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A thiocyanopalladation/carbocyclization transformation identified through enzymatic screening: stereocontrolled tandem C–SCN and C–C bond formation†

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Herein we describe a formal thiocyanopalladation/carbocyclization transformation and its parametrization and optimization using a new elevated temperature plate-based version of our visual colorimetric enzymatic screening method for reaction discovery. The carbocyclization step leads to C–SCN bond formation in tandem with C–C bond construction and is highly stereoselective, showing nearly absolute 1,2-*anti*-stereinduction (5 examples) for substrates bearing allylic substitution, and nearly absolute 1,3-*syn*-stereinduction (16 examples) for substrates bearing propargylic substitution. Based upon these high levels of stereinduction, the dependence of the 1,2-stereinduction upon cyclization substrate geometry, and the generally high preference for the transoid vinyl thiocyanate alkene geometry, a mechanistic model is proposed, involving (i) Pd(II)-enyne coordination, (ii) thiocyanopalladation, (iii) migratory insertion and (iv) β -elimination. Examples of transition metal-mediated C–SCN bond formation that proceed smoothly on unactivated substrates and allow for preservation of the SCN moiety are lacking. Yet, the thiocyanate functionality is of great value for biophysical chemistry (vibrational Stark effect) and medicinal chemistry (S,N-heterocycle construction). The title transformation accommodates C-, O-, N- and S-bridged substrates (6 examples), thereby providing the corresponding carbocyclic or heterocyclic scaffolds. The reaction is also shown to be compatible with a significant range of substituents, varying in steric and electronic demand, including a wide range of substituted aromatics, fused bicyclic and heterocyclic systems, and even biaryl systems. Combination of this new transformation with asymmetric allylation and Grubbs ring-closing metathesis provides for a streamlined enantio- and diastereoselective entry into the oxabicyclo[3.2.1]octyl core of the natural products massarilactone and annuonone A, as also evidenced by low temperature X-ray crystal structure determination. Utilizing this bicyclic scaffold, we demonstrate the versatility of the thiocyanate moiety for structural diversification post-cyclization. Thus, the bridging vinyl thiocyanate moiety is smoothly elaborated into a range of derivative functionalities utilizing transformations that cleave the S–CN bond, add the elements of RS–CN across a π -system and exploit the SCN moiety as a cycloaddition partner (7 diverse examples). Among the new functionalities thereby generated are thiotetrazole and sulfonyl tetrazole heterocycles that serve as carboxylate and phosphate surrogates, respectively, highlighting the potential of this approach for future applications in medicinal chemistry or chemical biology.

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Introduction

Described herein are our efforts to develop new chemistry for the introduction of the thiocyno functionality into natural product

core structures with control of stereochemistry. While important strides have been made in developing transition metal (TM)-mediated bond constructions with simple thiolates,¹ TM-based C–S bond formation still lags well behind the corresponding C–C, C–N and C–O-bond formation chemistry. In particular, there are few such reaction manifolds for installing valuable C–SCN bonds.^{2,3} Two issues loom large here, the longstanding documentation of TM-catalyst poisoning by thiol species^{1a,4} especially thiocyanate anion,⁵ and the lability of the SCN functionality.⁶ The title transformation emerged from a wider search for transformations of synthetic utility employing our *in situ* enzymatic screening (ISES) approach to reaction discovery.

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The ISES approach is part of a recent expansion of research in both the synthetic organic and process chemistry communities that explores the interplay of enzymatic chemistry and traditional synthetic chemistry. While there is an important spectrum of activity at this biocatalysis/synthesis interface,^{7–12} we have been particularly focused on the use of enzymes to screen potential catalytic organic and organometallic combinations across chemical transformations of interest with attention to both throughput¹³ and information content.^{14,15} In ISES, enzymes report directly to the experimentalist upon the course of a matrix of test reactions of synthetic organic interest, without the need to draw aliquots, quench or chromatograph samples.^{16,17} Particularly useful in these endeavors has been the development of visual colorimetric ISES;¹⁸ as depicted in Fig. 1A. Note that in this system, successful turnover of an alkyl carbonate substrate by a transition metal catalyst, for example, leads to diffusion of the alcohol-presumably following spontaneous decarboxylation of the alkyl carbonate leaving group-into the adjacent aqueous layer where two ‘reporting enzymes’ are present. In the aqueous ‘reporting layer’, alcohol oxidase oxidizes the alcohol to the corresponding aldehyde, thereby generating an H₂O₂ equivalent that is itself captured by a second reporting enzyme, peroxidase, and reduced to H₂O with the assistance of an ABTS dye cofactor. This reporting chemistry is reminiscent of ELISA technology, with the redox active dye generating two equivalents of the ABTS radical cation per substrate turnover event. This chromophore is both intense ($\epsilon_{404-414} \sim 70\,000\text{ M}^{-1}\text{ cm}^{-1}$ for two equivalents) and absorbs in

the visible, leading to the appearance of green color in the enzymatic reporting layer, easily to visible the naked eye.

Results and discussion

Utilizing enzymatic screening as a validation/parametrization tool

Utilizing the ISES platform illustrated in Fig. 1A, a broad-based search for (pseudo)halo-carbocyclization chemistry was previously conducted across a matrix of 64 transition metal (TM) complexes \times 6 (pseudo)halides \times 3 substrates.^{18b} This type of transformation is of particular interest to our group as it results in the installation of (pseudo)halovinyl moiety in the product, in keeping with our longstanding interest in developing halovinyl functionalities for mechanism-based enzyme inhibitors.¹⁹ Of the 1152 combinations screened, perhaps the most interesting and unprecedented discovery was the formal transition metal-mediated thiocyno-carbocyclization through which 5-*exo*-trig ether **1** yields furanoid system **2** upon treatment with LiSCN/PdCl₂(PhCN)₂. To our knowledge, this single hit in our laboratory is the only example of such a TM-mediated carbocyclization in which C–SCN and C–C bonds are formed concomitantly. Indeed, the only precedents for TM-mediated C–SCN bond formation of which we are aware require special activation conditions. For example, there are a couple of recent examples of additions across π -systems with TMSCN and a highly activated electrophilic hypervalent iodine–CF₃ species.² And there is a report of a sequence involving TM-mediated

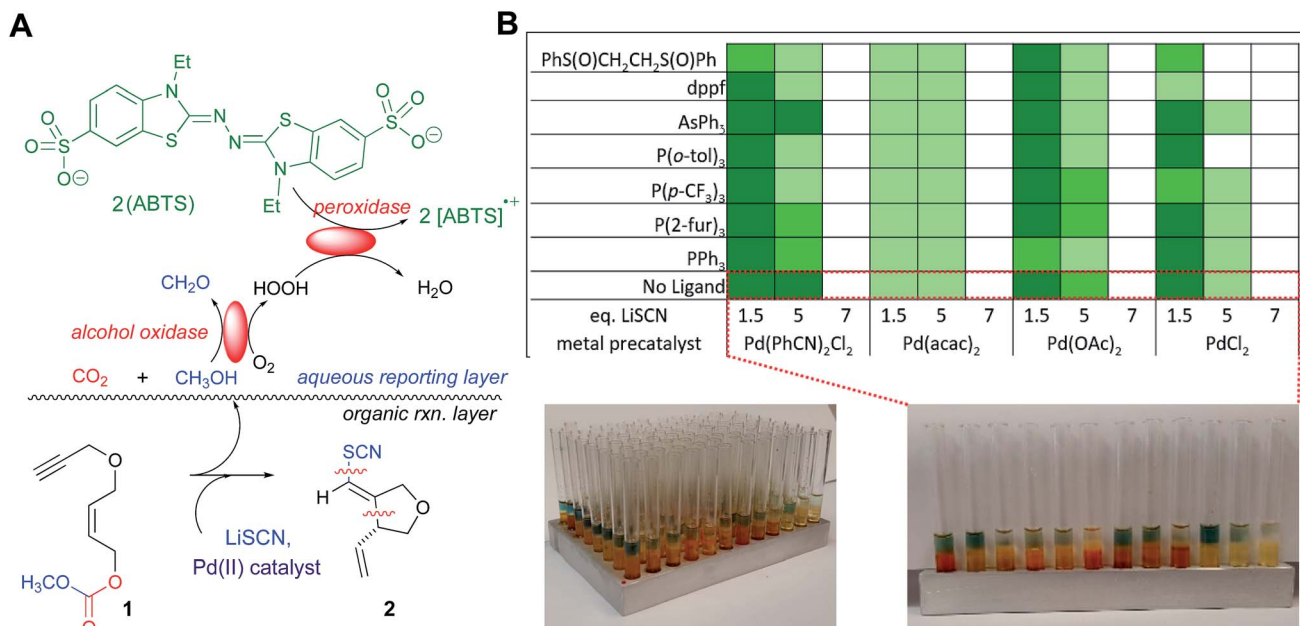


Fig. 1 A new elevated temperature format for the plate-based colorimetric enzymatic screen and its use in parametrizing and optimizing the new thiocyanopalladation/carbocyclization transformation. (A) Schematic of the colorimetric, enzyme-based screen for this new bond construction. Note: the green color is due to ABTS radical cation formed from the alcohol oxidase/peroxidase reporting enzyme couple, with intensity of the signal related to the efficiency of organometallic reaction screened. (B) Table illustrating the reaction parameters being probed here – nature of the Pd(II) catalyst, LiSCN loading, and ligand effects. Relative intensity of green shading indicates relative reaction progress after 15 min. (C) Images of the colorimetric enzymatic screen – entire 96-well aluminum plate after heating to 70 °C (sand bath). (D) Close-up view of the first row (white paper backing for clarity; dotted lines show how this row maps onto the schematic) highlighting the effect of LiSCN loading across the array of Pd(II) catalysts screened.



arene iodination, followed by non-metal-mediated substitution with thiocyanate.³

In this article, we describe the use of a new elevated temperature plate-based variant of colorimetric, enzymatic screening that allows us validate and parametrize this new transformation, and we report on the scope, stereochemical course and application of this chemistry to natural product core synthesis. Finally, by combining stereochemical probe substrates-varying alkene geometry and utilizing strategically placed resident stereocenters-we put forth a mechanistic postulate for this thiocyanopalladation/carbocyclization, consistent with the results of these studies.

Under the conditions of the initial screen, an excess of LiSCN was employed and Pd(II) catalyst-loading was at 10 mol% in an organic layer composed of THF/1,1,2-trichloroethane (TCE) at room temperature over ~20 min. Building on this single lead data point, we began our studies here with a rapid colorimetric screen across *T* (35–70 °C in 5 °C increments) and Pd-loading (2.5–10 mol%; see ESI† for details). This led to the conclusion that further screens should be at 60 °C and that 2.5 mol% Pd(II) would suffice. Given these observations, we set out to develop a thermal version of this visual colorimetric ISES. Accordingly, an aluminum 96 well tray (rapidly fabricated in any machine shop) was utilized as it was found that this platform could be conveniently heated in a sand bath. After some experimentation, it was established that the most efficient protocol here involved layering all of the organic/organometallic transformation samples to be screened into the respective wells/tubes in the tray, heating the 8 × 12 array to 60 °C for 15 min, cooling and then layering the aqueous reporting layers (100 μL each) thereupon, using a multichannel pipeter containing 8 arms/pipet tips. Following a 10 min enzymatic development period, the reporting well intensities were catalogued and color-coded accordingly (see Fig. 1C, and the ESI† for more details).

As highlighted in Fig. 1B, four palladium sources were screened: PdCl₂(PhCN)₂, Pd(acac)₂, Pd(OAc)₂, and PdCl₂. It was found that in addition to PdCl₂(PhCN)₂, Pd(OAc)₂, and PdCl₂ also support this chemistry, but Pd(acac)₂ appears to be an unacceptable Pd(II) source for this transformation. Most importantly, a significant effect of LiSCN loading on reaction efficiency was clearly visible across all viable Pd(II) catalysts screened. As can be seen in Fig. 1B and C, lower loadings of thiocyanate promote the reaction more effectively, perhaps owing to reduced catalyst poisoning. A preliminary ligand screen was also conducted. The results indicated that whereas sulfoxide ligands appear to inhibit the title transformation, mono- and bidentate phosphine ligands are at least compatible with the cyclization, opening up the possibility of more extensive ligand exploration in the future.

Overall then, this elevated temperature plate-based enzymatic screen led to the conclusion that the title transformation proceeds well at 2.5 mol% Pd(II) with 1.5 eq. of LiSCN in THF (or THF/TCE in the screening) with heating to ~60 °C (see ESI† for details). In this way, this initial screening-based optimization set the key parameters for studies described here in which we examine the scope of this new transformation with particular attention to functional group

tolerance, nature of the bridging functionality and stereochemical course.

We are pleased to report that the Pd(II)-mediated thiocyanocarboxylcyclization tolerates sulfur-, nitrogen- and carboxylate ester functionalities in the bridging position, and proceeds in very good yield, in general (Table 1). In most cases, high selectivity in favor of the transoid alkene is observed. Thus, the four C- and N-bridged systems, **4**, **6**, **8**, **10** all proceed with transoid selectivities >10 : 1, and as high as 30 : 1, in the case of the bridging trifluoroacetamide (**6**). Only for the bridging chalcogenides is this selectivity more modest, with the ether substrate **1** delivering geometric isomers **2t** and **2c** in a 4 : 1 ratio and thioether substrate **11** showing complete erosion of this selectivity. The general trend toward transoid products in these Pd(II)-mediated processes is consistent with initial delivery of the elements of SCN and Pd(II) across the alkyne in anti-fashion (*vide infra*). This anti-addition aligns with observations in other types metal-mediated enyne cyclizations reported by Lu and Ma,²⁰ Cook²¹ and by our group.^{18b}

The mixtures of transoid/cisoid products observed for substrates **1** and **11** may be indicative of either a combination of external and internal (*i.e.* from the metal center) thiocyanate delivery, in these cases, or could be the result of a rapid pre-equilibration in the thiocyanometallation step, prior to migratory insertion (*vide infra*). No cyclization product was observed with sulfone-bridged substrate **13**. Instead a conjugated sulfonyl butadiene species **14** was isolated [see ESI† for details of the formation of byproduct **14** = 1-(2'-thiocyanatoallylsulfonyl)buta-1E,3-diene; formula: C₈H₆NO₂S₂] presumably *via* 1,4-elimination of the carbonate facilitated by the enhanced C–H acidity of α to the sulfonyl functionality.

Structure elucidation

Alkene geometry was unambiguously established by a combination of X-ray crystallography and chemical shift correlation using ¹H and ¹³C NMR spectroscopy (Fig. 2). The crystal structure of the

Table 1 Variation of the bridging functionality

Substrate	Product	X	Yield (t : c) ^a
1	2	O	81% (4 : 1)
3	4	NTs	85% (11 : 1)
5	6	NCOCF ₃	87% (30 : 1)
7	8	C(COOEt) ₂	60 ^b % (24 : 1)
9	10	CH(COOEt)	80% (13 : 1) ^d
11	12	S	85% (1 : 1.5)
13	^c	SO ₂	Elimination byproduct 14

^a Ratio determined by crude ¹H NMR. ^b Based on recovered starting material. ^c See ESI materials for compound structure. ^d dr 1 : 1.5 ratio in the major transoid product.



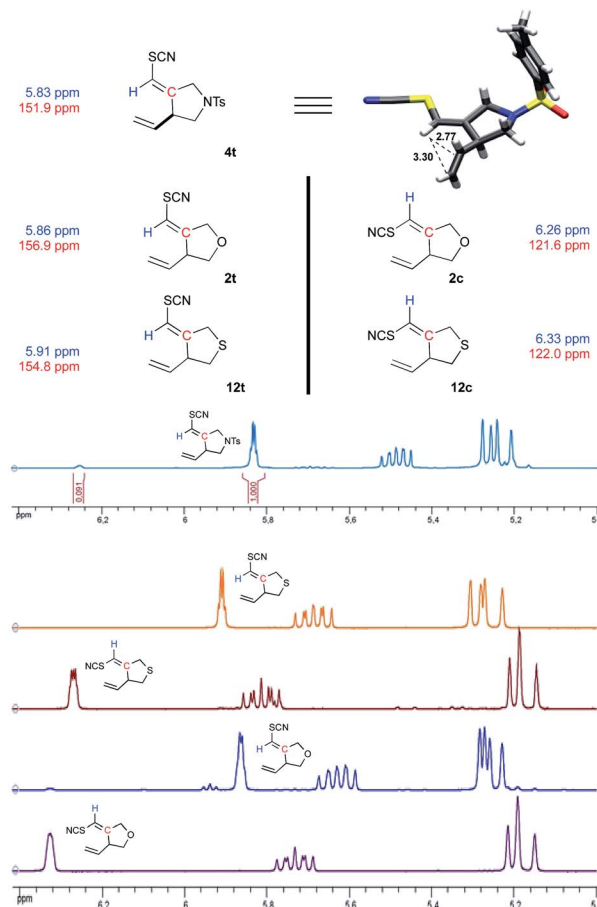


Fig. 2 Assignment of alkene geometry using a combination of X-ray crystallography and correlated ^1H and ^{13}C chemical shift patterns. Note the clear positioning of the vinyl thiocyanate methine proton in the shielding region above the π -system of the vinyl group in the transoid isomer, resulting in an ~ 0.4 ppm upfield shift, relative to the cisoid isomer.

NTs-bridged cyclized product **9** was solved and confirms the transoid alkene geometry in the predominant isomer. This result allowed us to align olefin geometry with the chemical shift trends seen in the NMR signatures, particularly of the vinyl thiocyanate functionality. Specifically, the cisoid cyclization products display upfield quaternary vinylic carbon signals (120–122 ppm) relative to those of their transoid counterparts (150–155 ppm). And the transoid cyclization products have upfield-shifted vinyl thiocyanate methine protons (5.8–5.9 ppm) relative to the corresponding methine signals in the cisoid isomers (6.2–6.3 ppm). The crystal structure of **4** also reveals the likely origin of this upfield shift; namely for the vinyl thiocyanate methane H appears to lie within the shielding cone of the π -system from the neighboring terminal vinyl group. The distances between this methine hydrogen and these vinylic carbons are 2.77 and 3.30 Å, respectively, within the range expected for π -shielding effects.²²

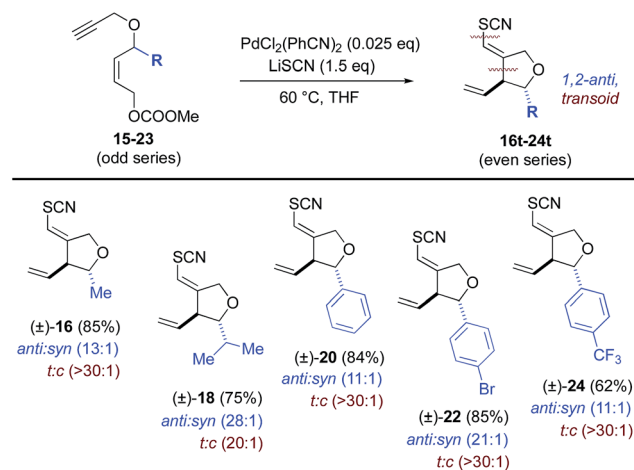
Stereocontrol and mechanism

Next, stereochemical course was examined, specifically, the use of extant stereocenters to set relative stereochemistry in the

thiocyanopalladation/carbocyclization product. The effects of substituents at both the allylic (Table 2) and propargylic (Table 3) positions upon induction of stereochemistry at the newly formed center were systematically examined. In the former allyl-substituted systems, the 1,2-*anti* product predominates in ratios greater than 10 : 1 for all examples tested. In the latter, propargyl-substituted systems, nearly absolute *syn*-1,3-stereocontrol is observed for all entries, in both geometric isomers of the product. For both substitution patterns, 2D NOESY NMR experiments were conducted to establish these relative stereochemistries. Functional groups accommodated by the title transformation include F (**42**), Br (**22**), OMe (**36–40**), CF_3 (**24**), OCH_2O (**52**), N-Ts (**4**), NCOCF_3 (**6**), CO_2Et (**8**, **10**) and SR (**12**) (see Tables 1–3). Simple alkyl substituents, such as Me (**16**, **26**) and the sterically demanding *i*-Pr (**18**, **28**) are well tolerated at both the allylic and propargylic positions. A positional survey was conducted with the methoxy substituent in the propargylic series, with *o*-, *m*- and *p*-OMe (**36–40**) substrates all giving highly efficient and stereocontrolled cyclization. Heterocycles examined include 2- (**50**) and 3-furyl (**46**), 3-thiophenyl (**48**) and 1,4-dioxane (**54**) systems. Fused systems are also tolerated, including 1-naphthyl (**34**) and benzodioxane (**54**), and the interesting, more highly extended, biaryl systems **44** and **54** also undergo efficient cyclization.

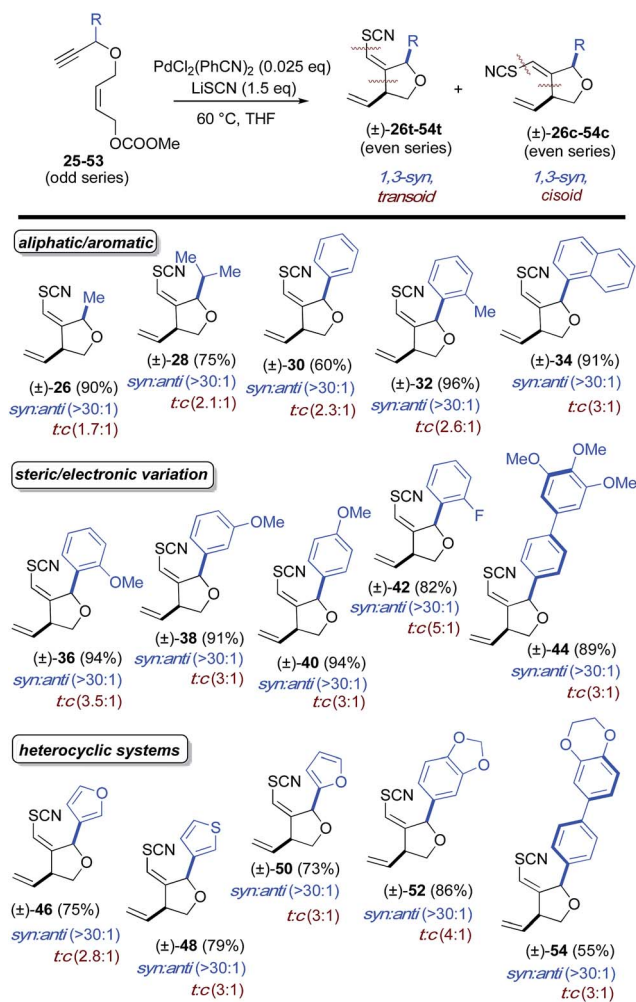
Selectivity toward the transoid alkene is seen throughout. Most notably, the allyl-substituted systems proceed with a *t* : *c* ratio greater than 20 : 1 while a more modest transoid preference is seen in the propargyl-substituted systems, ranging from 1.7 : 1 to 5 : 1. No reaction was observed for $R = t\text{Bu}$ in the propargylic position, possibly due to steric encumbrance of thiocyanate approach *anti* to Pd(II), although initial metal-coordination could also be retarded in this system. From

Table 2 Allylic substitution in the new thiocyanocarbocyclization: nearly absolute 1,2-*anti*-stereoinduction



^a Reaction conditions: 2.5 mol% of $\text{PdCl}_2(\text{PhCN})_2$ and 1.5 eq. of LiSCN. 5 mol% of $\text{PdCl}_2(\text{PhCN})_2$ were employed for substrates **15**, **17**, **19**. Ratios were determined by crude ^1H NMR. Ratios >30 : 1 assume an NMR detection limit of approximately 3%; *anti*:*syn* ratios were determined by crude NMR for the major alkene formed.

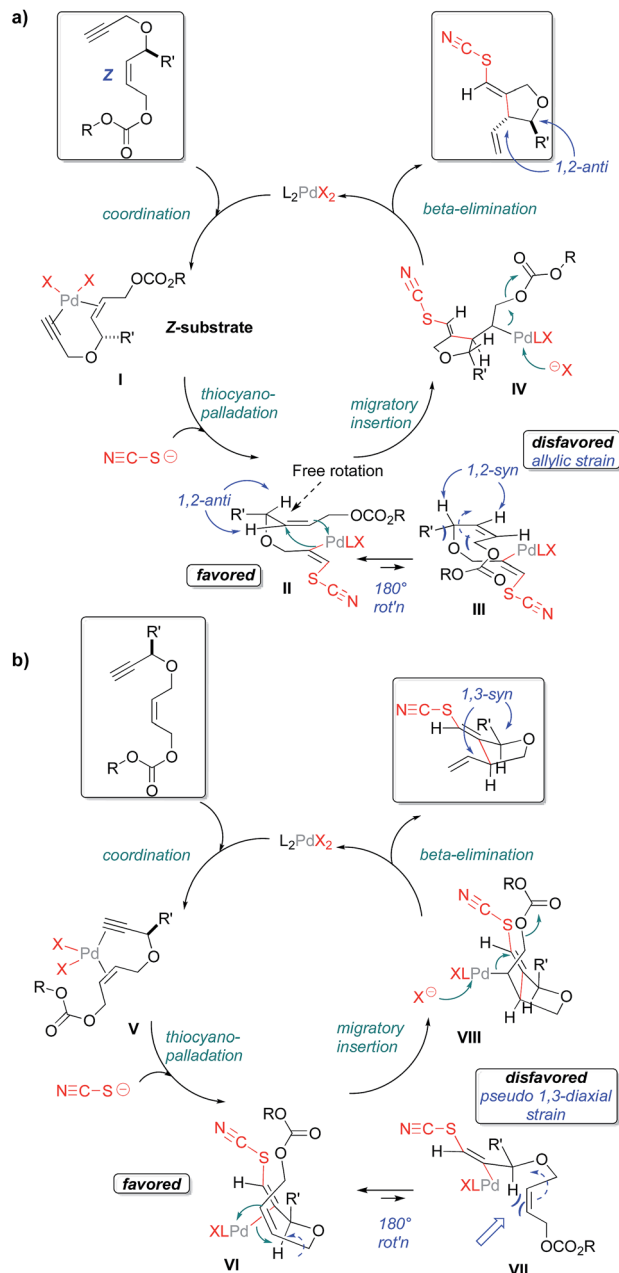


Table 3 Propargylic substitution in the new thiocyanocarbocyclization: nearly absolute 1,3-syn-stereinduction

^a Reaction conditions: 2.5 mol% of $\text{PdCl}_2(\text{PhCN})_2$ and 1.5 eq. of LiSCN . Ratios were determined by crude ^1H NMR. Ratios $>30:1$ assume an NMR detection limit of approximately 3%; anti:syn ratios were determined by crude NMR for the major alkene formed.

a mechanistic point of view the *p*-bromophenyl example (22) is intriguing. As will be seen, the mechanism proposed here employs a Pd(II) species throughout. The absence of products resulting from oxidative addition into the $\text{C}(\text{sp}^2)\text{-Br}$ bond in 22 is consistent with this mechanism. Moreover, the ability to carry aryl bromide functionality through this new thiocyanocarbocyclization opens up opportunities for additional elaboration of the cyclization products *via* cross-coupling reactions.

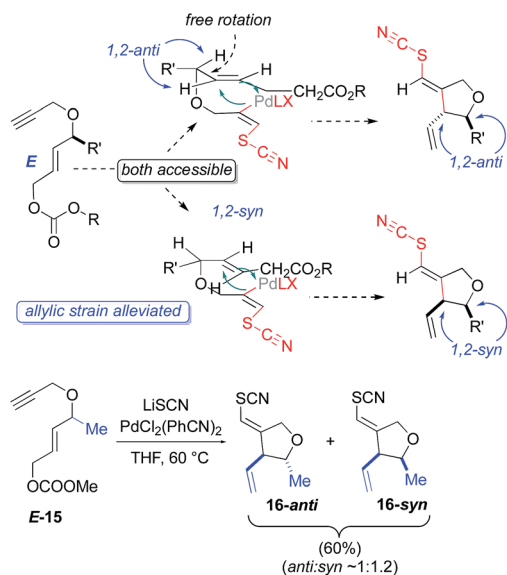
Scheme 1 depicts the mechanism that we currently favor for this thiocyanopalladation/carbocyclization transformation; one that appears to be consistent with the high levels of 1,2- and 1,3-stereinduction observed. For clarity, the mechanism is illustrated separately for substrates bearing allylic substitution in panel A and for substrates bearing propargylic substitution in panel B. As would be expected for an enyne cyclization, initial coordination of palladium to the 1,6-enyne is proposed,

**Scheme 1** Proposed mechanism for the thiocyanopalladation/carbocyclization.

followed by *anti*-thiocyanopalladation of the triple bond to give vinylpalladium intermediates II and VI, respectively. External nucleophilic delivery of thiocyanate to a Pd(II) -activated alkyne is in accordance with preponderance of transoid products seen across most test substrates. That said, note that Zhang *et al.* have reported internal halide delivery from a metal center in the case of rhodium-catalyzed cycloisomerizations. These reactions generally give cisoid halovinyl products.²³ As has already been noted, such competing mechanisms may be operative for the S-bridged substrate 11.

As an independent test of this mechanism, an enyne substrate bearing an *E*-configured allylic carbonate was prepared (Scheme 2) and subjected to the cyclization. In this





Scheme 2 Starting alkene geometry strongly influences relative stereochemistry.

case, a mixture of *anti*- and *syn*-1,2 products (*syn:anti* 1.2 : 1) is observed. This is to be expected, as the *E*-alkene geometry opens up a possible migratory insertion transition state leading to the 1,2-*syn*-configured oxacycle, as well as that leading to the previously seen 1,2-*anti*-configured oxacycle. This is because the *E*-olefin geometry in the cyclization substrate relieves the allylic strain inherent in the 1,2-*syn*-leading transition state for the *Z*-configured substrate (Scheme 2). The fact that the *Z*- and *E*-configured allylic carbonates give such different product distributions also

argues against mechanisms proceeding through rapidly equilibrating π -allyl intermediates.

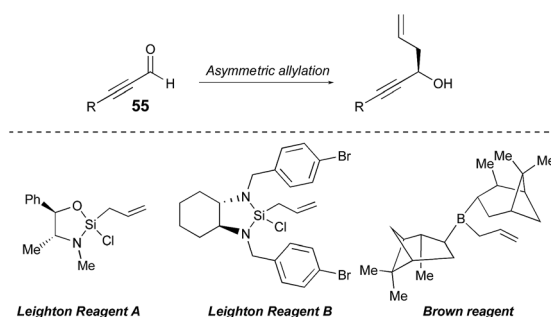
Stereocontrolled entry into NP core structures

We next set out to exploit the *syn*-1,3-selectivity observed for propargyl-substituted substrates to develop a streamlined entry into a functionalizable scaffold that maps onto the bridged, bicyclic core of the natural products (NPs) annuonone A²⁴ and massarilactone G²⁵ (Table 4 and Scheme 3). The plan here was to carry a terminal 'spectator alkene' into the new cyclization reaction, thereby positioning this alkene for a post-cyclization ring closing metathesis (RCM). There is interest in the annuonones as these NPs display allelopathic properties,²⁶ whereas massarilactones have shown neuraminidase inhibition activity²⁷ and some anti-cancer activity, *in vitro*.²⁸ This approach was motivated by the work of Waldmann²⁹ and Stockwell,³⁰ both of whom have argued for the 'privileged' nature of NP core structures when generating unnatural compound libraries for chemical biology.

Setting the absolute stereochemistry *via* asymmetric allylation

In this endeavor it was envisioned that the absolute stereochemistry could be set *via* asymmetric allylation of silyl propionaldehyde (55). There was some precedent for such an approach, albeit with modest success, utilizing either Soderquist's chiral B-allyl-borabicyclo[3.3.2]decane reagent³¹ (76%, 77% ee)³² or the Duthaler–Hafner allyl titanium TADDOLate reagent³³ (62%, 85% ee).³² Accordingly, a range of alternative, asymmetric allylation chemistries were examined here with variation of the metal, the chiral element, the silyl protecting group and the temperature (Table 4). Absolute configuration was determined using reported optical rotation where possible

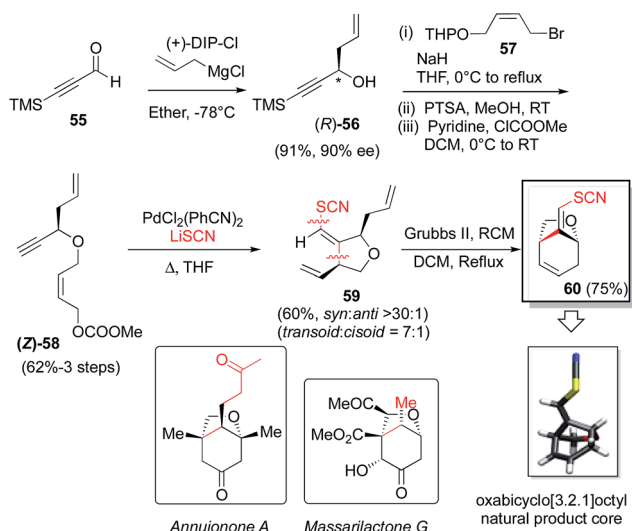
Table 4 Asymmetric allylation of the reactive silylated propynal substrate



Entry	R	Conditions	Yield	ee ^a
1	TMS	(<i>R</i>)-BINOL/Ti(OiPr) ₄ (10 mol%) tri- <i>n</i> -butylallylstannane, DCM, −20 °C	25%	+27%
2	TMS	(−)-DIP-OMe, allylmagnesium chloride, ether, −100 °C	85%	−64%
3	TMS	(−)-DIP-Cl, allylmagnesium chloride, ether, −78 °C	91%	−90%
4	TMS	Leighton reagent A, DCM, 0 °C	61%	+65%
5	TMS	Leighton reagent B, DCM, 0 °C	35%	+88%
6	TBS	Leighton reagent B, DCM, 0 °C	20%	+92%
7	TBS	(−)-DIP-Cl, allylmagnesium chloride, ether, −78 °C	77%	−90%

^a Sign translates to the optical rotation of the major enantiomer obtained.





Scheme 3 Rapid stereoselective access to oxabicyclo[3.2.1]octyl natural product cores. The new cyclization reaction affords core structures outfitted with a versatile thiocyanate functionality as a bio-orthogonal probe in and of itself or as a template for subsequent functionalization.

and ultimately confirmed *via* X-ray crystallography upon the eventual NP core structure.

Generation of the Keck-type allyltitanium species *via* *trans*-metallation from the allyl stannane *in situ* with allyl stannane gave low conversions and low ee as well (entry 1).³⁴ Preparation of allyl diisopinocampheylborane Brown reagent from (–)-DIP-OMe and (–)-DIP-Cl and allylmagnesium chloride showed promise (entries 2 and 3). We then turned our attention to the first and second generation allylsilane reagents developed by Leighton from pseudoephedrine³⁵ and *trans*-1,2-diaminocyclohexane,³⁶ respectively. These reagents presumably act *via* ‘strain-release Lewis acidity’ as first articulated by Denmark.³⁷ In the event, we were able to achieve 88% ee and 92% ee by pairing the TMS- and TBS-protected propionaldehyde substrates, respectively, with Leighton reagent **B** (entries 5 and 6). However, very modest yields were observed in both cases ($\leq 35\%$) and moving to TBS-protected propionaldehyde with DIP-Cl and allyl magnesium chloride did not improve the ee. Therefore, we chose to retain TMS protection. This is also synthetically expedient, as this silyl ether is cleaved in the course of the subsequent allyl bromide displacement step, presumably due to attack of the released bromide, thereby obviating an extra deprotection step. Thus, utilizing the optimal conditions found, and adjusting for the desired absolute stereochemistry, then, the Brown reagent³⁸ was generated *in situ* from (+)-DIP-Cl and allylmagnesium chloride following the protocol of Brimble³⁹ and provided the properly configured allylic alcohol **56**³² in 91% yield and 90% ee.

Next, deprotonation of alcohol **56**, followed by allylic bromide displacement, *O*-THP-deprotection and carbonate installation gave substrate **58**. Thiocyanopalladation/carbocyclization then furnished the desired *syn*-1,3-tetrahydrofuranoid system, **59** which, upon Grubbs II-catalyzed ring-closing metathesis,

provided the oxabicyclo[3.2.1]octyl NP core **60**. The absolute stereochemistry of **60** was confirmed by X-ray crystallography utilizing anomalous dispersion with sulfur serving as the requisite heavy atom (see ESI† for details).

Tailoring chemistry – opportunities for DOS

The new reaction modality reported here for introduction of the vinyl thiocyanate functionality into potential protein ligands represents a valuable tool for chemical biology. There has been great interest in exploiting the SCN functional group as a bio-orthogonal probe of active site electrostatic environment *via* vibrational Stark effect studies⁴⁰ as described by Boxer⁴¹ and others⁴² [for an interesting recent example combining the GFP fluorophore with a Stark effect IR probe, see ref. 43]. A particularly elegant recent example from Hammes-Schiffer and Benkovic utilizes a surgically embedded SCN reporter group in DHFR to demonstrate how the active site microenvironment subtly changes dielectric as the enzyme proceeds along the reaction coordinate for dihydrofolate reduction.²⁸ The title Pd(II)-mediated transformation should allow practitioners to place SCN IR-reporter groups into ligand scaffolds as well, as has been done for nitrile functional groups.⁴⁴

In addition to being a useful bioorthogonal probe for such vibrational Stark effect studies, the thiocyanate moiety is also an outstanding platform for diversity oriented synthesis (DOS). Thus, as is demonstrated herein, each library member bearing an SCN can be selectively diversified by tapping into the unique reactivity of this underexploited functional group. Briefly, the value of such diversification *via* SCN-tailoring chemistry is significant given current interest in methods to rapidly access novel chemical space.^{45–47}

In the event, the thiocyanate functionality could be utilized as a vehicle to generate aromatic or aliphatic thioethers by condensation with the appropriate Grignard reagent (Scheme 4). These reactions proceeded with clean S–CN cleavage; no competing conjugate addition was observed. In a particularly noteworthy example, selective magnesiation of 4-chlorobromobenzene, followed by condensation with the vinyl thiocyanate **60** providing an aromatic thioether **61**, bearing aryl chloride functionality, itself amenable to further elaboration *via* cross-coupling chemistry.⁴⁸ Reaction of **60** with isobutylmagnesium chloride and with methylenedioxyphenylmagnesium bromide, led cleanly to thioethers **62** and **63**, respectively, demonstrating the generality of this homologation chemistry. *S*-trifluoromethylation was also achieved upon treatment of **60** with TMSCF₃ and TBAF, providing a facile method for accessing the interesting vinyl trifluoromethyl thioether functionality (**64**).⁴⁹ In an alternative approach to ‘cleaving’ the SCN group, one may ‘split’ the SCN moiety through the cyanothiolation of alkynes under Pd catalysis. In this manner, vinyl thiocyanate **60** was successfully added across 1-hexyne⁵⁰ to regioselectively provide tri-substituted alkene **65**. Note that, in our hands, this cyanothiolation reaction shows a strong ligand dependence, as investigated by comparing the reactivity of various Pd₂dba₃-ligand combinations. It was observed that whereas PPh₃, Ph₂P(CH₂)₃PPh₂ (DPPP) and Pfu₃ all support this chemistry well, AsPh₃ gives greatly attenuated reactivity and the P(C₆F₅)₃ ligand





The ability to install an SCN functional group onto complex natural product-like ligand scaffolds is important as this will provide chemical biologists with a new tool for probing active site environments. The thiocyanate group provides an ideal spectroscopic window for vibrational Stark effect studies that are very sensitive to local electrostatic fields in the environment surrounding the functional group. This technique has been used to study protein ligand interactions, most notably by the groups of Boxer,^{41,44,60} Hammes-Schiffer,^{28,42c} Londergan^{42a,42b} and Webb.^{43,61} The most common approach is to generate an active site S-CN functionality by cyanating a cysteine residue with CN-Br. The complementary experiment, wherein the small molecule ligand carries the infrared probe is much rarer; we are

aware of an example in which the nitrile functionality on an inhibitor of human aldose reductase was used to probe key substrate–ligand interactions.⁴⁴ The example utilized a cyano substituent built into the aldose reductase inhibitor. Similar experiments with thiocyanate-bearing ligands are expected to be forthcoming in the future; the title transformation is expected to promote such studies by providing a streamlined entry into functionally dense ligand arrays bearing the SCN moiety, in short order and with control of stereochemistry. The potential for such ligand-based vibrational Stark effect experiments is high, as the observed SCN IR frequency is highly sensitive to the hydration sphere around the SCN functionality and to the hydrogen-bonding environment of the ligand.

Beyond their utility as an IR- or Raman-probes,⁴⁰ in medicinal chemistry circles, thiocyanates have served as synthetic intermediates to useful drug-like heterocyclic scaffolds, particularly fused cyclic thioureas⁶² and substituted benzothiazoles^{61a,63} such as the 2-amino-benzothiazole-based peptidomimetic oligomers recently described by Hamilton that disrupt amyloid peptide fibrillation.⁶⁴ In this study, we have highlighted the chemical versatility of the thiocyanate moiety, by generating an array of structural variants from a common NP core structure (Scheme 4),^{29,30} an approach that clearly holds promise for DOS applications.^{45,46,65} In surveying the literature, it seems clear that the thiocyanate moiety is an under-utilized functionality for chemical diversification in library development.^{45,46,65,66} In this regard, the new, versatile and stereoselective thiopalladation/carbocyclization transformation described herein will likely open up new vistas to the chemical biology community for both ligand-centered biophysical studies, and for exploiting the rich chemical potential of the SCN moiety.

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Author contributions

G. M., R. A. S., V. T. K., X. F. and G. A. A. performed the research; all authors analyzed and interpreted the results; D. B. B., G. M. and R. A. S. wrote the manuscript while V. T. K., X. F. and G. A. A. contributed to compiling the ESI.

Conflict of interests

The authors declare that they have no competing financial interests.

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Notes and references

- (a) C. Shen, P. Zhang, Q. Sun, S. Bai, T. S. A. Hor and X. Liu, *Chem. Soc. Rev.*, 2015, **44**, 291–314; (b) M. A. Fernandez-Rodriguez and J. F. Hartwig, *Chem.–Eur. J.*, 2010, **16**, 2355–2359; (c) M. A. Fernandez-Rodriguez and J. F. Hartwig, *J. Org. Chem.*, 2009, **74**, 1663–1672; (d) J. F. Hartwig, *Acc. Chem. Res.*, 2008, **41**, 1534–1544; (e) M. A. Fernandez-Rodriguez, Q. Shen and J. F. Hartwig, *J. Am. Chem. Soc.*, 2006, **128**, 2180–2181.
- (a) N. Zhu, F. Wang, P. Chen, J. Ye and G. Liu, *Org. Lett.*, 2015, **17**, 3580–3583; (b) Z. Liang, F. Wang, P. Chen and G. Liu, *Org. Lett.*, 2015, **17**, 2438–2441.
- F. Wang, X. Yu, Z. Qi and X. Li, *Chem.–Eur. J.*, 2016, **22**, 511–516.
- (a) N. Gao, S. Zheng, W. Yang and X. Zhao, *Org. Lett.*, 2011, **13**, 1514–1516; (b) I. P. Beletskaya and V. P. Ananikov, *Chem. Rev.*, 2011, **111**, 1596–1636; (c) T. Kondo and T.-a. Mitsudo, *Chem. Rev.*, 2000, **100**, 3205–3220.
- D. V. Sokol'skii and A. Y. Matveichuk, *Izv. Akad. Nauk Kaz. SSR, Ser. Khim.*, 1967, **17**, 39–43.
- F. Ke, Y.-Y. Qu, Z.-Q. Jiang, Z.-K. Li, D. Wu and X.-G. Zhou, *Org. Lett.*, 2011, **13**, 454–457.
- (a) S. P. France, S. Hussain, A. M. Hill, L. J. Hepworth, R. M. Howard, K. R. Mulholland, S. L. Flitsch and N. J. Turner, *ACS Catal.*, 2016, **6**, 3753–3759; (b) T. Sehl, H. C. Hailes, J. M. Ward, R. Wardenga, E. von Lieres, H. Offermann, R. Westphal, M. Pohl and D. Rother, *Angew. Chem., Int. Ed.*, 2013, **52**, 6772–6775.
- N. J. Turner and E. O'Reilly, *Nat. Chem. Biol.*, 2013, **9**, 285–288.
- (a) C. K. Savile, J. M. Janey, E. C. Mundorff, J. C. Moore, S. Tam, W. R. Jarvis, J. C. Colbeck, A. Krebber, F. J. Fleitz, J. Brands, P. N. Devine, G. W. Huisman and G. J. Hughes, *Science*, 2010, **329**, 305–309; (b) S. Lutz, *Science*, 2010, **329**, 285–287; (c) U. Arnold, M. P. Hinderaker, B. L. Nilsson, B. R. Huck, S. H. Gellman and R. T. Raines, *J. Am. Chem. Soc.*, 2002, **124**, 8522–8523.
- (a) Y. Okamoto, V. Kohler and T. R. Ward, *J. Am. Chem. Soc.*, 2016, **138**, 5781–5784; (b) T. K. Hyster, L. Knoerr, T. R. Ward and T. Rovis, *Science*, 2012, **338**, 500–503.
- (a) J. B. Siegel, A. Zanghellini, H. M. Lovick, G. Kiss, A. R. Lambert, J. L. St. Clair, J. L. Gallaher, D. Hilvert, M. H. Gelb, B. L. Stoddard, K. N. Houk, F. E. Michael and D. Baker, *Science*, 2010, **329**, 309–313; (b) M. M. Muller, M. A. Windsor, W. C. Pomerantz, S. H. Gellman and D. Hilvert, *Angew. Chem., Int. Ed.*, 2009, **48**, 922–925; (c) L. Jiang, E. A. Althoff, F. R. Clemente, L. Doyle, D. Roethlisberger, A. Zanghellini, J. L. Gallaher, J. L. Betker, F. Tanaka, C. F. Barbas III, D. Hilvert, K. N. Houk, B. L. Stoddard and D. Baker, *Science*, 2008, **319**, 1387–1391.
- (a) K. Panigrahi, G. A. Applegate, G. Malik and D. B. Berkowitz, *J. Am. Chem. Soc.*, 2015, **137**, 3600–3609;



- (b) H. Groeger and W. Hummel, *Curr. Opin. Chem. Biol.*, 2014, **19**, 171–179.
- 13 (a) M. Shevlin, *ACS Med. Chem. Lett.*, 2017, **8**, 601–607; (b) A. Buitrago Santanilla, E. L. Regalado, T. Pereira, M. Shevlin, K. Bateman, L.-C. Campeau, J. Schneeweis, S. Bertritt, Z.-C. Shi, P. Nantermet, Y. Liu, R. Helmy, C. J. Welch, P. Vachal, I. W. Davies, T. Cernak and S. D. Dreher, *Science*, 2015, **347**, 49–53; (c) J. L. Treece, J. R. Goodell, D. Vander Velde, J. A. Porco Jr and J. Aube, *J. Org. Chem.*, 2010, **75**, 2028–2038; (d) E. Wolf, E. Richmond and J. Moran, *Chem. Sci.*, 2015, **6**, 2501–2505.
- 14 (a) M. Teders, L. Pitzer, S. Buss and F. Glorius, *ACS Catal.*, 2017, **7**, 4053–4056; (b) D. S. Mannel, M. S. Ahmed, T. W. Root and S. S. Stahl, *J. Am. Chem. Soc.*, 2017, **139**, 1690–1698; (c) W. Bentley Keith, P. Zhang and C. Wolf, *Sci. Adv.*, 2016, **2**, e1501162; (d) K. D. Collins, T. Gensch and F. Glorius, *Nat. Chem.*, 2014, **6**, 859–871; (e) J. R. Cabrera-Pardo, D. I. Chai, S. Liu, M. Mrksich and S. A. Kozmin, *Nat. Chem.*, 2013, **5**, 423–427.
- 15 (a) A. Hamberg, S. Lundgren, E. Wingstrand, C. Moberg and K. Hult, *Chem.–Eur. J.*, 2007, **13**, 4334–4341; (b) A. Hamberg, S. Lundgren, M. Penhoat, C. Moberg and K. Hult, *J. Am. Chem. Soc.*, 2006, **128**, 2234–2235; (c) C. M. Sprout and C. T. Seto, *Org. Lett.*, 2005, **7**, 5099–5102.
- 16 (a) D. B. Berkowitz and G. Maiti, *Org. Lett.*, 2004, **6**, 2661–2664; (b) D. B. Berkowitz, M. Bose and S. Choi, *Angew. Chem., Int. Ed.*, 2002, **41**, 1603–1607.
- 17 (a) K. R. Karukurichi, X. Fei, R. A. Swyka, S. Broussy, W. Shen, S. Dey, S. K. Roy and D. B. Berkowitz, *Sci. Adv.*, 2015, **1**(6), e1500066; (b) S. Dey, D. R. Powell, C. Hu and D. B. Berkowitz, *Angew. Chem., Int. Ed.*, 2007, **46**, 7010–7014; (c) S. Dey, K. R. Karukurichi, W. Shen and D. B. Berkowitz, *J. Am. Chem. Soc.*, 2005, **127**, 8610–8611.
- 18 (a) S. K. Ginotra, J. A. Friest and D. B. Berkowitz, *Org. Lett.*, 2012, **14**, 968–971; (b) J. A. Friest, S. Broussy, W. J. Chung and D. B. Berkowitz, *Angew. Chem., Int. Ed.*, 2011, **50**, 8895–8899.
- 19 (a) C. D. McCune, M. L. Beio, J. M. Sturdivant, R. de la Salud-Bea, B. M. Darnell and D. B. Berkowitz, *J. Am. Chem. Soc.*, 2017, **139**, DOI: 10.1021/jacs.7b04690; (b) K. R. Karukurichi, R. de la Salud-Bea, W. J. Jahng and D. B. Berkowitz, *J. Am. Chem. Soc.*, 2007, **129**, 258–259; (c) D. B. Berkowitz, R. de la Salud-Bea and W.-J. Jahng, *Org. Lett.*, 2004, **6**, 1821–1824.
- 20 (a) For pioneering work on Pd(II)-mediated carbocyclization reactions that inspired these studies, see: W. Xu, A. Kong and X. Lu, *J. Org. Chem.*, 2006, **71**, 3854–3858; (b) L. Zhao, X. Lu and W. Xu, *J. Org. Chem.*, 2005, **70**, 4059–4063; (c) X. Xie, X. Lu, Y. Liu and W. Xu, *J. Org. Chem.*, 2001, **66**, 6545–6550; (d) X. Lu, G. Zhu, Z. Wang, S. Ma, J. Ji and Z. Zhang, *Pure Appl. Chem.*, 1997, **69**, 553–558; (e) H. Jiang, S. Ma, G. Zhu and X. Lu, *Tetrahedron*, 1996, **52**, 10945–10954; (f) X. Lu, S. Ma, J. Ji, G. Zhu and H. Jiang, *Pure Appl. Chem.*, 1994, **66**, 1501–1508.
- 21 G. R. Cook and R. Hayashi, *Org. Lett.*, 2006, **8**, 1045–1048.
- 22 C. S. Wannere and P. v. R. Schleyer, *Org. Lett.*, 2003, **5**, 605–608.
- 23 X. Tong, D. Li, Z. Zhang and X. Zhang, *J. Am. Chem. Soc.*, 2004, **126**, 7601–7607.
- 24 F. A. Macías, R. M. Varela, A. Torres, R. M. Oliva and J. G. Molinillo, *Phytochemistry*, 1998, **48**, 631–636.
- 25 H. Oh, D. C. Swenson, J. B. Gloer and C. A. Shearer, *Tetrahedron Lett.*, 2001, **42**, 975–977.
- 26 (a) F. A. Macías, A. López, R. M. Varela, A. Torres and J. M. G. Molinillo, *Phytochemistry*, 2004, **65**, 3057–3063; (b) T. Anjum and R. Bajwa, *Phytochemistry*, 2005, **66**, 1919–1921.
- 27 G. F. Zhang, W. B. Han, J. T. Cui, S. W. Ng, Z. K. Guo, R. X. Tan and H. M. Ge, *Planta Med.*, 2012, **78**, 76–78.
- 28 S. Chen, F. Ren, S. Niu, X. Liu and Y. Che, *J. Nat. Prod.*, 2014, **77**, 9–14.
- 29 (a) R. Narayan, M. Potowski, Z.-J. Jia, A. P. Antonchick and H. Waldmann, *Acc. Chem. Res.*, 2014, **47**, 1296–1310; (b) H. Lachance, S. Wetzler, K. Kumar and H. Waldmann, *J. Med. Chem.*, 2012, **55**, 5989–6001.
- 30 M. E. Welsch, S. A. Snyder and B. R. Stockwell, *Curr. Opin. Chem. Biol.*, 2010, **14**, 347–361.
- 31 E. Canales, K. G. Prasad and J. A. Soderquist, *J. Am. Chem. Soc.*, 2005, **127**, 11572–11573.
- 32 S. B. Kamptmann and R. Brückner, *Eur. J. Org. Chem.*, 2013, **2013**, 6584–6600.
- 33 A. Hafner, R. O. Duthaler, R. Marti, G. Rihs, P. Rothe-Streit and F. Schwarzenbach, *J. Am. Chem. Soc.*, 1992, **114**, 2321–2336.
- 34 G. E. Keck, K. H. Tarbet and L. S. Geraci, *J. Am. Chem. Soc.*, 1993, **115**, 8467–8468.
- 35 X. Wang, Q. Meng, A. J. Nation and J. L. Leighton, *J. Am. Chem. Soc.*, 2002, **124**, 10672–10673.
- 36 (a) L. M. Suen, M. L. Steigerwald and J. L. Leighton, *Chem. Sci.*, 2013, **4**, 2413–2417; (b) W. A. Chalifoux, S. K. Reznik and J. L. Leighton, *Nature*, 2012, **487**, 86–89; (c) X. Zhang, K. N. Houk and J. L. Leighton, *Angew. Chem., Int. Ed.*, 2005, **44**, 938–941.
- 37 S. E. Denmark, R. T. Jacobs, G. Dai-Ho and S. Wilson, *Organometallics*, 1990, **9**, 3015–3019.
- 38 (a) H. C. Brown and P. V. Ramachandran, *Pure Appl. Chem.*, 1994, **66**, 201–212; (b) U. S. Racherla and H. C. Brown, *J. Org. Chem.*, 1991, **56**, 401–404.
- 39 M. A. Brimble, P. Bachu and J. Sperry, *Synthesis*, 2007, **2007**, 2887–2893.
- 40 H. Kim and M. Cho, *Chem. Rev.*, 2013, **113**, 5817–5847.
- 41 (a) A. T. Fafarman, L. J. Webb, J. I. Chuang and S. G. Boxer, *J. Am. Chem. Soc.*, 2006, **128**, 13356–13357; (b) P. A. Sigala, A. T. Fafarman, P. E. Bogard, S. G. Boxer and D. Herschlag, *J. Am. Chem. Soc.*, 2007, **129**, 12104–12105.
- 42 (a) L. Edelstein, M. A. Stetz, H. A. McMahon and C. H. Londergan, *J. Phys. Chem. B*, 2010, **114**, 4931–4936; (b) H. A. McMahon, K. N. Alfieri, K. A. A. Clark and C. H. Londergan, *J. Phys. Chem. Lett.*, 2010, **1**, 850–855; (c) J. P. Layfield and S. Hammes-Schiffer, *J. Am. Chem. Soc.*, 2013, **135**, 717–725.
- 43 J. D. Slocum and L. J. Webb, *J. Am. Chem. Soc.*, 2016, **138**, 6561–6570.
- 44 L. Xu, A. E. Cohen and S. G. Boxer, *Biochemistry*, 2011, **50**, 8311–8322.



- 45 S. L. Schreiber, J. D. Kotz, M. Li, J. Aubé, C. P. Austin, J. C. Reed, H. Rosen, E. L. White, L. A. Sklar, C. W. Lindsley, B. R. Alexander, J. A. Bittker, P. A. Clemons, A. de Souza, M. A. Foley, M. Palmer, A. F. Shamji, M. J. Wawer, O. McManus, M. Wu, B. Zou, H. Yu, J. E. Golden, F. J. Schoenen, A. Simeonov, A. Jadhav, M. R. Jackson, A. B. Pinkerton, T. D. Y. Chung, P. R. Griffin, B. F. Cravatt, P. S. Hodder, W. R. Roush, E. Roberts, D.-H. Chung, C. B. Jonsson, J. W. Noah, W. E. Severson, S. Ananthan, B. Edwards, T. I. Oprea, P. J. Conn, C. R. Hopkins, M. R. Wood, S. R. Stauffer, K. A. Emmitte, L. S. Brady, J. Driscoll, I. Y. Li, C. R. Loomis, R. N. Margolis, E. Michelotti, M. E. Perry, A. Pillai and Y. Yao, *Cell*, 2015, **161**, 1252–1265.
- 46 (a) T. D. Davis, C. J. Gerry and D. S. Tan, *ACS Chem. Biol.*, 2014, **9**, 2535–2544; (b) D. S. Tan, *Nat. Chem. Biol.*, 2005, **1**, 74–84; (c) M. D. Burke and S. L. Schreiber, *Angew. Chem., Int. Ed.*, 2004, **43**, 46–58; (d) S. L. Schreiber, *Science*, 2000, **287**, 1964–1969.
- 47 B. D. Charette, R. G. MacDonald, S. Wetzler, D. B. Berkowitz and H. Waldmann, *Angew. Chem., Int. Ed.*, 2006, **45**, 7766–7770.
- 48 (a) J. Tang, A. Biafora and L. J. Goossen, *Angew. Chem., Int. Ed.*, 2015, **54**, 13130–13133; (b) T. Iwai, T. Harada, K. Hara and M. Sawamura, *Angew. Chem., Int. Ed.*, 2013, **52**, 12322–12326; (c) R. J. Lundgren, B. D. Peters, P. G. Alsabeh and M. Stradiotto, *Angew. Chem., Int. Ed.*, 2010, **49**, 4071–4074.
- 49 (a) T. Billard, S. Large and B. R. Langlois, *Tetrahedron Lett.*, 1997, **38**, 65–68; (b) M.-N. Bouchu, S. Large, M. Steng, B. Langlois and J.-P. Praly, *Carbohydr. Res.*, 1998, **314**, 37–45.
- 50 (a) I. Kamiya, J.-i. Kawakami, S. Yano, A. Nomoto and A. Ogawa, *Organometallics*, 2006, **25**, 3562–3564; (b) M. Pawliczek, L. K. B. Garve and D. B. Werz, *Org. Lett.*, 2015, **17**, 1716–1719.
- 51 S. Bhattacharya and P. K. Vemula, *J. Org. Chem.*, 2005, **70**, 9677–9685.
- 52 S. Otsuki, S. Nishimura, H. Takabatake, K. Nakajima, Y. Takasu, T. Yagura, Y. Sakai, A. Hattori and H. Kakeya, *Bioorg. Med. Chem. Lett.*, 2013, **23**, 1608–1611.
- 53 B. M. Trost and M. Lautens, *J. Am. Chem. Soc.*, 1985, **107**, 1781–1783.
- 54 (a) J. Song, Q. Shen, F. Xu and X. Lu, *Tetrahedron*, 2007, **63**, 5148–5153; (b) Q. Zhang and X. Lu, *J. Am. Chem. Soc.*, 2000, **122**, 7604–7605; (c) G. Zhu and X. Lu, *Organometallics*, 1995, **14**, 4899–4904.
- 55 (a) X. Tong, Z. Zhang and X. Zhang, *J. Am. Chem. Soc.*, 2003, **125**, 6370–6371; (b) A. Lei, M. He and X. Zhang, *J. Am. Chem. Soc.*, 2002, **124**, 8198–8199; (c) P. Cao, B. Wang and X. Zhang, *J. Am. Chem. Soc.*, 2000, **122**, 6490–6491.
- 56 O. Jackowski, J. Wang, X. Xie, T. Ayad, Z. Zhang and V. Ratovelomanana-Vidal, *Org. Lett.*, 2012, **14**, 4006–4009.
- 57 L. L. Welbes, T. W. Lyons, K. A. Cychosz and M. S. Sanford, *J. Am. Chem. Soc.*, 2007, **129**, 5836–5837.
- 58 (a) M.-C. P. Yeh, H.-F. Chen, Y.-Y. Huang and Y.-T. Weng, *J. Org. Chem.*, 2015, **80**, 10892–10903; (b) A. Saxena, F. Perez and M. J. Krische, *Angew. Chem., Int. Ed.*, 2016, **55**, 1493–1497; (c) G. M. R. Canlas and S. R. Gilbertson, *Chem. Commun.*, 2014, **50**, 5007–5010; (d) F. Grillet and K. M. Brummond, *J. Org. Chem.*, 2013, **78**, 3737–3754; (e) S. Mazumder, D. Shang, D. E. Negru, M.-H. Baik and P. A. Evans, *J. Am. Chem. Soc.*, 2012, **134**, 20569–20572; (f) A. D. Jenkins, A. Herath, M. Song and J. Montgomery, *J. Am. Chem. Soc.*, 2011, **133**, 14460–14466; (g) A. Fürstner, K. Majima, R. Martin, H. Krause, E. Kattnig, R. Goddard and C. W. Lehmann, *J. Am. Chem. Soc.*, 2008, **130**, 1992–2004; (h) M. Chen, Y. Weng, M. Guo, H. Zhang and A. Lei, *Angew. Chem.*, 2008, **120**, 2311–2314; (i) K. M. Brummond, H. Chen, B. Mitasev and A. D. Casarez, *Org. Lett.*, 2004, **6**, 2161–2163; (j) A. Lei, J. P. Waldkirch, M. He and X. Zhang, *Angew. Chem., Int. Ed.*, 2002, **41**, 4526–4529; (k) A. Lei, M. He, S. Wu and X. Zhang, *Angew. Chem., Int. Ed.*, 2002, **41**, 3457–3460.
- 59 B. M. Trost, M. C. Ryan, M. Rao and T. Z. Markovic, *J. Am. Chem. Soc.*, 2014, **136**, 17422–17425.
- 60 L. J. Webb and S. G. Boxer, *Biochemistry*, 2008, **47**, 1588–1598.
- 61 (a) M. P. Hay, S. Turcotte, J. U. Flanagan, M. Bonnet, D. A. Chan, P. D. Sutphin, P. Nguyen, A. J. Giaccia and W. A. Denny, *J. Med. Chem.*, 2010, **53**, 787–797; (b) A. J. Stafford, D. M. Walker and L. J. Webb, *Biochemistry*, 2012, **51**, 2757–2767.
- 62 (a) M. Buchholz, U. Heiser, S. Schilling, A. J. Niestroj, K. Zunkel and H.-U. Demuth, *J. Med. Chem.*, 2006, **49**, 664–677; (b) C. A. Parrish, N. D. Adams, K. R. Auger, J. L. Burgess, J. D. Carson, A. M. Chaudhari, R. A. Copeland, M. A. Diamond, C. A. Donatelli, K. J. Duffy, L. F. Faucette, J. T. Finer, W. F. Huffman, E. D. Hugger, J. R. Jackson, S. D. Knight, L. Luo, M. L. Moore, K. A. Newlander, L. H. Ridgers, R. Sakowicz, A. N. Shaw, C.-M. M. Sung, D. Sutton, K. W. Wood, S.-Y. Zhang, M. N. Zimmerman and D. Dhanak, *J. Med. Chem.*, 2007, **50**, 4939–4952.
- 63 Q. Chao, K. G. Sprankle, R. M. Grotzfeld, A. G. Lai, T. A. Carter, A. M. Velasco, R. N. Gunawardane, M. D. Cramer, M. F. Gardner, J. James, P. P. Zarrinkar, H. K. Patel and S. S. Bhagwat, *J. Med. Chem.*, 2009, **52**, 7808–7816.
- 64 H. Peacock, J. Luo, T. Yamashita, J. Luccarelli, S. Thompson and A. D. Hamilton, *Chem. Sci.*, 2016, **7**, 6435–6439.
- 65 J. E. Biggs-Houck, A. Younai and J. T. Shaw, *Curr. Opin. Chem. Biol.*, 2010, **14**, 371–382.
- 66 H. E. Pelish, N. J. Westwood, Y. Feng, T. Kirchhausen and M. D. Shair, *J. Am. Chem. Soc.*, 2001, **123**, 6740–6741.

