

CrossMark
click for updatesCite this: *RSC Adv.*, 2017, 7, 535

A highly sensitive metronidazole sensor based on a Pt nanospheres/polyfurfural film modified electrode

Jianzhi Huang,^a Xiaolei Shen,^a Ruili Wang,^a Qiang Zeng^{*ab} and Lishi Wang^{*a}

In this contribution, we combine the advantages of both Pt nanospheres and polyfurfural film and successfully develop a novel Pt nanospheres/polyfurfural film modified glassy carbon electrode (GCE) for electrochemical sensing of a nitro group containing pentatomic cyclic compound, metronidazole. In particular, the polyfurfural film and Pt nanospheres were handily obtained by a one-step electropolymerization method and a potential step method, respectively. Benefiting from their excellent synergistic catalytic properties, the Pt nanospheres/polyfurfural film/GCE shows significantly enhanced electrocatalytic activity towards metronidazole. A series of experimental parameters including the electropolymerization cycles of furfural, the deposition time of platinum, accumulation time, accumulation potential and pH of the supporting electrolyte for metronidazole was also investigated and optimized. The proposed sensor exhibited excellent selectivity, stability and reproducibility for the determination of metronidazole, providing a wide linear detection range from 2.5 to 500 $\mu\text{mol dm}^{-3}$ and a low detection limit of 50 nmol dm^{-3} ($S/N = 3$) under optimal experimental conditions. When the proposed sensor was applied to determine metronidazole in real human serum samples, it gave a satisfactory result.

Received 12th October 2016
Accepted 7th November 2016

DOI: 10.1039/c6ra25106d

www.rsc.org/advances

1. Introduction

Metronidazole, 1-(2-hydroxyethyl)-2-methyl-5-nitroimidazole (Chart 1), is the most important nitroimidazole derivative and is well-known for its anti-microbial properties.^{1,2} Because it is effective against anaerobic bacteria, trichomonas, amoeba and giardia, nowadays it has been extensively used for the treatment of anaerobic bacterial diseases^{3,4} and protozoal diseases such as trichomoniasis,⁵ amoebiasis^{6,7} and giardiasis.⁸ However, metronidazole can be genotoxic and carcinogenic when the

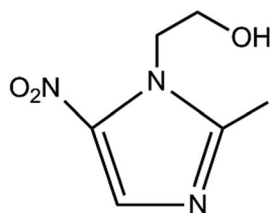


Chart 1 Chemical structure of metronidazole.

accumulated dose of metronidazole exceeds a 2 g daily dosage in human beings,^{9–11} presenting with some toxic reactions like seizures,¹² peripheral neuropathy,¹³ optic neuropathy¹⁴ and cerebellar ataxia.¹⁵ Therefore, it is significantly important and necessary to develop a highly sensitive and rapid method for the determination of metronidazole.

Several techniques such as high performance liquid chromatography,^{16,17} gas chromatography,¹⁸ supercritical fluid chromatography,¹⁹ ultraviolet spectrophotometry,²⁰ fluorescence spectrophotometry,²¹ capillary electrophoresis²² have been used for the detection and quantification of metronidazole, but high cost and time-consuming experimental processes limit their extensive application. In contrast with these methods, electrochemical method features high sensitivity, simplicity of operation, low cost and short analysis time. Therefore, sensors based on different chemically modified electrodes have been developed for the quantitative determination of metronidazole and achieved success.^{23–33}

Polymer-modified electrodes have attracted great attention in sensing applications.^{34–41} In particular, the strategy of electropolymerization is to immobilize a polymer film onto an electrode surface with multiple active sites and advantages of rapid synthesis, controllable film thickness, flexible charge transport and permeation characteristics by easily adjusting the electrochemical parameters.^{42,43} Previously, we have successfully prepared a polyfurfural film modified glassy carbon electrode (GCE) by a one-step electropolymerization

^aSchool of Chemistry and Chemical Engineering, South China University of Technology, Guangzhou 510641, People's Republic of China. E-mail: ceqzeng@scut.edu.cn; wanglish@scut.edu.cn; Tel: +86 20 87112906

^bState Key Laboratory of Pulp and Paper Engineering, South China University of Technology, Guangzhou 510641, People's Republic of China



method, which exhibits excellent electrocatalytic activity to the reduction of the nitro group in hexatomic aromatic compounds^{44,45} due to the promoted electron transfer by the conjugated π -electron backbones of polyfurfural film.^{44–46} Herein, we seek the possibility of the polyfurfural film for the catalytic reduction of the nitro group in a pentatomic cyclic compound of metronidazole. In order to improve the catalytic activity and sensitivity of the polyfurfural film modified electrode, a potential step method⁴⁷ was also used to prepare Pt nanospheres on the surface of the polyfurfural film, resulting in a proposed metronidazole sensor based on a Pt nanospheres/polyfurfural film modified GCE. With the excellent catalytic contributions of both Pt nanospheres and polyfurfural film, the Pt nanospheres/polyfurfural film/GCE shows superior electrocatalytic activity towards the nitro group in metronidazole. In contrast with the published electrochemical sensors, the proposed electrochemical sensor possesses a wider detection linear range and a lower detection limit for metronidazole, which successfully used in the quantitative determination of metronidazole in real human serum samples with a good performance. It is also highly sensitive, stable and reproducible for the proposed sensor. This developed technique would thus have an enormous meaning to pharmaceutical and biological analysis.

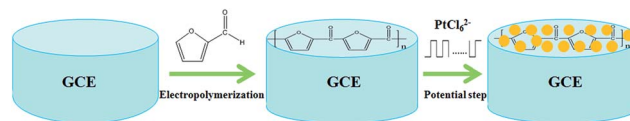
2. Experimental

2.1 Reagents

Metronidazole (99%, analytical grade) was purchased from Macklin Biochemical Co. Ltd. (Shanghai, China). Chloroplatinic acid hexahydrate (99.9%, analytical grade) was purchased from J&K Chemical (Beijing, China). Furfural (GC, 98%) was obtained from TCI Company. Sodium perchlorate and acetonitrile were obtained from Fuchen Chemical Reagent Company (Tianjin, China). Normal human serum samples were received from Huayueyang Biotechnology Co. Ltd (Beijing, China). All other reagents were of analytical grade and used without further purification. Britton–Robinson (BR) buffer solutions was prepared by mixing 0.04 mol dm⁻³ of boric acid, phosphoric acid and acetic acid, and then adjusted to the required pH value with 0.2 mol dm⁻³ of NaOH solution. All aqueous solutions were prepared with doubly distilled water.

2.2 Fabrication of Pt nanospheres/polyfurfural film/GCE

Before the fabrication of the polyfurfural film, a bare glassy carbon electrode (GCE, 3 mm diameter) was firstly polished with 0.3 μm alumina powders, and then successively polished with 0.05 μm alumina powders. After polishing, the bare GCE was rinsed ultrasonically with absolute alcohol and doubly distilled water, and then dried in the nitrogen stream. As reported previously,^{44–46} the electropolymerization of furfural was performed in acetonitrile containing furfural (0.01 mol dm⁻³) and sodium perchlorate (0.06 mol dm⁻³) by cyclic voltammetry at a scan rate of 100 mV s⁻¹ between -0.8 V and +2.8 V for 6 cycles. After electropolymerization, the polyfurfural film



Scheme 1 Schematic representation of preparation of Pt nanospheres/polyfurfural film/GCE.

modified glassy carbon electrode was rinsed with water to remove all physically adsorbed chemicals.

As followed, a potential step method⁴⁷ was employed to obtain Pt nanospheres in 0.5 mmol dm⁻³ H₂PtCl₆ solution containing 0.25 mol dm⁻³ H₂SO₄, where potential stepped repeatedly between 0.23 V and 0.60 V at 100 Hz for 450 s. These conditions allow proper amount of monodispersed Pt nanospheres to be formed without the overlapping effect of diffusion layer between Pt nuclei. The preparation of Pt nanospheres/polyfurfural film/GCE electrochemical sensor is concluded as Scheme 1.

2.3 Apparatus and method

Cyclic voltammetry (CV), electrochemical impedance spectroscopy (EIS) and differential pulse voltammetry (DPV) experiments were performed on a CHI 660D electrochemical workstation (Chenhua, Shanghai, China), a three-electrode configuration was used in all electrochemical experiments. In the process of electropolymerization, a bare GCE as the working electrode, a silver wire electrode as the reference electrode and a platinum wire as the counter electrode were used. While the Pt nanospheres/polyfurfural film/GCE working electrode and the saturated calomel reference electrode (SCE) were applied instead in the process of determination. The differential pulse voltammetry scans from -0.4 V to -1.0 V with an amplitude of 0.05 V, pulse width was 0.2 s, pulse period was 0.5 s, sampling width was 0.0167, increment was 0.004 V. For cyclic voltammetry, scan rate was 50 mV s⁻¹, sample interval was 0.001 V. Electrochemical impedance spectroscopy was obtained in 5 mmol dm⁻³ of K₃[Fe(CN)₆]/K₄[Fe(CN)₆] solution containing 0.1 mol dm⁻³ of KCl under open circuit potential with a frequency range from 0.1 Hz to 100 kHz and 50 mV amplitude. The electrochemical experiment parameters were same in all experiments. The surface morphology was characterized using a field emission scanning electron microscope (FE-SEM; Zeiss Ultra55, Germany). The working solution was deaerated with N₂ for 15 min prior to data acquisition during the experiments.

For the determination of metronidazole, the detection limit, C_m , was obtained using equation eqn (1):

$$C_m = 3S_b/m \quad (1)$$

where m is the slope of the calibration plot in the linear range, and S_b is the standard deviation of the blank response which is obtained from 20 replicate measurements of the blank BR buffer solution.



3. Results and discussion

3.1 Characterization of Pt nanospheres/polyfurfural film modified glassy carbon electrode

By contrast to a bare GCE (Fig. 1A), the SEM image of the polyfurfural film modified GCE shows a clear view of the polymer layer with a rough surface (Fig. 1B). After further modification, well dispersed Pt nanospheres are observed on the polymer surface and display uniform morphology and size distribution (Fig. 1C and D). For the high magnification SEM image of Pt nanospheres in Fig. 1E, the diameter of a Pt nanosphere is about 250 nm, showing a rough surface which makes Pt nanosphere possess a large surface area.

Fig. 2A shows the cyclic voltammograms of different electrodes in $5.0 \text{ mmol dm}^{-3} [\text{Fe}(\text{CN})_6]^{3-/4-}$ solution containing $0.1 \text{ mol dm}^{-3} \text{ KCl}$. The redox-label $[\text{Fe}(\text{CN})_6]^{3-/4-}$ displayed a reversible CV curve at the bare GCE (curve a). As seen from curve b, the cathodic and anodic peak currents were decreased obviously with a larger peak-to-peak separation (ΔE_p) when polyfurfural film was formed onto the surface of the bare GCE, suggesting that polyfurfural film hinders the electron transfer. Further modification of Pt nanospheres onto the surface of the polyfurfural film/GCE (curve c) results in the recovery of the cathodic and anodic peak currents and ΔE_p , indicating that the Pt nanospheres can improve the electron transfer rate due to

their excellent conductivity. Moreover, the EIS plots obtained after each modification process (Fig. 2B) are in accordance with the results concluded by CV and provide the confirmation that polyfurfural film and Pt nanospheres have been successfully modified on the electrode surface stepwise.

3.2 Electrochemical behavior of metronidazole at the Pt nanospheres/polyfurfural film/GCE

By cyclic voltammetry, the reduction of metronidazole was irreversible (Fig. 3A). In particular, two obvious reduction peaks were observed on a bare GCE (curve a), but only one reduction process was respectively observed on the Pt nanospheres/GCE (curve b), the bare platinum electrode (curve f), the polyfurfural film/GCE (curve c) and the Pt nanospheres/polyfurfural film/GCE (curve d), indicating two different reaction mechanisms. Comparing to the bare GCE, the electrodes respectively modified with Pt nanospheres, polyfurfural film and Pt nanospheres/polyfurfural film show a gradual increase on the reduction peak current of metronidazole due to the enhanced catalytical activities towards metronidazole. Interestingly, the current increase resulted by the polyfurfural film is more significant than that by Pt nanospheres. Therefore, this observation suggests that polyfurfural film plays the most important role in the catalytical reduction of metronidazole in this system.

In order to better explain the electrocatalytical reaction mechanism, differential pulse voltammetry was employed in the following studies, possessing highly sensitive. As reported

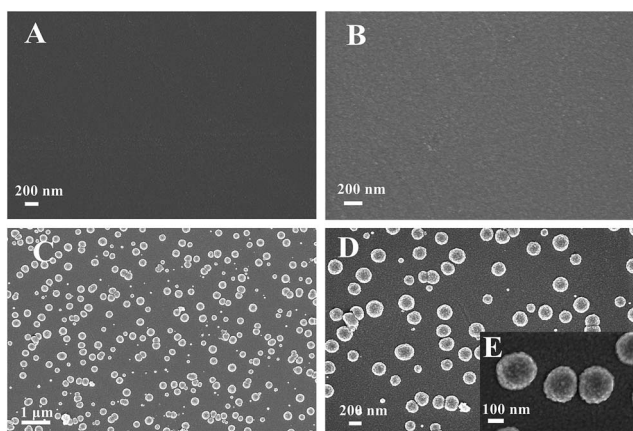


Fig. 1 SEM images of (A) bare GCE, (B) polyfurfural film/GCE, (C) and (D) Pt nanospheres/polyfurfural film/GCE, (E) the high magnification SEM image of the Pt nanospheres/polyfurfural film/GCE.

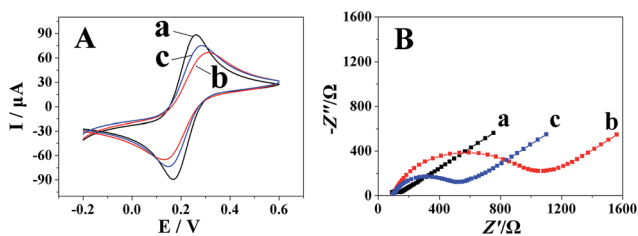


Fig. 2 (A) CVs and (B) EIS of (a) bare GCE, (b) polyfurfural film/GCE and (c) Pt nanospheres/polyfurfural film/GCE in $5.0 \text{ mM } [\text{Fe}(\text{CN})_6]^{3-/4-}$ containing 0.1 M KCl .

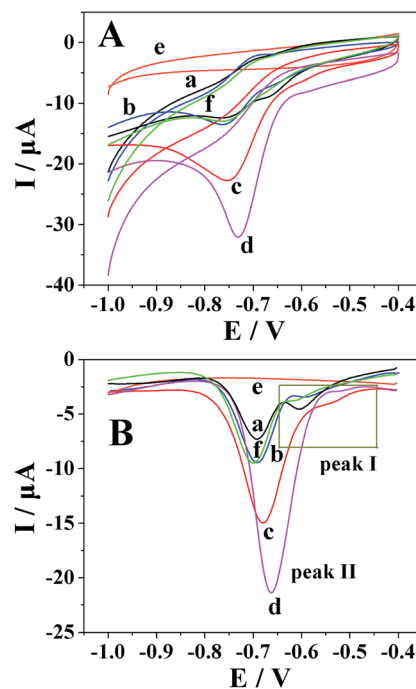


Fig. 3 CVs (A) and DPVs (B) of $500 \mu\text{mol dm}^{-3}$ metronidazole in BR (pH 10) buffer solution at (a) bare GCE, (b) Pt nanospheres/GCE, (f) bare platinum electrode, (c) polyfurfural film/GCE and (d) Pt nanospheres/polyfurfural film/GCE. CVs (A) and DPVs (B) of Pt nanospheres/polyfurfural film/GCE in BR (pH 10) buffer solution without metronidazole (e).



previously,^{23,33,48–51} the electrochemical reduction of metronidazole undergoes two possible reaction mechanisms (Scheme 2), the metronidazole molecules are initially adsorbed at the electrode surface and then reduced by two possible reaction mechanisms, where the $-\text{NO}_2$ group is initially reduced to $-\text{NO}_2^{\cdot-}$ (I-1) and then to $-\text{NHOH}$ (I-2) in the first mechanism and the $-\text{NO}_2$ group is directly converted to $-\text{NHOH}$ (II) in the second mechanism, respectively.

As shown in Fig. 3B, two obvious reduction peaks were observed on the bare GCE at -0.604 V and -0.696 V, respectively (curve a), indicating that the reduction processes of metronidazole mainly follow the first mechanism.^{23,33} By contrast, two reduction peaks were still observed on the Pt nanospheres/GCE (-0.592 V and -0.692 V, curve b), the bare platinum electrode (-0.614 V and -0.700 V, curve f) and the polyfurfural film/GCE (-0.536 V and -0.680 V, curve c), but the peak I became much weaker and the peak II is clearly enhanced. As the reduction of $-\text{NO}_2$ to $-\text{NO}_2^{\cdot-}$ (peak I) is a slow reaction process,^{23,33,51} the reduction of metronidazole is now mainly contributed by the mechanism II rather than the mechanism I, where the $-\text{NO}_2$ group can be directly reduced to $-\text{NHOH}$ due to the presence of Pt nanospheres or polyfurfural film. This observation is much more pronounced for the Pt nanospheres/polyfurfural film/GCE where the peak I is nearly disappeared, suggesting a one-step reaction mechanism triggered by both Pt nanospheres and polyfurfural film. The detailed descriptions about electrocatalytic reduction mechanism of metronidazole can be seen in Fig. 4. The green and red arrows represent two kinds of reaction mechanisms (mechanism I and II in Scheme 2), respectively. In particular, the majority of metronidazole molecules follow the primary mechanism and the minority was

involved in the reaction by the secondary mechanism under different conditions. The catalytic activity trend of different electrodes is also a consequence of the introduction of Pt nanospheres and polyfurfural film in order: Pt nanospheres/polyfurfural film/GCE > polyfurfural film/GCE > Pt nanospheres/GCE > bare GCE.

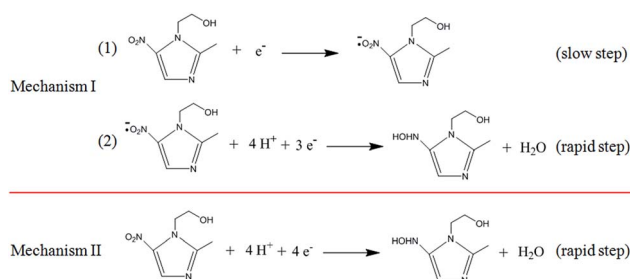
3.3 The effect of electropolymerization cycles of furfural and deposit time of platinum

The effects of electropolymerization cycles of furfural and deposit time of platinum on the reduction of $500 \mu\text{mol dm}^{-3}$ metronidazole in BR (pH 10) buffer solution have been investigated by differential pulse voltammetry. As shown in Fig. 5A, the reduction peak current reached the maximum value when 6 cycles were chosen. The polyfurfural film can be gradually formed on the GCE surface during the electropolymerization and its thickness and catalytic activity will be determined by the electropolymerization cycles of furfural. Less than 6 cycles, the polyfurfural film is probably not fully produced and not able to provide sufficient catalytic active sites to metronidazole. By contrast, if over 6 cycles, the formed polyfurfural film would be too thick and increase the resistance of the electrode surface to hinder electron transfer. Therefore, the optimized furfural electropolymerization cycles have been chosen at 6 cycles and as the optimal condition in subsequent experiments.

In order to improve the catalytic properties and electronic conductivity of polyfurfural film, Pt nanospheres were allowed to grow at the surface of polyfurfural film/GCE by a potential step method, resulting in Pt nanospheres/polyfurfural film/GCE. As shown in Fig. 5B, the reduction peak current of metronidazole increased significantly with the increase of the Pt deposit time and reached the maximum value when the deposit time was 450 s. However, after 450 s, the reduction peak current gradually decreased. Therefore, the Pt deposit time of 450 s was chosen in subsequent experiments.

3.4 Effect of scan rate

Fig. 6 shows the cyclic voltammograms of Pt nanospheres/polyfurfural film/GCE in $500 \mu\text{mol dm}^{-3}$ metronidazole at different scan rates. In Fig. 6, the reduction peak current showed linearity with the scan rate, and the linear regression equation can be expressed as $I_p (\mu\text{A}) = -0.0767v (\text{mV s}^{-1}) - 6.1189$ ($R = 0.9984$), which indicates the reduction of the



Scheme 2 The proposed electrocatalytic reactions of metronidazole.

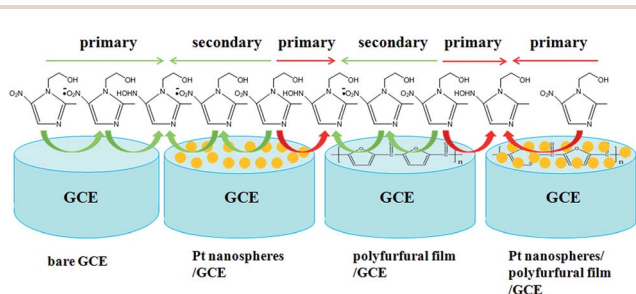


Fig. 4 The electrocatalytic reaction mechanisms of metronidazole.

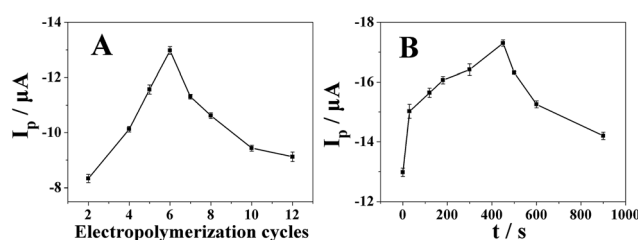


Fig. 5 The effect of (A) electropolymerization cycles of furfural and (B) deposit time of platinum on the reduction peak current of $500 \mu\text{mol dm}^{-3}$ metronidazole in BR (pH 10) buffer solution ($n = 3$).



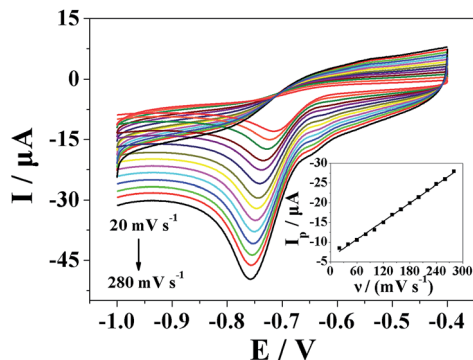


Fig. 6 CVs of metronidazole ($500 \mu\text{mol dm}^{-3}$) at the Pt nanospheres/polyfurfural film/GCE in BR (pH 10) buffer solution at different scan rates (20, 40, 60, 80, 100, 120, 140, 160, 180, 200, 220, 240, 260, 280 mV s^{-1}). Inset: the relationship between the reduction peak current and scan rate.

metronidazole is typical adsorption controlled process. The pre-peak at *ca.* -0.65 V in the CV represents a reaction process (I-1) in mechanism I, where the $-\text{NO}_2$ group is initially reduced to $-\text{NO}_2^{\cdot-}$ (I-1) and then to $-\text{NHOH}$ (I-2) in the first mechanism, the $-\text{NO}_2$ group is initially reduced to $-\text{NO}_2^{\cdot-}$ (I-1) is at *ca.* -0.65 V . It is proposed that metronidazole molecules are initially adsorbed at the surface-active sites of Pt nanospheres/polyfurfural film/GCE and then quickly reduced. Because the reduction of the metronidazole is typical adsorption controlled process, it is necessary to study the effect of accumulation time and accumulation potential.

3.5 The effect of accumulation time and accumulation potential

As shown in Fig. 7A, the reduction peak current by differential pulse voltammetry increased gradually with the accumulation time and reached the maximum value when the accumulation time is 360 s (accumulation potential was -0.4 V). The reduction peak current almost remained the same after 360 s due to the saturation of surface active catalytical sites of Pt nanospheres/polyfurfural film/GCE. Thus, the optimal accumulation time of 360 s was employed in further experiments.

With the optimal accumulation time determined above, we further studied the effect of accumulation potential on reduction peak current of metronidazole. As shown in Fig. 7B, the

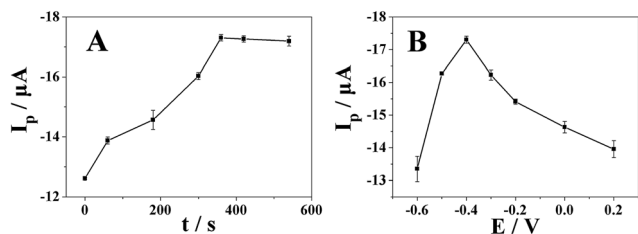


Fig. 7 The effect of accumulation time (A) and accumulation potential (B) on the reduction peak current of $500 \mu\text{mol dm}^{-3}$ metronidazole in BR (pH 10) buffer solution at the Pt nanospheres/polyfurfural film/GCE ($n = 3$).

reduction peak current increased gradually with the increase of accumulation potential and reached the maximum value when the accumulation potential is -0.4 V . After -0.4 V , the reduction peak current decreased with the increase of accumulation potential. Therefore, an accumulation potential at -0.4 V was chosen for determination of metronidazole in subsequent experiments.

3.6 pH effect on electrochemical behavior of metronidazole

The effect of solution pH on the electrochemical behavior of metronidazole at the Pt nanospheres/polyfurfural film/GCE was investigated with BR buffer solution in the pH range from 6 to 11 (Fig. 8A). In Fig. 8A, the reduction peak potential (E_p) shift to more negative values with the increase of pH value. Fig. 8B shows good linearity between the reduction peak potential of metronidazole and pH values range from 6 to 11, with the linear regression equation $E_p = -0.05424 \text{ pH} - 0.14137$ ($R = 0.9998$). The slope of the E_p -pH plot is $-54.24 \text{ mV pH}^{-1}$, which closes to -59.2 mV pH^{-1} , indicated that numbers of the transferring proton and electron in the reaction were same, the result further confirmed the proposed reaction mechanism in Fig. 4 that the reduction of metronidazole at the Pt nanospheres/polyfurfural film/GCE follows mechanism II, where the $-\text{NO}_2$ group of metronidazole can be directly reduced to $-\text{NHOH}$ group involving four electrons and four protons transfer. As shown in Fig. 8C, pH values influenced the reduction peak current of metronidazole at the Pt nanospheres/polyfurfural film/GCE vary from 6 to 11, when the pH value was 10, the reduction peak current reached to a maximum value. Therefore, a pH value of 10 was chosen as the optimum pH value in subsequent quantification.

3.7 The quantitative determination of metronidazole

The quantitative determination of metronidazole at the Pt nanospheres/polyfurfural film/GCE under optimal conditions

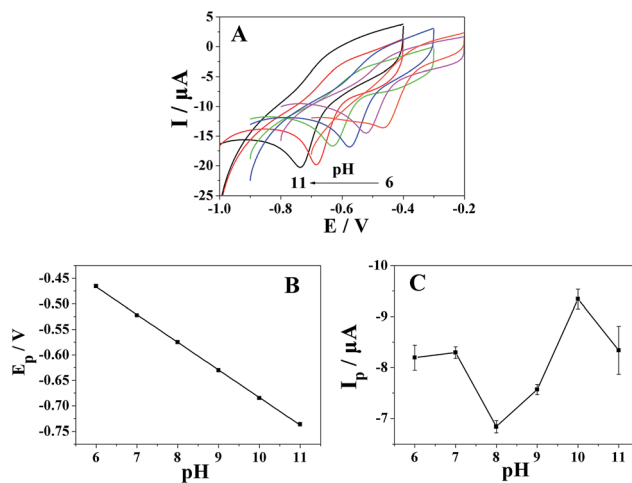


Fig. 8 CVs of (A) metronidazole ($200 \mu\text{mol dm}^{-3}$) in BR buffer solution at different pH values on the Pt nanospheres/polyfurfural film/GCE. The relationship of (B) pH vs. reduction peak potential and (C) pH vs. reduction peak current (I_p) for metronidazole ($n = 3$).



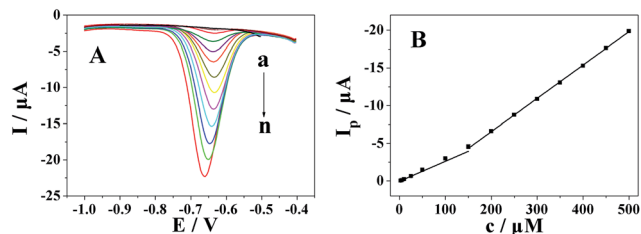


Fig. 9 (A) DPVs of metronidazole at the Pt nanospheres/polyfurfural film/GCE in BR (pH 10) buffer solution. Concentrations: 2.5, 5, 10, 25, 50, 100, 150, 200, 250, 300, 350, 400, 450 and 500 $\mu\text{mol dm}^{-3}$ (from a to n). (B) Linear relationship between reduction peak current and concentration.

addressed above was achieved by DPV. As shown in Fig. 9, the reduction peak current of metronidazole at the Pt nanospheres/polyfurfural film/GCE increased linearly with concentration ranges of 2.5–150 $\mu\text{mol dm}^{-3}$ and 150–500 $\mu\text{mol dm}^{-3}$, and the linear regression equations were $I_p (\mu\text{A}) = -0.02653c (\mu\text{M}) + 0.04097$ ($R = 0.9948$) and $I_p (\mu\text{A}) = -0.04418c (\mu\text{M}) + 2.28704$ ($R = 0.9999$), respectively. The detection limit for metronidazole was 50 nmol dm^{-3} ($S/N = 3$).

Moreover, the Pt nanospheres/polyfurfural film/GCE showed a wider linear range and a lower detection limits compared with the reported electrochemical sensors^{48,49,52–57} which were shown in Table 1.

3.8 Selectivity, reproducibility and stability of the Pt nanospheres/polyfurfural film/GCE

Selectivity is one of the key factors which can evaluate the practical application of the proposed sensor. In this study, various possible interferences from some inorganic ions, organic ions and organic compounds have been investigated by differential pulse voltammetry. In Table 2, the response of proposed sensor to metronidazole was not affected by additions of 1000-fold concentrations of inorganic ions (such as NO_3^- , Cl^- , PO_4^{3-} , SO_4^{2-} , F^- , CO_3^{2-} , Na^+ , Ca^{2+} , K^+ , Mg^{2+} , Zn^{2+} , NH_4^+ and K^+), organic ions (such as Ac^-) (signal change below 4%) and 100-fold concentrations of some organic compounds such

Table 1 Comparison of performances of the Pt nanospheres/polyfurfural film/GCE with other electrodes

Electrode	Linear range ($\mu\text{mol dm}^{-3}$)	Limit of detection (nmol dm^{-3})	Ref.
Gr-IL/GCE	0.1–25	47	48
Activated GCE	2–600	1100	49
Co/GCE	0.4–100	200	52
SWCNT/GCE	0.1–200	63	53
Carbon fiber/MDE	1–22	500	54
Poly(chromotrope 2B)/GCE	10–400	330	55
HMDE	0.23–1.8	36	56
P-AgSA-CE	20–100	2000	57
Pt nanospheres/polyfurfural film/GCE	2.5–500	50	This work

Table 2 Effect of different interferences on the reduction peak current signal of metronidazole ($200 \mu\text{mol dm}^{-3}$) at the Pt nanospheres/polyfurfural film/GCE in BR (pH 10) buffer solution

Interference	Concentration (mol dm^{-3})	Signal change (%)
NaNO_3	0.2	−2.49
CaCl_2	0.2	3.07
K_3PO_4	0.2	−3.83
MgSO_4	0.2	−3.16
$\text{Zn}(\text{Ac})_2$	0.2	−3.83
NH_4F	0.2	−1.70
K_2CO_3	0.2	−2.86
Ascorbic acid	0.02	2.84
Oxalic acid	0.02	−0.81
Dopamine	0.02	4.89
Cystine	0.02	0.41
Alanine	0.02	4.93
Glucose	0.02	−2.15
Citric acid	0.02	0.83
Tartaric acid	0.02	3.54
Imidazole	0.002	1.89
Benzimidazole	0.002	2.10
2-Methylimidazole	0.002	1.52
Nifedipine	0.002	2.50
Nitrendipine	0.002	3.48
Nitrophenol	0.002	3.25

as ascorbic acid, oxalic acid, dopamine, cystine, alanine, glucose, citric acid and tartaric acid (signal change below 5%). Moreover, interferences resulted from 10-fold concentrations of other imidazoles (imidazole, benzimidazole and 2-methylimidazole, signal change below 3%) and other molecules containing nitro group (nifedipine, nitrendipine and nitrophenol, signal change below 4%) were negligible. The results showed that the proposed sensor is superior for the determination of metronidazole with excellent selectivity.

Reproducibility of the electrochemical sensors is a key factor in their application. To estimate the method's reproducibility, six modified electrodes were fabricated and the reduction peak current of metronidazole at each electrode was obtained under the same conditions. The relative standard deviation (RSD) of measurements was calculated to be 0.57% for metronidazole, indicated that the proposed sensor possesses high reproducibility. Furthermore, stability of the electrochemical sensor is another important factor in their application and development. The reduction peak current of metronidazole remained 98.51% of its initial value when the as-prepared electrode was stored at 4 °C in a refrigerator for 1 month, suggested that the proposed sensor possesses excellent stability. Therefore, from the results above, with excellent selectivity, reproducibility and stability, the Pt nanospheres/polyfurfural film/GCE is promising for determination of metronidazole.

3.9 Analytical application for real samples

To investigate the analytical reliability and application potential of the Pt nanospheres/polyfurfural film/GCE for determination of metronidazole in real sample, human serum samples were



Table 3 Determination of metronidazole in real human serum samples

Sample	Added ($\mu\text{mol dm}^{-3}$)	Found ^a ($\mu\text{mol dm}^{-3}$)	Recovery (%)	RSD (%)
1	250	253.11 \pm 0.37	101.24	0.06
2	150	153.30 \pm 1.76	102.20	0.46
3	75	76.33 \pm 2.23	101.77	1.18
4	35	33.77 \pm 0.74	96.49	0.89
5	15	14.44 \pm 0.99	96.27	2.77

^a Sample responses are expressed as a confidence interval of 95% probability ($n = 3$).

used for quantitative analysis by the standard-addition technique. Three parallel experiments were performed on all measurements. As shown in Table 3, the average recoveries were between 96.27% and 102.20%, indicating that the Pt nanospheres/polyfurfural film/GCE can be successfully used for the determination of metronidazole in real samples.

4. Conclusions

In summary, a Pt nanospheres/polyfurfural film modified glassy carbon electrode has been fabricated for electrochemical sensing of metronidazole, by a one-step electropolymerization method and a potential step method, respectively. The proposed sensor was successfully applied to the determination of metronidazole in real human serum samples, which exhibited excellent electrocatalytic activity to the reduction of metronidazole. Moreover, the Pt nanospheres/polyfurfural film/GCE showed better sensitivity, stability and reproducibility compared with other sensors, which also had a wider linear ranges (2.5–500 $\mu\text{mol dm}^{-3}$) and a lower detection limit (50 nmol dm^{-3}). This work may provide a new and effective analytical platform for determination of metronidazole potentially in applications of real pharmaceutical and biological samples.

Acknowledgements

Financial support from the National Natural Science Foundation of China (Grant No. 21475046, 21427809, 21645004) and State Key Laboratory of Pulp and Paper Engineering (201623) are gratefully acknowledged. We also acknowledge the Fundamental Research Funds for the Central Universities (No. 2015ZM050 and 2015ZP028).

References

- N. C. Desai, A. S. Maheta, K. M. Rajpara, V. V. Joshi, H. V. Vaghani and H. M. Satodiya, *J. Saudi Chem. Soc.*, 2014, **18**, 963–971.
- A. M. Jarrad, T. Karoli, A. Debnath, C. Y. Tay, J. X. Huang, G. Kaeslin, A. G. Elliott and Y. Miyamoto, *Eur. J. Med. Chem.*, 2015, **101**, 96–102.
- N. Dione, S. Khelaifia, J. C. Lagier and D. Raoult, *Int. J. Antimicrob. Agents*, 2015, **45**, 537–540.
- A. Katsandri, A. Avlami, A. Pantazatou, G. L. Petrikkos, N. J. Legakis, J. Papaparaskevas and N. Hellenic, *Diagn. Microbiol. Infect. Dis.*, 2006, **55**, 231–236.
- L. A. Dunn, K. T. Andrews, J. S. McCarthy, J. M. Wright, T. S. Skinner, P. Upcroft and J. A. Upcroft, *Int. J. Antimicrob. Agents*, 2007, **29**, 98–102.
- B. Swami, D. Lavakusulu and C. S. Devi, *Curr. Med. Res. Opin.*, 1977, **5**, 152–156.
- H. Qureshi, A. Ali, R. Baqai and W. Ahmed, *J. Int. Med. Res.*, 1997, **25**, 167–170.
- A. V. Scorza and M. R. Lappin, *J. Feline Med. Surg.*, 2004, **6**, 157–160.
- M. W. Carroll, D. Jeon, J. M. Mountz, J. D. Lee, Y. J. Jeong, N. Zia, M. Lee, J. Lee, L. E. Via and S. Lee, *Antimicrob. Agents Chemother.*, 2013, **57**, 3903–3909.
- J. Han, L. Zhang, S. Yang, J. Wang and D. Tan, *Bull. Environ. Contam. Toxicol.*, 2014, **92**, 196–201.
- A. Kuriyama, J. L. Jackson, A. Doi and T. Kamiya, *Clin. Neuropharmacol.*, 2011, **34**, 241–247.
- J. P. Ferroir, C. Corpechot, A. Freudenreich and A. Khalil, *Rev. Neurol.*, 2009, **165**, 828–830.
- A. Etxeberria, S. Lonnaville, M. P. Rutgers and M. Gille, *Rev. Neurol.*, 2012, **168**, 193–195.
- N. M. McGrath, B. Kent-Smith and D. M. Sharp, *Clin. Exp. Ophthalmol.*, 2007, **35**, 585–586.
- M. P. Prabhakaran, M. Zamani, B. Felice and S. Ramakrishna, *Mater. Sci. Eng., C*, 2015, **56**, 66–73.
- A. Turcant, A. Premel-Cabic, A. Cailleux and P. Allain, *Clin. Chem.*, 1991, **37**, 1210–1215.
- B. M. Tashtoush, E. L. Jacobson and M. K. Jacobson, *Drug Dev. Ind. Pharm.*, 2008, **34**, 840–844.
- C. Ho, D. W. Sin, K. M. Wong and H. P. Tang, *Anal. Chim. Acta*, 2005, **530**, 23–31.
- V. R. Bari, U. J. Dhorda and M. Sundaresan, *Anal. Chim. Acta*, 1998, **376**, 221–225.
- A. K. Mishra, A. Kumar, A. Mishra and H. V. Mishra, *J. Nat. Sci., Biol. Med.*, 2014, **5**, 261–264.
- S. Z. Tan, J. H. Jiang, B. N. Yan, G. L. Shen and R. Q. Yu, *Anal. Chim. Acta*, 2006, **560**, 191–196.
- Y. Lin, Y. Su, X. Liao, N. Yang, X. Yang and M. F. Choi, *Talanta*, 2012, **88**, 646–652.
- H. B. Ammar, M. B. Brahim, R. Abdelhedi and Y. Samet, *Mater. Sci. Eng., C*, 2016, **59**, 604–610.
- Y. Gu, X. Yan, W. Liu, C. Li, R. Chen, L. Tang, Z. Zhang and M. Yang, *Electrochim. Acta*, 2015, **152**, 108–116.
- G. Yang, F. Zhao and B. Zeng, *Electrochim. Acta*, 2014, **135**, 154–160.
- E. Roy, S. K. Maity, S. Patra, R. Madhuri and P. K. Sharma, *RSC Adv.*, 2014, **4**, 32881–32893.
- S. Sadeghi, M. Hemmati and A. Garmroodi, *Electroanalysis*, 2013, **25**, 316–322.
- Y. Gu, X. Yan, C. Li, B. Zheng, Y. Li, W. Liu, Z. Zhang and M. Yang, *Biosens. Bioelectron.*, 2016, **77**, 393–399.
- Y. B. Mollamahale, M. Ghorbani, M. Ghalkhani, M. Vossoughi and A. Dolati, *Electrochim. Acta*, 2013, **106**, 288–292.



- 30 W. Liu, J. Zhang, C. Li, L. Tang, Z. Zhang and M. Yang, *Talanta*, 2013, **104**, 204–211.
- 31 B. Rezaei and S. Damiri, *Electrochim. Acta*, 2010, **55**, 1801–1808.
- 32 Y. Liu, J. Liu, H. Tang, J. Liu, B. Xu, F. Yu and Y. Li, *Sens. Actuators, B*, 2015, **206**, 647–652.
- 33 Y. Gu, W. Liu, R. Chen, L. Zhang and Z. Zhang, *Electroanalysis*, 2013, **25**, 1209–1216.
- 34 H. Li, H. Guan, H. Dai, Y. Tong, X. Zhao, W. Qi, S. Majeed and G. Xu, *Talanta*, 2012, **99**, 811–815.
- 35 K. C. Lin, C. Y. Yin and S. M. Chen, *Sens. Actuators, B*, 2011, **157**, 202–210.
- 36 M. R. Ganjali, P. Norouzi, F. Faridbod and H. Pirelahi, *J. Chin. Chem. Soc.*, 2007, **54**, 55–61.
- 37 K. Giribabu, R. Suresh, R. Manigandan, S. Munusamy, S. P. Kumar, S. Muthamizh and V. Narayanan, *Analyst*, 2013, **138**, 5811–5818.
- 38 F. Ismail and S. B. Adeloju, *Electroanalysis*, 2014, **26**, 2701–2709.
- 39 M. Li, W. Liu, J. P. Correia, A. C. Mourato, A. S. Viana and G. Jin, *Electroanalysis*, 2014, **26**, 374–381.
- 40 S. He, Z. Chen, Y. Yu and L. Shi, *RSC Adv.*, 2014, **4**, 45185–45190.
- 41 S. Chitravathi, B. E. Swamy, G. P. Mamatha and B. S. Sherigara, *J. Electroanal. Chem.*, 2012, **667**, 66–75.
- 42 H. L. Yan, H. L. Xiao, Q. J. Xie, J. L. Liu, L. G. Sun, Y. P. Zhou, Y. Zhang, L. Chao, C. Chen and S. Z. Yao, *Sens. Actuators, B*, 2015, **207**, 167–176.
- 43 X. Chen, Q. Zhang, C. Qian, N. Hao, L. Xu and C. Yao, *Biosens. Bioelectron.*, 2015, **64**, 485–492.
- 44 T. Y. Wei, X. J. Huang, Q. Zeng and L. S. Wang, *J. Electroanal. Chem.*, 2015, **743**, 105–111.
- 45 Q. Zeng, T. Y. Wei, M. Wang, X. J. Huang, Y. S. Fang and L. S. Wang, *Electrochim. Acta*, 2015, **186**, 465–470.
- 46 Y. H. Fu, Y. P. Lin, T. S. Chen and L. S. Wang, *J. Electroanal. Chem.*, 2012, **687**, 25–29.
- 47 N. Tian, Z. Y. Zhou, S. G. Sun, Y. Ding and Z. L. Wang, *Science*, 2007, **316**, 732–735.
- 48 J. Peng, C. Hou and X. Hu, *Sens. Actuators, B*, 2012, **169**, 81–87.
- 49 S. A. Ozkan, Y. Ozkan and Z. Senturk, *J. Pharm. Biomed. Anal.*, 1998, **17**, 299–305.
- 50 S. F. Lu, K. B. Wu, X. P. Dang and S. S. Hu, *Talanta*, 2004, **63**, 653–657.
- 51 V. Vyskocil and J. Barek, *Curr. Org. Chem.*, 2011, **15**, 3059–3076.
- 52 Y. Zhi, J. B. Hu, Z. D. Wu and Q. L. Li, *Anal. Lett.*, 1998, **31**, 429–437.
- 53 A. Salimi, M. Izadi, R. Hallaj and M. Rashidi, *Electroanalysis*, 2007, **19**, 1668–1676.
- 54 P. N. Bartlett, E. Ghoneim, G. El-Hefnawy and I. El-Hallag, *Talanta*, 2005, **66**, 869–874.
- 55 X. Li and G. Xu, *J. Food Drug Anal.*, 2014, **22**, 345–349.
- 56 Y. Gui, Y. N. Ni and S. Kokot, *Chin. Chem. Lett.*, 2011, **22**, 591–594.
- 57 V. Vyskocil, T. Navratil, A. Danhel, J. Dedik, Z. Krejcova, L. Skvorova, J. Tvrdivkova and J. Barek, *Electroanalysis*, 2011, **23**, 129–139.

