RSC Advances

PAPER

Cite this: RSC Adv., 2020, 10, 19454

Oxidative radical coupling of hydroquinones and thiols using chromic acid: one-pot synthesis of quinonyl alkyl/aryl thioethers†

T. P. Adarsh Krishna, D [Sa](http://orcid.org/0000-0003-2703-8562)kthivel Pandaram[,](http://orcid.org/0000-0003-2042-9624) D Suresh Chinnasamy and Andivelu Ilangovan \mathbb{D}^*

An efficient, simple and practical protocol for one-pot sequential oxidative radical C–H/S–H crosscoupling of thiols with hydroquinones (HQs) and oxidation leading to the formation of quinonyl alkyl/aryl thioethers using H_2CrO_4 was developed. This cross-coupling of thiyl and aryl radicals offers mono thioethers in good to moderate yield and works well with a wide variety of thiols. Similarly, this method works well for coupling of 2-amino thiophenol and HQs to form phenothiazine-3-ones 5a–c. C–S bond formation via thioether synthesis was observed using a chromium reagent for the first time. Theoretical studies on the pharmacokinetic properties of compounds $5a-c$ revealed that due to drug-like properties, compound 5b strongly binds with Alzheimer's disease (AD) associated AChE target sites. **PAPER**
 EXERCT CONSULTER SET CONSULTER AND 2000 CON

Received 17th February 2020 Accepted 11th May 2020

DOI: 10.1039/d0ra01519a

rsc.li/rsc-advances

Introduction

The quinonoid class of compounds finds applications in several fields such as synthetic chemistry,¹ medicinal chemistry,² natural products,³ and functional materials.⁴ The chemical reactivity and catalytic performance of quinones depends upon their electron-accepting character, redox behavior,⁵ and the nature of functional group(s) present.⁶ Hetero-functionalized quinones, especially quinonyl thioethers, display good structural stability,⁷ and useful pharmaceutical⁸ and other biological activities.⁹ Around thirty-one currently marketed drugs are thioether derivatives (Fig. 1),¹⁰ used for the treatment of cancer, HIV, diabetes, Alzheimer's, inflammatory diseases, etc. In the case of materials, the thioether group contributes to physical, electronic and surface properties.¹¹

In general, number of methods known for $C-S¹²$ bond formation via C–H functionalization only less compared to C–N or C–O bond formation. One of the potential reasons could be lack of transition metal catalysts and reagents that can tolerate sulfur poisoning. Classical approaches for the construction of C–S bond are the direct coupling of organic halides or acids with thiols/disulfides/arylsulfonyl halides and addition of thiols to unsaturated C–C bonds under free radical or metal (Mn, Fe, Ni, Cu, Ru, Rh, Pd, Ag and Cs) catalyzed conditions.¹²

A good number of methods are known for the introduction of the thioether group on quinones (Q–S–R). Aryl quinonyl thioethers (Q–S–R) were prepared by reaction of arylsulfonyl

chlorides with quinone-CuI-PPh₃,^{13a} aryl disulfides (R-S-S-R) with quinone-AgOAc-dpp^{13b} and CuI catalyzed oxidative addition of aryl boronic acids,^{14a} and aryl iodides with aryl thiols.^{14b} We recently demonstrated I_2 -DMSO¹⁵ mediated thiomethylation (–SMe) of quinone. Moreover, the conjugate addition of thiol on quinone was demonstrated using CuI–O₂,¹⁶ and Co(OAc)₂-O₂ (ref. 7) system.

Conjugate addition of thiols or its derivatives to quinones,¹⁷ usually leads to a mixture of thiolated hydroquinones (HQs).¹⁸ This followed by re-oxidation provides quinonyl thioethers. A popular method for synthesis of quinone is the oxidation of HQ. In direct conversion of HQs into quinonyl thioethers, two such transformations should take place in one pot. To the best of our knowledge, there are only two such reports known. In one case, carcinogenic S-alkylisothiouronium salt $19a$ was used for thioalkylation and in another study, an enzyme laccase was used to catalyse thiol addition on naphthohydroquinone^{19b} and $HQ^{19c,d}$ (Scheme 1). Formation of poly thioalkylated products and poor yield are the main drawbacks of the enzymatic reaction. C–S bond formation on HQ via oxidative radical cross-coupling (ORCC), to form quinonyl thioethers was seldom explored.

Fig. 1 Bioactive molecules containing a thioether linkage.

School of Chemistry, Bharathidasan University, Tiruchirappalli, Tamilnadu-620024, India. E-mail: ilangovanbdu@yahoo.com

[†] Electronic supplementary information (ESI) available: Experimental, spectral data and copies of spectra. See DOI: 10.1039/d0ra01519a

Chromium reagents are powerful oxidants which finds application in polymerization, aldol and Diels–Alder reactions.^{20a} However, compared to other transition metals, its application in organic synthesis is undeveloped. To avoid poisonous effects due to contamination, chromium reagents are immobilized on zeolite.^{20b} It is well known that chromic acid formed from $CrO₃$ dimerizes thiol oxidatively to form disulfides through thio radical,^{20c} and also oxidizes hydroquinone to quinone.²⁰^d Based on this and in continuation of our research interest on C–H functionalization of quinones, $15,21$ we hypothesized that HQ may undergo cross dehydrogenative oxidative radical cross-coupling (CDORCC) with thiol in the presence of CrO3. CDORCC is highly advantageous because it is atom economic,²² no need for pre-functionalization, and could be used for C–S bond forming reaction.²³ Under the present study, we observed the formation of quiononyl thioether in the presence of in situ generated chromic acid from $CrO₃$. To the best of our knowledge, this is the first example of C–S bond-forming reaction observed using a chromium reagent. Herein we present the results.

Result and discussion

To check our hypothesis, a solution of HQ (1a, 1.0 equiv.) and ethyl 3-mercaptopropanoate (2a, 1.5 equiv.) in DCM : $H_2O(1:1)$ v/v, 4 mL) was treated with different oxidizing agents (1.0 equiv.) and the results are summarized in Table 1. In the case of oxidants such as $K MnO₄$, NBS, Na₂O₂ (entry 1) and iodine reagents like PIDA, PIFA, IBX or NaIO₄ (entry 2) the reaction failed to deliver the expected product 3a. Interestingly, with $K_2S_2O_8$ (1.0 equiv., entry 3) both oxidation of HQ to quinone as well as C–S bond-forming reaction took place to afford the product 3a in 10% isolated yield. Based on this encouraging result, we tried the same reaction using $(NH_4)_2S_2O_8$ (1.0 equiv., entry 4) and $CrO₃$ (1.0 equiv., entry 5), to get product 3a in 20% and 30% respectively. Further, when 3.0 equiv. of $CrO₃$ (entry 5) was used the yield improved to 75%. However, with higher equivalents (3.0 equiv.) of $K_2S_2O_8$ (35%) and $(NH_4)_2S_2O_8$ (48%) the product 3a was obtained in less isolated yield as compared with $CrO₃$ (entries 3–5). By decreasing (2.0 equiv., entry 6) or increasing (4.0 equiv., entry 6) the quantity of $CrO₃$ the yield did

 a Reaction conditions: hydroquinone ${\bf 1a}$ (1.0 mmol), thiol ${\bf 2a}$ (1.5 mmol) oxidant (1 equiv.), additive (1 equiv.), solvent 1 : 1 ratio (4.0 mL) at rt for 1 h. b Isolate yield. c Oxidant (3.0 equiv.). d Oxidant (2.0 equiv.). c Oxidant (4.0 equiv.). f CrO₃ (0.1 equiv.) + (NH₄)₂S₂O₈ (3.0 equiv.); NR $=$ no reaction.

not improve. Additionally, when the reaction was carried out with only DCM (no reaction) or $H₂O$ (12%, entry 7), the result was not encouraging. This proves that solvent combinations are more favorable and water is essential for this reaction. Further, when the reaction was carried out in a combination of $H₂O$ with various solvents such as DCE, DMF and ACN (Table 1, entries 8– 10) the reaction yield diminished signicantly. Finally, the effects of different acids as additive was evaluated.

When TFA $(25\%$, entry 11) or p-TSA $(38\%$, entry 12) was used the product 3a was obtained only in very less yield. Furthermore, when HCl and $H₂SO₄$ were used as an additive, hydroquinone $(1a)$ was simply converted into *p*-bezoquinone $(6a)$ (entry 13). Finally, a catalytic amount of $CrO₃$ (0.1 equiv.) with $(NH_4)_2S_2O_8(3.0$ equiv.) reaction was performed (entry 14), under this condition only 52% of desired product was obtained. The systematic screening study revealed that 1a (1.0 equiv.), 2a (1.5 equiv.) and CrO₃ (3.0 equiv.) in DCM : H₂O (1 : 1 v/v, 4 mL) at room temperature could be the optimum condition for oxidative direct C–S bond formation and synthesis of quinonyl thioethers from hydroquinone.

With the optimum condition in hand, substrate scope of various alkyl and aryl thiols was examined under the optimized reaction condition and the results are depicted in Scheme 2. The reaction of a series of linear long-chain (C1–C5) mercaptopropanoates (2b–2e) with HQ afforded corresponding thioethers 3b– 3e in good to moderate yield (75% to 48%). As the chain length of thioesters increased, the reactivity decreased. Mercaptopropanoate (2f) containing isopropyl group provided product 3f in best isolated yield (85%). Similarly, mercaptopropanoates 2g–2h,

Scheme 2 Scope of C-H/S-H cross-coupling between HQ and thiols.^{a,b a}Reaction conditions: hydroquinone (1.0 equiv., 100 mg), thiol $(1.5 \text{ equiv.}), \text{CrO}_3$ (3.0 equiv.), DCM : H₂O (4 mL, 1 : 1) at rt, ^bisolated .
yield; cexpected product.

containing substituents such as 2-methoxyethyl (2g), benzyl (2h) and 3-phenylpropyl (2i) esters provided corresponding products 3g–3i in 1 h in good to moderate (82–40%). As noted earlier, with an increase in chain length, the yield decreased. In the case of free acids, such as 3-mercaptopropanonic acid (2j), amino-3 sulfhydrylpropanoic acid $(2k)$ the reaction failed to deliver the expected product 3j and 3k. Furthermore, simple thiols like butanethiol (2l) containing no ester group also reacted quickly with HQ to offer product 3l, albeit in moderate yield (45%).

As the next part of our study, the suitability of various aryl thiols was examined. The unsubstituted thiophenol (2m) gave aryl thioether 4a in a very good yield (85%). Similarly, thiophenols bearing electron-donating groups such as 4-Me (2n) and 4-OMe (2o) showed good reactivity and yielded corresponding aryl thioethers 4b (80%) and 4c (78%) in very good yield. The thiophenol 2p containing electron-withdrawing 4- NO2 group provided product 4d in moderate yield 70%. However, thiophenols 2q–s, substituted with deactivating

substituents like 4-Cl, 4-Br, 4-F gave low to the moderate yield of desired products 4e–g (40–43%). However, 2-mercaptopyridine (2t) failed to deliver the expected product 4h. Further, the reaction of substituted hydroquinones with 4-methylthiophenol $(2n)$ was examined. The 2-methoxy HQ $(1b)$ and 2-acelyl HQ $(1c)$ underwent reaction very smoothly to afford desired products in 68% (4i) and 44% (4j) respectively. HQ 1d containing deactivating –COOH group and HQ 1e containing highly substituted, 2,3,5-trimethyl failed to deliver the expected product (4k–l). Finally, the reactivity of ortho and meta substituted aryl thiol was also evaluated. Accordingly, 2-chlorothiophenol $(2u)$ and 3-fluorothiophenol (2v) underwent reaction smoothly and the desired products 4m and 4n were obtained in 45% and 48% respectively. This result when combined with the result obtained from compounds 4e–f reveals that the position of the substituent at $o/m/p$ did not influence the reactivity of aromatic thiols much under this oxidative condition.

Having demonstrated the synthesis of mono derivatives of quinonyl thioether in one pot using 1.5 equiv. of thiols, the effect of using a super stoichiometric quantity of thiols was examined (Scheme 3). Interestingly, when the reaction was carried out under standard condition using 3.0 equiv. aryl thiol 2n, bis-thiolated product 4o was obtained in 60% yield.

Vitamin K family consists of 2-methyl-1,4-naphthoquinone derivatives substituted with a long alkyl chain at 3-position. Thioether analogs of vitamin K were found to inhibit proteintyrosine phosphatases and induce protein-tyrosine phosphorylation in a human hepatoma cell line (Hep3B).²⁴ Utility of our method for the synthesis of Hep3B inhibitor was examined (Scheme 4). Accordingly, under the optimized condition, mercaptoethanol underwent reaction with menadione (6b) to provide corresponding product 7a, a Hep3B inhibitor, in 56% (Scheme 4).²⁴

As the next part of our study, we examined the reactivity of thiophenols with an additional functional group. Interestingly, under the optimal condition, when HQ was treated with 2 amino thiophenol derivatives 2w and 2x, we observed the formation of biologically important phenothiazines 5a (–Cl) and $5b$ (-CF₃) through tandem C–S and C–N bond formation and oxidation of hydroquinone to quinone in 80% and 70% yield

Scheme 3 One-pot C-H difunctionalization of HQ

Scheme 4 Synthesis of Hep3B inhibitor.

Scheme 5 Domino reaction for phenothiazine-3-one. Reaction conditions: hydroquinone (1.0 equiv., 100 mg), 2-aminothiolphenol (1.5 equiv.), CrO₃ (3.0 equiv.), DCM : H₂O (4 mL, 1 : 1) at rt, isolated yield.

respectively (Scheme 5). However, reaction with sterically substituted HQ (1f) provides the expected phenothiazines 5c only in trace quantity.

In general, phenothiazines are excellent chromogenic molecules that find application as coloring agents.^{25a} Moreover, this moiety is used as a common building block for the synthesis of pharmaceuticals including promazine drugs, antitubercular agents and cholinesterase inhibitors etc.^{25b} The computer modeling of Absorption, Distribution, Metabolism, and Excretion (ADME) properties of compounds provides an idea about structure–property relationships and drug metabolism and pharmacokinetics (DMPK) properties based on the compound structure. In silico ADME-PK is applied at an early phase of the drug development process, in order to remove molecules with poor ADME-PK properties and leads to significant savings in research and development costs.

In this contest, we tried to understand ADME-PK properties of phenothiazine-3-one derivatives 5a–c in silicon study was carried out using Swiss-ADME,²⁶ web tool (see ESI[†]). Results revealed that all compounds showed high gastrointestinal (GI) absorption, good blood–brain barrier (BBB) permeability and also compounds do not have P-glycoprotein (P-gp) permeability. The high lipophilicity (log $P_{\text{o/w}}$) and less skin permeation (log K_{p}) of compounds were observed in the range of 3.11 to 3.61 and -5.08 to -5.96 cm s⁻¹ respectively. Furthermore, compounds 5a-c showed inhibition of cytochrome P450 isomers such as CYP1A2, CYP2C19 and CYP2C9. In addition topological polar surface area (TPSA) of 5a–c was found to 58.20 Å (\leq 140 Å), indicating that compounds have appropriate oral bioavailability (0.55). All compounds 5a–c meet the criteria of drug-likeness assessment based on Lipinski, Ghose and Veber rules. The drug lead-likeness shows, compound 5**b** with trifluromethyl $(-CF_3)$ is the most druggable substance without any violation. A combination of fragment contributions and a complexity penalty of 5b indicated good synthetic accessibility (2.81).

Further, to establish interaction mode with Alzheimer's disease (AD) associated AChE (Fig. 2A) target (PDB ID:1EVE) compound 5b was selected and in silico docking studies of 5a was performed by using Glide 10.7 Schrödinger²⁷ software (see ESI†). Compound 5**b** exhibited G-score -7.30 kcal mol⁻¹ and glide model score -44.18. Moreover, compound 5b showed strong hydrophobic interaction (Fig. 2B and C) with the amino acid residue Tyr121. These results reveal that the compound 5b strongly binds with the AChE active site. Thus compound 5b has good potential to show AChE inhibition.

Fig. 2 (A) Acetylcholinesterase target; (B) 3D structure of protein– ligand interaction; (C) 2D structure of protein–ligand interaction.

Next, we examined the conversion efficiency of hydroquinone to benzoquinone under optimized reaction conditions. The tert-butyl benzoquinone (6b) obtained in excellent yield 91%. It reveals that the optimized condition was suitable for the conversion of HQ to BQ (Scheme 6).

To gain an understanding of the plausible mechanism for the C–S coupling reaction, a series of control experiments were conducted (Scheme 7). The reaction of isopropyl 3-mercaptopropanoate (2f) with benzoquinone (6a) under optimal condition provided only product 3f in low yield (36% equiv. 1), which is much less than the yield (86%, Scheme 2) obtained with hydroquinone (1a). This result indicated the benzoquinone may not the intermediate in the formation of product 3f and a sequential oxidative C–H/S–H cross-coupling between HQ and thiol should have taken place. Furthermore, the reaction carried out in the absence of $CrO₃$ failed to deliver the desired product 3f (equiv. 2), which revealed that, $CrO₃$ plays the role of oxidant in the oxidative cross-coupling reaction. **Paper Conserting Consertion Cons**

Additionally, to determine whether radical intermediate was involved, the reaction was carried out between 1a and 2f in the presence of radical scavenger TEMPO (2.0 equiv.). The yield of product 3f decreased from 86% to 10% which clearly indicated that radical intermediates are involved in the reaction. Further, unlike the formation of poly thioalklylated quinones^{18,19} in nucleophilic addition of thiol to quinones, under our reaction

Scheme 6 Conversion of HQ to BQ.

Scheme 7 Verification experiments for the mechanism.

Scheme 8 A Plausible mechanism

conditions, only mono thioalkylated quinones were obtained. If it is radical cross-coupling the steric effect did not impact severely on the reactivity. From this view of point, we carried out the reaction with o/m substituted aryl thiophenol (Scheme 2, entry 4m–4n) and desired products obtained in moderately. This result reveals that the steric hindrance did not much influence the reactivity. This clearly shows that no conjugate addition of thiol to quinone. The formation of thiyl radicals from thiol is known.²⁸ As it was mentioned earlier, without the presence of water (Table 1, entries 6 and 7) no reaction took place. Based on the verification experiments and background information, a plausible mechanism is proposed as shown in Scheme 8.

 $CrO₃$ on reaction with water is expected to produce chromic acid, a powerful oxidant used in controlled oxidation of organic compounds. After the formation of chromate thioester by the reaction between thiol and chromic acid, it is expected to undergo one-electron transfer reaction to afford thiyl radical A. Similarly, hydroquinone is expected to form corresponding chromate ester C via deprotonation of intermediate B, which is then transformed into a phenoxy followed by semi-quinone radical D via oxidation. Further, thiyl radical A and semi-quinone radical D might undergo radical cross-coupling to form intermediate E, which reoxidizes to form more stable quinonyl thioether.

Conclusion

A simple and efficient method for oxidative radical C–H/S–H cross-coupling of hydroquinones with alkyl thiols/thiophenol/ amino thiophenol to form quinonyl thioethers in monoselective fashion under mild condition is demonstrated. A series of quinonyl alkyl/aryl thioethers and phenothiazines-3 ones were obtained in moderate to good yield at rt. To the best of knowledge, this is the first example of C-S bond formation using a chromium reagent such as H_2CrO_4 . Furthermore, in silico analysis of ADME-PK properties of phenothiazines established that compound 5b meets the criteria of drug and lead likeness and also posses good bioavailability. Moreover, it was also observed that compound 5b strongly binds with Alzheimer's disease (AD) associated AChE target.

Experimental section

General information

All the reagents were purchased commercially and used without further purification. 1 H NMR (400 MHz) and 13 C NMR

(100 MHz) were recorded with Bruker 400 MHz spectrometer in CDCl₃ with tetramethylsilane (TMS) as the internal standard. Multiplicities are reported using the following abbreviations: $s = singlet, d = doublet, t = triplet, q = quartet,$ $m =$ multiplet, sep = septet, br = broad resonance. All the NMR spectra were acquired at ambient temperature. Analytical thin-layer chromatography (TLC) was performed using Silica Gel 60 \AA F254 pre-coated plates (0.25 mm thickness). Visualization was accomplished by irradiation with a UV lamp and staining with I_2 on silica gel. High-resolution mass spectra (HRMS) were recorded on the Thermo Executive Plus spectrometer.

General method-A

To a slowly stirred solution of hydroquinone (1.0 equiv.) and a suitable thiols (1.5 equiv.) in DCM : $H₂O(1:1, 4 mL)$, followed by which CrO_3 (3.0 equiv.) was added portion wise at room temperature. The progress of the reaction was monitored by TLC. Upon the complete consumption of starting materials it was diluted with ethyl acetate and water. The organic phase was separated, extracted with two or more times with ethyl acetate, dried over $Na₂SO₄$, filtered and concentrated. The crude product was purified by silica gel column chromatography using hexane/ethyl acetate as eluent to get product. The disposal of chromic acid starts with adding H_2SO_4 or $CaCO_3$ to the aqueous phase (pH = 1). Further, solid $\text{Na}_2\text{S}_2\text{O}_3$ was added while stirring until the solution turns blue and cloudy. For MSDS information of CrO3 (http://www.labchem.com/tools/msds/msds/ LC13090.pdf). **PSC** Advances Compounds (1990 MLE) were recorded on Paper and Source the Creative Commons and the Creative Commons are the creative Commons and the Creative Commons Attribution-Non-Commons Articles. Commons are the commo

Ethyl 3-((3,6-dioxocyclohexa-1,4-dien-1-yl)thio)propanoate (3a). The reaction was carried out according to general method A using hydroquinone 1a (100 mg, 0.90 mmol), thiol 2a (182 mg, 1.36 mmol) and CrO_3 (267 mg, 2.7 mmol). The reaction mixture was purified using eluent, hexane/ethyl acetate $=$ 90 : 10 to get the product as an orange yellow solid, 162.0 mg, 75.0% yield, mp 82–84 °C; ¹H NMR (400 MHz, CDCl₃): $\delta = 6.71$ $(d, J = 10.0$ Hz, 1H), 6.63 $(d, J = 10.0$ Hz, 1H) 6.33 (s, 1H), 4.08– 4.05 (q, 2H), 2.99–2.96 (t, 2H), 2.64–2.60 (t, 2H); 1.17–1.14 (s, 3H), ¹³C NMR (100 MHz, CDCl₃): $\delta = 183.8, 183.6, 170.6, 151.8,$ 137.4, 136.1, 124.9, 61.0, 32.1, 24.9, 14.1; HRMS (ESI): m/z [M + $[H]^+$ calcd for C₁₁H₁₂NaO₄S: 263.0354; found: 263.0348.

Methyl 3-((3,6-dioxocyclohexa-1,4-dien-1-yl)thio)propanoate (3b). The reaction was carried out according to general method A using hydroquinone 1a (100 mg, 0.90 mmol), thiol 2b $(162 \text{ mg}, 1.36 \text{ mmol})$ and CrO₃ $(267 \text{ mg}, 2.7 \text{ mmol})$. The reaction mixture was purified using eluent, hexane/ethyl acetate $=$ 90 : 10 to get the product as an orange yellow solid, 160.2 mg, in 78.0% yield, mp 84–86 °C; ¹H NMR (400 MHz, CDCl₃): $\delta = 6.73$ $(d, J = 10.0$ Hz 1H), 6.65 $(d, J = 10.0$ Hz, 1H) 6.35 (s, 1H), 3.64 (s, 3H), 3.01-2.98 (t, 2H), 2.68-2.64 (t, 2H); ¹³C NMR (100 MHz, CDCl₃): $\delta = 183.8, 183.6, 171.2, 151.8, 137.4, 136.1, 124.9, 52.1,$ 31.9, 24.9; HRMS (ESI): m/z [M + H]⁺ calcd for C₁₀H₁₀NaO₄S: 249.0197; found: 249.0200.

Butyl 3-((3,6-dioxocyclohexa-1,4-dien-1-yl)thio)propanoate (3c). The reaction was carried out according to general method A using hydroquinone 1a (100 mg, 0.90 mmol), thiol 2c

(220.5 mg, 1.36 mmol) and CrO_3 (267 mg, 2.7 mmol). The reaction mixture was purified using eluent, hexane/ethyl acetate $= 90 : 10$ to get the product as an orange yellow solid, 144.8 mg, 60.0% yield, mp 87–89 °C; ¹H NMR (400 MHz, CDCl₃): $\delta = 6.75$ $(d, J = 10.0 \text{ Hz}, 1\text{H}), 6.68-6.65 \text{ (dd, } J_1 = 10.0 \text{ Hz}, J_2 = 10.0 \text{ Hz},$ 1H) 6.37 (s, 1H), 4.07–4.04 (t, 2H), 3.02–3.00 (t, 2H), 2.68–2.65 (t, 2H), 1.59–1.52 (m, 2H), 1.35–1.26 (m, 2H), 0.88–0.85 (t, 3H); 13C NMR (100 MHz, CDCl₃): $\delta = 183.8, 183.6, 170.8, 151.9, 137.4,$ 136.1, 124.9, 65.4, 32.2, 30.5, 25.0, 19.0, 13.7; HRMS (ESI): m/z $[M + H]^{+}$ calcd for $C_{13}H_{16}O_4S$: 268.0269; found: 268.0774.

Pentyl 3-((3,6-dioxocyclohexa-1,4-dien-1-yl)thio)propanoate (3d). The reaction was carried out according to general method A using hydroquinone 1a (100 mg, 0.90 mmol), thiol 2d (239.5 mg, 1.36 mmol) and CrO_3 (267 mg, 2.7 mmol). The reaction mixture was purified using eluent, hexane/ethyl acetate $= 90 : 10$ to get the product as an orange yellow solid, 132.0 mg, 52.0% yield, mp 86–88 °C; ¹H NMR (400 MHz, CDCl₃): $\delta = 6.76$ $(d, J = 10.0$ Hz, 1H), 6.76 $(d, J = 10.0$ Hz, 1H), 6.38 $(d, J = 1.6$ Hz, 1H), 4.07–4.04 (t, 2H), 3.04–3.00 (t, 2H), 2.69–2.65 (t, 2H), 1.58 (s, 2H), 1.28 (d, 4H), 0.89 (s, 3H); ¹³C NMR (100 MHz, CDCl₃): δ = 183.8, 183.6, 170.7, 151.9, 137.4, 136.1, 124.9, 65.4, 32.2, 28.2, 27.9, 25.1, 22.2, 13.9; HRMS (ESI): m/z [M + H]⁺ calcd for $C_{14}H_{18}NaO_4S: 305.0823$; found: 305.0824. Paper

Paper Was Articles. Published on 21 May 2021 2:24:54 May 2021 2:24:54 PM. This article is licensed under a method on 21 May 2020 2:24:54

CREAT CREAT ARTICLE IS A CREAT ARTICLE IS A CREAT ARTICLE IS A CREAT ARTICLE

Hexyl 3-((3,6-dioxocyclohexa-1,4-dien-1-yl)thio)propanoate (3e). The reaction was carried out according to general method A using hydroquinone 1a (100 mg, 0.90 mmol), thiol 2e $(258.5 \text{ mg}, 1.36 \text{ mmol})$ and $CrO₃$ $(267 \text{ mg}, 2.7 \text{ mmol})$. The reaction mixture was purified using eluent, hexane/ethyl acetate $= 90 : 10$ to get the product as an orange yellow solid, 127.9 mg, 48.0% yield, mp 87–89 °C; ¹H NMR (400 MHz, CDCl₃): $\delta = 6.79$ $(d, J = 10.0$ Hz, 1H), 6.72-6.69 $(dd, J_1 = 10.0$ Hz, $J_2 = 10.0$ Hz, 1H), 6.40 (s, 1H), 4.10–4.07 (t, 2H), 3.06–3.02 (t, 2H), 2.72–2.68 (t, 2H), 1.64–1.57 (m, 2H), 1.29 (s, 6H), 0.87–0.84 (t, 3H); 13C NMR (100 MHz, CDCl₃): $\delta = 183.8, 183.6, 170.8, 152.0, 137.4, 136.1,$ 124.9, 65.4, 32.2, 31.4, 28.5, 25.5, 25.1, 22.5, 13.9; HRMS (ESI): m/z [M + H]⁺ calcd for C₁₅H₂₀NaO₄S: 319.0980; found: 319.0973.

Isopropyl 3-((3,6-dioxocyclohexa-1,4-dien-1-yl)thio) propanoate (3f). The reaction was carried out according to general method A using hydroquinone 1a (100 mg, 0.90 mmol), thiol 2f (201.4 mg, 1.36 mmol) and $C₁₀$ (267 mg, 2.7 mmol). The reaction mixture was purified using eluent, hexane/ethyl acetate $= 90 : 10$ to get the product as an orange yellow solid, 194.4 mg, 85.0% yield, mp 81–83 °C; $^1\mathrm{H}$ NMR (400 MHz, CDCl $_3$): δ = 7.10 (d, J = 10.0 Hz, 1H), 7.01 (d, J = 10.0 Hz, 1H), 6.71 (s, 1H), 5.36–5.27 (m, 1H), 3.36–3.33 (t, 2H), 2.99–2.96 (t, 2H), 1.53– 1.52 (d, 6H); ¹³C NMR (100 MHz, CDCl₃): δ = 183.8, 183.6, 170.2, 151.9, 137.4, 136.1, 124.9, 32.4, 29.6, 25.0, 21.7; HRMS (ESI): m/z $[M + H]^{+}$ calcd for C₁₂H₁₄NaO₄S: 277.0510; found: 277.0502.

2-Methoxyethyl 3-((3,6-dioxocyclohexa-1,4-dien-1-yl)thio) propanoate (3g). The reaction was carried out according to general method A using hydroquinone 1a (100 mg, 0.90 mmol), thiol 2g (223.1 mg, 1.36 mmol) and $C₁$ (267 mg, 2.7 mmol). The reaction mixture was purified using eluent, hexane/ethyl acetate $= 90 : 10$ to get the product as an orange yellow solid, 199.5 mg, 82.0% yield, mp 76–78 °C; 1 H NMR (400 MHz, CDCl3): $\delta = 6.59$ (d, $J = 10.0$ Hz, 1H), 6.50 (d, $J = 10.0$ Hz, 1H), 6.21 (s, 1H), 4.03–4.00 (t, 2H), 3.36–3.34 (t, 2H), 3.11 (s, 3H), 2.87–2.84 (t,

2H), 2.56–2.53 (m, 2H); ¹³C NMR (100 MHz, CDCl₃): $\delta = 183.7$, 183.6, 170.7, 151.6, 137.2, 136.0, 124.9, 70.0, 63.8, 58.6, 31.8, 24.7; HRMS (ESI): m/z [M + H]⁺ calcd for C₁₂H₁₄NaO₄S: 293.0460; found: 293.0451.

Benzyl 3-((3,6-dioxocyclohexa-1,4-dien-1-yl)thio)propanoate (3h). The reaction was carried out according to general method A using hydroquinone 1a (100 mg, 0.90 mmol), thiol 2h $(266.6 \text{ mg}, 1.36 \text{ mmol})$ and $CrO₃$ (267 mg, 2.7 mmol). The reaction mixture was purified using eluent, hexane/ethyl acetate $= 90 : 10$ to get the product as an orange yellow solid, 223.0 mg, 82.0% yield, mp 82–84 °C; ¹H NMR (400 MHz, CDCl₃): $\delta = 7.24$ – 7.21 (m, 5H), 6.66 (d, $J = 10.0$ Hz, 1H), 6.58 (d, $J = 10.0$ Hz, 1H), 6.30 (s, 1H), 5.03 (s, 2H), 2.97–2.93 (t, 2H), 2.67–2.64 (t, 2H); ¹³C NMR (100 MHz, CDCl₃): $\delta = 183.8, 183.7, 170.7, 151.8, 137.4,$ 136.1, 135.5, 128.7, 128.5, 128.4, 66.9, 32.2, 24.9; HRMS (ESI): m/ z [M + H]⁺ calcd for C₁₆H₁₄NaO₄S: 325.0510; found: 325.0514.

3-Phenylpropyl 3-((3,6-dioxocyclohexa-1,4-dien-1-yl)thio) propanoate (3i). The reaction was carried out according to general method A using hydroquinone 1a (100 mg, 0.90 mmol), thiol 2i (304.6 mg, 1.36 mmol) and $CrO₃$ (267 mg, 2.7 mmol). The reaction mixture was purified using eluent, hexane/ethyl acetate $= 90 : 10$ to get the product as an orange yellow semi solid, 118.8 mg, 40.0% yield; ¹H NMR (400 MHz, CDCl₃): δ = 7.29–7.16 (m, 5H), 6.80 (d, $J = 10.0$ Hz, 1H), 6.71 (d, $J = 10.0$ Hz, 1H), 6.41 (s, 1H), 4.16–4.13 (t, 2H), 3.05–3.01 (t, 2H), 2.71–2.66 $(q, 4H)$, 1.99–1.96 (t, 2H); ¹³C NMR (100 MHz, CDCl₃): $\delta = 183.8$, 183.7, 170.7, 151.9, 141.0, 137.4, 136.2, 128.5, 126.1, 124.9, 64.6, 32.2, 32.1, 30.0, 25.0; HRMS (ESI): m/z [M + H]⁺ calcd for C18H18NaO4S: 353.0823; found: 353.0816.

2-(Butylthio)cyclohexa-2,5-diene-1,4-dione (3k). The reaction was carried out according to general method A using hydroquinone 1a (100 mg, 0.90 mmol), thiol 2l (122.5 mg, 1.36 mmol) and CrO_3 (267 mg, 2.7 mmol). The reaction mixture was purified using eluent, hexane/ethyl acetate $= 90 : 10$ to get the product as an orange yellow solid, 79.5 mg, 45.0% yield, mp 80-82 °C; 1 H NMR (400 MHz, CDCl₃): $\delta = 7.11$ (d, $J = 10.0$ Hz, 1H), 7.05–7.02 $(dd, J_1 = 10.0 \text{ Hz}, J_2 = 10.0 \text{ Hz}, 1H$, 6.69 (s, 1H), 3.10–3.06 (t, 2H), 2.06–1.99 (q, 2H), 1.85–1.77 (q, 2H), 1.28–1.25 (t, 3H); ¹³C NMR (100 MHz, CDCl₃): $\delta = 183.9, 183.8, 153.1, 137.4, 136.1,$ 124.7, 30.1, 29.2, 22.1, 13.5.

2-(Phenylthio)cyclohexa-2,5-diene-1,4-dione (4a). The reaction was carried out according to general method A using hydroquinone 1a (100 mg, 0.90 mmol), thiol 2m (150.0 mg, 1.36 mmol) and $CrO₃$ (267 mg, 2.7 mmol). The reaction mixture was purified using eluent, hexane/ethyl acetate $= 90 : 10$, orange red solid, 150.0 mg, 85.0% yield; ¹H NMR (400 MHz, CDCl₃): $\delta =$ 7.50 (s, 5H), 6.83 (d, $J = 10.4$ Hz, 1H), 6.70–6.66 (dd, $J_1 = 10.4$ Hz, $J_2 = 10.4$ Hz, 1H), 5.88 (d, $J = 2.4$ Hz, 1H); ¹³C NMR (100 MHz, CDCl₃): δ = 184.5, 183.9, 154.5, 137.5, 135.8, 135.6, 130.6, 130.4, 126.9, 125.9. The spectral data of the compound 4a was complies with the values reported in the literature.²⁹

2-(p-Tolylthio)cyclohexa-2,5-diene-1,4-dione (4b). The reaction was carried out according to general method A using hydroquinone 1a (100 mg, 0.90 mmol), thiol 2n (168.7 mg, 1.36 mmol) and $CrO₃$ (267 mg, 2.7 mmol). The reaction mixture was purified using eluent, hexane/ethyl acetate $= 90 : 10$ to get the product as an orange red solid, 165.6 mg, 80.0% yield; ¹H NMR

(400 MHz, CDCl₃): $\delta = 7.38$ (d, $J = 8.4$ Hz, 2H), 7.28 (d, $J =$ 8.0 Hz, 2H), 6.81 (d, $J = 10.0$ Hz, 1H), 6.68-6.65 (dd, $J_1 = 10.0$ Hz, $J_2 = 10.0$ Hz, 1H), 5.87 (d, $J = 2.4$ Hz, 1H), 2.4 (s, 3H); ¹³C NMR $(100 \text{ MHz}, \text{CDCl}_3): \delta = 184.5, 184.1, 154.9, 141.1, 137.5, 135.5,$ 131.2, 125.8, 123.2, 21.4. The spectral data of the compound 4b was complies with the values reported in the literature.²⁹

2-((4-Methoxyphenyl)thio)cyclohexa-2,5-diene-1,4-dione (4c). The reaction was carried out according to general method A using hydroquinone 1a (100 mg, 0.90 mmol), thiol 2o (190.5 mg, 1.36 mmol) and $CrO₃$ (267 mg, 2.7 mmol). The reaction mixture was purified using eluent, hexane/ethyl acetate $= 90 : 10$ to get the product as an orange red solid, 168.3 mg, 76.0% yield; ¹H NMR (400 MHz, CDCl₃): $\delta = 7.37$ (d, $J = 8.8$ Hz, 2H), 6.98 (d, $J =$ 8.8 Hz, 2H), 6.78 (d, $J = 10.0$ Hz, 1H), 6.66-6.63 (dd, $J_1 = 10.0$ Hz, $J_2 = 10.0$ Hz, 1H), 5.84 (d, $J = 2.4$ Hz, 1H), 3.84 (s, 3H); ¹³C NMR $(100 \text{ MHz}, \text{CDCl}_3)$: $\delta = 184.5, 184.1, 161.5, 155.3, 137.5, 137.1,$ 135.9, 125.8, 116.9, 116.0, 55.5. The spectral data of the compound 4c was complies with the values reported in the literature.²⁹ **PSC** Advances Veweralcomes Veweralcomes Articles. Published on 21 May 2020. The published on 21 May 2020. Downloaded on 21 May 2021 2:24:54 PM. This article is likely and the set of $\frac{1}{2}$ PM. This article is likely a

2-((4-Nitrophenyl)thio)cyclohexa-2,5-diene-1,4-dione (4d). The reaction was carried out according to general method A using hydroquinone 1a (100 mg, 0.90 mmol), thiol 2p (210.8 mg, 1.36 mmol) and $CrO₃$ (267 mg, 2.7 mmol). The reaction mixture was purified using eluent, hexane/ethyl acetate $= 90 : 10$ to get the product as a yellow solid, 165.0 mg, 70.0% yield; $^1\mathrm{H}$ NMR (400 MHz, CDCl₃): $\delta = 8.34$ (d, $J = 8.0$ Hz, 2H), 7.73 (d, $J =$ 8.0 Hz, 2H), 6.88 (d, $J = 10.0$ Hz, 1H), 6.74 (d, $J = 10.0$ Hz, 1H), 5.95 (s, 1H); ¹³C NMR (100 MHz, CDCl₃): $\delta = 184.0, 183.2, 152.3$, 149.0, 137.5, 136.3, 135.9, 135.7, 126.7, 125.3, 124.1. The spectral data of the compound 4d was complies with the values reported in the literature.²⁹

2-((4-Chlorophenyl)thio)cyclohexa-2,5-diene-1,4-dione (4e). The reaction was carried out according to general method A using hydroquinone $1f(100 \text{ mg}, 0.60 \text{ mmol})$ and $CrO₃(180 \text{ mg},$ 1.8 mmol). The reaction mixture was purified using eluent, hexane/ethyl acetate $= 90 : 10$ to get the product as an yellow solid, 92.3 mg, 41.0% yield; ¹H NMR (400 MHz, CDCl₃): δ = 7.50–7.43 (dd, $J_1 = 8.4$ Hz, $J_2 = 8.4$ Hz, 2H), 6.85 (d, $J = 10.0$ Hz, 2H), 6.71–6.68 (dd, $J_1 = 10.0$ Hz, $J_2 = 10.0$ Hz, 1H), 5.87 (d, $J =$ 2.0 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃): $\delta = 184.3, 183.8, 153.9,$ 137.5, 137.3, 136.9, 135.8, 130.7, 126.0, 125.3. The spectral data of the compound 4e was complies with the values reported in the literature.²⁹

2-((4-Bromophenyl)thio)cyclohexa-2,5-diene-1,4-dione (4f). The reaction was carried out according to general method A using hydroquinone 1a (100 mg, 0.90 mmol), thiol 2r (255.5 mg, 1.36 mmol) and CrO_3 (267 mg, 2.7 mmol). The reaction mixture was purified using eluent, hexane/ethyl acetate $= 90 : 10$ to get the product as an orange solid, 113.8 mg, 43.0% yield; $^1\rm H$ NMR (400 MHz, CDCl₃): $\delta = 7.65 - 7.61$ (m, 2H), 7.39-7.36 (m, 2H), 6.84 (d, $J = 10.0$ Hz, 1H), 6.72-6.68 (dd, $J_1 = 10.0$ Hz, $J_2 =$ 10.0 Hz, 1H), 5.88 (d, $J = 2.4$ Hz, 1H); ¹³C NMR (100 MHz, CDCl₃): $\delta = 184.3, 183.7, 153.8, 137.5, 137.2, 135.9, 133.7, 126.1,$ 126.0, 125.9, 125.6. The spectral data of the compound 4f was complies with the values reported in the literature.²⁹

2-((4-Fluorophenyl)thio)cyclohexa-2,5-diene-1,4-dione (4g). The reaction was carried out according to general method A using hydroquinone 1a (100 mg, 0.90 mmol), thiol 2s (174.1 mg, 1.36 mmol) and $CrO₃$ (267 mg, 2.7 mmol). The reaction mixture was purified using eluent, hexane/ethyl acetate $= 90 : 10$ to get the product as an orange solid, 84.3 mg, 40.0% yield; ¹H NMR (400 MHz, CDCl₃): $\delta = 7.51 - 7.48$ (q, 2H), 7.22-7.18 (t, 2H), 6.83 $(d, J = 10.0$ Hz, 1H), 6.71–6.68 (dd, $J_1 = 10.0$ Hz, $J_2 = 10.0$ Hz, 1H), 5.85 (d, $J = 2.4$ Hz, 1H); ¹³C NMR (100 MHz, CDCl₃): $\delta =$ 184.4, 183.9, 165.9, 162.9, 154.4, 137.9, 137.8, 137.5, 125.9, 117.9, 117.7. The spectral data of the compound 4g was complies with the values reported in the literature.²⁹

2-Methoxy-5-(p-tolylthio)cyclohexa-2,5-diene-1,4-dione (4i). The reaction was carried out according to general method A using hydroquinone 1b (100 mg, 0.71 mmol), thiol 2n (132.0 mg, 1.06 mmol) and CrO_3 (210.9 mg, 2.13 mmol). The reaction mixture was purified using eluent, hexane/ethyl acetate $= 90 : 10$ to get the product as an orange red solid, 125.8 mg, 68.0% yield; ¹H NMR (400 MHz, CDCl₃): $\delta = 7.36$ (d, J = 8.0, 2H), 7.28 (d, $J = 8.0$, 2H), 5.97 (s, 1H), 5.78 (s, 1H), 3.84 (s, 3H), 2.41 (s, 3H); ¹³C NMR (100 MHz, CDCl₃): $\delta = 184.8, 184.0, 178.8,$ 159.6, 157.0. 141.1, 135.5, 135.4, 131.2, 123.4, 107.6, 106.7, 56.6, 21.4.

2-Acetyl-3-(p-tolylthio)cyclohexa-2,5-diene-1,4-dione (4j). The reaction was carried out according to general method A using hydroquinone 1c (100 mg, 0.65 mmol), thiol 2n (122.3 mg, 0.98 mmol) and $CrO₃$ (193.0 mg, 1.95 mmol). The reaction mixture was purified using eluent, hexane/ethyl acetate $= 90 : 10$ to get the product as an orange red solid, 77.8 mg, 44.0% yield; 1 H NMR (400 MHz, CDCl₃): $\delta = 7.33$ (d, $J = 8.0$, 2H), 7.15 (d, $J = 8.0$, 2H), 6.79 (d, $J = 8.0$ 1H), 6.72 (d, $J = 8.0$, 1H), 2.35 (s, 3H), 2.12 (s, 3H); ¹³C NMR (100 MHz, CDCl₃): δ = 198.8, 183.5, 183.1, 145.4, 142.6, 139.9, 136.7, 136.5, 134.3, 130.3, 126.3, 31.3, 21.4.

2-((2-Chlorophenyl)thio)cyclohexa-2,5-diene-1,4-dione (4m). The reaction was carried out according to general method A using hydroquinone 1a (100 mg, 0.90 mmol), thiol 2u (197.0 mg, 1.36 mmol) and $CrO₃$ (267 mg, 2.7 mmol). The reaction mixture was purified using eluent, hexane/ethyl acetate $= 90 : 10$ to get the product as an orange yellow solid, 101.3 mg, 45.0% yield; $^1\mathrm{H}$ NMR (400 MHz, CDCl₃): $\delta = 7.52-7.41$ (m, 4H), 6.84 (d, $J = 10$, 1H), 6.72–6.69 (dd, $J_1 = 2.4$, $J_2 = 2.4$, 1H), 5.90 (d, $J_1 = 2.4$, 1H); ¹³C NMR (100 MHz, CDCl₃): $\delta = 184.3, 183.6, 153.6, 137.5,$ 135.9, 135.3, 133.8, 131.9, 130.9, 128.8, 126.1.

2-((2-Chlorophenyl)thio)cyclohexa-2,5-diene-1,4-dione (4n). The reaction was carried out according to general method A using hydroquinone 1a (100 mg, 0.90 mmol), thiol 2v (175.0 mg, 1.36 mmol) and $CrO₃$ (267 mg, 2.7 mmol). The reaction mixture was purified using eluent, hexane/ethyl acetate $= 90 : 10$ to get the product as an orange yellow solid, 112.5 mg, 48.0% yield; ¹H NMR (400 MHz, CDCl₃): $\delta = 7.55$ -7.51 (m, 2H), 7.29-7.23 (m, 2H), 6.85 (d, $J = 10$, 1H), 6.72-6.69 (dd, $J_1 = 2.4$, $J_2 = 2.4$, 1H), 5.88 (q, 1H); ¹³C NMR (100 MHz, CDCl₃): δ = 184.3, 183.7, 163.9, 161.4, 151.7, 137.5, 137.4, 135.9, 133.5, 133.4, 126.0, 125.9, 125.8, 117.2, 116.9, 114.2, 114.0.

2,6-Bis(p-tolylthio)cyclohexa-2,5-diene-1,4-dione (4o). The reaction was carried out according to general method A using hydroquinone 1a (100 mg, 0.90 mmol), thiol 2n (334.8 mg, 2.7 mmol) and $CrO₃$ (267 mg, 2.7 mmol). The reaction mixture was purified using eluent, hexane/ethyl acetate $= 90 : 10$, to get the

product as an orange red solid, 190.1 mg, 60.0% yield; ¹H NMR (400 MHz, CDCl₃): $\delta = 7.36$ (d, $J = 8.0, 4H$), 7.27–7.25 (t, 4H), 5.57 (s, 2H), 2.39 (s, 6H); ¹³C NMR (100 MHz, CDCl₃): $\delta = 182.4$, 181.3, 154.0, 141.1, 135.5, 131.2, 126.5, 123.5, 21.4.

8-Chloro-3H-phenothiazin-3-one (5a). The reaction was carried out according to general method A using hydroquinone 1a (100 mg, 0.90 mmol), thiol 2u (216.2 mg, 1.36 mmol) and $CrO₃$ (267 mg, 2.7 mmol). The reaction mixture was purified using eluent, hexane/ethyl acetate $= 90 : 10$ to get the product as a red solid, 177.8 mg, 80.0% yield; $^1\rm H$ NMR (400 MHz, CDCl3): δ = 7.90 (d, 1H), 7.60 (d, 1H), 7.44–7.37 (m, 2H), 6.95–6.75 (m, 1H), 6.74 (s, 1H); ¹³C NMR (100 MHz, CDCl₃): $\delta = 182.4, 147.5,$ 139.9, 139.7, 135.6, 134.6, 133.4, 133.1, 131.0, 125.9, 121.9, 120.5.

8-(Trifluoromethyl)-3H-phenothiazin-3-one (5b). The reaction was carried out according to general method A using hydroquinone 1a (100 mg, 0.90 mmol), thiol 2v (189.0 mg, 1.36 mmol) and $CrO₃$ (267 mg, 2.7 mmol). The reaction mixture was purified using eluent, hexane/ethyl acetate $= 90 : 10$ to get the product as a red solid, 177.0 mg, 70.0% yield, mp 76–78 °C; $^1\mathrm{H}$ NMR (400 MHz, CDCl₃): $\delta = 8.13$ (s, 1H), 7.67-7.54 (m, 3H), 6.97–6.76 (m, 1H), 6.76 (s, 1H); ¹³C NMR (100 MHz, CDCl₃): δ = 182.4, 147.8, 139.8, 139.1, 135.8, 134.1, 130.8, 130.7, 127.5, 126.9, 126.8, 125.8, 121.1. Paper

Published on 21 May 2020. The Research of Common State Article is licensed on 21 May 2021-23:24:54 PM. The material including the CREMA, New Data Creative Common State Article is licensed under a CREMA Creative Com

2-(tert-Butyl)-8-chloro-3H-phenothiazin-3-one (5c). The reaction was carried out according to general method A using hydroquinone 1f (100 mg, 0.60 mmol), thiol 2h (143.5 mg, 0.90 mmol) and $CrO₃$ (178.2 mg, 1.8 mmol). The reaction mixture was purified using eluent, hexane/ethyl acetate $= 90 : 10$, orange red solid, trace yield; ¹H NMR (400 MHz, CDCl₃): $\delta =$ 7.85 (d, 1H), 7.50 (s, 1H), 7.14–7.33 (m, 2H), 6.66 (s, 1H), 1.38 (s, 9H).

2-(tert-Butyl)cyclohexa-2,5-diene-1,4-dione (6b). The reaction was carried out according to general method A using 2-(tertbutyl)benzene-1,4-diol 6b (100 mg, 0.58 mmol), thiol 2w (68.0 mg, 0.87 mmol) and CrO_3 (172.3 mg, 1.74 mmol). The reaction mixture was purified using eluent, hexane/ethyl acetate $= 90 : 10$ to get the product as a yellow solid, 90.0 mg, 91.0% yield; ¹H NMR (400 MHz, CDCl₃): $\delta = 6.65$ (s, 2H), 6.56 (2, 1H), 1.25 (s, 9H); ¹³C NMR (100 MHz, CDCl₃): δ = 188.4, 187.4, 156.0, 138.7, 135.0, 131.5, 35.3, 29.2, 29.0.

2-((2-Hydroxyethyl)thio)-3-methylnaphthalene-1,4-dione

(7a). The reaction was carried out according to general method A using menadione 6b (100 mg, 0.58 mmol), thiol 2w (68.0 mg, 0.87 mmol) and $CrO₃$ (172.3 mg, 1.74 mmol). The reaction mixture was purified using eluent, hexane/ethyl acetate $=$ 90 : 10 to get the product as a yellow solid, 80.5 mg, 56.0% yield; ¹H NMR (400 MHz, CDCl₃): $\delta = 8.08$ -8.04 (m, 2H), 7.70-7.67 (t, 2H), 3.80–3.78 (t, 2H), 3.35–3.32 (t, 2H), 2.39 (s, 3H); 13C NMR $(100 \text{ MHz}, \text{CDCl}_3)$: $\delta = 182.3, 181.6, 148.3, 145.7, 133.8, 133.5,$ 132.7, 132.0, 126.9, 126.7, 62.0, 37.3, 15.5. The spectral data of the compound 7a was complies with the values reported in the literature.²⁴

Conflicts of interest

The authors declare no conflict of interest.

Acknowledgements

AKTP and SP thanks UGC-RFSMS, New Delhi for the award of the fellowship. We thank DST-FIST and DST-PURSE New Delhi, India for NMR and HRMS facilities at School of Chemistry, Bharathidasan University, Tiruchirappalli, India.

References

- 1 (a) G. A. Kraus and J. Mengwasser, Molecules, 2009, 14, 2857– 2861 ; (b) A. E. Wendlandt and S. S. Stahl, Angew. Chem., Int. Ed., 2015, 54, 14638–14658; (c) B. Hosamani, M. F. Ribeiro, E. N. da Silva Junior and I. N. N. Namboothiri, Org. Biomol. Chem., 2016, 14, 6913–6931; (d) C. Song, X. Dong, H. Yi, C. W. Chiang and A. Lei, ACS Catal., 2018, 8, 2195–2199.
- 2 (a) P. R. Dandawate, A. C. Vyas, S. B. Padhye, M. W. Singh and J. B. Baruah, Mini-Rev. Med. Chem., 2010, 10, 436–454; (b) M. K. Hadden, S. A. Hill, J. Davenport, R. L. Matts and B. S. J. Blagg, Bioorg. Med. Chem., 2009, 17, 634–640.
- 3 (a) J. Madeo, A. Zubair and F. Marianne, SpringerPlus, 2013, 2(139), 1–8; (b) S. Park, E. Yun, I. H. Hwanf, S. Yoon, D. E. Kim, M. Na, G. Y. Song and S. Oh, Mar. Drugs, 2014, 12, 3231–3244; (c) M. Arai, T. Kawachi, H. Sato, A. Setiawan and M. Kobayashi, Bioorg. Med. Chem. Lett., 2014, 24, 3155–3157; (d) J. L. Bolton and T. Dunlap, Chem. Res. Toxicol., 2017, 30, 13-37; (e) P. A. Garcia, A. P. Hernández, A. S. Feliciano and M. A. Castro, Mar. Drugs, 2018, 16(292), 1–51.
- 4 (a) V. J. Litchfield, R. B. Smith, A. M. Franklin and J. Davis, Synth. Commun., 2008, 38, 3447–3455; (b) S. Er, C. Suh, M. P. Marshaka and A. A. Guzik, Chem. Sci., 2015, 6, 885– 893; (c) E. J. Son, J. H. Kim, K. Kim and C. B. Park, J. Mater. Chem. A, 2016, 4, 11179–11202; (d) Y. Ding, Y. Li and G. Yu, Chem, 2016, 1, 790–801; (e) O. Taran, Front. Chem., 2017, 5(49), 1–13.
- 5 (a) D. Sladic and M. J. Gasic, Molecules, 2006, 11(1), 1–33; (b) N. E. Najjar, H. G. Muhtasib, R. A. Ketola, P. Vuorela, A. Urtti and H. Vuorela, Phytochem. Rev., 2011, 10, 353–370; (c) P. S. Guin, S. Das and P. C. Mandal, Int. J. Electrochem., 2011, 816202, 1–22.
- 6 (a) J. Fourie, C. J. Oleschuk, F. Guziec Jr, L. Guziec, D. J. Filterman, C. Monterrosa and A. Begleiter, Canc. Chemother. Pharmacol., 2002, 49, 101–110; (b) C. Frontana, A. V. Mayagoitia, J. Garza, R. Vargas and I. Gonzalez, J. Phys. Chem. A, 2006, 110, 9411–9419; (c) L. Zhou, Y. Chen, X. Yang, Y. Su, W. Zhang and J. Xu, Catal. Lett., 2008, 125, 154–159; (d) X. Yang, Y. Wang, C. Zhang, T. Fang, L. Zhou, W. Zhang and J. Xu, J. Phys. Org. Chem., 2011, 24, 693–697.
- 7 W. Yu, P. Hjerrild, K. M. Jacobsen, H. N. Tobiesen, L. Clemmensen and T. B. Poulsen, Angew. Chem., Int. Ed., 2018, 57, 9805–9809 and references cited therein.
- 8 (a) K. Mozaina, C. L. Cantrell, A. B. Mims, A. R. Lax, M. R. Tellez and W. L. A. Osbrink, J. Agric. Food Chem., 2008, 56, 4021–4026; (b) C. K. Ryu, S. Y. Lee, N. Y. Kim, J. A. Hong, J. H. Yoon and A. Kim, Bioorg. Med. Chem. Lett., 2011, 21, 427–430; (c) V. K. Tandon, S. Kumar, N. N. Mishra and P. K. Shukla, Eur. J. Med. Chem., 2012,

56, 375–386; (d) A. Kacmaz, E. T. Acar, G. Atun, K. Kaya, B. D. Sigirci and F. Bagcigil, ChemistrySelect, 2018, 3, 8615– 8623; (e) H. Yıldırım, N. Bayrak, M. Yıldız, E. M. Kara, B. O. Celik and A. F. Tuyun, J. Mol. Struct., 2019, 1195, 681– 688.

- 9 Z. Wang, E. C. Southwick, M. Wang, S. Kar, K. S. Rosi, C. S. Wilcox, J. S. Lazo and B. I. Car, Cancer Res., 2001, 61, 7211–7216.
- 10 (a) H. O. Sjögren, M. Isaksson, D. Willner, I. Hellström, K. E. Hellström and P. A. Cancer, Res, 1997, 57, 4530-4536; (b) A. M. Keith Walker, A. C. Braemer, S. Hitt, R. E. Jones and T. R. Matthews, J. Med. Chem., 1978, 21, 840–842; (c) A. J. Sedman, Am. J. Med., 1984, 76, 109–114; (d) D. M. Livermore, A. M. Sefton and G. M. Scott, J. Antimicrob. Chemother., 2003, 52, 331–344; (e) X. Y. Zhang, G. S. Liu and J. Yuan, Chin. J. Clin. Rehabil., 2013, 7(24), 3340–3342; (f) T. F. Luscher, S. Taddei, J. C. Kaski, J. W. Jukema, D. Kallend, T. Munzel, J. J. P. Kastelein and J. E. Deanfield, Eur. Heart J., 2012, 33, 857-865.
- 11 (a) A. S. Thigpen, S. T. Nestor, R. A. O'Brien, S. Minkowicz, Y. Sheng, J. H. Davis Jr, K. N. West and A. Mirjafari, New J. Chem., 2017, 41, 1625–1630; (b) A. Bhadani and S. Singh, Langmuir, 2011, 27(23), 14033–14044.
- 12 (a) T. Kondo and T. Mitsudo, Chem. Rev., 2000, 100, 3205– 3220 and references cited therein; $; (b)$ C. Shen, P. Zhang, Q. Sun, S. Bai, T. S. Andy Hor and X. Liu, Chem. Soc. Rev., 2015, 44, 291–314 and references cited therein; For latest papers-; (c) W. H. Bao, C. Wu, J. T. Wang, W. Xia, P. Chen, Z. T. X. Xu and W. M. He, Org. Biomol. Chem., 2018, 16, 8403–8407; (d) G. S. Sorabad and M. R. Maddani, Asian J. Org. Chem., 2019, 8, 1–9; (e) W. H. Bao, Z. Wang, X. Tang, Y. F. Zhang, J. X. Tana, Q. Zhu, Z. Cao, Y. W. Lin and W. M. He, Chin. Chem. Lett., 2019, 30, 2259–2262; (f) L. Y. Xie, Y. L. Chen, L. Qin, Y. Wen, J. W. Xie, J. X. Tan, Y. Huang, Z. Cao and W. M. He, Org. Chem. Front., 2019, 6, 3950–3955. Open Access Article. Published on 21 May 2020. Downloaded on 9/24/2024 2:24:54 PM. This article is licensed under a [Creative Commons Attribution-NonCommercial 3.0 Unported Licence.](http://creativecommons.org/licenses/by-nc/3.0/) **[View Article Online](https://doi.org/10.1039/d0ra01519a)**
	- 13 (a) X. Yu, Q. Wu, H. Wan, Z. Xu, X. Xu and D. Wang, RSC Adv., 2016, 6, 62298–62301; (b) C. Zhang, J. Mc-Clure and C. J. Chou, J. Org. Chem., 2015, 80, 4919–4927.
	- 14 (a) D. Wang, X. Yu, L. Wang, W. Yao, Z. Xu and H. Wan, Tetrahedron Lett., 2016, 57, 5211–5214; (b) D. Wang, X. Yu, W. Yao, W. Hu, C. Ge and X. Shi, Chem. - Eur. J., 2016, 22, 5543–5546.
	- 15 S. Rajasekar, T. P. A. Krishna, N. Tharmalingam, A. Ilangovan and E. Mylonakis, ChemistrySelect, 2019, 4, 2281–2287.
	- 16 F. L. Zeng, X. L. Chen, S. Q. He, K. Sun, Y. Liu, R. Fu, L. B. Qu, Y. F. Zhao and B. Yu, Org. Chem. Front., 2019, 6, 1476–1480.
	- 17 (a) V. K. Tandon and H. K. Maurya, Tetrahedron Lett., 2009, 50, 5896–5902; (b) V. K. Tandon, H. K. Maurya, S. Kumar, A. Rashid and D. Panda, RSC Adv., 2014, 4, 12441–12447.
- 18 (a) I. H. Spinner, W. D. Raper and W. Metanomski, Can. J. Chem., 1963, 41, 483–494; (b) A. R. Katritzky, D. Fedoseyenkoa, P. P. Mohapatraa and P. J. Steel, Synthesis, 2008, 5, 777–787; (c) V. K. Tandona, D. B. Yadava, R. V. Singh, M. Vaisha, A. K. Chaturvedi and P. K. Shukla, Bioorg. Med. Chem. Lett., 2005, 15, 3463–3466; (d) D. E. Allgeier, S. A. Herbert, R. Nee, K. D. Schlecht and K. T. Finley, J. Org. Chem., 2003, 68, 4988–4990.
- 19 (a) Y. Lu, Y. Zhao, S. Wang, X. Wang, Z. Ge and R. Li, RSC Adv., 2016, 6, 11378-11381; (b) K. W. Wellington, G. E. Gordon, L. A. Ndlovu and P. Steenkamp, ChemCatChem, 2013, 5, 1570–1577; (c) K. W. Wellington, R. Bokako, N. Raseroka and P. Steenkamp, Green Chem., 2012, 14, 2567–2576; (d) M. Schlippert, A. Mikolasch, V. Hahn and F. Schauer, J. Mol. Catal. B Enzym., 2016, 126, 106–114.
- 20 (a) X. Zeng and X. Cong, Org. Chem. Front., 2015, 2, 69–72; (b) M. Yadava and X. Qiang, Chem. Commun., 2013, 49, 3327– 3329; (c) A. Levina, L. Zhang and P. A. Lay, *J. Am. Chem.* Soc., 2010, 132, 8720–8731; (d) M. Juaristi, J. M. Aizpurua, B. Lecea and C. Palomo, Can. J. Chem., 1984, 62, 2941–2944.
- 21 (a) A. Ilangovan, S. Saravanakumar and S. Malayappasamy, Org. Lett., 2013, 15(19), 4968–4971; (b) A. Ilangovan, A. Polu and G. Satish, Org. Chem. Front., 2015, 12, 1616–1620; (c) A. Polu and A. Ilangovan, Tetrahedron Lett., 2018, 59, 438– 441; (d) T. P. A. Krishna, P. Sakthivel and A. Ilangovan, Org. Chem. Front., 2019, 6, 3244–3251; (e) P. Sakthivel, T. P. A. Krishna and A. Ilangovan, Org. Biomol. Chem., 2020, 18, 3027–3031.
- 22 Y. Hong, G. Zhang, H. Wang, Z. Huang, J. Wang, A. K. Singh and A. Lei, Chem. Rev., 2017, 117, 9016–9085 and references cited therein.
- 23 Z. Huang, D. Zhang, X. Qi, Z. Yan, M. Wang, H. Yan and A. Lei, Org. Lett., 2016, 18, 2351–2354.
- 24 Y. Nishikawa, Z. Wang, J. Kerns, C. S. Wilcox and B. I. Carr, J. Biol. Chem., 1999, 274, 34803–34810.
- 25 (a) V. Venkatraman, M. Foscato, V. R. Jensenb and B. K. Alsberg, J. Mater. Chem. A, 2015, 3, 9851–9860; (b) Y. Liaoa, P. Jianga, S. Chena, F. Xiaoa and G. J. Deng, RSC Adv., 2013, 3, 18605–18608.
- 26 (a) A. Daina, O. Michielin and V. Zoete, Sci. Rep., 2017, 7, 42717; (b) T. Sander, J. Freyss, M. Von-Korff and C. Rufener, J. Chem. Inf. Model., 2015, 55, 460–473; (c) A. Daina and V. Zoete, Curr. Med. Chem., 2016, 11(11), 1117–1121.
- 27 J. R. Greenwood, D. Calkins, A. P. Sullivan and J. C. Shelley, J. Comput. Aided Mol. Des., 2010, 24, 591–604.
- 28 F. Denes, M. Pichowicz, G. Povie and P. Renaud, Chem. Rev., 2014, 114, 2587–2693 and cited therein.
- 29 D. Wang, X. Yu, L. Wang, W. Yao, Z. Xu and H. Wan, Tetrahedron Lett., 2016, 57, 5211–5214 and references cited therein.