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Magnetic surface molecularly imprinted poly(3-aminophenylboronic acid) for selective capture and determination of diethylstilbestrol

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Diethylstilbestrol (DES) is considered a representative example of an exogenous endocrine disrupting compound (EDC). It can retard development in infants, lead to serious metabolic regulation disorders, and even result in distortion and cancer in the reproductive system. Therefore, achieving rapid and accurate analysis of trace amounts of DES in complex environments is of great importance to human health and for environmental protection. Novel magnetic molecularly imprinted polymers (MIPs) with excellent molecular recognition ability and super water-compatibility were developed for the selective capture of DES in water samples. Fe₃O₄@SiO₂ magnetic nanoparticles (NPs) were synthesized and used as support cores. Molecularly imprinted poly(3-aminophenylboronic acid) (poly(APBA)), synthesized on magnetic cores based on a surface-imprinting strategy, can preferentially bind DES molecules in water samples. The magnetic core-shell MIPs (denoted as Fe₃O₄@SiO₂@APBA/MIPs) exhibited high binding capacity and favorable recognition specificity for DES in water. The adsorption kinetics and experimental isotherm data of DES on magnetic MIPs can be well described by the pseudo-second-order kinetic model and the Langmuir isotherm, respectively. The imprinted nanoparticles were subjected to magnetic solid-phase extraction (MSPE) of DES from water samples. The DES content in the samples was determined by high-performance liquid chromatography (HPLC). The peak area increased linearly with increasing DES concentration over the range 0.08–150 μg L⁻¹, with a detection limit of 0.03 μg L⁻¹. The recoveries for spiked lake water samples were in the range 97.1–103.2%, with relative standard deviation (RSD) of 2.8–4.3% (*n* = 6).

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1. Introduction

Diethylstilbestrol (DES) is considered a representative example of an exogenous endocrine disrupting compound (EDC), which directly interferes with the endocrine function by simulating or antagonizing the normal endogenous hormones.¹ DES can be bio-accumulated in the food chain and remain in organisms for a long time, having a serious impact on the organism even at very low concentrations. Once the human body is exposed to DES, the secretion and transport of natural hormones will be destroyed. DES can retard development in infants, lead to serious metabolic regulation disorders, and even result in distortion and cancer in the reproductive system.^{1,2} In recent years, there has been a dramatic increase in the use of hormonal cosmetics and drugs, while DES has been misused in order to promote the growth of animals. Through use of liquid chromatography coupled with mass spectrometry (LC-MS), researchers have found DES residues in foods (*e.g.* eggs, meat,

and milk), as well as in the soil and water environment.³ Currently, the use of DES is banned or restricted in China, the United States, and in many countries of Europe. The development of new technologies for the detection of DES has also been a matter of international concern. Therefore, achieving rapid and accurate analysis of trace amounts of DES in complex environments is of great importance to human health and for environmental protection.

Numerous methods have been used for the detection of DES in water, including LC-MS,³ gas chromatography coupled with mass spectrometry (GC-MS),⁴ high-performance liquid chromatography (HPLC) equipped with diode-array detector (HPLC-DAD),^{5,6} immunoassay,⁷ and capillary electrochromatography.⁸ Due to the generally low concentration of DES in real environmental samples, high-performance detection using these traditional methods usually requires an efficient sample preparation step for rapid pre-concentration, such as solid-phase extraction, liquid-phase extraction, and liquid-phase micro-extraction. The main challenges associated with these techniques for DES determination are poor selectivity and low recovery.^{5,6} However, by using magnetic molecularly imprinted polymer (MMIP) particles as the solid-phase extraction agent, DES can not only be selectively extracted from water samples,

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but also separated quickly under an external magnetic field. Therefore, the pretreatment process can be performed quickly and easily.

In general, MIPs are synthesized using templates, with a suitable monomer and cross-linking agent, with an initiator to initiate the polymerization. The template molecules are then removed to create recognition cavities with many functional recognition sites. These cavities can match the size, shape, and spatial structure of the template molecule. Thus, MIPs with a specific ability for molecular recognition can selectively rebind the target. MIPs have been widely used in sensors,⁹ and for catalysis,¹⁰ separation, and purification.¹¹ Recent studies have focused on the preparation of MIPs for the enriching and detection of DES, bisphenol A (BPA), or other estrogens^{6,12,13} as well as an evaluation of MIPs toward DES in the organic phase due to their excellent adsorption properties.^{1,6,13–17} In particular, the application of MIPs in the construction of sensors with a high affinity and selectivity for the target is highly promising. Recently, a series of novel electrochemical sensors combined MIPs with various new nano-materials, and excellent performances have been reported.^{18–22} The application of MIPs for the detection of trace amounts of DES in water is particularly promising.

It is difficult to synthesize MIPs directly in the aqueous phase because the formation of hydrogen bonds between the template and the functional monomer can be easily interfered with by water molecules.²³ Furthermore, template molecules of estrogens have poor solubility in the aqueous phase. In order to improve the water-compatibility of MIPs, Wu *et al.* grafted hydrophilic 2-hydroxyethyl methacrylate brushes onto the surface of the MIPs.²⁴ Other typical methods include the use of hydrophilic functional monomers, such as α -methacrylic acid,^{1,17} 2-acrylamido-2-methylpropanesulfonic acid,^{25,26} 4-vinylpyridine,²⁵ acryloyl- β -cyclodextrin,^{27,28} and 3-aminophenylboronic acid,²⁹ in the synthesis of the MIPs. These methods are simple and can improve the surface hydrophilicity of the MIPs. As a water-soluble functional monomer, aminophenylboronic acid (APBA) can be used to prepare MIPs of DES due to the presence of multiple functional groups including amino, hydroxyl, and phenyl groups, and can be polymerized both in aqueous and organic phase solution. Thin film polymers of APBA (poly(APBA)) have been used as the coating substrate on solid supports such as polystyrene nanoparticles (NPs), microspheres, and the gold surface of quartz crystal microbalance electrodes.^{29–31}

MMIPs^{15,17} can be prepared by synthesizing MIPs on the surface of Fe₃O₄ magnetic nanoparticles. Therefore, MMIPs can not only specially capture target molecules, but can also be rapidly magnetically separated from the solution. There are many methods for the synthesis of MMIPs, such as emulsion polymerization, the sol-gel method, and suspension polymerization, for example. In general, the magnetic properties of MIP microspheres obtained by traditional emulsion polymerization are usually weak because of the nucleation of micelles leading to low encapsulation efficiency.³² The imprinted film on particles synthesized by the sol-gel method, the residue of the hydrophobic portion of the silane coupling agent, is hard to

avoid resulting in adhesion between the particles.^{26,33} However, suspension polymerization with water as the continuous phase can be used to prepare imprinted nanoparticles with a small particle size, strong magnetic properties, and good dispersion in the aqueous phase.¹²

In this study, we developed super water-soluble DES-imprinted MMIPs (Fe₃O₄@SiO₂@APBA/MIPs) with a multi-layer core-shell structure for the selective recognition and extraction of DES from the aqueous phase. APBA was used as the hydrophilic monomer and cross-linking reagent. The binding properties, including molecular binding capacity and specific recognition ability, were investigated in detail. The MIP NPs exhibited much higher binding capability for DES in water than previously reported.^{5,27} The MIPs as adsorbents were used in enriching trace DES from lake water samples by magnetic solid-phase extraction (MSPE). The preparation procedure and working principle of Fe₃O₄@SiO₂@APBA/MIPs (Fig. 1A) and their applications to MSPE-HPLC (Fig. 1B) are schematically illustrated in Fig. 1.

2. Experimental

2.1 Reagents

DES, APBA, 3-aminopropyl triethoxy silane (APTES), and tetraethyl orthosilicate (TEOS) were purchased from J&K Scientific Ltd. Glutaraldehyde (GA), FeCl₃·6H₂O, FeCl₂·4H₂O, potassium peroxydisulfate, and other reagents were obtained from Beijing Chemical Reagent Company (Beijing, China). Bisphenol A was from Chengdu Xiya Chemical Co., Ltd. (Chengdu, China). Phenol, bisphenol F, estrone, and estradiol were provided by Tianjin Chemical Reagent Co. Ltd (Tianjing, China). All reagents were of analytical grade. Acetonitrile (ACN) for HPLC was of HPLC-reagent grade and was supplied by J&K Scientific Ltd. (Beijing, China). All solutions were prepared with ultrapure water (Milli-Q Advantage A10 Water Purification System, Millipore Corporation, France). DES (100 mg) was dissolved in 100 mL of ethanol for the preparation of 1000 mg L⁻¹ of DES stock solution, and stored at 4 °C until use. DES solutions with required concentration could be diluted with ultrapure water for further use. The elution solution was a mixture of methanol-0.1 M acetic acid (5.0 mL, v/v, 9/1).

2.2 Instruments

Scanning electron microscopy (SEM) and transmission electron microscopy (TEM) images of functionalized Fe₃O₄ NPs were obtained by SU-8010 (Hitachi) and HT7700 (Hitachi), respectively. X-ray energy dispersive spectroscopy (EDS) was used to obtain the chemical composition of the samples. Fourier transform infrared (FTIR) spectra were recorded on an IR Prestige-21 FTIR spectrometer (Shimadzu). The thermal stability of the imprinted NPs was analyzed using a TG 209 F3 thermogravimetric analyzer (TGA; Netzsch, Germany) at a heating rate of 10 °C min⁻¹ under an air atmosphere. Magnetization measurements of magnetic nanoparticles were performed using a vibration sample magnetometer (VSM; Lake Shore 7410). The static water contact angles (CA) of functionalized Fe₃O₄ NPs



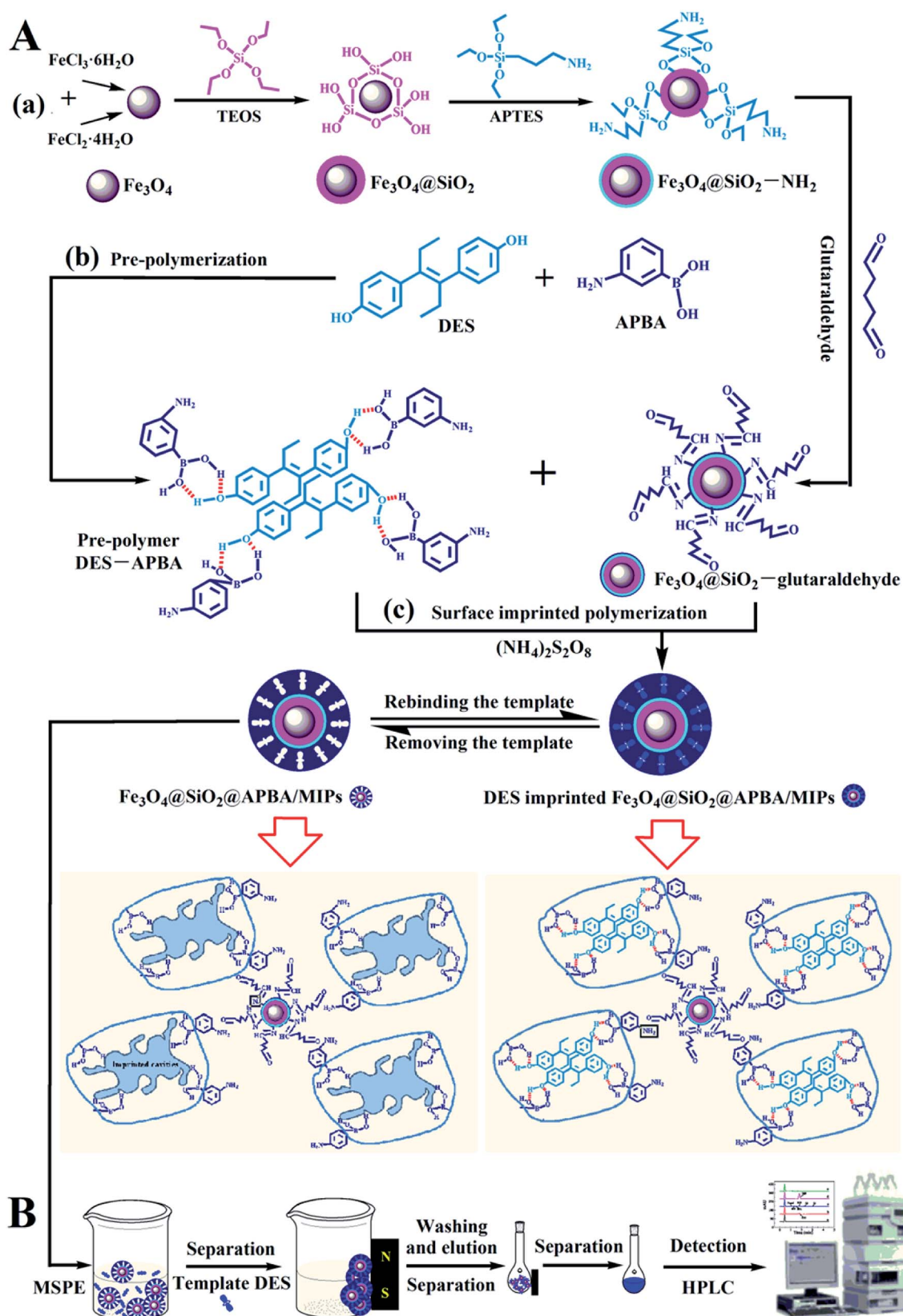


Fig. 1 Schematic procedure for (A) the preparation of DES-imprinted MIPs and (B) application to MIPs-MSPE-HPLC method for detection of DES.



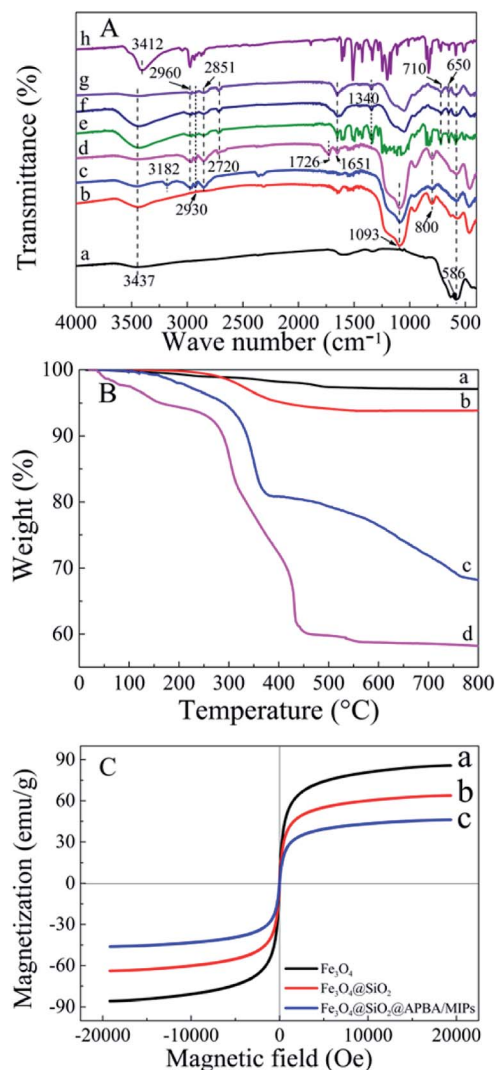


Fig. 2 (A) FTIR spectra of (a) as-prepared Fe₃O₄, (b) Fe₃O₄@SiO₂, (c) Fe₃O₄@SiO₂-NH₂, (d) glutaraldehyde-capped Fe₃O₄@SiO₂, (e) Fe₃O₄@SiO₂@APBA-DES/MIPs, (f) Fe₃O₄@SiO₂@APBA/MIPs, (g) Fe₃O₄@SiO₂@APBA/NIPs and (h) DES. (B) TGA curves of (a) Fe₃O₄, (b) Fe₃O₄@SiO₂, (c) Fe₃O₄@SiO₂@APBA/MIPs and (d) Fe₃O₄@SiO₂@APBA/NIPs. (C) VSM magnetization curves of (a) Fe₃O₄, (b) Fe₃O₄@SiO₂ and (c) Fe₃O₄@SiO₂@APBA/MIPs.

were measured using an OCA 15 Pro video optical measurement instrument of CA (Data Physic, Germany) with 2.5 μ L of deionized water droplets. Spectrophotometric experiments were carried out using a UV-2450 spectrophotometer (Shimadzu). Chromatographic analyses were performed using a Model 1260 HPLC instrument (Agilent Technologies Co., Ltd., USA), mainly equipped with a diode-array detector and a chromatographic column (150 mm \times 4.6 mm C₁₈). Optimized HPLC conditions were injection volume, 20 μ L; mobile phase, acetonitrile/ultrapure water (6 : 4, v/v); flow rate, 0.8 mL min⁻¹; temperature of the column, 25 $^{\circ}$ C; DAD detection wavelength, 240 nm.

2.3 Preparation of functionalized Fe₃O₄@SiO₂ NPs

Magnetic Fe₃O₄ NPs were synthesized according to the coprecipitation method that we reported previously.³⁴ In brief,

a mixture of FeCl₂·4H₂O (2.0 g), FeCl₃·6H₂O (5.2 g), 12 M HCl (0.85 mL), and 25 mL of water was degassed with high-pure nitrogen with stirring before use. Then, the mixed solution was added dropwise into 250 mL of 1.5 M NaOH solution in a water bath at 80 $^{\circ}$ C, and was stirred for 1 h under N₂ protection. After cooling down, the obtained Fe₃O₄ NPs were washed repeatedly five times with water and ethanol, and then collected magnetically, before being dried under an N₂ atmosphere.

Fe₃O₄@SiO₂ NPs functionalized with amino-groups were prepared based the Stöber process,^{24,35,36} with minor modifications. As-prepared Fe₃O₄ NPs (100 mg) were dispersed in a mixture of ethanol and ultrapure water (180 mL, 8 : 1, v/v) and ultrasonicated for 15 min. Then, 1.0 mL of ammonia aqueous solution (28%, w/w) was added into the suspension under vigorous stirring for 30 min in a water bath at 30 $^{\circ}$ C. After adding 1 mL of TEOS dropwise, the reaction proceeded continuously for 45 min; then, 0.5 mL amino-propyltriethoxysilane (APTES) was added dropwise into the suspension. The reaction between Fe₃O₄@SiO₂ NPs and APTES lasted for 4 h at 30 $^{\circ}$ C. The obtained Fe₃O₄@SiO₂-NH₂ NPs were collected using a magnet and washed with ethanol and ultrapure water three times, followed by drying under nitrogen gas protection overnight.

Fe₃O₄@SiO₂-NH₂ NPs were modified with glutaraldehyde¹² as the bridging agent to introduce free aldehyde groups for further covalent anchoring of MIPs grafted tightly on the surface of the support substrates. Briefly, 50 mg of Fe₃O₄@SiO₂-NH₂ NPs was dispersed in 50 mL of excess glutaraldehyde aqueous solution (5%, v/v) with slow stirring to form a homogeneous suspension and allowed to react for 12 h at room temperature under continuous stirring. It is necessary here to ensure that the concentration and volume of the glutaraldehyde solution are sufficiently in excess, and the magnetic particles are added into the glutaraldehyde solution. The order cannot be reversed to avoid aminated nanoparticles from being cross-linked by an insufficient amount of glutaraldehyde. The obtained Fe₃O₄@SiO₂-glutaraldehyde NPs were magnetically separated and then rinsed with equal volumes of ultrapure water three times, and finally collected magnetically.

2.4 Preparation of water-compatible MMIPs

The DES imprinted water-compatible magnetic MIPs (denoted as Fe₃O₄@SiO₂@APBA/MIPs) were prepared *via* a surface-imprinting polymerization process.^{30,37} The Fe₃O₄@SiO₂-glutaraldehyde NPs were redispersed in 50 mL of 20 mM APBA aqueous solution, stirred for 30 minutes, and statically aged for 12 h to allow self-assembly on the Fe₃O₄@SiO₂ NPs surface. For prepolymerization, 50 mL of the template-monomer solution containing 20 mM APBA and 5 mM DES was shaken for 30 min at room temperature and set aside for 12 h. Then, the self-assembly suspension and 20 mg of K₂S₂O₈ were added. The mixture was stirred at reflux at 60 $^{\circ}$ C for 24 h under an N₂ atmosphere for polymerization of poly(APBA). After magnetic separation, the obtained DES-loaded MIPs (denoted as Fe₃O₄@SiO₂@APBA/MIPs-DES) were rinsed with ethanol and ultrapure water and then eluted with the mixture of methanol-



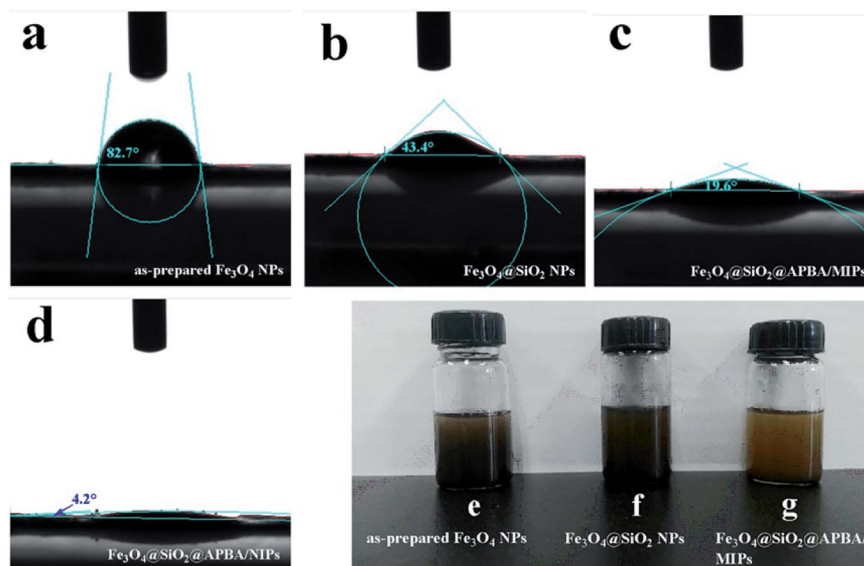


Fig. 3 Static water contact angles of the as-prepared Fe_3O_4 NPs (a), $\text{Fe}_3\text{O}_4@SiO_2$ NPs (b), $\text{Fe}_3\text{O}_4@SiO_2@APBA/MIPs$ (c), $\text{Fe}_3\text{O}_4@SiO_2@APBA/NIPs$ (d), and photographs showing the dispersion in pure water at 298 K of 1 mg mL^{-1} of the as-prepared Fe_3O_4 NPs (e), $\text{Fe}_3\text{O}_4@SiO_2$ NPs (f), $\text{Fe}_3\text{O}_4@SiO_2@APBA/MIPs$ (g) after settling down for 24 h.

0.1 M acetic acid (5 mL, 9 : 1, v/v) repeatedly with shaking to remove DES, until the eluent was free from DES as detected by UV-vis spectrometry at 240 nm.^{12,24} Finally, the resulting $\text{Fe}_3\text{O}_4@SiO_2@APBA/MIPs$ were washed thoroughly with ethanol and ultrapure water and dried at 40 °C under nitrogen gas protection overnight. Thus, recognition cavities complementary to DES in shape, size, and chemical functionality were formed in imprinted layers, which could selectively rebind DES. For comparison, non-imprinted polymers ($\text{Fe}_3\text{O}_4@SiO_2@APBA/NIPs$) were prepared using the same procedures in the absence of DES.

2.5 Adsorption experiments

Static adsorption experiments were performed at 288, 293, 298, 308, and 318 K to investigate the effect of temperature on the adsorption capacities of $\text{Fe}_3\text{O}_4@SiO_2@APBA/MIPs$ toward DES. MIPs or NIPs (20.0 mg) were suspended in a series of 50 mL DES aqueous solutions with various initial concentrations (C_0 , mg L^{-1}) ranging from 0.0500 to 100 mg L^{-1} . After a series of adsorbent-adsorbate mixtures were mechanically shaken for 3 h at different temperatures, the MIP or NIP NPs were separated magnetically, and then the equilibrium adsorption concentration of DES (C_e , mg L^{-1}) in the collected supernatant was measured by UV-vis spectrophotometer operating at 240 nm. The binding amounts of DES on MIPs or NIPs at equilibrium, defined as the equilibrium adsorption capacity (Q_e , mg g^{-1}), could be calculated using eqn (1):²⁵

$$Q_e = \frac{C_0 - C_e}{m} V \quad (1)$$

where V (L) represents the volume of DES solution and m (g) denotes the mass of $\text{Fe}_3\text{O}_4@SiO_2@APBA/MIPs$ (or NIPs) used.

The binding kinetics experiment procedure was similar to the static adsorption study for the monitoring of the minimum adsorption equilibrium time. $\text{Fe}_3\text{O}_4@SiO_2@APBA/MIPs$ or NIPs (20.0 mg) were added to 50 mL of DES solution with an initial concentration (C_0) of 60 mg L^{-1} . The suspension was shaken continuously for a series of time intervals (t) from 5 to 200 min at 298 K. The temporal concentration of DES (C_t , mg L^{-1}) in the supernatants was analyzed by UV. The binding amounts for DES with different contact time t , defined as the temporal adsorption capacity (Q_t , mg g^{-1}), was calculated as (eqn (2)):²⁵

$$Q_t = \frac{C_0 - C_t}{m} V \quad (2)$$

2.6 Application for analysis of DES in lake water samples

The $\text{Fe}_3\text{O}_4@SiO_2@APBA/MIPs$ were applied to extraction and then analysis of DES from lake water samples using the MSPE method coupled with HPLC.²⁴ The process is illustrated in Fig. 1B. First, 2000 mL of water samples collected from Moon Lake located in Beijing University of Technology (China) were filtered with a 0.45 μm filter membrane three times under vacuum. Then the filtered samples were stored at 4 °C for further experiments.³⁸ Before the first use, MIPs or NIPs were conditioned sequentially by immersion in ethanol (3 mL), elution solution (3 mL), and ultrapure water (3 mL) for 3 min, respectively. Subsequently, 80.0 mg MIPs were dispersed in 500 mL of the filtered samples or standard aqueous solutions, and then shaken for 160 min at 298 K, to achieve complete adsorption. MIPs or NIPs were collected using a magnet. After the MSPE step, saturated MIPs or NIPs were washed in sequence with 5.0 mL of ethanol and water, and followed by 5.0 mL of



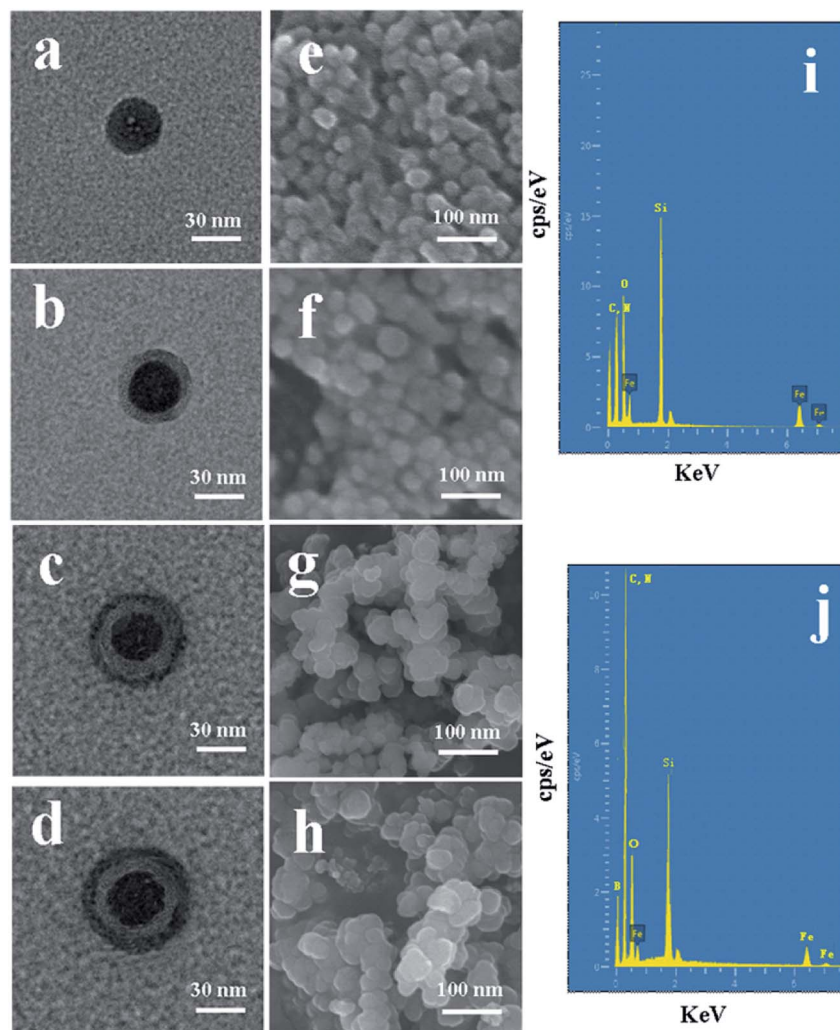


Fig. 4 TEM images of (a) as-prepared Fe_3O_4 , (b) $\text{Fe}_3\text{O}_4@SiO_2$, (c) $\text{Fe}_3\text{O}_4@SiO_2@APBA/MIPs$, and (d) $\text{Fe}_3\text{O}_4@SiO_2@APBA/NIPs$. SEM images of (e) as-prepared Fe_3O_4 , (f) $\text{Fe}_3\text{O}_4@SiO_2$, (g) $\text{Fe}_3\text{O}_4@SiO_2@APBA/MIPs$, and (h) $\text{Fe}_3\text{O}_4@SiO_2@APBA/NIPs$. EDS spectra of (i) as-prepared $\text{Fe}_3\text{O}_4@SiO_2$ and (j) $\text{Fe}_3\text{O}_4@SiO_2@APBA/MIPs$.

elution solution, and then separated magnetically. Finally, the collected eluents were determined by HPLC.

3. Results and discussion

3.1 Optimizing preparation conditions for MIPs

The molar ratio of the template-functional monomer is important in a successful imprinting process because of its effect on the number of recognition sites formed in MIPs and the quality of the MIPs. Adsorption tests of $\text{Fe}_3\text{O}_4@SiO_2@APBA/MIPs$ prepared with different molar ratios (1 : 3, 1 : 4, and 1 : 5) were carried out at 298 K in 60 mg L⁻¹ DES aqueous solution. The results indicated that the highest adsorption capacity of 18.85 mg g⁻¹ was achieved at a ratio of 1 : 4, while only 40% and 65% were achieved at ratios of 1 : 3 and 1 : 5, respectively. Therefore, a molar ratio of 1 : 4 was adopted in subsequent experiments. Moreover, the amount of MIP coating on the surface of $\text{Fe}_3\text{O}_4@SiO_2$ can also influence the adsorption capacity. Different quantities of $\text{Fe}_3\text{O}_4@SiO_2$ (25, 50, 75, and 100 mg) were used for the preparation of MIPs with fixed amounts

of the other reactant. The results demonstrated that 50 mg of $\text{Fe}_3\text{O}_4@SiO_2$ was optimal for further study.

3.2 Characterization of $\text{Fe}_3\text{O}_4@SiO_2@APBA/MIPs$

The FTIR (KBr) spectra of the functionalized Fe_3O_4 and imprinted NPs are presented in Fig. 2A. For Fe_3O_4 (curve a), the strong absorption peaks located at 586 and 3437 cm⁻¹ could be attributed to stretching vibrations of Fe–O and Fe–OH bonds. The bands around 1093 and 800 cm⁻¹ were caused by asymmetric and symmetric stretching vibrations of the Si–O groups (curve b), demonstrating that covalent bonds between the silane coupling agent and magnetite surface were generated on $\text{Fe}_3\text{O}_4@SiO_2$ NPs.³⁹ Characteristic peaks at 2930 and 2851 cm⁻¹ (stretching vibration of –CH₂–), 2960 cm⁻¹ (stretching vibration of –CH₃) and 3182 cm⁻¹ (corresponding to –NH₂) could be ascribed to the APTES successfully modified on silica shells (curve c). The new bands at 1726 and 2720 cm⁻¹ were, respectively, the characteristic of the C=O stretch and the C–H stretch of the aldehyde group from glutaraldehyde (curve d). Moreover,



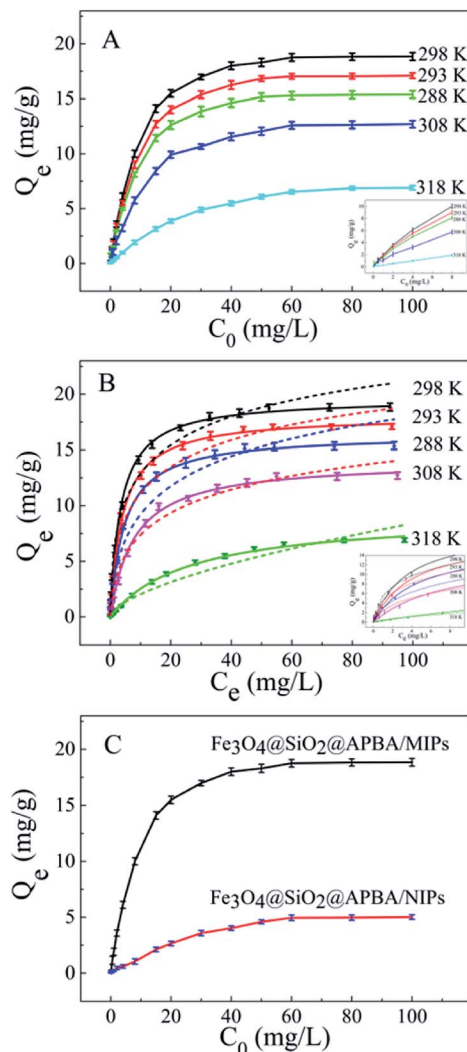


Fig. 5 (A) Equilibrium adsorption isotherms of DES on $\text{Fe}_3\text{O}_4\text{-SiO}_2\text{-APBA/MIPs}$ at five different temperatures (20 mg of $\text{Fe}_3\text{O}_4\text{-SiO}_2\text{-APBA/MIPs}$ in 50 mL of DES solution shaken for 3 h); (B) fitted adsorption isotherms with two adsorption isotherm models; the solid line is the Langmuir model simulation and the dotted line is the Freundlich model simulation; (C) equilibrium adsorption curves of $\text{Fe}_3\text{O}_4\text{-SiO}_2\text{-APBA/MIPs}$ and $\text{Fe}_3\text{O}_4\text{-SiO}_2\text{-APBA/NIPs}$ for DES at 298 K. Inserts of (A) and (B): adsorption isotherms of DES solution with initial concentrations ranging from 0.0500 to 8.00 mg L^{-1} .

the disappearance of the $-\text{NH}_2$ stretching vibration peak and the appearance of a new band of $\text{C}=\text{N}$ at 1651 cm^{-1}

demonstrated that glutaraldehyde was grafted onto the surface of $\text{Fe}_3\text{O}_4\text{-SiO}_2\text{-NH}_2$ by condensation with dehydration.^{25,40} Bands centered at 710 and 1340 cm^{-1} can be assigned to $-\text{B}-\text{OH}$ bending and stretching vibrations of APBA, respectively (curves e–g). The new bands centered at 650 cm^{-1} might result from the $\text{C}-\text{B}$ bond. Moreover, the increment of peak intensity at 1651 cm^{-1} from $\text{C}=\text{N}$, and the absent peak of $\text{C}=\text{O}$ at 1726 cm^{-1} observed in Fig. 2A (e–g) can be attributed the contribution of APBA. The results proved that poly(APBA) was bound to the $\text{Fe}_3\text{O}_4\text{-SiO}_2$ surface by covalent bonds. However, the expected bands around 586 , 1093 , and 1340 cm^{-1} corresponding to $\text{Fe}-\text{O}$, $\text{Si}-\text{O}$ and $\text{Si}-\text{O}$ bonds, respectively, in $\text{Fe}_3\text{-O}_4\text{-SiO}_2\text{-APBA-DES/MIPs}$ were overlapped with new peaks at $500\text{--}1600\text{ cm}^{-1}$ (curve e). These new bands were in accordance with bands from the FTIR spectrum of DES (curve h). Furthermore, the shifted stretching frequency of the $\text{O}-\text{H}$ group from 3437 to 3412 cm^{-1} (curve h) was due to the formation of hydrogen bonding between hydroxyl groups. After the sample was eluted, no characteristic bands of DES could be observed (curve f), indicating that the DES molecules were removed from the MIP composite. All results indicated that MIP layers were grafted on the surface of $\text{Fe}_3\text{O}_4\text{-SiO}_2$ NPs.

Thermogravimetric analysis (TGA) curves for Fe_3O_4 , $\text{Fe}_3\text{-O}_4\text{-SiO}_2$, $\text{Fe}_3\text{O}_4\text{-SiO}_2\text{-APBA/MIPs}$ and NIPs are shown in Fig. 2B. Weight loss at a temperature less than $200\text{ }^\circ\text{C}$ can be attributed to the elimination of water. The weight loss for Fe_3O_4 NPs and $\text{Fe}_3\text{O}_4\text{-SiO}_2$ NPs was approximately 1% (curve a) and 4% (curve b), respectively, when heated to $800\text{ }^\circ\text{C}$. The weight loss can be attributed to the decomposition of some contaminations and the grafted silane agent. Moreover, the weight loss of 25.9% for $\text{Fe}_3\text{O}_4\text{-SiO}_2\text{-APBA/MIPs}$ suggested that the imprinted polymers were grafted on $\text{Fe}_3\text{O}_4\text{-SiO}_2$ (curve c). Significant weight loss for $\text{Fe}_3\text{O}_4\text{-SiO}_2\text{-APBA/NIPs}$ (35.8%, curve d) could be observed. The slight difference between the imprinted NPs and non-imprinted NPs may be due to the different grafting density caused by DES. The difference in thermal stability between these NPs showed that the imprinted polymers successfully grafted onto the Fe_3O_4 .

The magnetic saturation test was performed at room temperature using a VSM to characterize the magnetic properties of Fe_3O_4 core-based nanoparticles. The three magnetic hysteresis loops with similar general shape are illustrated in Fig. 2C. As shown, the saturation magnetization of Fe_3O_4 , $\text{Fe}_3\text{-O}_4\text{-SiO}_2$, and $\text{Fe}_3\text{O}_4\text{-SiO}_2\text{-APBA/MIPs}$ at a field of 2.0×10^4 Oe decreased from 85.8 to 63.9 , and then to 46.3 emu g^{-1} , with increasing thickness of the modified layer on the surface of

Table 1 Adsorption isotherm parameters for DES adsorption on $\text{Fe}_3\text{O}_4\text{-SiO}_2\text{-APBA/MIPs}$

Temperature	$Q_{e(\text{exp})}$ (mg g^{-1})	Langmuir			Freundlich		
		Q_m (mg g^{-1})	K_L (L mg^{-1})	R^2	K_F	n	R^2
288 K	15.76	16.42	0.2167	0.9976	3.8734	2.2534	0.9594
293 K	17.48	18.08	0.2503	0.9990	4.4906	2.4193	0.9614
298 K	18.85	19.62	0.2917	0.9986	5.5139	2.7566	0.9721
308 K	12.65	14.04	0.1285	0.9989	2.4821	1.8384	0.9824
318 K	6.52	7.19	0.0382	0.9975	0.4953	1.6263	0.9691



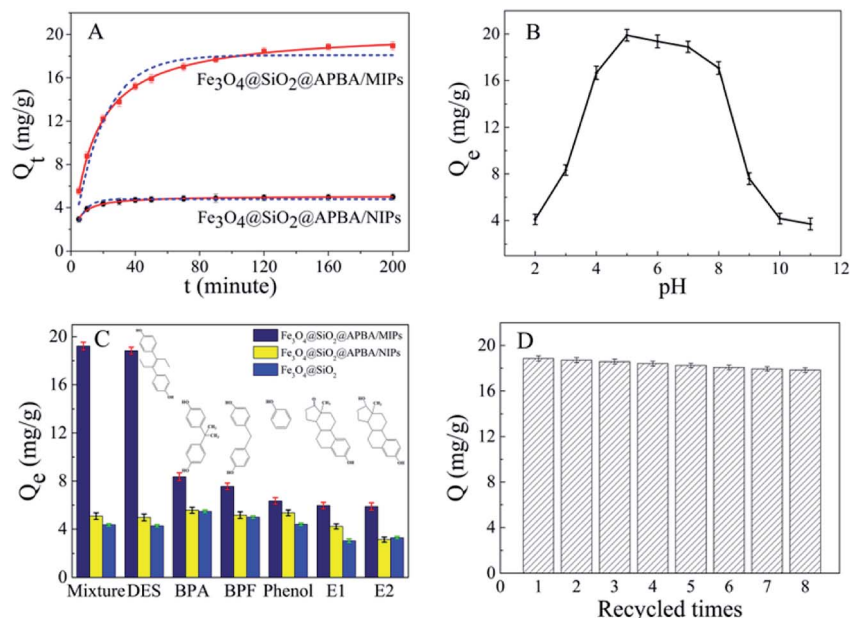


Fig. 6 (A) Kinetic adsorption curves of DES on MIPs and NIP NPs at 298 K. The dotted line and solid line correspond to the pseudo-first-order and pseudo-second-order fitting, respectively. (B) Effect of solution pH on DES adsorption on MIP NPs. (C) Selective adsorption capacities of MIPs, NIPs, and $\text{Fe}_3\text{O}_4@SiO_2$ NPs for DES, BPA, BPF, phenol, E1 and E2. (D) Regeneration cycles for MIP NPs.

Fe_3O_4 . However, the decrease did not clearly affect the magnetic performance of the $\text{Fe}_3\text{O}_4@SiO_2@APBA/MIPs$ particles; their rapid magnetic responsivity was demonstrated by the achievement of rapid adsorption within 5 s under an applied magnetic field. The adsorption experiments confirmed that MIP NPs can be used for effective magnetic separation.

Water contact angle experiments were performed to accurately evaluate the surface hydrophilicity of the nanoparticles.^{25,26} Fig. 3 shows the profiles of water droplets on compacted films of the Fe_3O_4 nanoparticles, $\text{Fe}_3\text{O}_4@SiO_2$ nanoparticles, $\text{Fe}_3\text{O}_4@SiO_2@APBA/MIPs$, and $\text{Fe}_3\text{O}_4@SiO_2@APBA/NIPs$, respectively. The as-prepared Fe_3O_4 NPs were hydrophobic, with a contact angle of 82.7° (Fig. 3a). The static water contact angle of the $\text{Fe}_3\text{O}_4@SiO_2$ NP film was 43.4° (Fig. 3b), indicating that $\text{Fe}_3\text{O}_4@SiO_2$ NPs were more hydrophilic than Fe_3O_4 NPs because of the presence of rich polar functional groups on their surface. Furthermore, the contact angle of the $\text{Fe}_3\text{O}_4@SiO_2@APBA/MIP$ film was much smaller at 19.6° (Fig. 3c), demonstrating that the hydrophilic MIPs were successfully grafted onto the surface of the $\text{Fe}_3\text{O}_4@SiO_2$ NPs. In addition, the contact angle of the $\text{Fe}_3\text{O}_4@SiO_2@APBA/NIPs$ was 4.2° (Fig. 3d), indicating their super hydrophilicity, which can be explained by the preparation through the grafting of APBA

polymers directly onto $\text{Fe}_3\text{O}_4@SiO_2$, with no DES. The improved surface hydrophilicity was beneficial to good dispersibility of the materials in water. As shown in Fig. 3(e) and (f), many more floats or sedimentations were present in Fe_3O_4 and silanized Fe_3O_4 suspensions than in the $\text{Fe}_3\text{O}_4@SiO_2@APBA/MIPs$ suspension. The excellent dispersion of $\text{Fe}_3\text{O}_4@SiO_2@APBA/MIPs$ in water provided greater opportunity for the DES molecules to access the imprinted cavities.

The morphological structure and particle size of the synthesized nanoparticles can be observed by TEM and SEM. It can be observed from Fig. 4a–d that the mean diameter sizes of Fe_3O_4 , $\text{Fe}_3\text{O}_4@SiO_2$, $\text{Fe}_3\text{O}_4@SiO_2@APBA/MIPs$, and $\text{Fe}_3\text{O}_4@SiO_2@APBA/NIPs$ were approximately 30, 40, 50 and 55 nm, respectively. An SiO_2 shell with a thickness of approximately 5 nm was clearly seen to be uniformly coated over the Fe_3O_4 dark core (Fig. 4b), forming the first layer of the core-shell structure, indicating the success of the fully coated silica shell. After imprinting, another external polymer layer with a thickness of approximately 5 nm appeared around $\text{Fe}_3\text{O}_4@SiO_2$ micro-particles (Fig. 4c), which suggests that the second imprinted shell had been successfully grafted.⁴¹ As seen from the SEM images, initially the Fe_3O_4 has a rough surface and a regular spherical shape, but relatively severe agglomeration

Table 2 Parameters of the two adsorption kinetic models for DES on MIPs and NIP NPs

Absorbent	$Q_{e(\text{exp})}$ (mg g^{-1})	Pseudo-first-order			Pseudo-second-order		
		R^2	k_1 (min^{-1})	$Q_{e(\text{cal})}$ (mg g^{-1})	R^2	k_2 (min^{-1})	$Q_{e(\text{cal})}$ (mg g^{-1})
$\text{Fe}_3\text{O}_4@SiO_2@APBA/MIPs$	18.85	0.9604	0.05446	18.08	0.9993	0.00364	18.83
$\text{Fe}_3\text{O}_4@SiO_2@APBA/NIPs$	4.96	0.9399	0.17418	4.80	0.9939	0.05699	4.92



Table 3 Imprinting factors and relative selectivity constants of DES and analogs for Fe₃O₄@SiO₂@APBA/MIPs and Fe₃O₄@SiO₂@APBA/NIPs

Analyte	Q _{MIPs} (mg g ⁻¹)	Q _{NIPs} (mg g ⁻¹)	IF	SC
DES	18.85	4.96	3.80	—
BPA	8.31	5.59	1.49	2.55
BPF	7.59	5.18	1.47	2.59
Phenol	6.33	5.35	1.18	3.22
E1	5.96	4.22	1.41	2.70
E2	5.88	3.12	1.88	2.02

(Fig. 4e). After grafting by the silane coupling agent, the agglomeration of Fe₃O₄@SiO₂ was alleviated, the surface became slightly smooth, and the morphology of the sphere became more regular (Fig. 4f). The chemical composition and elemental mapping of Fe₃O₄@SiO₂ and Fe₃O₄@SiO₂@APBA/MIPs were characterized by X-ray EDS analysis. In the EDS spectrum of Fe₃O₄@SiO₂ (Fig. 4i), the presence of iron, silicon, carbon, and nitrogen suggested that the silane coupling agent was grafted onto the surface of Fe₃O₄. After imprinting, the microspheres became larger, due to the coated imprinted polymers, and more uniform in size distribution (Fig. 4g). In addition, boron from APBA was observed in the EDS spectrum (Fig. 4j), while the peak for nitrogen overlaps with that of carbon. These results further confirm that imprinted polymers were coated on Fe₃O₄. For comparison, there was no remarkable difference in morphology and diameter between Fe₃O₄@SiO₂@APBA/NIPs (Fig. 4d and h) and Fe₃O₄@SiO₂@APBA/MIPs (Fig. 4c and g). Both MIP and NIP particles possessed similar uniform core-shell structures. However, the MIP NPs appeared to have a more uniform size distribution than the NIP NPs. There was a slight difference of 2.5 nm in the shell thickness of the NIPs compared with that of the MIPs (Fig. 4d), which might be due to the absence of DES molecules in the formation of the imprinting polymers shell. As shown, these results were in agreement with the above discussion on the FTIR spectra, static water contact angle, VSM, and TG analyses. Compared with other reports³⁴ based on coprecipitation reactions or using the solvothermal method,⁴² the hydrophilic Fe₃O₄ magnetic nanoparticles synthesized in this paper possessed higher magnetization or smaller particle sizes, respectively. The encapsulation of Fe₃O₄ with a nonporous SiO₂ shell could improve their dispersion in water, easily be modified with various groups, prevent the oxidization and agglomeration of Fe₃O₄, and then increase their reusability.^{43,44}

3.3 Adsorption isotherm studies of Fe₃O₄@SiO₂@APBA/MIPs

The adsorption isotherms of DES on Fe₃O₄@SiO₂@APBA/MIPs at five different temperatures are shown in Fig. 5A. The equilibrium adsorption capacities at 298 K were the highest among those at 288, 293, 308, and 318 K. When the temperature was higher than 298 K, the equilibrium adsorption capacities increased with decreasing temperature, which is consistent with previous findings showing that the imprinting cavities of

MIPs prepared at low temperatures possess a similar three-dimensional structure at low temperature, such that MIPs are more effective at low temperatures.⁴⁵ However, the capacities decreased with decrements in temperature below 298 K. The reason might be that the low temperature resulted in a slower diffusion rate of DES between the solution and the MIP film.²⁴ Furthermore, it can be seen that the adsorption capacity of Fe₃O₄@SiO₂@APBA/MIPs increased with increasing DES equilibrium concentration. The increase in DES concentration can accelerate the diffusion of DES molecules onto Fe₃O₄@SiO₂@APBA/MIPs. Therefore, 298 K was chosen as the appropriate temperature for subsequent experiments.

The Langmuir and Freundlich adsorption isotherm models were used for the nonlinear fitting of experimental data and evaluation of the adsorption isotherms of Fe₃O₄@SiO₂@APBA/MIPs (Fig. 5B). The Langmuir model is suitable for monolayer adsorption on uniform energy surfaces. The model equation is described in eqn (3):⁴⁶

$$\frac{C_e}{Q_e} = \frac{1}{Q_m} C_e + \frac{1}{Q_m K_L} \quad (3)$$

where Q_e (mg g⁻¹) represents the equilibrium adsorption capacity of DES, C_e (mg L⁻¹) is the equilibrium concentration of DES in solution, and Q_m (mg g⁻¹) is the maximum adsorption capacity of the adsorbent. The Langmuir constant of K_L (L mg⁻¹) is related to the affinity of the binding sites. The Freundlich model is suitable for multilayer adsorption occurred on heterogeneous surfaces. The model equation is expressed in eqn (4):⁴⁷

$$\ln Q_e = \frac{1}{n} \ln C_e + \ln K_F \quad (4)$$

where the Freundlich constants K_F and n represent the adsorption capacity and adsorption favorability, respectively. Table 1 listed the fitted values of Q_m , K_L , K_F , n , and R^2 calculated from the above two nonlinear regression isotherm models. Adsorption isotherms of Fe₃O₄@SiO₂@APBA/MIPs can be better described by the Langmuir model at five temperatures because their R^2 values all exceed 0.99, and are much higher than those of the Freundlich model. Moreover, it is seen that the Q_m calculated from the Langmuir equation were very close to the $Q_{e(\text{exp})}$ values obtained from experiments.

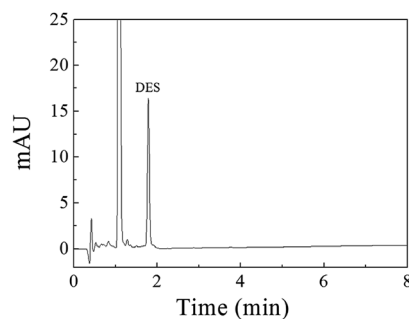


Fig. 7 Chromatogram of the eluate obtained using 80 mg of Fe₃O₄@SiO₂@APBA/MIPs to extract 500 mL of the DES standard solution (0.42 μg L⁻¹) for 160 min at 298 K.



Table 4 Analytical performances comparison of this MIPs-MSPE based on Fe₃O₄@SiO₂@APBA/MIPs with other MIPs based methods for estrogens by HPLC^a

Support	Functional monomer	Solvent	Evaluation solution	Target estrogens	Adsorption capacity (mg g ⁻¹)	Extraction method	Analytical method	Linearity range (μg L ⁻¹)	LODs (μg L ⁻¹)	Real sample	Ref.
RAFTPP	MAA, NIPAM	Acetonitrile	Aqueous	BPA	8.292	Packed SPE	HPLC	—	41.0 ng L ⁻¹	Seawater	24
—	MAA	Chloroform	Methanol	DES	8.43	MISPE	HPLC-DAD	—	1.8	Seawater	6
Silica gel	APTES	Methanol	Methanol	DES	62.58	SPE	HPLC	—	60	Fish samples	13
ATP	AA, MBAA	Ethanol	Ethanol	DES	105.14	SPE packing	HPLC	—	3	Pond water, fish samples	14
Silica-coated Fe ₃ O ₄ CNTs@SiO ₂	APTES, PTMOS	Ethanol	Ethanol	DES, E3, E2	6.57, 5.38, 3.82	SPE	HPLC	0.3–100	0.08–0.27	Lake, river water	15
—	Silica	Acetate buffer	Chloroform	DES, E3, E1	30.46	Removing	HPLC	5.0–1 × 10 ⁴	10.2–16.1	River, lake, tap water	16
Fe ₃ O ₄ @nafion	Dopamine	Tris-HBS	Aqueous	DES	9.74	DSPE	HPLC-DAD	3.5–1000 ng g ⁻¹	0.99 ng g ⁻¹	Milk	5
Fe ₃ O ₄ @SiO ₂	MAA	Acetonitrile	Acetonitrile	DES	3.086	MMIPs	HPLC-UV	20–8000	3.6/6.3	Pond water, milk	17
HFT	MAA	Acetonitrile	Acetonitrile	DES	—	Microextraction	HPLC	7.5–200	2.5	Milk	1
M-ATP	Cyd	Aqueous	Aqueous	E1, E2, E3, DES	0.04–0.1 mmol g ⁻¹	On-line SPE	HPLC-UV	3.0–2500 ng g ⁻¹	1–8 ng g ⁻¹	Milk	27
—	—	—	—	—	10.73 (DES)	—	—	—	—	—	—
Fe ₃ O ₄ @SiO ₂	AA	MeOH, ACN	Aqueous	E1, E2, E3, DES	0.216–0.16	SPME	HPLC-UV	8–2000 ng g ⁻¹	1.5–5.5 ng g ⁻¹	Milk powder	51
Fe ₃ O ₄ @SiO ₂	APBA	Aqueous	Aqueous	DES	18.85	MSPE	HPLC-DAD	0.08–150	0.03	Lake water	This work

^a PP: precipitation polymerization; MAA: α-methacrylic acid; NIPAM: N-isopropyl acrylamide; MISPE: molecularly imprinted solid-phase extraction; ATP: attapulgite nanofibrillar clay; MBAA: N,N'-methylene-bisacrylamide; AA: acrylamide; PTMOS: phenyltrimethoxysilane; CNTs: carbon nanotube; Tris-HBS: Tris-HCl buffer solution; HFT: hollow fiber tube; M-ATP: methylacryloxypropyl modified attapulgite; CyD: acryloyl-β-cyclodextrin; SPME: solid-phase microextraction.

Next, static adsorption experiments of NIP NPs were performed at 298 K for comparison with MIP NPs. At first, Q_e of the two adsorbents increased remarkably as the initial concentration increased from 0.500 to 50.0 mg L⁻¹, and then reached saturation adsorption at 50.0 mg L⁻¹ (Fig. 5C). However, Q_e of MIP NPs (18.85 mg g⁻¹) was approximately 3.8 times of that of NIP NPs (4.96 mg g⁻¹). These results suggest that NIP NPs have no specific adsorption properties.

3.4 Adsorption kinetic studies

The adsorption kinetics investigation showed that the minimum required time for the adsorption equilibrium for Fe₃O₄@SiO₂@APBA/MIPs was 160 min at 298 K (Fig. 6A). From this finding, 160 min was chosen as the optimal extraction time. The adsorption process was quite fast in comparison with traditional imprinted materials which would take 12–24 h to reach equilibrium state.⁴⁸ The reason for this can be attributed to the fact that the surface poly(APBA) imprinting films wrapped on Fe₃O₄@SiO₂ nanoparticles provided more binding sites at their surface and achieved faster mass transfer. NIP NPs showed a similar trend, but with much lower adsorption capacities. Two kinds of adsorption kinetic models were applied to fitting the experimental kinetic data of MIP NPs for DES to study the rate control and mass transfer mechanism of the adsorption process of DES at Fe₃O₄@SiO₂@APBA/MIPs according to previous reports.^{38,39,42} The pseudo-first-order model can be described as follows (eqn (5)):⁴⁹

$$\ln(Q_e - Q_t) = \ln Q_e - k_1 t \quad (5)$$

The pseudo-second-order model comprises all the steps of adsorption including external film diffusion, adsorption, and internal particle diffusion, and can be described as follows (eqn (6)):⁵⁰

$$\frac{t}{Q_t} = \frac{1}{k_2 Q_e^2} + \frac{t}{Q_e} \quad (6)$$

where Q_t (mg g⁻¹) is the instantaneous adsorption amount at various times t , and k_1 and k_2 (min⁻¹) are the adsorption rate constants. The plotted nonlinear regression fitting curves present a comparison of the two kinetic models (Fig. 6A). Corresponding fitting parameters and R^2 are summarized in Table 2, and show that the pseudo-second-order model was a better fit for the higher regression coefficient R^2 (>0.99, Table 2). Furthermore, the calculated adsorption capacity ($Q_{e(\text{cal})}$, Table 2) from the pseudo-second-order model agreed well with experimental data ($Q_{e(\text{exp})}$, Fig. 6A). Similar results have previously been reported for the adsorption of hydroxybenzoic acids on magnetic MIPs⁴² and estrogens on MIPs.³⁸ Therefore, the pseudo-second-order model was more suitable for describing the mass transfer process of DES molecules on Fe₃O₄@SiO₂@APBA/MIPs particles in solution. The adsorption process can be divided into three steps; *i.e.*



boundary diffusion, intra-particle diffusion, and adsorption reaction. The complex effect of multiple adsorption mechanisms is suitable for adsorption process with saturation sites.⁴⁰

3.5 Effect of solution pH

The pH experiments were performed in 50 mL of 60 mg L⁻¹ DES solutions with different pH values, with 20.0 mg of Fe₃O₄@SiO₂@APBA/MIPs dispersed in them for adsorption for 160 min. The competitive adsorption experiments were carried out in 50 mL of suspension with 20.0 mg of Fe₃O₄@SiO₂@APBA/MIPs or NIPs and 60 mg L⁻¹ of DES, BPA, BPF, phenol, E1, and E2 for adsorption for 160 min.

Fig. 6B shows the adsorption capacities of Fe₃O₄@SiO₂@APBA/MIPs toward DES at different pH values. The capacities in a broad pH range (pH 4 to 8) were attractive, although decreased rapidly at pH values lower than 4 or higher than 8. This is because the net charge of DES differs from that of the adsorbent at different pH values. When the pH value is greater than 8, DES molecules possess a negative charge value because of the phenolic hydroxyl anions.²⁵ Meanwhile, APBA, with its weak boric acid groups, may dissociate in the high pH range.²⁹ Thus, electrostatic repulsion between negatively charged MIPs and DES might result in the decreased adsorptivity of MIPs. Furthermore, the amino protonation might occur to APBA when the pH value of the solution was less than pH 4. As a result, hydrogen bonds were partially broken between APBA and DES, resulting in the decreased adsorptivity of MIPs to DES.²⁵

3.6 Binding selectivity for DES

Five reference compounds (BPA, BPF, phenol, E1, and E2) were used for the evaluation of binding selectivity. Fig. 6C demonstrates the clear differences in capacity for Fe₃O₄@SiO₂@APBA/MIPs between DES and reference compounds. In contrast, Fe₃O₄@SiO₂@APBA/NIPs and Fe₃O₄@SiO₂ NPs exhibited similar and poor adsorption toward the six reference compounds.

To further demonstrate the selectivity differences between imprinted and non-imprinted polymers, the parameters including the imprinting factor (IF), defined as $Q_{\text{MIPs}}/Q_{\text{NIPs}}$, and the relative selectivity constant (SC), defined as $IF_{\text{DES}}/IF_{\text{analogy}}$, were calculated.²⁰ The larger IF value demonstrated that MIPs for the analytes exhibited a higher selectivity. As presented in Table 3, the values of Q_{MIPs} for DES and IF_{DES} were larger than those of the other five reference compounds, indicating that Fe₃O₄@SiO₂@APBA/MIPs possessed relatively higher affinity for DES than those of its reference compounds. The similar Q_{NIPs} and SC values indicated that the adsorption of six compounds on Fe₃O₄@SiO₂@APBA/NIPs was non-specific. The above results confirm that the imprinting process was successfully achieved, and that Fe₃O₄@SiO₂@APBA/MIPs exhibits excellent recognition ability and high selectivity toward DES, even in the mixture of DES and five reference compounds with the same concentrations.

3.7 Reusability

The regeneration of the adsorbent is important in terms of practical applications. Saturated Fe₃O₄@SiO₂@APBA/MIPs (20 mg) was regenerated following consecutive steps of rinsing with ethanol three times, eluting under repeated shaking, and finally washing thoroughly with ethanol and ultrapure water. Regenerated MIPs were reused to extract 60 mg L⁻¹ of DES standard aqueous solutions, with the adsorption test was repeated over seven successive adsorption-regeneration cycles. The reusability was investigated by monitoring the adsorption capacity recovery. As shown in Fig. 6D, the adsorption capacity remained at 17.83 mg g⁻¹ after seven recycling procedures, and the adsorption efficiency lost was only 5.4% compared with the initial capacity (relative standard deviation [RSD] = 2.0%, $n = 8$). This excellent reusability and stability may be attributed to the properties of high chemical stability and good magnetic separation of the core-shell magnetic MIP, as well as the rapid mass transfer process.

3.8 Extraction performance for DES

An HPLC method for DES was established by using the Fe₃O₄@SiO₂@APBA/MIPs as adsorbents of MSPE. Different amounts of Fe₃O₄@SiO₂@APBA/MIPs, ranging from 20 to 100 mg, were used to extract DES from 500 mL extraction solvent when 10 µg L⁻¹ DES solution and 160 min of shaken-auxiliary extraction were adopted. The results show that recoveries could be higher than 95% when 80 mg adsorbent was used. However, when the amount of adsorbent was further increased, there was no clear increase in recovery. When the extraction time was increased from 30 min to 160 min, the recovery increased correspondingly from 45% to 95%. However, on further increase in extraction time, there was nearly no further increase in recovery. Therefore, shaken-auxiliary extraction for 3 h was adopted. Methanol, 0.1 M acetic acid, and different ratios of their mixture were tested as eluting

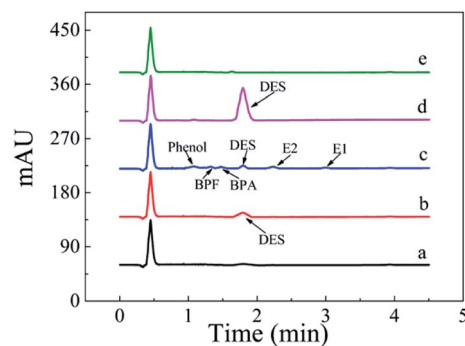


Fig. 8 HPLC-DAD chromatograms of the lake water sample. From the bottom to the top: (a) lake water sample without any pretreatment; (b) non-spiked sample extracted with Fe₃O₄@SiO₂@APBA/MIPs; (c) lake water sample spiked with DES, BPA, BPF, phenol, E1, and E2 individual at 1.5 µg L⁻¹ without extraction; spiked samples extracted for 160 min at 298 K with 80 mg of (d) Fe₃O₄@SiO₂@APBA/MIPs or (e) Fe₃O₄@SiO₂@APBA/NIPs.



Table 5 Results of the determination of DES in the spiked lake water sample by HPLC method

Sample	Detected ($\mu\text{g L}^{-1}$)	DES added ($\mu\text{g L}^{-1}$)	Found (average \pm SD) ($\mu\text{g L}^{-1}$)	RSD (%, $n = 6$)	Recovery (%, $n = 6$)
Lake water	0.08	0.1000	0.1858 ± 0.0052	2.8	103.2
		1.5000	1.5740 ± 0.0614	3.9	99.6
		10.0000	9.7877 ± 0.4249	4.3	97.1

solvents. The best recovery was obtained when 5 mL of a mixture of methanol-0.1 M acetic acid (9 : 1, v/v) was used.

A linear regression analysis was performed to obtain the calibration curves for detection of DES, and the ratios of HPLC peak areas (A , mA U s) versus corresponding concentrations of DES (C , $\mu\text{g L}^{-1}$) showed good linearity from 0.080 to $150 \mu\text{g L}^{-1}$ with correlation coefficients of R^2 value of 0.9992. The regression equation was $A = 49.6C - 2.8$. The limit of detection ($S/N = 3$) was $0.03 \mu\text{g L}^{-1}$ DES. The chromatogram of the eluate obtained using $\text{Fe}_3\text{O}_4@\text{SiO}_2@\text{APBA}/\text{MIPs}$ to extract DES standard solution is shown in Fig. 7. The method accuracy was studied by examining recoveries of spiked water samples at three levels (1.0, 10, and $100 \mu\text{g L}^{-1}$), and the recovery values were in the range of 95.6–103.4%. The intra-day and inter-day precisions of the method were given by the calculated RSD of extraction and analyses of DES at different spiked concentrations. The spiked concentrations at the above three levels were performed on the same day six times per day and on different days for consecutive six days, respectively.³⁸ The RSD values representing intra-day precision were 3.6%, 3.2% and 2.4% for the three concentrations, respectively ($n = 6$). The RSD for inter-day precision over 6 days were all less than 5.0% (4.8%, 4.2% and 3.8%, respectively, $n = 6$). Therefore, the results show that the proposed MSPE-HPLC method was applicable for rapid, sensitive, accurate, and quantitative determination of DES from water samples.

3.9 Method performance comparison

The proposed MIPs-MSPE-HPLC method for DES was compared with the other MIP-based pretreatment methods toward estrogens (Table 4). These reported methods were related to DES imprinted polymers synthesized and adsorption evaluated in organic solution, such as methanol,^{6,13} ethanol,^{14,15} acetonitrile,^{1,17} and chloroform.¹⁶ However, only a few studies on MIP adsorption for DES in aqueous phase have been reported.^{5,27,51} As seen from Table 4, compared with previous reports,^{1,6,15,17,24} the method in this paper not only created higher sensitivity, lower LODs and higher adsorption capacity to DES in aqueous solution, but also provided simple and fast pretreatment method.

3.10 Applications

The MSPE-HPLC method was applied to determine DES in lake water samples. There was almost no DES peak in HPLC-DAD chromatogram obtained from natural sample without enrichment or spiking (Fig. 8a). However, DES in the sample could be detected when analyzed by MSPE-HPLC based on $\text{Fe}_3\text{O}_4@\text{SiO}_2@\text{APBA}/\text{MIPs}$ under optimized conditions, and the

concentration value was $0.08 \mu\text{g L}^{-1}$ (Fig. 8b). Then the sample was spiked several times with $1.5 \mu\text{g L}^{-1}$ standard solutions of DES, BPA, BPF, phenol, E1, and E2, and the peak signals of analogs were all very weak (Fig. 8c). Therefore, quantitative analysis of trace DES in spiked samples by HPLC method without selective pretreatment process was difficult. After being enriched and extracted by MSPE based on MIPs and NIPs respectively, DES can be selectively adsorbed and then concentrated remarkably (Fig. 8d), and the peak of DES appeared distinctly. No obvious DES peak was observed in the eluted solution from NIPs (Fig. 8e), which also demonstrated the selectivity effect of the MIPs. Furthermore, the enrichment factor calculated was approximately 1900 for DES. The value of the enrichment factor demonstrated that $\text{Fe}_3\text{O}_4@\text{SiO}_2@\text{APBA}/\text{MIPs}$ possessed high pre-concentration ability for DES (Fig. 8d).³⁸ The lake water sample was then spiked at three levels (0.100 , 1.50 and $10.0 \mu\text{g L}^{-1}$) to validate the accuracy of the method in practical applications. Satisfactory recoveries of 97.1–103.2%, with RSD ranging from 2.8 to 4.3% ($n = 6$), were obtained (Table 5). The results indicated that the developed MIPs were ideal extraction adsorbents for MSPE, and thereby the proposed MIPs-MSPE-HPLC method was potentially applicable for highly efficient extraction and trace-determination of DES in real aqueous samples.

4. Conclusions

In this study, novel MIPs with excellent molecular recognition abilities and super water-compatibility (water contact angle of 19.6°) for the specific adsorption of DES in the aqueous phase were successfully prepared. The $\text{Fe}_3\text{O}_4@\text{SiO}_2@\text{APBA}/\text{MIPs}$ showed excellent features, such as high adsorption capacity (up to 18.85 mg g^{-1} at 298 K), rapid rebinding kinetics (only 160 min for adsorption equilibrium), good selectivity (imprinting factor of 3.80) and stability, as well as simple rapid magnetic separation. It was proven that $\text{Fe}_3\text{O}_4@\text{SiO}_2@\text{APBA}/\text{MIPs}$ provides great potential for pre-concentration of analyte samples in an environmentally friendly manner.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

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References

- M. Liu, M. Li, B. Qiu, X. Chen and G. Chen, *Anal. Chim. Acta*, 2010, **663**, 33–38.
- J. Wang, H. Ye, Z. Jiang, N. Chen and J. Huang, *Anal. Chim. Acta*, 2004, **508**, 171–176.
- W. Yan, Y. Li, L. Zhao and J.-M. Lin, *J. Chromatogr. A*, 2009, **1216**, 7539–7545.
- F. Huo, N. Li and X. Lin, *J. Northeast Agric. Univ.*, 2016, **23**, 44–49.
- L. Qiao, N. Gan, F. Hu, D. Wang, H. Lan, T. Li and H. Wang, *Microchim. Acta*, 2014, **181**, 1341–1351.
- X. He, X. Mei, J. Wang, Z. Lian, L. Tan and W. Wu, *Mar. Pollut. Bull.*, 2016, **102**, 142–147.
- Z. Yan, P. Xiong, N. Gan, J. He, N. Long, Y. Cao, F. Hu and T. Li, *J. Electroanal. Chem.*, 2015, **736**, 30–37.
- S. Liu, Z. Xie, X. Wu, X. Lin, L. Guo and G. Chen, *J. Chromatogr. A*, 2005, **1092**, 258–262.
- S. Yaqub, U. Latif and F. L. Dickert, *Sens. Actuators, B*, 2011, **160**, 227–233.
- Y. Liu, R. Liu, C. Liu, S. Luo, L. Yang, F. Sui, Y. Teng, R. Yang and Q. Cai, *J. Hazard. Mater.*, 2010, **182**, 912–918.
- T. Muhammad, L. Cui, W. Jide, E. V. Piletska, A. R. Guerreiro and S. A. Piletsky, *Anal. Chim. Acta*, 2012, **709**, 98–104.
- W.-R. Zhao, T.-F. Kang, L.-P. Lu, F.-X. Shen and S.-Y. Cheng, *J. Electroanal. Chem.*, 2017, **786**, 102–111.
- X. Jiang, C. Zhao, N. Jiang, H. Zhang and M. Liu, *Food Chem.*, 2008, **108**, 1061–1067.
- C. Zhao, Y. Ji, Y. Shao, X. Jiang and H. Zhang, *J. Chromatogr. A*, 2009, **1216**, 7546–7552.
- R. Gao, Y. Hao, S. Zhao, L. Zhang, X. Cui, D. Liu, Y. Tang and Y. Zheng, *RSC Adv.*, 2014, **4**, 56798–56808.
- R. Gao, X. Su, X. He, L. Chen and Y. Zhang, *Talanta*, 2011, **83**, 757–764.
- X. Xie, X. Pan, S. Han and S. Wang, *Anal. Bioanal. Chem.*, 2015, **407**, 1735–1744.
- M. Beytur, F. Kardaş, O. Akyıldırım, A. Özkan, B. Bankoğlu, H. Yüksek, M. L. Yola and N. Atar, *J. Mol. Liq.*, 2018, **251**, 212–217.
- M. L. Yola and N. Atar, *J. Electrochem. Soc.*, 2018, **165**, H1–H9.
- M. L. Yola, C. Göde and N. Atar, *Electrochim. Acta*, 2017, **246**, 135–140.
- M. L. Yola and N. Atar, *Ind. Eng. Chem. Res.*, 2017, **56**, 7631–7639.
- N. Atar, M. L. Yola and T. Eren, *Appl. Surf. Sci.*, 2016, **362**, 315–322.
- Y. Ren, W. Ma, J. Ma, Q. Wen, J. Wang and F. Zhao, *J. Colloid Interface Sci.*, 2012, **367**, 355–361.
- X. Wu, X. Wang, W. Lu, X. Wang, J. Li, H. You, H. Xiong and L. Chen, *J. Chromatogr. A*, 2016, **1435**, 30–38.
- F. Duan, C. Chen, L. Chen, Y. Sun, Y. Wang, Y. Yang, X. Liu and Y. Qin, *Ind. Eng. Chem. Res.*, 2014, **53**, 14291–14300.
- Y.-S. Chang, T.-H. Ko, T.-J. Hsu and M.-J. Syu, *Anal. Chem.*, 2009, **81**, 2098–2105.
- C. Zhao, X. Guan, X. Liu and H. Zhang, *J. Chromatogr. A*, 2012, **1229**, 72–78.
- Y. Guo, X. Liang, Y. Wang, Y. Liu, G. Zhu and W. Gui, *J. Appl. Polym. Sci.*, 2012, **128**, 4014–4022.
- L. Li, X. He, L. Chen and Y. Zhang, *Sci. China, Ser. B: Chem.*, 2009, **52**, 1402–1411.
- C.-L. Yan, Y. Lu and S.-Y. Gao, *J. Polym. Sci., Part A: Polym. Chem.*, 2007, **45**, 1911–1919.
- A. Bossi, S. A. Piletsky, E. V. Piletska, P. G. Righetti and A. P. F. Turner, *Anal. Chem.*, 2001, **73**, 5281–5286.
- L. Zhu, X. Yang and Y. Cao, *Anal. Lett.*, 2013, **46**, 982–998.
- J. Bai, X. Zhang, Y. Peng, X. Hong, Y. Liu, S. Jiang, B. Ning and Z. Gao, *Sens. Actuators, B*, 2017, **238**, 420–426.
- N.-N. Li, T.-F. Kang, J.-J. Zhang, L.-P. Lu and S.-Y. Cheng, *Anal. Methods*, 2015, **7**, 5053–5059.
- B. Luo, X. Song, F. Zhang, A. Xia, W. Yang, J. Hu and C. Wang, *Langmuir*, 2010, **26**, 1674–1679.
- H. Jiang, D. Jiang, J. Shao and X. Sun, *Biosens. Bioelectron.*, 2016, **75**, 411–419.
- Q.-P. You, M.-J. Peng, Y.-P. Zhang, J.-F. Guo and S.-Y. Shi, *Anal. Bioanal. Chem.*, 2014, **406**, 831–839.
- H. Lu and S. Xu, *Talanta*, 2015, **144**, 303–311.
- L. Xie, J. Guo, Y. Zhang, Y. Hu, Q. You and S. Shi, *Food Chem.*, 2015, **178**, 18–25.
- J. Liu, W. Wang, Y. Xie, Y. Huang, Y. Liu, X. Liu, R. Zhao, G. Liu and Y. Chen, *J. Mater. Chem.*, 2011, **21**, 9232–9238.
- J. Li, X. Zhang, Y. Liu, H. Tong, Y. Xu and S. Liu, *Talanta*, 2013, **117**, 281–287.
- S. Shi, J. Guo, Q. You, X. Chen and Y. Zhang, *Chem. Eng. J.*, 2014, **243**, 485–493.
- Y. Zhang, M. Nie, S. Shi, Q. You, J. Guo and L. Liu, *Food Chem.*, 2014, **146**, 56–64.
- X. Guo, F. Mao, W. Wang, Y. Yang and Z. Bai, *ACS Appl. Mater. Interfaces*, 2015, **7**, 14983–14991.
- Y. Lu, C. Li, X. Wang, P. Sun and X. Xing, *J. Chromatogr. B: Anal. Technol. Biomed. Life Sci.*, 2004, **804**, 53–59.
- J. Luo, S. Jiang and X. Liu, *J. Phys. Chem. C*, 2013, **117**, 18448–18456.
- X. Wang, J. Pan, W. Guan, J. Dai, X. Zou, Y. Yan, C. Li and W. Hu, *J. Chem. Eng. Data*, 2011, **56**, 2793–2801.
- W. J. Cheong, S. H. Yang and F. Ali, *J. Sep. Sci.*, 2013, **36**, 609–628.
- M. Meng, Z. Wang, L. Ma, M. Zhang, J. Wang, X. Dai and Y. Yan, *Ind. Eng. Chem. Res.*, 2012, **51**, 14915–14924.
- T. S. Anirudhan, S. R. Rejeena and J. Binusree, *J. Chem. Eng. Data*, 2013, **58**, 1329–1339.
- H. Lan, N. Gan, D. Pan, F. Hu, T. Li, N. Long and L. Qiao, *J. Chromatogr. A*, 2014, **1331**, 10–18.

