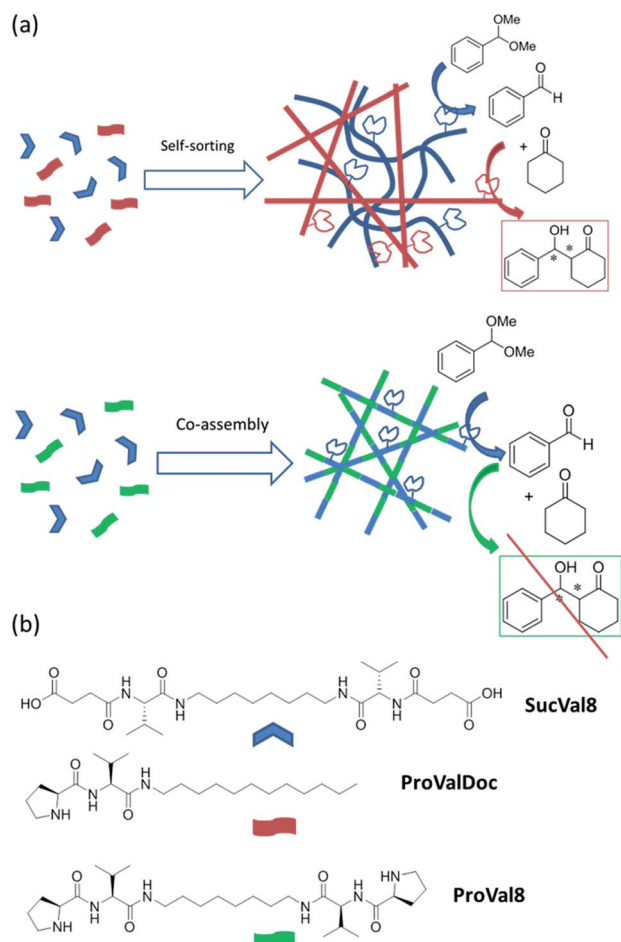
In this communication we present for the first time the use of two orthogonally self-assembled catalytic molecular hydrogel networks for performing one-pot tandem catalysis. In contrast, an analogous co-assembled network is shown to be catalytically inactive (Scheme 1a). For this purpose, we selected three low-molecular weight hydrogelators with different catalytic groups and different self-assembly motives, which were previously described in our group (Scheme 1b). Compounds **SucVal8** and **ProVal8** present a bolaamphiphilic structure of similar molecular dimensions bearing carboxyl groups and L-proline residues, respectively, as end-groups.<sup>10</sup> Compound **ProValDoc** is an amphiphilic dipeptide with an L-proline residue at the N-terminus.<sup>5b</sup>

<sup>a</sup>Departament de Química Inorgànica i Orgànica, Universitat Jaume I, E-12071 Castelló, Spain. E-mail: escuder@uji.es

<sup>b</sup>Department of Chemical Engineering, Delft University of Technology, Julianalaan 136, 2628 BL Delft, The Netherlands. E-mail: j.h.vanEsch@tudelft.nl

† Electronic supplementary information (ESI) available: Synthesis and characterization of compounds, technical details and additional figures are provided. See DOI: 10.1039/c6sc01268j





Scheme 1 (a) Reaction scheme and self-sorting vs. co-assembly of different self-assembling components based on the degree of interaction and recognition between them; (b) structures of hydrogelators SucVal8, ProValDoc and ProVal8.

## Results and discussion

Two different hydrogels were prepared by mixing **SucVal8** and the two L-proline derivatives (**ProValDoc** and **ProVal8**). We envisaged that the structural differences between **SucVal8** and **ProValDoc** would lead to their orthogonal self-assembly into two self-sorted networks, whereas the similar backbone structure of **SucVal8** and **ProVal8** would render a single co-assembled network. Then, these two mixed networks were studied for the one-pot deacetalization–direct aldol tandem reaction (Scheme 1a). All the individual gels and systems of two hydrogels were prepared by gentle heating to completely dissolve the compounds in water followed by sonication for 1 min, and left to stand overnight at 25 °C. For catalysis, the reactants were added to the gels and vortexed. Upon reaction, the products were extracted at the desired time by extracting twice with 1 mL  $\text{CDCl}_3$ , dried over  $\text{MgSO}_4$ , and observed by  $^1\text{H}$  NMR.

The hydrogels of **ProValDoc**, on the one hand, are white turbid looking gels with a minimum gel concentration (m.g.c) of 5.7 mM and temperature for the gel to solution transition,  $T_{\text{gel}} = 40$  °C (see ESI† for gel preparation methods;  $T_{\text{gel}}$  was

determined by inverted vial test). The catalytic activity of **ProValDoc** for the direct aldol condensation of aldehydes and ketones has already been reported by our group.<sup>5b,d</sup> On the other hand, **SucVal8** is a bolaamphiphilic hydrogelator with carboxylic acid groups at the two ends, and is able to form transparent self-standing hydrogels showing a m.g.c of 7 mM with  $T_{\text{gel}} = 70$  °C.<sup>10b</sup> The use of Brønsted acids for deprotection of acetal groups is well known.<sup>11</sup> First, we checked the catalytic activity of **SucVal8** fibers for the deacetalisation reaction. Complete conversion of 35 mM benzaldehyde dimethyl acetal into benzaldehyde was observed in 10 hours when 20 mol% (7 mM gel) of catalyst was used, while a lower  $K_{\text{obs}}$  was observed in samples of **SucVal8** in solution (Table S1 in ESI†). **ProValDoc** did not catalyse this reaction, either in solution or as a gel.

Subsequently, a mixture of both **SucVal8** + **ProValDoc** at their respective m.g.c values was used to obtain a strong turbid translucent gel. The AFM (Fig. 1) and TEM (Fig. S1–S5 in ESI†) images of this gel revealed the presence of two different kinds of fibers (Fig. 1c and S8c†). The larger, more crystalline fibers were attributed to **ProValDoc**, whereas the coexisting twisted thinner fibers were assigned to **SucVal8**, based on their similarity with the fiber morphologies observed for the single component gels (Fig. 1a and b). The height and width profiles provided in the ESI (Fig. S7 and S8†) show the presence of two different kinds of fibers with profiles similar to the individual gelators.

To further confirm the orthogonal self-assembly of the catalytic network components, we performed wide angle powder X-ray diffraction (WAXD) of the xerogels of the two different gelators at their m.g.c and of their mixture. The WAXD data for **SucVal8** reflected an amorphous structure with no crystallinity (Fig. 2a), whereas that for **ProValDoc** alone showed more crystallinity, which can be attributed to one of the



Fig. 1 AFM images of hydrogels of (A) **SucVal8**, (B) **ProValDoc**, (C) **ProValDoc** + **SucVal8** and (D) **ProVal8** + **SucVal8**.



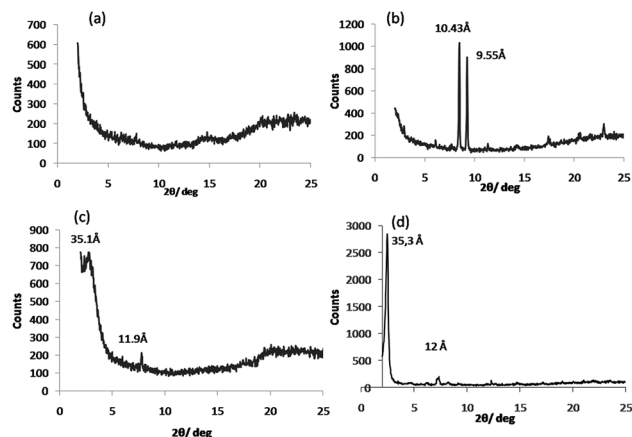


Fig. 2 WAXD patterns of the xerogels of (a) SucVal8, (b) ProValDoc and (c) SucVal8 + ProValDoc. (d) ProValDoc gel formed by pH change.

previously described polymorphs of this compound, the most thermostable one (Fig. 2b).<sup>12</sup> Interestingly, the WAXD of the mixture showed an overlap of the patterns of amorphous **SucVal8** and a different polymorph of **ProValDoc**, which is dominant when gels are formed by pH change from acidic solutions (Fig. 2c and d). Even though gels of this compound alone and in the mixture of two hydrogelators were both prepared by heating followed by sonication, we obtained different polymorphs. This result suggests, as already mentioned in our previous report, a seeding effect caused by a small amount of protonated **ProValDoc** crystals which could be formed in the presence of the acidic counterpart. Nevertheless, as we reported before, all the polymorphs are catalytically active with only slight differences in the respective reaction rates.<sup>12</sup> To further confirm the self-sorting of the two fibers, rheological experiments were done, which revealed the  $G'_{\max}$  for the mixed system (**SucVal8** + **ProValDoc**) to be 11 430 Pa, which is one order of magnitude higher than the individual gelators **SucVal8** ( $G'_{\max} = 1578$  Pa) and **ProValDoc** ( $G'_{\max} = 650$  Pa) (Fig. 3 and S10†).

This falls in line with the previous reports of orthogonal systems being mechanically stronger than the individual

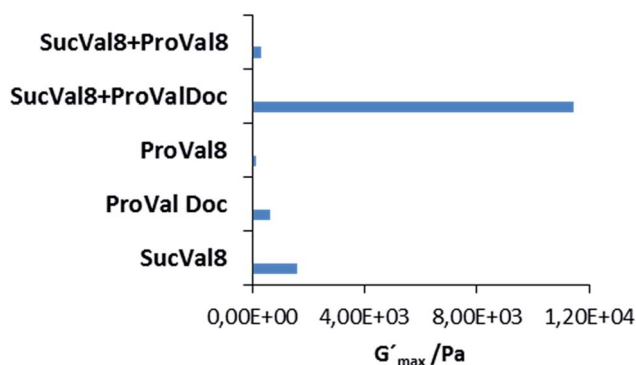


Fig. 3 Comparison of  $G'_{\max}$  of different gels. The plateau region of the graph was taken as the  $G'_{\max}$  of each system (Fig. S9†).

hydrogelators owing to the higher density of fibers.<sup>13</sup> Additionally, DSC experiments revealed the thermal imprints of both **SucVal8** and **ProValDoc** in the mixed system, suggesting the self-sorted assembly of both the hydrogelators (Fig. S11†).

From the  $T_{\text{gel}}$  experiments we know that **SucVal8** forms more thermally stable gels than **ProValDoc**. Therefore, we formed different gels by mixing the two gelators and varying the concentration of **ProValDoc** to see if this induced an effect on the overall thermal stability of the gelator (Fig. 4). The gels of the mixture proved to be slightly more ( $\sim 5$  °C higher) thermally stable than **SucVal8**, probably due to the higher density of fibers in the system owing to the presence of two different self-sorted fiber networks, making the gel stiffer and more stable.<sup>13,9b</sup> Thus, the self-sorting of molecules in a gel system consisting of two structurally different hydrogelators was suggested by the AFM, TEM, rheology,  $T_{\text{gel}}$ , WAXD and DSC experiments.

The orthogonally assembled catalytic fibers were then examined for catalytic activity for the two previously mentioned reactions in tandem and in one pot. The **SucVal8** + **ProValDoc** gel was first tested for deprotection of the acetal group. A quantitative yield of benzaldehyde from benzaldehyde dimethyl acetal (35 mM) was obtained in 10 h using 7 mM (20 mol%) of the catalyst. To examine the viability of the **SucVal8** + **ProValDoc** gel for the tandem reaction, the two substrates, benzaldehyde dimethyl acetal and cyclohexanone, were simultaneously introduced to the system at  $t = 0$ . In 72 hours more than 85% of the aldehyde obtained by deacetalisation was converted to the final aldol product with an *anti* : *syn* ratio of 84 : 16 (Fig. 5a, 90% ee).  $K_{\text{obs}} = (2.1 \pm 0.4) \times 10^{-5} \text{ s}^{-1}$  for the tandem reaction. No final product was observed when both the gelators were mixed when completely soluble (see ESI† for solubility procedure). These results confirm that both networks maintain their catalytic activity without interfering with each other. Moreover, when the direct aldol reaction between benzaldehyde and cyclohexanone was performed in the **SucVal8** + **ProValDoc** gel, a 91% yield was obtained in 72 hours (*anti* : *syn* ratio of 90 : 10; 91% ee).  $K_{\text{obs}} = (2.2 \pm 0.3) \times 10^{-5} \text{ s}^{-1}$ , suggesting that the acetal deprotection was not the rate limiting step.

Orthogonal assembly is mandatory for the successful isolation of catalytic groups with opposite acid–base characteristics for dual catalysis in one pot. To highlight this fact, we

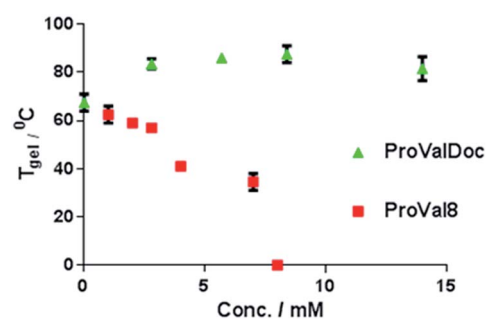


Fig. 4  $T_{\text{gel}}$  of the different gels of **SucVal8** + **ProValDoc** and **SucVal8** + **ProVal8** formed by varying the concentration of the proline component. **SucVal8** was present in the mixtures at its m.g.c.



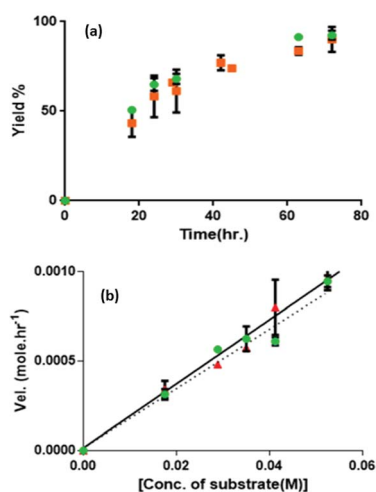


Fig. 5 (a) Kinetics of the tandem (●) and direct aldol reaction (▲) in the mixture of **SucVal8** + **ProValDoc** at their respective m.g.c values; (b) velocity of product formation versus substrate concentration for the tandem and direct aldol reaction.

performed similar experiments to those described above but this time using compound **ProVal8**, which is structurally very similar to **SucVal8**. **ProVal8** is a bolaamphiphilic hydrogelator like **SucVal8**, but with a proline group at each end (Scheme 1), that forms a white turbid weak gel at an m.g.c of 6 mM with  $T_{gel} = 37\text{ }^{\circ}\text{C}$ . As we have reported before for other bolaamphiphilic analogues,<sup>14</sup> we expected **ProVal8** to co-assemble with **SucVal8**. **ProVal8** forms large crystalline fibers with a width varying from 40–50 nm (see Fig. S3 and S6 in ESI†). However, the TEM images of the mixed gel **SucVal8** + **ProVal8** revealed a homogeneous system of only one kind of fiber with a width of 20–30 nm (Fig. S5†). On the other hand, AFM of the mixture (Fig. 1d) showed the presence of twisted thin fibrils (Fig. 1a). A closer analysis of the AFM images revealed that the fibrils in the mixture **SucVal8** + **ProVal8** had a pitch value of 70 nm, which is almost twice the value of the pure **SucVal8** (45 nm) (see Fig. S7 in ESI†). Therefore, fibers of the mixture are different from fibers constituting the individual gels, alluding towards the co-assembly of the two molecules to form a different kind of homogenous network.

The WAXD of the xerogel of this mixture also showed a different pattern of packing to the individual hydrogelators (Fig. 2a and 6a, b). Moreover, the gel of this mixture seemed to be weaker than **SucVal8** alone. This was further confirmed by  $T_{gel}$  experiments with a varying concentration of **ProVal8** in the mixture. With an increase in the concentration of **ProVal8**, a decrease in  $T_{gel}$  was observed. On increasing the concentration of **ProVal8** above 7 mM in the mixture, we could no longer obtain self-standing gels but only suspended aggregates (Fig. 4). The disruptive co-assembly of **ProVal8** and **SucVal8** was further vindicated by rheology and DSC experiments.  $G'_{max}$  for **ProVal8** + **SucVal8** was measured to be 290 Pa, which was lower than that of **SucVal8** (1578 Pa) and slightly higher than **ProVal8** (138 Pa) (Fig. 3 and S10†). The thermal imprint of the mixed system was also different from the individual

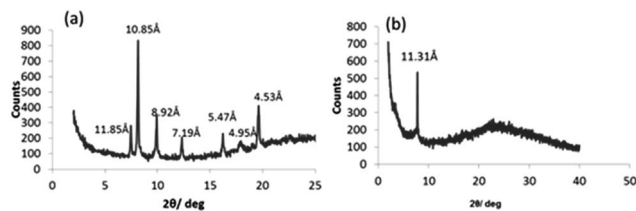


Fig. 6 WAXD patterns of the xerogels of (a) **ProVal8** and (b) **SucVal8** + **ProVal8**.

systems (Fig. S11†). All of the above experiments suggested that the molecules of **ProVal8** and **SucVal8** showed a disruptive co-assembly that interferes with the gelation ability of each hydrogelator.<sup>15</sup> Subsequently, catalytic studies were performed for this co-assembled mixture. Single component gels of **ProVal8** were able to catalyze the aldol condensation of benzaldehyde and cyclohexanone in water with a yield of 55% in 96 hours. However, when the tandem deacetalization/aldol reaction was tested with the mixture of **SucVal8** + **ProVal8**, only 58% conversion of benzaldehyde dimethyl acetal to benzaldehyde was observed in 96 hours, and no final aldol product was obtained even after much longer times. These results suggest a specific interaction between the acidic and proline groups of the two molecules while co-assembling, presumably involving a salt bridge, and thus rendering them catalytically inactive for the enamine-based aldol reaction, which is hampered if the proline amino group is protonated. FTIR did not show significant differences in the position of bands between the different samples, however, broad bands were obtained for the xerogel of the mixture **SucVal8** + **ProVal8**, suggesting the formation of ammonium salts to a greater extent (see Fig. S14†).

## Conclusions

In conclusion, we have reported for the first time two catalytic molecular hydrogelators that, owing to their structural differences, are able to orthogonally self-assemble into a two-network gel and perform two reactions in one pot which would be otherwise incompatible, as exemplified by a related co-assembled network. We have shown that it is possible, by the sole use of molecularly instructed non-covalent interactions, to construct spatially separated catalytic sites able to work in tandem, mimicking multi-enzymatic complexes found in cells. This work is an example of a tailored design of self-assembled supramolecular catalysts for one-pot tandem reactions, which is often not easy to achieve when working in homogeneous catalysis, and it could be extended to other subsets of incompatible reactions.<sup>16</sup>

## Acknowledgements

This work was supported by the EU (Marie Curie ITN-Smartnet). N. S. thanks EU for a Marie Curie ESR contract.



## Notes and references

- 1 T. L. Lohr and T. J. Marks, *Nat. Chem.*, 2015, **7**, 477–482.
- 2 D. Voet, J. G. Voet and C. W. Pratt, *Fundamentals of Biochemistry: Life at the Molecular Level*, Wiley, 2nd Ed, 2006.
- 3 (a) B. Helms, S. J. Guillaudeu, Y. Xie, M. McMurdo, C. J. Hawker and J. M. J. Fréchet, *Angew. Chem., Int. Ed.*, 2005, **44**, 6384–6387; (b) P. Li, C. -Y. Cao, Z. Chen, H. Liu, Y. Yu and W. -G. Song, *Chem. Commun.*, 2012, **48**, 10541–10543; (c) D. Vernekar and D. Jagadeesan, *Catal. Sci. Technol.*, 2015, **5**, 4029–4038; (d) J. Gao, X. Zhang, Y. Lu, S. Liu and J. Liu, *Chem.-Eur. J.*, 2015, **21**, 7403–7407; (e) L. M. Aguirre-Díaz, F. Gandara, M. Iglesias, N. Snejkó, E. Gutierrez-Puebla and M. A. Monge, *J. Am. Chem. Soc.*, 2015, **137**, 6132–6135.
- 4 (a) M. Raynal, P. Ballester, A. Vidal-Ferran and P. W. N. M. van Leeuwen, *Chem. Soc. Rev.*, 2014, **43**, 1660–1733; (b) M. Raynal, P. Ballester, A. Vidal-Ferran and P. W. N. M. van Leeuwen, *Chem. Soc. Rev.*, 2014, **43**, 1734–1787.
- 5 For instance: (a) M. O. Guler and S. I. Stupp, *J. Am. Chem. Soc.*, 2007, **129**, 12082; (b) F. Rodríguez-Llansola, B. Escuder and J. F. Miravet, *J. Am. Chem. Soc.*, 2009, **131**, 11478–11484; (c) F. Rodríguez-Llansola, J. F. Miravet and B. Escuder, *Chem. Commun.*, 2009, 7303–7305; (d) C. Berdugo, J. F. Miravet and B. Escuder, *Chem. Commun.*, 2013, **49**, 10608–10610; (e) Z. Huang, S. Guan, Y. Wang, G. Shi, L. Cao, Y. Gao, Z. Dong, J. Xu, Q. Luo and J. Liu, *J. Mater. Chem. B*, 2013, **1**, 2297–2304; (f) C. Zhang, X. Xue, Q. Luo, Y. Li, K. Yang, X. Zhuang, Y. Jiang, J. Zhang, J. Liu, G. Zou and X.-J. Liang, *ACS Nano*, 2014, **8**, 11715; (g) N. Singh, M. P. Conte, R. V. Ulijn, J. F. Miravet and B. Escuder, *Chem. Commun.*, 2015, **51**, 13213; (h) K. S. Lee and J. R. Parquette, *Chem. Commun.*, 2015, **51**, 15653; (i) M. Tena-Solsona, J. Nanda, S. Díaz-Oltra, A. Chotera, G. Ashkenasy and B. Escuder, *Chem.-Eur. J.*, 2016, **22**, 6687.
- 6 M. M. Safont-Sempere, G. Fernández and F. Würthner, *Chem. Rev.*, 2011, **111**, 5784–5814.
- 7 (a) M. Kölbél and F. M. Menger, *Langmuir*, 2001, **17**, 4490–4492; (b) A. Heeres, C. van der Pol, M. Stuart, A. Friggeri, B. L. Feringa and J. van Esch, *J. Am. Chem. Soc.*, 2003, **125**, 14252–14253; (c) A. Brizard, M. Stuart, K. van Bommel, A. Friggeri, M. de Jong and J. van Esch, *Angew. Chem., Int. Ed.*, 2008, **47**, 2063–2066.
- 8 D. K. Kumar and J. W. Steed, *Chem. Soc. Rev.*, 2014, **43**, 2080–2088.
- 9 (a) J. R. Moffat and D. K. Smith, *Chem. Commun.*, 2009, 316–318; (b) M. M. Smith and D. K. Smith, *Soft Matter*, 2011, **7**, 4856–4860; (c) K. J. Nagy, M. C. Giano, A. Jin, D. J. Pochan and J. P. Schneider, *J. Am. Chem. Soc.*, 2011, **133**, 14975–14977; (d) K. L. Morris, L. Chen, J. Raeburn, O. R. Sellick, P. Cotanda, A. Paul, P. C. Griffiths, S. M. King, R. K. O'Reilly, L. C. Serpell and D. J. Adams, *Nat. Commun.*, 2013, **4**, 1480; (e) D. Li, Y. Shi and L. Wang, *Chin. J. Chem.*, 2014, **32**, 123–127; (f) J. Raeburn, B. Alston, J. Kroeger, T. O. McDonald, J. R. Howse, P. J. Cameron and D. J. Adams, *Mater. Horiz.*, 2014, **1**, 241–246; (g) E. R. Draper, E. G. B. Eden, T. O. McDonald and D. J. Adams, *Nat. Chem.*, 2015, **7**, 848–852; (h) D. J. Cornwell, O. J. Daubney and D. K. Smith, *J. Am. Chem. Soc.*, 2015, **137**, 15486–15492; (i) C. C. Horgan, A. L. Rodriguez, R. Li, K. F. Bruggeman, N. Stupka, J. K. Raynes, L. Day, J. W. White, R. J. Williams and D. R. Nisbet, *Acta Biomater.*, 2016, DOI: 10.1016/j.actbio.2016.04.038.
- 10 (a) F. Rodríguez-Llansola, B. Escuder and J. F. Miravet, *Org. Biomol. Chem.*, 2009, **7**, 3091–3094; (b) M. Fontanillo, C. A. Angulo-Pachón, B. Escuder and J. F. Miravet, *J. Colloid Interface Sci.*, 2013, **412**, 65–71.
- 11 P. G. M. Wuts and T. W. Greene, Protection for the Carbonyl Group, in *Greene's Protective Groups in Organic Synthesis*, John Wiley & Sons: Hoboken, 2006, pp. 431–532.
- 12 S. Díaz-Oltra, C. Berdugo, J. F. Miravet and B. Escuder, *New J. Chem.*, 2015, **39**, 3785–3791.
- 13 C. Colquhoun, E. R. Draper, E. G. B. Eden, B. N. Cattoz, K. L. Morris, L. Chen, T. O. McDonald, A. E. Terry, P. C. Griffiths, L. C. Serpell and D. J. Adams, *Nanoscale*, 2014, **6**, 13719–13725.
- 14 B. Escuder, S. Martí and J. F. Miravet, *Langmuir*, 2005, **21**, 6776–6787.
- 15 S. Fleming, S. Debnath, P. W. J. M. Frederix, N. T. Hunt and R. V. Ulijn, *Biomacromolecules*, 2014, **15**, 1171–1184.
- 16 J. Lu, J. Dimroth and M. Weck, *J. Am. Chem. Soc.*, 2015, **137**, 12984–12989, and references cited therein.

