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1. Introduction

1,2,3-Triazole, a five-membered heterocyclic ring system, is a very well-known biologically active pharmacophore constructed by the copper-catalyzed azide–alkyne cycloaddition (CuAAC) reaction, which is popular as a click chemistry reaction.¹ Over the last decade, 1,2,3-triazole has become one of the key structural motifs and is used in numerous fields including polymer chemistry,² material science,³ and drug discovery.⁴ 1,2,3-triazole-based molecules display various biological activities such as anti-fungal,^{5,6} anti-bacterial,⁶ anti-tubercular,⁷ antiinflammatory,⁸ anti-allergic,⁹ anti-HIV,¹⁰ anti-cancer,¹¹ and antiphytopathogenic.¹² Some marketed drugs have the 1,2,3-triazole unit in their structure, and these include Cefatrizine (a broadspectrum antibiotic), Tazobactam (an antibiotic), and Carboxyamidotriazole (CAI) (a calcium channel blocker) (Fig. 1).

Azole drugs have broad-spectrum activities against most yeasts and filamentous fungi and are mostly used in antifungal

New *N*-phenylacetamide-incorporated 1,2,3triazoles: [Et₃NH][OAc]-mediated efficient synthesis and biological evaluation[†]

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A facile, highly efficient, and greener method for the synthesis of new 1,4-disubstituted-1,2,3-triazoles was conducted using [Et₃NH][OAc] as a medium by the implementation of ultrasound irradiation *via* click chemistry, affording excellent yields. The present synthetic method exhibited numerous advantages such as mild reaction conditions, excellent product yields, minimal chemical waste, operational simplicity, shorter reaction time, and a wide range of substrate scope. The synthesized compounds were further evaluated for *in vitro* antifungal activity against five fungal strains, and some of the compounds displayed equivalent or greater potency than the standard drug. A molecular docking study against the modelled three-dimensional structure of cytochrome P450 lanosterol 14*α*-demethylase was also performed to understand the binding affinity and binding interactions of the enzyme. Furthermore, the synthesized compounds were evaluated for DPPH radical scavenging activity and antitubercular activity against *Mycobacterium tuberculosis* H37Rv strain.

chemotherapy.13 Some well-known antifungal agents including fluconazole, voriconazole, ravuconazole, and itraconazole contain a 1,2,4-triazole ring in their structure, as shown in Fig. 2. However, their clinical uses have been restricted by their relatively high risk of toxicity, pharmacokinetic deficiencies, emergence of drug resistance, and undesirable side effects. These antifungal drugs inhibit CYP51, a key enzyme in the biosynthesis of ergosterol, through a mechanism in which the antifungal agent having a triazole scaffold is positioned perpendicular to the porphyrin plane with a ring nitrogen atom (N-4 of triazole) coordinated with a heme iron atom.¹⁴ Over a couple of decades, the incidence of systemic fungal infections has increased due to cancer chemotherapy, organ transplantation, tuberculosis, and immunodeficiency virus (HIV) infection.15 However, the extensive use of antifungal drugs has led to an increase in the resistance to these drugs.¹⁶ Hence, there is an urgent need for developing new antifungal agents with effective activities, low toxicity, and minimum side effects.

Ionic liquids (ILs) are environment-friendly solvents because of their interesting properties and they can be used as alternatives to harmful organic solvents. Furthermore, they are useful in catalytic reactions¹⁷ and organic synthesis¹⁸ because of their



Fig. 1 Structures of drugs containing the 1,2,3-triazole unit.

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Fig. 2 1,2,4-Triazole-based marketed antifungal drugs.

unique properties, making them superior media for increasing selectivity, reactivity, and recyclability. Various ionic liquids have been significantly used in heterocyclic synthesis as solvents or catalysts.¹⁹ The copper-catalyzed 1,3-dipolar cycloaddition of organic azides and terminal alkynes has been reported by using ionic liquids such as [bmim][BF₄],^{20a,b} [Bmim]OH,^{20c} (SNIL-Cu(π)),^{20d} ([Hmim]TFA),^{20e} and [C₈dabco][N(CN)₂],^{20f} and DBU-based ionic liquids.^{20g} The ionic liquid [Et₃NH][OAc] has been used extensively in various organic transformations.²¹

Considering the importance of ionic liquids in organic synthesis and in continuation of our work^{7d,22} as well as for the development of new synthetic methodologies using ionic liquids,²³ herein, we reported the efficient synthesis of new 1,4-disubstituted-1,2,3-triazoles *via* the click chemistry approach using [Et₃NH][OAc] as the medium under ultrasonic irradiation. Furthermore, the newly synthesized compounds were investigated for their antifungal, antioxidant, and antitubercular activities.

2. Results and discussion

2.1 Chemistry

There are several reports on the synthesis of 1,4-disubstituted-1,2,3-triazoles bearing amide functionality and displaying various biological activities;²⁴ recently, Ferroni *et al.* developed triazoles as non-steroidal anti-androgens for prostate cancer treatment.²⁵ On the basis of these findings, we designed small



Scheme 1 Synthesis of alkynes 2a-c and 2-azido-N-phenyl-acetamides 5a-g. Reagents and conditions: (a) propargyl bromide, K_2CO_3 , DMF, 2 h, 90–95%; (b) chloroacetyl chloride, NEt₃, CH₂Cl₂, 0 °C to rt, 3–5 h, 85–95%; (c) NaN₃, toluene, reflux, 5–7 h, 88–96%.

1,2,3-triazoles with amide linkage in their structures. Initially, the starting materials, *i.e.*, (prop-2-yn-1-yloxy)benzenes **2a–c** were prepared by a previously reported method.^{7d} 2-Azido-*N*-phenylacetamides **5a–g** were synthesized from their corresponding anilines *via* chloroacetylation using chloroacetyl chloride, followed by nucleophilic substitution with sodium azide in excellent yields (Scheme 1).

After the synthesis of alkynes **2a–c** and azides **5a–g**, we considered the model substrates (prop-2-yn-1-yloxy)benzenes **2a** (1.0 mmol) and 2-azido-*N*-phenylacetamide **5a** (1.0 mmol) for the click reaction (Scheme 2).

The literature survey reveals the use of t-BuOH-H₂O, THF-H₂O, and DMF-H₂O as solvents in combination with copper salts for the click reaction. Accordingly, we performed the model reactions of 2a and 5a by using $CuSO_4 \cdot 5H_2O_4$. $Cu(OAc)_2 \cdot H_2O$, CuI, and CuCl with or without sodium ascorbate as a reducing agent in solvents such as t-BuOH and THF (Table 1). In our initial attempt, the model reaction was carried out by using the conventional method, *i.e.*, the reaction mixture was stirred at room temperature for 10 h using various copper salts in t-BuOH or THF as a solvent (Table 1, entries 2-7). The product 2-(4-(phenoxymethyl)-1H-1,2,3-triazol-1-yl)-N-phenylacetamide 6a was obtained in 28-72% yield. Furthermore, the model reaction was carried out using copper salts and t-BuOH/ THF as a solvent under ultrasound irradiation (40 kHz, 30 °C) for 7 h, which resulted in the product 6a in 32-83% yield (Table 1, entries 2-7). These results indicate a longer reaction time with a lower yield of the product 6a.

Ionic liquids have unique properties as solvents and as media; herein, we attempted to use various ionic liquids for the synthesis of new amide-linked 1,4-disubstituted-1,2,3-triazole 6a via the click chemistry approach. Freshly synthesized appropriate ionic liquids such as [Et₃NH][OAc], [Et₃NH][Cl], [Et₃NH][HSO₄], [HDBU][OAc], and [HDBU][HSO₄] in combination with copper salts such as $CuSO_4 \cdot 5H_2O$ and $Cu(OAc)_2 \cdot H_2O$ were used along with sodium ascorbate as a reducing agent to carry out the model reaction (Table 1, entries 9-16) at room temperature for 7 h. The maximum yield (90%) of the product **6a** was obtained by using $Cu(OAc)_2 \cdot H_2O$ as the catalyst and [Et₃NH][OAc] as the solvent (Table 1, entries 9 and 10). In our further attempt, the model reaction was carried out under ultrasound irradiation for appropriate times (35 and 45 min) (Table 1, entries 9-16). The product 6a was obtained in 95% yield under ultrasound irradiation using Cu(OAc)₂·H₂O as the catalyst and [Et3NH][OAc] as the solvent (Table 1, entries 9 and 10). In short, it was observed from the optimization of the best reaction conditions that the Cu(OAc)2·H2O-catalyzed click reaction in [Et₃NH][OAc] ionic liquid under ultrasound



Scheme 2 Standard model reaction.

Table 1 Optimization of the reaction conditions for the synthesis of 6a



Entry		Solvent/medium	Conventional method		US method	
	Copper salt		Time ^a	$\operatorname{Yield}^{b}(\%)$	Time ^a	$\operatorname{Yield}^{b}(\%)$
1	_	<i>t</i> -BuOH : H ₂ O	10 h	0	6 h	0
2	$CuSO_4 \cdot 5H_2O$	t -BuOH : H_2O	10 h	28	6 h	32
3	$CuSO_4 \cdot 5H_2O + Na ascorbate$	t-BuOH : H ₂ O	10 h	70	6 h	78
4	$Cu(OAc)_2 \cdot H_2O$	t-BuOH : H ₂ O	10 h	72	6 h	82
5	$Cu(OAc)_2 \cdot H_2O + Na ascorbate$	t-BuOH : H ₂ O	10 h	72	6 h	83
6	CuI	THF	10 h	70	6 h	80
7	CuCl	THF	10 h	60	6 h	65
8	_	[Et ₃ NH][OAc]	10 h	0	6 h	0
9	$Cu(OAc)_2 \cdot H_2O$	[Et ₃ NH][OAc]	7 h	90	35 min	95
10	$Cu(OAc)_2 \cdot H_2O + Na ascorbate$	[Et ₃ NH][OAc]	7 h	90	35 min	95
11	$CuSO_4 \cdot 5H_2O + Na ascorbate$	[Et ₃ NH][OAc]	7 h	81	35 min	84
12	$Cu(OAc)_2 \cdot H_2O$	[Et ₃ NH][Cl]	7 h	Trace	35 min	10
13	$Cu(OAc)_2 \cdot H_2O$	[Et ₃ NH][HSO ₄]	7 h	Trace	35 min	12
14	$Cu(OAc)_2 \cdot H_2O$	[HDBU][OAc]	7 h	84	45 min	88
15	$CuSO_4 \cdot 5H_2O + Na ascorbate$	HDBU	7 h	81	45 min	84
16	$Cu(OAc)_2 \cdot H_2O$	[HDBU][HSO4]	7 h	10	45 min	14

^a Reaction conditions: 2a (1 mmol), 5a (1 mmol), and 10 mol% [Cu], 1 mL solvent, 30 °C; reaction progress monitored by TLC. ^b Isolated yield, US: ultrasonication.

irradiation at 30 °C efficiently gives the desired 1,4disubstituted-1,2,3-triazole derivative 6a as compared to the conventional method with respect to the reaction time and yield (Table 1, entry 9).

In order to evaluate the amount of the $Cu(OAc)_2 \cdot H_2O$ catalyst for the model reaction for different concentrations such as 5, 10, 15, and 20 mol% (Table 2, entries 1-4), the reaction was carried out in the presence of 1 mL of [Et₃NH][OAc] as a solvent at 30 °C under ultrasound irradiation. It was observed that 10 mol% of $Cu(OAc)_2 \cdot H_2O$ was sufficient to achieve an excellent yield of the product 6a in the shortest reaction time (Table 2, entry 2), while an excess amount of the catalyst (up to 20 mol%) did not increase the yield of the product. The above results reveal that 10 mol% of Cu(OAc)₂·H₂O in [Et₃NH][OAc] is the sufficient catalyst amount in terms of an excellent yield and short reaction time (Table 2, entry 2).

The recyclability of the [Et₃NH][OAc] solvent was also explored for the model reaction. The reaction was conducted for four successive cycles on the same recycled solvent. After completion of the first reaction with 95% yield, the reaction mass was poured into ice cold water to get the solid product, which was isolated by filtration. The filtrate was subjected to evaporation of water to get a viscous liquid, which on cooling gave the ionic liquid. Then, the recovered [Et₃NH][OAc] was reused for four more successive cycles without significant loss in the yield of the product 6a. The yields achieved were 94%,

92%, 91%, and 88% after the successful runs (Fig. 3). The result demonstrates the high stability of the ionic liquid [Et₃NH][OAc] under the reaction conditions, which could be recycled and reused without a significant loss in the product yield.

In a further attempt, we also examined a one-pot reaction for the selected model reaction with 2a (1.0 mmol), 4a (1.0 mmol), and sodium azide (1.5 mmol) for the synthesis of 6a, which was optimized for different copper sources (Table 3) for the one-pot, three-component click reaction in [Et₃NH][OAc] as the solvent, thus producing 2-(4-(phenoxymethyl)-1H-1,2,3-triazol-1-yl)-Nphenylacetamide 6a (Scheme 3). The optimization result of this one-pot model reaction revealed that 15 mol% of $Cu(OAc)_2 \cdot H_2O$ in [Et₃NH][OAc] as the solvent was a sufficient catalyst loading amount to obtain an excellent yield. However, it required a longer reaction time and higher concentration of the Cu catalyst (Table 3, entry 2) than the two-component reaction, as discussed earlier.

After optimization of the reaction parameters and to define the present methodology, as discussed earlier, the reaction was assessed with a variety of substituted alkynes 2a-c and azides **5a-g** in the presence of $Cu(OAc)_2 \cdot H_2O$ in $[Et_3NH][OAc]$ under ultrasound irradiation. The reaction conditions were favorable for the electron-donating and electron-withdrawing groups on alkynes 2a-c and azides 5a-g, resulting in the corresponding 1,4-disubstituted-1,2,3-triazole derivatives 6a-g, 7a-g, and 8a-g in excellent yields (Scheme 4).

Table 2 The influence of the amount of $\mathsf{Cu}(\mathsf{OAc})_2\!\cdot\!\mathsf{H}_2\mathsf{O}$ on the synthesis of $\mathbf{6a}$

2a	N ₃ , NH + 5a	Cu(OAc) ₂ .H ₂ O	
Entry	mol%	Time (min)	Isolated yield (%)
1	5	55	70
1 2	5 10	55 35	70 95

All the synthesized compounds, namely, **6a–g**, **7a–g**, and **8a– g** were confirmed by physical data and spectral analysis.

2.2 Biological activity

2.2.1. Antifungal activity. All the newly synthesized compounds were screened for their in vitro antifungal activities against five different fungal strains, namely, Candida albicans, Fusarium oxysporum, Aspergillus flavus, Aspergillus niger, and Cryptococcus neoformans. The minimum inhibitory concentration (MIC, $\mu g m L^{-1}$) values of all the newly synthesized compounds were determined by the standard agar dilution method as per the CLSI (formerly, NCCLS) guidelines²⁶ (Table 4). DMSO was used as the negative control and Miconazole was used as the standard antifungal drug for the comparison of antifungal activities. Among the series, the triazole derivatives 6 and 7 derived from alkynes 2a and 2b (particularly from phenol and 4-methoxyphenol) did not show any activity against all the fungal strains. However, the triazoles derived from alkyne 2c (from 4-nitrophenol) displayed excellent antifungal activity against all the strains. The compounds 8b, 8c, and 8f having MIC of 12.5 μ g mL⁻¹ were found to be more potent; compounds 8a, 8d, and 8g having MIC of 25 μ g mL⁻¹ were found to be equipotent as compared to the standard drug Miconazole against the fungal strain Candida albicans. The compounds 8b, 8c, 8d, 8e, and 8g with MIC of 25 μ g mL⁻¹ were found to be equivalent to the standard drug Miconazole and 8f with MIC of



Fig. 3 Reusability of [Et₃NH][OAc].

Table 3 The effect of the amount of copper salts for the one-pot synthesis of 6a

Entry	Catalyst	mol%	Time (h)	Isolated yield (%)
1	$Cu(OAc)_2 \cdot H_2O$	10	17	90
2	$Cu(OAc)_2 \cdot H_2O$	15	16	95
3	$Cu(OAc)_2 \cdot H_2O$	20	16	95
4	$CuSO_4 \cdot 5H_2O + Na ascorbate$	10	18	81
5	$CuSO_4 \cdot 5H_2O + Na$ ascorbate	15	17	87
6	CuI	10	18	50
7	CuI	15	17	61
8	CuCl	10	17	40
9	CuCl	15	17	45

12.5 μ g mL⁻¹ was more potent than Miconazole against the fungal strain *F. oxysporum*. Compounds **8e** and **8g** with MIC of 12.5 μ g mL⁻¹ exhibited equivalent activities as compared to Miconazole against *A. flavus*. For the fungal strain *A. niger*, the compounds **8b**, **8d**, **8e**, and **8g** with MIC of 25 μ g mL⁻¹ displayed equivalent antifungal activities as compared to Miconazole. However, **8f** having MIC of 12.5 μ g mL⁻¹ was more active than Miconazole. For the fungal strain *C. neoformans*, the compounds **8e**, **8f**, and **8g** with MIC of 12.5 μ g mL⁻¹ were more potent than the standard drug and **8c** with MIC of 25 μ g mL⁻¹ was equivalent to Miconazole.

The remaining compounds displayed moderate to low antifungal activities against all the tested fungal strains. The antifungal activity depends on the various substituents present on phenyl rings. Among the series, the compounds with nitro groups (**8a–g**) were the most active against all the tested fungal strains. Compounds **8f** and **8g** were the most active in the series as they possessed a nitro group at R and chloro groups at R² and R³; the chloro group at the R¹ position for **8e** resulted in less activity against one fungal strain. The methyl group at the R¹/R²/R³ position and the nitro group at the R position in **8b**, **8c**, and **8d** resulted in good activity than that of the others in the series.

2.2.2. Antitubercular activity. All the newly synthesized 1,4disubstituted-1,2,3-triazoles (**6a–g**, **7a–g**, and **8a–g**) were screened for their *in vitro* antitubercular activities against the MTB H37Rv (ATCC 27294) strain using the microplate Alamar Blue assay method with the determination of MIC in triplicate; the results are mentioned in Table 4. The newly synthesized 1,2,3-triazoles exhibited MICs ranging from 25 to \geq 25 µg mL⁻¹. Among the series, compounds **6a**, **6c**, and **6f** exhibited good antitubercular activities with MIC of 25 µg mL⁻¹ as compared to other compounds.



Scheme 3 Synthesis of 1,4-disubstituted-1,2,3-triazole 6a via the one-pot three-component click reaction.



Scheme 4 Synthesis of 1,4-disubstituted-1,2,3-triazole derivatives 6a-g, 7a-g, and 8a-g.

Table 4	In vitro biological evaluation	of the synthesized	compounds 6a–g, 7a–g	, and $8a-g$ MIC (μ g mL ⁻¹) ^a
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	Antifungal activity							
Entry	CA	FO	AF	AN	CN	Anti TB MTB H37Rv	DPPH IC_{50} (µg mL ⁻¹)	
6a	87.5	100	175	125	187.5	25	55.37	
6b	150	125	150	150	*	>25	26.64	
6c	175	200	150	175	200	25	20.54	
6d	187.5	162.5	187.5	200	*	>25	*	
6e	200	*	*	187.5	175	>25	29.64	
6f	150	175	162.5	200	*	25	74.28	
6g	162.5	175	175	187.5	150	>25	74.41	
7a	137.5	125	100	137.5	125	>25	11.44	
7b	175	200	100	175	200	>25	16.79	
7c	*	*	200	*	175	>25	12.30	
7d	200	175	150	150	200	>25	19.46	
7e	175	175	200	*	*	>25	70.33	
7f	100	150	100	100	75	>25	*	
7g	75	100	87.5	100	200	>25	49.60	
8a	25	37.5	25	75	75	>25	97.29	
8b	12.5	25	25	25	37.5	>25	*	
8c	12.5	25	25	50	25	>25	48.02	
8d	25	25	37.5	25	50	>25	22.17	
8e	50	25	12.5	25	12.5	>25	62.01	
8f	12.5	12.5	25	12.5	12.5	>25	*	
8g	25	25	12.5	25	12.5	>25	16.99	
MA	25	25	12.5	25	25	NA	NA	
INH	NA	NA	NA	NA	NA	0.1	NA	
RIF	NA	NA	NA	NA	NA	0.2	NA	
EMB	NA	NA	NA	NA	NA	1.56	NA	
CIP	NA	NA	NA	NA	NA	1.56	NA	
BHT	NA	NA	NA	NA	NA	NA	16.47	

^{*a*} CA: *Candida albicans*; FO: *Fusarium oxysporum*; AF: *Aspergillus flavus*; AN: *Aspergillus niger* and CN: *Cryptococcus neoformans*. MA: Miconazole, DPPH: 2,2-diphenyl-1-picrylhydrazyl; INH: Isoniazid; RIF: Rifampicin; EMB: Ethambutol; CIP: Ciprofloxacin; BHT: butylated hydroxytoluene; *no activity was observed, NA: not applicable.

2.2.3. Antioxidant activity. The antioxidant activities of the synthesized compounds **6a–g**, **7a–g**, and **8a–g** were screened using a 2,2-diphenyl-1-picrylhydrazyl (DPPH) radical scavenging assay.²⁷ The DPPH radical scavenging assay is the most commonly used procedure for screening the antioxidant activities of different natural and synthetic compounds. A lower IC₅₀ value demonstrates more antioxidant activity. The IC₅₀ (concentration required to scavenge 50% of the radicals) values were calculated to assess the potential antioxidant activities.

Butylated hydroxytoluene (BHT) was used as the standard drug for the comparison of antioxidant activities; the observed results are summarized in Table 4. According to the DPPH assay, compounds 7a and 7c exhibited excellent radical scavenging activities with IC_{50} values of 11.44 and 12.30 µg mL⁻¹, respectively, compared to the synthetic antioxidant BHT. Similarly, compounds 7b, 7d, and 8g having IC_{50} values of 16.79, 19.46, and 16.99 µg mL⁻¹, respectively, showed comparable antioxidant activities with that of BHT; in contrast, the

Table 5	Molecular docking	score for the	compounds	6a-g, 7a-g	, and 8a–g
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Molecular docking score								
Total score $(-\log K_i)$	Crash score	Polar score	Entry	Total score $(-\log K_i)$	Crash score	Polar score		
7.0377	-0.6933	1.9115	7e	6.4127	-1.2494	1.4225		
6.4564	-2.9621	0.0004	7 f	7.2492	-1.4724	2.5093		
6.8229	-0.7726	2.0682	7g	7.2394	-1.4033	3.0192		
6.0468	-1.4517	1.3586	8a	7.0868	-1.1801	3.6314		
6.5873	-0.9241	2.8329	8b	7.5441	-2.8665	0.9019		
6.7343	-0.8582	3.0543	8c	7.4658	-1.0993	0.7391		
7.0332	-1.0463	1.6968	8d	7.3945	-1.5222	1.9024		
6.0045	-0.7865	2.8105	8e	7.3314	-0.9292	0.7808		
6.6907	-0.9183	4.5244	8f	7.9971	-1.9946	0.0031		
6.5748	-2.7834	0.0090	8g	7.5972	-1.3842	3.0509		
7.0239	-1.2982	3.2802	MA	6.4895	-1.4009	1.0311		
	Total score (-log K _i) 7.0377 6.4564 6.8229 6.0468 6.5873 6.7343 7.0332 6.0045 6.6907 6.5748	Total score $(-\log K_i)$ Crash score7.0377 -0.6933 6.4564 -2.9621 6.8229 -0.7726 6.0468 -1.4517 6.5873 -0.9241 6.7343 -0.8582 7.0332 -1.0463 6.0045 -0.7865 6.6907 -0.9183 6.5748 -2.7834	Total score $(-\log K_i)$ Crash scorePolar score7.0377 -0.6933 1.9115 6.4564 -2.9621 0.0004 6.8229 -0.7726 2.0682 6.0468 -1.4517 1.3586 6.5873 -0.9241 2.8329 6.7343 -0.8582 3.0543 7.0332 -1.0463 1.6968 6.0045 -0.7865 2.8105 6.6907 -0.9183 4.5244 6.5748 -2.7834 0.0090	Total score $(-\log K_i)$ Crash scorePolar scoreEntry7.0377 -0.6933 1.9115 7e6.4564 -2.9621 0.0004 7f6.8229 -0.7726 2.0682 7g6.0468 -1.4517 1.3586 8a6.5873 -0.9241 2.8329 8b6.7343 -0.8582 3.0543 8c7.0332 -1.0463 1.6968 8d6.0045 -0.7865 2.8105 8e6.6907 -0.9183 4.5244 8f6.5748 -2.7834 0.0090 8g	Total score $(-\log K_i)$ Crash scorePolar scoreEntryTotal score $(-\log K_i)$ 7.0377 -0.6933 1.9115 7c 6.4127 6.4564 -2.9621 0.0004 7f 7.2492 6.8229 -0.7726 2.0682 7g 7.2394 6.0468 -1.4517 1.3586 8a 7.0868 6.5873 -0.9241 2.8329 8b 7.5441 6.7343 -0.8582 3.0543 8c 7.4658 7.0332 -1.0463 1.6968 8d 7.3945 6.0045 -0.7865 2.8105 8e 7.3314 6.6907 -0.9183 4.5244 8f 7.9971 6.5748 -2.7834 0.0090 8g 7.5972	Total score $(-\log K_i)$ Crash scorePolar scoreEntryTotal score $(-\log K_i)$ Crash score7.0377 -0.6933 1.9115 7e 6.4127 -1.2494 6.4564 -2.9621 0.0004 7f 7.2492 -1.4724 6.8229 -0.7726 2.0682 7g 7.2394 -1.4033 6.0468 -1.4517 1.3586 8a 7.0868 -1.1801 6.5873 -0.9241 2.8329 8b 7.5441 -2.8665 6.7343 -0.8582 3.0543 8c 7.4658 -1.0993 7.0332 -1.0463 1.6968 8d 7.3945 -1.5222 6.0045 -0.7865 2.8105 8e 7.3314 -0.9292 6.6907 -0.9183 4.5244 8f 7.9971 -1.9946 6.5748 -2.7834 0.0090 8g 7.5972 -1.3842		



Fig. 4 Binding pose and molecular interactions in the active sites of *C*. *albicans* lanosterol 14α -demethylase: (a) **8f** and (b) **8g**.

compounds **6b**, **6c**, **6e**, and **8d** showed moderate activities with IC_{50} values of 26.64, 20.54, 29.64, and 22.17 µg mL⁻¹, respectively.

2.2.4. Molecular docking. The molecular docking study of all the synthesized triazole derivatives **6a–g**, **7a–g**, and **8a–g** was performed against the modelled three-dimensional structure of cytochrome P450 lanosterol 14α -demethylase of *C. albicans* to understand the binding affinity and binding interactions of the enzyme and the synthesized derivatives using the SYBEL 2.1.1 package following the standard procedures.^{28,29} The molecular docking results are presented in Table 5.

The docking results indicated that the triazole scaffold of the synthesized derivatives 6a-g, 7a-g, and 8a-g was incorporated appropriately in the active site of CYP450 and through various kinds of non-bonded interactions such as H-bond interactions and π bond interactions with the active sites of the amino acid residues. Among all the synthesized derivatives, 8f (7.9971), 8g (7.597), and 8b (7.544) were the most active triazole derivatives when compared with the standard Miconazole drug (6.4895). The most active triazole derivative 8f (7.9971) contained hydrophilic as well as hydrophobic amino acids in the active sites such as isoleucine, valine, methionine, cysteine, leucine, and phenylalanine. The hydrophobic amino acids Cys506 and Ile507 formed conventional hydrogen bonds with the oxygen (-O) of the nitro group on the phenyl ring with the distances of 1.95 and 2.02 Å, respectively. Met336 contains non-polar side chain forms conventional hydrogen bond interaction with a hydrogen atom (-H) attached to the nitrogen atom and methylene group with distance 2.41 and 2.45 Å, respectively. The hydrophobic amino acids such as Val166, Ile167, Ala182, Leu186, Leu240, Leu316, and Ala185 interacted with the π electron cloud to form weak π -interactions such as π -alkyl and

alkyl interactions with the chloro-phenyl (–Cl) ring; the π electron cloud with the phenyl ring and the π electron cloud of the triazole ring are shown in Fig. 4a. The second most active triazole derivative **8g** (7.597) interacted with the active site of amino acids by forming non-covalent bonded interactions such as H-bond interaction and π bond interaction. The hydrophobic amino acid Ala501 contains a non-polar side chain that interacts with the oxygen (–O) atom of the nitro group to form a conventional hydrogen bond interaction with distance of 2.02 Å. The polar aromatic amino acid Tyr168 forms hydrogen bond interactions with the nitrogen atom of the triazole ring with a distance of 1.84 Å.

The amino acid Gly500 formed a carbon-hydrogen bond with the oxygen (–O) of the nitro group on the phenyl ring with a distance of 2.39 Å. The hydrophobic amino acid Try154 with an aromatic ring interacted with the triazole π cloud to form π - π stacked interactions, while the amino acids Leu412, Met544, Met413, and His546 interacted with the π cloud of the chloro group on the phenyl ring (–Cl) and the phenyl ring to form weak π -alkyl and alkyl interactions, as shown in Fig. 4b.

3. Conclusion

In conclusion, we demonstrated [Et₃NH][OAc]- and ultrasoundmediated synthesis of new *N*-phenylacetamide-incorporated 1,2,3-triazole derivatives **6(a–g)**, **7(a–g)**, and **8(a–g)** in excellent yields *via* the click chemistry approach for the first time. This safer method reduced the use of harmful solvents and reaction time. We also studied the recovery and reusability of [Et₃NH] [OAc] as a medium. The synthesized triazole derivatives were evaluated for their antifungal, antioxidant, and antitubercular activities. Among the series, the compounds **8a**, **8b**, **8c**, **8d**, **8e**, **8f**, and **8g** displayed excellent antifungal activities as compared to the standard drug Miconazole with lower MIC values.

The *in silico* molecular docking study supported the experimental results and demonstrated that the 1,2,3-triazole derivatives **8f** and **8g** are the most active and have the potential to inhibit the survival of fungal species. The predictions of the pharmacokinetic parameters suggest that the synthesized derivatives have the potential to have high oral drug bioavailability.

4. Experimental

4.1 General methods

All the solvents and reagents were purchased from commercial suppliers, namely, Spectrochem Pvt. Ltd., Avra, Sigma Aldrich, and Alfa Aesar and were used without further purification. The progress of each reaction was monitored by thin layer chromatography (TLC) using silica gel F_{254} precoated TLC aluminium sheets (Merck, Germany) and by locating the spots using a UV light as the visualizing agent or iodine vapours. The melting points were obtained by the open capillary method and were uncorrected. The IR spectra were recorded on a Bruker FT-IR spectrometer. The ¹H NMR spectra were recorded on Bruker AC-200 MHz and Bruker Advance III 500 MHz NMR spectrometers. The ¹³C NMR spectra were recorded on Bruker AC-50 MHz

Paper

and Bruker Advance III 125 MHz NMR in DMSO- d_6 , CDCl₃ + DMSO- d_6 , and CDCl₃ + CD₃OD using tetramethylsilane (TMS) as the internal standard; the chemical shifts (δ) were recorded in parts per million (ppm). The splitting pattern abbreviations are designed as singlet (s), doublet (d), double doublet (dd), triplet (t), quartet (q), and multiplet (m). High-resolution mass spectra (HRMS) were recorded on an Agilent 6520 (QTOF) ESI-HRMS instrument and Agilent Technologies 1200 series electron spray ionization (ESI) was used for the LC-MS data.

4.2 General experimental procedure for the synthesis of *N*-phenylacetamide-incorporated 1,2,3-triazole derivatives (6a–g, 7a–g, and 8a–g)

The mixture of appropriate (prop-2-yn-1-yloxy) benzenes **2a–c** (1 mmol), substituted 2-azido-*N*-phenylacetamide **5a–g** (1 mmol) and Cu(OAc)₂·H₂O (10 mol%) in [NEt₃H][OAc] (1 mL) was placed in a round bottom flask. The mixture was sonicated (40 kHz) at 30 °C for an appropriate time until the completion of the reaction. The progress of the reaction was monitored by TLC. After the completion of the reaction, the mixture was quenched with crushed ice. The obtained solid was filtered and washed with water. The crude solid was crystallized in ethanol to afford the corresponding pure product. The synthesized compounds **6a–g**, **7a–g**, and **8a–g** were characterized by IR, ¹H NMR, ¹³C NMR, and mass spectroscopy.

4.2.1. 2-(4-(Phenoxymethyl)-1*H*-1,2,3-triazol-1-yl)-*N*-phenylacetamide (6a). The compound 6a was obtained *via* a 1,3dipolar cycloaddition reaction between azide 5a and alkyne 2a in 35 min as a white solid; mp: 166–168 °C; FT-IR (cm⁻¹): 3263 (N–H stretching), 1667 (C=O stretching); ¹H NMR (200 MHz, DMSO- d_6 , δ ppm): 5.17 (s, 2H, –NCH₂CO–), 5.36 (s, 2H, –OCH₂), 6.96 (t, *J* = 8.0 Hz, 1H, Ar–H), 7.08 (t, *J* = 8.0 Hz, 3H, Ar), 7.28– 7.38 (m, 4H, Ar–H), 7.59 (d, *J* = 8.0 Hz, 2H, Ar–H), 8.26 (s, 1H, triazole), and 10.49 (s, 1H, NH); ¹³C NMR (50 MHz, DMSO- d_6 , δ ppm): 51.8, 60.5, 114.3, 118.9, 120.5, 123.5, 125.9, 128.6, 129.2, 138.0, 142.2, 157.7, and 163.8; HRMS calcd for C₁₇H₁₇N₄O₂ [M + H]⁺: 309.1346 and found 309.1340.

4.2.2. 2-(4-(Phenoxymethyl)-1*H*-1,2,3-triazol-1-yl)-*N*-(*o*-tolyl) acetamide (6b). The compound 6b was obtained *via* a 1,3-dipolar cycloaddition reaction between azide 5b and alkyne 2a in 37 min as a white solid; mp: 184–186 °C; FT-IR (cm⁻¹): 3271 (N–H stretching), 1665 (C=O stretching); ¹H NMR (200 MHz, DMSO-*d*₆, δ ppm): 2.24 (s, 3H, -CH₃), 5.17 (s, 2H, -NCH₂CO–), 5.41 (s, 2H, -OCH₂), 6.95 (t, *J* = 8.0 Hz, 1H, Ar–H), 7.05 (d, *J* = 8.0 Hz, 2H, Ar–H), 7.10–7.35 (m, 5H, Ar–H), 7.43 (d, *J* = 8.0 Hz, 1H, Ar–H), 8.26 (s, 1H, triazole), and 9.84 (s, 1H, NH); ¹³C NMR (50 MHz, DMSO-*d*₆, δ ppm): 17.5, 51.3, 60.5, 114.3, 120.4, 124.8, 125.8, 129.1, 130.1, 131.2, 135.1, 142.2, 157.7, and 164.0; HRMS calcd for C₁₈H₁₉N₄O₂ [M + H]⁺: 323.1508 and found 323.1510.

4.2.3. 2-(4-(Phenoxymethyl)-1*H*-1,2,3-triazol-1-yl)-*N*-(*m*-tolyl)acetamide (6c). The compound 6c was obtained *via* a 1,3-dipolar cycloaddition reaction between azide 5c and alkyne 2a in 38 min as a yellow solid; mp: 130–132 °C; FT-IR (cm⁻¹): 3302 (N–H stretching), 1676 (C=O stretching); ¹H NMR (500 MHz, CDCl₃ + DMSO- d_6 , δ ppm): 2.21 (s, 3H, –CH₃), 5.10 (s, 2H, –NCH₂CO–), 5.13 (s, 2H, –OCH₂), 6.79–6.93 (m,

4H, Ar–H), 7.08 (d, J = 8.0 Hz 1H), 7.13–7.30 (m, 3H, Ar–H), 7.34 (s, 1H, Ar–H), 7.85 (s, 1H, triazole), and 9.70 (s, 1H, NH); ¹³C NMR (125 MHz, CDCl₃ + DMSO- d_6 , δ ppm): 21.1, 52.7, 61.4, 114.3, 116.7, 120.2, 124.8, 124.9, 128.3, 129.1, 137.3, 138.4, 143.7, 157.8, and 163.0; LC-MS for C₁₈H₁₈N₄O₂: [M]⁺ 323.1.

4.2.4. 2-(4-(Phenoxymethyl)-1*H*-1,2,3-triazol-1-yl)-*N*-(*p*-tolyl)acetamide (6d). The compound 6d was obtained *via* a 1,3-dipolar cycloaddition reaction between azide 5d and alkyne 2a in 36 min as a yellow solid; mp: $202-204 \degree C$; FT-IR (cm⁻¹): 3271 (N-H stretching), 1665 (C=O stretching).

4.2.5. 2-(4-(Phenoxymethyl)-1*H*-1,2,3-triazol-1-yl)-*N*-(2chlorophenyl)-acetamide (6e). The compound 6e was obtained *via* a 1,3-dipolar cycloaddition reaction between azide 5e and alkyne 2a in 36 min as a white solid; mp: 178– 180 °C; FT-IR (cm⁻¹): 3267 (N-H stretching), 1671 (C=O stretching); ¹H NMR (500 MHz, CDCl₃ + DMSO-*d*₆, δ ppm): 5.19 (s, 2H, -NCH₂CO-), 5.31 (s, 2H, -OCH₂), 6.94 (d, *J* = 8.0 Hz, 3H, Ar-H), 7.05 (d, *J* = 8.0 Hz, 1H, Ar-H), 7.16-7.34 (m, 4H, Ar-H), 7.89 (s, 1H, triazole), 8.02 (d, *J* = 8.0 Hz, 1H, Ar-H), and 9.01 (s, 1H, NH); ¹³C NMR (125 MHz, CDCl₃ + DMSO-*d*₆, δ ppm): 52.8, 61.3, 114.3, 120.8, 123.1, 124.3, 125.5, 127.1, 129.0, 133.7, 141.9, 157.7, and 162.6. LC-MS for C₁₇H₁₅ClN₄O₂: [M]⁺ 343.

4.2.6. 2-(4-(Phenoxymethyl)-1*H*-1,2,3-triazol-1-yl)-*N*-(3chlorophenyl)-acetamide (6f). The compound 6f was obtained *via* a 1,3-dipolar cycloaddition reaction between azide 5f and alkyne 2a in 37 min as a white solid; mp: 140– 142 °C; FT-IR (cm⁻¹): 3257 (N–H stretching), 1672 (C=O stretching); ¹H NMR (500 MHz, CDCl₃ + DMSO-*d*₆, δ ppm): 5.45 (s, 2H, –NCH₂CO–), 5.46 (s, 2H, –OCH₂), 7.15–7.57 (m, 7H Ar–H), 7.67 (s, 1H, Ar–H), 7.92 (s, 1H, triazole), 8.15 (s, 1H, Ar–H), and 10.16 (s, 1H, NH). ¹³C NMR (125 MHz, CDCl₃ + DMSO-*d*₆, δ ppm): 53.1, 61.7, 114.7, 117.9, 120.0, 121.2, 124.5, 129.5, 129.9, 134.4, 138.9, 158.1, and 163.5; LC-MS for C₁₇H₁₅ClN₄O₂: [M]⁺ 343.1.

4.2.7. 2-(4-((4-Methoxyphenoxy)methyl)-1*H*-1,2,3-triazol-1yl)-*N*-phenylacetamide (7a). The compound 7a was obtained *via* a 1,3-dipolar cycloaddition reaction between azide 5a and alkyne 2b in 37 min as a white solid; mp: 180–182 °C; FT-IR (cm⁻¹): 3271 (N–H stretching), 1674 (C=O stretching); ¹H NMR (500 MHz, CDCl₃, δ ppm): 3.76 (s, 3H, OCH₃), 5.19 (s, 2H, –NCH₂CO–), 5.20 (s, 2H, –OCH₂), 6.83 (d, *J* = 8.0 Hz, 2H, Ar–H), 6.92 (d, *J* = 8.0 Hz, 2H, Ar–H), 7.14 (t, *J* = 8.0 Hz, 1H, Ar–H), 7.31 (t, *J* = 8.0 Hz, 2H, Ar–H), 7.43 (d, *J* = 8.0 Hz, 2H, Ar–H), 7.81 (s, 1H, triazole), and 7.87 (s, 1H, NH); ¹³C NMR (125 MHz, CDCl₃, δ ppm): 53.9, 55.7, 62.5, 114.8, 115.9, 120.3, 125.4, 129.1, 137.0, 139.3, 152.1, and 162.3; LC-MS for C₁₈H₁₈N₄O₃: [M]⁺ 339.1.

4.2.8. 2-(4-((4-Methoxyphenoxy)methyl)-1*H*-1,2,3-triazol-1yl)-*N*-(*o*-tolyl)acetamide (7b). The compound 7b was obtained *via* a 1,3-dipolar cycloaddition reaction between azide 5b and alkyne 2b in 35 min as a white solid; mp: 176–178 °C; FT-IR (cm⁻¹): 3266 (N–H stretching), 1674 (C=O stretching); ¹H NMR (500 MHz, CDCl₃, δ ppm): 2.10 (s, 3H, CH₃), 3.75 (s, 3H, –OCH₃), 5.21 (s, 2H, –NCH₂CO–), 5.22 (s, 2H, –OCH₂), 6.82 (d, *J* = 8.0 Hz, 2H, Ar–H), 6.91 (d, *J* = 8.0 Hz, 2H, Ar–H), 7.08 (t, *J* = 8.0 Hz, 1H, Ar–H), 7.15 (d, *J* = 8.0 Hz, 1H, Ar–H), 7.20 (t, *J* = 8.0 Hz, 1H, Ar–H), 7.80 (s, 1H, triazole), and 7.82 (s, 1H, NH); $^{13}\mathrm{C}$ NMR (125 MHz, CDCl₃, δ ppm): 17.4, 53.9, 55.7, 62.4, 114.8, 115.9, 122.3, 124.5, 125.9, 126.9, 127.9, 137.6, and 162.8; LC-MS for $\mathrm{C_{19}H_{20}N_4O_3}$: $[M]^+$ 353.1.

4.2.9. 2-(4-((4-Methoxyphenoxy)methyl)-1*H*-1,2,3-triazol-1yl)-*N*-(*m*-tolyl)acetamide (7c). The compound 7c was obtained *via* a 1,3-dipolar cycloaddition reaction between azide 5c and alkyne 2b in 35 min as a yellow solid; mp: 140–142 °C; FT-IR (cm⁻¹): 3271 (N–H stretching), 1665 (C=O stretching); ¹H NMR (500 MHz, CDCl₃ + DMSO-*d*₆, δ ppm): 2.18 (s, 3H, CH₃), 3.62 (s, 3H, OCH₃), 5.01 (s, 2H, –NCH₂CO–), 5.10 (s, 2H, –OCH₂), 6.68 (d, *J* = 6 Hz, 2H, Ar–H), 6.79 (d, *J* = 8.0 Hz, 2H, Ar–H), 7.04 (t, *J* = 8.0 Hz, 1H, Ar–H), 7.18 (d, *J* = 8.0 Hz, 1H, Ar–H), 7.26 (d, *J* = 8.0 Hz, 1H, Ar–H), 7.31 (s, 1H, Ar–H), 7.79 (s, 1H, triazole), and 9.73 (s, 1H, NH); ¹³C NMR (125 MHz, CDCl₃ + DMSO-*d*₆, δ ppm): 21.3, 52.9, 55.5, 62.4, 114.5, 115.7, 116.8, 120.4, 124.4, 125.1, 128.5, 137.6, 138.5, 144.1, 152.2, 153.9, and 163.3. LC-MS for C₁₉H₂₀N₄O₃: [M]⁺ 353.1.

4.2.10. 2-(4-((4-Methoxyphenoxy)methyl)-1H-1,2,3-triazol-

1-yl)-*N***-(***p***-tolyl)acetamide (7d). The compound 7d was obtained** *via* **a 1,3-dipolar cycloaddition reaction between azide 5d and alkyne 2b in 36 min as a white solid; mp: 196–198 °C; FT-IR (cm⁻¹): 3273 (N–H stretching), 1675 (C=O stretching); ¹H NMR (500 MHz, CDCl₃ + DMSO-***d***₆, \delta ppm): 2.10 (s, 3H, CH₃), 3.55 (s, 3H, OCH₃), 4.94 (s, 2H, –NCH₂CO–), 5.02 (s, 2H, –OCH₂), 6.62 (d,** *J* **= 8.0 Hz, 2H, Ar–H), 6.72 (d,** *J* **= 8.0 Hz, 2H, Ar–H), 6.89 (d,** *J* **= 8.0 Hz, 2H, Ar–H), 7.24 (d,** *J* **= 8.0 Hz, 2H, Ar–H), 7.73 (s, 1H, triazole), and 9.68 (s, 1H, NH); ¹³C NMR (125 MHz, CDCl₃ + DMSO-***d***₆, \delta ppm): 20.4, 52.6, 55.2, 62.1, 114.2, 115.4, 119.5, 124.2, 128.9, 133.5, 134.9, 151.9, 153.6, and 162.8; LC-MS for C₁₉H₂₀N₄O₃: [M]⁺ 353.1.**

4.2.11. *N*-(2-Chlorophenyl)-2-(4-((4-methoxyphenoxy)methyl)-**1***H*-1,2,3-triazol-1-yl)acetamide (7e). The compound 7e was obtained *via* a 1,3-dipolar cycloaddition reaction between azide 5e and alkyne 2b in 35 min as a white solid; mp: 168–170 °C; FT-IR (cm⁻¹): 3260 (N–H stretching), 1683 (C=O stretching); ¹H NMR (500 MHz, CDCl₃ + DMSO-*d*₆, δ ppm): 3.89 (s, 3H, OCH₃), 5.32 (s, 2H, –NCH₂CO–), 5.47 (s, –OCH₂), 6.96 (d, *J* = 8.0 Hz, 2H, Ar–H), 7.06 (d, *J* = 8.0 Hz, 2H, Ar–H), 7.23 (t, *J* = 8.0 Hz, 1H, Ar–H), 7.39 (t, *J* = 8.0 Hz, 1H, Ar–H), 7.50 (d, *J* = 8.0 Hz, 1H, Ar–H), 8.03 (s, 1H, triazole), 8.26 (d, *J* = 8.0 Hz, 1H, Ar–H), and 8.98 (s, 1H, NH); ¹³C NMR (125 MHz, CDCl₃ + DMSO-*d*₆, δ ppm): 53.0, 55.4, 62.3, 114.4, 115.6, 122.8, 124.3, 125.6, 127.3, 129.1, 133.4, 144.7, 152.0, 153.9, and 163.3; LC-MS for C₁₈H₁₇ClN₄O₃: [M]⁺ 373.

4.2.12. *N*-(2-Chlorophenyl)-2-(4-((4-methoxyphenoxy)methyl)-1*H*-1,2,3-triazol-1-yl)acetamide (7f). The compound 7f was obtained *via* a 1,3-dipolar cycloaddition reaction between azide 5f and alkyne 2b in 35 min as a green solid; mp: 160–162 °C; FT-IR (cm⁻¹): 3266 (N–H stretching), 1675 (C=O stretching); ¹H NMR (500 MHz, CDCl₃ + DMSO-*d*₆, δ ppm): 3.57 (s, 3H, OCH₃), 4.96 (s, 2H, –NCH₂CO–), 5.05 (s, 2H, –OCH₂), 6.63 (d, *J* = 8.0 Hz, 2H, Ar–H), 6.73 (d, *J* = 8.0 Hz, 2H, Ar–H), 6.87 (d, *J* = 8.0 Hz, 1H, Ar–H), 7.03 (t, *J* = 8.0 Hz, 1H, Ar–H), 7.24 (d, *J* = 8.0 Hz, 1H, Ar–H), 7.53 (s, 1H, Ar– H), 7.73 (s, 1H, triazole), and 10.04 (s, 1H, NH); ¹³C NMR (125 MHz, CDCl₃ + DMSO-*d*₆, δ ppm): 52.4, 55.1, 62.0, 114.1, 115.27, 117.3, 119.3, 123.7, 124.1, 129.4, 133.8, 138.7, 151.8, 153.5, and 163.2; LC-MS for C₁₈H₁₇ClN₄O₃ [M + H]⁺ 373. **4.2.13.** *N*-(**4-Chlorophenyl**)-2-(**4-((4-methoxyphenoxy)methyl**)-**1***H*-**1**,**2**,**3-triazol-1-yl**)**acetamide** (7g). The compound 7g was obtained *via* a **1**,**3-**dipolar cycloaddition reaction between azide **5g** and alkyne **2b** in 36 min as a white solid; mp: 192–194 °C; FT-IR (cm⁻¹): 3266 (N-H stretching), 1672 (C=O stretching).

4.2.14. 2-(4-((4-Nitrophenoxy)methyl)-1*H***-1,2,3-triazol-1-yl)-***N***-phenylacetamide (8a). The compound 8a was obtained** *via* **a 1,3-dipolar cycloaddition reaction between azide 5a and alkyne 2c** in 40 min as a yellow solid; mp: 180–182 °C; FT-IR (cm⁻¹): 3250 (N–H stretching), 1682 (C=O stretching); ¹H NMR (500 MHz, CDCl₃ + CD₃OD, δ ppm): 5.18 (s, 2H, –NCH₂CO–), 5.27 (s, 2H, –OCH₂), 7.05 (d, J = 8.0 Hz, 3H, Ar–H), 7.27 (t, J = 8.0 Hz, 3H, Ar–H), 7.49 (d, J = 8.0 Hz, 2H, Ar–H), 7.98 (s, 1H, triazole), and 8.17 (d, J = 8 Hz, 2H, Ar–H); ¹³C NMR (125 MHz, CDCl₃ + CD₃OD, δ ppm): 52.7, 61.9, 114.5, 125.1, 125.7, 128.8, 137.5, 141.7, and 162.8; LC-MS for C₁₇H₁₅N₅O₄: [M]⁺ 354.

4.2.15. 2-(4-((4-Nitrophenoxy)methyl)-1*H*-1,2,3-triazol-1-yl)-*N*-(*o*-tolyl)acetamide (8b). The compound 8b was obtained *via* a 1,3-dipolar cycloaddition reaction between azide 5b and alkyne 2c in 39 min as a white solid; mp: 178–180 °C; FT-IR (cm⁻¹): 3257 (N–H stretching), 1668 (C=O stretching); ¹H NMR (500 MHz, CDCl₃ + CD₃OD, δ ppm): 2.15 (s, 3H, CH₃), 5.23 (s, 2H, –NCH₂CO–), 5.25 (s, 2H, –OCH₂), 7.03–7.13 (m, 6H, Ar–H), 7.47 (s, 1H, NH), 7.98 (s, 1H, triazole), and 8.14 (s, 2H, Ar–H); ¹³C NMR (125 MHz, CDCl₃ + CD₃OD, δ ppm): 17.2, 52.6, 62.1, 114.9, 124.9, 125.4, 125.9, 126.4, 126.5, 127.4, 130.7, 131.2, and 163.7; LC-MS for C₁₈H₁₇N₅O₄ [M]⁺ 368.1.

4.2.16. 2-(4-((4-Nitrophenoxy)methyl)-1*H*-1,2,3-triazol-1-yl)-*N*-(*m*-tolyl)acetamide (8c). The compound 8c was obtained *via* a 1,3-dipolar cycloaddition reaction between azide 5c and alkyne 2c in 38 min as a white solid; mp: 168–170 °C; FT-IR (cm⁻¹): 3280 (N–H stretching), 1662 (C=O stretching); ¹H NMR (200 MHz, DMSO- d_6 , δ ppm): 2.28 (s, 3H, CH₃), 5.36 (s, 4H, –NCH₂CO, –OCH₂), 6.91 (d, *J* = 8.0 Hz, 1H, Ar–H), 7.19 (d, *J* = 8.0 Hz, 1H, Ar–H), 7.27 (t, *J* = 8 Hz, 1H, Ar–H), 7.33 (d, *J* = 8.0 Hz, 2H, Ar–H), 7.42 (s, 1H, Ar–H), 8.24 (d, *J* = 8.0 Hz, 2H, Ar–H), 8.32 (s, 1H, triazole), and 10.42 (s, 1H, NH); ¹³C NMR (50 MHz, DMSO- d_6 , δ ppm): 21.1, 52.2, 61.9, 115.3, 116.4, 119.8, 124.5, 125.9, 126.7, 128.7, 138.1, 138.3, 141.0, 141.5, 163.3, and 164.0; HRMS calcd for C₁₈H₁₈N₅O₄ [M + H]⁺: 368.1359 and found 368.1364.

4.2.17. 2-(4-((4-Nitrophenoxy)methyl)-1*H*-1,2,3-triazol-1-yl)-*N*-(*p*-tolyl)acetamide (8d). The compound 8d was obtained *via* a 1,3-dipolar cycloaddition reaction between azide 5d and alkyne 2c in 40 min as a white solid; mp: 210–212 °C; FT-IR (cm⁻¹): 3260 (N–H stretching), 1696 (C=O stretching); ¹H NMR (500 MHz, CDCl₃ + CD₃OD, δ ppm): 2.26 (s, 3H, CH₃), 5.14 (s, 2H, –NCH₂CO–), 5.26 (s, 2H, –OCH₂), 7.0 (d, *J* = 8.0 Hz, 2H, Ar–H), 7.07 (d, *J* = 8.0 Hz, 2H, Ar–H), 7.35 (d, *J* = 8.0 Hz, 2H, Ar–H), 7.96 (s, 1H, triazole), and 8.16 (d, *J* = 8.0 Hz, 2H, Ar–H); ¹³C NMR (125 MHz, CDCl₃ + CD₃OD, δ ppm): 17.7, 53.1, 62.6, 115.4, 124.8, 125.9, 126.5, 126.9, 131.2, 143.5, 152.0, and 164.2; LC-MS for C₁₈H₁₇N₅O₄: [M + H]⁺ 368.1.

4.2.18. *N*-(2-Chlorophenyl)-2-(4-((4-nitrophenoxy)methyl)-1H-1,2,3-triazol-1-yl)acetamide (8e). The compound 8e was obtained *via* a 1,3-dipolar cycloaddition reaction between azide 5e and alkyne **2c** in 39 min as a red solid; mp: 169–171 °C; FT-IR (cm⁻¹): 3243 (N–H stretching), 1672 (C=O stretching); ¹H NMR (500 MHz, CDCl₃ + CD₃OD, δ ppm): 5.31 (s, 2H, –NCH₂CO–), 5.33 (s, 2H, –OCH₂), 7.08 (d, *J* = 8.0 Hz, 3H, Ar–H), 7.29 (d, *J* = 8.0 Hz, 2H), 7.37 (d, *J* = 8.0 Hz, 1H, Ar–H), 7.96 (s, 1H, triazole), 8.15 (s, 1H, NH), and 8.20 (d, *J* = 8.0 Hz, 2H, Ar–H); ¹³C NMR (125 MHz, CDCl₃ + CD₃OD, δ ppm): 53.2, 62.1, 114.8, 125.1, 126.0, 126.1, 127.7, 129.4, 133.5, 136.5, 146.4, 152.3, and 163.0; LC-MS for C₁₇H₁₄ClN₅O₄: [M]⁺ 388.

4.2.19. *N*-(3-Chlorophenyl)-2-(4-((4-nitrophenoxy)methyl)-1*H*-1,2,3-triazol-1-yl)acetamide (8f). The compound 8f was obtained *via* a 1,3-dipolar cycloaddition reaction between azide 5f and alkyne 2c in 40 min as a white solid; mp: 190–192 °C; FT-IR (cm⁻¹): 3327 (N–H stretching), 1683 (C=O stretching); ¹H NMR (200 MHz, DMSO-*d*₆, δ ppm): 5.36 (s, 2H, –NCH₂CO–), 5.40 (s, 2H, –OCH₂), 7.15 (d, *J* = 8.0 Hz, 1H, Ar–H), 7.29 (d, *J* = 8.0 Hz, 2H, Ar–H), 7.42 (d, *J* = 8.0 Hz, 2H, Ar–H), 7.78 (s, 1H, Ar–H), 8.23 (d, *J* = 8.0 Hz, 2H, Ar–H), 8.33 (s, 1H, triazole), and 10.81 (s, 1H, NH); ¹³C NMR (50 MHz, DMSO-*d*₆, δ ppm): 52.3, 61.9, 115.4, 117.7, 118.9, 123.6, 125.9, 126.8, 130.7, 133.3, 139.9, 141.1, 141.7, 163.4, and 164.7; HRMS calcd for C₁₇H₁₅ClN₅O₄ [M + H]⁺: 388.0813 and found 388.0814.

4.2.20. *N*-(**4**-Chlorophenyl)-2-(**4**-((**4**-nitrophenoxy)methyl)-**1***H*-**1**,**2**,**3**-triazol-**1**-yl)acetamide (8g). The compound 8g was obtained *via* a 1,3-dipolar cycloaddition reaction between azide 5g and alkyne **2c** in 37 min as a white solid; mp: 188–190 °C; FT-IR (cm⁻¹): 3243 (N–H stretching), 1672 (C=O stretching); ¹H NMR (200 MHz, DMSO-*d*₆, δ ppm): 5.36 (s, 4H, –NCH₂CO, –OCH₂), 7.29 (d, *J* = 8.0 Hz, 2H, Ar–H), 7.39 (d, *J* = 8.0 Hz, 2H, Ar–H), 7.61 (d, *J* = 8.0 Hz, 2H, Ar–H), 8.23 (d, *J* = 8.0 Hz, 2H, Ar–H), 8.32 (s, 1H, triazole), and 10.64 (s, 1H, NH); ¹³C NMR (50 MHz, DMSO*d*₆, δ ppm): 52.2, 61.9, 115.3, 120.8, 125.8, 126.7, 127.4, 128.8, 137.3, 141.0, 141.6, 163.3, and 164.3; HRMS calcd for C₁₇H₁₅ClN₅O₄ [M + H]⁺: 388.0813 and found 388.0819.

4.3 Experimental protocol for biological activity

4.3.1. Antifungal activity. Antifungal activity was determined by the standard agar dilution method as per the CLSI (formerly, NCCLS) guidelines.²⁶ The newly synthesized compounds were screened for their in vitro antifungal activity against five human pathogenic fungal strains including Candida albicans (NCIM 3471), Fusarium oxysporum (NCIM 1332), Aspergillus flavus (NCIM 539), Aspergillus niger (NCIM 1196), and Cryptococcus neoformans (NCIM 576). The synthesized compounds and the standard Miconazole were dissolved in DMSO. The medium yeast nitrogen base was dissolved in phosphate buffer of pH 7 and was autoclaved at 110 °C for 10 min. With each set, a growth control without the antifungal agent and solvent control DMSO were included. The fungal strains were freshly subcultured onto sabouraud dextrose agar (SDA) and incubated at 25 °C for 72 h. The fungal cells were suspended in sterile distilled water and diluted to get 105 cells per mL. Ten μ L of the standardized suspension was inoculated onto the control plates and the media incorporated with the antifungal agents. The inoculated plates were incubated at 25 °C for 48 h. The readings were taken at the end of 48 and 72 h. MIC is the lowest

concentration of the drug preventing the growth of macroscopically visible colonies on the drug-containing plates when there is visible growth on the drug-free control plates.

4.3.2. *In vitro* **Mtb MABA assay.** The newly synthesized compounds were screened for their *in vitro* antitubercular activity against *Mtb H37Rv* (ATCC 27294) by using the Microplate Alamar Blue Assay (MABA)³⁰ for the determination of MIC in triplicate. The MIC (in μ g mL⁻¹) was recorded as the lowest concentration/highest dilution of the compounds/control drugs that completely inhibited the growth of the *Mtb* cultures. The MIC values of the compounds were compared with those of the standard drugs (Isoniazid, Rifampicin, Ethambutol, and Ciprofloxacin). The experimental method for antitubercular activity is briefly described as follows.

Initially, the inoculum was prepared from a fresh LJ medium re-suspended in the 7H9-S medium (7H9 broth, 0.1% casitone, 0.5% glycerol, supplemented oleic acid, albumin, dextrose, and catalase [OADC]), adjusted to a McFarland tube no. 1, and diluted 1:20; 100 µL was used as the inoculum. Each drug stock solution was thawed and diluted in 7H9-S at four-fold the final highest concentration tested. Serial two-fold dilutions of each drug were prepared directly in a sterile 96-well microtiter plate using 100 µL 7H9-S. A growth control containing no antibiotic and a sterile control was also prepared on each plate. Sterile water was added to all perimeter wells to avoid evaporation during incubation. The plate was covered, sealed in plastic bags, and incubated at 37 °C in normal atmosphere. After 7 days incubation, 30 µL of alamar blue solution was added to each well, and the plate was re-incubated overnight. A change in the colour from blue (oxidised state) to pink (reduced) indicated the growth of bacteria, and the MIC was defined as the lowest concentration of the drug that prevented this change in colour.

4.3.3. Antioxidant activity (DPPH radical scavenging). The hydrogen atom or electron donation ability of some of the compounds was measured from the bleaching of the purple coloured methanol solution of 1,1-diphenyl-1-picrylhydrazyl (DPPH). The spectrophotometric assay uses the stable radical DPPH as a reagent. One mL of various concentrations of the test compounds (5, 10, 25, 50, and 100 μ g mL⁻¹) in methanol was added to 4 mL of 0.004% IJ w/v methanol solution of DPPH. The reaction mixture was incubated at 37 °C. The scavenging activity on DPPH was determined by measuring the absorbance at 517 nm after 30 min. All tests were performed in triplicate, and the mean values were obtained. The percent of inhibition of free radical production from DPPH was calculated by using the following equation:

% of scavenging = $[(A_{\text{control}} - A_{\text{sample}})/\text{IJA}_{\text{sample}} \times 100)],$

where A_{control} is the absorbance of the control (DPPH radical without test sample) and A_{sample} is the absorbance of the test sample (DPPH radical with test sample). The control contained all the reagents except the test samples.

4.3.4. Homology modeling. The three-dimensional homology model structure of cytochrome P450 lanosterol 14α -demethylase of *C. albicans* was predicted using the theoretical methods of protein structure prediction, *i.e.*, comparative

modelling using the Biopredicta module of VLifeMDS 4.3 Pro-Model. The protein sequence of cytochrome P450 lanosterol 14ademethylase for C. albicans was retrieved from the protein knowledge database Universal Protein Resource (UniProtKB) (http://www.uniprot.org/) (accession code: P10613) and its homologous template was identified from the protein structural database, i.e., Protein Data Bank (PDB) using Basic Local Alignment Search Tool for proteins (BLASTp). The homologous template identified using BLASTp solved the three-dimensional crystal structure of human lanosterol 14a-demethylase (CYP51) complex with azole (PDB ID: 3LD6). To construct a basic framework and to build the loop regions of C. albicans cytochrome P450 lanosterol 14α -demethylase, pairwise sequence alignment and loop refinement of CA-CYP51 (P10613) and human CYP51 (3LD6_B) were performed. Three-dimensional modelled protein structures were subjected to structural quality check programs such as PRO-CHECK and its other structural validation parameters.³¹

Conflicts of interest

There are no conflicts to declare.

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Notes and references

- (a) D. Dheer, V. Singh and R. Shankar, *Bioorg. Chem.*, 2017, 71, 30; (b) A. Massarotti, S. Aprile, V. Mercalli, E. D. Grosso, G. Grosa, G. Sorba and G. C. Tron, *ChemMedChem*, 2014, 9, 2497; (c) S. G. Agalave, S. R. Maujan and V. S. Pore, *Chem.-Asian J.*, 2011, 6, 2696; (d) M. Whiting, J. Muldoon, Y. C. Lin, S. M. Silverman, W. Lindstrom, A. J. Olson, H. C. Kolb, M. G. Finn, K. B. Sharpless, J. H. Elder and V. V. Fokin, *Angew. Chem., Int. Ed.*, 2006, 45, 1435; (e) V. V. Rostovtsev, L. G. Green, V. V. Fokin and K. B. Sharpless, *Angew. Chem., Int. Ed.*, 2002, 41, 2596; (f) C. W. Tornoe, C. Christensen and M. Meldal, *J. Org. Chem.*, 2002, 67, 3057.
- 2 (*a*) K. Kempe, A. Krieg, C. R Becer and U. S. Schubert, *Chem. Soc. Rev.*, 2012, **41**, 176; (*b*) N. Akeroyd and B. Klumperman, *Eur. Polym. J.*, 2011, **47**, 1207.
- 3 (a) J. J. Bryant and U. H. F. Bunz, *Chem.-Asian J.*, 2013, 8, 1354; (b) K. D. Hanni and D. A. Leigh, *Chem. Soc. Rev.*, 2010, 39, 1240.
- 4 (a) G. C. Tron, T. Pirali, R. A. Billington, P. L. Canonico, G. Sorba and A. A. Genazzani, *Med. Res. Rev.*, 2008, 28, 278;
 (b) A. D. Moorhouse and J. E. Moses, *ChemMedChem*, 2008, 3, 715;
 (c) J. F. Lutz and Z. Zarafshani, *Adv. Drug Delivery Rev.*, 2008, 60, 958.

- 5 (a) N. Fu, S. Wang, Y. Zhang, C. Zhang, D. Yang, L. Weng, B. Zhao and L. Wang, *Eur. J. Med. Chem.*, 2017, 136, 596;
 (b) H. J. Chen, Y. J. Jiang, Y. Q. Zhangc, Q. W. Jingb, N. L. Y. Wang, W. N. Zhangb and C. Q. Shenga, *Chin. Chem. Lett.*, 2017, 28, 913; (c) Z. C. Dai, Y. F. Chen, M. Zhang, S. K. Li, T. T. Yang, L. Shen, J. X. Wang, S. S. Qian, H. L. Zhu and Y. H. Ye, *Org. Biomol. Chem.*, 2015, 13, 477; (d) Z. Jiang, J. Gu, C. Wang, S. Wang, N. Liu, Y. Jiang, G. Dong, Y. Wang, Y. Liu, J. Yao, Z. Miao, W. Zhang and C. Sheng, *Eur. J. Med. Chem.*, 2014, 82, 490; (e) N. G. Aher, V. S. Pore, N. N. Mishra, A. Kumar, P. K. Shukla, A. Sharma and M. K. Bhat, *Bioorg. Med. Chem. Lett.*, 2009, 19, 759.
- 6 (a) B. Zhang, Eur. J. Med. Chem., 2019, 168, 357; (b)
 X. L. Wang, K. Wan and C. H. Zhou, Eur. J. Med. Chem., 2010, 45, 4631.
- 7 (a) K. S. Raju, S. AnkiReddy, G. Sabitha, V. S. Krishna, D. Sriram, K. B. Reddy and S. R. Sagurthi, *Bioorg. Med. Chem. Lett.*, 2019, 29, 284; (b) S. Zhan, Z. Xu, C. Gao, Q. C. Ren, L. Chang, Z. S. Lv and L. S. Feng, *Eur. J. Med. Chem.*, 2017, 138, 501; (c) Z. Xu, X. F. Song, Y. Q. Hu, M. Qiang and Z. S. Lv, *Eur. J. Med. Chem.*, 2017, 138, 66; (d) M. H. Shaikh, D. D. Subhedar, L. Nawale, D. Sarkar, F. A. K. Khan, J. N. Sangshetti and B. B. Shingate, *Med. Chem. Commun.*, 2015, 6, 1104; (e) S. R. Patpi, L. Pulipati, P. Yogeeswari, D. Sriram, N. Jain, B. Sridhar, R. Murthy, D. T. Anjana, S. V. Kalivendi and S. Kantevar, *J. Med. Chem.*, 2012, 55, 3911.
- 8 (a) P. S. Rao, C. Kurumurthy, B. Veeraswamy, G. S. Kumar,
 Y. Poornachandra, C. G. Kumar, S. B. Vasamsetti,
 S. Kotamraju and B. Narsaiah, *Eur. J. Med. Chem.*, 2014, 80,
 184; (b) R. D. Simone, M. G. Chini, I. Bruno, R. Riccio,
 D. Mueller, O. Werz and G. Bifulco, *J. Med. Chem.*, 2011,
 54, 1565.
- 9 D. R. Buckle, D. J. Outred, C. J. M. Rockell, H. Smith and B. A. Spicer, *J. Med. Chem.*, 1983, **26**, 251.
- 10 (a) Y. Tian, Z. Liu, J. Liu, B. Huang, D. Kang, H. Zhang, E. D. Clercq, D. D. mans, C. Pannecouque, K. H. Lee, C. H. Chen, P. Zhan and X. Liu, *Eur. J. Med. Chem.*, 2018, 151, 339; (b) G. Wu, W. A. Zalloum, M. E. Meuser, L. Jing, D. Kang, C. Chen, Y. Tian, F. Zhang, S. Cocklin, K. H. Lee, X. Liu and P. Zhan, *Eur. J. Med. Chem.*, 2018, 158, 478; (c) M. J. Giffin, H. Heaslet, A. Brik, Y. C. Lin, G. Cauvi, C. H. Wong, D. E. McRee, J. H. Elder, C. D. Stout and B. E. Torbett, *J. Med. Chem.*, 2008, 51, 6263.
- 11 (a) M. Allam, A. K. D. Bhavani, A. Mudiraj, N. Ranjan, M. Thippana and P. P. Babu, Eur. J. Med. Chem., 2018, 156, 43; (b)V. Prachayasittikul, R. Pingaew, N. Anuwongcharoen, A. Worachartcheewan, C. Nantasenamat, S. Prachayasittikul, S. Ruchirawat and V. Prachayasittikul, *SpringerPlus*, 2015, 4, 571; (c) Y. C. Duan, Y. C. Ma, E. Zhang, X. J. Shi, M. M. Wang, X. W. Ye and H. M. Liu, Eur. J. Med. Chem., 2013, 62, 11; (d) Y. C. Zheng, Y. C. Duan, J. L. Ma, R. M. Xu, X. Zi, W. L. Lv, M. M. Wang, X. W. Ye, S. Zhu, D. Mobley, Y. Y. Zhu, J. W. Wang, J. F. Li, Z. R. Wang, W. Zhao and H. M. Liu, J. Med. Chem., 2013, 56, 8543.

- 12 X. Wang, Z. C. Dai, Y. F. Chen, L. L. Cao, W. Yan, S. K. Li, J. X. Wang, Z. G. Zhang and Y. H. Ye, *Eur. J. Med. Chem.*, 2017, **126**, 171.
- 13 R. Cha and J. D. Sobel, *Expert Rev. Anti-Infect. Ther.*, 2004, 2, 357.
- 14 N. H. Georgopapadakou and T. J. Walsh, *Antimicrob. Agents Chemother.*, 1996, **40**, 279.
- 15 D. A. Enoch, H. A. Ludlam and N. M. Brown, J. Med. Microbiol., 2006, 55, 809.
- 16 (a) G. I. Lepesheva, N. G. Zaitseva, W. D. Nes, et al., J. Biol. Chem., 2006, 281, 3577; (b) L. Jeu, F. J. Piacenti, A. G. Lyakhovetskiy and H. B. Fung, Clin. Ther., 2003, 25, 1321; (c) M. A. Pfaller, S. A. Messer, R. J. Hollis and R. N. Jones, Antimicrob. Agents Chemother., 2001, 45, 2862.
- 17 (a) T. L. Greaves and C. J. Drummond, *Chem. Rev.*, 2008, 108, 206; (b) R. Sheldon, *Chem. Commun.*, 2001, 2399; (c) P. Wasserscheid and W. Keim, *Angew. Chem., Int. Ed.*, 2000, 39, 3772; (d) T. Welton, *Chem. Rev.*, 1999, 99, 2071.
- 18 (a) C. Rub and B. Konig, *Green Chem.*, 2012, 14, 2969; (b)
 W. Miao and T. H. Chan, *Org. Lett.*, 2003, 5, 5003.
- 19 M. A. P. Martins, C. P. Frizzo, A. Z. Tier, D. N. Moreira, N. Zanatta and H. G. Bonacorso, *Chem. Rev.*, 2014, 114, PR1-PR70.
- 20 (a) A. A. Ali, M. Konwar, M. Chetia and D. Sarma, *Tetrahedron Lett.*, 2016, 57, 5661; (b) M. Tavassoli, A. L. Isfahani, M. Moghadam, S. Tangestaninejad, V. Mirkhani and I. M. Baltork, ACS Sustainable Chem. Eng., 2016, 4, 1454; (c) M. Dabiri, S. K. Movahed and D. I. MaGee, Res. Chem. Intermed., 2015, 41, 3335; (d) H. Singh, J. Sindhu, J. M. Khurana, C. Sharma and K. R. Aneja, Eur. J. Med. Chem., 2014, 77, 145; (e) A. Z. Ahmady, F. Heidarizadeh and M. Keshavarz, Synth. Commun., 2013, 43, 2100; (f) A. Marra, A. Vecchi, C. Chiappe, B. Melai and A. Dondoni, J. Org. Chem., 2008, 73, 2458; (g) Y. B. Zhao, Z. Y. Yan and Y. M. Liang, Tetrahedron Lett., 2006, 47, 1545.
- 21 (a) N. J. Parmar, H. A. Barad, B. R. Pansuriya and N. P. Talpada, RSC Adv., 2013, 3, 8064; (b) N. J. Parmar, B. R. Pansuriya, H. A. Barad, B. D. Parmar, R. Kant and V. K. Gupta, Monatsh. Chem., 2013, 144, 865; (c) N. J. Parmar, R. A. Patel, B. D. Parmar and N. P. Talpada, Bioorg. Med. Chem. Lett., 2013, 23, 1656; (d) P. Attri and M. Pal, Green Chem. Lett. Rev., 2010, 3, 249; (e) T. R. Sutariya, B. M. Labana, N. J. Parmar, R. Kant, V. K. Gupta, G. B. Plata and J. M. Padroón, New J. Chem., 2015, 39, 2657; (f) A. K. Verma, P. Attri, V. Chopra, R. K. Tiwari and R. Chandra, Monatsh. Chem., 2008, 139, 1041.
- 22 (a) A. B. Danne, A. S. Choudhari, S. Chakraborty, D. Sarkar,
 V. M. Khedkar and B. B. Shingate, *Med. Chem. Commun.*,
 2018, 9, 1114; (b) S. P. Khare, T. R. Deshmukh,
 J. N. Sangshetti, V. S. Krishna, D. Sriram, V. M. Khedkar and B. B. Shingate, *ChemistrySelect*, 2018, 3, 13113; (c)
 A. B. Danne, A. S. Choudhari, D. Sarkar, J. N. Sangshetti,
 V. M. Khedkar and B. B. Shingate, *Res. Chem. Intermed.*,
 2018, 44, 6283; (d) M. H. Shaikh, D. D. Subhedar,
 M. Arkile, V. M. Khedkar, N. Jadhav, D. Sarkar and

B. B. Shingate, *Bioorg. Med. Chem. Lett.*, 2016, **26**, 561; (e) M. H. Shaikh, D. D. Subhedar, B. B. Shingate, F. A. K. Khan, J. N. Sangshetti, V. M. Khedkar, L. Nawale, D. Sarkar, G. R. Navale and S. S. Shinde, *Med. Chem. Res.*, 2016, **25**, 790; (f) M. H. Shaikh, D. D. Subhedar, F. A. K. Khan, J. N. Sangshetti and B. B. Shingate, *Chin. Chem. Lett.*, 2016, **27**, 295; (g) M. H. Shaikh, D. D. Subhedar, V. M. Khedkar, P. C. Jha, F. A. K. Khan, J. N. Sangshetti and B. B. Shingate, *Chin. Chem. Lett.*, 2016, **27**, 1058.

- 23 (a) D. D. Subhedar, M. H. Shaikh, B. B. Shingate, L. Nawale, D. Sarkar, V. M. Khedkar, F. A. K. Khan and J. N. Sangshetti, Eur. J. Med. Chem., 2017, 125, 385; (b) D. D. Subhedar, M. H. Shaikh, F. A. K. Khan, J. N. Sangshetti, V. M. Khedkar and B. B. Shingate, New J. Chem., 2016, 40, 3047; (c) D. D. Subhedar, M. H. Shaikh, B. B. Shingate, L. Nawale, D. Sarkar and V. M. Khedkar, Med. Chem. Commun., 2016, 7, 1832; (d) D. D. Subhedar, M. H. Shaikh, L. Nawale, A. Yeware, D. Sarkar, F. A. K. Khan, I. N. Sangshetti and B. B. Shingate, Bioorg. Med. Chem. Lett., 2016, 26, 2278; (e) D. D. Subhedar, M. H. Shaikh, M. A. Arkile, A. Yeware, D. Sarkar and B. B. Shingate, Bioorg. Med. Chem. Lett., 2016, 26, 1704; (f) M. H. Shaikh, D. D. Subhedar, F. A. Kalam Khan, J. N. Sangshetti and B. B. Shingate, Res. Chem. Intermed., 2016, 42, 5115; (g) D. D. Subhedar, M. H. Shaikh, L. Nawale, A. Yeware, D. Sarkar and B. B. Shingate, Res. Chem. Intermed., 2016, 42, 6607.
- 24 (a) S. T. Dhumal, A. R. Deshmukh, K. R. Kharat, B. R. Sathe,
 S. S. Chavan and R. A. Mane, New J. Chem., 2019, 43, 7663; (b)
 C. P. Kaushik, R. Luxmi, M. Kumar, D. Singh, K. Kumar and
 A. Pahwa, Synth. Commun., 2019, 49, 118; (c)
 G. Wang,
 Z. Peng, J. Wang, X. Li and J. Li, Eur. J. Med. Chem., 2017,
 125, 423; (d) P. Neeraja, S. Srinivas, K. Mukkanti,
 P. K. Dubey and S. Pal, Bioorg. Med. Chem. Lett., 2016, 26,
 5212; (e) G. Wang, Z. Peng, J. Wang, J. Li and X. Li, Bioorg.
 Med. Chem. Lett., 2016, 26, 5719.
- 25 C. Ferroni, A. Pepe, Y. Sang Kim, S. Lee, A. Guerrini, M. D. Parenti, A. Tesei, A. Zamagni, M. Cortesi, N. Zaffaroni, M. De Cesare, G. L. Beretta, J. B. Trepel, S. V. Malhotra and G. Varchi, *J. Med. Chem.*, 2017, **60**, 3082.
- 26 (a) K. Khan, In vitro and In vivo Screening Techniques for Bioactivity Screening and Evaluation, *Proc. Int. Workshop UNIDO-CDRI*, 1997, p. 210; (b) A. H. Collins, *Microbiological Methods*, Butterworth, London, 2nd edn, 1976.
- 27 M. Burits and F. Bucar, Phytother. Res., 2000, 14, 323.
- 28 S. Li, D. Li, T. Xiao, S. Zhang, Z. Song and H. Ma, *J. Agric. Food Chem.*, 2016, **23**, 8927.
- 29 V. S Dofe, A. P. Sarkate, D. K. Lokwani, S. H. Kathwate and C. H. Gill, *Res. Chem. Intermed.*, 2017, **43**, 15.
- 30 S. G. Franzblau, R. S. Witzig and J. C. McLaughlin, J. Clin. Microbiol., 1998, 36, 362.
- 31 (a) J. N. Sangshetti, F. A. Kalam Khan, R. S. Chouthe, M. G. Damale and D. B. Shinde, *Chin. Chem. Lett.*, 2014, 25, 1033; (b) R. W. Hooft, G. Vriend, C. Sander and E. E. Abola, *Nature*, 1996, 381, 272.