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# Synthesis and stabilities of peptide-based [1]rotaxanes: molecular grafting onto lasso peptide scaffolds†

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The chemical synthesis of peptide-based [1]rotaxanes (lasso peptides) was achieved by [2]rotaxane formation followed by two chemoselective ligation reactions. Our approach enabled incorporation of various peptide sequences into a common rotaxane structure. The synthetic lasso peptides were characterized by NMR, chromatography, and partial degradation by proteases. A linear peptide epitope grafted onto the rotaxane scaffold showed significantly improved proteolytic stability.

## Introduction

Microcin J25 (MccJ25) and related structures are bacterially produced lasso peptides with a unique, interlocked structure that imparts remarkable thermal and proteolytic stabilities (Fig. 1A).<sup>1,2</sup> These peptides, consisting of 15–24 amino acids, are characterized by a threaded structure where a C-terminal region of the peptide threads through an N-terminal macrolactam ring. Amazingly, they are ribosomally synthesized followed by post-translational, enzymatic cyclization and cleavage of a leader peptide. The steric bulk of amino acid side chains maintains the lasso topology.

Lasso peptides represent an extreme example of the many strategies employed to stabilize  $\alpha$ -peptide-based natural products. Unmodified linear peptides are generally unstable *in vivo*, and this intrinsic disadvantage hampers the use of peptides as drugs despite of their high selectivity and potency.<sup>3</sup> A number of methods have been proposed to enhance the stability of peptides including cyclization, backbone *N*-methylation, and incorporation of unnatural amino acids. To take advantage of these strategies, scientists have recently adopted a molecular grafting approach where peptides are incorporated into a stable peptide/protein scaffold.<sup>4</sup> Marahiel *et al.* have demonstrated a molecular grafting of the RGD-motif onto the MccJ25 scaffold using bacterial biosynthetic machinery.<sup>5</sup> This approach made the otherwise unstable RGD peptide resistant to proteolytic degradation while showing integrin binding in the nanomolar range. Though the molecular grafting on MccJ25 was achieved

by biosynthesis, there has been no report on chemical synthesis of natural lasso peptides simply because formation of the threaded structure is extremely difficult.

In this manuscript, we report chemical syntheses of peptide-based [1]rotaxanes (lasso peptides) reminiscent of natural lasso peptides (Fig. 1B). Chromatography, NMR, and analysis of partial enzymatic degradation products confirmed the successful formation of the threaded structure and evaluation

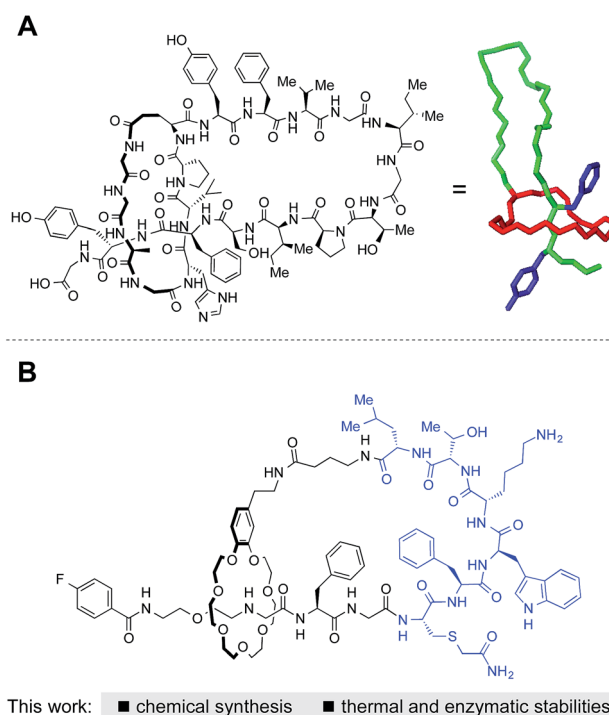


Fig. 1 Lasso peptides. (A) Structure of microcin J25 and (right) 3D structure adopted from PDB 1Q71. (B) This work: synthesis of lasso peptides by grafting with chemoselective ligations.

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of the physicochemical properties of the synthetic lasso peptides revealed excellent thermal and proteolytic stabilities.

## Results and discussion

### Synthetic strategy to prepare lasso peptides

Initially, we considered two strategies to construct lasso peptides (Fig. 2):<sup>6</sup> (1) formation of unthreaded peptide  $A_{un}$  resulting from a covalent link between a macrocycle and a peptide-containing segment.  $A_{un}$  is in a thermodynamic equilibrium with pseudo[1]rotaxane **A** where the successful trapping of **A** with a capping agent **B** would lead to lasso peptide **C**; (2) formation of [2]rotaxane **D** followed by the attachment of a linear peptide **E** by two sequential chemical ligations to afford lasso peptide **C**.

Pioneering studies from Leigh, Aucagne and others on the preparation and properties of peptide rotaxanes provided promising precedent for the planned studies.<sup>7</sup> Importantly, the synthesis of lasso peptides has been accomplished by Coutrot *et al.*,<sup>8</sup> who incorporated triglycine residues into a [1]rotaxane scaffold by a self-entanglement strategy. However, the method for locking a molecule into the lasso structure was not practical, and the diversity of peptide sequences limited.

After careful consideration, we adopted the second approach, as with the first strategy the topology-determining step potentially produces  $C_{un}$  and **C** that are difficult to differentiate. In contrast, in the latter approach, the threaded structure of [2]rotaxane **D** – which can be easily characterized – is retained through the synthesis provided that the [2]rotaxane structure is thermally and chemically stable. More importantly, the latter approach enables modular synthesis of lasso peptides. Once an appropriate synthetic scheme is established, the “stitching” strategy enables incorporation of an almost unlimited number of peptide sequences into the stable lasso peptide scaffold.

### Synthesis of [2]rotaxane with a potassium acyltrifluoroborate (KAT) as capping agent

We initially focused on preparation of [2]rotaxanes by a simple protocol. We selected the molecular recognition between benzo-

21-crown-7 (B21C7) and secondary dialkylammoniums, as they have been frequently used for rotaxane syntheses.<sup>9</sup> A preliminary study revealed the coupling of a potassium acyltrifluoroborate (KAT) and an azide in the presence of  $\text{HBF}_4 \cdot \text{OEt}_2$  (ref. 10) was suitable for forming a [2]rotaxane.<sup>11</sup> This result prompted us to prepare 4-fluorophenyl KAT **1**, crown ether **2** and azide **3** (Fig. 3). **2** and **3** are pre-functionalized with a protected hydroxylamine and thioester, respectively, which are the key functional groups for the subsequent ligation steps. These functional groups did not interfere with the formation of the [2]rotaxane, and **4** was obtained in 21% yield as a mixture of diastereomers **4a** and **4b**. The axle component **5** was concomitantly formed in the same reaction in 31% yield and used for synthesis of branched-cyclic peptides.

### Synthesis of lasso peptides from [2]rotaxane **4**

With a reliable protocol for the preparation of [2]rotaxanes in hand, we derivatized [2]rotaxane **4** to form lasso peptide by two amide-forming ligations, the  $\alpha$ -ketoacid-hydroxylamine (KAHA) ligation<sup>12</sup> and native chemical ligation (NCL)<sup>13</sup> (Fig. 4). Fmoc cleavage by 5%  $\text{Et}_2\text{NH}$  in DMSO revealed unprotected hydroxylamine **6** in 78% yield, poised for KAHA ligation. Importantly, the thioester remained intact under the Fmoc cleavage condition employed. Peptide  $\alpha$ -ketoacid **7** was prepared by Fmoc SPPS following the protocol recently reported by our group.<sup>14</sup> The peptide sequence of **7** was adapted from a commercially available peptide drug, octreotide, a somatostatin analog used for the treatment of diseases such as acromegaly.<sup>15</sup> We found that a cysteine side chain protecting group was necessary; in this case the *S*-unprotected variant of **7** did not undergo clean KAHA ligation, presumably because of intramolecular cyclization between the N-terminal cysteine and the C-terminal leucine  $\alpha$ -ketoacid.

Rotaxane **6** and peptide **7** were ligated in DMSO/ $\text{H}_2\text{O}$  (6 : 4) at 60 °C to form peptido[2]rotaxane **8**. Unfortunately, the *O*-diethylcarbamoyl hydroxylamine was somewhat unstable under the KAHA ligation conditions (Fig. S4†). Nonetheless, the ligation product **8** was isolated in 23% yield as a major product.

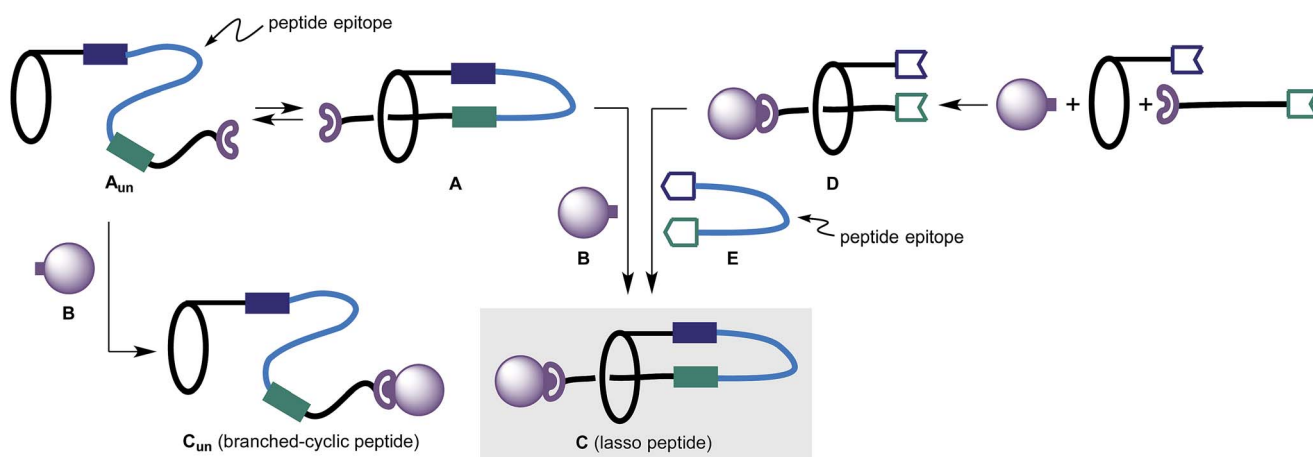


Fig. 2 Strategies for synthesis of lasso peptides. un = unthreaded.



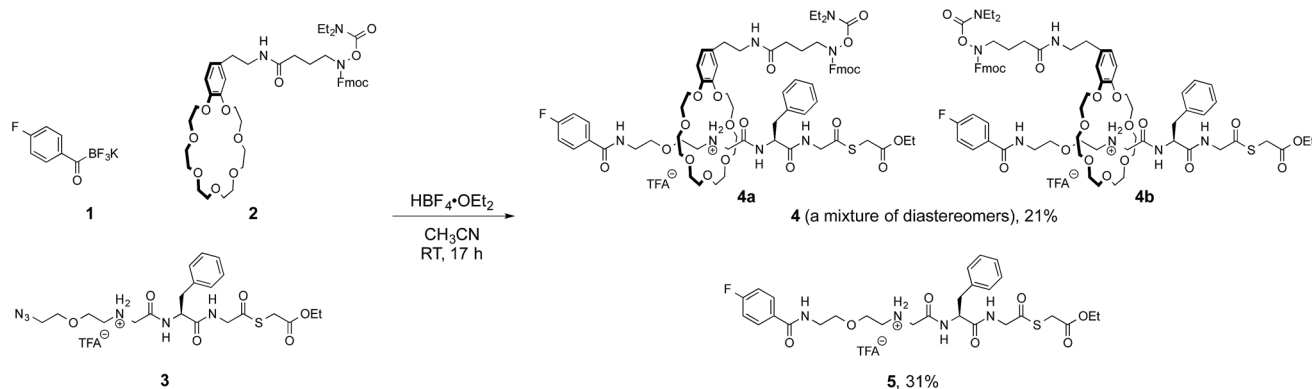


Fig. 3 Formation of [2]rotaxane 4 by KAT-azide coupling.

The deprotection-KAHA ligation sequence could be carried out in a one-pot manner, reducing the number of purification steps and avoiding possible decomposition of unprotected hydroxylamines upon lyophilization.

The cyclization step was performed by NCL in the presence of an excess amount of 4-mercaptophenylacetic acid (MPAA) and TCEP. *In situ* deprotection of cysteine followed by ligation afforded the lasso peptide **9** in 71% yield. Alkylation of the sulfhydryl group present in **9** to suppress oxidation or dimerization of the cysteine residue gave **L1** in 66% yield.

This reaction sequence was exploited to prepare branched-cyclic peptide **B1**; Fmoc-cleavage, KAHA ligation, NCL followed by cysteine alkylation afforded **B1** (Scheme S4<sup>†</sup>). Both **L1** and **B1** were analyzed by NMR. Although we could not obtain compelling evidence for the threaded structure solely from NMR experiments, including <sup>1</sup>H-<sup>15</sup>N HSQC and ROESY spectra, the <sup>1</sup>H NMR spectrum of **L1** was clearly different from that of **B1**. In addition, HRMS, HPLC retention time, and analysis of partial proteolytic degradation products further confirmed the formation of the threaded structure in **L1**.<sup>16</sup>

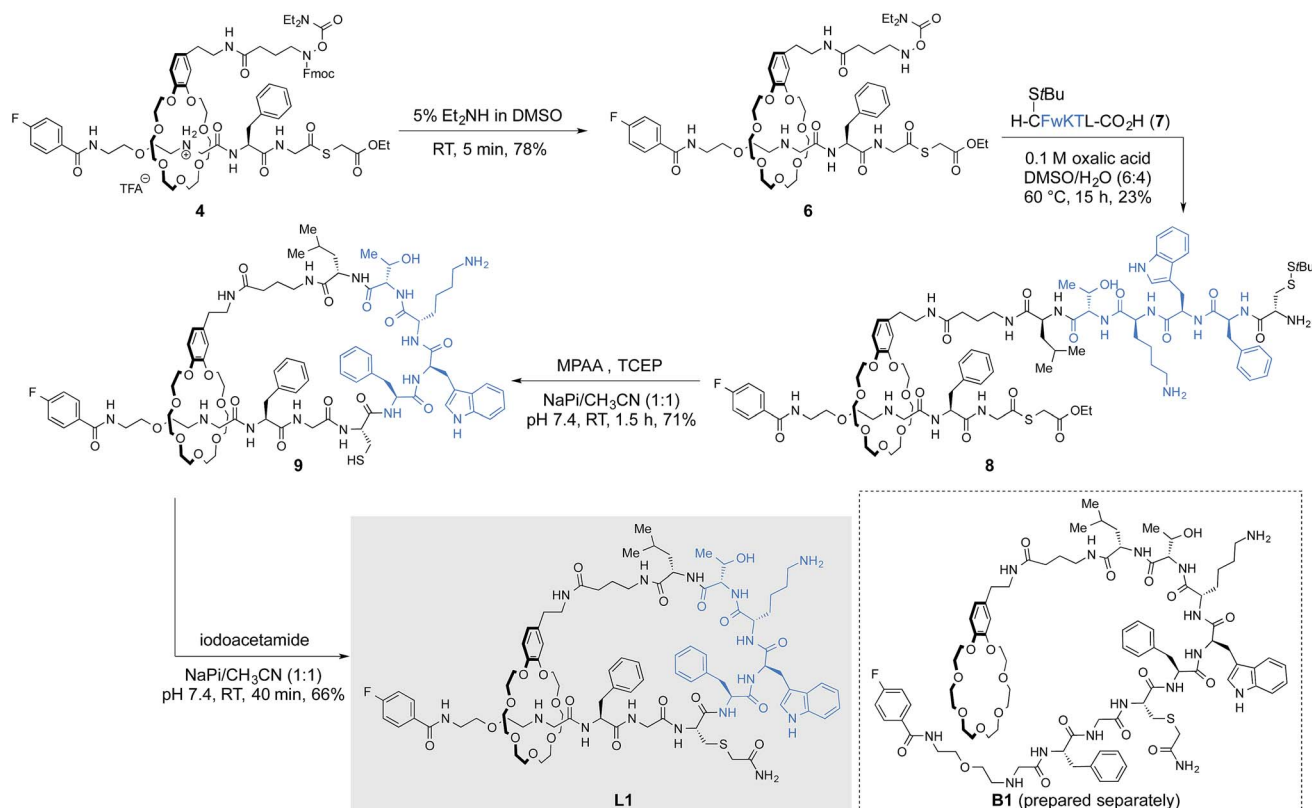


Fig. 4 Synthesis of lasso peptide **L1**. For simplicity, only one of the diastereomers is shown. All products were obtained as TFA salts after HPLC purification. TFAs were omitted from the structures for clarity. NaPi = sodium phosphate buffer.



We investigated the possibility of incorporating various peptide sequences into our rotaxane scaffold. The peptide sequences were taken from an antibody Fc region binder,<sup>17</sup> lassomycin,<sup>18</sup> and somatostatin, which gave 9, 10 and 12-mer peptides, respectively, including the N-terminal cysteine and the C-terminal leucine  $\alpha$ -ketoacid. The same reaction sequence was applicable and provided lasso peptides **10–12** (Fig. 5A).

The threaded compounds were subjected to trypsin digestion and degraded into [2]rotaxanes (Fig. 5B, S76 and S90<sup>†</sup>). The results established the existence of the threaded structure in **10–12**.

### Structure modification of the crown ether

With the synthetic protocol in hand, we began to modify the core structure [2]rotaxane. We were especially concerned about

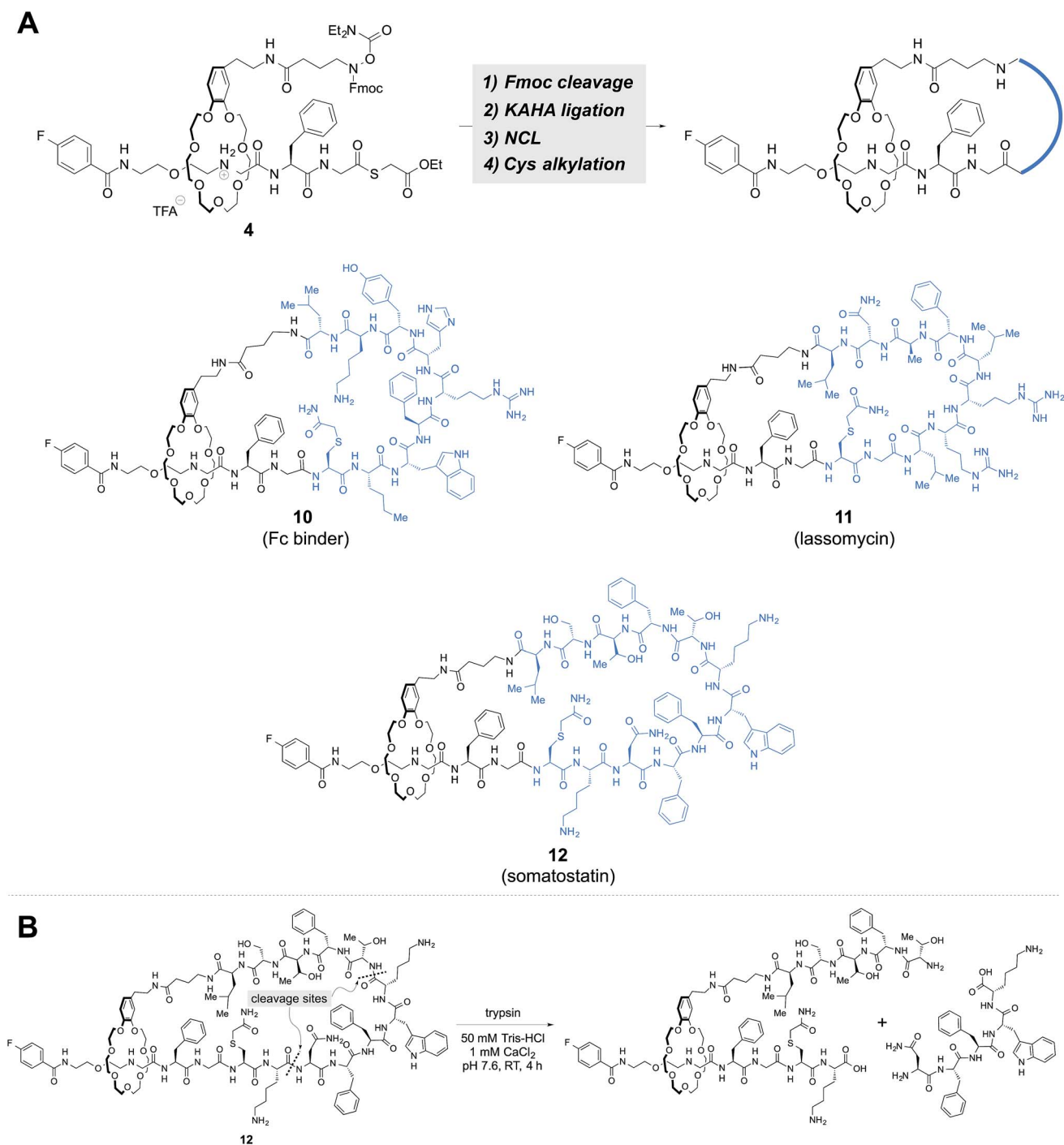


Fig. 5 Peptide substrate scope. (A) Insertion of linear peptides with different length and side chain functional groups. (B) Trypsin digestion of **12**. Degradation products were analyzed by HRMS (MALDI). The threaded structure of lasso peptides were retained in the form of [2]rotaxane.



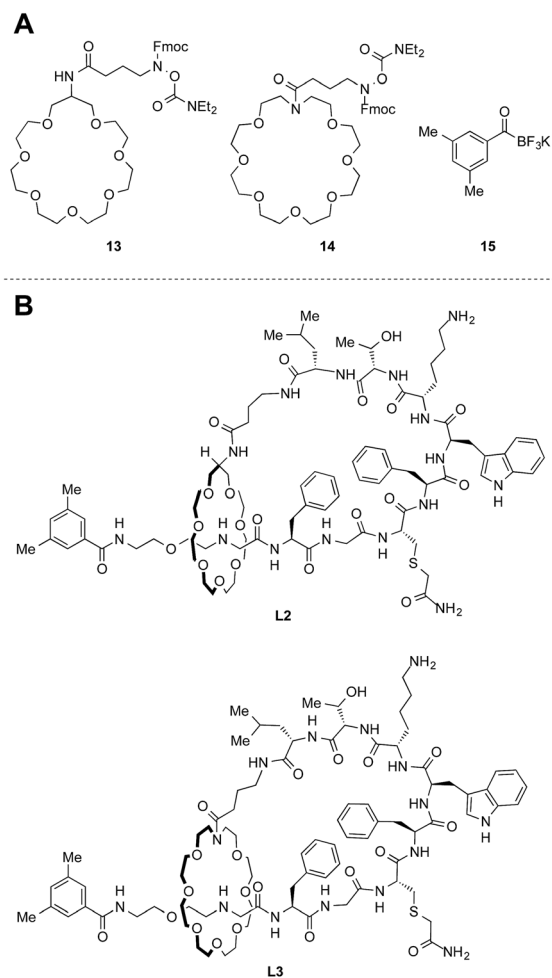


Fig. 6 Modification of the core [2]rotaxane structure. (A) Structures of crown ethers **13** and **14** and KAT **15**. (B) Structures of lasso peptides **L2** and **L3**. Note that **L2** was obtained as a mixture of diastereomers.

the electron-rich nature of the catechol moiety in the crown ether. This moiety could be susceptible to enzymatic oxidation and might pose other stability issues. Based on this consideration, we prepared two crown ethers **13** and **14** without electron-rich moieties (Fig. 6A). Since the new crown ethers have larger ring sizes, a larger capping agent **15** was needed to form thermally stable [2]rotaxanes (Fig. S23<sup>†</sup>). The established protocol was applied to the synthesis of peptides **L2** and **L3** (Fig. 6B). Branched-cyclic peptides **B2** and **B3** (*i.e.* unthreaded variants of **L2** and **L3**) were also prepared and used in the stability tests for comparison (*vide infra*).

### Chromatographic characterization of lasso peptides

It has long been known that lasso peptides and their branched-cyclic counterparts are chromatographically distinct and show different retention times upon analysis by HPLC.<sup>19</sup> This was true for our compounds; the retention times of the lasso peptides **L1–L3** were found to be different from those of the branched-cyclic peptides **B1–B3**, respectively (Fig. S62–S64<sup>†</sup>).

### Thermal and enzymatic stabilities of lasso peptides

Many natural lasso peptides are also characterized by their thermal stability. For example, MccJ25 did not denature at 95 °C and maintained its threaded structure.<sup>20</sup> As expected, the stopper groups used for the [2]rotaxane formation were large enough, and **L1–L3** did not show dethreading even at 95 °C for 8 h (Fig. S117–S119<sup>†</sup>).

We tested the proteolytic stability of lasso peptides **L1–L3**, linear peptides **B1–B3**, and the representative peptide macrocycle **C1** against chymotrypsin, trypsin, and proteinase K.<sup>11</sup> We were pleased to find that **L1–L3** were significantly more stable than **B1–B3**. Lasso peptides **L1–L3** and peptide macrocycle **C1** were similarly stable against trypsin and proteinase K; but in case of chymotrypsin, **L1–L3** were even more stable than

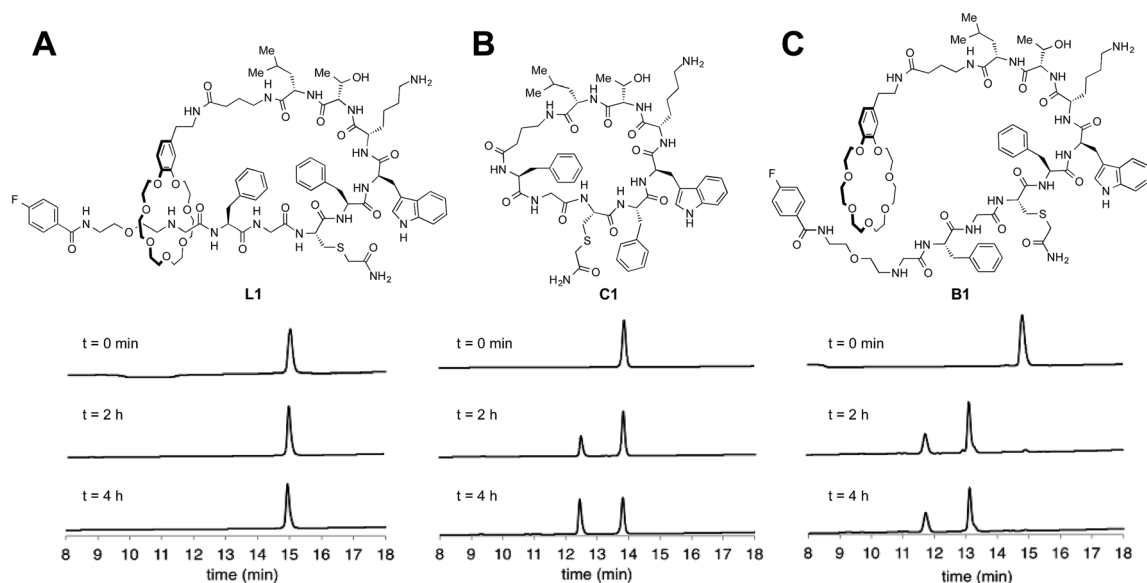


Fig. 7 HPLC monitoring of degradation by chymotrypsin. (A) Lasso peptide **L1**. (B) Peptide macrocycle **C1**. (C) Branched-cyclic peptide **B1**.



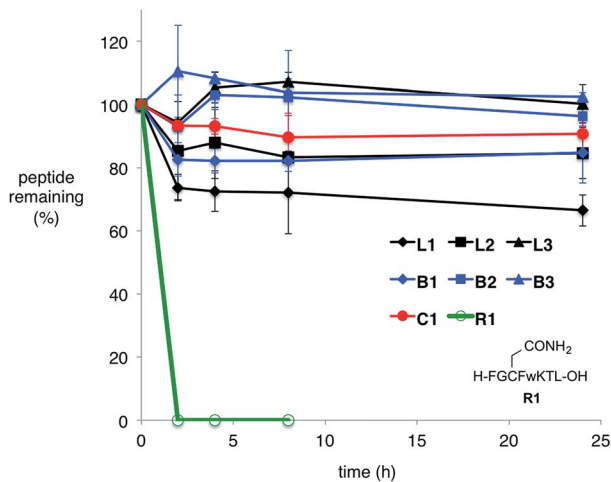


Fig. 8 Serum stability tests. Stability is evaluated based on remaining peptide substrate monitored by HPLC. Data is averaged from two independent experiments and error bars show standard deviations. R1 is an N- and C-terminal unmasked linear peptide.

macrocycle C1 (Fig. 7). In particular, L1 and L2 showed excellent resistance to degradation by chymotrypsin even after 24 h.

Serum stability tests were also conducted (Fig. 8). For the peptide sequence used in this study, N-, and C-terminal masking was effective enough to stabilize the peptides; N-, and C-terminal unmasked linear peptide R1 was degraded within 2 h in human serum. In contrast to lability against specific proteases mentioned above, branched-cyclic peptides B1–B3 were stable in human serum. All cyclic compounds except for L1 were stable under the condition for 24 h. L1, though reasonably stable, was gradually degraded possibly due to the catechol moiety.

## Conclusions

In summary, we achieved the molecular grafting of linear peptide epitopes onto a lasso peptide scaffold. The combination of KAHA ligation and NCL enabled attachment of various peptide sequences from a common, readily prepared rotaxane. The synthetic lasso peptides were thermally stable, and the peptide epitope gained significant proteolytic stability upon grafting. A merger of our molecular grafting approach and other stability enhancement techniques would provide further improvements of physicochemical properties of peptides.

## Notes

A patent application covering portions of this work has been filed.

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## Notes and references

- 1 Thermally unstable lasso peptides have been recently discovered: J. D. Hagemann, M. Zimmermann, X. Xie and M. A. Marahiel, *J. Am. Chem. Soc.*, 2013, **135**, 210–222.
- 2 (a) M. O. Maksimov, S. J. Pan and A. J. Link, *Nat. Prod. Rep.*, 2012, **29**, 996–1006; (b) J. D. Hagemann, M. Zimmermann, X. Xie and M. A. Marahiel, *Acc. Chem. Res.*, 2015, **48**, 1909–1919; (c) Y. Li, S. Zirah and S. Rebuffat, *Lasso Peptides: Bacterial Strategies to make and maintain Bioactive Entangled Scaffolds*, Springer, New York, 2015.
- 3 D. J. Craik, D. P. Fairlie, S. Liras and D. Price, *Chem. Biol. Drug Des.*, 2013, **81**, 136–147.
- 4 (a) S. Gunasekera, F. M. Foley, R. J. Clark, L. Sando, L. J. Fabri, D. J. Craik and N. L. Daly, *J. Med. Chem.*, 2008, **51**, 7697–7704; (b) C. T. T. Wong, D. K. Rowlands, C.-H. Wong, T. W. C. Lo, G. K. T. Nguyen, H.-Y. Li and J. P. Tam, *Angew. Chem., Int. Ed.*, 2012, **51**, 5620–5624; (c) C. K. Wang, C. W. Gruber, M. Cemazar, C. Siatskas, P. Tagore, N. Payne, G. Sun, S. Wang, C. C. Bernard and D. J. Craik, *ACS Chem. Biol.*, 2014, **9**, 156–163; (d) A. C. Conibear, S. Chaousis, T. Durek, K. J. Rosengren, D. J. Craik and C. I. Schroeder, *Biopolymers*, 2016, **106**, 89–100.
- 5 (a) T. A. Knappe, F. Manzenrieder, C. Mas-Moruno, U. Linne, F. Sasse, H. Kessler, X. Xie and M. A. Marahiel, *Angew. Chem., Int. Ed.*, 2011, **50**, 8714–8717; (b) J. D. Hagemann, M. D. Simone, M. Zimmermann, T. A. Knappe, X. Xie, F. S. D. Leva, L. Marinelli, E. Novellino, S. Zahler, H. Kessler and M. A. Marahiel, *J. Med. Chem.*, 2014, **57**, 5829–5834.
- 6 For discussion about synthesis of [1]rotaxanes, see: P. Waelès, C. Clavel, K. Fournel-Marotte and F. Coutrot, *Chem. Sci.*, 2015, **6**, 4828–4836.
- 7 (a) A. Fernandes, A. Viterisi, F. Coutrot, S. Potok, D. A. Leigh, V. Aucagne and S. Papot, *Angew. Chem., Int. Ed.*, 2009, **48**, 6443–6447; (b) A. Fernandes, A. Viterisi, V. Aucagne, D. A. Leigh and S. Papot, *Chem. Commun.*, 2012, **48**, 2083–2085; (c) B. Lewandowski, G. De Bo, J. W. Ward, M. Pappmeyer, S. Kuschel, M. J. Aldegunde, P. M. E. Gramlich, D. Heckmann, S. M. Goldup, D. M. D'Souza, A. E. Fernandes and D. A. Leigh, *Science*, 2013, **339**, 189–193; (d) G. De Bo, S. Kuschel, D. A. Leigh, B. Lewandowski, M. Pappmeyer and J. W. Ward, *J. Am. Chem. Soc.*, 2014, **136**, 5811–5814.
- 8 C. Clavel, K. Fournel-Marotte and F. Coutrot, *Molecules*, 2013, **18**, 11553–11575.
- 9 (a) C. Zhang, S. Li, J. Zhang, K. Zhu, N. Li and F. Huang, *Org. Lett.*, 2007, **9**, 5553–5556; (b) Q. Zhou, P. Wei, Y. Zhang, Y. Yu and X. Yan, *Org. Lett.*, 2013, **15**, 5350–5353.
- 10 G. A. Molander, J. Raushel and N. M. Ellis, *J. Org. Chem.*, 2010, **75**, 4304–4306.



- 11 See ESI† for detailed procedures.
- 12 J. W. Bode, R. M. Fox and K. D. Baucom, *Angew. Chem., Int. Ed.*, 2006, **45**, 1248–1252.
- 13 P. E. Dawson, T. W. Muir, I. Clark-Lewis and S. B. H. Kent, *Science*, 1994, **266**, 776–779.
- 14 T. G. Wucherpfennig, V. R. Pattabiraman, F. R. P. Limberg, J. Ruiz-Rodríguez and J. W. Bode, *Angew. Chem., Int. Ed.*, 2014, **53**, 12248–12252.
- 15 G. Weckbecker, I. Lewis, R. Albert, H. A. Schmid, D. Hoyer and C. Bruns, *Nat. Rev. Drug Discovery*, 2003, **2**, 999–1017.
- 16 See ESI† for detailed characterization of **L1**.
- 17 S. Menegatti, M. Hussain, A. D. Naik, R. G. Carbonell and B. M. Rao, *Biotechnol. Bioeng.*, 2013, **110**, 857–870.
- 18 E. Gavrish, C. S. Sit, S. Cao, O. Kandrór, A. Spoering, A. Peoples, L. Ling, A. Fetterman, D. Hughes, A. Bissell, H. Torrey, T. Akopian, A. Mueller, S. Epstein, A. Goldberg, J. Clardy and K. Lewis, *Chem. Biol.*, 2014, **21**, 509–518.
- 19 K. J. Rosengren, R. C. Clark, N. L. Daly, U. Göransson, A. Jones and D. J. Craik, *J. Am. Chem. Soc.*, 2003, **125**, 12464–12474.
- 20 A. Blond, M. Cheminant, D. Destoumieux-Garzon, I. Segalas-Milazzo, J. Peduzzi, C. Goulard and S. Rebuffat, *Eur. J. Biochem.*, 2002, **269**, 6212–6222.

