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Introduction

Alkynones are attractive motifs in organic chemistry involved in the synthesis of medicinally valuable heteroaromatic compounds.¹ These molecules are also important intermediates in the synthesis of natural products² and as part of biologically active molecules (Fig. 1).³

Consequently, a rich variety of methodologies targeting their synthesis has been reported, some of which involve the addition of borylated terminal alkynes to acyl chlorides,⁴ the addition of hypervalent alkynyl iodides to aldehydes *via* C–C bond cleavage, metal-catalyzed C–H bond activation of aldehydes⁵ or the oxidation of propargylic alcohols.⁶ While impressive, these methodologies present some drawbacks, such as excessive generation of chemical waste, instability of some of the substrates required and poor functional group tolerance. The



Synthesis of D-glyco-alkynone derivatives via

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A carbonylative Sonogashira coupling approach to the synthesis of glyco-alkynones is described. Eighteen

examples were obtained in moderate do nearly quantitative yields under mild conditions employing

 $MO(CO)_6$ as a safe carbon monoxide source. Functionalization of the alkynyl moiety via cycloaddition

carbonylative Sonogashira reaction[†]

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with organic azides provided six examples of glyco-triazoles.

In a previous report,^{8a} we explored the synthesis of amidoglucals and glucal esters *via* the carbonylative coupling reaction of 2-iodo-D-glucal. Herein, we describe the synthesis of glycoalkynones *via* carbonylative Sonogashira coupling reaction, expanding the spectrum of reactions involving this important substrate (Scheme 1).

Taking advantage of the alkynyl group readily installed by this reaction, we also explored the synthesis of glyco-substituted triazoles *via* click chemistry. This approach has been of pivotal importance for carbohydrate chemistry as a tool to efficiently connect a sugar moiety to a molecule of interest *via* a triazole linker, improving the hydrophilicity, bioavailability and chemical profile of these fragments.⁹ Moreover, the biological activity demonstrated by several alkynone derivatives (*e.g.* triazoles) make new routes to these structures synthetically relevant (Fig. 2).¹⁰



Fig. 1 Alkynones in biologically active compounds.

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Scheme 1 2-lodoglucal carbonylative coupling reactions.

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Scheme 2 Synthesis of 2-iodo-tri-O-acetyl-D-glucal

Screening of reaction conditions

Results and discussion

We commenced our study by synthesizing 2-iodo-tri-*O*-acetyl-D-glucal (1) from tri-*O*-acetyl-D-glucal, *N*-iodosuccinimide (NIS) and AgNO₃ (Scheme 2).¹¹

With substrate 1 in hand, we next screened the reaction conditions for the carbonylation of 2-iodo-tri-O-acetyl-D-

glucal (1) with $Mo(CO)_6$ and 4-ethynyltoluene. Reactions were followed by TLC to ensure full conversion of the starting material 1 (Table 1).

We started by screening the effect of the catalyst on the reaction outcome. $PdCl_2$, $Pd(PhCN)_2Cl_2$, and $Pd(Prol)_2$ (Table 1, entries 1, 2 and 4, respectively) led to the formation of alkynone **3a** in moderate yields. Catalysts containing ligands that are at the same time electron-rich and sterically demanding, such as xantphos and PEPPSI,¹² delivered alkynone **3a** in good to nearly quantitative yield (Table 1, entries 3, 5 and 6), with the combination $PdCl_2$ /xantphos being the best. In order to seek other high-yielding set of conditions, the effect of the base was next examined. Organic and inorganic bases such as DIPEA, DBU, NaOAc and K₂CO₃ gave **3a** in lower yields, with inorganic K₂CO₃ delivering the desired product in only 25%. Different solvents were also screened, however, only poor to modest yields of **3a** were obtained.

With the optimized reaction conditions in hand, we set out to investigate the generality of this reaction (Scheme 3).

Terminal alkynes bearing electron-neutral and electrondonating groups delivered the desired alkynones in good to excellent yields (**3a–g**). Electron-withdrawing groups such as the difluorinated moiety present in **2h** and *meta*-chloro substituted **2i** gave **3h** and **3i** in good yields, while *meta*fluorinated **2j** gave **3j** in 67%. Incorporation of an heteroaromatic substituent was also tolerated, and alkynone **3k** was obtained in 65% yield. Pleasingly, both cyclopropyl and TMS groups proved to be stable under the reaction

| $\begin{array}{c} A_{CO} & & \\ A_{CO} & & \\ A_{CO} & & \\ A_{CO} & & \\ 0 A_{C} & & \\ 1 & & 2a \end{array} \xrightarrow{(catalyst (5.0 mol\%) \\ base (3.0 equiv.) \\ Mo(CO)_e (1.0 equiv.) \\ solvent, 70 °C \end{array} \xrightarrow{A_{CO} & 0 \\ A_{CO} & & \\ OA_{C} & \\ $ | | | | | |
|---|---|----------------------|-------------|-------------------|-----------|
| Entry ^a | Catalyst/ligand | Base (3.0 equvi.) | Solvent | Reaction time (h) | Yield (%) |
| Effect of cat | alyst | | | | |
| 1 | PdCl ₂ | Et ₃ N | 1,4-Dioxane | 12 | 66 |
| 2 | $Pd(PhCN)_2Cl_2$ | Et_3N | 1,4-Dioxane | 12 | 58 |
| 3 | Pd(PhCN) ₂ Cl ₂ /xantphos | Et_3N | 1,4-Dioxane | 12 | 73 |
| 4 | $Pd(Prol)_2$ | Et_3N | 1,4-Dioxane | 12 | 63 |
| 5 | PEPPSI-IPr | Et_3N | 1,4-Dioxane | 12 | 75 |
| 6 | PdCl ₂ /xantphos | Et_3N | 1,4-Dioxane | 2 | 99 |
| Effects of ba | se | | | | |
| 7 | PdCl ₂ /xantphos | DIPEA | 1,4-Dioxane | 16 | 55 |
| 8 | PdCl ₂ /xantphos | DBU | 1,4-Dioxane | 16 | 43 |
| 9 | PdCl ₂ /xantphos | NaOAc | 1,4-Dioxane | 16 | 32 |
| 10 | PdCl ₂ /xantphos | K_2CO_3 | 1,4-Dioxane | 16 | 25 |

Effect of solvent 11 PdCl₂/xantphos Et_3N Toluene 16 55 12 PdCl₂/xantphos THF 43 Et₃N 16 13 PdCl₂/xantphos DMF 16 32 Et₃N 14 PdCl₂/xantphos Et₃N MeCN 16 25

^a Reaction condition: 1 (0.2 mmol), catalyst (5 mol%), ligand (5 mol%), 4-ethynyltoluene (1.5 equvi.), base (3.0 equvi.), solvent (3 mL).





Reaction conditions: 1 (0.2 mmol), terminal alkyne (1.5 equiv.), PdCl₂ (5.0 mol%), Xantphos (5.0 mol%), El₃N (3.0 equiv.), Mo(CO)₆ (1.0 equiv.), 1,4-dioxane, 2 h, 70 °C. ^a Reaction time: 4 h. ^b gram-scale reaction.

Scheme 3 Sonogashira carbonylative coupling reaction of 2-iodo-p-glucal and terminal alkynes.

conditions, with products **31** and **3m** being isolated in 72% and 80%, respectively, both leaving useful handles for further functionalization (see Scheme 4).¹³ Incorporation of terminal alkynes bearing alkyl moieties provided mixed results, with **2n** and **2o** delivering alkynones in moderate yields, while **2p** and **2q**, bearing a tertiary alcohol, provided **3p** and **3q** in good yields. 1,4-Diethynylbenzene **2r** was subjected to the reaction conditions, giving the symmetrical alkynone **3r** in 70%. Finally, the reaction with **2a** was repeated on a gram scale, providing **3a** in 80% isolated yield (Scheme 4).

In order to demonstrate the usefulness of this methodology, we decided to explore the formation of 1,2,3-triazoles *via* click chemistry. An *in situ*-generated terminal alkyne provided the desired triazoles **5a–f** in the presence of organic azides, PMDTA and copper iodide (conditions found after a quick



Scheme 4 Gram-scale reaction.

screening).¹⁴ A variety of moieties were tolerated at the position 1 of the newly formed ring: a benzylic substituent (5a, 67%), heteroaromatic substituents (5b, 72% and 5c, 70%) and



Reaction conditions: **3m** (0.25 mmol), Cul (1.0 equiv.), RN₃ (1.2 equiv.), PMDTA (1.2 equiv.), TBAF (1.2 equiv.), THF (3 mL), 0 °C for 2 h. PMDTA = *N*,*N*,*N*,*N*'',*N*''.pentamethyldiethylenetriamine.

Scheme 5 Synthesis of D-glyco-1,2,3-triazoles.

unactivated (5d, 65% and 5e, 58%) and activated aromatic rings (5f, 86%) (Scheme 5).

Conclusions

In conclusion, we have described a convenient palladiumcatalyzed Sonogashira carbonylative coupling reaction for the synthesis of D-glyco-alkynones. This approach permitted the synthesis of 18 examples in moderate to nearly quantitative yields under mild conditions, employing $Mo(CO)_6$ as a safe carbon monoxide source. Further functionalization of a masked terminal alkynone allowed the synthesis of D-glyco-1,2,3triazoles in moderate yields, demonstrating one of the potential applications of the alkynones described herein.

Experimental section

General considerations

The compounds were all identified by usual analytical methods: ¹H NMR, ¹³C NMR, IR, and HR-MS (ESI). ¹H and ¹³C NMR spectra were measured in CDCl₃, in a Bruker DPX-300 instrument. ¹H chemical shifts were reported in ppm referenced relative to TMS internal standard (0.00 ppm) or the residual chloroform peak (7.26 ppm). Abbreviations to denote the multiplicity of a particular signal are: m (multiplet), s (singlet), d (doublet), t (triplet) and dd (doublet of doublets). ¹³C chemical shift were reported in ppm relative to the CDCl₃ triplet (77.16 ppm). IR spectra were measured on an Agilent Technologies Cary 630 and were reported in wavenumbers (cm⁻¹). High-resolution mass spectra (HRMS) were recorded on a Shimadzu LCMS-TOF, using ESI with 50% solution of acetonitrile/H2O and 0.1% formic acid as ionization method. Thin layer chromatography (TLC) was performed using silica gel UV254 0.20 mm thickness. For visualization, TLC plates were either placed under ultraviolet light, or stained with iodine or acidic vanillin solution. The solvents were purified by distillation or used without any purification in the case of HPLC-grade material. All other compounds were used as received.

General procedure for the synthesis of 3a-r

To a vial equipped with a magnetic stirrer bar and sealed with a rubber septum connected to a deflated balloon with a needle were added the tri-*O*-acetylated iodoglucal (0.2 mmol), 1,4dioxane (3.0 mL), PdCl₂ (5 mol%), xantphos (5 mol%), Mo(CO)₆ (0.2 mmol, 1 equiv.), the alkyne (0.3 mmol, 1.5 equiv.) and Et₃N (0.6 mmol, 3 equiv.). The reaction mixture was vigorously stirred at 70 °C for 2 to 4 h. The resulting mixture was washed with water and extracted with ethyl acetate. The organic layers were then combined and evaporated. The crude products were purified by flash chromatography using hexane and ethyl acetate as eluent (7 : 3).

General procedure for the synthesis of 5a-f

To a vial (20 mL) equipped with a magnetic stirrer bar under a nitrogen atmosphere containing CuI (0.25 mmol, 1 equiv.), THF (4 mL), an organic azide (0.3 mmol, 1.2 equiv.) and **3m** (0.25 mmol, 1 equiv.) was added PMDETA (0.3 mmol, 1.2 equiv.) and the reaction mixture was stirred at 0 $^{\circ}$ C for 2 h. After this period, the reaction mixture was diluted with ethyl acetate and washed with aqueous NaCl. The organic phase was collected, dried over MgSO₄, filtered and the solvent was evaporated under reduced pressure. Purification was performed using flash chromatography (ethyl acetate/hexane, 4 : 6).

Analytical data of compounds 3a-r/5a-f

Product **3a** was obtained as a yellow oil (83 mg, 0.20 mmol, 99%). ¹H NMR (300 MHz, CDCl₃): δ = 8.00 (s, 1H), 7.40 (d, *J* = 7.9 Hz, 2H), 7.12 (d, *J* = 7.8 Hz, 2H), 5.73 (dd, *J* = 3.1, 1.6 Hz, 1H), 5.15 (t, *J* = 3.0 Hz, 1H), 4.70–4.51 (m, 1H), 4.40 (dd, *J* = 12.1, 7.8 Hz, 1H), 4.14 (dd, *J* = 12.1, 4.4 Hz, 1H), 2.31 (s, 3H), 2.15–1.89 (m, 9H). ¹³C NMR (75 MHz, CDCl₃): δ = 174.5, 170.2, 169.4, 169.1, 160.7, 141.3, 132.8, 129.4, 116.7, 114.9, 91.4, 84.8, 75.6, 65.6, 61.2, 60.9, 21.6, 20.7, 20.6, 20.6. IR (ν , cm⁻¹) = 2877, 2112, 1685, 1564, 1177, 1328, 1197, 1154, 1143, 991. HRMS (ESI-TOF) calc. [C₂₂H₂₂O₈Na⁺] 437.1212, found 437.1212.

Product **3b** was obtained as a yellow oil (72 mg, 0.18 mmol, 92%). ¹H NMR (300 MHz, CDCl₃): δ = 8.08 (s, 1H), 7.65–7.55 (m, 2H), 7.49–7.34 (m, 3H), 5.82 (d, *J* = 1.8 Hz, 1H), 5.23 (t, *J* = 3.0 Hz, 1H), 4.74–4.60 (m, 1H), 4.48 (dd, *J* = 12.1, 7.8 Hz, 1H), 4.21 (dd, *J* = 12.1, 4.5 Hz, 1H), 2.18–2.01 (m, 9H). ¹³C NMR (75 MHz, CDCl₃): δ = 174.4, 170.2, 169.4, 169.1, 160.9, 132.8, 130.6, 128.6, 119.8, 114.9, 90.8, 84.9, 75.6, 65.6, 61.2, 60.9, 20.7, 20.6, 20.6. IR (ν , cm⁻¹) = 2959, 2864, 2127, 1682, 1566, 1324, 1266, 1175, 1151, 992. HRMS (ESI-TOF) calc. [C₂₁H₂₀O₈Na⁺] 423.1056, found 423.1051.

Product **3c** was obtained as a yellow oil (84 mg, 0.18 mmol, 88%). ¹H NMR (300 MHz, CDCl₃): δ = 8.10 (s, 1H), 7.74–7.53 (m, 6H), 7.49–7.36 (m, 3H), 5.93–5.79 (m, 1H), 5.24 (t, *J* = 3.1 Hz, 1H), 4.75–4.64 (m, 1H), 4.49 (dd, *J* = 12.0, 7.8 Hz, 1H), 4.22 (dd, *J* = 12.1, 4.5 Hz, 1H), 2.19–1.98 (m, 9H). ¹³C NMR (75 MHz, CDCl₃): δ = 174.4, 170.2, 169.4, 169.1, 160.8, 143.5, 139.7, 133.3, 128.9, 128.1, 127.3, 127.1, 118.5, 114.9, 90.9, 85.7, 75.7, 65.6, 61.2, 60.9, 20.7, 20.6, 20.6. IR (ν , cm⁻¹) = 2959, 2931, 2123, 1685, 1566, 1438, 1324, 1264, 1175, 1151, 991. HRMS (ESI-TOF) calc. [C₂₇H₂₄O₈Na⁺] 499.1363, found 499.1361.

Product **3d** was obtained as a yellow oil (78 mg, 0.18 mmol, 90%). ¹H NMR (300 MHz, CDCl₃): δ = 8.06 (s, 1H), 7.54 (d, *J* = 8.8 Hz, 2H), 6.90 (d, *J* = 8.8 Hz, 2H), 5.87–5.78 (m, 1H), 5.23 (t, *J* = 3.0 Hz, 1H), 4.72–4.62 (m, 1H), 4.47 (dd, *J* = 12.1, 7.9 Hz, 1H), 4.21 (dd, *J* = 12.2, 4.5 Hz, 1H), 3.84 (s, 3H), 2.19–1.99 (m, 9H). ¹³C NMR (75 MHz, CDCl₃): δ = 174.5, 170.2, 169.4, 169.1, 161.6, 160.3, 134.8, 114.7, 114.4, 111.6, 91.9, 84.8, 75.5, 65.7, 61.3, 60.9, 55.4, 20.7, 20.6, 20.6. IR (ν , cm⁻¹) = 2866, 2747, 2119, 1685, 1549, 1460, 1175, 1151, 1134, 991. HRMS (ESI-TOF) calc. [C₂₂H₂₂O₉Na^{+]} 453.1156, found 453.1159.

Product **3e** was obtained as a yellow oil (88 mg, 0.19 mmol, 99%). ¹H NMR (300 MHz, CDCl₃): δ = 7.88 (s, 1H), 7.29 (d, *J* = 8.5 Hz, 1H), 6.61–6.47 (m, 2H), 5.67–5.59 (m, 1H), 5.05 (t, *J* = 3.4 Hz, 1H), 4.54–4.41 (m, 1H), 4.33–4.23 (m, 1H), 4.12–3.95 (m, 1H), 3.62 (s, 3H), 2.29 (s, 3H), 1.96–1.80 (m, 9H). ¹³C NMR (75 MHz, CDCl₃): δ = 174.5, 170.2, 169.4, 169.1, 161.5, 160.4, 144.1, 135.2, 115.4, 115.0, 111.8, 111.6, 90.8, 88.4, 75.6, 65.6, 61.2, 60.9, 55.3, 21.0, 20.7, 20.6, 20.5. IR (ν , cm⁻¹) = 2821, 2756, 2112, 1685,

1566, 1549, 1324, 1259, 1179, 1151, 992. HRMS (ESI-TOF) calc. $[\rm C_{23}H_{24}O_9Na^+]$ 467.1313, found 453.1311.

Product **3f** was obtained as a yellow oil (91 mg, 0.19 mmol, 95%). ¹H NMR (300 MHz, CDCl₃): $\delta = 8.05$ (s, 1H), 7.98 (s, 1H), 7.64 (m, 2H), 7.45 (d, J = 8.4 Hz, 1H), 7.11 (dd, J = 9.0, 2.4 Hz, 1H), 7.04 (d, J = 2.6 Hz, 1H), 5.85–5.70 (m, 1H), 5.28–5.06 (m, 1H), 4.64–4.56 (m, 1H), 4.52–4.35 (m, 1H), 4.24–4.11 (m, 1H), 3.85 (s, 3H), 2.19–1.96 (m, 9H). ¹³C NMR (75 MHz, CDCl₃): $\delta = 174.5$, 170.2, 169.4, 169.2, 160.7, 159.3, 135.4, 133.9, 129.7, 128.9, 128.1, 127.2, 119.9, 114.9, 114.4, 105.9, 92.1, 85.0, 75.6, 65.7, 61.3, 61.0, 55.4, 20.7, 20.6, 20.6. IR (ν , cm⁻¹) = 2913, 2866, 2117, 1680, 1560, 1436, 1324, 1177, 1151, 991. HRMS (ESI-TOF) calc. [C₂₆H₂₄O₉Na⁺] 503.1313, found 503.1312.

Product **3g** was obtained as a yellow oil (73 mg, 0.17 mmol, 85%). ¹H NMR (300 MHz, CDCl₃): δ = 9.21 (s, 1H), 7.70 (s, 1H), 7.55–7.33 (m, 4H), 5.93 (s, 1H), 5.37 (s, 2H), 5.23 (t, *J* = 3.4 Hz, 1H), 4.60–4.52 (m, 1H), 4.47 (dd, *J* = 11.6, 7.7 Hz, 1H), 4.20 (dd, *J* = 11.8, 4.1 Hz, 1H), 2.12–2.03 (m, 9H). ¹³C NMR (75 MHz, CDCl₃): δ = 171.5, 170.3, 169.6, 169.3, 154.2, 144.0, 131.9, 130.9, 128.3, 128.0, 120.4, 114.5, 96.0, 90.3, 74.3, 73.9, 66.2, 62.5, 61.0, 20.7, 20.7, 20.6. IR (ν , cm⁻¹) = 2861, 2080, 1680, 1574, 1527, 1324, 1177, 1145, 981, 732. HRMS (ESI-TOF) calc. [C₂₂H₂₂O₉Na⁺] 453.1156, found 453.1156.

Product **3h** was obtained as a yellow oil (67 mg, 0.15 mmol, 77%). ¹H NMR (300 MHz, CDCl₃): δ = 7.93 (s, 1H), 7.45–7.30 (m, 1H), 6.80–6.68 (m, 2H), 5.63–5.56 (m, 1H), 5.05 (t, *J* = 2.8 Hz, 1H), 4.54–4.45 (m, 1H), 4.34–4.21 (m, 1H), 4.13–3.94 (m, 1H), 2.01–1.79 (m, 9H). ¹³C NMR (75 MHz, CDCl₃): δ = 173.9, 170.2, 169.3, 169.1, 164.4, 164.3 (dd, *J* = 253.5 Hz, *J* = 7.5 Hz), 161.6, 135.7, 135.6, 115.0, 112.3, 112.2 (dd J = 22.2 Hz, 3.3 Hz), 105.2 (dd, *J* = 3.7 Hz), 104.7 (t, *J* = 24.7 Hz) 89.4, 75.7, 65.5, 61.0, 60.9, 20.7, 20.6. IR (ν , cm⁻¹) = 2976, 2136, 1685, 1560, 1456, 1326, 1175, 1151, 992, 937, 711. HRMS (ESI-TOF) calc. [C₂₁H₁₈F₂O₈Na⁺] 436.0862, found 436.0869.

Product **3i** was obtained as a yellow oil (74 mg, 0.17 mmol, 85%). ¹H NMR (300 MHz, CDCl₃): δ = 7.57 (s, 1H), 7.51–7.39 (m, 1H), 7.35 (d, J = 7.8 Hz, 2H), 5.85–5.75 (m, 1H), 5.23 (t, J = 3.0 Hz, 1H), 4.70–4.68 (m, 1H), 4.56–4.42 (m, 1H), 4.23 (d, J = 4.5 Hz, 1H), 2.22–1.92 (m, 9H). ¹³C NMR (75 MHz, CDCl₃): δ = 174.1, 170.2, 169.3, 169.1, 161.2, 134.5, 132.4, 130.9, 130.8, 129.9, 121.5, 114.9, 88.7, 85.5, 75.8, 65.5, 61.0, 60.9, 20.7, 20.6, 20.6. IR (ν , cm⁻¹) = 2975, 2130, 1682, 1562, 1426, 1365, 1266, 1173, 1151, 991. HRMS (ESI-TOF) calc. [C₂₁H₁₉ClO₈Na⁺] 457.0661, found 457.0660.

Product **3j** was obtained as a yellow oil (56 mg, 0.13 mmol, 67%). ¹H NMR (300 MHz, CDCl₃): $\delta = 8.07$ (s, 1H), 7.43–7.37 (m, 2H), 7.32–7.23 (m, 1H), 7.24–7.07 (m, 1H), 5.88–5.78 (m, 1H), 5.23 (t, J = 3.0 Hz, 1H), 4.82–4.64 (m, 1H), 4.49 (dd, J = 12.1, 7.9 Hz, 1H), 4.21 (dd, J = 12.2, 4.5 Hz, 1H), 2.18–2.01 (m, 9H). ¹³C NMR (75 MHz, CDCl₃): $\delta = 174.1$, 170.2, 169.3, 169.12, 162.53 (d, J = 246.7 Hz), 161.1, 130.4 (d, J = 8.5 Hz), 128.7 (d, J = 3.2 Hz), 121.6 (d, J = 3.4 Hz), 85.2, 75.8, 65.5, 61.0, 60.9, 20.7, 20.6, 20.5. IR (ν , cm⁻¹) = 2970, 2132, 1685, 1564, 1326, 1268, 1177, 1151, 1113, 985, 849. HRMS (ESI-TOF) calc. [C₂₁H₁₉FO₈Na⁺] 441.0956, found 441.0956.

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Product **3k** was obtained as a yellow oil (52 mg, 0.13 mmol, 65%). ¹H NMR (300 MHz, CDCl₃): δ = 8.74 (s, 1H), 8.60 (d, *J* = 4.0 Hz, 1H), 8.01 (s, 1H), 7.87–7.73 (m, 1H), 7.28 (dd, *J* = 7.9, 5.0 Hz, 1H), 5.74 (s, 1H), 5.16 (s, 1H), 4.68–4.58 (m, 1H), 4.42 (dd, *J* = 12.2, 7.9 Hz, 1H), 4.13 (dd, *J* = 12.1, 4.5 Hz, 1H), 2.09–1.90 (m, 9H). ¹³C NMR (75 MHz, CDCl₃): δ = 173.9, 170.2, 169.3, 169.1, 161.3, 153.0, 150.6, 139.6, 123.2, 117.2, 114.9, 87.5, 86.8, 75.8, 65.5, 61.0, 60.8, 20.7, 20.6, 20.6. IR (ν , cm⁻¹) = 2859, 2119, 1680, 1566, 1326, 1181, 992. HRMS (ESI-TOF) calc. [C₂₀H₁₉NO₈Na⁺] 424.1003, found 444.1002.

Product **3l** was obtained as a yellow oil (53 mg, 0.14 mmol, 72%). ¹H NMR (300 MHz, CDCl₃): δ = 7.90 (s, 1H), 5.72 (s, 1H), 5.18 (s, 1H), 4.64–4.54 (m, 1H), 4.44 (dd, *J* = 12.1, 7.8 Hz, 1H), 4.19 (d, *J* = 4.5 Hz, 1H), 2.19–1.91 (m, 9H), 1.45–1.42 (m, 1H), 1.07–0.86 (m, 4H). ¹³C NMR (75 MHz, CDCl₃): δ = 174.8, 174.8, 170.5, 169.7, 169.5, 160.8, 115.1, 98.5, 75.8, 66.0, 61.5, 61.3, 21.0, 21.0, 20.9, 9.8, 9.8. IR (ν , cm⁻¹) = 2915, 2138, 1682, 1566, 1365, 1175, 1149, 991, 864. HRMS (ESI-TOF) calc. [C₁₈H₂₀O₈Na⁺] 387.1050, found 387.1051.

Product **3m** was obtained as a yellow oil (64 mg, 0.16 mmol, 80%). ¹H NMR (300 MHz, CDCl₃): δ = 7.82 (s, 1H), 5.60–5.48 (m, 1H), 5.02 (t, *J* = 3.1 Hz, 1H), 4.51–4.39 (m, 1H), 4.26 (dd, *J* = 12.2, 7.9 Hz, 1H), 4.01 (dd, *J* = 12.2, 4.5 Hz, 1H), 1.99–1.81 (m, 9H), 0.08 (s, 9H). ¹³C NMR (75 MHz, CDCl₃): δ = 174.8, 170.9, 170.03, 169.8, 162.1, 115.6, 99.7, 98.6, 76.4, 66.3, 61.7, 61.7, 21.4, 21.3, 21.3, 0.0. IR (ν , cm⁻¹) = 2864, 2028, 1914, 1685, 1566, 1324, 1261, 1175, 1151, 987, 817. HRMS (ESI-TOF) calc. [C₁₈H₂₄O₈-SiNa⁺] 419.1133, found 419.1135.

Product **3n** was obtained as a yellow oil (53 mg, 0.14 mmol, 70%). ¹H NMR (300 MHz, CDCl₃): δ = 7.95 (s, 1H), 5.73 (dd, *J* = 3.1, 1.7 Hz, 1H), 5.19 (t, *J* = 3.1 Hz, 1H), 4.69–4.58 (m, 1H), 4.45 (dd, *J* = 12.1, 7.8 Hz, 1H), 4.17 (dd, *J* = 12.1, 4.5 Hz, 1H), 2.39 (t, *J* = 7.0 Hz, 2H), 2.18–1.96 (m, 9H), 1.69–1.36 (m, 4H), 0.94 (t, *J* = 7.3 Hz, 3H). ¹³C NMR (75 MHz, CDCl₃): δ = 174.7, 170.2, 169.3, 169.1, 160.7, 114.9, 94.1, 77.0, 75.5, 65.6, 61.1, 60.9, 29.7, 22.0, 20.7, 20.6, 20.6, 18.6, 13.4. IR (ν , cm⁻¹) = 2838, 2862, 2147, 1685, 1566, 1365, 1324, 1175, 1149, 991, 864. HRMS (ESI-TOF) calc. [C₁₉H₂₄O₈Na⁺] 403.1363, found 403.1361.

Product **30** was obtained as a yellow oil (45 mg, 0.12 mmol, 62%). ¹H NMR (300 MHz, CDCl₃): δ = 7.76 (s, 1H), 5.53 (dd, *J* = 3.1, 1.7 Hz, 1H), 4.99 (t, *J* = 3.1 Hz, 1H), 4.49–4.37 (m, 1H), 4.25 (dd, *J* = 12.1, 7.8 Hz, 1H), 3.97 (dd, *J* = 12.1, 4.5 Hz, 1H), 2.17 (t, *J* = 7.1 Hz, 2H), 1.94–1.81 (m, 9H), 1.44 (h, *J* = 7.2 Hz, 2H), 0.84 (t, *J* = 7.4 Hz, 3H). ¹³C NMR (75 MHz, CDCl₃): δ = 174.7, 170.2, 169.3, 169.1, 160.7, 114.9, 93.9, 77.8, 75.5, 65.6, 61.1, 60.9, 21.2, 20.8, 20.7, 20.6, 20.6, 13.5. IR (ν , cm⁻¹) = 2916, 2879, 1680, 1560, 1141989, 836, 724. HRMS (ESI-TOF) calc. [C₁₈H₂₂O₈Na⁺] 389.1207, found 403.1361.

Product **3p** was obtained as a yellow oil (67 mg, 0.15 mmol, 78%). ¹H NMR (300 MHz, CDCl₃): δ = 7.61 (s, 1H), 7.32–7.11 (m, 5H), 5.65–5.52 (m, 1H), 5.16–5.04 (m, 1H), 4.57–4.49 (m, 1H), 4.36 (dd, *J* = 11.8, 7.8 Hz, 1H), 4.07 (dd, *J* = 12.0, 4.5 Hz, 1H), 2.84 (t, *J* = 7.3 Hz, 2H), 2.63 (t, 2H), 2.05–1.94 (m, 9H). ¹³C NMR (75 MHz, CDCl₃): δ = 174.5, 170.2, 169.3, 169.1, 161.1, 139.5, 128.5, 128.3, 126.7, 114.9, 92.7, 78.3, 75.4, 65.6, 60.9, 60.8, 33.8, 21.0, 20.7, 20.6, 20.6. IR (ν , cm⁻¹) = 2926, 2840, 2149, 1685,

1566, 1324, 1261, 1175, 1151, 1017, 991, 678. HRMS (ESI-TOF) calc. $[C_{23}H_{24}O_8Na^+]$ 451.1363, found 451.1361.

Product **3q** was obtained as a yellow oil (73 mg, 0.17 mmol, 86%). ¹H NMR (300 MHz, CDCl₃): δ = 7.91 (s, 1H), 5.68 (d, *J* = 2.3 Hz, 1H), 5.12 (t, *J* = 3.0 Hz, 1H), 4.62–4.52 (m, 1H), 4.37 (dd, *J* = 12.2, 7.8 Hz, 1H), 4.12 (dd, *J* = 12.1, 4.4 Hz, 1H), 2.68 (s, 1H), 2.09–1.98 (m, 9H), 1.93–1.84 (m, 2H), 1.72–1.43 (m, 8H). ¹³C NMR (75 MHz, CDCl₃): δ = 174.3, 170.2, 169.5, 169.1, 161.0, 114.7, 95.5, 79.9, 75.6, 65.5, 61.1, 60.9, 39.1, 39.1, 24.9, 22.9, 20.7, 20.6, 20.6. IR (ν , cm⁻¹) = 3363, 2840, 2766, 2136, 1691, 1568, 1326, 1182, 1156, 996. HRMS (ESI-TOF) calc. [C₂₁H₂₆O₉Na⁺] 445.1469, found 445.1467.

Product **3r** was obtained as a yellow oil (101 mg, 0.14 mmol, 70%). ¹H NMR (300 MHz, CDCl₃): $\delta = 8.07$ (s, 2H), 7.61 (d, J = 2.6 Hz, 4H), 5.95–5.73 (m, 2H), 5.29–5.10 (m, 2H), 4.77–4.58 (m, 2H), 4.58–4.46 (m, 2H), 4.23 (dd, J = 9.7, 5.6 Hz, 2H), 2.25–1.97 (m, 18H). ¹³C NMR (75 MHz, CDCl₃): $\delta = 174.0$, 170.2, 169.3, 169.1, 161.2, 132.8, 122.0, 114.9, 89.0, 87.0, 75.8, 65.5, 61.0, 60.8, 20.7, 20.6, 20.6. IR (ν , cm⁻¹) = 2870, 2129, 1685, 1564, 1324, 1264, 1177, 1151, 989. HRMS (ESI-TOF) calc. [C₃₆H₃₄O₁₆Na⁺] 745.1739, found 745.1735.

Product **5a** was obtained as a yellow oil (61 mg, 0.13 mmol, 67%). ¹H NMR (300 MHz, CDCl₃): δ = 9.15 (s, 1H), 8.02 (s, 1H), 7.46–7.34 (m, 5H), 5.96–5.85 (m, 1H), 5.55 (d, *J* = 1.4 Hz, 1H), 5.30 (s, 2H), 4.64 (d, *J* = 5.3 Hz, 1H), 4.54–4.44 (m, 1H), 4.23 (dd, *J* = 12.2, 4.7 Hz, 1H), 2.19–2.05 (m, 9H). ¹³C NMR (75 MHz, CDCl₃): δ = 180.3, 169.3, 168.4, 168.2, 161.0, 132.6, 128.3, 128.1, 127.3, 126.5, 126.5, 111.5, 73.6, 64.9, 60.5, 60.1, 53.4, 19.7, 19.6, 19.6. IR (ν , cm⁻¹) = 3261, 2866, 2836, 1680, 1560, 1475, 1324, 1179, 1149, 989, 706. HRMS (ESI-TOF) calc. [C₂₂H₂₃N₃O₈Na⁺] 480.1377, found 480.1375.

Product **5b** was obtained as a yellow oil (69 mg, 0.14 mmol, 72%). ¹H NMR (300 MHz, CDCl₃): δ = 9.15 (s, 1H), 8.67 (s, 1H), 8.45 (s, 1H), 7.87 (s, 1H), 7.55–7.42 (m, 2H), 7.30 (d, *J* = 2.8 Hz, 1H), 6.68–6.52 (m, 1H), 5.97–5.84 (m, 1H), 5.23 (t, *J* = 2.9 Hz, 1H), 4.65–4.56 (m, 1H), 4.46 (m, 1H), 4.21 (m, 1H), 2.06–1.97 (m, 9H). ¹³C NMR (75 MHz, CDCl₃): δ = 181.5, 170.3, 169.5, 169.3, 162.0, 148.1, 135.90, 128.1, 126.7, 126.2, 126.2, 115.5, 113.5, 112.6, 112.1, 103.5, 74.7, 66.0, 61.6, 61.2, 20.8, 20.7, 20.6. IR (ν , cm⁻¹) = 2834, 2862, 2779, 1682, 1560, 1475, 1460, 1324, 1162, 1011, 989, 700. HRMS (ESI-TOF) calc. [C₂₃H₂₂N₄O₈Na⁺] 505.1330, found 505.1329.

Product **5c** was obtained as a yellow oil (70 mg, 0.14 mmol, 70%). ¹H NMR (300 MHz, CDCl₃): δ = 9.13 (s, 1H), 9.07 (s, 1H), 8.58 (s, 1H), 8.37 (t, 1H), 8.24 (d, *J* = 8.0 Hz, 1H), 7.82 (d, *J* = 8.9 Hz, 1H), 5.93–5.89 (m, 1H), 5.24–5.22 (m, 1H), 4.64–4.57 (m, 1H), 4.47 (dd, *J* = 12.0, 7.8 Hz, 1H), 4.20 (dd, *J* = 12.0, 4.5 Hz, 1H), 2.08–1.94 (m, 9H). ¹³C NMR (75 MHz, CDCl₃): δ = 181.1, 170.3, 169.4, 169.2, 162.2, 162.1, 156.2, 148.6, 135.2, 126.0, 124.9, 120.0, 119.3, 115.1, 112.7, 74.8, 65.9, 61.5, 61.1, 20.8, 20.7, 20.6. IR (ν , cm⁻¹) = 2985, 2864, 2037, 1687, 1562, 1186, 1154, 994, 838, 855. HRMS (ESI-TOF) calc. [C₂₂H₂₀N₄O₈SNa⁺] 523.0894, found 523.0890.

Product **5d** was obtained as a yellow oil (89 mg, 0.13 mmol, 65%). ¹H NMR (300 MHz, CDCl₃): δ = 9.09 (s, 1H), 8.48 (s, 1H), 7.95 (d, *J* = 8.4 Hz, 1H), 7.49 (s, 1H), 7.40–7.21 (m, 11H), 6.28 (d, *J* = 3.5 Hz, 1H), 5.98–5.85 (m, 1H), 5.29 (s, 2H), 5.18 (s, 2H),

4.68–4.54 (m, 1H), 4.50–4.41 (m, 1H), 4.18 (dd, J = 12.1, 4.5 Hz, 1H), 2.10–1.92 (m, 9H). ¹³C NMR (75 MHz, CDCl₃): $\delta = 201.9$, 180.2, 170.4, 169.8, 169.1, 164.9, 163.5, 159.3, 139.6, 135.6, 135.5, 133.7, 128.7, 128.5, 128.2, 128.2, 128.2, 127.2, 125.6, 121.5, 113.7, 112.7, 111.6, 105.9, 71.8, 71.1, 67.1, 65.3, 61.1, 60.5, 20.8, 20.6, 20.5. IR (ν , cm⁻¹) = 2967, 2931, 1687, 1559, 1195, 1169, 1046, 998. HRMS (ESI-TOF) calc. [C₃₆H₃₃N₃O₁₁Na⁺] 706.2007, found 706.2004.

Product **5e** was obtained as a yellow oil (57 mg, 0.12 mmol, 58%). ¹H NMR (300 MHz, CDCl₃) δ = 9.07 (s, 1H), 8.69–8.59 (m, 2H), 8.30 (d, *J* = 8.0 Hz, 1H), 8.11 (d, *J* = 8.5 Hz, 1H), 7.74 (t, *J* = 8.1 Hz, 1H), 6.28 (d, *J* = 3.5 Hz, 1H), 5.92–5.83 (m, 1H), 4.66–4.57 (m, 1H), 4.52–4.41 (m, 1H), 4.19 (dd, *J* = 12.1, 4.5 Hz, 1H), 2.09–1.98 (m, 9H). ¹³C NMR (75 MHz, CDCl₃): δ = 180.1, 170.4, 169.8, 169.1, 163.6, 137.0, 131.2, 126.0, 125.7, 123.9, 115.8, 115.8, 113.7, 112.7, 74.9, 71.8, 61.5, 61.1, 20.7, 20.7, 20.6. IR (ν , cm⁻¹) = 2868, 1687, 1562, 1486, 1309, 1188, 1171, 994, 717. HRMS (ESI-TOF) calc. [C₂₁H₂₀N₄O₁₀Na⁺] 511.1072, found 511.1071.

Product **5f** was obtained as a yellow oil (81 mg, 0.17 mmol, 86%). ¹H NMR (300 MHz, CDCl₃): δ = 9.11 (s, 1H), 8.41 (s, 1H), 7.58 (d, *J* = 9.0 Hz, 2H), 6.98 (d, *J* = 8.9 Hz, 2H), 5.89 (t, 2H), 5.21 (t, *J* = 3.1 Hz, 1H), 4.64–4.55 (m, 1H), 4.44 (dd, *J* = 12.1, 7.7 Hz, 1H), 4.19 (dd, *J* = 12.0, 4.8 Hz, 3H), 2.08–1.94 (m, 9H). ¹³C NMR (75 MHz, CDCl₃): δ = 181.3, 170.3, 169.4, 169.2, 162.0, 160.3, 129.6, 127.8, 125.7, 122.3, 115.0, 112.6, 74.7, 65.9, 61.6, 61.1, 55.6, 20.7, 20.7, 20.6. IR (ν , cm⁻¹) = 2902, 2875, 1687, 1564, 1471, 1326, 1262, 1184, 1153, 998, 838. HRMS (ESI-TOF) calc. [C₂₂H₂₃N₃O₉Na⁺] 496.1327, found 496.1329.

Conflicts of interest

There are no conflicts to declare.

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