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Peptide GEQQQPGM derived from rice α -globulin reduces the risk of atherosclerosis in hamsters by improving vascular endothelial cells injury†

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A previous study has confirmed that oral administration of rice α -globulin (100 mg per day per kg bodyweight) efficiently prevents atherosclerosis disease in male apolipoprotein E-deficient (ApoE^{-/-}) mice. However, the key peptide in rice α -globulin after gastrointestinal digestion and absorption as well as its anti-atherosclerosis mechanism still remain unclear. We obtained the absorbable part in the simulated digestive products of rice α -globulin by everting the intestinal sacs followed by purification using gel filtration chromatography and high performance liquid chromatography. The peptide GEQQQPGM was confirmed to be one of the key fragments in rice α -globulin by LC-MS/MS (liquid chromatography-mass spectrometry/mass spectrometry). We investigated the hypocholesterolemic effect of the peptide in hamsters fed a hypercholesterolemic diet. The synthetic peptide GEQQQPGM was orally administrated to hamsters at a dose of 100 mg per kg bodyweight, and deionized water was orally administrated to the normal and control hamsters for 30 days. In the peptide group, plasma total and LDL cholesterol concentrations and atherogenic index were significantly lower than those in the control group. To investigate the anti-atherosclerosis mechanism of the peptide, the effects of the GEQQQPGM peptide derived from rice α -globulin on vascular endothelial cells injury were studied. The cell model was established using 10 ng mL⁻¹ of tumor necrosis factor- α (TNF- α). The study was performed on HUVECs that were pretreated with 50 μ g mL⁻¹ of GEQQQPGM. The cell viability and indicators of inflammation and oxidative stress were measured. The decreased viability of the HUVECs induced by TNF- α was significantly increased by GEQQQPGM. The peptide obviously reduced the levels of monocyte chemoattractant protein-1 (MCP-1), intercellular adhesion molecule-1 (ICAM-1), and vascular cell adhesion molecule-1 (VCAM-1). It also effectively affected the antioxidant indices such as it decreased the levels of nitric oxide (NO), inducible nitric oxide synthase (iNOS), malondialdehyde (MDA), and reactive oxygen species (ROS), whereas increased the activities of glutathione peroxidase (GSH-Px) and superoxide dismutase (SOD). Furthermore, GEQQQPGM inhibited nuclear translocation and expression of p65 protein by decreasing the inhibitor of κ B kinase α (IKK α) protein and up-regulating the inhibitor of nuclear factor κ B kinase (I κ B) protein. These results demonstrated that GEQQQPGM reduced TNF- α -induced vascular endothelial cell injury by inhibiting the inflammatory response and oxidative stress, thereby reducing the risk of atherosclerosis.

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1. Introduction

Vascular endothelial dysfunction is considered as an important risk factor for the development of atherosclerosis, which is

significantly involved in the pathogenesis of cardiovascular diseases.^{1,2} Inflammatory response and oxidative stress are the major causes of reduced endothelial nitric oxide (NO) availability in injured vascular endothelial cells, which can be triggered by the tumor necrosis factor- α (TNF- α).^{3,4} TNF- α , as the most common pro-inflammatory cytokine in atherosclerosis, induces inflammation and oxidative stress, thereby regulating the expression of some cell molecules in vascular endothelial cells.^{5,6} Therefore, a TNF- α -induced cell injury model is widely used to investigate the protective effects of food functional components on vascular endothelial cells.

Numerous studies on experimental animals and humans have proven that the intake of food proteins, such as rice,

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soybean, and oat proteins, can significantly reduce the risk of atherosclerosis by lowering plasma lipids.^{7–9} Food proteins are digested into amino acids and peptides, and some of these peptides can be directly absorbed by the body such that to prevent lifestyle diseases such as atherosclerosis, hypertension, and hyperlipidemia.^{10,11}

Matsui *et al.* reported that the oral administration of the dipeptide WH (10 or 100 mg per day per kg bodyweight) derived from soybean globulin inhibited progressive atherosclerotic lesions in ApoE^{-/-} mice without any alteration of the lipid profiles.¹² They further confirmed that WH improved atherosclerosis by inhibiting the phosphorylation of voltage-dependent L-type Ca²⁺ channels in rat vascular smooth muscle cells.¹³ Similarly, the tripeptide HGI and HGK derived from soybean glycinin have been reported to suppress the intracellular Ca²⁺ concentration in vascular smooth muscle cells.¹⁴ Huang *et al.* reported that the tripeptide IRW derived from egg protein reduced the TNF- α -induced inflammatory response and oxidative stress and reduced the levels of superoxide ions in human umbilical vein endothelial cells (HUVECs).⁶

Endothelial dysfunction in atherosclerosis generates reactive oxygen species (ROS) that can be detoxified by anti-oxidative enzymes.⁷ Moreover, pro-inflammatory nuclear factor κ B (NF- κ B) has been recognized as a major transcription factor that mediates the key steps in the development of atherosclerosis. The tripeptide IRW, as a signal transduction messenger, reduced the production of ROS by inhibiting the inflammatory activation of NF- κ B.⁶

A previous study has confirmed that the oral administration of rice α -globulin (100 mg per day per kg bodyweight) efficiently reduces the en face lesion area in the aorta and the lesion size in the aortic root and thus prevents atherosclerosis disease in male apolipoprotein E-deficient (ApoE^{-/-}) mice without any alteration of the plasma lipid levels.¹⁵ We speculate that there may be functional peptide fragments in rice α -globulin and these peptides play a role in protecting vascular cell injury. However, the key peptide in rice α -globulin after gastrointestinal digestion and absorption together with its anti-atherosclerosis mechanism still remains unclear.

2. Materials and methods

2.1. Materials

TNF- α , fetal calf serum (FBS), and penicillin-streptomycin antibiotics were purchased from Invitrogen-Gibco (USA). A protein quantitative kit (BCA) was purchased from Sainobo Co., Ltd. (Beijing, China). A cell counting kit-8 (CCK-8) was purchased from Dojindo Co. Ltd. (Shanghai, China). The assay kits for NO, iNOS, ROS, SOD, GSH-PX, and MDA were purchased from Beyotime Biotechnology Co., Ltd. (Shanghai, China).

2.2. Preparation of the peptide

The α -globulin digestion experiments were carried out using a pepsin trypsin sequential digestion *in vitro* digestion model. First, the protein was digested using pepsin in HCl at pH 2.0

and 37 °C for 2 h. The pH value was adjusted to 5.3 using a 0.09 mol L⁻¹ NaHCO₃ solution. After trypsin was added, the pH value was adjusted to 7.8 using a 1 mol L⁻¹ NaOH solution. The sample was digested at 37 °C for 2 h. The sample was heated at 100 °C for 10 min to terminate the reaction. After centrifugation at 2000 \times g for 20 min, the supernatant was obtained and frozen for storage.

The peptide absorption experiments were carried out using everted intestinal sacs *in vitro*. Male hamsters (four-week-old) were obtained from Vital River Lab Animal Technology Co, Ltd. (Beijing, China). The hamsters were fasted for 16 h and then anesthetized. After the small intestine was removed, it was washed with Krebs-Ringer solution (118.4 mmol L⁻¹ NaCl, 4.7 mmol L⁻¹ KCl, 1.2 mmol L⁻¹ KH₂PO₄, 1.2 mmol L⁻¹ MgSO₄·7H₂O, 2.5 mmol L⁻¹ CaCl₂·2H₂O, 11.7 mmol L⁻¹ C₆H₁₂O₆, and 26.5 mmol L⁻¹ NaHCO₃) to remove the contents. The small intestine was everted with glass, and its lower part was tightened. Then, 2 mL of Krebs-Ringer solution was added to the small intestine, and it was placed in a test tube at 37 °C. The test tube contained Krebs-Ringer solution and was incorporated with a mixture of gases (95% O₂ and 5% CO₂, 2 mL min⁻¹). After adding the protein digestive juice, the absorption experiments were carried out at 37 °C for 90 min and then the samples were taken from the test tubes.

The 30 mg mL⁻¹ absorbed product was preliminarily separated by a glass chromatography column (Sephadex G-15: 1.6 \times 50 cm, Bio-Rad, Hercules, CA, USA) using ultrapure water at a flow rate of 1 mL min⁻¹. The elution peaks were obtained at 220 nm (UV detector) and freeze-dried. Next, the 20 mg mL⁻¹ samples were purified using RP-HPLC (Agilent Prostar 218, Agilent Co., USA). The chromatographic conditions were as follows: Innoval C18 (21.2 \times 250 mm, 10 μ m); sample volume = 1 mL; flow rate = 5 mL min⁻¹; and detection wavelength = 220 nm.

After this, the amino acid sequence of the peptides was determined by LC-mass spectrometry (MS)/MS (Agilent 1100, Agilent Co., USA; LCQ Decap, Thermo Co., USA). The LC-MS conditions were as follows: chromatographic column (Zorbax SB-C18: 2.1 \times 150 mm); mobile phase A: water (0.1% TFA) and B: acetonitrile (0.1% TFA); gradient: 0–40 min, 5–95% B; 40–50 min; 95–5% B; injection volume = 20 μ L; and flow rate = 0.2 mL min⁻¹. The MS conditions were as follows: ESI spray voltage = 4.5 kV; capillary temperature = 300 °C; positive ion detection; MS/MS dynamic scanning was data-dependent scan; dynamic elimination times = 1 and 0.5 min; collision energy for MS 2 = 35%; and collision energy and scanning range: *m/z* 900–1400 and *m/z* 1400–2000, respectively.

The LC-MS/MS data were compared using the fragment ion calculator software. After entering the target amino acid sequence in rice α -globulin, the following parameters were set up: mass type and selected MONO; charge state = +1; and ion type = B and Y. The calculated results were compared *via* the two stage mass spectrometry, and the matching results were positive; otherwise, the peptide segment was not detected.

The peptides in α -globulin were prepared using solid-phase synthesis. The peptide was synthesized according to the sequence obtained from China Peptides Co. Ltd. (Shanghai, China), and its purity (>95%) was verified using LC-MS/MS.



2.3. Animals and diets

In this study, four-week-old male hamsters (bodyweight = 100 ± 3 g) were obtained from Vital River Lab Animal Technology Co, Ltd. (Beijing, China) and kept in cages in an air-conditioned room. The peptide and α -globulin were orally administrated to the hamsters at a dose of 5 mg in 1 mL of deionized water and 50 mg in 1 M NaCl, respectively, and deionized water was orally administrated to the control hamsters. The hamsters were fasted for 4 h and then sacrificed by removal of the whole blood from abdominal aorta. Acetonitrile (containing 0.1% formic acid) was added to 500 μ L of plasma and allowed to stand for 30 min at 4 °C. The plasma was centrifuged at $8000 \times g$ at 4 °C for 20 min, and then, supernatant was obtained (with filtration through a 0.45 μ m membrane).

Male hamsters (four-week-old) were obtained from Vital River Lab Animal Technology Co, Ltd. (Beijing, China) and kept in cages in an air-conditioned room (temperature = 21–23 °C; humidity = $55 \pm 5\%$; and 12 hours light/dark cycle). After 3 days of acclimation, the hamsters were divided into 3 groups (normal, control, and GEQQQPGM) of 7 hamsters. The peptide was orally administrated to the hamsters at a dose of 100 mg per kg bodyweight, and deionized water was orally administrated to the normal and control hamsters at 10:00 each day for 30 days. The composition of the experimental diet in the normal group was prepared according to the AIN-93G (American Institute of Nutrition, 1993) standard diet. The composition of the experimental diet in the control and GEQQQPGM groups was prepared according to AIN-93G with some modification. The composition (in g kg^{-1}) of the diet was 200 casein, 397 corn starch, 70 soybean oil, 10 cholesterol, 50 lard, 2 bile salt, 38.486 sucrose, 50 cellulose, 3 DL-methionine, 132 maltodextrin-10, 0.014 *t*-butylhydroquinone, 2.5 choline bitartrate, 35 mineral mix, and 10 vitamin mix. The experimental diet was prepared by Beijing Nuokangyuan Biotechnology Co., Ltd. (Beijing, China). The food intake was noted and monitored every day to ensure the similar content of diet in all the groups. The bodyweight of the animals was obtained every three days. The hamsters were fasted for 16 h and then sacrificed by removal of the whole blood from the abdominal aorta. The plasma lipid concentration was analyzed using an automatic chemistry analyzer (Hitachi, Tokyo, Japan).

All experiments were carried out according to the P. R. China legislation and approved by the Bioethics Committee of the Institute of Medicinal Plant Development, Chinese Academy of Medical Sciences and Peking Union Medical College.

2.4. Cell culture

HUVECs (Lot: 63505670), endothelial cell growth Kit-BBE, and vascular cell basal medium were acquired from the ATCC (Manassas VA, USA). The medium containing 10% FBS, Kit-BBE, and penicillin–streptomycin (1 : 1000) were prepared according to the method of ATCC. The cells were cultured in a humidified atmosphere containing 5% CO_2 at 37 °C. The cells were treated with 0.25% trypsin/1 mM EDTA after approximately 80% confluence. The HUVECs were serum-starved overnight prior to

stimulation. The DMEM with 10% of FBS medium was used for the cell subcultures.

2.5. Analysis of the cell viability

The cell viability was analyzed using the CCK-8 kit. Cells were seeded in 96-well plates (1×10^4 per well, 100 μ L per well) for the CCK-8 assay. The cells were pretreated with the peptide (25, 50, 100, and 200 $\mu\text{g mL}^{-1}$) for 1 h. In the presence of the peptide, the cells were stimulated with or without TNF- α (10 ng mL^{-1}) for 18 h. Then, the CCK-8 solution (10 μ L per well) was added followed by incubation at 37 °C for 2 h. Since the absorbance is proportional to the number of viable cells, the cell viability has been calculated by the absorbance at 450 nm using a microplate reader (Micro Reader 4, Hyperion, USA).

2.6. ELISA analysis

HUVECs were plated at a density of 1×10^5 cells per mL in 6-well plates and pretreated with or without 50 $\mu\text{g mL}^{-1}$ of peptide for 2 h before stimulation with 10 ng mL^{-1} of TNF- α . To determine the content of iNOS, the cells were digested and fragmented by repeating freeze-thaw three times (at –80 °C for 5 min and at 37 °C for 5 min). The GSH-PX and SOD activities in the cells were measured using an ELISA assay kit. The MDA content was measured using a commercially available kit.

2.7. Measurement of the NO production

Nitrite and nitrate as the stable products were converted from NO and used to determine the NO content in the HUVECs. The cells were plated at a density of 1×10^5 cells per mL in 6-well plates and pretreated with or without 50 $\mu\text{g mL}^{-1}$ of peptide for 2 h before stimulation with 10 ng mL^{-1} of TNF- α . The nitrite and nitrate concentrations in the HUVECs were measured using Griess reagent. The cell supernatant was added to 96-wells (50 μ L per well) followed by addition of 50 μ L of Griess reagent I and II. The nitrite and nitrate concentrations were determined using a microplate reader at 540 nm, and the NO content was calculated.

2.8. Western blotting

After pre-incubation with GEQQQPGM (50 $\mu\text{g mL}^{-1}$) for 18 h, the cells (1×10^7 cells per mL) were lysed using RIPA buffer containing trypsin inhibitor, centrifuged at $8000 \times g$ for 20 min, and quantified using the BCA protein assay. The proteins in the cell lysate were resolved on SDS-PAGE and transferred to the membranes. The target proteins were immunoblotted with primary antibodies and then secondary goat anti rabbit IgG HRP (H + L) antibodies conjugated to horseradish peroxidase. The antibody bindings were revealed using an ECL plus Western blotting detection reagent kit (GE Healthcare, Buckinghamshire, UK). The band density was measured by densitometry and quantified using the Image J software. MCP-1 antibody (#2027), ICAM-1 antibody (#4915), VCAM-1 (E1E8X) rabbit mAb (#13662), IKK α antibody (#2682), and I κ B- α antibody (#4812) were used to detect the protein expression.



2.9. Measurement of ROS production

After pre-incubation with GEQQQPGM ($50 \mu\text{g mL}^{-1}$) for 18 h, the HUVECs were incubated in TNF- α (10 ng mL^{-1}) for 2 h followed by incubation with DCFH-DA ($20 \mu\text{M}$) for 30 min. Then, the cells were observed and imaged using a fluorescence microscope (ECLIPSE TE-2000-S, Nikon Co., Japan). The mean fluorescence intensity was measured using the Image J software ($n = 6$).

2.10. Measurement of NF- κ B activation and nuclear translocation

Cells were immunofluorescence stained to detect the main subunit of NF- κ B, p65, such that to confirm whether NF- κ B was activated or not. Confluent cell monolayers in 96-well plates were pretreated with $50 \mu\text{g mL}^{-1}$ peptide for 6 h prior to incubation with 10 ng mL^{-1} TNF- α for 1 h. The cells were fixed, permeabilized, and immunostained for p65 (anti-rabbit Cy3) and nucleus (DAPI). Using the fluorescence microscope, it was observed that NF- κ B exhibited red fluorescence, whereas the nucleus exhibited blue fluorescence.

2.11. Statistical analysis

Data are expressed as the mean with the standard error (SE) and have been analyzed by Tukey–Kramer multiple comparison post hoc test for the animal experiment. In the cell experiment, all data are presented as the mean value \pm standard deviation (SD). The data are expressed as the fold change over the untreated control. One-way analysis of variance with an appropriate post test was used for the determination of statistical significance. Statistical significance was defined as $P < 0.05$. All statistical analysis was performed using the SPSS software.

3. Results

3.1. Identification of the peptide in rice α -globulin after gastrointestinal digestion and absorption

The absorbable part in the simulated digestive products of rice α -globulin was obtained by everting intestinal sacs and purified by gel filtration chromatography and high performance liquid chromatography. After this, the amino acid sequence of the peptides was measured and compared with that of α -globulin to identify the specific sequence of rice α -globulin. As the result, GEQQQPGM was confirmed to be one of the key fragments in rice α -globulin (Fig. 1). Moreover, further serum tests showed that after the α -globulin and peptide were administrated, the peptide GEQQQPGM was able to enter the plasma (ESI 1, $\dagger m/z$ 1002.2).

3.2. Metabolism-related indicators in hamsters

Metabolism-related indicators were measured in the 3 groups (Table 1). The initial bodyweight, bodyweight gain, food intake, liver weight, plasma HDL cholesterol, and triacylglycerol were not significantly different among the 3 groups. The hamsters orally administrated with GEQQQPGM have significantly lower plasma total and LDL cholesterol concentrations than the

control group ($P < 0.05$). Furthermore, the atherogenic index was reduced in the GEQQQPGM group when compared with that in the control group ($P < 0.05$).

3.3. GEQQQPGM increased cell viability in TNF- α -injured HUVECs

To evaluate whether the peptide GEQQQPGM protects endothelial cells from TNF- α -induced injury, the HUVEC viability was measured (Fig. 2). GEQQQPGM ($200 \mu\text{g mL}^{-1}$) had no significant effect on the cell viability in normal HUVECs; however, the viability of the HUVECs was significantly reduced by the TNF- α treatment ($69.8 \pm 6.3\%$) when compared with that in the control (100%) ($P < 0.01$). The reduction of HUVEC viability induced by TNF- α was significantly increased by GEQQQPGM at a dose of $50 \mu\text{g mL}^{-1}$ ($103.7 \pm 7.0\%$) ($P < 0.01$). The pretreatment with GEQQQPGM obviously restored the cell viability in TNF- α -injured HUVECs, and the most effective dose was 50 and $100 \mu\text{g mL}^{-1}$ ($103.7 \pm 7.0\%$ and $105.6 \pm 6.1\%$) ($P < 0.01$).

3.4. Regulation of MCP-1, VCAM-1, and ICAM-1 expression with GEQQQPGM in TNF- α -injured HUVECs

The MCP-1 concentration was measured, and the results are shown in Fig. 3A. The peptide had no significant effect on the MCP-1 concentration in the HUVECs, whereas the TNF- α -injured HUVECs showed a significantly higher MCP-1 concentration than the control and GEQQQPGM groups ($P < 0.01$). The HUVECs treated with both TNF- α and GEQQQPGM showed significantly lower MCP-1 concentrations than the TNF- α -injured HUVECs ($P < 0.01$). As shown in Fig. 3B and C, the protein expression of VCAM-1 and ICAM-1 in the HUVECs were greatly increased by TNF- α treatment, whereas GEQQQPGM suppressed the TNF- α -induced up-regulation significantly ($P < 0.01$). In addition, the peptide had no significant effect on the protein expression of VCAM-1 and ICAM-1 in the TNF- α -injured HUVECs.

3.5. Regulation of the NO and iNOS concentrations with GEQQQPGM in TNF- α -injured HUVECs

The NO and iNOS concentrations are shown in Table 2. The concentrations of NO and iNOS in the TNF- α -injured HUVECs were significantly higher than those in the control and GEQQQPGM groups ($P < 0.01$). The peptide had no significant effect on the NO and iNOS concentrations in the normal HUVECs; however, GEQQQPGM decreased the NO and iNOS concentrations significantly ($P < 0.05$) in the TNF- α -injured HUVECs.

3.6. The effects of GEQQQPGM on the oxidative stress indexes in TNF- α -injured HUVECs

As shown in Table 2, the HUVECs treated with TNF- α showed significantly lower the GSH-Px and SOD activities and higher MDA content when compared with the control and GEQQQPGM groups ($P < 0.01$). GEQQQPGM did not affect the activities of GSH-Px and SOD, and the MDA content in the



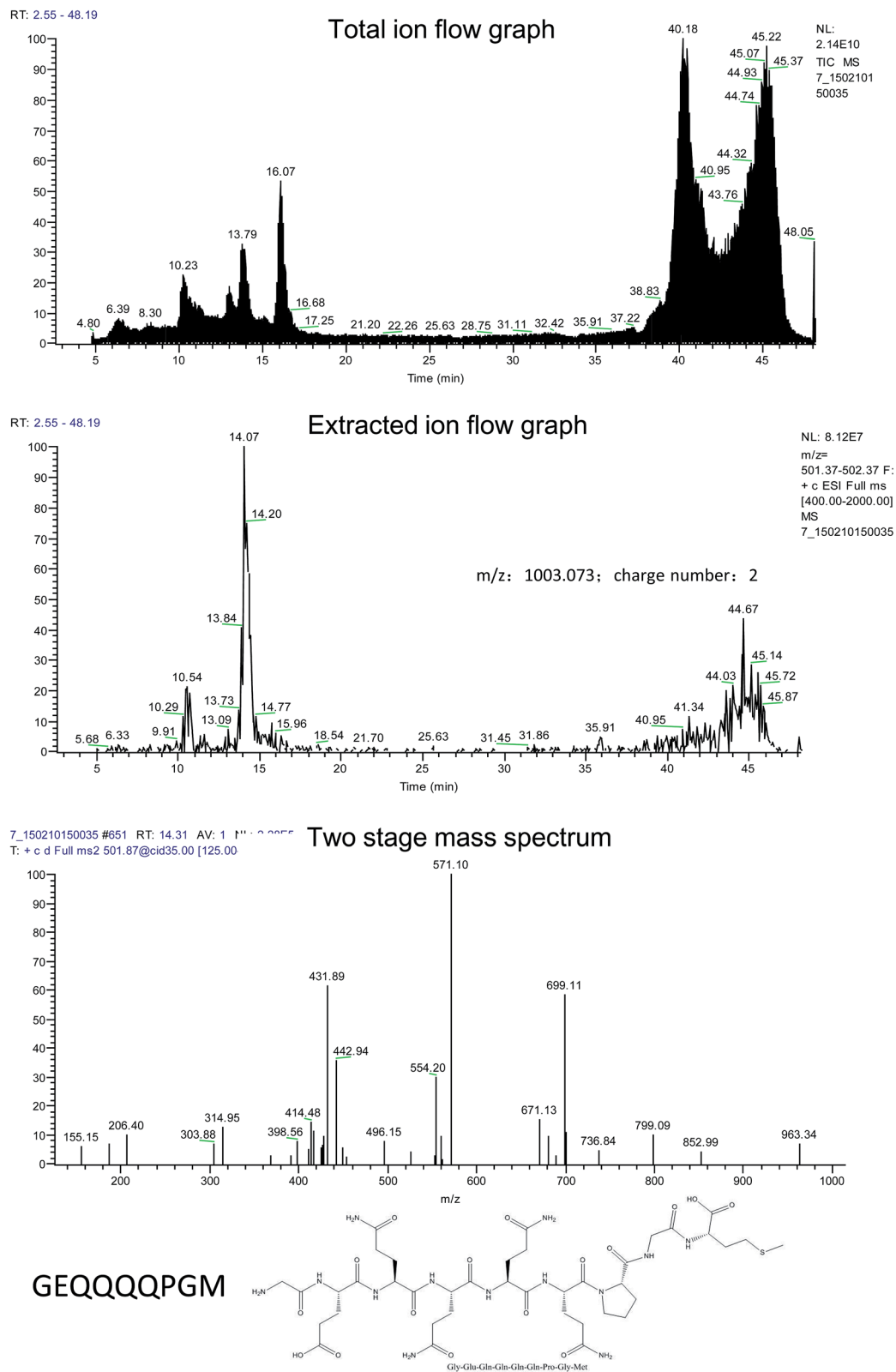


Fig. 1 The amino acid sequence of the peptides was determined by LC-mass spectrometry (MS)/MS.

Table 1 The effects of oral administration of GEQQQQPGM on the metabolic parameters of hamsters fed with a hypercholesterolemic diet

	Normal	Control	GEQQQQPGM
Initial bodyweight (g)	108.7 ± 2.4	108.6 ± 2.2	108.9 ± 2.3
Final bodyweight (g)	121.1 ± 2.8	121.6 ± 2.7	121.7 ± 3.0
Bodyweight gain (g)	12.4 ± 1.4	13.0 ± 1.5	12.8 ± 1.6
Food intake (g day ⁻¹)	7.85 ± 0.22	7.88 ± 0.22	7.86 ± 0.20
Liver weight (g)	6.02 ± 0.34	6.06 ± 0.29	6.05 ± 0.23
Plasma lipids (mmol L ⁻¹)			
Total cholesterol	3.28 ± 0.23 ^c	6.85 ± 0.27 ^a	5.09 ± 0.25 ^b
LDL cholesterol	1.47 ± 0.15 ^c	2.75 ± 0.14 ^a	1.98 ± 0.16 ^b
HDL cholesterol	1.56 ± 0.17	1.62 ± 0.18	1.71 ± 0.14
Triglycerides	2.01 ± 0.53	2.40 ± 0.64	2.47 ± 0.47
Atherogenic index	2.28 ± 0.31 ^c	5.85 ± 0.33 ^a	4.09 ± 0.34 ^b

The mean ± SD were determined from 7 hamsters per group. The different superscript letters a, b and c indicate the significant differences at $P < 0.05$ (Tukey–Kramer's multiple comparison post hoc test).

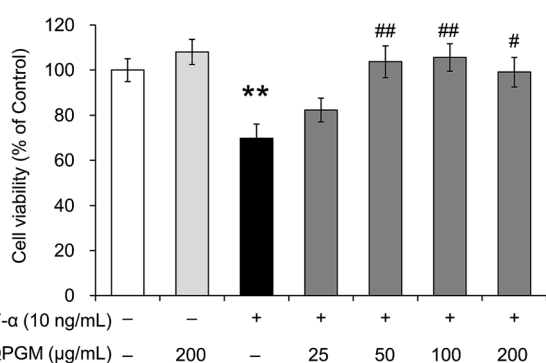


Fig. 2 GEQQQQPGM increased the cell viability in TNF- α -injured HUVECs. Confluent HUVECs were pretreated with different concentrations of GEQQQQPGM for 1 h prior to incubation with 10 ng mL⁻¹ TNF- α for 6 h. Cell apoptosis was analyzed using CCK-8 according to the standard operation instructions. The cell viability levels are expressed as the fold over the control. The bars represent the mean value ± SE, $n = 6$. * indicates $P < 0.05$, compared to the control; # and ## indicate $P < 0.05$ and $P < 0.01$, respectively, compared to TNF- α alone.

normal HUVECs; however, it increased the GSH-Px ($P < 0.01$) and SOD ($P < 0.01$) activities and decreased the MDA content ($P < 0.05$) significantly in the TNF- α -injured HUVECs.

3.7. The effects of GEQQQQPGM on the ROS in TNF- α -injured HUVECs

As shown in Fig. 4, basal ROS production in the HUVECs was reduced by GEQQQQPGM alone ($P < 0.05$) without the TNF- α treatment. The peptide GEQQQQPGM markedly inhibited the increase in TNF- α -induced ROS production ($P < 0.01$).

3.8. GEQQQQPGM inhibited the nuclear translocation of p65 protein in TNF- α -injured HUVECs

The p65 protein in the HUVECs was observed by staining (Fig. 5). The stimulation of TNF- α caused the nuclear translocation of p65 protein. Moreover, pretreatment with GEQQQQPGM significantly inhibited the TNF- α induced-nuclear localization of p65 protein.

3.9. GEQQQQPGM inhibited the IKK-NF- κ B pathway in TNF- α -injured HUVECs

To confirm the effects of GEQQQQPGM on the IKK-NF- κ B pathway, the protein expression of p65, IKK α , and I κ B α in the TNF- α -injured HUVECs was evaluated. As shown in Fig. 6, the protein expressions of p65 and IKK α were greatly increased by the TNF- α treatment, whereas GEQQQQPGM suppressed the TNF- α -induced up-regulation significantly ($P < 0.01$). The protein expression of I κ B α was greatly reduced by the TNF- α treatment, whereas GEQQQQPGM also suppressed the TNF- α -induced down-regulation significantly ($P < 0.05$).

4. Discussion

It has been widely reported that the intake of food proteins can significantly reduce the lesion of atherosclerosis that is closely involved in the pathogenesis of CVDs.^{1,2,9} Moreover, functional peptides, such as IRW, WH, HGI, and HGK, derived from food proteins have been reported to prevent atherosclerosis by improving the function of vascular cells.^{6,13,14} Previous animal experiments have indicated that rice α -globulin (100 mg per day per kg body weight) can efficiently prevent atherosclerosis in ApoE^{-/-} mice; however, the mechanism is not clear.¹⁵ In the present study, GEQQQQPGM was confirmed to be one of the key fragments in rice α -globulin after gastrointestinal digestion and absorption, and the peptide was able to enter the plasma (Fig. 1 and ESI 1[†]). Moreover, we demonstrated that orally administered GEQQQQPGM had significantly lower plasma total and LDL cholesterol concentrations and atherogenic index in hypercholesterolemic hamsters fed with a hypercholesterolemic diet ($P < 0.05$) (Table 1). This result indicated that GEQQQQPGM as one of the functional peptide fragments in rice α -globulin was directly responsible for its anti-atherogenic effect. This was consistent with a previous report on the anti-atherogenic effect of peptides stating that oral administration of the dipeptide WH derived from soybean globulin inhibited progressive atherosclerotic lesions in ApoE^{-/-} mice at 100 mg per day per kg bodyweight.¹²



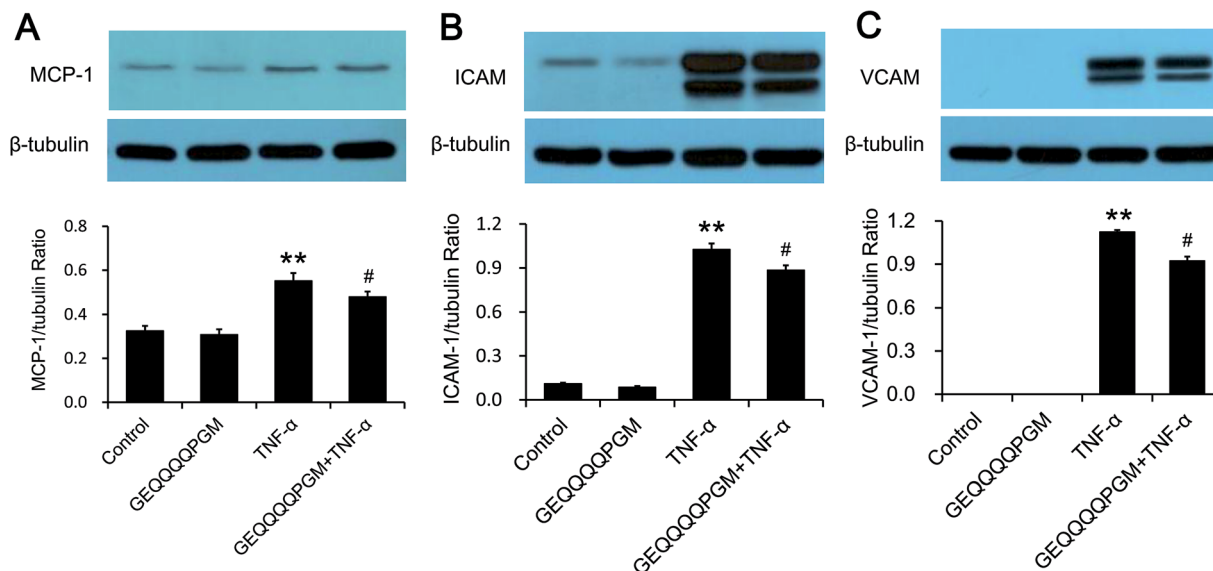


Fig. 3 The regulation of VCAM-1, ICAM-1 and MCP-1 protein expression with GEQQQPGM in TNF- α -injured HUVECs. Confluent HUVECs were pretreated with GEQQQPGM ($25 \mu\text{g mL}^{-1}$) for 18 h prior to incubation with 5 ng mL^{-1} TNF- α for 6 h. The protein expressions of MCP-1 (A), ICAM-1 (B) and VCAM-1 (C) were measured by Western blot analysis. The bars represent the mean value \pm SE, $n = 6$. Representative Western blots are shown. ** indicates $P < 0.01$; # indicates $P < 0.05$, compared to the control.

Table 2 The effects of GEQQQPGM on the NO and iNOS concentrations and oxidative stress indexes in the TNF- α -injured HUVECs^a

	Control	GEQQQPGM	TNF- α	GEQQQPGM + TNF- α
NO ($\mu\text{mol L}^{-1}$)	21.8 ± 2.9	23.7 ± 2.4	$44.7 \pm 3.2^{**}$	$35.0 \pm 3.6^{\#}$
iNOS (pg mL^{-1})	463 ± 27	438 ± 25	$658 \pm 26^{**}$	$572 \pm 18^{\#}$
MDA (mmol L^{-1})	1.20 ± 0.06	$0.68 \pm 0.05^{**}$	$1.71 \pm 0.08^{**}$	$0.85 \pm 0.06^{\#\#}$
GSH-Px (U per mg protein)	83.6 ± 3.1	86.7 ± 3.7	$65.7 \pm 3.4^{**}$	$78.2 \pm 4.9^{\#}$
SOD (U per mg protein)	258 ± 8	260 ± 11	$215 \pm 9^{**}$	$246 \pm 10^{\#\#}$

^a The means \pm SD were determined from 6 separate experiments. * and ** indicate $P < 0.05$ and $P < 0.01$, respectively, compared to the control; # and ## indicate $P < 0.05$ and $P < 0.01$, compared to TNF- α alone.

To clarify the mechanism(s) of the anti-atherogenic effect, in the present study, we determined the effect of the peptide GEQQQPGM digested and absorbed from rice α -globulin in the human body on TNF- α -induced HUVECs injury. As a result, the viability of the TNF- α -injured HUVECs was significantly increased by GEQQQPGM at the doses of 50, 100, and $200 \mu\text{g mL}^{-1}$; this showed that the peptide could play a role in protecting vascular cell injury (Fig. 2). This result suggests that GEQQQPGM, as one of the functional peptide fragments in rice α -globulin, reduces the atherosclerosis risk due to its protective effects on vascular endothelial cell injury. Similarly, the tripeptide IRW derived from egg protein reduced the TNF- α -induced cell injury by reducing the inflammatory response, oxidative stress, and the levels of superoxide ions in HUVECs.⁶

TNF- α stimulation can cause an inflammatory response and oxidative stress in vascular endothelial cell injury; this leads to the up regulation of MCP-1, ICAM-1, and VCAM-1 expression.¹⁶ In the present study, MCP-1, ICAM-1, and VCAM-1 were over expressed in the TNF- α -injured HUVECs, which was consistent with the previous studies.^{6,17} The peptide significantly suppressed the up-regulation of these cell molecules in the

HUVECs treated with TNF- α (Fig. 3). Moreover, the TNF- α induced the excessive synthesis of NO as an indicator of inflammation by the up-regulation of iNOS expression in the HUVECs; this led to endothelial cell injury and atherosclerotic plaque formation.^{18–20} In this study, GEQQQPGM significantly lowered the NO and iNOS concentrations in the TNF- α -injured HUVECs (Table 2). These results showed that GEQQQPGM improved TNF- α -induced vascular endothelial cell injury by inhibiting the inflammatory response.

Endothelial cell injury in atherosclerosis leads to the mitochondrial overproduction of ROS, which are traditionally regarded as the toxic by-products of aerobic metabolism because the stability of ROS, such as superoxide anion, hydrogen peroxide, and hydroxyl radicals, is very important for redox balance.^{21–23} Our results showed that GEQQQPGM pretreatment significantly inhibited ROS production in the TNF- α -injured HUVECs (Fig. 4). In addition, the peptide significantly reduces the MDA content, which is an important index for evaluating the degree of lipid peroxidation (Table 2). It has been widely reported that ROS can be detoxified by GSH-Px and SOD as the anti-oxidative enzymes.⁵ In the present study, we



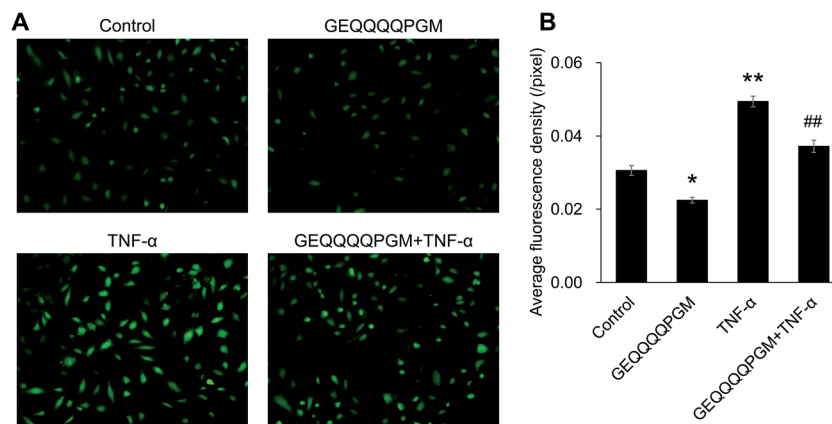


Fig. 4 The effect of GEQQQQPGM on oxidative stress in the TNF- α -injured HUVECs. Confluent HUVECs were pretreated with GEQQQQPGM ($25 \mu\text{g mL}^{-1}$) for 18 h prior to incubation with 10 ng mL^{-1} for 6 h. The fluorescence intensity was measured using Image J software. The bars represent the mean value \pm SE, $n = 6$. * and ** indicate $P < 0.05$ and $P < 0.01$, respectively, compared to the control; ## indicates $P < 0.01$, compared to TNF- α alone. (B) Represents the statistical results of (A).

demonstrated that GEQQQQPGM significantly increased the GSH-Px and SOD activities in TNF- α -injured HUVECs (Table 2). Therefore, the protective effect of GEQQQQPGM could be partially attributed to the amelioration of oxidative stress in TNF- α -induced HUVEC injury. A similar mechanism for improving TNF- α -induced vascular endothelial cell injury has also been observed in the previous studies on the peptide IRW derived from egg protein.⁶

Numerous studies have confirmed that the mechanism of a self-amplifying signal in the inflammatory response is iNOS regulated by NF- κ B in endothelial cells.^{20,24} Moreover, NF- κ B is the main target for ROS, and its activation has been linked to

endothelial dysfunction.^{25,26} Therefore, the IKK-NF- κ B pathway was involved in the pathogenesis of atherosclerosis *via* regulation of the transcription of inflammation and oxidative stress genes. In this pathway, activated IKK α leads to phosphorylation and degradation of I κ B α . Then, the p65 proteins are induced and released from the cytosol and migrate into the cell nucleus, where they interact with the promoter regions of various proteins that are up-regulated during inflammation.²⁷ In the present study, we investigated whether GEQQQQPGM exerted beneficial effects on the IKK-NF- κ B pathway. The results show that GEQQQQPGM can obviously inhibit the TNF- α -induced nuclear localization of p65 protein (Fig. 5). In addition, the

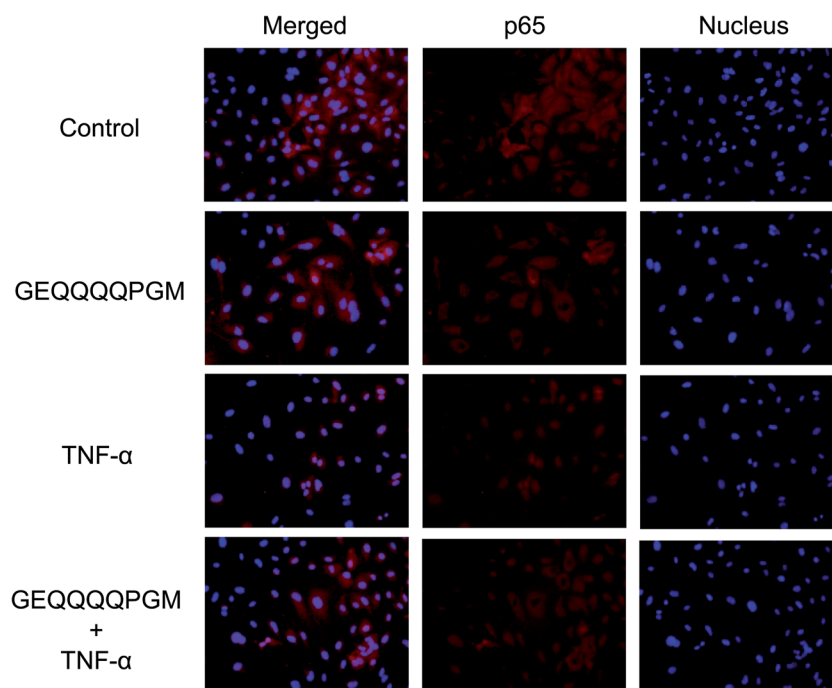


Fig. 5 The effects of GEQQQQPGM on p65 translocation in the TNF- α -injured HUVECs. Confluent HUVECs were pretreated with GEQQQQPGM ($25 \mu\text{g mL}^{-1}$) for 6 h prior to incubation with 10 ng mL^{-1} TNF- α for 1 h. The nucleus and p65 protein were dyed using blue and red fluorescence, respectively ($n = 6$).



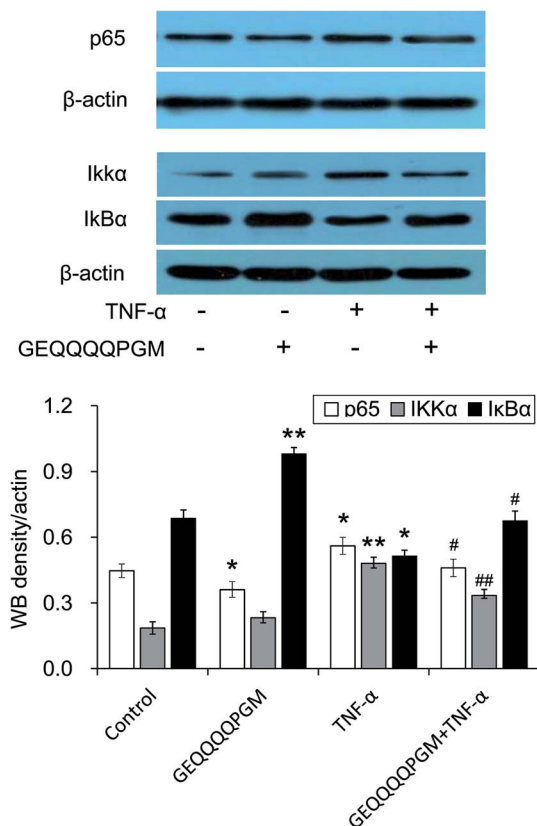


Fig. 6 GEQQQQPGM inhibited the IKK-NF- κ B pathway in TNF- α -injured HUVECs. Confluent HUVECs were pretreated with GEQQQQPGM ($25 \mu\text{g mL}^{-1}$) for 6 h prior to incubation with 10 ng mL^{-1} TNF- α for 1 h. The protein expressions of nuclear p65, IKK, and I κ B α were measured by Western blot analysis. The bars represent the mean value \pm SE, $n = 6$. * and ** indicate $P < 0.05$ and $P < 0.01$, respectively, compared to the control; # and ## indicates $P < 0.01$, compared to TNF- α alone.

peptide exerted a beneficial effect on the protein expression of p65, IKK α , and I κ B α in the TNF- α -injured HUVECs (Fig. 6). This finding showed that GEQQQQPGM reduced the inflammatory response and oxidative stress by inhibiting the inflammatory activation of NF- κ B.

5. Conclusions

In conclusion, the present study suggested that GEQQQQPGM, as one of functional peptide fragments in rice α -globulin, was directly responsible for its anti-atherogenic effect. GEQQQQPGM reduces the atherosclerosis risk due to its protective effects on vascular endothelial cell injury. Moreover, this study clearly shows that GEQQQQPGM at a dose of $50 \mu\text{g mL}^{-1}$ reduced 10 ng mL^{-1} of TNF- α -induced inflammatory response and oxidative stress in HUVECs by inhibiting the inflammatory activation of NF- κ B, thus preventing vascular endothelial cell injury. In addition, our results indicated that the peptide GEQQQQPGM as one of functional peptide fragments was directly responsible for the improvement of rice α -globulin in cardiovascular disease.

Conflicts of interest

There are no conflicts to declare.

Abbreviations

HUVECs	Human umbilical vein endothelial cells
TNF- α	Tumor necrosis factor- α
CCK-8	Cell counting kit-8
ICAM-1	Intercellular adhesion molecule-1
VCAM-1	Vascular cell adhesion molecule-1
MCP-1	Monocyte chemoattractant protein-1
ROS	Reactive oxygen species
NO	Nitric oxide
iNOS	Inducible nitric oxide synthase
SOD	Superoxide dismutase
GSH-PX	Glutathione peroxidase
MDA	Malondialdehyde
GFC	Gel filtration chromatography
HPLC	High performance liquid chromatography
LC-MS	Liquid chromatography-mass spectrometry
NF- κ B	Nuclear factor κ B
IKK	Inhibitor of nuclear factor κ B kinase
I κ B	Inhibitor of NF- κ B
FBS	Fetal calf serum
EDTA	Ethylenediaminetetraacetic acid
PMSF	Phenylmethanesulfonyl fluoride

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