






Cite this: *Chem. Commun.*, 2020, 56, 5747

Received 22nd March 2020,
Accepted 13th April 2020

DOI: 10.1039/d0cc02120b

rsc.li/chemcomm

Dynamic modulation of proximity-induced enzyme activity using supramolecular polymers†

Eva Magdalena Estirado,‡^a Bas J. H. M. Rosier, ^{‡a} Tom F. A. de Greef ^{abc} and Luc Brunsveld ^{*a}

Synthetic supramolecular polymers are used as dynamic nanoscaffolds for the activation of the apoptotic signalling enzyme caspase-9. Recruitment of caspase-9 to the nanoscaffold results in an increase in enzymatic activity due to enhanced proximity, with a bell-shaped response as a function of nanoscaffold concentration. The modularity of the system allows for dynamic regulation of enzyme activity through variation of the recruitment-motif density along the supramolecular polymer.

Spatial and temporal coordination of biomolecular components are of key importance for efficient intracellular communication and faithful signal transduction.^{1,2} Stable organization of functionally related components occurs on the microscale through vesicle-based compartmentalization,³ or on the nanoscale using dedicated scaffold proteins that assemble enzymes into signalling cascades.⁴ Transient organization of components based on non-covalent interactions allows the living cell to finely tune regulatory behaviours with examples including rapidly exchanging membraneless organelles,^{5,6} open-ended receptor clustering on lipid membranes,^{7,8} and dynamic rearrangement of cytoskeletal polymers.⁹

Synthetic supramolecular polymers are similarly based on non-covalent interactions and could therefore be used to mimic specific features of transient cellular organization in a controlled setting.^{10–12} Examples include the assembly of quasi-one-dimensional fibrillar aggregates with tunable kinetic characteristics, hierarchical assemblies, and branched networks such as hydrogels and coacervates.^{13–15} In addition, supramolecular

polymers can be used as modular templates for the assembly of various biomolecules, including nanoparticles,^{16,17} nucleic acids,^{18,19} and proteins.^{20–23}

Recent work has revealed that intracellular signalling pathways heavily rely on the dynamic assembly of higher-order fibril-like aggregates that are structurally well-defined but open-ended and sometimes quasi-infinite.^{24,25} As such, these assemblies function as platforms for the transient recruitment of signalling proteins that undergo proximity-induced activation,^{26,27} and subsequently elicit immune or inflammatory responses, or apoptosis. The nature of the non-covalent interactions involved in these assemblies allows for nucleated growth resulting in signal amplification,²⁶ the appearance of threshold concentrations and cooperative incorporation of other enzymes.²⁸ Pathway inhibition can be dynamically regulated by co-assembly of non-functional fiber monomers.²⁹ Bottom-up construction of synthetic systems mimicking these natural systems has remained very difficult, especially because of the combined need for synthetically accessible systems with physical chemical properties, regarding assembly and dynamics, matching the biological systems at hand.

Here, we present a supramolecular polymeric nanoscaffold as a synthetic model system for the dynamic recruitment, organization and activation of the signalling enzyme caspase-9 (cysteine-dependent aspartic protease) as model system. The synthetic recruitment nanoscaffold consists of amphiphilic monomers made of bis-pyridine-based C₃-symmetrical molecules decorated with single-stranded DNA handles (**DNA-Disc**, Fig. 1a).¹⁹ The hybrid DNA-decorated monomers self-assemble in water into columnar assemblies, displaying multiple copies of DNA recruitment handles (Fig. 1b).³⁰ The handles are accessible for interacting through DNA-duplex formation with complementary DNA anti-handle strands located at the enzyme of interest, in our case caspase-9 (Fig. 1b). Full-length caspase-9 consists of a catalytic domain responsible for its protease activity, and a caspase recruitment domain (CARD) that, by interacting with the apoptosome, brings multiple caspases together to initiate proximity-induced activation.^{31–33} In the present system,

^a Laboratory of Chemical Biology and Institute for Complex Molecular Systems, Eindhoven University of Technology, P.O. Box 513, 5600 MB Eindhoven, The Netherlands. E-mail: L.brunsveld@tue.nl

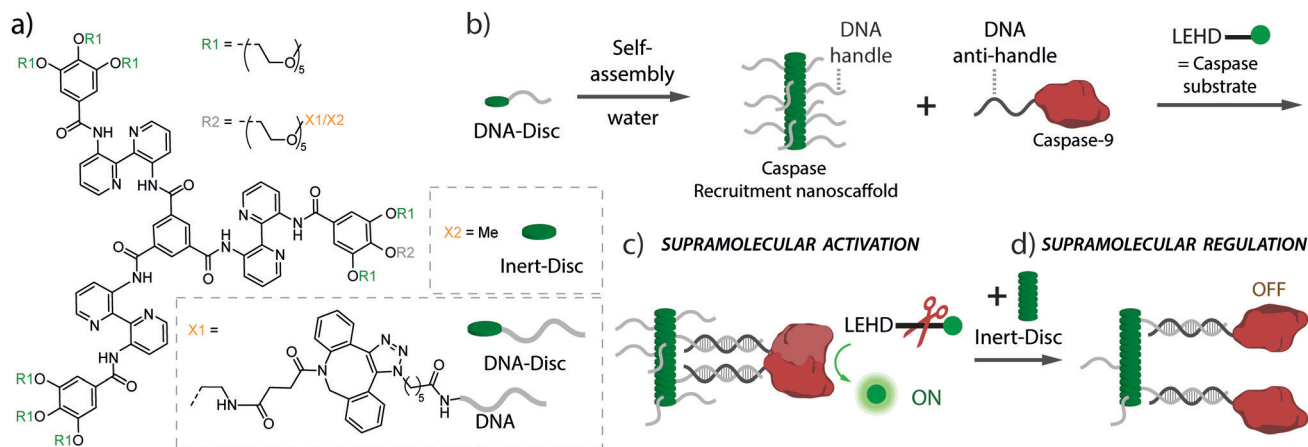
^b Computational Biology Group, Department of Biomedical Engineering, Eindhoven University of Technology, P.O. Box 513, 5600 MB Eindhoven, The Netherlands

^c Institute for Molecules and Materials, Radboud University, Nijmegen, The Netherlands

† Electronic supplementary information (ESI) available: Experimental details and supporting figures. See DOI: 10.1039/d0cc02120b

‡ These authors contributed equally.





the CARD domain has been replaced by a DNA strand, that recruits caspase-9 onto the nanoscaffold through DNA-duplex formation.³⁴ The catalytic domain was expressed with an N-terminal non-canonical amino acid (*p*-azidophenylalanine), introduced by using amber codon suppression in *E. coli*, in order to site-specifically decorate the protein with the DNA anti-handle (ESI[†]).³⁴ The DNA contains a 5'-end nucleotide linker between the enzyme and the recruitment recognition sequence (anti-handle) in order to avoid undesired interactions with the nanoscaffold. A detailed quantitative analysis has previously demonstrated that the kinetic characteristics of the caspase-9 enzyme-DNA conjugate is similar to wildtype caspase-9.³⁴

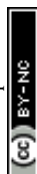
The modularity and dynamic nature of monomer intermixing in the supramolecular nanoscaffold allows straightforward tuning of the nanoscaffold characteristics, such as the systematic variation of the recruiter handle density in the nanoscaffold (Fig. 1c and d), which potentially regulates enzymatic activity through the variation of inter-enzyme distances, as seen in natural systems.

First, the ability of the supramolecular nanoscaffold to recruit caspase-9 was characterized. A series of control experiments were carried out to rule out possible unspecific interactions between the components in the system. A constant concentration of caspase-9 (25 nM, well below its dimerization affinity³⁴) was incubated with non-DNA-functionalized monomers (**Inert-Disc**) at different concentrations (Fig. S1, ESI[†]). Upon the addition of Ac-LEHD-AFC substrate, the fluorescence intensity of the samples was monitored over time and the enzymatic activity was calculated using the initial slope of the fluorescence curves (monitored between the first 20 to 60 minutes after addition of the fluorogenic substrate, unless stated otherwise). The enzymatic activity (Fig. S1, ESI[†]) exhibited similar background levels when the experiment was performed in the presence and absence of the **Inert-Disc**, indicating the lack of interference

between the supramolecular nanoscaffold and the substrate or caspase-9.

Next, the active assembly to the supramolecular nanoscaffold and resultant proximity-induced caspase-9 activation was analysed. Therefore, nanoscaffolds were assembled using a series of concentrations of **DNA-Disc** ranging from 0 to 500 nM, and subsequently incubating with caspase-9 for 2 hours at 4 °C, after which substrate was added to the sample. Fluorescence intensity traces at the different **DNA-Disc** concentrations are shown in Fig. S2 (ESI[†]), revealing that the presence of supramolecular recruitment nanoscaffold effectively enhances proteolytic cleavage, in comparison with the background activity of non-scaffolded enzymes. The enzymatic activities derived from the intensity traces for the entire concentration range (see Fig. 2a) showed a step-wise enhancement in enzyme activity with increasing concentration of **DNA-Disc**. Optimal enzyme activity was reached at equimolar amounts of both caspase-9 and **DNA-Disc**. Importantly, the nearly 5-fold increase in enzyme activity seen at 20 nM of **DNA-Disc** agrees with the enhancement seen in previous studies where caspase-9 was dimerized on a DNA origami-based nanoscaffold at roughly the same caspase-9 concentration,³⁴ or with other *in vitro* systems that typically do not reach the activity of apoptosome activated caspase-9.^{31–33,35}

A further increase in **DNA-Disc** concentration, beyond the optimum, diminished enzyme activity (Fig. 2a), which can be attributed to combinatorial inhibition.^{36,37} That is, supramolecular levels of DNA-decorated supramolecular monomers promote sequestration of caspase-9 on the nanoscaffold in the inactive monomeric state, and therefore reduce proximity-induced formation of enzymatically active dimers. Note that full suppression of the activity to background levels was not observed, even at high **DNA-Disc** concentrations (500 nM). The intrinsic



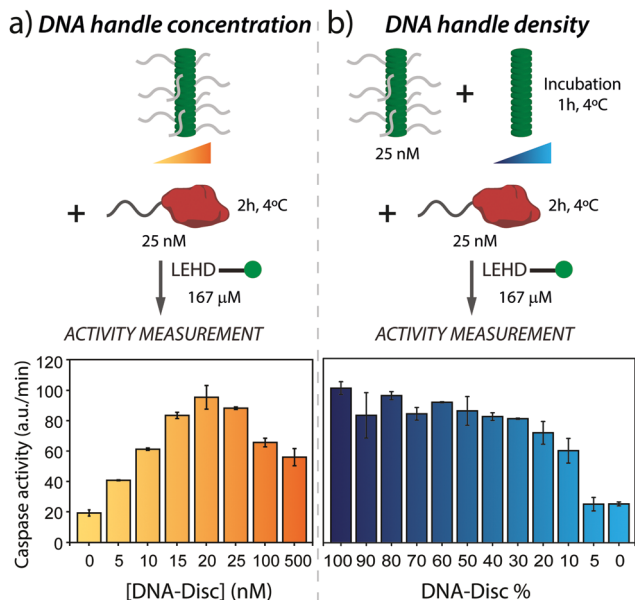


Fig. 2 (a) Effect of nanoscaffold concentration on the enzymatic activity of caspase-9. Enzymatic activity increases with **DNA-Disc** concentration up to a 5-fold at equimolar concentrations of enzyme and **DNA-Disc**. At high nanoscaffold densities, combinatorial inhibition leads to a decrease in enzymatic activity. Experiments were conducted with supramolecular nanoscaffolds containing 100% **DNA-Disc**. In the cartoon, DNA handles were partially omitted for clarity. (b) Effect on caspase-9 activity of the variation in DNA handle density displayed by the nanoscaffold. Varying DNA handle densities is achieved by keeping the total concentration of **DNA-Disc** at 25 nM, and adding varying amounts of **Inert-Disc**, resulting in nanoscaffolds in which the DNA handle density varies between 100 and 0%. In both graphs, the enzyme activity is based on the initial slope of the kinetic traces (Fig. S2 and S3, ESI†). The error bars represent the standard deviation of the measurements performed in triplicates.

affinity between caspase-9 monomers (K_D of dimerization of around 50 μM)³⁸ in combination with a high effective concentration on the nanoscaffold could lead to a stabilization of the active caspase-9 dimer complex and therefore explain the enhanced enzymatic activity at high **DNA-Disc** concentrations.

Next, we evaluated the effect of changing the DNA handle density on the nanoscaffold on enzymatic activity. This can be achieved in a straightforward fashion by co-assembly of **Inert-Disc** and **DNA-Disc**, which results in homogeneous intermixing of both monomers along the supramolecular polymer, as previously shown.¹⁹ To this end, **DNA-Disc** at a constant concentration of 25 nM was co-assembled with varying concentrations of **Inert-Disc** (from 0 to 500 nM), leading to nanoscaffolds with handle densities ranging from 100 to 5% (Fig. 2b). The samples were incubated for one hour to ensure full co-assembly, followed by incubation with 25 nM of caspase-9 at 4 °C for 2 hours. After addition of enzyme substrate, the fluorescence was monitored over time (Fig. S3, ESI†) and the activity was determined. The control experiment with non-scaffolded caspase-9 showed typical values for background activity (0% **DNA-Disc**, Fig. 2b).

The samples featuring DNA handles at different densities showed a non-linear variation of enzymatic activity. A decrease

from 100 to 50% in handle density did not have a significant effect on activity, however, activity gradually diminished between 50 to 10%, and dropped sharply to background activity levels after 10% (Fig. 2b). To gain insights into these results, it is useful to estimate the expected distance between DNA handles for a given DNA handle density. The distance between the monomer cores forming the supramolecular nanoscaffold has been estimated to be 0.34 nm.³⁹ Assuming homogeneous co-assembly of **Inert-Disc** and **DNA-Disc**, the distance between DNA handles is ≤ 3.4 nm for nanoscaffolds containing $\geq 10\%$ **DNA-Disc**. Given previous work that demonstrated markedly enhanced activity at a DNA separation of 6 nm,³⁴ we conclude that the handle density is sufficiently high to enable efficient caspase-9 dimerization across the range of nanoscaffolds containing $\geq 20\%$ **DNA-Disc**, explaining the constant enzyme activity in the first part of the graph in Fig. 2b. In nanoscaffolds with 5% **DNA-Disc** the average inter-handle distance is 6.8 nm, which does not result in enzymatic activity. An explanation can be that, compared to the DNA origami-based system which employs a fixed two-dimensional scaffold, the columnar supramolecular nanoscaffold displays DNA handles in three dimensions, and therefore might lower effective caspase-9 dimerization at this **DNA-Disc** concentration. These experiments demonstrate that co-assembly with high concentrations of **Inert-Disc** affect enzymatic activity by lowering the DNA handle density on the supramolecular nanoscaffold.

Finally, we explored dynamic regulation of enzymatic activity by addition of **Inert-Disc** after equilibration of DNA-functionalized nanoscaffolds. The enzyme activity of an equimolar solution at 25 nM of **DNA-Disc** and caspase-9 was followed over time in the presence of substrate (Fig. 3a). After approximately 60 minutes, a solution of **Inert-Disc** was added to a final concentration of 10 μM. Note that the instant increase of the fluorescence for both the non-scaffolded and scaffolded caspase-9 (Fig. 3a, green and orange curves) is a consequence of the characteristic absorption peak at 350 nm and emission peak at 520 nm of the nanoscaffold in the aggregated state, which does not affect the slope of the curve (enzyme activity). The histogram in Fig. 3b represents the enzymatic activity extracted from Fig. 3a before and after the addition of the **Inert-Disc**. The addition of **Inert-Disc** to the non-scaffolded caspase-9 showed a negligible effect on the enzyme activity. In contrast, the scaffolded version displayed a decrease in enzyme activity of roughly 35%. The intrinsic affinity between caspase-9 monomers could explain the residual enzymatic activity after the addition of **Inert-Disc**, similar to the observations in Fig. 2. These results indicate the potential of dynamic supramolecular polymers to regulate enzymatic activity by simple addition of a non-functional monomer, which results in an increased distance between the individual enzymes and a concomitant decrease in proximity-induced dimerization.

In this communication, we mimic proximity-induced activation of enzymes with a fully synthetic and programmable nanoscaffold, making use of DNA-decorated supramolecular polymers and engineered signalling proteins (caspase-9). Tethering of enzymes on the nanoscaffold *via* hybridization between their complementary DNA handle strands resulted in their activation due to proximity-induced dimerization. The extent of enzyme



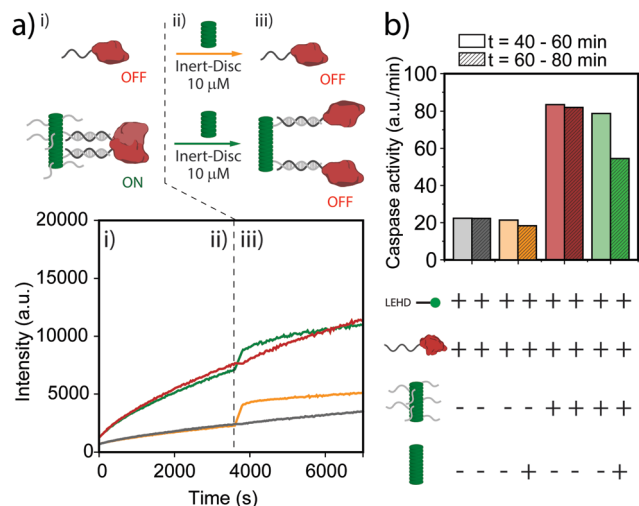


Fig. 3 Supramolecular regulation of caspase activity. (a) Schematic representation of the supramolecular regulation of caspase activity through the segregation of the enzymes along the supramolecular nanoscaffolds by addition of **Inert-Disc**. First, the supramolecular nanoscaffold (**DNA-Disc** at 25 nM) was incubated with equimolar amounts of caspase-9 for 2 hours at 4 °C. (i) After equilibration, 167 μM of Ac-LEHD-AFC substrate was added to the mixture and the fluorescence was recorded over time. The increasing fluorescence upon substrate cleavage was measured every 30 seconds, exciting the sample at $\lambda_{\text{ex}} = 400$ nm, and recording at $\lambda_{\text{em}} = 505$ nm. (ii) Approximately after one hour, **Inert-Disc** was added to the sample reaching a final concentration of 10 μM. (iii) The addition of **Inert-Disc** enlarged the distance between enzymes regulating their activity via the supramolecular element (for color coding see (b)). (b) Enzyme activity extracted from (a). The slopes are determined from the 20 minutes before (plain columns) and after addition (dashed columns) of **Inert-Disc**. The presence of the components for each experiment is represented with a (+).

activity could be modulated by varying the concentration of DNA-functionalized monomers, resulting in a bell-shaped activity response with a 500% increase in activity at optimal equimolar concentrations of enzyme and supramolecular monomer. Moreover, the enzymatic activity could be inhibited by decreasing the average density of DNA handles displayed on the nanoscaffold, achieved by co-assembly of non-functionalized monomers either *a priori* or dynamically after equilibration.

Taken together, these results provide insights into the potential of the implementation of supramolecular polymers as general recruitment nanoscaffolds for the distance-dependent activation of signalling proteins. The modularity of the supramolecular polymer and the DNA element, together with their befitting assembly characteristics, opens many possibilities for the recruitment of other proteins of interest and the development of programmable enzyme assembling systems.

This work was supported by The Netherlands Organization for Scientific Research (NWO) through Gravity Program 024.001.035, and through the European Union's program Horizon 2020 Marie Curie Innovative Training Network MULTI APP (2015-2019).

Conflicts of interest

There are no conflicts of interest to declare.

Notes and references

- 1 R. P. Bhattacharyya, A. Reményi, B. J. Yeh and W. A. Lim, *Annu. Rev. Biochem.*, 2006, **75**, 655–680.
- 2 B. N. Kholodenko, J. F. Hancock and W. Kolch, *Nat. Rev. Mol. Cell Biol.*, 2010, **11**, 414–426.
- 3 A. H. Chen and P. A. Silver, *Trends Cell Biol.*, 2012, **22**, 662–670.
- 4 M. C. Good, J. G. Zalatan and W. A. Lim, *Science*, 2011, **332**, 680–686.
- 5 S. F. Banani, H. O. Lee, A. A. Hyman and M. K. Rosen, *Nat. Rev. Mol. Cell Biol.*, 2017, **18**, 285–298.
- 6 Y. Shin and C. P. Brangwynne, *Science*, 2017, **357**, eaaf4382.
- 7 N. C. Hartman and J. T. Groves, *Curr. Opin. Cell Biol.*, 2011, **23**, 370–376.
- 8 J. B. Casaletto and A. I. McClatchey, *Nat. Rev. Cancer*, 2012, **12**, 387–400.
- 9 D. A. Fletcher and R. D. Mullins, *Nature*, 2010, **463**, 485–492.
- 10 K. Petkau-Milroy, M. H. Sonntag and L. Brunsveld, *Chem. – Eur. J.*, 2013, **19**, 10786–10793.
- 11 R. Freeman, J. Boekhoven, M. B. Dickerson, R. R. Naik and S. I. Stupp, *MRS Bull.*, 2015, **40**, 1089–1100.
- 12 O. J. G. M. Goor, S. I. S. Hendrikse, P. Y. W. Dankers and E. W. Meijer, *Chem. Soc. Rev.*, 2017, **46**, 6621–6637.
- 13 S. Van Dun, C. Ottmann, L. G. Milroy and L. Brunsveld, *J. Am. Chem. Soc.*, 2017, **139**, 13960–13968.
- 14 C. Facciotti, V. Saggiomo, A. Bunschoten, R. Fokkink, J. B. Ten Hove, J. Wang and A. H. Velders, *Soft Matter*, 2018, **14**, 9542–9549.
- 15 S. Hirayama, K. Oohora, T. Uchihashi and T. Hayashi, *J. Am. Chem. Soc.*, 2020, **142**, 1822–1831.
- 16 S. H. Jung, J. Jeon, H. Kim, J. Jaworski and J. H. Jung, *J. Am. Chem. Soc.*, 2014, **136**, 6446–6452.
- 17 Y. Vyborna, M. Vybornyi and R. Häner, *Chem. Commun.*, 2017, **53**, 5179–5181.
- 18 S. P. W. Wijnands, E. W. Meijer and M. Merks, *Bioconjugate Chem.*, 2019, **30**, 1905–1914.
- 19 M. Á. Alemán García, E. Magdalena Estirado, L. G. Milroy and L. Brunsveld, *Angew. Chem., Int. Ed.*, 2018, **57**, 4976–4980.
- 20 Q. Luo, Z. Dong, C. Hou and J. Liu, *Chem. Commun.*, 2014, **50**, 9997–10007.
- 21 M. K. Müller, K. Petkau and L. Brunsveld, *Chem. Commun.*, 2011, **47**, 310–312.
- 22 K. Petkau-Milroy, D. A. Uhlenheuer, A. J. H. Spiering, J. A. J. M. Vekemans and L. Brunsveld, *Chem. Sci.*, 2013, **4**, 2886–2891.
- 23 S. P. W. Wijnands, W. Engelen, R. P. M. Lafleur, E. W. Meijer and M. Merks, *Nat. Commun.*, 2018, **9**, 65.
- 24 J. C. Kagan, V. G. Magupalli and H. Wu, *Nat. Rev. Immunol.*, 2014, **14**, 821–826.
- 25 M. Bienz, *Trends Biochem. Sci.*, 2014, **39**, 487–495.
- 26 Q. Qiao, C. Yang, C. Zheng, L. Fontán, L. David, X. Yu, C. Bracken, M. Rosen, A. Melnick, E. H. Egelman and H. Wu, *Mol. Cell*, 2013, **51**, 766–779.
- 27 Y. Tan and J. C. Kagan, *Cell*, 2019, **177**, 384–398.e11.
- 28 L. David, Y. Li, J. Ma, E. Garner, X. Zhang and H. Wu, *Proc. Natl. Acad. Sci. U. S. A.*, 2018, **115**, 1499–1504.
- 29 T.-M. Fu, Y. Li, A. Lu, Z. Li, P. R. Vajjhala, A. C. Cruz, D. B. Srivastava, F. DiMaio, P. A. Penczek, R. M. Siegel, K. J. Stacey, E. H. Egelman and H. Wu, *Mol. Cell*, 2016, **64**, 236–250.
- 30 E. Magdalena Estirado, M. A. Alemán García, J. Schill and L. Brunsveld, *J. Am. Chem. Soc.*, 2019, **141**, 18030–18037.
- 31 M. Renatus, H. R. Stennicke, F. L. Scott, R. C. Liddington and G. S. Salvesen, *Proc. Natl. Acad. Sci. U. S. A.*, 2001, **98**, 14250–14255.
- 32 Y. Chao, E. N. Shiozaki, S. M. Srinivasula, D. J. Rigotti, R. Fairman and Y. Shi, *PLoS Biol.*, 2005, **3**, e183.
- 33 Y. Li, M. Zhou, Q. Hu, X. Bai, W. Huang, S. H. W. Scheres and Y. Shi, *Proc. Natl. Acad. Sci. U. S. A.*, 2017, **114**, 1542–1547.
- 34 B. J. H. M. Rosier, A. J. Markvoort, B. Gumí Audenis, J. A. L. Roodhuizen, A. den Hamer, L. Brunsveld and T. F. A. de Greef, *Nat. Catal.*, 2020, **3**, 295–306.
- 35 D. T. Dang, H. D. Nguyen, M. Merks and L. Brunsveld, *Angew. Chem., Int. Ed.*, 2013, **52**, 2915–2919.
- 36 A. Levchenko, J. Bruck and P. W. Sternberg, *Proc. Natl. Acad. Sci. U. S. A.*, 2000, **97**, 5818–5823.
- 37 J. J. McCann, U. B. Choi and M. E. Bowen, *Structure*, 2014, **22**, 1458–1466.
- 38 S. J. Riedl and G. S. Salvesen, *Nat. Rev. Mol. Cell Biol.*, 2007, **8**, 405–413.
- 39 T. Metzroth, A. Hoffmann, R. Martín-Rapún, M. M. J. Smulders, K. Pieterse, A. R. A. Palmans, J. A. J. M. Vekemans, E. W. Meijer, H. W. Spiess and J. Gauss, *Chem. Sci.*, 2011, **2**, 69–76.

