

Cite this: *RSC Adv.*, 2019, 9, 12784

Convenient and efficient synthesis of novel 11*H*-benzo[5,6][1,4]thiazino[3,4-*a*]isoindol-11-ones derived from 2-bromo-(2/3-substitutedphenyl)-1*H*-indene-1,3(2*H*)-diones†

Satbir Mor * and Suchita Sindhu

An unprecedented formation of 11*H*-benzo[5,6][1,4]thiazino[3,4-*a*]isoindol-11-ones through a one-step reaction of differently substituted 2-aminobenzenethiols and 2-bromo-(2/3-substitutedphenyl)-1*H*-indene-1,3(2*H*)-diones in freshly dried ethanol under reflux conditions has been investigated. This unique transformation probably occurs through an initial nucleophilic substitution followed by ring opening and subsequent intramolecular cyclization. The structures of all the synthesized benzo[1,4]thiazino isoindolinones were established by FTIR, ¹H NMR, ¹³C NMR, HRMS, and X-ray crystallographic analysis. This approach was found to be simple and convenient and provides several advantages such as substantial atom economy, short reaction time and operational simplicity.

Received 30th March 2019
Accepted 16th April 2019

DOI: 10.1039/c9ra02403d

rsc.li/rsc-advances

Introduction

Benzothiazine represents a class of heterocyclic compounds in which benzene is fused with a six-membered thiazine ring containing nitrogen and sulphur atoms. Depending on the positions of nitrogen and sulphur atoms in the ring, benzothiazines exist as 1,2-benzothiazines, 1,3-benzothiazines, 1,4-benzothiazines, 2,3-benzothiazines, 2,4-benzothiazines, 3,4-benzothiazines, *etc.* Among them, 1,4-benzothiazines have gained a lot of interest worldwide due to numerous pharmaceutical activities, such as antidepressant,¹ antitubercular,² antimalarial,³ anti-HIV,⁴ anti-rheumatic,⁵ antihypertensive,⁶ anti-inflammatory,⁷ antidiabetic,⁸ anticancer,⁹ antagonists,¹⁰ *etc.* associated with them. With a unique structural feature due to the presence of a fold along nitrogen–sulphur axis, the benzo[1,4]thiazine skeleton shows some biological activities and structural specificities similar to the phenothiazines.^{11–13} Moreover, benzo[1,4]thiazine occurs naturally as pheomelanin in red hairs and feathers.¹⁴ Additionally, it is a main component of trichochromes (dimers of benzothiazines).¹⁵ Moreover, the synthesis of benzo[1,4]thiazines has attracted the attention of various medicinal and organic chemists for decades. In the 19th century, the synthesis of several benzo[1,4]thiazines was carried out by Unger.¹⁶ Later on, various methods of synthesis of benzo[1,4]thiazines have been developed, which include the condensation of 2-aminobenzenethiol

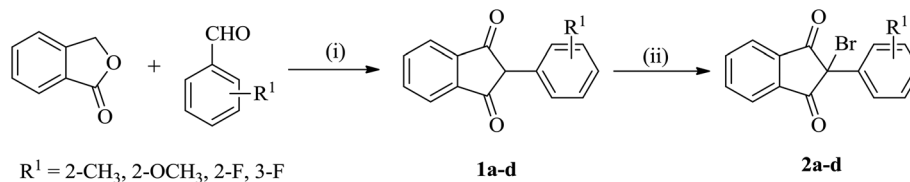
with 2,5-dihydro-2,5-dimethoxyfuran,¹⁷ 1,3-dicarbonyl compounds,¹⁸ α -cyano- β -alkoxy carbonyl epoxides,¹⁹ 1,2-diaroylacetylene with silica supported perchloric acid,²⁰ and many more. Moreover, benzo[*b*][1,4]thiazine-4-carbonitriles were synthesized by copper(i)-catalyzed oxidative coupling of 2-aminobenzothiazole with alkynes followed by intramolecular cyclization.²¹ Recently, the synthesis of benzo[1,4]thiazines was reported by an environmentally benign and efficient method using iron-catalyst, focused on the suitability of iron in comparison with other metals like palladium for reactions involving the preparation of therapeutic agents for human consumption.²²

Isoindolinone moiety is a heterocyclic framework that is present as the core unit in several natural products such as isoindolobenzazocine, chilenine, lennoxamine and nuevamine isolated from various *Berberis* species,²³ fumaridine,²⁴ isoquinoline fumadensine,²⁵ isoindolobenzazepine,²⁶ pictonamine,²⁷ pazinaclo-²⁸ and aristolactam alkaloids.²⁹ The isoindolinone and its derivatives exhibit a wide range of biological activities such as sedatives and hypnotics,³⁰ anxiolytic agents,³¹ antibacterial,³² MDM2-p53 protein–protein interaction and tumor necrosis factor production,³³ antihypertensive,³⁴ antileukemic,³⁵ anti-inflammatory,³⁶ vasodilatory,³⁷ antiviral,³⁸ *etc.* Furthermore, isoindolo[2,1-*a*]quinolones were found to be active against N₂-induced hypoxia.³⁹ Due to extensive biological activities of isoindolinones, consequently, considerable efforts have been made toward their synthesis. In recent years, various new synthetic methodologies have been developed for the preparation of isoindoline derivatives, of which, the most common methods involve the use of transition metal-catalyst such as [CpRhCl₂]₂,⁴⁰ Pd-catalyzed reactions,⁴¹ ultrathin Pt nanowire as catalysts,⁴² (cyclooctadiene)(pentamethylcyclopentadiene) ruthenium

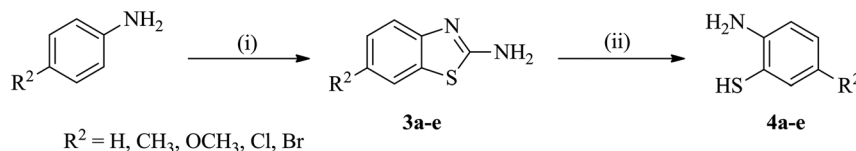
Department of Chemistry, Guru Jambheshwar University of Science & Technology, Hisar-125001, Haryana, India. E-mail: satbir_mor@yahoo.co.in; Fax: +91-1662-276240; Tel: +91-1662-263397

† Electronic supplementary information (ESI) available. CCDC 1897439. For ESI and crystallographic data in CIF or other electronic format see DOI: 10.1039/c9ra02403d

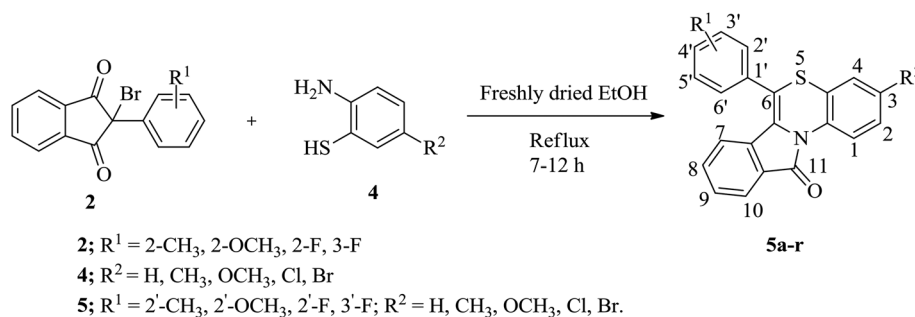




Scheme 1 Synthesis of 2-bromo-(2/3-substitutedphenyl)-1H-indene-1,3(2H)-diones (**2a-d**). Reaction conditions: (i) Na/CH₃OH, CH₃-COOC₂H₅, reflux, 65 °C; (ii) Br₂/CHCl₃, stirring, r.t.



Scheme 2 Synthesis of 5-substituted 2-aminobenzenethiols (**4a-e**). Reaction conditions: (i) NaSCN, Br₂/glacial CH₃COOH, stirring, 0–5 °C; (ii) KOH/H₂O, reflux, 80 °C.



Scheme 3 Synthesis of 11H-benzo[5,6][1,4]thiazino[3,4-a]isoindol-11-ones (**5a-r**).

chloride catalyst,⁴³ microwave method using Cu(OAc)₂·H₂O/DBU,⁴⁴ polymethylhydrosiloxane and fluoride ions as catalyst,⁴⁵ phase transfer catalyst,⁴⁶ lithiation procedures,⁴⁷ electrochemical method,⁴⁸ *etc.* However, these methods have intrinsic drawbacks, which include harsh reaction conditions, employing transition-metal-based catalysts which are often toxic, expensive, difficult to remove, and are normally obtained from limited natural resources. Therefore, it is necessary to develop novel, convenient and efficient approaches for overcoming these problems and readily available starting materials for the synthesis of isoindolinones. Therefore, Zhou *et al.* reported a metal-free strategy for the preparation of isoindolinones with high regioselectivity.⁴⁹

Stimulated by the above observation for benzo[1,4]thiazines and isoindolinones, herein, we report the preparation of several novel 11H-benzo[5,6][1,4]thiazino[3,4-a]isoindol-11-ones (**5**) by the reaction of 2-bromo-(2/3-substitutedphenyl)-1H-indene-1,3(2H)-diones (**2**) and 2-aminobenzenethiols (**4**) with an advantage of good yields and simple work-up procedure.

Results and discussion

The starting reactants *i.e.* 2-bromo-(2/3-substitutedphenyl)-1H-indene-1,3(2H)-diones (**2**) and 5-substituted 2-aminobenzenethiols (**4**) were obtained as outlined in Schemes 1 and 2, respectively. The synthesis of **2** was carried out by the reaction of phthalide with

appropriate 2/3-substituted benzaldehyde in the presence of ethyl acetate and sodium methoxide,^{50,51} which upon subsequent bromination in Br₂/chloroform afforded the corresponding 2-bromo-(2/3-substitutedphenyl)-1H-indene-1,3(2H)-diones (**2**) in good yields.⁵² The synthesis of 5-substituted 2-aminobenzenethiols (**4**) began with 4-substituted anilines as the starting materials, which on reaction with sodium thiocyanate in Br₂/glacial CH₃-COOH afforded the corresponding 2-amino-6-substituted

Table 1 Optimization of the reaction conditions^{a,b}

Entry	Solvent	Time (h)	Temp (°C)	Yield ^c (%)
1	EtOH	12	80	30
2	Freshly dried EtOH	7	80	78
3	DMF	16	85	0
4	Toluene	16	98	0
5	THF	16	85	0
6	Dry EtOH + dry DMF	20	90	Trace
7	Dry EtOH + dry toluene	20	90	0
8	Dry EtOH + dry THF	20	90	Trace
9	MeOH	20	70	24
10	DMSO	20	80	0
11	MeCN	20	80	0

^a Bold represents the most optimal reactions conditions for the synthesis. ^b Reaction conditions: **2** ($R^1 = 3\text{-F}$) (3 mmol), **4** ($R^2 = \text{CH}_3$) (3 mmol), solvent (20 mL). ^c Yield after chromatography on silica gel.



benzothiazoles (3) in good yields. Thereafter, the benzothiazoles (3) were subjected to base-catalyzed hydrolytic fission to give the respective 5-substituted 2-aminobenzenethiols (4) in moderate to good yields following the procedure as described in literature.^{53–55}

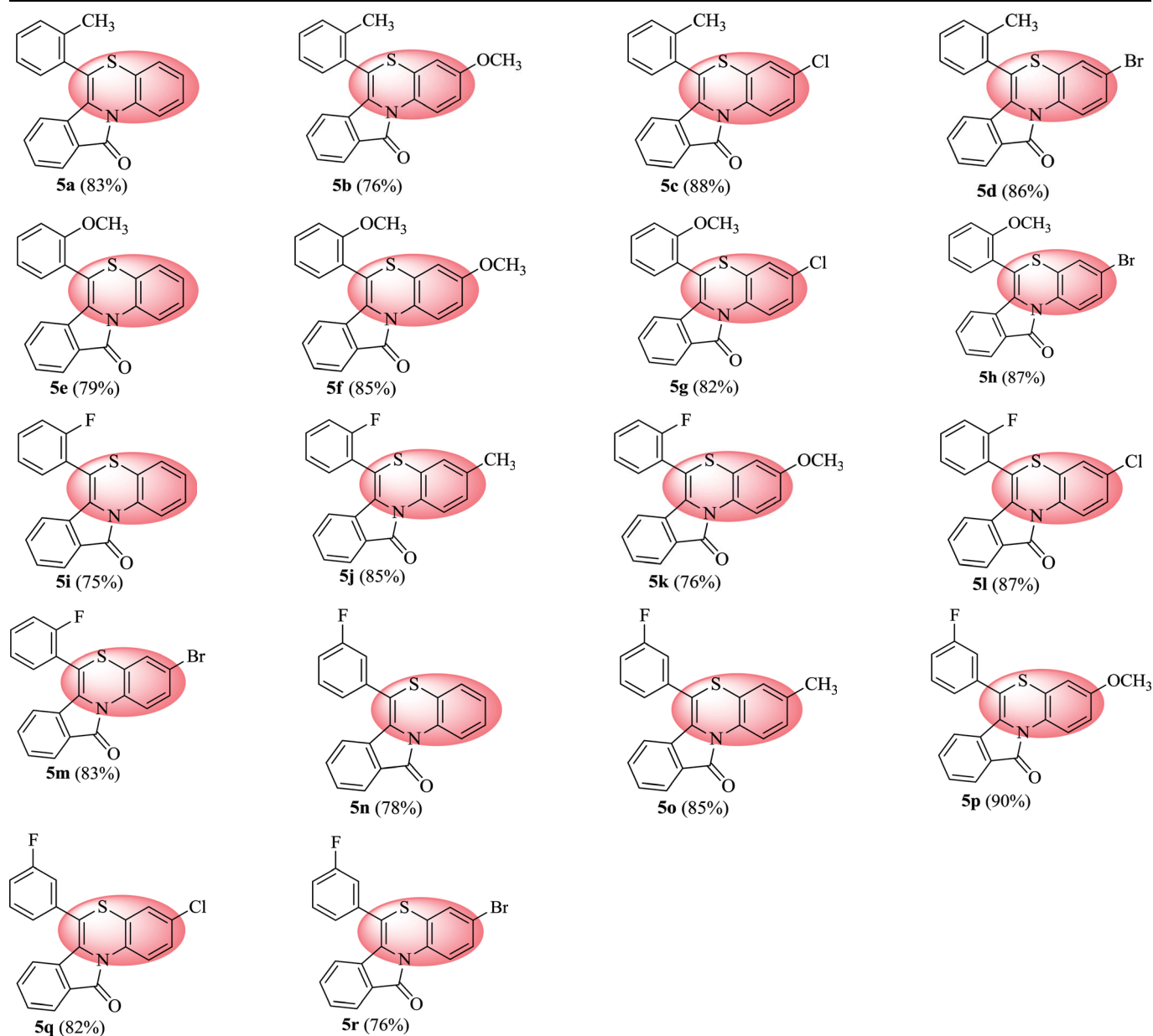
The synthesis of 11*H*-benzo[5,6][1,4]thiazino[3,4-*a*]isoindol-11-ones (5a–r) was carried out by the reaction of 2-bromo-(2/3-substitutedphenyl)-1*H*-indene-1,3(2*H*)-diones (2) and 5-substituted 2-aminobenzenethiols (4) in presence of freshly dried ethanol under reflux conditions on a water bath for 7–12 hours as outlined in Scheme 3.

To establish the optimal reaction conditions, 2 ($R^1 = 3\text{-F}$) and 4 ($R^2 = \text{CH}_3$) were used as a model substrates for the formation of the benzo[1,4]thiazino isoindolinone (5o). The reaction was carried out in an assortment of different solvents

to study the impact of solvent on the outcome of the reaction. The results of standardized study are depicted in Table 1.

The results for the optimization of reaction conditions summarized in Table 1 revealed that freshly dried ethanol has been proved the best solvent for the reaction because it provided highest yield of the product (5o) in minimum time as compared to utilizing other solvents/mixture of solvents. With the optimization conditions in hand, we investigated the scope of this strategy for the construction of a small library of benzo[1,4]thiazino isoindolinones (5) (Table 2). The reaction was tested with diversely substituted 2-bromo-(2/3-substitutedphenyl)-1*H*-indene-1,3(2*H*)-diones (2) and 5-substituted 2-amino-benzenethiols (4) in freshly dried ethanol. We were delighted to note that all the substrates responded positively to give desired

Table 2 Structures of various 11*H*-benzo[5,6][1,4]thiazino[3,4-*a*]isoindol-11-ones (5a–r) with their yields (%)



products (**5a-r**) in 7–12 h in good to excellent yields. A silica gel column chromatographic purification/separation was required which was performed by using hexane : ethyl acetate (9 : 1, v/v) as an eluent. It was observed that changing the substitution pattern in (**2**) and (**4**) did not affect the outcome of the reaction.

The structures of all the synthesized benzo[1,4]thiazino isoindolinones (**5a-r**) were ascertained by FTIR, ^1H NMR, ^{13}C NMR, HRMS, and single-crystal X-ray crystallography analysis. For instance, the FTIR spectrum of **5b** displayed an absorption band at 1703 cm^{-1} which could be easily assigned to C=O stretching, however C=C stretching was observed at 1649 cm^{-1} . ^1H NMR spectrum of **5b** exhibited a doublet integrating for one proton centered at δ 6.22 ($J = 7.96\text{ Hz}$) due to $\text{C}_7\text{-H}$ and another one-proton doublet at δ 7.93 ($J = 7.68\text{ Hz}$) due to $\text{C}_{10}\text{-H}$. A one-proton doublet appeared in the most downfield region centered at δ 9.20 ($J = 9.32\text{ Hz}$) was expediently assigned to $\text{C}_1\text{-H}$. The ^{13}C NMR spectrum of **5b** exhibited a signal in the most downfield region at δ 164.82 ppm, which was safely attributed to 5-membered ring carbonyl carbon *i.e.* C_{11} . These assignments were further confirmed by 2D NMR spectral studies *i.e.* $^1\text{H}\text{-}^1\text{H}$ COSY, $^1\text{H}\text{-}^{13}\text{C}$ HMQC, HSQC and DEPT. The $^1\text{H}\text{-}^1\text{H}$ correlations between δ 9.20 ($\text{C}_1\text{-H}$) and δ 6.81 ($\text{C}_2\text{-H}$); δ 6.81 ($\text{C}_2\text{-H}$) and δ 6.64 ($\text{C}_4\text{-H}$); δ 7.93 ($\text{C}_{10}\text{-H}$) and δ 7.35–7.47 ($\text{C}_9\text{-H}$); δ 7.35–7.47 ($\text{C}_9\text{-H}$) and δ 7.23–7.27 ($\text{C}_8\text{-H}$); δ 7.23–7.27 ($\text{C}_8\text{-H}$) and δ 6.22 ($\text{C}_7\text{-H}$) were established through COSY spectrum. The HSQC experiments revealed the assignment of carbon signals at δ 120.58 (C_1), δ 112.62 (C_2), δ 110.84 (C_4), δ 121.30 (C_7), δ 131.61 (C_8), δ 122.86 (C_{10}) because the key correlation ($^1\text{H}\text{-}^{13}\text{C}$) was observed as: $\delta_{\text{H}} 9.20$ ($\text{C}_1\text{-H}$) \rightarrow δ_{C} 120.58 (C_1); $\delta_{\text{H}} 6.81$ ($\text{C}_2\text{-H}$) \rightarrow δ_{C} 112.62 (C_2), $\delta_{\text{H}} 6.64$ ($\text{C}_4\text{-H}$) \rightarrow δ_{C} 110.84 (C_4), $\delta_{\text{H}} 6.22$ ($\text{C}_7\text{-H}$) \rightarrow δ_{C} 121.30 (C_7), $\delta_{\text{H}} 7.23\text{-}7.27$ ($\text{C}_8\text{-H}$) \rightarrow δ_{C} 131.61 (C_8) and $\delta_{\text{H}} 7.93$ ($\text{C}_{10}\text{-H}$) \rightarrow δ_{C} 122.86 (C_{10}). Additionally, HMBC and ^{13}C -DEPT spectra of **5b** proved helpful towards chemical shift assignments of proton and carbon signals. However, the remaining aliphatic and aromatic protons (^1H NMR) and carbons (^{13}C NMR) resonated in the expected regions. Further,

HRMS analysis result of **5b** was found in full agreement with its molecular weight (*vide* experimental). The structure of **5b** was further confirmed by X-ray crystallography study. Purification of **5b** from a solution of hexane : ethyl acetate (9 : 1, v/v) at room temperature yielded suitable yellow coloured single crystals for X-ray analysis. The crystal structure data of **5b** has been deposited at the Cambridge Crystallographic Data Centre with deposition number CCDC 1897439. X-ray crystal structure (ORTEP diagram) of representative compound **5b** is depicted in Fig. 1. The summary of crystal structure data is provided in ESI (Tables 3 and 4 \dagger).

Similar to **5b**, the structure of the remaining derivatives (**5a**, **5c-r**) have been ascertained on the basis of spectral (FTIR, ^1H NMR, ^{13}C NMR and HRMS) results. The FTIR spectra of all these compounds (**5a**, **5c-r**), the presence of C=O group, in each case, was confirmed by the appearance of an absorption band in the region at $1681\text{-}1710\text{ cm}^{-1}$ while C=C stretching frequencies were observed in the range of $1637\text{-}1660\text{ cm}^{-1}$. The ^1H NMR spectra of compounds (**5a**, **5c-r**), followed the same pattern as observed in **5b**. The ^1H NMR spectra of compounds (**5a**, **5c-r**), in each case, exhibited a one-proton doublet in the region at δ 6.22–6.66 ($J = 7.92\text{-}8.64\text{ Hz}$) undoubtedly assignable to $\text{C}_7\text{-H}$ and another doublet integrating for one proton in the region at δ 7.92–8.07 ($J = 6.52\text{-}7.76\text{ Hz}$) due to $\text{C}_{10}\text{-H}$. The most characteristic feature of ^1H NMR spectra of the compounds (**5a**, **5c-r**) was appearance of a doublet integrating for one proton in the most downfield region at δ 9.04–9.23 ($J = 8.44\text{-}9.40\text{ Hz}$) due to $\text{C}_1\text{-H}$. The remaining aliphatic and aromatic protons, in each case, showed their signals in the expected regions. The salient feature of ^{13}C NMR spectra of compounds (**5a**, **5c-r**) is the downfield shifting of signal due to C_{11} (carbonyl carbon of 5-membered ring) which was observed in the region at δ 164.65–165.26 as compared to remaining carbon atoms. However, the remaining carbons resonated in the expected regions. Further, the results obtained from HRMS analysis were found in consistent with their molecular formulae (*vide* experimental).

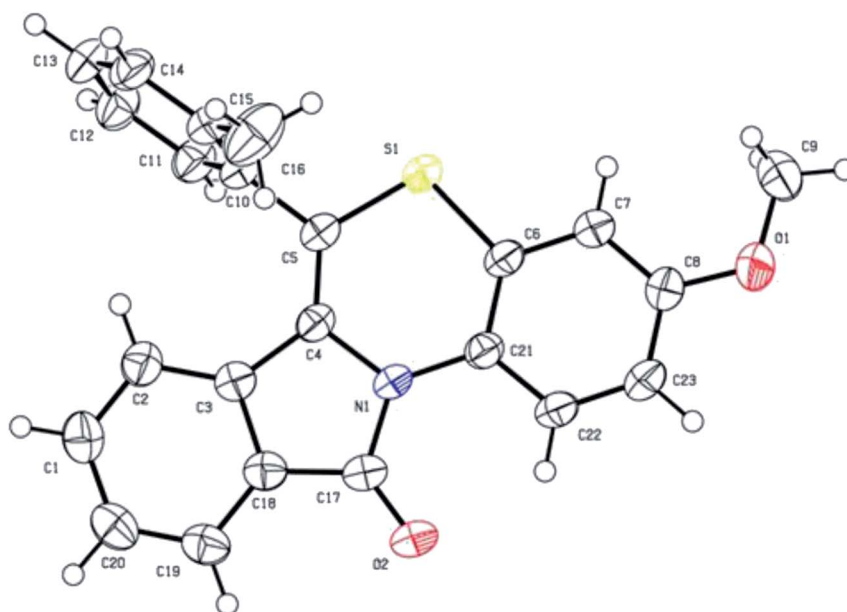
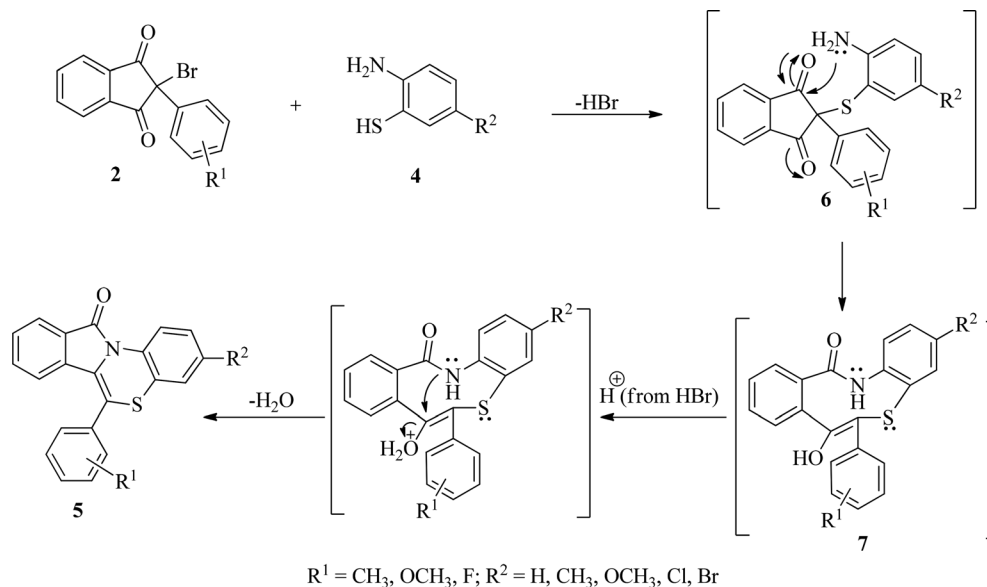


Fig. 1 X-ray crystal structure of 3-methoxy-6-(*o*-tolyl)-11*H*-benzo[5,6][1,4]thiazino[3,4-*a*]isoindol-11-one (**5b**).





Scheme 4 Plausible mechanism for the formation of 11H-benzo[5,6][1,4]thiazino[3,4-a]isoindol-11-ones (5).

On the basis of results of the studies, a plausible mechanism for the formation of benzo[1,4]thiazino isoindolinones (5) has been presented in Scheme 4. It has been proposed that initially formation of 6 occur by nucleophilic attack of SH group of 2-aminobenzenethiols (4) on C₂ of 2-bromo-(2/3-substitutedphenyl)-1H-indene-1,3(2H)-diones (2) with elimination of HBr which is accompanied by intramolecular rearrangement to give intermediate 7. A subsequent cyclodehydration of intermediate 7 occurs to produce 11H-benzo[5,6][1,4]thiazino [3,4-a]isoindol-11-ones (5).

Conclusion

In conclusion, we have successfully developed a simple and efficient protocol for the synthesis of biologically important novel heterocyclic frameworks *i.e.* 11H-benzo[5,6][1,4]thiazino[3,4-a]isoindol-11-ones (5). The reaction possibly proceeds through the elimination of hydrogen bromide followed by sequential intramolecular cyclization to afford the target products. This methodology offers several advantages like short reaction time, appreciable atom economy and easy work up procedure for the synthesis of these interesting molecules. Hopefully, these findings will prove helpful to molecular architectures in the field of organic synthesis. The biological evaluation studies of the synthesized benzo[1,4]thiazino isoindolinones (5) are underway and results will be reported in due course of work.

Experimental

General section

All the reagents and solvents were utilized as obtained from chemical suppliers without further additional purification and 2-aminothiophenol was obtained from Sigma-Aldrich. NMR spectra were recorded in CDCl₃ at 400 MHz (for ¹H NMR) and at 100 MHz (for ¹³C NMR) on a 400 MHz Bruker Avance-III

spectrometer and using tetramethylsilane (TMS) as internal standard at room temperature. Chemical shifts (δ) are reported in parts per million (ppm) and coupling constants (J) are expressed in hertz (Hz), and abbreviations in NMR spectra are s – singlet, d – doublet, dd – doublet of doublet, t – triplet and m – multiplet. The Fourier Transform Infrared (FTIR) spectra were recorded on IR affinity-1 FTIR (Shimadzu) spectrophotometer by using KBr, and abbreviations in FTIR spectra are s – strong, m – medium, w – weak. HRMS analysis was performed using LC-MS on SCIEX 5600⁺ QTOF operating at positive full scan mode (120 000 FWHM) in the range of 100–1000 *m/z*. The settings for electrospray ionization were as follows: oven temperature of 150 °C and source voltage of 3.6 kV. Samples were diluted to a final concentration of 0.1 mg mL⁻¹ in MeCN. The samples were injected by direct infusion into the mass spectrometer using an autosampler. The X-ray single crystal structure was performed on a Bruker Kappa Apex II diffractometer using radiation source from fine-focus sealed tube in the ω and ϕ -scan mode. An empirical absorption correction was applied using the SADABS (Bruker, 1999) program and data processing was accomplished with the SAINT (Bruker, 2004) processing program. The structure was solved in the space group *P*-1 by the direct methods and refined on F^2 by full-matrix least-squares using SHELXL-97 (Sheldrick, 1997). Thin-layer chromatography (TLC) was checked on precoated TLC plates (SIL G/UV₂₅₄, ALUGRAM) as the stationary phase and a mixture of ethyl acetate and hexane was used as mobile phase. Melting points (mp) were observed in open glass capillaries with Electrothermal Melting Point apparatus, LABCO Co., India and are uncorrected.

Experimental section

General method for the preparation of 2-bromo-(2/3-substitutedphenyl)-1H-indene-1,3(2H)-diones (2). The compound (2) was prepared by the reaction of phthalide with appropriately substituted benzaldehyde in presence of ethyl acetate and



sodium methoxide,^{50,51} which upon subsequent bromination in Br₂/chloroform afforded the corresponding 2-bromo-(2/3-substitutedphenyl)-1*H*-indene-1,3(2*H*)-diones (2) in good yields.⁵²

General procedure for the preparation of 5-substituted-2-aminobenzenethiols (4). The reaction of 4-substituted anilines with sodium thiocyanate in Br₂/glacial CH₃COOH afforded the corresponding 2-amino-6-substituted benzothiazoles (3), which upon subsequent base-catalyzed hydrolytic fission afforded the corresponding 5-substituted 2-aminobenzenethiols (3) in moderate to good yields as described in the literature.^{53–55}

General procedure for the preparation of 11*H*-benzo[5,6][1,4]thiazino[3,4-*a*]isoindol-11-ones (5a–r). A mixture of equimolar quantities of 2-bromo-(2/3-substitutedphenyl)-1*H*-indene-1,3(2*H*)-dione (3.0 mmol) and 5-substituted 2-amino-benzenethiol (3.0 mmol) taken in a round bottomed flask was charged with freshly dried ethanol (20 mL) and heated to reflux on a water bath for 7–12 h. Progress of the reaction was monitored by TLC by withdrawing the aliquots from reaction mixture at different intervals of time. After completion of the reaction, the contents were cooled at ambient temperature and the solid thus separated out was filtered, and purified by column chromatography on silica gel (60–120 mesh). Elution of the column with hexane : ethyl acetate (90 : 10, v/v) afforded the corresponding 11*H*-benzo[5,6][1,4]thiazino[3,4-*a*]isoindol-11-ones (5a–r) in good yields.

6-(*o*-Tolyl)-11*H*-benzo[5,6][1,4]thiazino[3,4-*a*]isoindol-11-one (5a). Yield: 83% as yellow solid, mp 164–170 °C; FTIR (KBr): ν_{\max} = 3068 (s), 2871 (s), 1689 (s), 1649 (m), 1557 (m), 1477 (s) cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 9.23 (d, *J* = 8.44 Hz, 1H, H-1), 7.95 (d, *J* = 7.64 Hz, 1H, H-10), 7.35–7.47 (m, 6H, H-8, H-9, H-3', H-4', H-5', H-6'), 7.25–7.29 (m, 2H, H-3, H-4), 7.00–7.07 (m, 1H, H-2), 6.23 (d, *J* = 8.00 Hz, 1H, H-7), 2.35 (s, 3H, CH₃) ppm; ¹³C NMR (400 MHz, CDCl₃): δ 165.23 (C-11), 139.08, 137.59, 133.50, 133.39, 133.28, 131.88, 131.11, 130.02, 129.96, 128.30, 127.76, 127.23, 125.64, 125.44, 123.82, 123.05, 121.36, 119.83, 119.38, 117.60, 18.96 (CH₃) ppm; HRMS (ESI⁺) *m/z* calcd for C₂₂H₁₅NOS [M + H]⁺ 342.0953, found 342.0925.

3-Methoxy-6-(*o*-tolyl)-11*H*-benzo[5,6][1,4]thiazino[3,4-*a*]isoindol-11-one (5b). Yield: 76% as a yellow solid, mp 210–214 °C; FTIR (KBr): ν_{\max} = 3068 (s), 2927 (s), 1703 (s), 1649 (m), 1588 (m), 1496 (m) cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 9.20 (d, *J* = 9.32 Hz, 1H, H-1), 7.93 (d, *J* = 7.68 Hz, 1H, H-10), 7.35–7.47 (m, 5H, H-9, H-3', H-4', H-5', H-6'), 7.23–7.27 (m, 1H, H-8), 6.81 (dd, *J* = 9.20 and 2.92 Hz, 1H, H-2), 6.64 (d, *J* = 2.88 Hz, 1H, H-4), 6.22 (d, *J* = 7.96 Hz, 1H, H-7), 3.81 (s, 3H, OCH₃), 2.37 (s, 3H, CH₃) ppm; ¹³C NMR (400 MHz, CDCl₃): δ 164.82 (C-11), 156.58 (C-3), 137.65, 133.44, 131.61, 131.08, 130.72, 130.03, 130.01, 128.41, 128.28, 127.32, 127.19, 126.86, 122.86, 121.30, 121.10, 120.58, 116.79, 112.62, 110.84, 55.56 (OCH₃), 19.00 (CH₃) ppm; HRMS (ESI⁺) *m/z* calcd for C₂₃H₁₇NO₂S [M + H]⁺ 372.1058, found 372.1018.

3-Chloro-6-(*o*-tolyl)-11*H*-benzo[5,6][1,4]thiazino[3,4-*a*]isoindol-11-one (5c). Yield: 88% as a yellow solid, mp 180–187 °C; FTIR (KBr): ν_{\max} = 3072 (s), 2972 (s), 1705 (s), 1651 (m), 1602 (w), 1566 (w), 1475 (s) cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 9.19 (d, *J* = 9.08 Hz, 1H, H-1), 7.93 (d, *J* = 7.60 Hz, 1H, H-10), 7.33–7.49 (m, 6H, H-8, H-9, H-3', H-4', H-5', H-6'), 7.01–7.08 (m, 2H, H-2, H-4), 6.23 (d, *J* = 8.00 Hz, 1H, H-7), 2.34 (s, 3H, CH₃) ppm; ¹³C NMR

(400 MHz, CDCl₃): δ 165.14 (C-11), 159.49, 137.49, 133.60, 133.48, 132.11, 131.19, 130.78, 130.21, 128.56, 128.10, 127.52, 127.31, 126.79, 124.94, 123.79, 123.12, 121.88, 121.44, 120.34, 116.69, 19.28 (CH₃) ppm; HRMS (ESI⁺) *m/z* calcd for C₂₂H₁₄ClNOS [M + H]⁺ 376.0563, found 376.0528.

3-Bromo-6-(*o*-tolyl)-11*H*-benzo[5,6][1,4]thiazino[3,4-*a*]isoindol-11-one (5d). Yield: 86% as a yellow solid; mp 177–184 °C; FTIR (KBr): ν_{\max} = 3066 (s), 2962 (s), 1699 (s), 1641 (m), 1597 (m), 1573 (w), 1558 (w), 1475 (s) cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 9.12 (d, *J* = 9.08 Hz, 1H, H-1), 7.93 (d, *J* = 7.64 Hz, 1H, H-10), 7.34–7.45 (m, 6H, H-8, H-9, H-3', H-4', H-5', H-6'), 7.01–7.08 (m, 2H, H-2, H-4), 6.23 (d, *J* = 8.00 Hz, 1H, H-7), 2.34 (s, 3H, CH₃) ppm; ¹³C NMR (400 MHz, CDCl₃): δ 165.14 (C-11), 159.53, 136.45, 133.60, 133.50, 132.11, 131.19, 130.47, 130.20, 129.94, 128.56, 128.52, 127.84, 126.75, 124.95, 123.80, 123.12, 122.19, 121.44, 120.06, 116.76, 19.28 (CH₃) ppm; HRMS (ESI⁺) *m/z* calcd for C₂₂H₁₄BrNOS [M + H]⁺ 418.9979, found 418.9942.

6-(2-Methoxyphenyl)-11*H*-benzo[5,6][1,4]thiazino[3,4-*a*]isoindol-11-one (5e). Yield: 79% as a yellow solid, mp 150–156 °C; FTIR (KBr): ν_{\max} = 3103 (s), 2962 (s), 1689 (s), 1654 (m), 1593 (m), 1483 (m) cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 9.19 (d, *J* = 8.44 Hz, 1H, H-1), 7.93 (d, *J* = 7.64 Hz, 1H, H-10), 7.49–7.53 (m, 1H, H-9), 7.36–7.43 (m, 2H, H-3', H-8), 7.22–7.29 (m, 2H, H-4', H-5'), 7.05–7.12 (m, 4H, H-2, H-3, H-4, H-6'), 6.46 (d, *J* = 8.00 Hz, 1H, H-7), 3.78 (s, 3H, OCH₃) ppm; ¹³C NMR (400 MHz, CDCl₃): δ 165.26 (C-11), 157.57, 133.98, 131.62, 131.53, 131.41, 131.07, 128.27, 128.13, 127.53, 125.57, 125.37, 122.95, 121.62, 121.51, 120.29, 119.39, 114.90, 111.92, 55.92 (OCH₃) ppm; HRMS (ESI⁺) *m/z* calcd for C₂₂H₁₅NO₂S [M + H]⁺ 358.0902, found 358.0867.

3-Methoxy-6-(2-methoxyphenyl)-11*H*-benzo[5,6][1,4]thiazino[3,4-*a*]isoindol-11-one (5f). Yield: 85% as a yellow solid, mp 155–158 °C; FTIR (KBr): ν_{\max} = 3060 (s), 2939 (s), 1707 (s), 1595 (m), 1543 (m), 1495 (s) cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 9.16 (d, *J* = 9.28 Hz, 1H, H-1), 7.92 (d, *J* = 6.52 Hz, 1H, H-10), 7.46–7.59 (m, 2H, H-8, H-9), 7.37 (dd, *J* = 7.48 and 1.72 Hz, 1H, H-2), 7.28–7.34 (m, 1H, H-3'), 7.06–7.18 (m, 3H, H-4', H-5', H-6'), 6.79 (dd, *J* = 9.36 and 2.96 Hz, 1H, H-4), 6.44 (d, *J* = 7.96 Hz, 1H, H-7), 3.80 (s, 3H, OCH₃), 3.79 (s, 3H, CH₃) ppm; ¹³C NMR (400 MHz, CDCl₃): δ 164.87 (C-11), 157.58, 156.51, 131.54, 131.49, 131.39, 130.93, 130.12, 128.40, 128.14, 127.35, 123.81, 123.17, 122.78, 121.60, 121.46, 120.58, 114.15, 112.39, 111.86, 110.75, 55.92 (OCH₃), 55.54 (OCH₃) ppm; HRMS (ESI⁺) *m/z* calcd for C₂₃H₁₇NO₃S [M + H]⁺ 388.1007, found 388.0982.

3-Chloro-6-(2-methoxyphenyl)-11*H*-benzo[5,6][1,4]thiazino[3,4-*a*]isoindol-11-one (5g). Yield: 82% as a yellow solid, mp 185–190 °C; FTIR (KBr): ν_{\max} = 3107 (s), 2931 (s), 1693 (s), 1645 (m), 1579 (m), 1479 (m) cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 9.17 (d, *J* = 9.08 Hz, 1H, H-1), 7.94 (d, *J* = 7.68 Hz, 1H, H-10), 7.52–7.56 (m, 1H, H-9), 7.42–7.46 (m, 1H, H-8), 7.37 (dd, *J* = 7.48 and 1.72 Hz, 1H, H-4), 7.27–7.32 (m, 1H, H-3'), 7.19 (dd, *J* = 9.04 and 2.44 Hz, 1H, H-2), 7.07–7.14 (m, 3H, H-4', H-5', H-6'), 6.48 (d, *J* = 8.00 Hz, 1H, H-7), 3.80 (s, 3H, OCH₃) ppm; ¹³C NMR (400 MHz, CDCl₃): δ 165.16 (C-11), 157.55, 132.56, 131.83, 131.71, 131.38, 131.02, 130.32, 128.38, 128.09, 127.51, 127.27, 124.96, 123.01, 122.43, 122.38, 121.65, 121.58, 120.33, 114.06, 111.93, 55.90 (OCH₃) ppm; HRMS (ESI⁺) *m/z* calcd for C₂₂H₁₄ClNO₂S [M + H]⁺ 392.0512, found 392.0485.



3-Bromo-6-(2-methoxyphenyl)-11H-benzo[5,6][1,4]thiazino[3,4-a]isoindol-11-one (5h). Yield: 87% as a yellow solid, mp 160–165 °C; FTIR (KBr): ν_{\max} = 3066 (s), 2987 (s), 1689 (s), 1646 (m), 1575 (m), 1482 (s) cm^{-1} ; $^1\text{H NMR}$ (400 MHz, CDCl_3): δ 9.09 (d, J = 9.04 Hz, 1H, H-1), 7.93 (d, J = 7.76 Hz, 1H, H-10), 7.51–7.55 (m, 1H, H-9), 7.41–7.45 (m, 1H, H-8), 7.26–7.37 (m, 4H, H-3', H-4', H-5', H-6'), 7.07–7.13 (m, 2H, H-2, H-4), 6.47 (d, J = 8.00 Hz, 1H, H-7), 3.79 (s, 3H, OCH_3) ppm; $^{13}\text{C NMR}$ (400 MHz, CDCl_3): δ 165.19 (C-11), 157.56 (C-2'), 133.07, 131.86, 131.73, 131.38, 131.04, 130.23, 128.40, 128.10, 127.77, 127.50, 124.45, 123.03, 122.73, 121.66, 121.60, 120.61, 117.86, 114.15, 111.94, 55.90 (OCH_3) ppm; HRMS (ESI^+) m/z calcd for $\text{C}_{22}\text{H}_{14}\text{BrNO}_2\text{S} [\text{M} + \text{H}]^+$ 434.9929, found 434.9899.

6-(2-Fluorophenyl)-11H-benzo[5,6][1,4]thiazino[3,4-a]isoindol-11-one (5i). Yield: 75% as a yellow solid, mp 220–224 °C; FTIR (KBr): ν_{\max} = 3061 (s), 1687 (s), 1649 (m), 1608 (m), 1573 (m), 1523 (w), 1479 (m) cm^{-1} ; $^1\text{H NMR}$ (400 MHz, CDCl_3): δ 9.06 (d, J = 9.04 Hz, 1H, H-1), 7.96 (d, J = 7.64 Hz, 1H, H-10), 7.44–7.50 (m, 2H, H-8, H-9), 7.24–7.36 (m, 3H, H-4', H-5', H-6'), 7.07–7.10 (m, 4H, H-2, H-4, H-3, H-2'), 6.49 (d, J = 8.00 Hz, 1H, H-7) ppm; $^{13}\text{C NMR}$ (100 MHz, CDCl_3): δ 165.21 (C-11), 160.16 (d, $J_{\text{C-F}}$ = 250.24 Hz), 133.81, 132.91, 132.26 (d, $J_{\text{C-F}}$ = 8.07 Hz), 131.89, 131.81 (d, $J_{\text{C-F}}$ = 1.83 Hz), 131.77, 130.71, 130.55, 128.90, 128.63, 127.82, 125.47 (d, $J_{\text{C-F}}$ = 1.83 Hz), 123.20, 122.12 (d, $J_{\text{C-F}}$ = 16.06 Hz), 121.26, 120.65, 119.52 (d, $J_{\text{C-F}}$ = 17.44 Hz), 118.09, 116.94 ppm; HRMS (ESI^+) m/z calcd for $\text{C}_{21}\text{H}_{12}\text{FNOS} [\text{M} + \text{H}]^+$ 346.0702, found 346.0717.

6-(2-Fluorophenyl)-3-methyl-11H-benzo[5,6][1,4]thiazino[3,4-a]isoindol-11-one (5j). Yield: 85% as a yellow solid, mp 240–244 °C; FTIR (KBr): ν_{\max} = 3059 (s), 1683 (s), 1649 (m), 1608 (m), 1576 (m), 1489 (s) cm^{-1} ; $^1\text{H NMR}$ (400 MHz, CDCl_3): δ 9.06 (d, J = 8.60 Hz, 1H, H-1), 7.95 (d, J = 7.64 Hz, 1H, H-10), 7.43–7.58 (m, 4H, H-8, H-9, H-3', H-4'), 7.25–7.36 (m, 4H, H-2, H-4, H-5', H-6'), 6.48 (d, J = 7.96 Hz, 1H, H-7), 2.30 (s, 3H, CH_3) ppm; $^{13}\text{C NMR}$ (100 MHz, CDCl_3): δ 165.04 (C-11), 160.20 (d, $J_{\text{C-F}}$ = 249.35 Hz), 135.46, 131.99 (d, $J_{\text{C-F}}$ = 7.91 Hz), 131.87 (d, $J_{\text{C-F}}$ = 1.76 Hz), 131.71, 131.34, 130.71, 128.89, 128.56, 128.43, 127.61, 125.84, 125.35 (d, $J_{\text{C-F}}$ = 3.82 Hz), 125.03, 123.10, 121.29 (d, $J_{\text{C-F}}$ = 11.62 Hz), 120.39, 119.28, 116.81 (d, $J_{\text{C-F}}$ = 21.27 Hz), 111.11, 20.64 (CH_3) ppm; HRMS (ESI^+) m/z calcd for $\text{C}_{22}\text{H}_{14}\text{FNOS} [\text{M} + \text{H}]^+$ 360.0858, found 360.0841.

6-(2-Fluorophenyl)-3-methoxy-11H-benzo[5,6][1,4]thiazino[3,4-a]isoindol-11-one (5k). Yield: 76% as a yellow solid, mp 266–270 °C; FTIR (KBr): ν_{\max} = 3060 (s), 1690 (s), 1649 (m), 1604 (m), 1568 (m), 1496 (s) cm^{-1} ; $^1\text{H NMR}$ (400 MHz, CDCl_3): δ 9.14 (d, J = 9.32 Hz, 1H, H-1), 7.94 (d, J = 7.64 Hz, 1H, H-10), 7.53–7.59 (m, 1H, H-9), 7.43–7.50 (m, 2H, H-8, H-3'), 7.25–7.35 (m, 3H, H-4', H-5', H-6'), 6.80 (dd, J = 9.28 and 2.92 Hz, 1H, H-4), 6.64 (d, J = 2.88 Hz, 1H, H-2), 6.48 (d, J = 8.60 Hz, 1H, H-7), 3.81 (s, 3H, OCH_3) ppm; $^{13}\text{C NMR}$ (100 MHz, CDCl_3): δ 164.84 (C-11), 160.23, 156.70, 132.04 (d, $J_{\text{C-F}}$ = 7.98 Hz), 131.91 (d, $J_{\text{C-F}}$ = 1.79 Hz), 131.63, 130.65, 128.62, 128.58, 128.49, 127.20, 125.36 (d, $J_{\text{C-F}}$ = 3.76 Hz), 123.03, 122.11 (d, $J_{\text{C-F}}$ = 15.91 Hz), 121.23, 120.88, 120.64, 116.82 (d, $J_{\text{C-F}}$ = 21.19 Hz), 112.63, 110.86, 110.37, 55.55 (OCH_3) ppm; HRMS (ESI^+) m/z calcd for $\text{C}_{22}\text{H}_{14}\text{FNO}_2\text{S} [\text{M} + \text{H}]^+$ 376.0808, found $[\text{M} + \text{H}]^+$ 376.0802.

3-Chloro-6-(2-fluorophenyl)-11H-benzo[5,6][1,4]thiazino[3,4-a]isoindol-11-one (5l). Yield: 87% as a yellow solid, mp 274–280 °C; FTIR (KBr): ν_{\max} = 3062 (s), 1689 (s), 1646 (m), 1568 (m), 1483 (s) cm^{-1} ; $^1\text{H NMR}$ (400 MHz, CDCl_3): δ 9.13 (d, J = 9.04 Hz, 1H, H-1), 7.95 (d, J = 7.64 Hz, 1H, H-10), 7.54–7.59 (m, 1H, H-9), 7.45–7.49 (m, 2H, H-8, H-3'), 7.19–7.36 (m, 4H, H-2, H-4', H-5', H-6'), 7.07 (d, J = 2.36 Hz, 1H, H-4), 6.49 (d, J = 8.00 Hz, 1H, H-7) ppm; $^{13}\text{C NMR}$ (100 MHz, CDCl_3): δ 165.13 (C-11), 160.17 (d, $J_{\text{C-F}}$ = 249.44 Hz), 132.43, 132.25 (d, $J_{\text{C-F}}$ = 7.96 Hz), 132.11, 131.78 (d, $J_{\text{C-F}}$ = 1.73 Hz), 130.70, 130.60, 128.90, 128.43, 128.25, 127.61, 125.46 (d, $J_{\text{C-F}}$ = 3.77 Hz), 125.03, 123.29, 121.72 (d, $J_{\text{C-F}}$ = 13.55 Hz), 121.35, 120.39, 118.17, 116.90 (d, $J_{\text{C-F}}$ = 21.44 Hz), 110.30 ppm; HRMS (ESI^+) m/z calcd for $\text{C}_{21}\text{H}_{11}\text{ClFNOS} [\text{M} + \text{H}]^+$ 380.0312, found 380.0305.

3-Bromo-6-(2-fluorophenyl)-11H-benzo[5,6][1,4]thiazino[3,4-a]isoindol-11-one (5m). Yield: 83% as a yellow solid, mp 254–260 °C; FTIR (KBr): ν_{\max} = 3061 (s), 1701 (s), 1660 (m), 1566 (m), 1487 (m), 1473 (m), 1456 (m) cm^{-1} ; $^1\text{H NMR}$ (400 MHz, CDCl_3): δ 9.07 (d, J = 9.08 Hz, 1H, H-1), 7.96 (d, J = 7.68 Hz, 1H, H-10), 7.55–7.60 (m, 1H, H-9), 7.45–7.49 (m, 2H, H-8, H-3'), 7.28–7.37 (m, 4H, H-2, H-4', H-5', H-6'), 7.22 (d, J = 2.32 Hz, 1H, H-4), 6.50 (d, J = 8.64 Hz, 1H, H-7) ppm; $^{13}\text{C NMR}$ (100 MHz, CDCl_3): δ 165.15 (C-11), 160.16 (d, $J_{\text{C-F}}$ = 249.55 Hz), 132.93, 132.25 (d, $J_{\text{C-F}}$ = 8.01 Hz), 132.13, 131.77 (d, $J_{\text{C-F}}$ = 1.83 Hz), 130.71, 130.57, 128.90, 128.41, 128.26, 127.82, 125.47 (d, $J_{\text{C-F}}$ = 3.71 Hz), 123.30, 121.97, 121.70 (d, $J_{\text{C-F}}$ = 15.93 Hz), 121.35, 120.66, 118.10, 116.91 (d, $J_{\text{C-F}}$ = 21.08 Hz), 110.37 ppm; HRMS (ESI^+) m/z calcd for $\text{C}_{21}\text{H}_{11}\text{BrFNOS} [\text{M} + \text{H}]^+$ 423.9807, found 423.9814.

6-(3-Fluorophenyl)-11H-benzo[5,6][1,4]thiazino[3,4-a]isoindol-11-one (5n). Yield: 78% as a yellow solid, mp 185–190 °C; FTIR (KBr): ν_{\max} = 3062 (s), 1693 (s), 1643 (m), 1606 (m), 1585 (m), 1481 (s) cm^{-1} ; $^1\text{H NMR}$ (400 MHz, CDCl_3): δ 9.17 (d, J = 8.80 Hz, 1H, H-1), 7.96 (d, J = 7.72 Hz, 1H, H-10), 7.44–7.56 (m, 2H, H-8, H-9), 7.23–7.31 (m, 3H, H-4', H-5', H-6'), 6.99–7.11 (m, 4H, H-2, H-3, H-4, H-2'), 6.55 (d, J = 7.96 Hz, 1H, H-7) ppm; $^{13}\text{C NMR}$ (100 MHz, CDCl_3): δ 165.23 (C-11), 163.30 (d, $J_{\text{C-F}}$ = 247.69 Hz), 136.64 (d, $J_{\text{C-F}}$ = 8.18 Hz), 133.80, 132.03, 131.95, 131.70, 131.45 (d, $J_{\text{C-F}}$ = 8.41 Hz), 128.92 (d, $J_{\text{C-F}}$ = 8.07 Hz), 128.43, 127.89, 125.42 (d, $J_{\text{C-F}}$ = 3.07 Hz), 123.22, 121.78, 119.44, 117.89, 117.65, 117.14, 116.80 (d, $J_{\text{C-F}}$ = 21.92 Hz), 116.00, 115.79 ppm; HRMS (ESI^+) m/z calcd for $\text{C}_{21}\text{H}_{12}\text{FNOS} [\text{M} + \text{H}]^+$ 346.0702, found 346.0673.

6-(3-Fluorophenyl)-3-methyl-11H-benzo[5,6][1,4]thiazino[3,4-a]isoindol-11-one (5o). Yield: 85% as a yellow solid, mp 220–224 °C; FTIR (KBr): ν_{\max} = 3062 (s), 1681 (s), 1645 (m), 1606 (m), 1583 (m), 1490 (s) cm^{-1} ; $^1\text{H NMR}$ (400 MHz, CDCl_3): δ 9.04 (d, J = 8.60 Hz, 1H, H-1), 7.93 (d, J = 7.64 Hz, 1H, H-10), 7.42–7.54 (m, 2H, H-8, H-9), 7.21–7.31 (m, 4H, H-2', H-4', H-5', H-6'), 7.06 (dd, J = 8.60 and 1.48 Hz, 1H, H-2), 6.90 (d, J = 1.56 Hz, 1H, H-4), 6.52 (d, J = 8.0 Hz, 1H, H-7) ppm; $^{13}\text{C NMR}$ (100 MHz, CDCl_3): δ = 165.03 (C-11), 163.27 (d, $J_{\text{C-F}}$ = 247.57 Hz), 136.74 (d, $J_{\text{C-F}}$ = 8.05 Hz), 135.45, 131.51, 131.39 (d, $J_{\text{C-F}}$ = 8.40 Hz), 131.29, 130.54, 128.50 (d, $J_{\text{C-F}}$ = 2.92 Hz), 127.15, 125.77, 125.44 (d, $J_{\text{C-F}}$ = 3.03 Hz), 123.10, 121.74, 119.28, 117.06, 116.93, 116.78 (d, $J_{\text{C-F}}$ = 14.11 Hz), 116.60, 116.58, 20.66 ppm; HRMS (ESI^+) m/z calcd for $\text{C}_{22}\text{H}_{14}\text{FNOS} [\text{M} + \text{H}]^+$ 360.0858, found 360.0890.



6-(3-Fluorophenyl)-3-methoxy-11H-benzo[5,6][1,4]thiazino[3,4-a]isoindol-11-one (**5p**). Yield: 90% as a yellow solid, mp 210–212 °C; FTIR (KBr): ν_{\max} = 3066 (m), 2964 (m), 1701 (s), 1637 (m), 1579 (m), 1554 (w), 1539 (w), 1475 (s) cm^{-1} ; $^1\text{H NMR}$ (400 MHz, CDCl_3): δ 9.13 (d, J = 9.40 Hz, 1H, H-1), 7.92 (d, J = 7.48 Hz, 1H, H-10), 7.51–7.54 (m, 2H, H-8, H-9), 7.22–7.30 (m, 4H, H-2', H-4', H-5', H-6'), 6.78 (dd, J = 9.04 and 2.68 Hz, 1H, H-2), 6.61 (d, J = 2.72 Hz, 1H, H-4), 6.52 (d, J = 7.96 Hz, 1H, H-7), 3.79 (s, 3H, OCH_3) ppm; $^{13}\text{C NMR}$ (100 MHz, CDCl_3): δ 164.83 (C-11), 163.27 (d, $J_{\text{C-F}}$ = 247.82 Hz), 156.63, 136.60 (d, $J_{\text{C-F}}$ = 7.95 Hz), 133.79, 132.92, 131.43, 131.33 (d, $J_{\text{C-F}}$ = 7.71 Hz), 130.47, 128.58, 127.11, 125.50 (d, $J_{\text{C-F}}$ = 3.05 Hz), 123.02, 121.73, 120.64, 117.25, 117.12, 116.97, 116.83 (d, $J_{\text{C-F}}$ = 15.68 Hz), 115.89, 55.55 (OCH_3) ppm; HRMS (ESI^+) m/z calcd for $\text{C}_{22}\text{H}_{14}\text{FNO}_2\text{S}$ [$\text{M} + \text{H}$] $^+$ 376.0808, found 376.0835.

3-Chloro-6-(3-fluorophenyl)-11H-benzo[5,6][1,4]thiazino[3,4-a]isoindol-11-one (**5q**). Yield: 82% as a yellow solid, mp 232–238 °C; FTIR (KBr): ν_{\max} = 3064 (s), 1685 (s), 1647 (m), 1583 (m), 1490 (s) cm^{-1} ; $^1\text{H NMR}$ (400 MHz, CDCl_3): δ 9.12 (d, J = 9.08 Hz, 1H, H-1), 7.93 (d, J = 7.68 Hz, 1H, H-10), 7.44–7.55 (m, 2H, H-8, H-9), 7.17–7.33 (m, 5H, H-2, H-2', H-4', H-5', H-6'), 7.05 (d, J = 2.40 Hz, 1H, H-4), 6.53 (d, J = 8.00 Hz, 1H, H-7) ppm; $^{13}\text{C NMR}$ (100 MHz, CDCl_3): δ 165.11 (C-11), 163.30 (d, $J_{\text{C-F}}$ = 247.92 Hz), 136.17 (d, $J_{\text{C-F}}$ = 8.06 Hz), 132.37, 131.90, 131.54 (d, $J_{\text{C-F}}$ = 8.39 Hz), 130.54 (d, $J_{\text{C-F}}$ = 3.17 Hz), 128.84, 128.24, 127.65, 127.18, 125.40 (d, $J_{\text{C-F}}$ = 3.09 Hz), 124.95, 123.27, 121.84, 121.63, 120.38, 117.32, 117.11, 116.79 (d, $J_{\text{C-F}}$ = 21.96 Hz), 115.73 ppm; HRMS (ESI^+) m/z calcd for $\text{C}_{21}\text{H}_{11}\text{ClFNO}_2\text{S}$ [$\text{M} + \text{H}$] $^+$ 380.0312, found 380.0336.

3-Bromo-6-(3-fluorophenyl)-11H-benzo[5,6][1,4]thiazino[3,4-a]isoindol-11-one (**5r**). Yield: 76% as a yellow solid, mp 210–213 °C; FTIR (KBr): ν_{\max} = 3062 (s), 2934 (s), 1691 (s), 1647 (m), 1579 (m), 1479 (s) cm^{-1} ; $^1\text{H NMR}$ (400 MHz, CDCl_3): δ 9.06 (d, J = 9.08 Hz, 1H, H-1), 7.93 (d, J = 6.72 Hz, 1H, H-10), 7.44–7.56 (m, 2H, H-8, H-9), 7.18–7.34 (m, 6H, H-2, H-2', H-4', H-5', H-6', H-4), 6.53 (d, J = 7.96 Hz, 1H, H-7) ppm; $^{13}\text{C NMR}$ (100 MHz, CDCl_3): δ 165.10 (C-11), 163.29 (d, $J_{\text{C-F}}$ = 247.98 Hz), 136.14 (d, $J_{\text{C-F}}$ = 8.01 Hz), 135.94, 132.85, 131.91, 131.54 (d, $J_{\text{C-F}}$ = 8.37 Hz), 130.59, 130.51, 128.83, 128.22, 127.72, 125.39 (d, $J_{\text{C-F}}$ = 3.08 Hz), 123.26, 121.88 (d, $J_{\text{C-F}}$ = 7.91 Hz), 120.62, 118.05, 117.32, 117.11, 116.78 (d, $J_{\text{C-F}}$ = 21.93 Hz), 115.80 ppm; HRMS (ESI^+) m/z calcd for $\text{C}_{21}\text{H}_{11}\text{BrFNO}_2\text{S}$ [$\text{M} + \text{H}$] $^+$ 423.9807, found 423.9838.

Conflicts of interest

The authors declare that there are no conflicts of interest in this work.

Acknowledgements

The authors are grateful to the Council of Scientific and Industrial Research, New Delhi, India for providing financial support (CSIR No. 09/752(0060)/2016-EMR-1).

References

- P. Li, Q. Zhang, A. J. Robichaud, T. Lee, J. Tomesch, W. Yao and J. P. Hendrick, *J. Med. Chem.*, 2014, **57**(6), 2670–2682.
- A. Guleria, D. K. Jangid, N. Gautam, R. Lakhota, A. Chowdhary and D. C. Gautam, *Comb. Chem. High Throughput Screening*, 2015, **18**(1), 48–52.
- A. Barazarte, G. Lobo, N. Gamboa, J. R. Rodrigues, M. V. Capparelli, A. Alvarez-Larena and J. E. Charris, *Eur. J. Med. Chem.*, 2009, **44**(3), 1303–1310.
- G. Grandolini, L. Perioli and V. Ambrogi, *Eur. J. Med. Chem.*, 1999, **34**(9), 701–709.
- H. Matsuoka, N. Ohi, M. Mihara, H. Suzuki, K. Miyamoto, N. Maruyama and K. Yano, *J. Med. Chem.*, 1997, **40**(1), 105–111.
- M. Jeleń, K. Pluta, M. Zimecki, B. Morak-Młodawska, J. Artym and M. Kocięba, *Eur. J. Med. Chem.*, 2015, **89**, 411–420.
- J. Gowda, A. M. A. Khader, B. Kalluraya, P. Shree and A. R. Shabaraya, *Eur. J. Med. Chem.*, 2011, **46**(9), 4100–4106.
- H. Tawada, Y. Sugiyama, H. Ikeda, Y. Yamamoto and K. Meguro, *Chem. Pharm. Bull.*, 1990, **38**(5), 1238–1245.
- J. Zhang, C. Ming, W. Zhang, P. N. Okechukwu, B. Morak-Młodawska, K. Pluta and K. K. Ooi, *Drug Des., Dev. Ther.*, 2017, **11**, 3045–3063.
- M. Fujita, S. Ito, A. Ota, N. Kato, K. Yamamoto, Y. Kawashima and J. Iwao, *J. Med. Chem.*, 1990, **33**(7), 1898–1905.
- S. Mor, S. Nagoria, S. Sindhu and V. Singh, *Chem. Biol. Interface*, 2017, **7**(1), 1–18.
- S. Mor, P. Pahal and B. Narasimhan, *Eur. J. Med. Chem.*, 2012, **53**, 176–189.
- S. Mor and S. Nagoria, *Synth. Commun.*, 2016, **46**(2), 169–178.
- X. Huang, N. Rong, P. Li, G. Shen, Q. Li, N. Xin and C. Zhao, *Org. Lett.*, 2018, **20**, 3332–3336.
- A. Napolitano, L. Panzella, L. Leone and M. d'Ischia, *Acc. Chem. Res.*, 2012, **46**(2), 519–528.
- O. Unger, *Ber. Dtsch. Chem. Ges.*, 1897, **30**(1), 607–610.
- A. Jaafar, A. Khalaf, F. Farès, D. Grée, H. Abdallah, T. Roisne and A. Hachem, *Mediterr. J. Chem.*, 2014, **3**(2), 831–837.
- N. Gautam, A. Garg and D. C. Gautam, *Nucleosides, Nucleotides Nucleic Acids*, 2015, **34**(1), 40–55.
- M. Saadouni, T. Ghailane, S. Boukhris, A. Hassikou, N. Habbadi, R. Ghailane and H. Amri, *Org. Commun.*, 2014, **7**(2), 77–84.
- O. Ponomarov, Z. Padělková and J. Hanusek, *J. Heterocycl. Chem.*, 2011, **48**(6), 1225–1228.
- S. Mitra, A. Chakraborty, S. Mishra, A. Majee and A. Hajra, *Org. Lett.*, 2014, **16**(21), 5652–5655.
- W. Hu and S. Zhang, *J. Org. Chem.*, 2015, **80**(12), 6128–6132.
- (a) V. Fajardo, V. Elango, B. K. Cassels and M. Shamma, *Tetrahedron Lett.*, 1982, **23**, 39–42; (b) E. Valencia, J. Freyer, M. Shamma and V. Fajardo, *Tetrahedron Lett.*, 1984, **25**, 599–602; (c) E. Valencia, V. Fajardo, A. J. Freyer and M. Shamma, *Tetrahedron Lett.*, 1985, **26**, 993–996.
- V. Rys, A. Couture, E. Deniau and P. Grandclaudeon, *Tetrahedron*, 2003, **59**, 6615–6619.
- M. H. Abu Zarga, S. S. Sabri, S. Firdous and M. Shamma, *Phytochemistry*, 1987, **26**(4), 1233–1234.
- (a) V. Fajardo, V. Elango, B. K. Cassels and M. Shamma, *Tetrahedron Lett.*, 1982, **23**, 39–42; (b) F. G. Fang and S. J. Danishefsky, *Tetrahedron Lett.*, 1989, **30**, 2747–2750.



- 27 E. Valencia, V. Fajardo, S. Firdous, A. J. Freyer and M. Shamma, *Tetrahedron Lett.*, 1985, **26**, 993–996.
- 28 T. Wada and N. Fukuda, *Psychopharmacology*, 1991, **103**, 314–322.
- 29 Z.-L. Chang and D.-Y. Zhu, in *The Alkaloids*, ed. Arnold Brossi, Academic Press, New York, 1st edn, 1978, vol. 31, pp. 29–65.
- 30 N. Kanamitsu, T. Osaki, Y. Itsuji, M. Yoshimura, H. Tsujimoto and M. Soga, *Chem. Pharm. Bull.*, 2007, **55**(12), 1682–1688.
- 31 M. Linden, D. Hadler and S. Hofmann, *Hum. Psychopharmacol.*, 1997, **12**(5), 445–452.
- 32 T. Lübbers, P. Angehrn, H. Gmünder and S. Herzig, *Bioorg. Med. Chem. Lett.*, 2007, **17**(16), 4708–4714.
- 33 A. F. Watson, J. Liu, K. Bennaceur, C. J. Drummond, J. A. Endicott, B. T. Golding, R. J. Griffin, K. Haggerty, X. Lu, J. M. McDonnell, D. R. Newell, M. E. M. Noble, C. H. Revill, C. Riedinger, Q. Xu, Y. Zhao, J. Lunec and I. R. Hardcastle, *Bioorg. Med. Chem. Lett.*, 2011, **21**, 5916–5919.
- 34 J. G. Topliss, L. M. Konzelman, N. Sperber and F. E. Roth, *J. Med. Chem.*, 1964, **7**(4), 453–456.
- 35 E. C. Taylor, P. Zhou, L. D. Jennings, Z. Mao, B. Hu and J. G. Jun, *Tetrahedron Lett.*, 1997, **38**, 521–524.
- 36 J. A. Al-Qaisi, T. M. Alhussainy, N. A. Qinna, K. Z. Matalka, E. N. Al-Kaissi and Z. A. Muhi-Eldeen, *Arabian J. Chem.*, 2014, **7**(6), 1024–1030.
- 37 K. Achinami, N. Ashizawa and F. Kobayasui, *Jpn. Pat.*, JP03, 1991, vol. 133, p. 955; *Chem. Abstr.*, 1991, **115**, 255977J.
- 38 I. Pendrak, S. Barney, R. Wittrock, D. M. Lambert and W. D. Kingsbury, *J. Org. Chem.*, 1994, **59**, 2623–2625.
- 39 Y. Ishihara, Y. Kiyota and G. Goto, *Chem. Pharm. Bull.*, 1990, **38**(11), 3024–3030.
- 40 S. Sharma, E. Park, J. Park and I. S. Kim, *Org. Lett.*, 2012, **14**(3), 906–909.
- 41 H. Cao, L. McNamee and H. Alper, *Org. Lett.*, 2008, **10**, 5281–5284.
- 42 L. Shi, L. Hu, J. Wang, X. Cao and H. Gu, *Org. Lett.*, 2012, **14**, 1876–1879.
- 43 R. W. Foster, C. J. Tame, H. C. Hailes and T. D. Sheppard, *Adv. Synth. Catal.*, 2013, **355**, 2353–2360.
- 44 L. Zhang, Y. Zhang, X. Wang and J. Shen, *Molecules*, 2013, **18**, 654–665.
- 45 S. Das, D. Addis, L. R. Knopke, U. Bentrup, K. Junge, A. Brukner and M. Beller, *Angew. Chem.*, 2011, **123**, 9346–9350.
- 46 Y. Zhou, Y. Zhai, J. Li, D. Ye, H. Jiang and H. Liu, *Green Chem.*, 2010, **12**, 1397–1404.
- 47 (a) H. Watanabe, C. L. Mao, I. T. Barnish and C. R. Hauser, *J. Org. Chem.*, 1969, **34**, 919–926; (b) W. E. Parham and L. D. Jones, *J. Org. Chem.*, 1976, **41**, 1187–1191.
- 48 V. More, R. Rohlmann, O. G. Mancheno, C. Petronzi, L. Palombi, V. Intintoli and A. Massa, *RSC Adv.*, 2012, **2**, 3592–3595.
- 49 Y. Zhou, Y. Zhai, J. Li, D. Ye, H. Jiang and H. Liu, *Green Chem.*, 2010, **12**(8), 1397–1404.
- 50 R. Mohil, D. Kumar and S. Mor, *J. Heterocycl. Chem.*, 2014, **51**, 203–211.
- 51 S. L. Shapiro, K. Geiger, J. Youlus and L. Freedman, *J. Org. Chem.*, 1961, **26**, 3580–3582.
- 52 N. E. Cundasawmy and D. B. Maclean, *Can. J. Chem.*, 1972, **50**, 3028–3036.
- 53 R. Q. Brewster and F. B. Dains, *J. Am. Chem. Soc.*, 1936, **58**, 1364–1366.
- 54 R. L. Mital and S. K. Jain, *J. Chem. Soc. C*, 1969, **16**, 2148–2150.
- 55 M. Matsui, Y. Marui, M. Kushida, K. Funabiki, H. Muramastu, K. Shibata, K. Hirota, M. Hosoda and K. Tai, *Dyes Pigm.*, 1998, **38**, 57–64.

