

Cite this: *Chem. Sci.*, 2018, 9, 7198

All publication charges for this article have been paid for by the Royal Society of Chemistry

Received 13th June 2018  
Accepted 12th July 2018

DOI: 10.1039/c8sc02610f

rsc.li/chemical-science

## Tetrazine-mediated bioorthogonal prodrug–prodrug activation†

Kevin Neumann,<sup>‡</sup> Alessia Gambardella,<sup>§</sup> Annamaria Lilienkamp and Mark Bradley<sup>\*</sup>

The selective and biocompatible activation of prodrugs within complex biological systems remains a key challenge in medical chemistry and chemical biology. Herein we report, for the first time, a dual prodrug activation strategy that fully satisfies the principle of bioorthogonality by the symbiotic formation of two active drugs. This dual and traceless prodrug activation strategy takes advantage of the  $_{\text{INV}}\text{DA}$  chemistry of tetrazines (here a prodrug), generating a pyridazine-based miR21 inhibitor and the anti-cancer drug camptothecin and offers a new concept in prodrug activation.

### Introduction

Conventional prodrug activation strategies typically rely on physiological changes *e.g.* pH around a tumour or a specific biological stimulus, for example the expression of an enzyme, to “switch-on” or activate a prodrug.<sup>1</sup> An alternative approach<sup>2,3</sup> is the application of chemical reactions that can take place within a biological environment with high selectivity and biocompatibility,<sup>4</sup> with such reactions typically being “unnatural” in origin. Bioorthogonal reactions have found applications in drug delivery and include examples of prodrug activation and even *in situ* drug synthesis.<sup>5</sup> Examples of bioorthogonal prodrug activation include application of the Staudinger reaction and strain-promoted alkene–azide cycloaddition that have been used to activate prodrugs of doxorubicin.<sup>6–8</sup> More broadly, bioorthogonal reactions have enabled the rapid and selective labelling of proteins,<sup>9,10</sup> glycans,<sup>11</sup> lipids<sup>12</sup> and DNA<sup>13</sup> under physiological conditions often in a pre-targeted imaging scenario.<sup>14–16</sup>

Since the inverse electron demand Diels–Alder ( $_{\text{INV}}\text{DA}$ ) reaction between tetrazines and electron-rich dienophiles was first described as a bioorthogonal reaction,<sup>17</sup> the tetrazine-promoted  $_{\text{INV}}\text{DA}$  reaction has been the subject of intense interest. This includes a series of studies where tetrazine-quenched profluorophores undergo “switch-on” of fluorescence upon treatment with a dienophile,<sup>19–21</sup> while tetrazine chemistry

has been used to label pre-targeted antibodies with PET isotopes.<sup>22–24</sup> Thus, tetrazine-mediated  $_{\text{INV}}\text{DA}$  chemistry has been shown to offer high chemical selectivity and to be rapid, efficient and biologically compatible, undoubtedly enhanced by the acceleration shown in water for all Diels–Alder chemistries.<sup>18,25</sup> Yet, despite their extensive use in imaging, examples of tetrazine-mediated prodrug activation are limited, but include a *trans*-cyclooctene–doxorubicin conjugate that liberates the decaged drug upon reaction with a tetrazine and subsequent oxidation of the resulting 1,4-dihydropyridazine to the pyridazine.<sup>26,27</sup> This approach was recently adapted to allow the release of carbonyl sulphide (OCS) that was converted, *via* carbonic anhydrase, to the gasotransmitter  $\text{H}_2\text{S}$ .<sup>28</sup> In addition, tetrazine has been used for the targeted degradation of proteins.<sup>29</sup> Recently, we and others, have shown that vinyl ethers undergo facile reaction with tetrazines resulting in elimination of the corresponding alkoxide or phenoxide.<sup>25,30–32</sup> Thus, polymeric nanoparticles, bearing a vinyl ether caged linker were shown to liberate doxorubicin upon treatment with a tetrazine, resulting in “switch-on” of cytotoxicity.<sup>30</sup>

Here, we report a new concept in prodrug activation with the simultaneous, dual, and fully traceless (except the loss of  $\text{N}_2$ ) activation/generation of two different drugs. This chemistry utilises tetrazine as a masked prodrug, which removes the vinyl ether from a second prodrug and incorporated the structural elements of the vinyl group into its own structure, giving rise to two active drugs (Fig. 1A). The chemistry explored used a tetrazine as a prodrug of a pyridazine (a common scaffold found in many drugs such as apresoline®, sulfamethoxy-pyridazine® and cadralazine®) and, in our case, generated the known microRNA 21 (miR21) inhibitor **2**,<sup>33,34</sup> leading to downregulation of oncogenic miR21 and consequently “switch-on” of apoptosis. The other prodrug (the dienophile) was the vinyl ether masked-camptothecin **3** that liberated the anticancer drug **4**, upon reaction with the tetrazine **1** (Fig. 1B). Notably, for the first time,

*EaStCHEM School of Chemistry, University of Edinburgh, Joseph Black Building, King's Buildings, David Brewster Road, EH9 3FJ Edinburgh, UK. E-mail: mark.bradley@ed.ac.uk*

† Electronic supplementary information (ESI) available. See DOI: 10.1039/c8sc02610f

‡ Current address: Laboratorium für Organische Chemie, Department of Chemistry and Applied Biosciences, ETH Zurich, 8093 Zurich, Switzerland, kevin.neumann@org.chem.ethz.ch

§ These authors contributed equally.



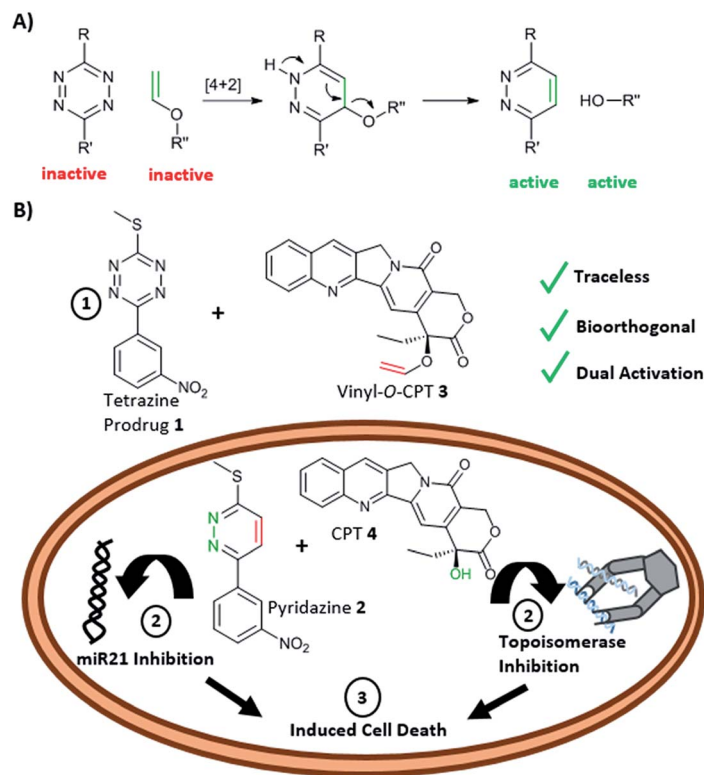


Fig. 1 (A)  $_{INVDA}$  reaction between a vinyl ether masked drug (inactive) and the tetrazine masked drug (inactive) leads to an active drug pair (a pyridazine and an alcohol). (B) Reaction between the tetrazine prodrug 1 (masked pyridazine-based miR21 inhibitor 2) and the vinyl-*O*-camptothecin 3 (caged camptothecin 4) showing the dual and traceless prodrug-prodrug generation of 2 and 4. The inhibition of microRNA 21 and topoisomerase would lead to cell death.

the tetrazine scaffold can be considered as a protecting group for bioactive pyridazines.

## Results and discussion

### Synthesis of tetrazine-prodrug

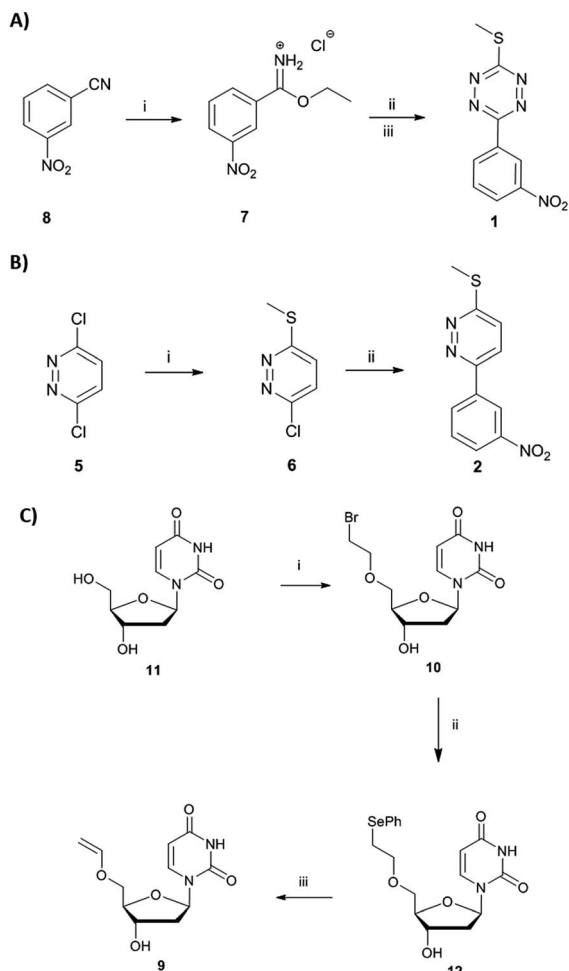
Short non-coding microRNAs (miRNA) play a critical role in several biological processes with dysregulation of levels miRNA being associated with numerous diseases, in particular cancer.<sup>35,36</sup> Oncogenic miR21 downregulates apoptosis with miRNA inhibition resulting in notable increase in apoptosis. Pyridazine 2, an miR21 inhibitor,<sup>33,34</sup> was readily synthesized in two steps, starting from 2,5-dichloropyridazine 5, via 2-chloro-5-thiomethoxydipyridazine 6 (generated by reaction with sodium thiomethoxide) followed by a Suzuki coupling with 3-nitrophenylboronic acid (Scheme 1B). Pyridazines<sup>37</sup> can also be formed via  $_{INVDA}$  reaction from the corresponding tetrazines and activated alkenes (Fig. 1). The synthesis of tetrazine 1 was achieved using 3-nitrophenyl imidoester 7 as a precursor, which was readily accessible from 3-nitrobenzotrile 8. In a facile route to tetrazines, compound 7 was treated with methyl thiocarbonylhydrazidium S7 to give 2,4-dihydrotetrazine that was oxidised *in situ* with amyl nitrite to give the tetrazine prodrug 1 (Scheme 1A). Importantly, in the case of 2, the corresponding tetrazine prodrug 1 bears electron withdrawing and donating moieties which are known to increase reactivity and elimination of the alkoxide in  $_{INVDA}$  chemistries.<sup>38</sup>

The miR21 inhibitor 2 and the tetrazine prodrug 1, were evaluated for their activity on breast, prostate and brain cancer cells (SK-BR3, PC3 and U87-MG, respectively), which all express miR21 providing a broad platform of cell lines for cytotoxicity assays.<sup>39–41</sup> Whereas miR21 is seen as an oncogenic factor in glioblastoma pathogenesis and breast cancer progression, its role in the progression of prostate cancer is not fully understood.<sup>42</sup> No influence on cell viability was observed when the cells were treated with up to 10  $\mu$ M of the tetrazine prodrug 1; however, the same concentration of miR21 inhibitor 2 resulted in reduced cell viability in all three cell lines (MTT assay, Fig. 2B). Additionally, an Annexin V assay confirmed the results by indicating early apoptotic or dead SK-BR3 cells after treatment with the miR21 inhibitor 2 (Fig. 2C).

The activation of the tetrazine prodrug 1 with a vinyl ether containing small molecule was initially investigated. We postulated that 5'-*O*-vinyl deoxyuridine 9 would be a biocompatible, non-toxic dienophile, since the resulting alcohol is a naturally occurring nucleoside. Thus, deoxyuridine 11 was selectively alkylated with 1,2-dibromoethane to give 5'-*O*-bromoethyldeoxyuridine 10. Substitution of the bromine with caesium phenylselenolate gave the phenylselenenyl ether 12,<sup>43</sup> with oxidation with periodate giving 5'-*O*-vinyl deoxyuridine 9 (Scheme 1C).

Cellular incubation of the 5'-*O*-vinyl nucleoside 9 (20  $\mu$ M) confirmed the biocompatibility of the vinyl ether with no apoptosis of SK-BR3 cells observed. The addition of tetrazine





**Scheme 1** (A) (i) HCl, EtOH/dioxane (1 : 1). (ii) Methyl thiocarbonyl drazidium S7, pyridine, DMF. (iii) Amyl nitrite, CH<sub>2</sub>Cl<sub>2</sub>. (B) (i) NaSCH<sub>3</sub>, NEt<sub>3</sub> (ii) 3-nitrophenylboronic acid, Na<sub>2</sub>CO<sub>3</sub>, Pd(dppf), dioxane/H<sub>2</sub>O (4 : 1). (C) (i) 1,2-dibromoethane, NaH, DMF. (ii) PhSeH, CsOH·H<sub>2</sub>O. (iii) (1) NaIO<sub>4</sub>, NaHCO<sub>3</sub>, CH<sub>3</sub>OH/H<sub>2</sub>O (5 : 1); (2) DIPEA, CH<sub>3</sub>CN.

prodrug **1** (10 μM) with **9** (20 μM), however, gave equivalent levels of cell death as induced by the addition of 10 μM of pure inhibitor **2** with 30% of cells being positive in an Annexin assay (Fig. 2 and S1†), thus demonstrating *in situ* prodrug activation. The biological results are in accordance with the literature,<sup>41</sup> showing a slightly enhanced effect of cancer progression by miR21 inhibition in glioblastoma and breast cancer cell lines compared with prostate cancer cell lines.

Hydrolytic stability is a critical parameter for any tetrazine targeted for biological applications and the half-life of prodrug **1** was determined to be  $2.2 \pm 0.04$  days in DMSO/PBS, some 10-fold higher than the widely used 3,6-di-2-pyridinyltetrazine **S5** ( $t_{1/2} = 0.31 \pm 0.03$  days in DMSO/PBS) (Fig. S2–S5†). Tetrazine **1** also exhibited reasonable stability in the presence of glutathione (5 mM GSH in DMSO/H<sub>2</sub>O) with 77% of **1** remaining after 3 days vs. 88% remaining without GSH (Fig. S6†).

The reactivity of tetrazine **1** in *INV*DA chemistry was investigated by determining the second order rates constants using two different dienophiles (see ESI, Scheme S2†). The reaction between tetrazine **1** and water-soluble vinyl ether **S1** displayed a significant water acceleration, which is in accordance with the literature and confirmed the biocompatibility of this reaction (Fig. S2–S5†).<sup>18,25</sup>

### Prodrug–prodrug activation

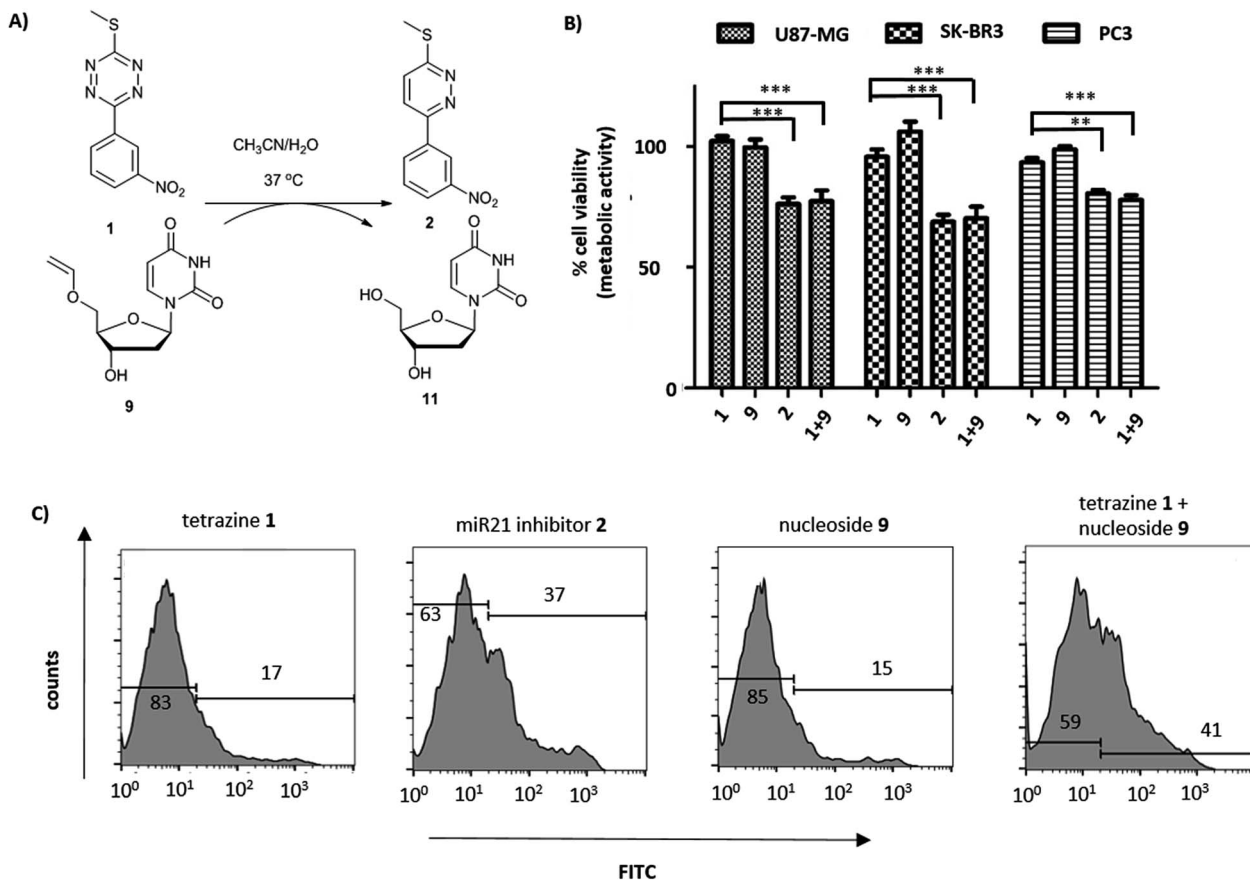
Camptothecin **4** is a topoisomerase I inhibitor that induces S-phase specific cell death. Since its discovery in the 1960's, several camptothecin derivatives and prodrugs have been reported with the aim of overcoming the drawbacks associated with camptothecin such as solubility and the stability of the lactone, which has been shown to play a crucial role in inhibiting topoisomerase I.<sup>43,44</sup> In particular, it has been shown that alkylation or acetylation of the hydroxy group at the C20 position enhances the stability of the lactone;<sup>45,46</sup> however, masking the hydroxy group of camptothecin causes a loss of its therapeutic efficiency with only a few examples known where the protecting group can be cleaved (usually by enzymatic triggering) without loss of activity.<sup>47,48</sup>

Vinyl-*O*-camptothecin **3** was synthesised in a single step procedure by slightly modifying a reported iridium-catalysed *trans*-vinylation reaction<sup>49</sup> using 1,4-dioxane to overcome the poor solubility of camptothecin **4** and an excess of vinylacetate (Fig. 3A). As postulated, masking the hydroxy group of camptothecin with a vinyl ether, caused a significant reduction in cytotoxicity, increasing the IC<sub>50</sub> from 0.15 μM to 4.6 μM for PC3 cells and from 0.18 μM to 4.9 μM for SK-BR3 cells (Fig. 3 and S11†).

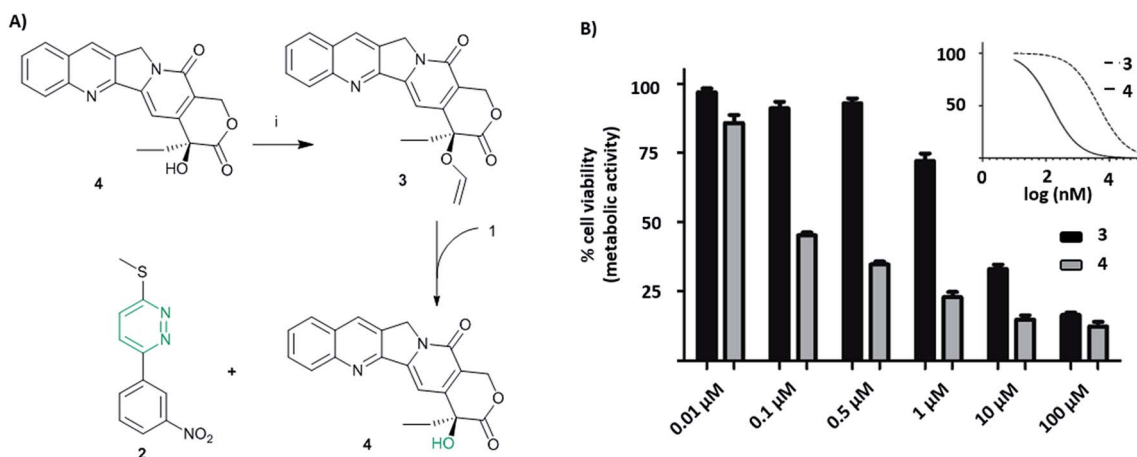
Treatment of vinyl-*O*-camptothecin **3** with the tetrazine prodrug **1** showed (monitored by HPLC) the generation of the active parent drug camptothecin **4** alongside the miR21 inhibitor **2**. HPLC analysis also indicated the formation of small quantities of the oxidised tetrazine and a peak assigned to the oxidised pyridazine (Scheme S3, Fig. S14†). Thus, this demasking generates two active drugs and resulted in controlled switch-on of cytotoxicity (Fig. 4 and S12†). Importantly, co-treatment of PC3 cells with **2** and **4** showed an additive effect beyond the decaging/activation of **1** alone with increased levels of dead cells compared to treatment with **2** or **4** (Fig. S15†). In addition, masking the hydroxyl moiety not only increased the IC<sub>50</sub> value but also compound stability. We assume that the enhanced stability of prodrug **3** leads eventually to a higher concentration of the active drug **4** at the target site, or at least longer duration of the active drug. Although the activation was slow in solution, the biological experiments suggest that the slow but constant release overtime together with the enhanced stability of **3** results in an efficient drug activation system that even with incomplete conversion delivers cytotoxicity comparable to that of the free drug. Future research will focus on prodrug pairs with even higher reactivity.

Thus, the herein presented prodrug–prodrug activation leads to an increased therapeutic window providing a concept that we believe will find wide application in drug delivery.





**Fig. 2** (A) Reaction between tetrazine 1 and 5'-O-vinyl deoxyuridine 9 (see ESI† for HPLC analysis and reaction kinetics). (B) U87-MG, SK-BR3 and PC3 cells incubated with tetrazine 1 (10  $\mu$ M), 5'-O-vinyl deoxyuridine 9 (20  $\mu$ M), miR21 inhibitor 2 (10  $\mu$ M) and tetrazine 1 (10  $\mu$ M) with 5'-O-vinyl deoxyuridine 9 (20  $\mu$ M). Cell viability measured after 72 h (MTT assay,  $n = 3$ ). \*\*\*  $P < 0.001$  and \*\*  $P < 0.01$  by one-way ANOVA with Tukey post-test. No cytotoxicity was observed for 9 up to 20  $\mu$ M; (C) flow cytometry histograms of Annexin V assay (FITC labelled) with tetrazine 1 (10  $\mu$ M), miR21 inhibitor 2 (10  $\mu$ M), 5'-O-vinyl deoxyuridine 9 (20  $\mu$ M) and tetrazine 1 (10  $\mu$ M) with 5'-O-vinyl deoxyuridine 9 (20  $\mu$ M) after 14 h of incubation with SK-BR3.



**Fig. 3** (A) (i) Camptothecin 4, vinyl acetate,  $\text{Na}_2\text{CO}_3$ ,  $[\text{Ir}(\text{cod})\text{Cl}]_2$ , 1,4-dioxane, 100  $^\circ\text{C}$ , 4 h. The reaction between tetrazine 1 and vinyl-O-camptothecin 3 gave >85% conversion ( $\text{CH}_3\text{OH}/\text{CH}_3\text{CN}/\text{H}_2\text{O}$ ) within 5 days as determined by HPLC. (B) Cell viability of PC3 cells after incubation with vinyl-O-camptothecin 3 ( $\text{IC}_{50} = 4.64 \pm 1.13 \mu\text{M}$ ) and camptothecin 4 ( $\text{IC}_{50} = 0.15 \pm 0.06 \mu\text{M}$ ) for 72 h at 37  $^\circ\text{C}$ ; insert is non-linear fit used to determine  $\text{IC}_{50}$  values (MTT assay,  $n = 3$ ).





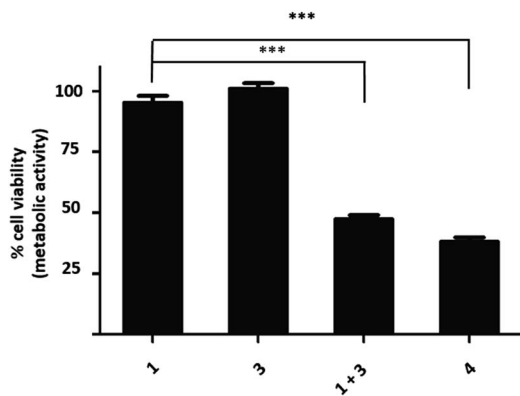


Fig. 4 Cell viability after treatment with tetrazine **1** (10  $\mu$ M) 95  $\pm$  14%, vinyl-*O*-camptothecin **3** (0.5  $\mu$ M) 101  $\pm$  10%, co-treatment of tetrazine **1** (10  $\mu$ M) and vinyl-*O*-camptothecin **3** (0.5  $\mu$ M) 47  $\pm$  8%, camptothecin **4** (0.5  $\mu$ M) 38  $\pm$  5%, (PC3, MTT-assay,  $n = 3$ ) \*\*\*  $P < 0.001$  by one-way ANOVA with Tukey post-test.

## Conclusions

In summary, we report for the first time a symbiotic prodrug-prodrug activation strategy that fully complies with the principle of bioorthogonality. To illustrate the power of this new strategy, we showed that a tetrazine prodrug scaffold was converted into a pyridazine-based miR21 inhibitor upon reaction and decaging of a vinyl ether masked camptothecin. This demasking takes advantage of the water acceleration effect (for water dependency of kinetics see Fig. S1†), which has been widely exploited and acknowledged in tetrazine chemistry<sup>18,25</sup> and results in the activation of two drugs without the generation of by-products, such as the phosphine oxide seen in the Staudinger ligation. Since drug resistance is a major concern in anti-cancer therapy, which has been linked to an overexpression of miRNA,<sup>50</sup> activation of a conventional anti-cancer drug such as camptothecin in concert with a miR21 inhibitor, offers a new bioorthogonal prodrug-prodrug activation strategy and is an exceptionally atom efficient method of prodrug activation. The dual/traceless prodrug-prodrug activation strategy opens up new possibilities and directions in the field of drug delivery, in particular in the field of combination therapy (administration of two or more drugs) that is the most common clinical used strategy in cancer therapy.

It should be noted that the herein presented prodrug-prodrug activation is not only suitable for hydroxyl and pyridazine containing drugs. One could imagine, for example that the traceless Staudinger ligation could be utilised in a similar manner leading to free drugs containing amines and organo phosphorous moieties, e.g. cyclophosphamides. In a broader context, the prodrug-prodrug approach presented here is not limited to the treatment of cancer and could be useful as a combination approach in other therapeutic areas.

## Conflicts of interest

There are no conflicts to declare.

## Acknowledgements

This work was supported by the European Research Council (Advanced Grant ADREEM ERC-2013-340469).

## Notes and references

- J. Rautio, H. Kumpulainen, T. Heimbach, R. Oliyai, D. Oh, T. Järvinen and J. Savolainen, *Nat. Rev. Drug Discovery*, 2008, **7**, 255–270.
- J. Prescher and C. R. Bertozzi, *Nat. Chem. Biol.*, 2005, **1**, 13–21.
- K. M. Dean and A. E. Palmer, *Nat. Chem. Biol.*, 2014, **10**, 512–523.
- E. M. Sletten and C. R. Bertozzi, *Angew. Chem., Int. Ed.*, 2009, **48**, 6974–6998.
- J. Li and P. R. Chen, *Nat. Chem. Biol.*, 2016, **12**, 129–137.
- K. Gorska, A. Manicardi, S. Barluenga, N. Winssinger, J. J. Hyldig-Nielsen, G. Zon, D. H. Ly, S. K. Kim, B. Norden and P. E. Nielsen, *Chem. Commun.*, 2011, **47**, 4364–4366.
- R. van Brakel, R. C. M. Vulders, R. J. Bokdam, H. Grüll and M. S. Robillard, *Bioconjugate Chem.*, 2008, **19**, 714–718.
- S. S. Matikonda, D. L. Orsi, V. Staudacher, I. A. Jenkins, F. Fiedler, J. Chen and A. B. Gamble, *Chem. Sci.*, 2015, **6**, 1212–1218.
- T. Peng and H. C. Hang, *J. Am. Chem. Soc.*, 2016, **138**, 14423–14433.
- M. C. Uzagare, I. Claußnitzer, M. Gerrits and W. Bannwarth, *ChemBioChem*, 2012, **13**, 2204–2208.
- S. T. Laughlin and C. R. Bertozzi, *Nat. Protoc.*, 2007, **2**, 2930–2944.
- A. B. Neef and C. Schultz, *Angew. Chem., Int. Ed.*, 2009, **48**, 1498–1500.
- C.-X. Song and C. He, *Acc. Chem. Res.*, 2011, **44**, 709–717.
- B. L. Oliveira, Z. Guo, O. Boutoureira, A. Guerreiro, G. Jiménez-Osés and G. J. L. Bernardes, *Angew. Chem., Int. Ed.*, 2016, **55**, 14683–14687.
- B. M. Zeglis, K. K. Sevak, T. Reiner, P. Mohindra, S. D. Carlin, P. Zanzonico, R. Weissleder and J. S. Lewis, *J. Nucl. Med.*, 2013, **54**, 1389–1396.
- L. Carroll, H. L. Evans, E. O. Aboagye and A. C. Spivey, *Org. Biomol. Chem.*, 2013, **11**, 5772–5781.
- M. L. Blackman, M. Royzen and J. M. Fox, *J. Am. Chem. Soc.*, 2008, **130**, 13518–13519.
- A. C. Knall and C. Slugovc, *Chem. Soc. Rev.*, 2013, **42**, 5131–5142.
- J. C. T. Carlson, L. G. Meimetis, S. A. Hilderbrand and R. Weissleder, *Angew. Chem., Int. Ed.*, 2013, **52**, 6917–6920.
- L. G. Meimetis, J. C. T. Carlson, R. J. Giedt, R. H. Kohler and R. Weissleder, *Angew. Chem., Int. Ed.*, 2014, **53**, 7531–7534.
- A. Wiczorek, P. Werther, J. Euchner and R. Wombacher, *Chem. Sci.*, 2017, **8**, 1506–1510.
- B. M. Zeglis, C. Brand, D. Abdel-Atti, K. E. Carnazza, B. E. Cook, S. Carlin, T. Reiner and J. S. Lewis, *Mol. Pharmaceutics*, 2015, **12**, 3575–3587.



- 23 J.-P. Meyer, J. L. Houghton, P. Kozłowski, D. Abdel-Atti, T. Reiner, N. V. K. Pillarsetty, W. W. Scholz, B. M. Zeglis and J. S. Lewis, *Bioconjugate Chem.*, 2016, **27**, 298–301.
- 24 C. Denk, D. Svatoněk, S. Mairinger, J. Stanek, T. Filip, D. Matscheko, C. Kuntner, T. Wanek and H. Mikula, *Bioconjugate Chem.*, 2016, **27**, 1707–1712.
- 25 M. Staderini, A. Gambardella, A. Lilienkamp and M. Bradley, *Org. Lett.*, 2018, **20**, 3170–3173.
- 26 R. M. Versteegen, R. Rossin, W. ten Hoeve, H. M. Janssen and M. S. Robillard, *Angew. Chem., Int. Ed.*, 2013, **52**, 14112–14116.
- 27 R. Rossin and M. S. Robillard, *Curr. Opin. Chem. Biol.*, 2014, **21**, 161–169.
- 28 A. K. Steiger, Y. Yang, M. Royzen and M. D. Pluth, *Chem. Commun.*, 2017, **53**, 1378–1380.
- 29 H. Lebraud, D. J. Wright, C. N. Johnson and T. D. Heightman, *ACS Cent. Sci.*, 2016, **2**, 927–934.
- 30 K. Neumann, S. Jain, A. Gambardella, S. E. Walker, E. Valero, A. Lilienkamp and M. Bradley, *ChemBioChem*, 2017, **18**, 91–95.
- 31 E. Jiménez-Moreno, Z. Guo, B. L. Oliveira, I. S. Albuquerque, A. Kitowski, A. Guerreiro, O. Boutureira, T. Rodrigues, G. Jiménez-Osés and G. J. L. Bernardes, *Angew. Chem., Int. Ed.*, 2017, **56**, 243–247.
- 32 H. Wu, S. C. Alexander, S. Jin and N. K. Devaraj, *J. Am. Chem. Soc.*, 2016, **138**, 11429–11432.
- 33 Q. Huang, A. Deiters and K. Gumireddy, *Microna modulators and method for identifying and using the same*, WO 2013019469 A1, 2012.
- 34 K. Gumireddy, D. D. Young, X. Xiong, J. B. Hogenesch, Q. Huang and A. Deiters, *Angew. Chemie Int. Ed.*, 2008, **47**, 7482–7484.
- 35 A. Esquela-Kerscher and F. J. Slack, *Nat. Rev. Cancer*, 2006, **6**, 259–269.
- 36 G. A. Calin and C. M. Croce, *Nat. Rev. Cancer*, 2006, **6**, 857–866.
- 37 C. G. Wermuth, C. Vergelli, C. Biancalani, N. Cesari, A. Graziano, P. Biagini, J. Gracia, A. Gavalda, V. D. Paz and C. Norton, *MedChemComm*, 2011, **2**, 935.
- 38 X. Fan, Y. Ge, F. Lin, Y. Yang, G. Zhang, W. S. C. Ngai, Z. Lin, S. Zheng, J. Wang and J. Zhao, *Angew. Chem., Int. Ed.*, 2016, **55**, 14046–14050.
- 39 G. Shi, D. Ye, X. Yao, S. Zhang, B. Dai, H. Zhang, Y. Shen, Y. Zhu, Y. Zhu and W. Xiao, *Acta Pharmacol. Sin.*, 2010, **31**, 867–873.
- 40 L. Shi, J. Chen, J. Yanga, T. Pan, S. Zhang and Z. Wang, *Brain Res.*, 2010, **1352**, 255–264.
- 41 L. X. Yan, Q. N. Wu, Y. Zhang, Y. Y. Li, D. Z. Liao, J. H. Hou, J. Fu, M. S. Zeng, J. P. Yun, Q. L. Wu, Y. X. Zeng and J. Y. Shao, *Breast Cancer Res.*, 2011, **13**, R2.
- 42 M. Folini, P. Gandellini, N. Longoni, V. Profumo, M. Callari, M. Pennati, M. Colecchia, R. Supino, S. Veneroni, R. Salvioni, R. Valdagni, M. Grazia Daidone and N. Zaffaroni, *Mol. Cancer*, 2010, **9**, 12.
- 43 R. J. Cohen, D. L. Fox and R. N. Salvatore, *J. Org. Chem.*, 2004, **69**, 4265–4268.
- 44 C. Jin, S. Wen, Q. Zhang, Q. Zhu, J. Yu and W. Lu, *ACS Med. Chem. Lett.*, 2017, **8**, 762–765.
- 45 M. Deshmukh, P. Chao, H. L. Kutscher, D. Gao and P. J. Sinko, *J. Med. Chem.*, 2010, **53**, 1038–1047.
- 46 R. P. Hertzberg, M. J. Caranfa, K. G. Holden, D. R. Jakas, G. Gallagher, M. R. Mattern, S. M. Mong, J. O. Bartus, R. K. Johnson and W. D. Kingsbury, *J. Med. Chem.*, 1989, **32**, 715–720.
- 47 A. Gopin, S. Ebner, B. Attali and D. Shabat, *Bioconjugate Chem.*, 2006, **17**, 1432–1440.
- 48 B. Schmid, D. Chung, A. Warnecke, I. Fichtner and F. Kratz, *Bioconjugate Chem.*, 2007, **18**, 702–716.
- 49 Y. Okimoto, S. Sakaguchi and Y. Ishii, *J. Am. Chem. Soc.*, 2002, **124**, 1590–1591.
- 50 J. Ma, C. Dong and C. Ji, *Cancer Gene Ther.*, 2010, **17**, 523–531.

