

Cite this: *RSC Adv.*, 2019, 9, 9709

Discovery of methylpyrimidine ring-fused diterpenoid analogs as a novel testosterone synthesis promoter†

Jie Bai,^{‡a} Jia Xie,^{‡b} Li-Ting Wang,^a Yajing Xing,^b Qian-Ru Jiang,^a Fan Yang,^{id a}
Jie Tang,^a Zhengfang Yi^{*b} and Wen-Wei Qiu^{id *a}

Herein we screened our small synthetic library of diterpenoid analogs for hit compounds on promoting testosterone synthesis and the methylpyrimidine ring-fused diterpenoid analog **7** was obtained as the hit. Based on the hit, a series of derivatives were designed, synthesized and evaluated for their effects on testosterone secretion in mouse Leydig TM3 cells. Most of the derivatives showed better activity in promoting testosterone synthesis than the positive control compound icariin, among which compound **17** has optimal activity and little cytotoxicity. Preliminary mechanism studies indicated that **17** significantly promoted the expression of testosterone synthesis-related marker genes (StAR, 3 β -HSD and CYP11A1). Further studies showed that **17** provided sufficient steroid materials for testosterone synthesis by stimulating autophagy in Leydig cells. Thus compound **17** emerged as a potential lead compound for further development of therapeutics for late onset of hypogonadism (LOH).

Received 26th January 2019

Accepted 20th March 2019

DOI: 10.1039/c9ra00702d

rsc.li/rsc-advances

Introduction

Diterpenoids are a major branch of natural products which exert many different biological activities.¹ For instance, anti-proliferative,² multi-drug resistance-reversing,³ antimicrobial,⁴ vasoactive,⁵ antidiabetic,⁶ anti-osteoporosis⁷ immunomodulatory,⁸ and anti-inflammatory⁹ effects. Therefore, diterpenoids play an important role in the discovery of novel bioactive agents.

Testosterone, which is primarily produced in Leydig cells as the major circulating androgen, plays important roles in sexual differentiation, secondary sex characteristics, reproductive function and sexual function.^{10,11} Serum testosterone deficiency causes late onset of hypogonadism (LOH), which is a clinical and biochemical syndrome.¹² The incidence rates of LOH are 13%, 30% and 47% for men aged 40–49, 50–59 and over 70 respectively.¹³ Previous studies indicated that LOH-related hypogonadal symptoms included sexual desire decrease, muscle mass and strength reduction, osteoporosis, increased

central body fat, forgetfulness, loss of memory, difficulty in concentration, insomnia and erectile dysfunction (ED).^{14,15}

Testosterone replacement therapy (TRT), as the main clinical treatment method for LOH at present, provides a wide range of benefits for hypogonadism, improving libido and sexual function, fertility, bone density, muscle mass, and quality of life.^{16,17} However the clinical efficacy and long-term safety of TRT remains controversial. TRT involves the direct administration of an exogenous hormone, and with this treatment, the androgen level in the serum is superphysiological and unstable.¹³ So, it has considerable side effects, for instance, testicular atrophy, erythropoiesis, prostate cancer, cardiovascular events, intrahepatic cholestasis, sleep apnea, liver failure and worsening of lower urinary tract symptoms from an enlarged prostate.^{18,19} Therefore, the discovery of novel agents as anti-LOH agents is urgently needed.

Testosterone production is regulated by luteinizing hormone (LH) and LH induces cAMP synthesis in Leydig cells by binding to the membrane LH receptors.²⁰ cAMP catalyzes the synthesis of PKA, which transports cholesterol from the cytoplasmic pool to mitochondria and promotes steroidogenesis by steroid-generating enzymes (CYP11A1, 3 β -HSD) and steroidogenic acute regulatory (StAR) protein.²¹ Additionally, autophagy in Leydig cells promotes lipid metabolism and provides sources such as triglycerides (TGs) and cholesterol for testosterone production, which plays an important role in testosterone synthesis.^{22,23}

Herein, we report the discovery of a hit compound as testosterone synthesis promoter by screening our small synthetic library of diterpenoid analogs. The methylpyrimidine

^aShanghai Engineering Research Center of Molecular Therapeutics and New Drug Development, School of Chemistry and Molecular Engineering, East China Normal University, 3663 North Zhongshan Road, Shanghai 200062, China. E-mail: wuqiu@chem.ecnu.edu.cn; Tel: +86-21-54340103

^bShanghai Key Laboratory of Regulatory Biology, Institute of Biomedical Sciences, School of Life Sciences, East China Normal University, Shanghai 200241, China. E-mail: zfyi@bio.ecnu.edu.cn; Tel: +86-21-54345016

† Electronic supplementary information (ESI) available: Spectrum of the corresponding compounds. ¹H and ¹³C NMR spectra of all final compounds. See DOI: 10.1039/c9ra00702d

‡ These authors contributed equally.



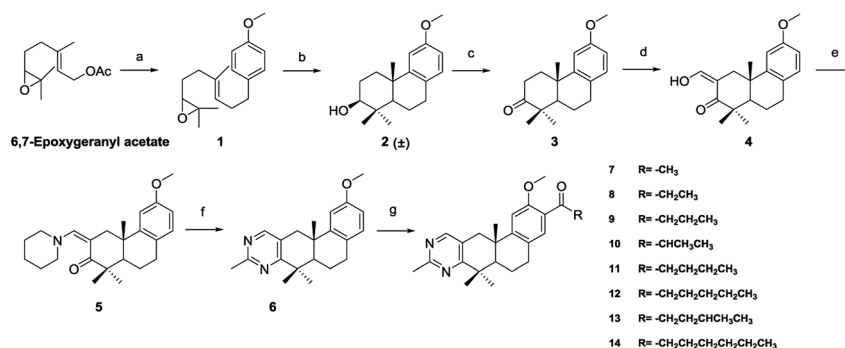
ring-fused diterpene analogue **7** was selected as the hit. Then a series of analogs were synthesized based on the hit and their activities in promoting testosterone production in mouse Leydig TM3 cells were evaluated. Results showed that compound **17** (**SH380**) was the most potent testosterone synthesis promoter. The mechanism by which the **17** increases testosterone production was preliminary explored.

Results and discussion

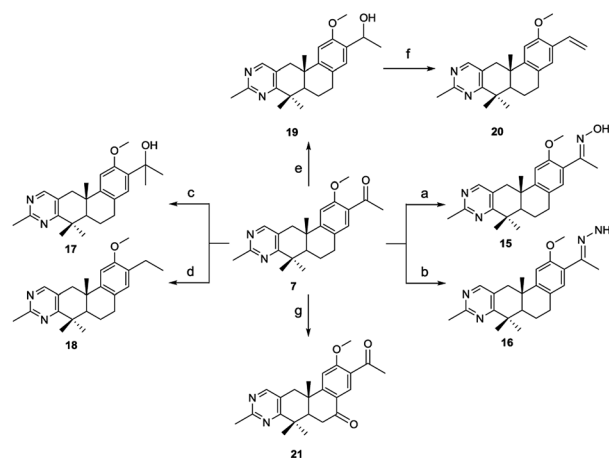
Chemistry

The methylpyrimidine acyl derivatives **7–14** were synthesized as shown in Scheme 1. Compound **6** was prepared according to our previously reported procedure^{24,25} and details were as follows. Coupling reaction of 6,7-epoxygeranyl acetate with (4-methoxybenzyl) magnesium chloride yielded compound **1**, Li₂-CuCl₄ was added as catalyst. Key intermediate **2** was obtained by the cyclization of **1** in presence of boron trifluoride etherate, and is a racemate according to the Stork–Eschenmoser hypothesis.^{26,27} Therefore, all the synthesized diterpenoids herein belong to racemates. Compound **3** was obtained by oxidation of **2** with 2-iodoxybenzoic acid (IBX). Claisen condensation of **3** with ethyl formate in the presence of NaH provided compound **4**. Compound **6** was furnished by reaction of **4** with piperidine in EtOH and then condensation with acetamide hydrochloride. Compound **7–14** were prepared by Friedel–Crafts acylation from **6** with various acyl chlorides respectively.

The synthetic route of 2-methylpyrimidine derivatives **15–22** is outlined in Scheme 2. Compound **15** was prepared from **7** with hydroxylamine hydrochloride in the presence of K₂CO₃. Condensation of **7** with hydrazine hydrate (85%) yielded **16**. Reaction of **7** with methylmagnesium chloride yielded **17**. Compound **18** was produced by Wolff–Kishner–Huang Min-lon reduction of **7** with hydrazine hydrate (85%) in the presence of NaOH. Reduction of **7** with sodium borohydride produced compound **19**, and **20** was provided by dehydration of **19** with *p*-toluenesulfonic acid. Oxidation of **7** with CrO₃ in AcOH produced **21**.



Scheme 1 Reagents and conditions: (a) (4-methoxybenzyl) magnesium chloride, LiCuCl₄, anhydrous THF, 0 °C; (b) boron trifluoride etherate, anhydrous CH₂Cl₂, -78 °C; (c) IBX, THF, DMSO, rt; (d) ethyl formate, NaH, anhydrous THF, rt; (e) piperidine, EtOH, reflux; (f) acetamide hydrochloride, sodium methoxide, EtOH, reflux; (g) acyl chloride, AlCl₃, DCM, 0 °C.



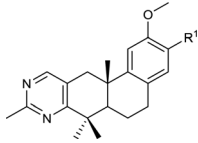
Scheme 2 Reagents and conditions: (a) hydroxylamine hydrochloride, K₂CO₃, EtOH, rt; (b) hydrazine hydrate (85%), TsOH, EtOH, reflux; (c) methylmagnesium chloride, anhydrous THF, 0 °C; (d) hydrazine hydrate (85%), NaOH, triglycol, 120 °C to 190 °C; (e) NaBH₄, DCM, MeOH, 0 °C; (f) TsOH, anhydrous THF, 60 °C; (g) CrO₃, AcOH, rt.









Activity and cytotoxicity evaluation of compound **7** and its derivatives

A hit was obtained by screening our small synthetic diterpenoids library. Mouse TM3 Leydig cells were treated with each tested compound at concentration of 20 μM for 24 h, and the testosterone levels in the culture medium were measured by ELISA (enzyme-linked immuno sorbent assay) kit. Among those analogs, compound **7** (QB364) was determined as a testosterone synthesis promoter with a testosterone concentration of 271.7 μM L⁻¹ and was more potent than the positive icariin.²⁸ Therefore, it was selected as the hit compound for further structure–activity relationship (SAR) optimization.

The first-round synthetic compounds **8–14** were obtained by Friedel–Crafts acylation with various acyl chlorides respectively and their activity in promoting testosterone production on mouse Leydig TM3 cells was determined by ELISA kit. As shown in Table 1, the activity decreased significantly when the acetyl group (**7**) was replaced by various acyl substituents. Thus,



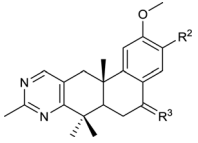
Table 1 Testosterone levels and cytotoxicity of compounds 7–14 in TM3 cells^a










Compd	R1	Testosterone Levels ^b (μM L ⁻¹)
Control	—	164.5 ± 0.2
Icariin	—	229.8 ± 4.8
7		271.7 ± 10.0
8		233.0 ± 0.5
9		252.3 ± 2.4
10		242.6 ± 6.0
11		245.4 ± 4.9
12		217.1 ± 9.9
13		174.1 ± 0.5
14		204.2 ± 6.0

^a See Experimental section. ^b Testosterone levels in the culture medium are measured by ELISA kit; data are expressed as mean ± SD of three independent assays.

compound 7 was selected for the second-round structure modification.

The second-round synthetic analogs 15–21 were obtained by modification of the acetyl group of 7 and their activity in promoting testosterone production was determined using ELISA kit. The results (Table 2) showed that the activity increased when the carbonyl group of 7 was substituted by oxime (15), while decreased as hydrazine substituent (16) was introduced into. If the carbonyl group of 7 was reduced to hydroxy (19) the activity decreased obviously. To our delight, the testosterone level in TM3 was increased significantly as the acetyl group was replaced by *tert*-butanol group (17). If the acetyl group was converted to the alkyl substituent such as ethyl (18) or vinyl (20), the activity of testosterone production was also decreased. For derivative modified at the C-6 and C-7 positions, the activity decreased as the carbonyl (21) substituent was introduced into C7 position. Further cytotoxicity evaluation in mouse TM3 cells with the SRB assay disclosed that all these analogs showed very little cytotoxicity (IC₅₀ > 100 μM). Considering the above results, the safe compound 17 exhibited most potent activity in promoting testosterone production, thus it was selected for further studies.

Table 2 Testosterone levels and cytotoxicity of compounds 15–21 in TM3 cells^a


Compd	R2	R3	Testosterone Levels ^b (μM/L)	IC ₅₀ ^c (μM)
7		—	271.7 ± 10.0	>100
15		—	282.2 ± 5.5	>100
16		—	188.5 ± 5.5	>100
17		—	303.5 ± 5.5	>100
18		—	237.0 ± 4.0	>100
19		—	222.8 ± 7.0	>100
20		—	227.1 ± 3.0	>100
21		O	259.5 ± 6.5	>100

^a See Experimental section. ^b Testosterone levels in the culture medium are measured by ELISA kit; data are expressed as mean ± SD of three independent assays. ^c Activities of compounds against the growth of TM3 cells are detected with the SRB assay and their IC₅₀ data are an average of at least three independent experiments.

Table 3 IC₅₀ of compound 24 against the growth of various normal cells^a

Cell line	Leydig cell				
	TM3	HAF	NCM460	L-02	PTN1A
IC ₅₀ (μM) ^c	112.6 ^d	111.8 ^d	>200	>200	>200

^a See Experimental section. ^b HAF, human fibroblast cell line; NCM460, human colon mucosal epithelial cell line; L-02, human fetal hepatocyte cell line; PTN1A, human prostate epithelial cell line. ^c Various cells were treated with indicated concentrations of compound 17 for 24 h; cell proliferation was determined by SRB assay; data were expressed as IC₅₀ of three independent assays. ^d Variation ± 10%.

Compound 17 has little cytotoxicity on TM3 and other normal cells within the effective concentrations

To further determination of the *in vitro* safety of 17, we detected its cytotoxicity in TM3 and several human normal cells including NCM460, HAF, L-02 and PTN1A by SRB assay. As shown in Table 3, compound 17 showed very little toxicity on cell proliferation of TM3 (IC₅₀ = 112.6 μM) and HAF (IC₅₀ = 112.6 μM) cells. In addition, 17 has almost no toxic effects on NCM460, L-02 and PTN1A cells (IC₅₀ > 200 μM). Therefore, compound 17 is a safe compound *in vitro*.



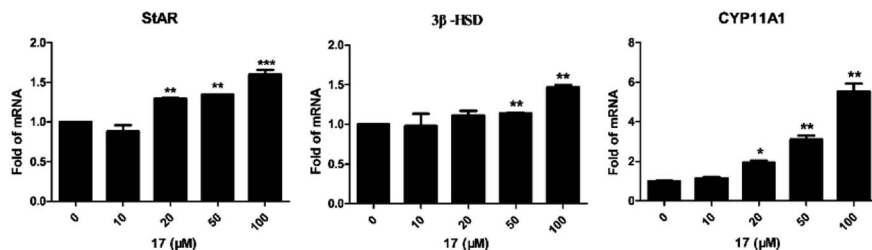


Fig. 1 Compound 17 advanced the mRNA expression levels of testosterone synthetase in Leydig cells.^a See Experimental section. Mouse Leydig TM3 cells were treated with different concentrations of compound 17 for 24 h. mRNA levels of steroidogenesis-related genes StAR, 3β-HSD and CYP11A1 were measured by quantitative-PCR. Data were expressed as mean ± SD of three independent assays; Student's *t*-tests were performed; **p* < 0.05, ***p* < 0.01, ****p* < 0.001.

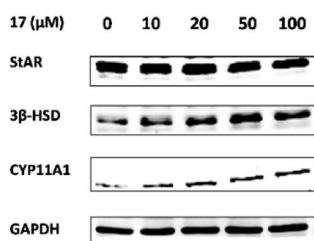


Fig. 2 Compound 17 advanced the protein expression levels of testosterone synthetase in Leydig cells.^a See Experimental section. Mouse Leydig TM3 cells were treated with different concentrations of 17 for 24 h and the protein levels of StAR, 3β-HSD and CYP11A1 were measured by Western blotting. GAPDH was used as a control.

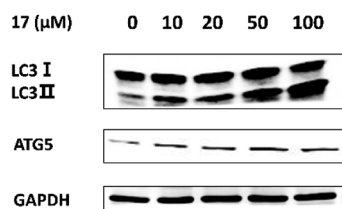


Fig. 3 Compound 17 promotes the autophagy-related protein expression levels in TM3 cells.^a See Experimental section. Mouse Leydig TM3 cells were treated with different concentrations of 17 for 24 h. The expression of LC3 I/II and ATG5 were detected by western blotting. GAPDH was used as a control.

Compound 17 enhanced the expression of testosterone synthesis-regulating genes

For determination of the primary mechanism of 17 on testosterone synthesis, the mRNA and protein expression levels of testosterone synthetase, including StAR, 3β-HSD and CYP11A1 were analyzed. The qPCR analysis showed that the mRNA expression levels of StAR, 3β-HSD and CYP11A1 were increased significantly in a concentration-dependent manner (Fig. 1). Furthermore, the protein expression levels of the three key enzymes in TM3 cells were validated by western blotting. Results indicated that 17 also increased the protein expression levels of StAR, 3β-HSD and CYP11A1 obviously as the concentration rised from 10 μM to 100 μM (Fig. 2). These primary

mechanism studies indicated that 17 increased testosterone levels in Leydig cells *via* promoting testosterone synthesis.

Compound 17 regulates the expression of autophagy-related proteins in Leydig cells

Autophagy dysfunction played a key role in the loss of testosterone production in some LOH patients.²¹ Therefore, we tried to identify if 17 promoted testosterone production *via* induction of autophagy. LC3-II serves as a reliable marker for mature autophagosomes and ATG5 is a gene product required for the formation of autophagosomes. We assessed the levels of the autophagy-related proteins by western blotting. As shown in Fig. 3, the expressions of ATG5, LC3-II and the ratio of LC3-II/LC3-I were increased significantly after treatment with 17. It suggested that the compound could promoted testosterone synthesis *via* activating autophagy in Leydig cells.

Conclusions

In this study, we designed and synthesized a number of 15 methylpyrimidine ring-fused tricyclic diterpene analogs. Most of the derivatives showed potent activity in promoting testosterone production in mouse Leydig TM3 cells, which was much better than that of the positive control icariin. Among these analogs, compound 17 exhibited the most potent activity and almost no toxic effect. Preliminary mechanism studies indicated that 17 could significantly enhance the expression of testosterone synthesis-related marker genes (StAR, 3β-HSD and CYP11A1) in a concentration-dependent manner. Further studies disclosed that 17 could also stimulate autophagy in Leydig cells and promote the expression of autophagy-related genes (LC3 and ATG 5). In conclusion, compound 17 was discovered as a promising compound, which could be further developed as a potential therapeutic agent for LOH.

Experimental section

Chemistry

General methods. Unless otherwise specified, starting materials, reagents, and solvents were purchased from commercial suppliers and used without further purification. Anhydrous THF was obtained by distillation over sodium wire.



All non-aqueous reactions were run under a nitrogen atmosphere and all reaction vessels were oven-dried. Thin-layer chromatography (TLC) was performed on silica gel plates (QingDao) with a layer thickness of 0.25 mm and spots were visualized by their quenching of the fluorescence of an indicator admixed to the SiO₂ layer. Column chromatographic purification was carried out using silica gel (200–300 mesh) and an EtOAc/hexane mixture or gradient was used unless otherwise stated. ¹H and ¹³C NMR spectra were recorded on a Bruker Advance III 400 spectrometer at 400 MHz (¹H) and 100 MHz (¹³C). Chemical shifts are reported in δ (ppm) using the δ 0 signal of tetramethylsilane (TMS) as internal standards. High resolution mass spectra were performed using a Bruker ESI-TOF high-resolution mass spectrometer.

General procedure for the Friedel–Crafts acylation (method A). To a solution of compound **6** (100 mg, 0.27 mmol) in anhydrous CH₂Cl₂ (20 mL) was added anhydrous AlCl₃ (108 mg, 0.81 mmol) and respective acyl chloride (0.54 mmol) at 0 °C under nitrogen. Then reaction mixture was stirred for 2 h at room temperature. After that the mixture was poured into water (50 mL) and extracted with CH₂Cl₂ (10 mL \times 3). The organic layer was washed with brine, dried over anhydrous Na₂SO₄, and concentrated under vacuum. The residue was purified by silica gel chromatography to afford the product.

General procedure for the reduction reaction (method B). NaBH₄ (3 mmol) was added into a solution of compound **7** or **21** (0.3 mmol) in anhydrous CH₂Cl₂ (10 mL) and MeOH (10 mL) with ice cooling under nitrogen. After stirring for 1 h at room temperature, the reaction was quenched by saturated aqueous NH₄Cl solution with ice cooling. The mixture was extracted with EtOAc (20 mL \times 3). The organic layer was washed with brine, dried over Na₂SO₄ and concentrated under vacuum. The residue was purified by silica gel chromatography to give the product.

General procedure for the oxidation reaction (method C). CrO₃ (0.6 mmol) was added into a solution of compound **6** (0.3 mmol) in AcOH (10 mL) with ice cooling. After stirring for 3 h at room temperature, the mixture was poured into water (50 mL) and extracted with EtOAc (20 mL \times 3). The organic layer was washed with sodium bicarbonate solution (5%) and brine, dried over anhydrous Na₂SO₄, and concentrated under vacuum. The residue was purified by silica gel chromatography to give the product.

General procedure for the Wolff–Kishner–Huang Min-lon reduction (method D). Hydrazine hydrate (85%) (1.2 mmol) was added into a stirred solution of compound **7** (0.3 mmol) in triglycol (10 mL) under nitrogen. The reaction mixture was stirred for 1 h at 120 °C. Then removed the condensing tube and heated the mixture to 190 °C. After the temperature is stable, the condensing tube was reinstalled and mixture was stirred for 3 h at 190 °C. After cooling, the mixture was poured into water (40 mL) and extracted with EtOAc (15 mL \times 3). The organic layer was washed with brine, dried over anhydrous Na₂SO₄, and concentrated. The residue was purified by silica gel chromatography to give the product.

General procedure for the dehydration reaction (method E). To a solution of compound **19** or **22** (0.3 mmol) in anhydrous EtOH (10 mL) was added TsOH (0.6 mmol) under nitrogen. The

reaction mixture was stirred for 6 h under reflux. Then the mixture was poured into water (30 mL) and extracted with EtOAc (10 mL \times 3). The organic layer was washed with sodium bicarbonate solution (5%) and brine, dried over anhydrous Na₂SO₄, and concentrated under vacuum. The residue was purified by silica gel chromatography to give the product.

Compound 7. This compound was obtained from compound **6** employing method A. White solid. Yield 92%. ¹H NMR (400 MHz, CDCl₃) δ 8.39 (s, 1H), 7.49 (s, 1H), 6.93 (s, 1H), 3.94 (s, 3H), 3.16 (d, J = 15.5 Hz, 1H), 2.98–2.93 (m, 1H), 2.85–2.75 (m, 2H), 2.69 (s, 3H), 2.60 (s, 3H), 2.03–1.96 (m, 1H), 1.85 (d, J = 12.3 Hz, 1H), 1.78–1.71 (m, 1H), 1.36 (s, 3H), 1.34 (s, 3H), 1.16 (s, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 199.41, 172.09, 166.09, 157.62, 157.34, 151.81, 131.11, 127.87, 126.34, 123.29, 108.57, 55.57, 49.14, 40.65, 39.69, 37.52, 31.79, 31.22, 30.12, 25.82, 23.96, 23.16, 20.44. HRMS (ESI): calcd for C₂₃H₂₉N₂O₂ [M + H]⁺, 365.2224; found, 365.2233.

Compound 8. This compound was obtained from compound **6** employing method A. White solid. Yield 90%. ¹H NMR (400 MHz, CDCl₃) δ 8.38 (s, 1H), 7.44 (s, 1H), 6.92 (s, 1H), 3.92 (s, 3H), 3.16 (d, J = 15.5 Hz, 1H), 3.00–2.93 (m, 3H), 2.85–2.75 (m, 2H), 2.69 (s, 3H), 2.00–1.96 (m, 1H), 1.85 (d, J = 12.4 Hz, 1H), 1.78–1.71 (m, 1H), 1.36 (s, 3H), 1.33 (s, 3H), 1.16–1.13 (m, 6H). ¹³C NMR (100 MHz, CDCl₃) δ 203.11, 172.21, 166.11, 157.64, 156.97, 151.25, 131.00, 127.94, 126.64, 123.42, 108.59, 55.65, 49.22, 40.72, 39.75, 37.52, 36.95, 31.27, 30.20, 25.84, 24.02, 23.21, 20.50, 8.48. HRMS (ESI): calcd for C₂₄H₃₁N₂O₂ [M + H]⁺, 379.2380; found, 379.2384.

Compound 9. This compound was obtained from compound **6** employing method A. White solid. Yield 87%. ¹H NMR (400 MHz, CDCl₃) δ 8.40 (s, 1H), 7.42 (s, 1H), 6.93 (s, 1H), 3.93 (s, 3H), 3.17 (d, J = 15.6 Hz, 1H), 2.99–2.92 (m, 3H), 2.86–2.76 (m, 2H), 2.70 (s, 3H), 2.02–1.97 (m, 1H), 1.87 (d, J = 12.5 Hz, 1H), 1.80–1.76 (m, 1H), 1.73–1.68 (m, 2H), 1.38 (s, 3H), 1.35 (s, 3H), 1.17 (s, 3H), 0.97 (t, J = 7.4 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 202.65, 172.22, 166.17, 157.66, 156.89, 151.18, 130.93, 128.00, 127.08, 123.37, 108.65, 55.68, 49.29, 45.67, 40.78, 39.76, 37.54, 31.29, 30.22, 25.85, 24.01, 23.21, 20.53, 17.86, 13.95. HRMS (ESI): calcd for C₂₅H₃₃N₂O₂ [M + H]⁺, 393.2537; found, 393.2560.

Compound 10. This compound was obtained from compound **6** employing method A. White solid. Yield 88%. ¹H NMR (400 MHz, CDCl₃) δ 8.39 (s, 1H), 7.29 (s, 1H), 6.91 (s, 1H), 3.91 (s, 3H), 3.53–3.46 (m, 1H), 3.16 (d, J = 15.5 Hz, 1H), 2.97–2.93 (m, 1H), 2.85–2.76 (m, 2H), 2.70 (s, 3H), 1.99 (dd, J = 12.2, 4.5 Hz, 1H), 1.86 (d, J = 12.3 Hz, 1H), 1.79–1.73 (m, 1H), 1.37 (s, 3H), 1.34 (s, 3H), 1.16–1.13 (m, 9H). ¹³C NMR (100 MHz, CDCl₃) δ 207.69, 172.18, 166.09, 157.63, 156.24, 150.62, 130.84, 127.96, 127.07, 123.40, 108.42, 55.68, 49.20, 40.72, 39.94, 39.71, 37.45, 31.25, 30.18, 25.85, 24.00, 23.18, 20.47, 18.68, 18.55. HRMS (ESI): calcd for C₂₅H₃₃N₂O₂ [M + H]⁺, 393.2537; found, 393.2557.

Compound 11. This compound was obtained from compound **6** employing method A. White solid. Yield 82%. ¹H NMR (400 MHz, CDCl₃) δ 8.39 (s, 1H), 7.41 (s, 1H), 6.92 (s, 1H), 3.92 (s, 3H), 3.16 (d, J = 15.5 Hz, 1H), 2.97–2.93 (m, 3H), 2.85–2.76 (m, 2H), 2.70 (s, 3H), 1.98 (dd, J = 12.4, 5.1 Hz, 1H), 1.85 (d, J = 12.4 Hz, 1H), 1.79–1.71 (m, 1H), 1.68–1.61 (m, 2H), 1.41–1.39 (m, 2H), 1.37 (s, 3H), 1.34 (s, 3H), 1.16 (s, 3H), 0.92 (t, J = 7.3 Hz,



3H). ^{13}C NMR (100 MHz, CDCl_3) δ 202.85, 172.13, 166.09, 157.63, 156.79, 151.14, 130.91, 127.92, 126.93, 123.35, 108.53, 55.61, 49.18, 43.42, 40.69, 39.70, 37.48, 31.24, 30.16, 26.53, 25.84, 23.98, 23.17, 22.51, 20.46, 13.97. HRMS (ESI): calcd for $\text{C}_{26}\text{H}_{35}\text{N}_2\text{O}_2$ $[\text{M} + \text{H}]^+$, 407.2693; found, 407.2692.

Compound 12. This compound was obtained from compound 6 employing method A. White solid. Yield 84%. ^1H NMR (400 MHz, CDCl_3) δ 8.40 (s, 1H), 7.42 (s, 1H), 6.92 (s, 1H), 3.93 (s, 3H), 3.17 (d, $J = 15.5$ Hz, 1H), 2.99–2.93 (m, 3H), 2.86–2.76 (m, 2H), 2.70 (s, 3H), 1.99 (dd, $J = 12.7, 5.8$ Hz, 1H), 1.86 (d, $J = 12.3$ Hz, 1H), 1.80–1.65 (m, 5H), 1.37 (s, 2H), 1.35 (s, 6H), 1.17 (s, 3H), 0.90 (t, $J = 6.9$ Hz, 3H). ^{13}C NMR (100 MHz, CDCl_3) δ 202.86, 172.15, 166.10, 157.64, 156.81, 151.15, 130.93, 127.93, 126.93, 123.36, 108.54, 55.62, 49.19, 43.70, 40.70, 39.71, 37.49, 31.63, 31.25, 30.17, 25.85, 24.11, 23.99, 23.18, 22.53, 20.47, 14.00. HRMS (ESI): calcd for $\text{C}_{27}\text{H}_{37}\text{N}_2\text{O}_2$ $[\text{M} + \text{H}]^+$, 421.2850; found, 421.2858.

Compound 13. This compound was obtained from compound 6 employing method A. White solid. Yield 91%. ^1H NMR (400 MHz, CDCl_3) δ 8.40 (s, 1H), 7.41 (s, 1H), 6.92 (s, 1H), 3.93 (s, 3H), 3.17 (d, $J = 15.5$ Hz, 1H), 2.97–2.94 (m, 3H), 2.86–2.76 (m, 2H), 2.70 (s, 3H), 1.99 (dd, $J = 12.8, 5.4$ Hz, 1H), 1.86 (d, $J = 12.5$ Hz, 1H), 1.80–1.72 (m, 1H), 1.63–1.61 (m, 1H), 1.59–1.53 (m, 2H), 1.37 (s, 3H), 1.35 (s, 3H), 1.17 (s, 3H), 0.92 (d, $J = 8.1$ Hz, 6H). ^{13}C NMR (100 MHz, CDCl_3) δ 203.12, 172.17, 166.11, 157.63, 156.76, 151.16, 130.97, 127.96, 126.94, 123.38, 108.53, 55.60, 49.20, 41.80, 40.71, 39.73, 37.50, 33.33, 31.26, 30.18, 27.94, 25.85, 24.00, 23.19, 22.48 (2C), 20.48. HRMS (ESI): calcd for $\text{C}_{27}\text{H}_{37}\text{N}_2\text{O}_2$ $[\text{M} + \text{H}]^+$, 421.2850; found, 421.2876.

Compound 14. This compound was obtained from compound 6 employing method A. White solid. Yield 80%. ^1H NMR (400 MHz, CDCl_3) δ 8.40 (s, 1H), 7.41 (s, 1H), 6.92 (s, 1H), 3.93 (s, 3H), 3.17 (d, $J = 15.4$ Hz, 1H), 2.99–2.93 (m, 3H), 2.86–2.76 (m, 2H), 2.70 (s, 3H), 1.99 (dd, $J = 12.5, 5.2$ Hz, 1H), 1.86 (d, $J = 12.3$ Hz, 1H), 1.79–1.73 (m, 1H), 1.68–1.64 (m, 2H), 1.37 (s, 3H), 1.34 (s, 6H), 1.31 (s, 3H), 1.17 (s, 3H), 0.88 (t, $J = 6.4$ Hz, 3H). ^{13}C NMR (100 MHz, CDCl_3) δ 202.88, 172.18, 166.10, 157.62, 156.81, 151.14, 130.93, 127.94, 126.96, 123.38, 108.55, 55.63, 49.21, 43.75, 40.71, 39.73, 37.49, 31.70, 31.26, 30.18, 29.11, 25.84, 24.40, 24.00, 23.19, 22.54, 20.48, 14.06. HRMS (ESI): calcd for $\text{C}_{28}\text{H}_{39}\text{N}_2\text{O}_2$ $[\text{M} + \text{H}]^+$, 435.3006; found, 435.3025.

Compound 15. To a solution of compound 7 (100 mg, 0.27 mmol) in anhydrous EtOH (10 mL) was added hydroxylamine hydrochloride (38 mg, 0.54 mmol) and K_2CO_3 (151 mg, 1.10 mmol). After stirring for 12 h at room temperature, the mixture was poured into water (50 mL) and extracted with DCM (20 mL \times 3). The organic layer was washed with brine, dried over anhydrous Na_2SO_4 , and concentrated under vacuum. The residue was purified by silica gel chromatography (petroleum ether/EtOAc, 2/1, v/v) to afford compound 15 (64 mg, 65%) as a white solid. ^1H NMR (400 MHz, CDCl_3) δ 8.40 (s, 1H), 7.04 (s, 1H), 6.89 (s, 1H), 3.87 (s, 3H), 3.17 (d, $J = 15.6$ Hz, 1H), 2.96–2.91 (m, 1H), 2.86–2.75 (m, 2H), 2.71 (s, 3H), 2.24 (s, 3H), 1.97 (dd, $J = 12.7, 5.8$ Hz, 1H), 1.86 (d, $J = 12.4$ Hz, 1H), 1.80–1.72 (m, 1H), 1.37 (s, 3H), 1.35 (s, 3H), 1.16 (s, 3H). ^{13}C NMR (100 MHz, CDCl_3) δ 172.39, 166.00, 157.57, 157.13, 155.91, 147.70, 129.90, 127.74, 125.16, 123.64, 108.29, 55.70, 49.39, 40.85, 39.72, 37.25, 31.27,

30.26, 25.79, 24.05, 23.17, 20.54, 15.16. HRMS (ESI): calcd for $\text{C}_{23}\text{H}_{30}\text{N}_3\text{O}_2$ $[\text{M} + \text{H}]^+$, 380.2333; found, 380.2311.

Compound 16. To a solution of compound 7 (109 mg, 0.30 mmol) in anhydrous EtOH (10 mL) was added TsOH (63 mg, 0.33 mmol) and hydrazine hydrate (85%, 0.03 mL, 0.6 mmol). After stirring for 6 h under reflux, the mixture was poured into water (50 mL) and pH was adjusted to 9 with sodium bicarbonate solution. Then the mixture was extracted with DCM (20 mL \times 3). The organic layer was washed with brine, dried over anhydrous Na_2SO_4 , and concentrated under vacuum. The residue was purified by silica gel chromatography (petroleum ether/EtOAc, 1/1, v/v) to afford compound 16 (85 mg, 75%) as a white solid. ^1H NMR (400 MHz, CDCl_3) δ 8.40 (s, 1H), 7.29 (s, 1H), 6.92 (s, 1H), 3.90 (s, 3H), 3.19 (d, $J = 15.6$ Hz, 1H), 3.00–2.96 (m, 1H), 2.89–2.77 (m, 2H), 2.71 (s, 3H), 2.22 (s, 3H), 1.99 (dd, $J = 12.5, 5.5$ Hz, 1H), 1.88 (d, $J = 12.4$ Hz, 1H), 1.82–1.74 (m, 1H), 1.61 (s, 2H), 1.38 (s, 3H), 1.36 (s, 3H), 1.18 (s, 3H). ^{13}C NMR (100 MHz, CDCl_3) δ 172.36, 165.98, 158.51, 157.59, 156.13, 147.71, 130.01, 127.89, 127.85, 123.67, 108.49, 55.81, 49.42, 40.88, 39.72, 37.28, 31.28, 30.33, 25.81, 24.08, 23.18, 20.59, 18.78. HRMS (ESI): calcd for $\text{C}_{23}\text{H}_{31}\text{N}_4\text{O}$ $[\text{M} + \text{H}]^+$, 379.2492; found, 379.2468.

Compound 17. To a solution of compound 7 (109 mg, 0.30 mmol) in anhydrous THF (15 mL) was added methylmagnesium chloride dropwise at 0 °C under nitrogen. After stirring for 6 h at room temperature, the reaction was quenched by saturated aqueous NH_4Cl solution with ice cooling. The mixture was extracted with EtOAc (20 mL \times 3). The organic layer was washed with brine, dried over Na_2SO_4 and concentrated under vacuum. The residue was purified by silica gel chromatography (petroleum ether/EtOAc, 5/1, v/v) to afford compound 17 (98 mg, 86%) as white solid. ^1H NMR (400 MHz, CDCl_3) δ 8.39 (s, 1H), 7.01 (s, 1H), 6.89 (s, 1H), 4.16 (s, 1H), 3.95 (s, 3H), 3.15 (d, $J = 15.5$ Hz, 1H), 2.94–2.81 (m, 2H), 2.76 (d, $J = 15.5$ Hz, 1H), 2.70 (s, 3H), 1.98 (dd, $J = 12.6, 5.8$ Hz, 1H), 1.86 (d, $J = 12.4$ Hz, 1H), 1.80–1.73 (m, 1H), 1.60 (d, $J = 2.8$ Hz, 6H), 1.37 (s, 3H), 1.34 (s, 3H), 1.15 (s, 3H). ^{13}C NMR (100 MHz, CDCl_3) δ 172.30, 165.98, 157.60, 155.40, 145.33, 133.94, 127.68, 126.45, 123.65, 108.27, 72.24, 55.41, 49.39, 40.85, 39.66, 36.94, 31.25, 30.56, 29.76, 29.69, 25.83, 24.05, 23.14, 20.63. HRMS (ESI): calcd for $\text{C}_{24}\text{H}_{33}\text{N}_2\text{O}_2$ $[\text{M} + \text{H}]^+$, 381.2537; found, 381.2514.

Compound 18. This compound was obtained from compound 7 employing method D. White solid. Yield 67%. ^1H NMR (400 MHz, CDCl_3) δ 8.39 (s, 1H), 6.88 (s, 1H), 6.82 (s, 1H), 3.86 (s, 3H), 3.17 (d, $J = 15.6$ Hz, 1H), 2.93–2.81 (m, 2H), 2.76 (d, $J = 15.8$ Hz, 1H), 2.70 (s, 3H), 2.60 (qd, $J = 7.4, 2.4$ Hz, 2H), 1.97 (dd, $J = 12.7, 5.8$ Hz, 1H), 1.87 (d, $J = 12.3$ Hz, 1H), 1.81–1.73 (m, 1H), 1.37 (s, 3H), 1.34 (s, 3H), 1.20 (t, $J = 7.5$ Hz, 3H), 1.16 (s, 3H). ^{13}C NMR (100 MHz, CDCl_3) δ 172.38, 165.89, 157.61, 155.92, 143.94, 130.78, 129.35, 127.23, 123.88, 107.18, 55.51, 49.52, 40.97, 39.66, 36.96, 31.27, 30.41, 25.86, 24.11, 23.15, 22.73, 20.69, 14.07. HRMS (ESI): calcd for $\text{C}_{23}\text{H}_{31}\text{N}_2\text{O}$ $[\text{M} + \text{H}]^+$, 351.2431; found, 351.2417.

Compound 19. This compound was obtained from compound 7 employing method B. White solid. Yield 93%. ^1H NMR (400 MHz, CDCl_3) δ 8.38 (s, 1H), 7.05 (d, $J = 5.2$ Hz, 1H), 6.85 (s, 1H), 5.08–5.02 (m, 1H), 3.90 (s, 3H), 3.16 (d, $J = 15.5$ Hz,



1H), 2.95–2.78 (m, 3H), 2.70 (s, 3H), 1.98 (dd, $J = 12.6, 5.6$ Hz, 1H), 1.88–1.70 (m, 3H), 1.51 (dd, $J = 6.5, 2.8$ Hz, 3H), 1.37 (s, 3H), 1.34 (s, 3H), 1.15 (s, 3H). ^{13}C NMR (100 MHz, CDCl_3) δ 172.34, 165.98, 157.59, 155.16, 145.60, 131.58, 127.70, 126.80, 123.69, 107.50, 66.42, 55.42, 49.47, 40.94, 39.68, 37.08, 31.27, 30.48, 25.82, 24.09, 23.15, 22.81, 20.61. HRMS (ESI): calcd for $\text{C}_{23}\text{H}_{31}\text{N}_2\text{O}_2$ $[\text{M} + \text{H}]^+$, 367.2380; found, 367.2382.

Compound 20. This compound was obtained from compound 19 employing method E. White solid. Yield 80%. ^1H NMR (400 MHz, CDCl_3) δ 8.39 (s, 1H), 7.20 (s, 1H), 6.99 (dd, $J = 17.8, 11.2$ Hz, 1H), 6.85 (s, 1H), 5.72 (dd, $J = 17.7, 1.6$ Hz, 1H), 5.24 (dd, $J = 11.1, 1.6$ Hz, 1H), 3.88 (s, 3H), 3.17 (d, $J = 15.5$ Hz, 1H), 2.98–2.92 (m, 1H), 2.87–2.74 (m, 2H), 2.70 (s, 3H), 1.98 (dd, $J = 12.6, 5.7$ Hz, 1H), 1.87 (d, $J = 13.3$ Hz, 1H), 1.81–1.74 (m, 1H), 1.37 (s, 3H), 1.35 (s, 3H), 1.16 (s, 3H). ^{13}C NMR (100 MHz, CDCl_3) δ 172.29, 165.96, 157.61, 155.33, 146.45, 131.29, 127.62, 127.03, 125.08, 123.69, 114.26, 107.97, 55.75, 49.43, 40.89, 39.69, 37.18, 31.27, 30.42, 25.85, 24.06, 23.17, 20.63. HRMS (ESI): calcd for $\text{C}_{23}\text{H}_{29}\text{N}_2\text{O}$ $[\text{M} + \text{H}]^+$, 349.2274; found, 349.2270.

Compound 21. This compound was obtained from compound 7 employing method C. White solid. Yield 70%. ^1H NMR (400 MHz, CDCl_3) δ 8.45 (s, 1H), 8.44 (s, 1H), 7.01 (s, 1H), 4.05 (s, 3H), 3.30 (d, $J = 15.5$ Hz, 1H), 2.91 (d, $J = 15.4$ Hz, 1H), 2.81–2.75 (m, 2H), 2.71 (s, 3H), 2.61 (s, 3H), 2.43 (dd, $J = 13.5, 4.1$ Hz, 1H), 1.39 (s, 3H), 1.38 (s, 3H), 1.29 (s, 3H). ^{13}C NMR (100 MHz, CDCl_3) δ 198.40, 195.80, 170.80, 166.61, 162.65, 157.85, 157.70, 130.59, 127.76, 124.13, 122.08, 107.23, 55.99, 47.33, 39.64, 39.43, 37.73, 36.46, 31.37, 30.72, 25.84, 23.70, 22.04. HRMS (ESI): calcd for $\text{C}_{23}\text{H}_{27}\text{N}_2\text{O}_3$ $[\text{M} + \text{H}]^+$, 379.2016; found, 379.2009.

Testosterone assays

TM3 cells were seeded in 24-well plates at a density of 1.0×10^5 cells per well and incubated overnight. The culture medium was then replaced with fresh medium containing various compounds (20 μM) respectively. After incubation for 24 h, the culture medium was collected and centrifuged at $1000 \times g$ for 5 min. Testosterone concentrations in the supernatant were determined with enzyme-linked immunosorbent assay (EISA) kits (Shanghai XinFan Biological Technology Co Ltd, Shanghai, China).

Cell lines and culture conditions

The cell lines used in this study were obtained from the American Type Culture Collection (ATCC). NCM460, L-02 and PTN1A cell lines were cultured in RPMI 1640 medium containing 10% foetal bovine serum (FBS), and TM3 mouse Leydig cell line was cultured in Dulbecco's modified eagle's medium (DMEM)/F12 medium, which was supplemented with 2.5% FBS and 5% horse serum. Moreover, HAF cell line was cultured in DMEM medium with 1% glutamine and 10% FBS. All cells were incubated at 37 °C and 5% CO_2 incubator.

Cell viability assay

The cell viability of cell lines in the presence of this series of compounds was determined by SRB (Sigma Aldrich) assay. In

brief, cells were plated at the appropriate cell densities in 96-well plates during the experiment. After incubation for 24 h, the cells were treated with different concentrations of various compounds for 24 h. Control group were exposed to DMSO at a concentration equivalent to that of the compound-treated cells. After treatment, 25 μL of 50% TCA was added for cell fixation at 4 °C. After 1 hour or more, the plates were washed by water for five times. The plates were allowed to dry using hair dryer followed by being dyed with 100 μL 0.4% SRB for 10 min. After dying, the plates were washed by 1% acetic acid to remove the dye and allowed to dry using hair dryer. 100 μL of 10 mM Tris-based solution was added to each well, and absorbance was measured using a 96-well plate reader at 515 nm. Three independent experiments were carried out in triplicate. The IC_{50} was calculated using GraphPad Software.

Western blot analysis

Cells were exposed to various concentrations of compounds for indicated time, and total cellular protein was lysed in RIPA buffer [50 mM Tris-HCl (pH 7.4), 150 mM NaCl, 5 mM EDTA, 1% Triton X-100, 1% sodium deoxycholic acid, 0.1% SDS, 2 mM phenylmethanesulfonyl fluoride (PMSF), 30 mM Na_2HPO_4 , 50 mM NaF, 1 mM Na_3VO_4] containing protease/phosphatase inhibitors (Roche). Lysates were combined with sample loading buffer and heated at 100 °C. Protein concentrations were measured by a Bicinchoninic acid assay (Thermo Scientific). Lysates were mixed with sample loading buffer and heated at 100 °C. for 15 min. After separated by 8–15% SDS-PAGE, extracted protein were transferred to nitrocellulose membranes. Membranes were incubated in 5% (w/v) bovine serum albumin (TBST/BSA) and stored overnight at 4 °C on a shaker with specific primary antibodies (1/1000 in TBST/BSA). Then membranes were washed with TBST and incubated for 45 min with secondary antibody (1/10 000 in TBST/BSA) at room temperature. Immunoreactive proteins were visualized using the Odyssey Fluorescence Scanner (LI-COR Bioscience, Inc., Lincoln, NE, USA). The StAR (ab133657; 1 : 3000), HSD3B1 (ab65156; 1 : 1000) and CYP11A1 (BS6578; 1 : 1000) antibodies were purchased from Abcam (Cambridge, MA) and Bioworld Technology, Inc (St. Paul, MN). LC3A/B (12741; 1 : 3000) and Atg5 (12994; 1 : 3000) antibodies were purchased from Cell Signaling Technology (Danvers, MA). The β -actin and GAPDH antibody (1 : 10 000) was purchased from Sigma (St Louis, MO). The secondary antibody was conjugated with IRDye 680/800 (Millennium Science; 926-32221, 926-32210; 1 : 10 000).

Quantitative real-time polymerase chain reaction (qRT-PCR)

Total RNA was extracted from cells with TRIZOL reagent (Invitrogen, Carlsbad, CA, USA) according to the manufacturer's protocols. Total RNA (1 μg) was converted to cDNA using a PrimeScript reverse transcription Master Mix kit (TaKaRa, Japan) according to the manufacturer's instructions. Quantitative RT-PCR was performed using SYBR Premix Ex Taq™ Kit (TaKaRa, Japan). The relative expression of StAR, β 3-HSD and CYP11A1 were analyzed by RT-PCR with β -actin as an internal control.



Three independent experiments were carried out in triplicate. The gene-specific primers are listed in Table S1.†

Statistical analysis

Grouped data are expressed as mean \pm s.d. Significance between groups was analysed by one-way analysis of variance or Student's *t*-test using GraphPad Prism 5.0 (GraphPad Software). Data were expressed as mean and S.D., and *P* < 0.05 was considered significant. All experiments were performed at least three times except for animal experiments.

Conflicts of interest

There are no conflicts of interest to declare.

Abbreviations

NPL	Natural product-like
LOH	Late-onset hypogonadism
StAR	Steroidogenic acute regulatory
3 β -HSD	3 β -Hydroxysteroid dehydrogenase
CYP11A1	Cytochrome P450 family 11 subfamily A member 1
TRT	Testosterone replacement therapy
ED	Erectile dysfunction
LH	Luteinizing hormone
cAMP	Cyclic adenosine monophosphate
PKA	Protein kinase A
TG	Triglyceride
IBX	2-Iodoxybenzoic acid
THF	Tetrahydrofuran
SAR	Structure–activity relationship
Rt	Room temperature
ELISA	Enzyme-linked immuno sorbent assay
IC ₅₀	Half maximal inhibitory concentration
SRB	Sulforhodamine B
RT-PCR	Real-time polymerase chain reaction
NMR	Nuclear magnetic resonance
GAPDH	Reduced glyceraldehyde-phosphate dehydrogenase
LC3	Light chain 3
ATG5	Autophagy related 5
H&E	Haematoxylin and eosin
IHC	Immunohistochemical
TMS	Tetramethylsilane
IF	Immunofluorescence
PMSF	Phenylmethanesulfonyl fluoride
SDS-PAGE	Sodium dodecyl sulfate polyacrylamide gel electrophoresis
TBST	Tris buffered saline tween
BSA	Bovine serum albumin
FITC	Fluorescein isothiocyanate

Acknowledgements

This work was supported by Shanghai Science and Technology Council (Grant 18ZR1411200), National Natural Science Foundation of China (No. 81773204, No. 81472788) and Major State

Basic Research Development Program of China (2015CB910400).

References

- 1 A. Vasas and J. Hohmann, *Chem. Rev.*, 2014, **114**, 8579–8612.
- 2 X. Rao, X. Huang, L. He, J. Song, Z. Song and S. Shang, *Comb. Chem. High Throughput Screening*, 2012, **15**, 840–844.
- 3 N. Duarte, A. Varga, G. Cherepnev, R. Radics, J. Molnar and M. J. U. Ferreira, *Bioorg. Med. Chem.*, 2007, **15**, 546–554.
- 4 K. Yoshikawa, N. Kokudo, M. Tanaka, T. Nakano, H. Shibata, N. Aragaki, T. Higuchi and T. Hashimoto, *Chem. Pharm. Bull.*, 2008, **56**, 89–92.
- 5 Y. Tian, W. Xu, C. Zhu, S. Lin, Y. Li, L. Xiong, S. Wang, L. Wang, Y. Yang, Y. Guo, H. Sun, X. Wang and J. Shi, *J. Nat. Prod.*, 2011, **74**, 1221–1229.
- 6 C. Lipina and H. S. Hundal, *Cell. Signalling*, 2014, **26**, 2343–2349.
- 7 T. T. Dao, K. Y. Lee, H. M. Jeong, P. H. Nguyen, T. L. Tran, P. T. Thuong, B. T. Nguyen and W. K. Oh, *J. Nat. Prod.*, 2011, **74**, 2526–2531.
- 8 M. Shahlaei, S. M. Ghanadian, A. M. Ayatollahi, M. A. Mesaik, O. M. Abdalla, S. Afsharypuor and M. Rabbani, *Med. Chem. Res.*, 2003, **22**, 1795–1803.
- 9 C. S. Kim, L. Subedi, S. Y. Kim, S. U. Choi, K. H. Kim and K. R. Lee, *J. Nat. Prod.*, 2016, **79**, 387–394.
- 10 E. J. Richmond and A. D. Rogol, *Nat. Clin. Pract. Endocrinol. Metab.*, 2007, **3**, 338–344.
- 11 S. G. Haider, *Endocrinology*, 2007, **148**, 2581–2582.
- 12 C. Wang, E. Nieschlag, R. Swerdloff, H. M. Behre, W. J. Hellstorm, L. J. Gooren, J. M. Kaufman, J. Legros, B. Lunenfeld, A. Morales, J. E. Morley, C. Schulman, I. M. Thompson, W. Weidner and F. C. W. Wu, *Eur. Urol.*, 2009, **55**, 121–130.
- 13 D. Sun, Y. Cui, B. Jin, X. Zhang, X. Yang and C. Gao, *Evid. Based Complement. Alternat. Med.*, 2012, **2012**, 1–10.
- 14 B. Lunenfeld, F. Saad and C. E. Hoesl, *Aging Male*, 2005, **8**, 59–74.
- 15 C. H. Liao, Y. N. Wu, F. Y. Lin, W. K. Tsai, S. P. Liu and H. S. Chiang, *Andrology*, 2013, **1**, 563–569.
- 16 N. Bassil, S. Alkaade and J. E. Morley, *Ther. Clin. Risk Manage.*, 2009, **5**, 427–448.
- 17 S. Kliesch, *Urologe A*, 2010, **49**, 32–36.
- 18 J. I. Makinen and I. Huhtaniemi, *Gerontology*, 2011, **57**, 193–202.
- 19 R. Vigen, C. I. O'Donnell, A. E. Baron, G. K. Grunwald, T. M. Maddox, S. M. Bradley, A. Barqawi, G. Woning, M. E. Wierman, M. E. Plomondon, J. S. Rumsfeld and P. M. Ho, *J. Am. Med. Assoc.*, 2013, **310**, 1829–1836.
- 20 M. L. Dufau, *Annu. Rev. Physiol.*, 1988, **50**, 483–508.
- 21 K. Henriksen, H. Hakovirta and M. Parvinen, *Endocrinology*, 1995, **136**, 3285–3291.
- 22 J. D. Rabinowitz and E. White, *Science*, 2010, **330**, 1344–1348.
- 23 F. Gao, G. Li, C. Liu, H. Gao, H. Wang, W. Liu, M. Chen, Y. Shang, L. Wang, J. Shi, W. Xia, J. Jiao, F. Gao, J. Li, L. Chen and W. Li, *J. Cell Biol.*, 2018, **217**, 2103–2119.



- 24 G. Wei, Y. Wu, X. L. He, T. Liu, M. Liu, J. Luo and W. W. Qiu, *Eur. J. Med. Chem.*, 2017, **131**, 48–67.
- 25 L. G. Yu, T. F. Ni, W. Gao, Y. He, Y. Y. Wang, H. W. Cui, C. G. Yang and W. W. Qiu, *Eur. J. Med. Chem.*, 2015, **90**, 10–20.
- 26 A. Eschenmoser and D. Arigoni, *Helv. Chim. Acta*, 2005, **88**, 3011–3050.
- 27 G. Stork and A. W. Burgstaher, *J. Am. Chem. Soc.*, 1955, **77**, 5068–5077.
- 28 Z. B. Zhang and Q. T. Yang, *Asian J. Androl.*, 2006, **8**, 601–605.

