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# Synthesis of novel 10,11-methylenedioxy-camptothecin glycoside derivatives and investigation of their anti-tumor effects *in vivo*†

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10,11-Methylenedioxy-camptothecin (FL118) is a novel camptothecin analogue that possesses exceptional antitumor efficacy in human tumor xenograft models. The aim of the current study was to develop novel 20-substituted FL118 derivatives coupled with glycosyl-succinic acid esters with improved antitumor efficacy. These FL118 glycoside derivatives were designed, synthesized and their cytotoxicity evaluated in three tumor cell lines (A-549, MDA-MB-231 and RM-1). All of the derivatives showed superior *in vitro* cytotoxic activity and were more potent than irinotecan in A549 and MDA-MB-231 cells. In mouse prostate cancer cells RM-1, 10,11-methylenedioxy-camptothecin rhamnoside **11b** displayed significant activities with IC<sub>50</sub> of 48.27 nM. Western blot analysis demonstrated that **11b** inhibited survivin expression and induced cancer cells apoptosis. Further cell cycle analyses clearly showed **11b** induced G2/M phase cell cycle arrest. Molecule docking studies suggested that the binding mode of **11b** was different from that of the crystal complex of ligand topotecan in Top1/DNA. Importantly, **11b** showed high *in vivo* antitumor efficacy in the RM-1 mouse model with transplantation of prostate cancer (TGI = 44.9%) at dose of 9 mg kg<sup>-1</sup> without apparent toxicity.

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## 1. Introduction

20(*S*)-Camptothecin (CPT) is a cytotoxic quinoline alkaloid (Fig. 1) that was discovered by Wall *et al.* in 1966.<sup>1</sup> Over the past five decades, many CPT derivatives were synthesized and tested, but most of the compounds designed so far are highly toxic to normal tissues or have other shortcomings making them unsuitable candidates for cancer treatment. To date, only two camptothecin analogues (*i.e.* irinotecan and topotecan, Fig. 1) have been commercially approved by the FDA for treatment of cancer in clinic.<sup>2,3</sup> However, resistance to irinotecan and topotecan is often observed in practice, especially in patients using these drugs for an extended period of time.<sup>4–9</sup>

The anticancer agent 10,11-methylenedioxy-camptothecin (FL118), a CPT analogue with a methylenedioxy group linked

to positions 10 and 11 of the A-ring (Fig. 1), was recently identified through small molecule inhibitor screening and shows much higher anticancer activities in several different cancer types both *in vitro* and *in vivo*.<sup>10</sup> It has been previously demonstrated that although FL118 is not a better Top1 inhibitor than the clinically used camptothecin analogues,<sup>10</sup> FL118 is able to selectively inhibit multiple cancer survival and proliferation-

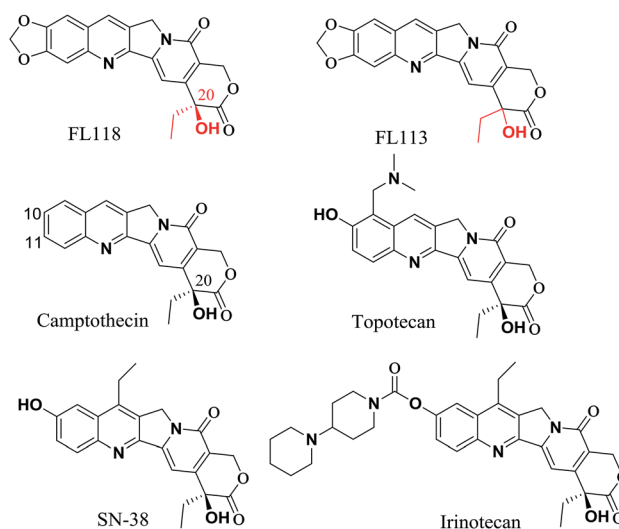


Fig. 1 Comparison chemical structure of FL118 with FL113, camptothecin, irinotecan, SN-38 (active metabolite of irinotecan) and topotecan.

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associated antiapoptotic proteins (survivin,<sup>11–21</sup> XIAP,<sup>22–26</sup> and cIAP2 (ref. 27 and 28)) and the Bcl-2 family (Mcl-1 (ref. 29–35)), which contribute to FL118 function and antitumor activity.<sup>11,36</sup>

The superior antitumor efficacy of FL118 has inspired our interest in the development of antitumor drugs using the core structure of FL118 as a promising scaffold for the generation of novel FL118 analogs. Results from previous studies suggest that modification of the free hydroxyl group at 20-position in camptothecin *via* ester bonds could be a promising way to improve *in vivo* antitumor efficacy and reduce gastrointestinal toxicity.<sup>37–40</sup> The idea of replacing the hydrogen atom in the hydroxyl group of FL118 with saccharide is to improve its water solubility and further decrease its normal tissue toxicity without affecting its antitumor activity.

In the present study, different configurations of 10,11-methylenedioxy-camptothecin glycosyl substituted analogs were synthesized and their antitumor activity was evaluated. The two fragments, including saccharide moiety and succinic acid ester, were incorporated into the structure at the 20-position of 10,11-methylenedioxy-camptothecin to improve its water solubility.

## 2. Results and discussion

### 2.1 Chemistry

According to published procedures,<sup>41,42</sup> the glycosyl donors 2,3,4,6-tetra-*O*-acetyl-*D*-glucopyranose **3a** or 2,3,4-tri-*O*-acetyl-*L*-rhamnose **3b** was prepared from *D*-glucose **1a** or *L*-rhamnose **1b** in 68% or 63% yield, respectively. The obtained compound **3a** or **3b** was then reacted with succinic anhydride to form  $\beta$ -*O*-(2,3,4,6-tetra-*O*-acetyl- $\beta$ -*D*-glucosyl)-succinic acid monoester **4a** by the yield of 89% or  $\alpha$ -*O*-(2,3,4-tri-*O*-acetyl- $\alpha$ -*L*-rhamnosyl)-succinic acid monoester **4b** by the yield of 87% (Scheme S1†).

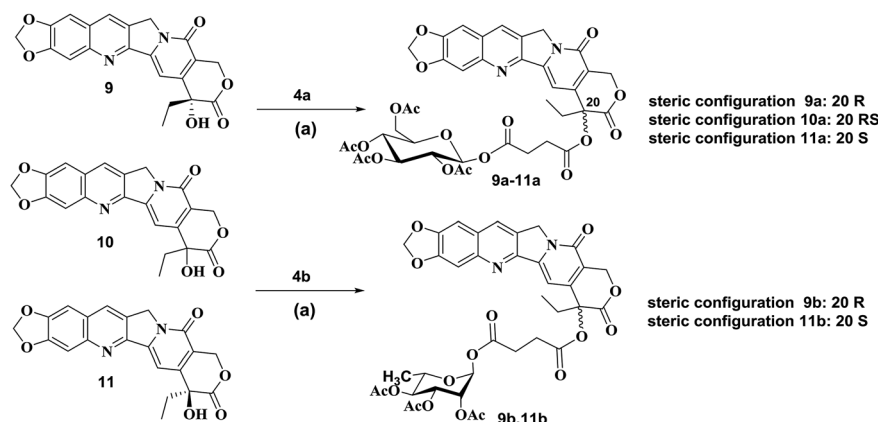
The synthetic routes to form target compounds (**9a–11a**, **9b**, **11b**) are outlined in Scheme 1. 2-Amino-4,5-methylenedioxybenzaldehyde **5** was prepared according to the published procedures.<sup>43,44</sup> Different configurations of compounds 20(*R*)-10,11-methylenedioxy-camptothecin **9**, 20(*RS*)-10,11-methylenedioxy-camptothecin **10** and 20(*S*)-10,11-methylenedioxy-camptothecin **11** was accomplished using Friedlander condensation with 80–85% yield between 6-amino

piperonal **5** and the known tricyclic keto lactone **6**, **7** or **8**, respectively (Scheme S2†).<sup>45,46</sup> Coupling of compound **9** with glycosyl donors **4a** or **4b**, catalyzed by EDCI and DMAP, resulted in pure *R* steric configuration 10,11-methylenedioxy-camptothecin glucoside **9a** ( $[\alpha]_D^{14} = 284.25^\circ$ ) or rhamnoside **9b** ( $[\alpha]_D^{14} = 98.75^\circ$ ) in 68% and 71% yield, respectively. Similarly, coupling of compound **10** with glycosyl donor **4a**, resulted in racemic *RS* configuration 10,11-methylenedioxy-camptothecin glucoside **10a** ( $[\alpha]_D^{14} = 32.75^\circ$ ) with a 63% yield. The pure *S* steric configuration 10,11-methylenedioxy-camptothecin glycoside **11a** ( $[\alpha]_D^{14} = -27.75^\circ$ ) and **11b** ( $[\alpha]_D^{14} = -166.50^\circ$ ) were obtained by coupling of compound **11** with glycosyl donors **4a** and **4b** in 61% and 73% yield, respectively (Scheme 1).

### 2.2 Cytotoxicity and IC<sub>50</sub>

The *in vitro* cytotoxicity of the newly synthesized compounds were determined in three tumor cell lines, A-549 (human lung carcinoma), MDA-MB-231 (human breast carcinoma) and RM-1 (mouse prostate carcinoma) while irinotecan and FL118 were used as positive controls. To obtain IC<sub>50</sub> values for each compound, all three cancer cell lines were treated with series of concentrations of each compound. As shown in Table 1, all target compounds exhibited significant cytotoxic activities against the three tumor cell lines *in vitro* with IC<sub>50</sub> values ranging from 2.32 nM to 4.53  $\mu$ M. All of the new designed compounds were less potent than FL118, the core structure of these synthesized compounds, against A549 and MDA-MB-231 cell lines. In comparison, all the new compounds exhibited superior cytotoxicity to irinotecan in the two human cell lines. In addition, all of the compounds were more potent against A549 cells, moreover, 10,11-methylenedioxy-camptothecin rhamnoside **11b** (the *S*-type enantiomer) showed the most significant cytotoxic effect with IC<sub>50</sub> of 83 nM. The IC<sub>50</sub> values in Table 1 indicated that the A-549 cell line was more sensitive to these compounds than the MDA-MB-231 cell lines.

For both human cancer cell lines, the compounds coupled with rhamnose (**9b** and **11b**) were generally more potent than those coupled with glucose (**9a** and **11a**), indicating that the rhamnose may be a better group in developing new antitumor agents. Furthermore, the configuration of the 20-position is



Scheme 1 Synthesis of compounds **9a–11a**, **9b**, **11b**. Reagents and conditions: (a) EDCI, DMAP, CH<sub>2</sub>Cl<sub>2</sub>, reflux.



**Table 1** IC<sub>50</sub> values for the compounds in two human cancer cell lines (A549, MDA-MB-231) and the mouse prostate cancer cell lines (RM-1)

Comp.	<i>In vitro</i> IC <sub>50</sub> (nM) <sup>a</sup>		
	A549	MDA-MB-231	RM-1
<b>9a</b>	1612.50 ± 276.48	4527.40 ± 174.66	>1000
<b>9b</b>	604.50 ± 208.17	1300.50 ± 276.48	>1000
<b>10a</b>	102.18 ± 33.41	213.00 ± 81.46	>1000
<b>11a</b>	135.80 ± 61.10	252.40 ± 70.75	>1000
<b>11b</b>	83.34 ± 11.04	154.50 ± 35.50	48.27 ± 6.25
FL118	8.94 ± 1.54	24.73 ± 13.82	69.19 ± 8.34
Irinotecan	9140.30 ± 1054.87	7817.50 ± 2386.18	>1000

<sup>a</sup> Each IC<sub>50</sub> value was calculated from 3 independent experiments performed in triplicate. Data are shown as mean ± SD.

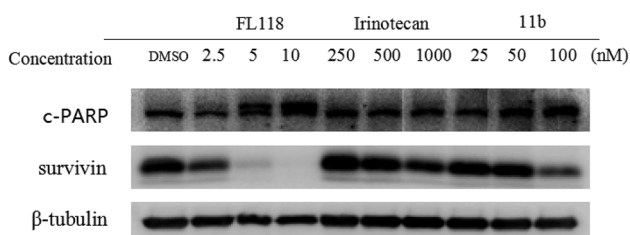
crucial for its activity. Activity of the pure *S*-configuration derivatives (**11a** and **11b**) was higher than the pure *R*-configuration derivatives (**9a** and **9b**) in the two human cancer cell lines. In A549 cells, the mixture of *R*- and *S*- configuration derivative (**10a**) was the most active compound among the three compounds (**9a**, **10a**, **11a**). However, these compounds were less effective than FL118 in A549 and MDA-MB-231 cells. For RM-1 mouse prostate cancer cells, compounds **11b** (IC<sub>50</sub>: 48.27 nM) showed the greatest cytotoxic effects, whilst the other glycoside compounds showed lower inhibitory activity at 1 μM (Table 1).

### 2.3 Effects of compound 11b on cell apoptosis

To study how **11b** affected the cell apoptosis, human lung cancer cells A549 were treated with FL118, irinotecan or **11b** for 48 h, respectively (Fig. 2). Specifically, the data revealed that treatment of cancer cells with **11b** and FL118 resulted in the downregulation of survivin, while irinotecan treatment showed minimal effect on survivin. Furthermore, **11b** and FL118 treatment increased the production of PARP cleavage in a concentration-dependent manner, which is the hallmark of apoptosis.

### 2.4 Effects of compound 11b on cell cycle

Since survivin is a central molecule for normal cell cycle progression in most of cancer cells, we next tested how the compound **11b** affected the cell cycles in the A549 lung cancer cells by flow cytometry analysis. When A549 cells were treated

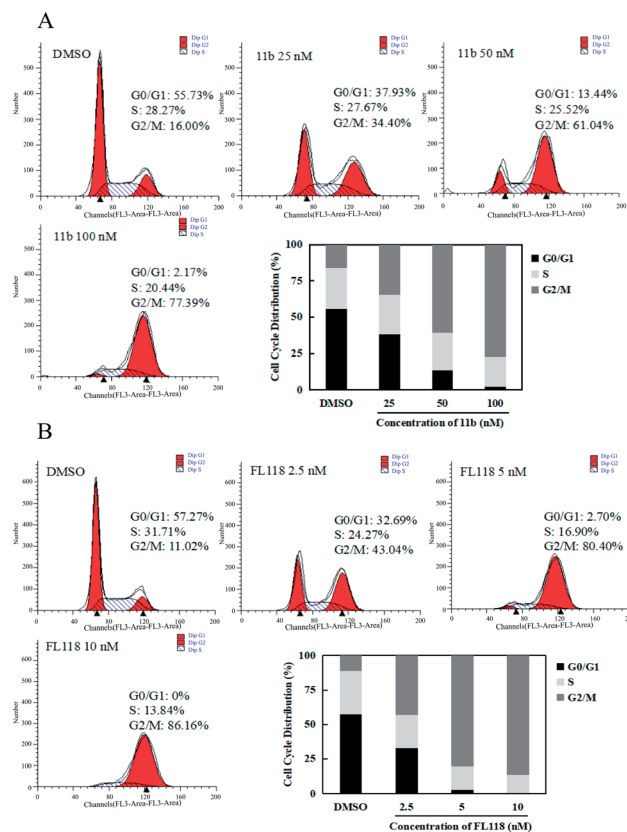


**Fig. 2** Effects of compounds irinotecan, FL118 and **11b** on cell apoptosis related proteins c-parp and survivin in A549 human lung cancer cells. The cells were seeded in 15 cm dishes for 24 h, and then cells were treated with serial concentrations of three compounds, respectively. After 48 h of incubation, cells were harvested for western blotting analysis. β-Actin was served as an equal loading control.

with **11b** from 25 nM to 100 nM for 48 h, the population of cells in G2/M phase dramatically increased as compared to that of vehicle (DMSO) group (Fig. 3A), along with concomitant losses in the G1 phase. As shown in Fig. 3B, compound FL118 at the concentrations of 2.5 nM, 5 nM, and 10 nM also increased G2/M cell population in a concentration-dependent manner. The cell cycle data clearly showed compound **11b** arrested A549 cells mainly at the G2/M phase.

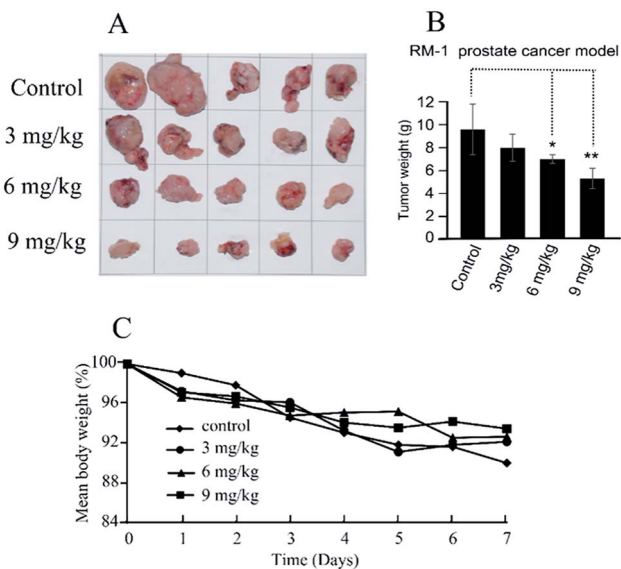
### 2.5 *In vivo* growth inhibition

The *in vivo* antitumor efficacy of compound **11b** was analyzed using RM-1 mouse model with transplantation of prostate cancer at doses of 3, 6 and 9 mg kg<sup>-1</sup> per dose. Treatment was initiated 10 days after subcutaneous tumor implantation when the individual tumors had grown to about 200–300 mm<sup>3</sup>. Compound **11b** was administered by intratumoral injection to the mice (three times, every other day) for 7 days. As shown in Fig. 4, compound **11b** showed superior antitumor activity *in vivo* at a dose of 9 mg kg<sup>-1</sup>, whilst this concentration was much lower than the clinical dose of irinotecan (200 mg kg<sup>-1</sup>) and topotecan (12.5 mg kg<sup>-1</sup>). Tumor growth inhibition (TGI) was calculated at the end of the treatment. Compound **11b** demonstrated significant antitumor activity (TGI = 44.9%) at



**Fig. 3** A cell cycle distribution of A549 human lung cancer cells after treatments with compound **11b** (A) and FL118 (B). The cells were seeded in 6-well plates for 24 h, and then the cells were treated with compound **11b** or FL118. After 48 h of treatments, cells were harvested and subject to cell cycle analyses as described in the Experimental section. Statistical analysis was conducted among control and treated groups in G0/G1, S, and G2/M phases, separately.



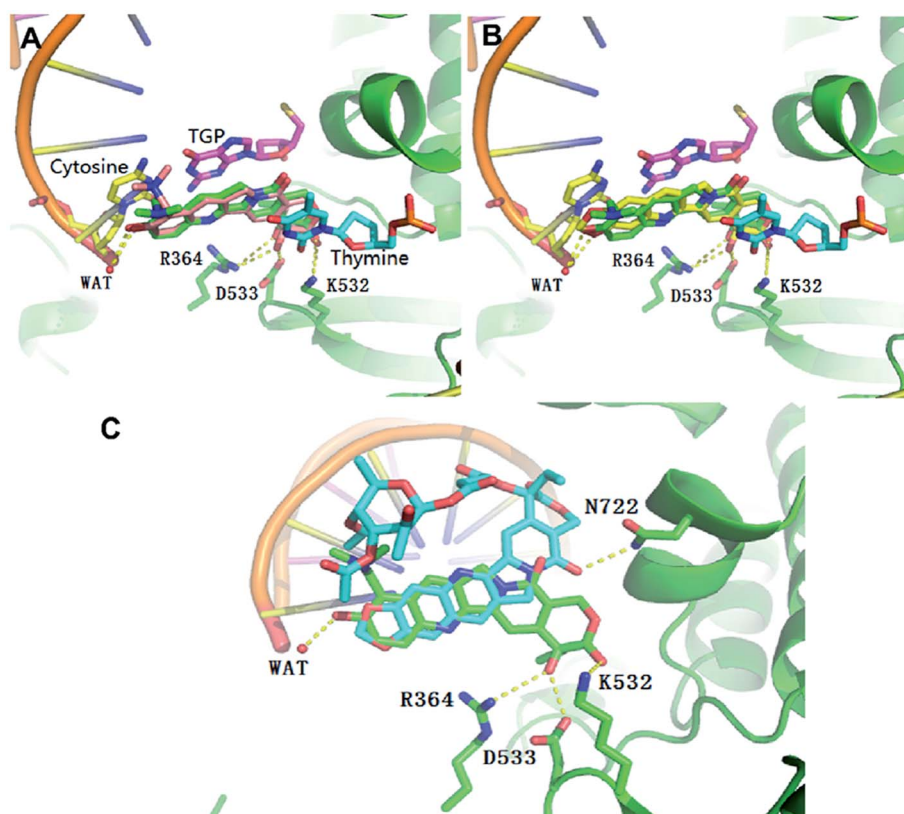


**Fig. 4** *In vivo* antitumor effects of compound **11b** in the mouse prostate tumor of RM-1 cancer model. (A) Images of excised tumors in each group. (B) Weight of the excised tumors in each group (\*\*\* $P < 0.001$ , \*\* $P < 0.01$ , \* $P < 0.05$ ; Student's *t*-test). Data are expressed as the mean  $\pm$  standard deviation. (C) The mean mouse body weight curves derived from five mice treated with vehicle (control) or with one of the three doses of **11b**. Of note, the standard error (SE) of body weight loss variation is within 10% among each group.

a dose of  $9 \text{ mg kg}^{-1}$ . It also can be seen from Fig. 4C that all the groups had nearly the same body weights, indicating the low toxicity of **11b** toward mice.

## 2.6 Molecule docking studies

In order to understand the different molecular binding mechanism of compounds topotecan, FL118 and **11b** in Top1/DNA complex, computational docking was performed using the Top1/DNA complex (PDB code: 1K4T).<sup>47</sup> The docking study showed that the binding mode of topotecan was similar to that of the crystallographic topotecan in Top1/DNA complex (Fig. 5A), with a stacking interaction between 5-thio-2-deoxyguanosine phosphonic acid (TGP)/cytosine on one side of the ligand and a thymine base pair on the opposite side of the ligand. The topotecan interacts with Asp533, Arg364, Lys532 and a water molecule through a hydrogen bonding interaction. As shown in Fig. 5B, the predicted binding pose of FL118 is shown to have a similar disposition with respect to the interacting residues. The MOE docking energy of FL118 is about  $1.28 \text{ kcal mol}^{-1}$  higher than the SN-38 in the crystal structure suggesting that FL118 is significantly less active than the SN-38 for targeting the DNA topoisomerase I (Table S1†).<sup>10</sup> 10,11-Methylenedioxy-camptothecin rhamnoside **11b** displayed an energetically less favourable binding mode with the Top1/DNA complex (Fig. 5C), which indicates **11b** may also show lower topoisomerase I inhibition activity than topotecan.



**Fig. 5** Overlap of the binding mode of compounds topotecan (pink) (A), FL118 (yellow) (B), **11b** (blue) (C) with the crystallographic topotecan (green) in Top1/DNA complex (PDB code: 1K4T). The oxygen, nitrogen, and sulfur atoms of compounds are shown in red, blue, and yellow, respectively. The side chains of the binding site are colored according to the atom types (carbon, green; oxygen, red; nitrogen, blue) and the hydrogen bonds are shown as dashed lines. The docking poses were visualized using PyMOL1.8.



Table 2 Calculated ADMET descriptors related to absorption and distribution properties<sup>a</sup>

Comp.	CNS	log HERG	PCaco	log B/B	PMDCK	F%
<b>9a</b>	-2	-5.213	159.608	-2.027	68.069	44.6
<b>11a</b>	-2	-6.569	39.646	-3.132	15.107	32.978
<b>9b</b>	-2	-6.666	68.091	-2.827	27.106	41.469
<b>11b</b>	-2	-7.012	43.907	-3.17	16.869	38.065

<sup>a</sup> CNS: -2 (inactive), +2 (active); log HERG > -5; PCaco: < 25 poor, > 500 great; log B/B (-3-1.2); PMDCK: < 25 poor, > 500 great.

### 2.7 *In silico* ADMET predictions

Nowadays, the computational prediction of descriptors representing absorption, distribution, metabolism, excretion and toxicity properties (ADMET) are considered useful *in silico* tools, decreasing the proportion of drug candidates that can fail in clinical trials for ADMET reasons. Owing to the excellent *in vitro* activity, we initiated *in silico* calculations for ADMET prediction of compounds. As shown in Table 2, the predicted central nervous system (CNS) activity was computed on a -2 (inactive) to +2 (active) scale and showed that all these four compounds (**9a**, **11a**, **9b** and **11b**) could be CNS inactive due to low values. The blood/brain partition coefficients (log B/B) were computed and also indicated all the four derivatives were difficult to access to the CNS. Human ether-a-go-go related gene (HERG) K<sup>+</sup> channel blockers are potentially toxic. The recommended range for predicted log IC<sub>50</sub> values for blockage of HERG K<sup>+</sup> channels is > -5. The log HERG values (-5.213 to -7.012) for compounds (**9a**, **11a**, **9b** and **11b**) indicate that these compounds could show cardiac toxicity. Madin-Darby canine kidney (MDCK) monolayers are widely used to make oral absorption estimates. The values showed that compounds (**9a**, **11a**, **9b** and **11b**) had poor MDCK cell permeability. However, all these four derivatives showed good Caco-2 permeability (PCaco).

## 3. Conclusions

In summary, a series of 20-substituted-10,11-methylenedioxy-camptothecin glycoside derivatives were synthesized and their cytotoxic activities were evaluated against three tumor cell lines (A-549, MDA-MB-231, RM-1). All of the synthesized compounds showed superior cytotoxic activity to the irinotecan, but were less potent than FL118 in the two human cancer cell lines (A549 and MDA-MB-231). In the mouse prostate cancer cell model RM-1, 10,11-methylenedioxy-camptothecin rhamnoside **11b** displayed excellent activity with IC<sub>50</sub> of 48.27 nM, which is lower than that for FL118 and irinotecan. Western blot analysis indicated **11b** inhibited survivin expression and increased the expression of apoptotic marker PARP cleavage. Cell cycle analyses showed that **11b** induced A549 lung cancer cell cycle arrest in a concentration-dependent manner. Molecular docking studies suggested that the binding mode of **11b** was different from that of the crystal ligand topotecan in Top1/DNA complex. In addition, compound **11b** exhibited significant antitumor effects in the RM-1 prostate cancer model *in vivo* (TGI = 44.9%) at a dose of 9 mg kg<sup>-1</sup>. Overall, our findings provide strong

evidence that FL118 is a promising scaffold for design of novel anti-tumor compounds with high anti-tumor efficacy.

## 4. Experimental

### 4.1 Chemistry

**Materials and methods.** All reagents used in the experiments were obtained from commercial sources and purified in a conventional manner. Column chromatography was performed using silica gel (200–300 mesh) purchased from MeiGao Ltd. (Qingdao, China). TLC was performed on a silica gel plate purchased from Merck Ltd. (Darmstadt, Germany). Melting points were determined with an X-4 digital micro melting point tester (Taike Ltd., Beijing, China) and were uncorrected. <sup>1</sup>H NMR, <sup>13</sup>C NMR spectra were acquired using Jeol JNM-ECP 600 spectrometer (Jeol Ltd., Tokyo, Japan) with tetramethylsilane (Me<sub>4</sub>Si) as the internal standard. Chemical shifts were recorded as δ values in ppm. The following abbreviations were used: s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet, dd = double-doublet, dt = double-triplet, tt = triple-triplet. Mass spectra were recorded on a Q-TOF global mass spectrometer and the ionization method for all compounds below was electrospray ionization (ESI).

**β-O-(2,3,4,6-Tetra-O-acetyl-β-D-glucosyl)-succinic acid monoester 4a.** To a stirred solution of **3a** (2 mmol) in dry THF (20 mL) was slowly added succinic anhydride (3 mmol) and DMAP (0.05 mmol) at room temperature. After being stirred for 24 h, the solvent was removed on a rotary evaporator and EtOAc (30 mL) was poured into the residue. The solution was washed with HCl (1 M), brine, dried and concentrated. The oil was purified by flash chromatography on silica gel (petroleum ether/ethyl acetate: 5/1) to afford **4a** as a colorless oily liquid with a 89% yield. <sup>1</sup>H NMR (CHLOROFORM-D, 600 MHz) δ: 12.87 (s, 1H), 5.64 (d, J = 7.6 Hz, 1H), 5.25 (dd, J = 11.0, 3.2 Hz, 1H), 5.15 (d, J = 3.7 Hz, 1H), 4.96 (dd, J = 11.0, 3.7 Hz, 1H), 4.35 (t, J = 6.4 Hz, 1H), 4.04 (m, 2H), 2.96 (t, J = 7.5 Hz, 2H), 2.48 (t, J = 7.5 Hz, 2H), 2.17 (s, 3H), 2.10 (s, 3H), 2.07 (s, 3H), 2.03 (s, 3H).

**α-O-(2,3,4-Tri-O-acetyl-α-L-rhamnosyl)-succinic acid monoester 4b.** According to the same synthesis procedure of compound **4a**, compounds **4b** were obtained as a colorless oily liquid in 87% yield. <sup>1</sup>H NMR (CHLOROFORM-D, 600 MHz) δ: 5.44 (dd, J = 10.6, 3.3 Hz, 1H), 5.22 (m, 1H), 5.09 (t, J = 10.6 Hz, 1H), 4.64 (d, J = 1.5 Hz, 1H), 3.98 (m, 1H), 2.97 (t, J = 6.5 Hz, 2H), 2.58 (t, J = 6.5 Hz, 2H), 2.16 (s, 3H), 2.06 (s, 3H), 2.00 (s, 3H), 1.23 (d, J = 5.9 Hz, 3H).

#### General procedure for the synthesis of 9–11

*Taking the 9 for example.* A round bottom flask, **5** (1.06 g, 4 mmol), **6** (1.2 g, 7.2 mmol) and 4-methylbenzenesulfonic acid (0.16 g, 0.8 mmol) were dissolved in toluene (200 mL). Then the reaction mixture was refluxed under N<sub>2</sub> atmosphere for 12 h at 110 °C. The solvent was evaporated under reduced pressure and the residue was purified by flash chromatography on silica gel (dichloromethane/acetone: 3/1) to afford pure product **9**.

**20-R-10,11-Methylenedioxcamptothecin 9.** Yield: 80%; mp > 250 °C; <sup>1</sup>H NMR (DMSO-*d*<sub>6</sub>, 500 MHz) δ 8.45 (s, 1H), 7.49 (s, 2H), 7.24 (s, 1H), 6.47 (s, 1H), 6.27 (s, 2H), 5.39 (d, J = 16.5 Hz, 2H), 5.20 (s, 2H), 1.90–1.79 (m, 2H), 0.91–0.81 (m, 3H); <sup>13</sup>C NMR



(DMSO-*d*<sub>6</sub>, 125 MHz)  $\delta$  172.3, 157.3, 151.8, 150.5, 150.4, 149.1, 147.0, 146.4, 130.6, 128.89, 126.1, 118.5, 105.3, 103.7, 103.0, 96.3, 72.9, 65.7, 50.6, 30.7, 8.2.

**20-RS-10,11-Methylenedioxycompotothecin 10.** Yield: 85%; mp > 250 °C; <sup>1</sup>H NMR (CF<sub>3</sub>COOD, 600 MHz)  $\delta$  9.03 (s, 1H), 8.17 (s, 1H), 7.70 (s, 1H), 7.57 (s, 1H), 6.43 (s, 2H), 5.72 (s, 2H), 5.56 (s, 2H), 2.14 (q, *J* = 7.1 Hz, 2H), 1.12 (t, *J* = 7.1 Hz, 3H); <sup>13</sup>C NMR (CF<sub>3</sub>COOD, 151 MHz)  $\delta$  172.5, 159.8, 158.7, 156.9, 151.0, 149.9, 145.5, 144.9, 143.7, 130.4, 128.7, 119.9, 106.1, 104.6, 101.2, 97.5, 72.9, 65.7, 51.2, 30.3, 7.7.

**20-S-10,11-Methylenedioxycompotothecin 11.** Yield: 81%; mp > 250 °C; <sup>1</sup>H NMR (CF<sub>3</sub>COOD, 600 MHz)  $\delta$  9.05 (s, 1H), 8.16 (s, 1H), 7.69 (s, 1H), 7.55 (s, 1H), 6.44 (s, 2H), 5.93 (d, *J* = 16.6 Hz, 1H), 5.73 (s, 2H), 5.60 (d, *J* = 16.6 Hz, 1H), 2.16 (d, *J* = 6.6 Hz, 2H), 1.15 (s, 3H); <sup>13</sup>C NMR (CF<sub>3</sub>COOD, 151 MHz)  $\delta$  178.8, 161.1, 155.5, 153.9, 143.5, 142.2, 141.5, 132.1, 131.9, 124.6, 107.7, 106.5, 102.4, 99.5, 76.4, 68.9, 54.3, 33.7, 8.4.

#### General procedure for the synthesis of 9a–11a, 9b, 11b

*Taking the 9a for example.* To a stirred solution of 9 (0.25 mmol) in dry DCM (40 mL) was added EDCI (2.2 mmol), DMAP (0.51 mmol) and 4a (1.0 mmol) at room temperature. Then the reaction mixture was refluxed under N<sub>2</sub> atmosphere for 12 h at 40 °C. The mixture was cooled down to room temperature and DCM (30 mL) was poured into the mixture. The solution was washed with HCl (1 M), brine, dried over anhydrous Na<sub>2</sub>SO<sub>4</sub> and concentrated under reduced pressure to provided colorless oily and concentrated under reduced pressure. The residue was purified by flash chromatography on silica gel (dichloromethane/acetone: 5/1) to afford white powder 9a.

**(2',3',4',6'-Tetra-O-acetyl- $\beta$ -D-glucosyl)-succinic acid-10,11-methylenedioxy-compotothecin-20(R)-O-ester 9a.** Yield: 68%; mp 138–139 °C;  $[\alpha]_D^{14} = 284.25^\circ$  (*c* = 0.4 mg mL<sup>-1</sup>, CH<sub>3</sub>OH); <sup>1</sup>H NMR (DMSO-*d*<sub>6</sub>, 500 MHz)  $\delta$  8.43 (s, 1H), 7.47 (d, *J* = 9.8 Hz, 2H), 6.98 (s, 1H), 6.28 (d, *J* = 4.8 Hz, 2H), 6.24 (d, *J* = 7.8 Hz, 1H), 5.45 (d, *J* = 2.6 Hz, 2H), 5.32 (t, *J* = 9.9 Hz, 1H), 5.18 (s, 2H), 5.06–4.97 (m, 2H), 4.17–4.06 (m, 2H), 3.90 (d, *J* = 11.6 Hz, 1H), 2.92–2.69 (m, 5H), 2.14 (tt, *J* = 13.9, 7.1 Hz, 2H), 1.97–1.93 (m, 10H), 1.26–1.20 (m, 1H), 0.90 (t, *J* = 7.3 Hz, 3H); <sup>13</sup>C NMR (DMSO-*d*<sub>6</sub>, 126 MHz)  $\delta$  171.3, 170.7, 170.4, 170.1, 170.0, 169.6, 167.6, 157.0, 151.8, 150.2, 149.1, 147.0, 146.9, 145.6, 130.6, 128.8, 126.1, 118.4, 105.2, 103.6, 103.1, 94.6, 89.0, 76.6, 69.7, 69.6, 69.1, 67.7, 66.8, 61.5, 50.6, 30.9, 28.8, 20.8, 20.8, 20.7, 20.6, 8.0. HRMS (ESI-TOF, [M + H]<sup>+</sup>): *m/z* calcd for C<sub>39</sub>H<sub>38</sub>N<sub>2</sub>O<sub>18</sub>, 823.7290; found, 823.2191.

**(2',3',4',6'-Tetra-O-acetyl- $\beta$ -D-glucosyl)-succinic acid-10,11-methylenedioxy-compotothecin-20(RS)-O-ester 10a.** Yield: 63%; mp 160–163 °C;  $[\alpha]_D^{14} = 32.75^\circ$  (*c* = 0.4 mg mL<sup>-1</sup>, CH<sub>3</sub>OH); <sup>1</sup>H NMR (DMSO-*d*<sub>6</sub>, 600 MHz)  $\delta$ : 8.41 (s, 1H), 7.50–7.46 (s, 1H), 7.42 (s, 1H), 6.96 (s, 1H), 6.29–6.22 (m, 2H), 6.20 (t, *J* = 4.4 Hz, 1H), 5.41 (dd, *J* = 13.7, 6.5 Hz, 2H), 5.25 (t, *J* = 9.9 Hz, 1H), 5.16 (s, 2H), 4.99 (t, *J* = 9.7 Hz, 1H), 4.81 (dd, *J* = 10.2, 3.7 Hz, 1H), 4.14–4.08 (m, 2H), 3.98–3.92 (m, 1H), 2.90–2.85 (m, 1H), 2.80–2.73 (m, 2H), 2.72–2.67 (m, 1H), 2.14–2.05 (m, 1H), 1.97–1.86 (m, 9H), 1.72–1.66 (m, 3H), 0.90–0.84 (m, 3H); <sup>13</sup>C NMR (DMSO-*d*<sub>6</sub>, 150 MHz)  $\delta$ : 171.3, 170.9, 170.4, 170.0, 169.7, 169.5, 167.6, 157.0, 151.9, 150.2, 149.2, 147.0, 146.8, 145.7, 130.6, 128.8, 126.2, 118.3, 105.1, 103.6, 103.1, 94.6, 89.0, 76.6, 69.6, 69, 68.9, 67.7, 66.7, 61.5, 50.5, 30.8, 28.9, 28.8, 20.9, 20.7, 20.2, 8.0, 0.6. HRMS

(ESI-TOF, [M + H]<sup>+</sup>): *m/z* C<sub>39</sub>H<sub>38</sub>N<sub>2</sub>O<sub>18</sub>, 823.2192; found, 823.2193.

**(2',3',4',6'-Tetra-O-acetyl- $\beta$ -D-glucosyl)-succinic acid-10,11-methylenedioxy-compotothecin-20(S)-O-ester 11a.** Yield: 61%; mp 196 °C;  $[\alpha]_D^{14} = -27.75^\circ$  (*c* = 0.4 mg mL<sup>-1</sup>, CH<sub>3</sub>OH); <sup>1</sup>H NMR (DMSO-*d*<sub>6</sub>, 500 MHz)  $\delta$  8.45 (s, 1H), 7.51 (s, 1H), 7.45 (s, 1H), 6.99 (s, 1H), 6.28 (d, *J* = 7.6 Hz, 2H), 6.22 (d, *J* = 7.8 Hz, 1H), 5.43 (dd, *J* = 17.8, 6.3 Hz, 2H), 5.28 (t, *J* = 9.9 Hz, 1H), 5.21 (s, 2H), 5.02 (t, *J* = 9.6 Hz, 1H), 4.84 (dd, *J* = 10.3, 3.6 Hz, 1H), 4.14 (d, *J* = 9.8 Hz, 2H), 3.99 (t, *J* = 10.7 Hz, 1H), 2.90 (dt, *J* = 18.8, 9.3 Hz, 1H), 2.84–2.68 (m, 3H), 2.18–2.08 (m, 2H), 2.01–1.86 (m, 9H), 1.73 (s, 3H), 0.90 (q, *J* = 7.2 Hz, 3H); <sup>13</sup>C NMR (DMSO-*d*<sub>6</sub>, 126 MHz)  $\delta$  171.3, 170.9, 170.4, 170.0, 169.7, 169.5, 167.6, 157.0, 151.9, 150.2, 149.2, 147.0, 146.8, 145.7, 130.6, 128.8, 126.2, 118.3, 105.1, 103.6, 103.1, 94.8, 89.1, 76.6, 69.7, 69.7, 69.0, 67.7, 66.7, 61.5, 50.6, 30.8, 28.9, 28.8, 20.9, 20.7, 20.2, 8.0. HRMS (ESI-TOF, [M + H]<sup>+</sup>): *m/z* calcd for C<sub>39</sub>H<sub>38</sub>N<sub>2</sub>O<sub>18</sub>, 823.7290; found, 823.2196.

**(2',3',4'-Tri-O-acetyl- $\alpha$ -L-rhamnosyl)-succinic acid-10,11-methylenedioxy-compotothecin-20(R)-O-ester 9b.** Yield: 71%; mp 177–178 °C;  $[\alpha]_D^{14} = 98.75^\circ$  (*c* = 0.4 mg mL<sup>-1</sup>, CH<sub>3</sub>OH); <sup>1</sup>H NMR (DMSO-*d*<sub>6</sub>, 500 MHz)  $\delta$  8.42 (s, 1H), 7.49–7.42 (m, 2H), 7.02 (d, *J* = 5.4 Hz, 1H), 6.28 (d, *J* = 9.3 Hz, 2H), 5.94 (s, 1H), 5.46 (s, 2H), 5.19–5.11 (m, 3H), 5.07 (s, 1H), 4.89 (t, *J* = 10.0 Hz, 1H), 3.97 (dt, *J* = 12.4, 6.2 Hz, 1H), 2.89–2.70 (m, 4H), 2.18–2.06 (m, 3H), 2.03–1.99 (m, 3H), 1.96 (s, 2H), 1.91 (d, *J* = 6.2 Hz, 3H), 1.09 (d, *J* = 6.1 Hz, 3H), 0.91 (t, *J* = 7.4 Hz, 3H); <sup>13</sup>C NMR (DMSO-*d*<sub>6</sub>, 126 MHz)  $\delta$  171.3, 170.2, 170.1, 169.9, 169.8, 167.6, 157.0, 151.8, 150.2, 149.1, 147.0, 146.8, 145.7, 130.6, 128.8, 126.1, 118.3, 105.2, 103.5, 103.1, 94.8, 90.7, 76.6, 70.3, 70.2, 68.6, 68.5, 68.4, 66.7, 50.5, 30.8, 28.9, 20.9, 20.8, 20.8, 17.7, 8.0. HRMS (ESI-TOF, [M + H]<sup>+</sup>): *m/z* calcd for C<sub>37</sub>H<sub>36</sub>N<sub>2</sub>O<sub>16</sub>, 765.6930; found, 765.2142.

**(2',3',4'-Tri-O-acetyl- $\alpha$ -L-rhamnosyl)-succinic acid-10,11-methylenedioxy-compotothecin-20(S)-O-ester 11b.** Yield: 73%; mp 202–203 °C;  $[\alpha]_D^{14} = -166.50^\circ$  (*c* = 0.4 mg mL<sup>-1</sup>, CH<sub>3</sub>OH); <sup>1</sup>H NMR (DMSO-*d*<sub>6</sub>, 500 MHz)  $\delta$  8.42 (s, 1H), 7.46 (d, *J* = 9.1 Hz, 2H), 6.94 (s, 1H), 6.28 (d, *J* = 9.0 Hz, 2H), 6.05 (s, 1H), 5.44 (d, *J* = 16.7 Hz, 2H), 5.20–5.12 (m, 4H), 4.90 (t, *J* = 9.7 Hz, 1H), 3.96 (dq, *J* = 12.3, 6.1 Hz, 1H), 2.89 (dt, *J* = 16.6, 6.4 Hz, 1H), 2.81 (dd, *J* = 14.5, 8.4 Hz, 1H), 2.79–2.71 (m, 2H), 2.20–2.08 (m, 2H), 2.04–1.89 (m, 9H), 1.02 (t, *J* = 7.0 Hz, 3H), 0.91 (t, *J* = 7.4 Hz, 3H); <sup>13</sup>C NMR (DMSO-*d*<sub>6</sub>, 126 MHz)  $\delta$  171.2, 170.2, 170.1, 170.0, 169.9, 167.6, 157.0, 151.8, 150.2, 149.1, 147.0, 146.8, 145.6, 130.6, 128.8, 126.1, 118.3, 105.1, 103.6, 103.1, 94.7, 90.6, 76.6, 70.1, 68.6, 68.6, 68.4, 66.7, 50.5, 30.8, 29.0, 29.0, 20.9, 20.9, 20.8, 17.5, 8.0. HRMS (ESI-TOF, [M + H]<sup>+</sup>): calcd for C<sub>37</sub>H<sub>36</sub>N<sub>2</sub>O<sub>16</sub>, 765.6930; found, 765.2138.

## 4.2 Bioactivity study

**Cell culture.** The human ovarian cancer cell line 2008, the human lung cancer cell line A549, the human breast cancer cell line MDA-MB-231 and the mouse prostate cancer cell line RM-1, were obtained from the Type Culture Collection of the Chinese Academy of Sciences (Shanghai, China). A2008, A549, MDA-MB-231 and RM-1 cells were maintained in RPMI 1640 media (Gibco) supplemented with 5% heat-inactivated FBS (Gibco, Australia), 100 U mL<sup>-1</sup> of penicillin, and 0.1 mg mL<sup>-1</sup> of



streptomycin (hyclone) at 37 °C with 5% CO<sub>2</sub> and 95% air. Cells were maintained at subconfluency and culture media were changed every other day. All cells used were between passages 3 to 30. DMSO was used as the vehicle to deliver the compounds at a final concentration of 0.1% in all of the experiments.

**Cellular viability assay and IC<sub>50</sub> calculations.** For cellular viability assay, a 96-wells plate was seeded with 2000 cells per well in 100 μL of complete cell culture medium. After 24 hours, 100 μL of complete medium containing serial concentrations of each compound was added to each well. Either cancer or CHO cells were cultured for 48 hours, followed by addition of 20 μL of resazurin (2 mg mL<sup>-1</sup> dissolved in water, catalog no. R7017-5 G, Sigma) to the media for 16 hours. The fluorescent signal was monitored at an excitation wavelength of 544 nm and an emission wavelength of 595 nm using a Spectramax M5 plate reader (Molecular Devices). The relative fluorescence unit (RFU) generated from the assay was proportional to the number of living cells in each well.<sup>48</sup> The IC<sub>50</sub> values for each drug were calculated using the Logit approach.

**Immunoblot analysis.** A549 cells were seeded in 15 cm-petri dishes (8 × 10<sup>4</sup> cells per mL, 20 mL per dish). After 24 h, cells were treated with serial concentrations of compound FL118, irinotecan and **11b**. After another 48 h of incubation at 37 °C, all the cells were collected with cell-scrappers, and incubated on ice for 30 min in cell lysis buffer (Cell Signaling Technology, USA). Cell suspensions were centrifugated at 10 000 × *g* for 10 min at 4 °C, then supernatants were collected. Proteins were quantified by BCA Protein Assay Kit (Beyotime, Beijing, China). For immunoblot analysis, equal amount of proteins (25–100 μg, depending on the proteins of interest) were resolved over 10% or 12% SDS-polyacrylamide gel electrophoresis and transferred to nitrocellulose membrane. The membranes containing the transferred protein were blocked in blocking buffer (5% nonfat dry milk, 1% Tween-20 in 20 mM Tris-buffered saline, pH 7.6) for 2 h at room temperature, and then incubated with appropriate monoclonal primary antibody in blocking buffer overnight at 4 °C. After incubation with appropriate secondary antibodies, the membranes were washed three times with Tris buffer with Tween-20, and then visualized using Western Lightning (PerkinElmer, Waltham, MA, USA). Antibody for survivin was purchased from Santa Cruz. Antibody for PARP was purchased from Cell Signaling.

**Cell cycle analyses.** A549 (6 × 10<sup>4</sup> cell per mL) cells were seeded in 6-well plates. Gentle mixing was required to avoid accumulation of cells in the center of each well. After 24 h of incubation for attachment, cells were treated with serial concentrations of compound **11b** or FL118 in 2 mL of serum complete media. After 48 h, media containing any floating cells were collected and combined with adherent cells that were detached by brief trypsinization (0.25% trypsin–EDTA; hyclone). Cell pellets were washed with 1 mL of ice-cold PBS and then resuspended in 1 mL of 70% ethanol and then kept at 4 °C overnight. After centrifugation (1600 × *g*, 5 min), the supernatant was removed and the cells were incubated with 0.5 mL PI/RNase staining buffer (BD Biosciences) for 15 min at room temperature. Single-cell suspension was generated by gentle pipetting. Cell cycle was analyzed using the Beckman cell

analyzer FC500-mpl at Marine Biodiversity and Evolutionary Institute (Ocean University of China), and data were processed using Beckman CXP software and Multicycle software.

**In Vivo antitumor activity.** The *in vivo* antitumor activity of compound **11b** was determined in the RM-1 mouse prostate cancer model. All animal procedures were performed in accordance with the Guidelines for Care and Use of Laboratory Animals of School of Medicine and Pharmacy, Ocean University of China and approved by the Animal Ethics Committee of School of Medicine and Pharmacy, Ocean University of China. Five-week-old male C57BL6 mice were subcutaneously inoculated with a small RM-1 tumor block on the back of the mouse. The animals were randomized into the appropriate experimental groups (five animals for the control and for each treatment group). When the tumor volume reached 200–300 mm<sup>3</sup>, drugs were administered by intratumoral injection (three doses, every other day on three occasions) for 7 days. The drug was formulated in a mixture of 5% DMSO, 4% Tween 80, and 81% normal saline. Body weights were recorded every day after drug treatment. At the end of the observation period, animals were euthanized by cervical dislocation and the tumor bulks were peeled off according to the Guide for the Care and Use of Laboratory Animals as published by the US National Institutes of Health (NIH Publication No. 85-23, revised 1996) and approved by the Institutional Ethics Review Board of Sun Yat-Sen University, and the case number is IACUC-DD-17-0505.

### 4.3 Molecular modeling

Molecular docking was performed using MOE using AMBER10:EHT forcefield. Compounds topotecan, FL118 and **11b** were drawn in Chem3D Pro saved the format as mol2 and minimized using 10 000 steps of steepest minimization in MOE. The X-ray crystal structures of the Top1/DNA complex (PDB code: 1K4T) was downloaded from the protein data bank (<http://www.rcsb.org>). In consideration of the flexibility of the side chains of the residues at the binding site, the induced fit docking approach was applied in the docking studies. The produced conformation of with the best score was selected for the analysis.

### 4.4 ADMET predictions

The prediction of ADMET properties were performed using the Advanced Chemistry Development (ACD) Percepta platform (<http://www.acdlabs.com>). Any ADMET descriptor was evaluated by Percepta based on training libraries implemented in the software, which include a consistent pool of molecules whose pharmacokinetic and toxicity profiles are experimentally known.

## Conflicts of interest

The authors declare no competing financial interest.

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