

Cite this: *RSC Adv.*, 2017, 7, 53117

# Sequential solvent fractionation of lignin for selective production of monoaromatics by Ru catalyzed ethanolysis†

Jae-Young Kim,<sup>a</sup> Shin Young Park,<sup>a</sup> Jae Hoon Lee,<sup>a</sup> In-Gyu Choi<sup>ab</sup> and Joon Weon Choi<sup>id</sup>\*<sup>c</sup>

The purpose of this study was to investigate the impact of lignin structure, especially its average molecular weight ( $M_w$ ) on the distribution of catalytically depolymerized products. Soda lignin (SL) was sequentially fractionated by organic solvents (ethyl acetate: F1, methanol: F2, acetone: F3, dioxane/water: F4, and insoluble fraction: F5). The  $M_w$  of SL, F1, F2, F3, and F4 were 2800, 1120, 2860, 5850, and 7200 Da, while that of F5 could not be determined due to its low solubility in GPC solvent. 2D-HSQC-NMR analysis revealed that the relative abundance of the  $\beta$ -O-4 linkage increased upon increasing the  $M_w$  of the lignin fraction. Each fraction was efficiently depolymerized into lignin-oil under the combination of supercritical ethanol (350 °C) and Ru/C (5 wt%). The yield of lignin-oil, mixture of monoaromatic compounds, as well as high molecular phenolics, ranged from 62.5 to 81.4 wt%. The yield of total monoaromatic compounds in lignin-oil varied from ca. 3.4 to 10 wt% depending on the lignin fraction. The selectivity of monoaromatic compounds produced from the lignin depolymerization process was clearly affected by the  $M_w$  value of lignin. Specifically, F1 and F2 yielded higher amounts of non-alkylated phenols, methylated-, and ethylated phenols compared to the other lignin fractions.

Received 19th October 2017  
Accepted 2nd November 2017

DOI: 10.1039/c7ra11541e

rsc.li/rsc-advances

## 1. Introduction

Lignin, the second most abundant polymeric material in nature next to cellulose, consists of three kinds of lignin monomer including *p*-coumaryl, coniferyl, and sinapyl alcohol with various C–O ( $\beta$ -O-4,  $\alpha$ -O-4, and 4-O-5) and C–C bonds ( $\beta$ - $\beta$ , biphenyl, and  $\beta$ -5). Interest in lignin valorization has increased in recent years, because only 2% of the 50 million tons of lignin produced from US pulping industries (2010) was used for high value products, whereas the rest was burned for power generation.<sup>1</sup> This underutilization of lignin evokes the desire to develop methodologies that enable depolymerization of lignin into bio-based chemicals.<sup>2</sup> However, given its structural complexity as a polymer and poor solubility in commonly used solvents, lignin was irregularly degraded into small molecular fragments during the depolymerization process, causing the formation of heterogeneous mixtures of alkylated and

oxygenated phenols.<sup>3–5</sup> For this reason, selective conversion of lignin into specific chemicals and fuels is still challenge in industrial biorefineries.

Due to this need, the development of a catalytic conversion process that can selectively degrade C–O and C–C bonds for the production of a manageably uniform product has been mainly studied. Deuss *et al.* reported an acid-catalyzed depolymerization process that can effectively degrade the  $\beta$ -O-4 linkage in lignin by suppressing undesired side reactions.<sup>6</sup> A solvent-free approach was developed by Kumar *et al.* for the depolymerization of kraft lignin to aromatic monomers using bimetallic sulfided NiMo and CoMo catalysts.<sup>7</sup> Efficient hydrogenolysis of lignin under subcritical water was reported by Onwudili and Williams using Pd/C, resulting in the high conversion of lignin into an oil phase mainly composed of monoaromatic compounds.<sup>8</sup> Supercritical alcohol has been also considered as a promising way for efficient lignin depolymerization.<sup>9–13</sup> Recently, Huang developed one-step valorization of lignin to high yield monoaromatic compounds (23 wt%) without char formation under the combination of supercritical ethanol and CuMgAlO<sub>x</sub> catalyst.<sup>14</sup>

In addition to process approaches, recent developments in this field have highlighted the need to investigate the structural impact of lignin. Ye *et al.* reported that lignin species played an important role in yield of monoaromatic compounds.<sup>15</sup> Also in our prior study, three kinds of technical lignins were depolymerized into alkylated phenols under supercritical *t*-butanol condition, which revealed that the yield, as well as type of

<sup>a</sup>Department of Forest Sciences, Research Institute for Agriculture and Life Science, Seoul National University, 599 Gwanak-ro, Gwanak-gu, Seoul, 151-921, Republic of Korea

<sup>b</sup>Research Institute of Agriculture and Life Sciences, Seoul National University, Seoul 151-742, Republic of Korea

<sup>c</sup>Graduate School of International Agricultural Technology, Institute of Green-Bio Science and Technology, Seoul National University, Pyeongchang 232-916, Republic of Korea. E-mail: [cjw@snu.ac.kr](mailto:cjw@snu.ac.kr); Fax: +82-33-339-5689; Tel: +82-33-339-5840

† Electronic supplementary information (ESI) available. See DOI: 10.1039/c7ra11541e



monoaromatic compounds, was clearly affected by the lignin source.<sup>11</sup> More recently, Bouxin *et al.*, performed catalytic conversion of four lignins with different contents of  $\beta$ -O-4 linkages at 300 °C with methanol/water and Pt/Al<sub>2</sub>O<sub>3</sub> catalyst.<sup>16</sup> They described that the proportion of  $\beta$ -O-4 linkages is the critical factor for both the yield and type of the monoaromatic compounds produced. Although some attempts to investigate structural impact of lignin on depolymerization process have been explored, there have been few reports discussing the effect of lignin molecular weight ( $M_w$ ) on this process. The  $M_w$  of lignin is one of the crucial factors determining the quality of valorized products (*e.g.* antioxidant, plastic, and carbon fiber).<sup>17–20</sup> There exists only one prior study by Toledano *et al.* who investigated the effect of lignin  $M_w$  on the distribution of catalytically depolymerized products.<sup>21</sup> They used ultrafiltration to separate lignin fractions with different  $M_w$  and each fraction was depolymerized by a hydrogen-free, mild, hydrogenolytic, heterogeneously catalyzed methodology assisted by microwave.

The objective of this study was to understand the yield and distribution of monoaromatic compounds in relation to lignin structure, especially the  $M_w$  of lignin. In the current study, lignin was sequentially fractionated by four kinds of organic solvents to obtain five lignin fractions with different molecular distributions. Separated fractions were structurally characterized by several analyses, and these were efficiently depolymerized into low molecular lignin-oil under the combination of supercritical ethanol (350 °C) and Ru/C catalyst. Lignin-oil from each fraction subjected to several analyses to characterize its physicochemical properties. Based on these results, the impact of lignin  $M_w$  on its suitability for conversion to fine chemicals was evaluated.

## 2. Materials and methods

### 2.1 Sequential solvent fractionation

Soda lignin (SL) produced from a mixture of wheat straw and sarkanda grass *via* the soda pulping process was purchased from Granit Research and Development SA. This lignin was sequentially fractionated by ethyl acetate, methanol, acetone, and dioxane/water (95 vol%), as modified from the method outlined in previous literature.<sup>22</sup> 10 g of lignin was dissolved in 100 ml of the first solvent (ethyl acetate), and then this mixture was agitated at room temperature for 2 h. The undissolved solid fraction was filtered using a funnel glass filter (2G4) under vacuum. The filtrate was subsequently evaporated under reduced pressure to remove most of the solvent, followed by suspension in water, and then freezing-drying to recover the lignin fraction (F1, ethyl acetate soluble). The undissolved lignin fractions were further extracted with methanol, acetone, and dioxane/water (95 vol%), and the recovered lignin fractions were noted as F2, F3, F4, and F5 (final insoluble fraction), respectively.

### 2.2 Lignin characterization

The yield of each lignin fraction was determined gravimetrically. The weight percentages of carbon, hydrogen, and oxygen

of the lignin fractions were determined using an elemental analyzer CHNS-932 from LECO Corp. The content of methoxyl (OMe) moieties was measured using a gas chromatographic method.<sup>23</sup> To quantify the hydroxyl group (OH) content in the lignin fraction, <sup>31</sup>P-NMR spectroscopy was performed.<sup>24</sup> During sample preparation, 25 mg of each lignin fraction and its derived lignin-oil was phosphorylated by 70  $\mu$ l of 2-chloro-4,4,5,5-tetramethyl-1,3,2-dioxaphospholane (TMDP). Cyclohexanol was used as an internal standard. This mixture was analyzed by a Bruker AVANCE 600 MHz instrument with an inverse-gated decoupling pulse sequence using a 45 degree pulse angle and 5.0 s relaxation delay. The resulting <sup>31</sup>P-NMR spectra were integrated using the Bruker TopSpin 3.5 software.

2D-HSQC-NMR were acquired with a Bruker AVANCE 600 spectrometer (Bruker, Germany) using 20 mg of lignin fraction dissolved in 0.75 ml DMSO-D<sub>6</sub> employing a standard Bruker pulse sequence “hsqcetgpsisp2.2” with a 90° pulse, 0.08 s acquisition time, 2.0 s pulse delay, 1JC-H of 150 Hz, 48 scans, and an acquisition of 1024 data points (for 1H) over 512 increments (for 13C). The 1H and 13C pulse widths observed were  $p_1 = 11.43 \mu$ s and  $p_2 = 10.00 \mu$ s, respectively. HSQC data processing and plots were carried out using the MestReNova v10.0 software's default processing template with automatic phase and baseline correction. Two regions of interest were considered: (1) aliphatic oxygenated side chain region ( $\delta_C/\delta_H$  50–90/2.5–5.9) and (2) aromatic/unsaturated region ( $\delta_C/\delta_H$  100–140/5.5–8.0). The number of inter-unit linkages was quantified by using the signal intensity from selected syringyl and guaiacyl peaks.<sup>25</sup> We choose C<sub>2,6</sub>-H<sub>2,6</sub> in H units ( $\delta_C/\delta_H$  127.9/7.19), C<sub>2</sub>-H<sub>2</sub> in G units (110.9/6.98), and C<sub>2,6</sub>-H<sub>2,6</sub> in S units (103.8/6.71) signals as an internal standard. The amount of linkages, expressed as a number per 100 aromatic units, was calculated on the basis of correspond to each signal ( $\beta$ -O-4:  $\delta_C/\delta_H$  71.9/4.9,  $\beta$ -5:  $\delta_C/\delta_H$  87.7/5.5, and  $\beta$ - $\beta$ :  $\delta_C/\delta_H$  85.5/4.6).

The chemical compositions of lignin fractions were estimated according to the National Renewable Energy Laboratory (NREL) standard biomass analytical protocol.<sup>26</sup>

### 2.3 Lignin depolymerization in supercritical ethanol state

Each fractionated lignin was introduced into catalytic depolymerization process under supercritical ethanol according to the procedure of our earlier study.<sup>9</sup> 5 wt% ruthenium catalyst supported on activated carbon (Ru/C) catalyst was purchased from Sigma-Aldrich and used without further treatment. Briefly, 0.50 g of lignin sample and 20 ml of EtOH (ACS grade,  $\geq 99.5\%$ , Sigma-Aldrich) were loaded into the reactor with 0.025 g of Ru/C catalyst. Before reaction, sufficient nitrogen gas (99.9%) was flowed, and then 3 MPa of hydrogen gas (99.9%) was pressurized. The reactor was heated to 350 °C with an average heating rate of 13.2 °C min<sup>-1</sup> and maintained at this temperature for 40 min. The maximum pressure in the reactor was approximately 17 MPa. After reaction, the reactor was rapidly quenched by ice-water with an average cooling rate of -32.2 °C min<sup>-1</sup>. Three lignin depolymerized products (lignin-oil, char, and gas) were recovered and the yield of them was calculated according to following equation:



$$\text{Lignin-oil yield (wt\%)} = \frac{\text{weight of oil fraction (g)}}{\text{weight of lignin (g)}} \times 100$$

$$\begin{aligned} \text{Char yield (wt\%)} \\ &= \frac{(\text{weight of solid fraction (g)}) - (\text{weight of catalyst (g)})}{\text{weight of lignin (g)}} \\ &\times 100 \end{aligned}$$

$$\text{Gas yield (wt\%)} = 100 - (\text{lignin oil yield} + \text{char yield})$$

## 2.4 Chemical properties of lignin-oil

**2.4.1 GC-MS/FID (gas chromatography-mass spectroscopy/flame ionization detector).** Monoaromatic compounds in lignin-oil was qualified and quantified by GC-MS/FID analysis using an Agilent HP7890B GC equipped with an Agilent HP5975A mass selective detector (MSD). A dilute lignin-oil of 1.0  $\mu\text{l}$  (in 5 ml acetone) with internal standard (IS, fluoranthene 50.5 mg/5 ml in acetone) was injected into the DB-5 capillary column (30 m  $\times$  0.25 mm ID  $\times$  0.25  $\mu\text{m}$  film thicknesses) with a split ratio of 1 : 15. In this system, the separated products were split equally (1 : 1) to (1) FID and (2) MS system. The initial oven temperature was set to 50  $^{\circ}\text{C}$  for 5 min, and then increased to 300  $^{\circ}\text{C}$  with a rate of 3  $^{\circ}\text{C min}^{-1}$ . The final temperature was held for ten min. The injector and detector temperatures were 220  $^{\circ}\text{C}$  and 300  $^{\circ}\text{C}$ , respectively, and He (99.9% purity) was used as the carrier gas. The mass spectra of each compound were identified using NIST MS Search 2.0 (NIST/EPA/NIH Mass Spectral Library; NIST 02). 16 standard compounds were injected into the GC-FID to investigate a response factor (RF) between each standard compound and the internal standard.<sup>9</sup> The yield of monoaromatic compounds was calculated by following equation:

$$\begin{aligned} \text{The yield of monoaromatic compounds (mg/g of lignin)} \\ &= \text{RF} \times \frac{\text{compound peak area (FID)}}{\text{IS peak area (FID)}} \times \frac{\text{IS amount } (\mu\text{g})}{\text{lignin (mg)}} \end{aligned}$$

**2.4.2 Gel permeation chromatography (GPC).** Each fractionated lignin was acetylated with acetic anhydride/pyridine (1 : 1 v/v) at 105  $^{\circ}\text{C}$  for 2 h before GPC analysis. Approximately 2 mg of acetylated lignin and lignin-oil dissolved in tetrahydrofuran (1 mg  $\text{ml}^{-1}$ ) was introduced into a GPC max instrument (Viscotek RImax, Viscotek, UK) equipped with PLgel 3  $\mu\text{m}$  MIXED-D columns (300  $\times$  7.5 mm, VARIAN, Inc.), a PLgel 3  $\mu\text{m}$  MIXED-E column (300  $\times$  7.5 mm, VARIAN, Inc.), and a PLgel 5  $\mu\text{m}$  guard column (50  $\times$  7.5 mm, VARIAN, Inc.), while the effluent (0.7  $\text{ml min}^{-1}$ ) was detected by UV-Vis detection (VE3210, Viscotek). 12 types of polystyrenes with different molecular weights ranging from 266 (styrene) to 66 000 Da were used to calculate the average molecular weight ( $M_w$ ) and the number average molecular weight ( $M_n$ ) of analytes in the effluent. The depolymerization degree (%) of lignin-oil was calculated using the following equation:

$$\begin{aligned} \text{Depolymerization degree (\%)} \\ &= \left( 1 - \frac{\text{The } M_w \text{ of lignin-oil}}{\text{The } M_w \text{ of lignin fraction}} \right) \times 100 \end{aligned}$$

**2.4.3 Elemental analysis.** Lignin-oil was injected into an elemental analyzer CHNS-932 (LECO Corp) to measure the weight percentages of carbon, hydrogen, and nitrogen. The oxygen content was calculated by subtraction.

## 3. Results and discussion

### 3.1 Chemical and structural characteristics of lignin fractions

SL was sequentially fractionated with four kinds of organic solvent with different solvation capacity. This approach is based on the solvation power of solvent, which is varied by both the cohesive energy and the hydrogen-bonding capacity of the solvent molecule.<sup>27</sup> The yields of the five fractions were 22.0, 52.5, 4.6, 14.9, and 6.1 wt%, respectively (Table 1). The  $M_w$  as well as  $M_n$  of fractionated lignin was investigated by GPC, and the resulted curves are described in Fig. 1. SL had a multimodal molecular distribution with an overall  $M_w$  of 2800 Da and PDI of 2.8. After fractionation, each fraction had a relatively narrow PDI compared to SL. The  $M_w$  value and PDI of the lignin fraction increased from 1120 (F1) to 7200 Da (F4) and 1.8 (F1) to 2.3 (F4), respectively, as sequential extraction progressed, which was in agreement with previous works.<sup>22,28–30</sup> Meanwhile, F5 was not totally dissolved in THF due to the presence of carbohydrate (30 wt%) and ash content (7.5 wt%). This is similar to the results from Wang *et al.* who reported that the purity of lignin determined by klason method decreased as fractionation step was progressed.<sup>29</sup>

Elemental composition and functional group content (OMe and OH) of the lignin fraction were quantitatively determined and shown in Table 1. As confirmed in Table 1, the carbon content of F5 was relatively lower than that of others (59.0 to 64.4 wt%) because F5 contained high carbohydrate (30 wt%) and ash content (7.5 wt%). With respect to functional groups in the lignin fraction, the content of phenolic OH including H, G, and S showed a decreasing trend with increasing  $M_w$  of the lignin fraction. However, the OMe content of the lignin fraction did not show a notable trend as a function of the  $M_w$  value. It is well known that a higher phenolic OH content in lignin resulted from more serious degradation of aryl-ether bond cleavage during the soda pulping process. Meanwhile, the content of aliphatic OH was the highest in F2, followed by F3, F4, and F1. The reason for this trend was unclear, but it might be related to the polarity index of solvents used in the extraction step (ethyl acetate: 4.4, methanol: 5.1, acetone: 5.1, and dioxane: 4.8).<sup>31</sup>

### 3.2 2D-HSQC-NMR analysis of lignin fractions

Further insight into the structural difference between lignin fractions was investigated by 2D-HSQC-NMR. Fig. 2 and 3 show the HSQC spectra of lignin fractions with the substructures corresponding to the assigned  $^{13}\text{C}$ - $^1\text{H}$  correlations.<sup>32,33</sup>



Table 1 Chemical and structural characteristics of lignin fractions

Lignin	Yield (wt%)	Elemental composition (wt%)					OMe (mmol g <sup>-1</sup> )	OH <sup>b</sup> (mmol g <sup>-1</sup> )			
		C	H	N	O <sup>a</sup>	Aliphatic		<i>p</i> -Hydroxyl (H)	Guaiacyl (G)	Syringyl (S)	
SL	—	62.7	5.8	0.4	31.1	3.77	1.13	0.28	0.75	0.74	
F1	22.0 ± 1.3	64.4	6.5	—	29.1	3.19	0.61	0.52	1.23	1.15	
F2	52.5 ± 1.6	63.0	5.8	0.5	30.7	3.55	1.27	0.40	0.98	0.88	
F3	4.6 ± 0.6	64.6	6.2	0.3	29.0	2.35	0.96	0.20	0.47	0.51	
F4	14.9 ± 1.8	59.0	5.6	1.0	34.4	2.58	0.81	0.12	0.32	0.39	
F5	6.1 ± 0.5	52.0	6.5	0.5	41.1	1.52	—	—	—	—	

<sup>a</sup> Calculated by difference. <sup>b</sup> Determined by <sup>31</sup>P-NMR analysis.

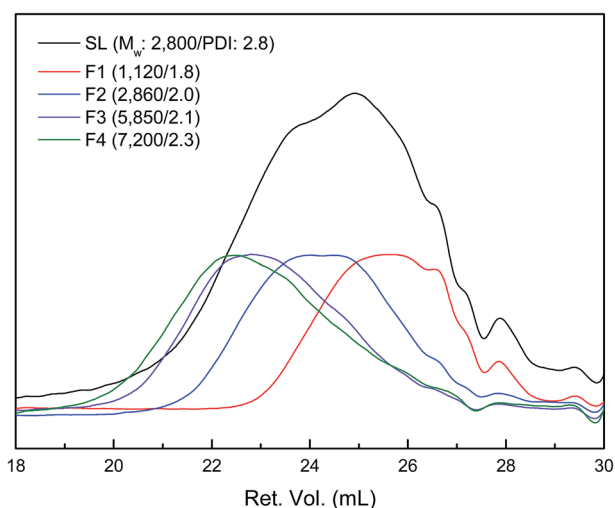


Fig. 1 GPC curves of lignin fractions (the peak maxima were normalized to same value except for SL).

Recorded spectra in this study displayed representative interunit linkages such as the  $\beta$ -aryl-ether (A:  $\beta$ -O-4), phenylcoumaran (B:  $\beta$ -5), and resinol (C:  $\beta$ - $\beta$ ) structure in lignin fractions. In addition, not only H, G, and S units, but also ferulates and *p*-coumarates were detected because SL was extracted from herbaceous crops (wheat straw and sarkanda grass). As shown in Fig. 2, the correlation of the  $\beta$ -O-4 structure (A) in the spectra of F1 is much less intense than other samples, which indicates F1 had the highest condensed structure. Interestingly, the relatively high xylan content in F5 was confirmed at  $\delta_C/\delta_H$  3.1/74.1, 3.3/75.9, and 3.5/77.2 ppm (black spot). This result is consistent with the result of above section.

The amount of interunit linkages in lignin fractions was quantified by integration of HSQC spectra with an aromatic unit as an internal standard; the result is given in Table 2.<sup>25</sup> The relative abundance of  $\beta$ -O-4,  $\beta$ -5, and  $\beta$ - $\beta$  of SL (per 100 aromatic units) was 6.1, 0.8, and 1.5, which were reasonable levels as compared with previous studies.<sup>32,33</sup> 2D-HSQC-NMR analysis was also used in previous work by Li *et al.* who reported that F2 (methanol soluble fraction) had the highest proportion of  $\beta$ -O-4 linkages. However, in this study, the amount of  $\beta$ -O-4 linkage gradually increased from 0 to 18.9 upon increasing the  $M_w$  of

the lignin fraction as shown in Table 2, except for F5. This result correlated with those observations of phenolic OH content, as discussed above.

### 3.3 Yield of lignin depolymerized products

Each lignin fraction was depolymerized into low molecular lignin-oil under the combination of supercritical ethanol (350 °C) and 5 wt% Ru/C catalyst for 40 min. Before this reaction, 3 MPa hydrogen gas was pressurized into the reactor to stabilize or quench radicals, and unstable intermediates formed during the lignin depolymerization process.<sup>34</sup> The yield of lignin-oil, mixture of monoaromatic compounds, as well as di-, tri- or higher molecular phenolics, ranged from 62.5 to 81.4 wt% (Fig. 4). Char and gas, mainly composed of H<sub>2</sub>, CH<sub>4</sub>, CO, and CO<sub>2</sub>, were produced as byproducts resulting from various side reactions, including carbonization, recondensation, and further decomposition of low molecular weight fragments in lignin-oil.<sup>9,11</sup> Our experiment revealed that the distribution of depolymerized lignin products varied depending on the lignin fraction. In this study, the maximum lignin-oil yield (81.4 wt%) was achieved in F1 with the smallest amount of char (1.6 wt%). F3 gave rise to the second highest lignin-oil yield in spite of its relative high  $M_w$ . This meant that not only  $M_w$  but also other structural characteristics of lignin have influence on the distribution of depolymerized lignin products. Meanwhile, according to a previous study, char formation during lignin depolymerization has a negative effect on catalyst activity, because char deposits on catalyst surfaces give rise to catalyst deactivation.<sup>35</sup> These results suggested that a lignin fraction having low  $M_w$  was advantageous for catalytic liquefaction processes.

### 3.4 Chemical properties of lignin-oils

**3.4.1 Yield of monoaromatic compounds in lignin-oils.** GC/MS analysis of the lignin-oil was performed to identify and quantify the major monoaromatic compounds derived from each lignin fraction, and detailed information of the GC/MS results is given in Table S1.† To better understand the effect of lignin fraction  $M_w$  on the distribution of monoaromatic compounds, GC/MS detectable monoaromatic compounds were classified into one of five categories (C6P: phenols without side chain, C1C6P: methylated phenols, C2C6P: ethylated phenols,



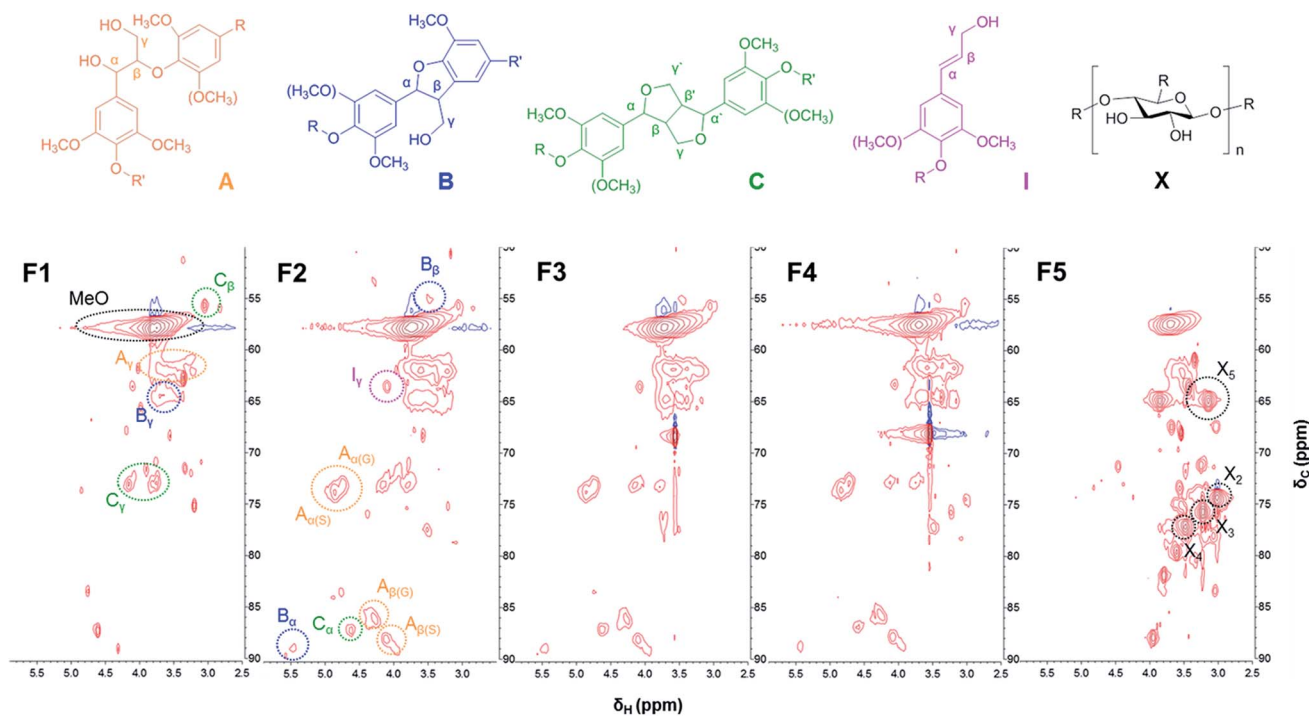


Fig. 2 Side chain ( $\delta_C/\delta_H$  50–90/2.5–5.9) regions in the 2D-HSQC-NMR spectra of lignin fractions.

C3C6P: propylated phenols, and OP: phenols with oxygenated functional groups) based on their structures (Fig. 5).

The yields of total and classified monoaromatic compounds in lignin-oil are presented in Fig. 6. As shown in Fig. 6(f), the yield of total monoaromatic compounds ( $\text{mg g}^{-1}$  of lignin) decreased from 100.3 (F1) to 69.6 (F4) upon increasing the  $M_w$  of

the lignin fraction. These values were relatively higher than the results from Toledano *et al.* who reported that 2.00 to 6.58 ( $\text{mg g}^{-1}$  of lignin) of monoaromatic compounds could be obtained from different  $M_w$  of lignin fractions *via* nickel catalyzed hydrogen-free microwave irradiation at 150 °C.<sup>21</sup> Meanwhile, F5 showed the lowest yield of total monoaromatic compounds, as well as lignin-

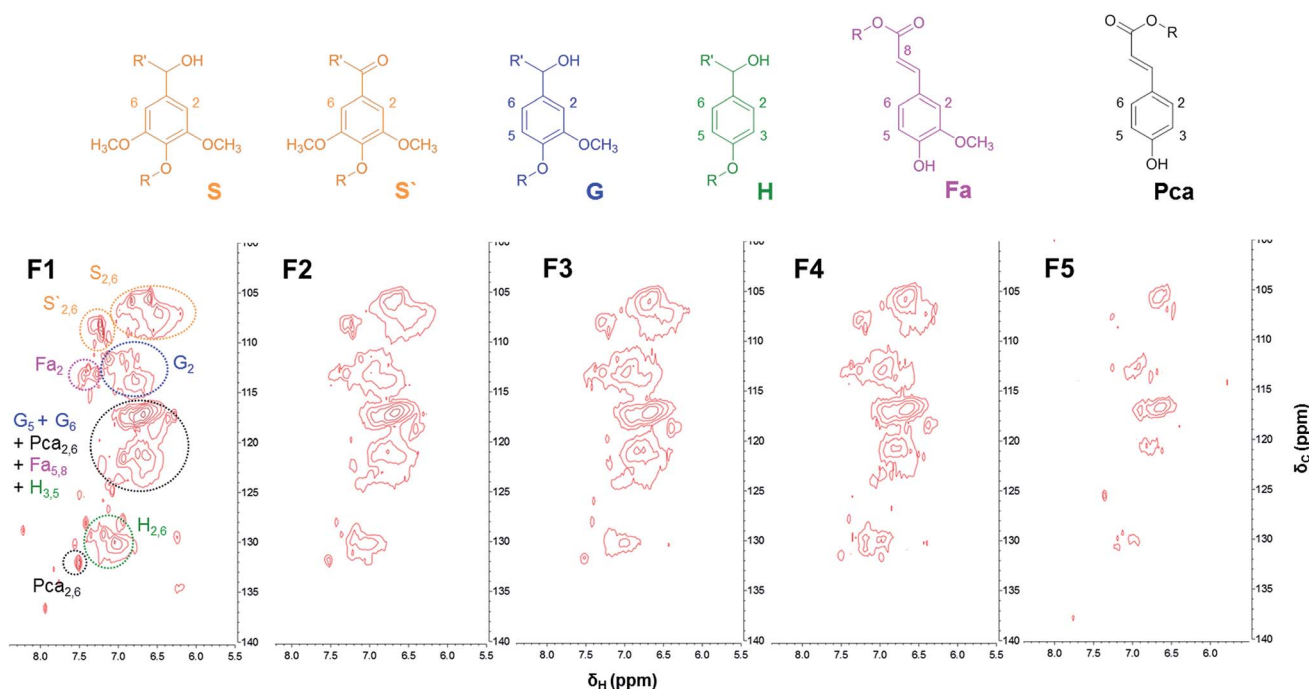


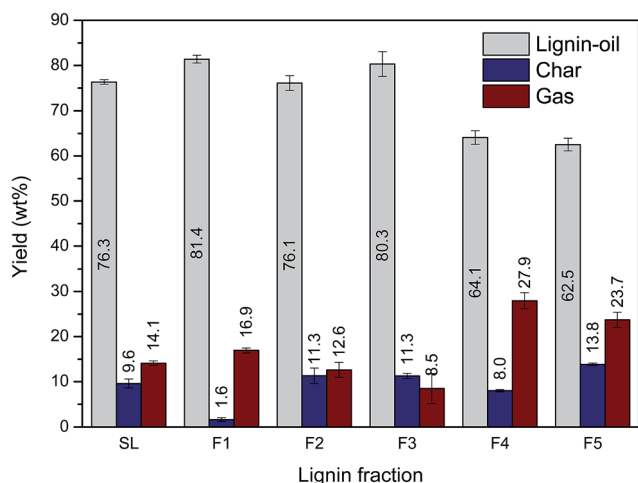
Fig. 3 Aromatic/unsaturated ( $\delta_C/\delta_H$  100–140/5.5–8.0) regions in the 2D-HSQC-NMR spectra of lignin fractions.



**Table 2** Relative amounts of main interunit linkages in lignin fractions (a number per 100 aromatic units)

Lignin	$\beta$ -O-4	$\beta$ - $\beta$	$\beta$ -5
SL <sup>a</sup>	6	1.4	0.5
SL <sup>b</sup>	3.4	0.7	—
SL	6.1	1.5	0.8
F1	—	1.5	—
F2	7.9	0.8	1.6
F3	16.5	3.7	0.4
F4	18.9	1.5	0.3
F5	5.8	—	—

<sup>a</sup> Data from previous study.<sup>32</sup> <sup>b</sup> Data from previous study.<sup>33</sup>



**Fig. 4** Yield of lignin depolymerized products as a function of lignin molecular weight.

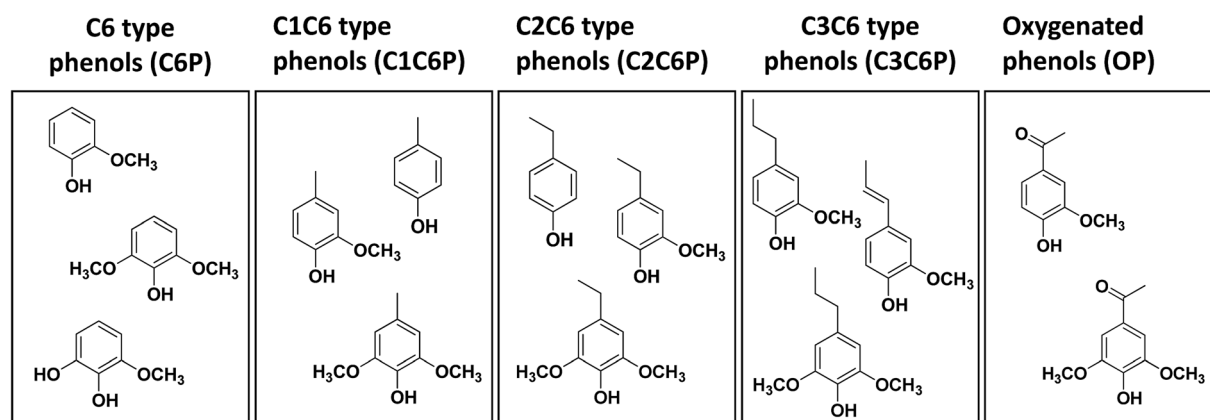
oil, because it contained some contaminant as mentioned above. Several compounds derived from carbohydrate were observed in F5 lignin-oil, which is well in line with high carbohydrate content typically observed in F5 (Fig. S1†).

Fig. 6(a) and (b) display the yield of C6P and C1C6P compounds as a function of the  $M_w$  value of the lignin fraction.

Guaiacol, methylated guaiacol, 3-methoxycatechol, syringol, and methylated syringol, mainly used as a precursor to chemical feedstocks and various flavorants were produced as main C6P and C1C6P compounds.<sup>36</sup> The sum of C6P and C1C6P showed the highest yields in F2 (19.8 and 16.8 mg g<sup>-1</sup> of lignin, respectively) followed by F3. On the other hand, the yield of C2C6P showed a different tendency (Fig. 6(c)). These C2C6P compounds, such as 4-ethylphenol, 4-ethylguaiacol, and 4-ethylsyringol, were potential chemical intermediates for the production of phenol, flavors, resins, polymers, medicines, and so on.<sup>37</sup> Many prior studies reported that such compounds were mainly obtained from the thermochemical conversion process of native or technical lignin (e.g. hydrogenolysis, solvolysis, and pyrolysis).<sup>11,15,33,38–40</sup> C2C6P compounds were possibly transformed from 4-vinylphenols, primary thermal degradation products from lignin, by selective hydrogenolysis of vinyl function.<sup>15,34,41</sup> As confirmed in Fig. 6(c), the sum of C2C6P was the highest at F1 (44.2 mg g<sup>-1</sup> of lignin) and decreased with increment of lignin fraction  $M_w$ . It was previously reported that non-alkylated phenols, methylated-, and ethylated phenols can result from the condensed fraction of lignin.<sup>16</sup> The results of the current study are partially consistent with previous results. F1 and F2 with lower amounts of  $\beta$ -O-4 linkages yielded these compounds (C6P, C1C6P, and C2C6P) more than other lignin fractions.

The yield of C3C6P compounds is illustrated in Fig. 6(d). It has been previously revealed that C3C6P compounds from lignin such as 4-propylguaiacol, 4-propylsyringol, and *trans*-isoeugenol obtained was directly related to the amount of  $\beta$ -O-4 linkages in the starting lignin material.<sup>16,42</sup> As mentioned above (2D-HSQC-NMR), the relative amount of  $\beta$ -O-4 linkages was the highest at F4 (18.9) followed by F3 (16.5) and F2 (7.9). However, the sum of C3C6P did not fully match as a function of the amount of  $\beta$ -O-4 linkages. This suggested that the yield of C3C6P compound from lignin depolymerization process was affected not only by the amount of  $\beta$ -O-4 linkages, but also other structural characteristics of lignin such as functional group contents and the  $M_w$ .

Meanwhile, as observed in Fig. 6(e), acetosyringone was the major product among OP compounds, and others, including



**Fig. 5** Classification monoaromatic compounds on the basis of their structures.



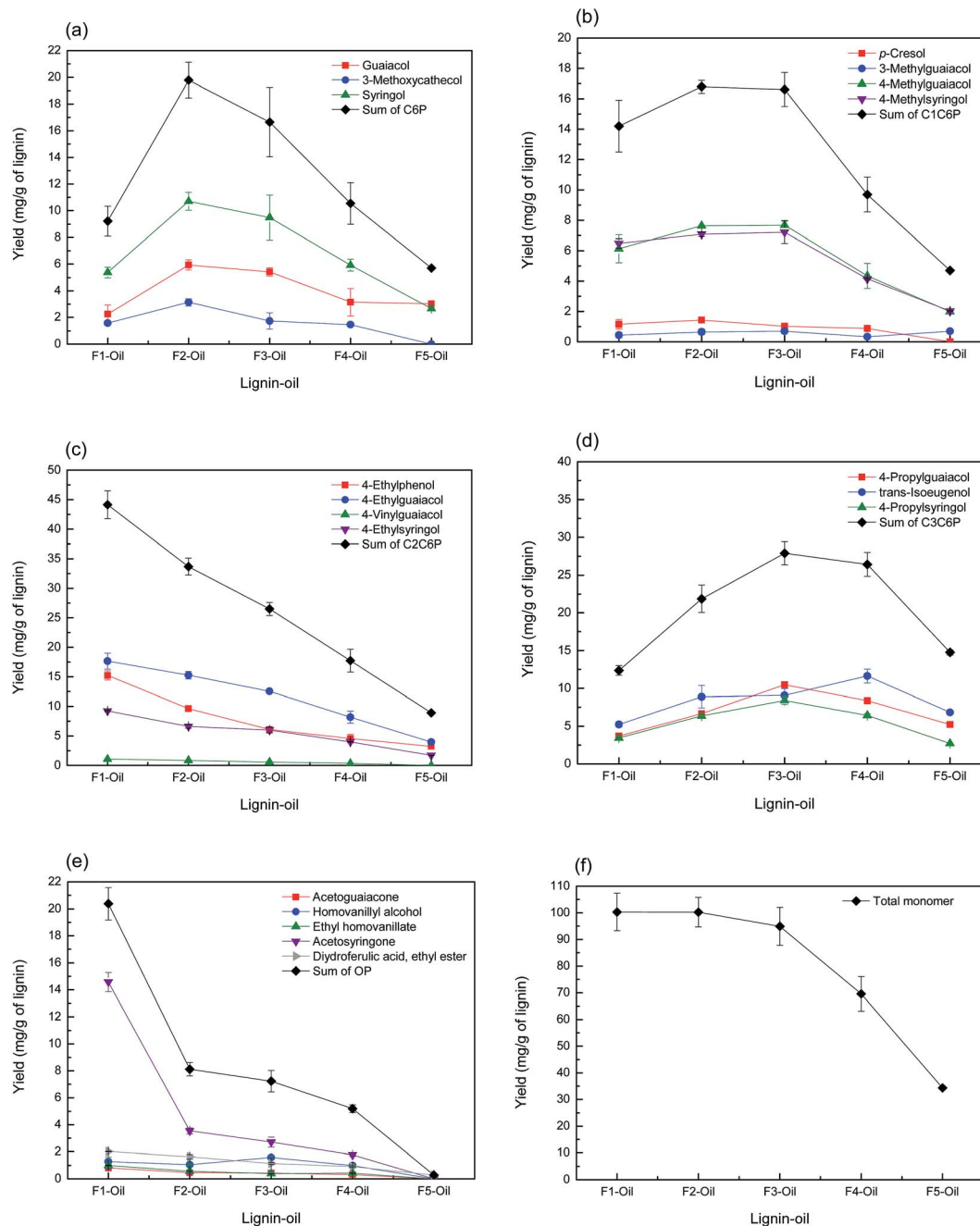


Fig. 6 Yield of monoaromatic compounds ((a) C6P, (b) C1C6P, (c) C2C6P, (d) C3C6P, (e) OP and (f) total monomer) in lignin-oils as a function of lignin molecular weight.

acetoguaiacone, homovanillyl alcohol, and ethyl homovanillate, showed a low yield ( $<2 \text{ mg g}^{-1}$  of lignin). Acetosyringone is specialized for the production of medicines such as non-steroidal anti-inflammatory drugs, anti-asthmatic, and non-narcotic analgesic.<sup>43</sup> The yield of this compound was the highest at F1 ( $14.6 \text{ mg g}^{-1}$  of lignin) and gradually decreased with increasing the  $M_w$  of lignin fraction. Sum of OP had a similar pattern of acetosyringone. This trend suggested that the lignin fraction with lower  $M_w$  value contained more OP-type monoaromatic compounds than higher  $M_w$  lignin fraction.

Our previous studies revealed that whole lignin (SL) had a low selectivity of each compound in lignin-oil due to its

structural complexity,<sup>9,11</sup> but the result of the current study suggested that the selectivity of monoaromatic compounds could be partly controlled by sequential fractionation of lignin before lignin depolymerization process.

**3.4.2 The average molecular weight of lignin-oils.** To investigate molecular distribution of lignin-oil from each lignin fraction, the  $M_w$  value of lignin-oil was also determined by GPC, and this result is provided in Fig. 7 and Table 3. As exhibited in Fig. 7, the GPC curve of lignin-oil had bimodal peaks. The  $M_w$  of the peak eluting at 28 ml was 150 to 160 Da, whereas the  $M_w$  of the peak eluting before 28 ml ranged from 690 to 1120 Da depending on lignin-fraction (Table 3). This result showed that



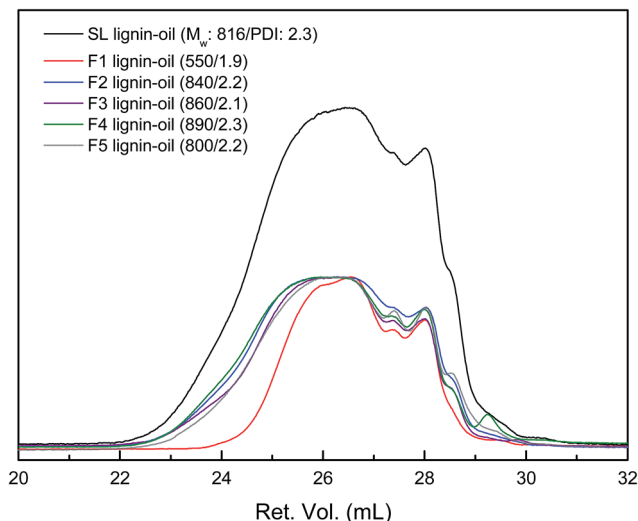


Fig. 7 GPC curves of lignin-oils produced from each lignin fraction (the peak maxima were normalized to same value except for SL lignin-oil).

lignin-oil consisted of both monoaromatic compounds and higher molecular phenolics, and is also obvious evidence of efficient lignin depolymerization for the reaction condition used in this study. The average  $M_w$  value of lignin-oil ranged from 550 to 890 Da, which was in accordance with the trend of the  $M_w$  of lignin fraction. These values are similar to the results of Cheng *et al.* who described lignin degraded product with the  $M_w$  values between 670 to 940 from alkaline lignin obtained at 200–450 °C using water/ethanol with various catalysts.<sup>44</sup> The depolymerization degree (%) of lignin-oil was the highest at F4 (88%) and the lowest at F1 (51%). The highest value at F4 presumably corresponded to the abundant part of lignin potentially degraded. Meanwhile, the PDI of lignin-oil shows similar or slightly increased values compared with that of the lignin fraction, except for the SL case. This result implies that lignin with a wide molecular distribution (SL) could be more easily degraded into narrow molecular range lignin-oil than other fractionated lignins beginning with narrow a molecular distribution. It was also explained that side reactions involving condensation, recondensation, and further decomposition occurred simultaneously with main lignin degradation reaction, leading to product variation in lignin-oil.<sup>6,10,14,45</sup>

Table 3 GPC results of lignin-oils produced from each lignin fraction

Lignin-oil	The $M_w$ (Da)/PDI		The average $M_w$ (Da)/PDI	Depolymerization degree (%)
	Before 28 ml	Near 28 ml		
SL	870/1.3	150/1.1	820/2.3	71
F1	690/1.2	160/1.1	550/1.9	51
F2	880/1.3	150/1.1	840/2.2	71
F3	1120/1.3	150/1.1	860/2.1	85
F4	1040/1.3	150/1.1	890/2.3	88
F5	890/1.2	150/1.1	800/2.2	—

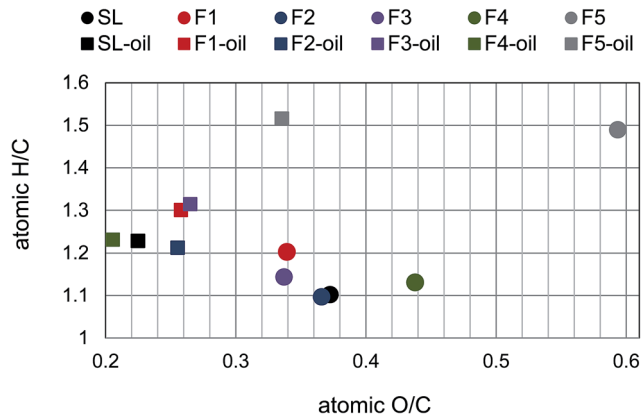


Fig. 8 van Krevelen diagram of lignin fractions and lignin-oils.

**3.4.3 Elemental composition of lignin-oils.** A Van Krevelen diagram showing the H/C and O/C ratios of lignin fractions and their derived lignin-oils is displayed in Fig. 8. Overall, the elemental composition of lignin fractions and lignin-oils did not show a significant difference depending on the  $M_w$  of lignin fraction. Lignin fractions had 1.1 to 1.2 of H/C ratios and 0.34 to 0.44 of O/C ratios, except for F5. F5 demonstrated relatively higher H/C and O/C ratios than those of other fractions. After depolymerization of each lignin fraction, the O/C ratios of lignin-oils considerably decreased to 0.21–0.26, while their H/C ratios increased to 1.2–1.3. The H/C ratios of lignin-oils obtained in this experiment are similar to the findings of Huang *et al.* who reported that the THF-soluble fraction produced from supercritical ethanolysis of SL at 300 °C with  $\text{CuMgAlO}_x$  catalyst has H/C ratios of approximately 1.2 with O/C ratios of 0.1–0.2. The difference in O/C ratios might be resulted from different reaction conditions (*e.g.* temperature, reaction time, and catalyst).<sup>14</sup> The change in H/C and O/C ratios after the depolymerization process was indicative of enhancing Ru/C catalyzed hydrodeoxygenation, as well as hydrogenation under high reaction temperature and  $\text{H}_2$  pressure.<sup>46,47</sup>

## 4. Conclusions

The lignin structure, especially the  $M_w$ , had a crucial impact on the yield and distribution of monoaromatic compounds produced after the catalytic depolymerization process. Lignin fractions with a relatively lower  $M_w$  value had lower amounts of  $\beta$ -O-4 linkages. These fractions yielded a relatively higher concentration of C6P, C1C6P, and C2C6P than other lignin fractions. In contrast, C3C6Ps were mainly produced in high  $M_w$  lignin fractions (F3 and F4). It can be envisioned from these results that pre-solvent fractionation of lignin will help in the selective production of specific monoaromatic compounds after the lignin depolymerization process. This study also investigated the chemical properties of lignin-oils from different lignin fractions, providing basic information for design of further application strategies.






## Conflicts of interest

There are no conflicts to declare.

## Acknowledgements

This study was supported by Korea Forest Service (Korea Forestry Promotion Institute) through 'R&D Program for Forest Science Technology (Project No. 2017052A00-1720-BB02)'.  


## References

- 1 D. Stewart, *Ind. Crop. Prod.*, 2008, **27**, 202–207.
- 2 X. Sun, C. Huang, Z. Xue, C. Yan and T. Mu, *Energy Fuels*, 2015, **29**, 2564–2570.
- 3 J. Zakzeski, P. C. Bruijninx, A. L. Jongerius and B. M. Weckhuysen, *Chem. Rev.*, 2010, **110**, 3552–3599.
- 4 A. J. Ragauskas, G. T. Beckham, M. J. Bidy, R. Chandra, F. Chen, M. F. Davis, B. H. Davison, R. A. Dixon, P. Gilna and M. Keller, *Science*, 2014, **344**, 1246843.
- 5 Z. Xue, X. Zhao, R.-c. Sun and T. Mu, *ACS Sustainable Chem. Eng.*, 2016, **4**, 3864–3870.
- 6 P. J. Deuss, M. Scott, F. Tran, N. J. Westwood, J. G. de Vries and K. Barta, *J. Am. Chem. Soc.*, 2015, **137**, 7456–7467.
- 7 C. R. Kumar, N. Anand, A. Kloekhorst, C. Cannilla, G. Bonura, F. Frusteri, K. Barta and H. J. Heeres, *Green Chem.*, 2015, **17**, 4921–4930.
- 8 J. A. Onwudili and P. T. Williams, *Green Chem.*, 2014, **16**, 4740–4748.
- 9 J.-Y. Kim, J. Park, U.-J. Kim and J. W. Choi, *Energy Fuels*, 2015, **29**, 5154–5163.
- 10 J.-Y. Kim, S. Oh, H. Hwang, T.-s. Cho, I.-G. Choi and J. W. Choi, *Chemosphere*, 2013, **93**, 1755–1764.
- 11 J.-Y. Kim, J. Park, H. Hwang, J. K. Kim, I. K. Song and J. W. Choi, *J. Anal. Appl. Pyrolysis*, 2015, **113**, 99–106.
- 12 K. Barta, T. D. Matson, M. L. Fettig, S. L. Scott, A. V. Iretskii and P. C. Ford, *Green Chem.*, 2010, **12**, 1640–1647.
- 13 A. Narani, R. K. Chowdari, C. Cannilla, G. Bonura, F. Frusteri, H. J. Heeres and K. Barta, *Green Chem.*, 2015, **17**, 5046–5057.
- 14 X. Huang, T. I. Korányi, M. D. Boot and E. J. Hensen, *ChemSusChem*, 2014, **7**, 2276–2288.
- 15 Y. Ye, Y. Zhang, J. Fan and J. Chang, *Bioresour. Technol.*, 2012, **118**, 648–651.
- 16 F. P. Bouxin, A. McVeigh, F. Tran, N. J. Westwood, M. C. Jarvis and S. D. Jackson, *Green Chem.*, 2015, **17**, 1235–1242.
- 17 X. Pan, J. F. Kadla, K. Ehara, N. Gilkes and J. N. Saddler, *J. Agric. Food Chem.*, 2006, **54**, 5806–5813.
- 18 X. Dong, M. Dong, Y. Lu, A. Turley, T. Jin and C. Wu, *Ind. Crops Prod.*, 2011, **34**, 1629–1634.
- 19 H. Yoshida, R. Möreck, K. P. Kringstad and H. Hatakeyama, *J. Appl. Polym. Sci.*, 1990, **40**, 1819–1832.
- 20 K. Sudo and K. Shimizu, *J. Appl. Polym. Sci.*, 1992, **44**, 127–134.
- 21 A. Toledano, L. Serrano, A. M. Balu, R. Luque, A. Pineda and J. Labidi, *ChemSusChem*, 2013, **6**, 529–536.
- 22 M.-F. Li, S.-N. Sun, F. Xu and R.-C. Sun, *Separ. Purif. Technol.*, 2012, **101**, 18–25.
- 23 S. Baker, *Holzforschung*, 1996, **50**, 573–574.
- 24 Y. Pu, S. Cao and A. J. Ragauskas, *Energy Environ. Sci.*, 2011, **4**, 3154–3166.
- 25 J.-L. Wen, S.-L. Sun, B.-L. Xue and R.-C. Sun, *Materials*, 2013, **6**, 359–391.
- 26 A. Sluiter, B. Hames, R. Ruiz, C. Scarlata, J. Sluiter, D. Templeton and D. Crocker, *Laboratory Analytical Procedure*, 2008.
- 27 C. Schuerch, *J. Am. Chem. Soc.*, 1952, **74**, 5061–5067.
- 28 A. Arshanitsa, J. Ponomarenko, T. Dizhbite, A. Andersone, R. J. Gosselink, J. van der Putten, M. Lauberts and G. Telysheva, *J. Anal. Appl. Pyrolysis*, 2013, **103**, 78–85.
- 29 G. Wang and H. Chen, *Sep. Purif. Technol.*, 2013, **120**, 402–409.
- 30 T.-Q. Yuan, J. He, F. Xu and R.-C. Sun, *Polym. Degrad. Stab.*, 2009, **94**, 1142–1150.
- 31 M. N. Gupta, R. Batra, R. Tyagi and A. Sharma, *Biotechnol. Prog.*, 1997, **13**, 284–288.
- 32 Z. Strassberger, P. Prinsen, F. van der Klis, D. S. van Es, S. Tanase and G. Rothenberg, *Green Chem.*, 2015, **17**, 325–334.
- 33 S. Constant, H. L. Wienk, A. E. Frissen, P. de Peinder, R. Boelens, D. S. Van Es, R. J. Grisel, B. M. Weckhuysen, W. J. Huijgen and R. J. Gosselink, *Green Chem.*, 2016, **18**, 2651–2665.
- 34 K. H. Kim, R. C. Brown, M. Kieffer and X. Bai, *Energy Fuels*, 2014, **28**, 6429–6437.
- 35 M. Saidi, F. Samimi, D. Karimipourfard, T. Nimmanwudipong, B. C. Gates and M. R. Rahimpour, *Energy Environ. Sci.*, 2014, **7**, 103–129.
- 36 J. N. Murwanashyaka, H. Pakdel and C. Roy, *Sep. Purif. Technol.*, 2001, **24**, 155–165.
- 37 J. Das and A. B. Halgeri, *Appl. Catal., A*, 2000, **194**, 359–363.
- 38 Z. Jiang, T. He, J. Li and C. Hu, *Green Chem.*, 2014, **16**, 4257–4265.
- 39 W. Xu, S. J. Miller, P. K. Agrawal and C. W. Jones, *ChemSusChem*, 2012, **5**, 667–675.
- 40 B. Loubinoux, M. Heitz, G. Coudert and G. Guillaumet, *Tetrahedron Lett.*, 1980, **21**, 4991–4994.
- 41 P. R. Patwardhan, R. C. Brown and B. H. Shanks, *ChemSusChem*, 2011, **4**, 1629–1636.
- 42 Q. Song, F. Wang, J. Cai, Y. Wang, J. Zhang, W. Yu and J. Xu, *Energy Environ. Sci.*, 2013, **6**, 994–1007.
- 43 <http://www.ebi.ac.uk/chebi/searchId.do?chebiId=CHEBI:2404>.
- 44 S. Cheng, C. Wilks, Z. Yuan, M. Leitch and C. C. Xu, *Polym. Degrad. Stab.*, 2012, **97**, 839–848.
- 45 Y. Jin, X. Ruan, X. Cheng and Q. Lü, *Bioresour. Technol.*, 2011, **102**, 3581–3583.
- 46 G. S. Macala, T. D. Matson, C. L. Johnson, R. S. Lewis, A. V. Iretskii and P. C. Ford, *ChemSusChem*, 2009, **2**, 215–217.
- 47 M. Kleinert, J. R. Gasson and T. Barth, *J. Anal. Appl. Pyrolysis*, 2009, **85**, 108–117.