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Synthesis and biological evaluation of fluorinated 3,4-dihydroquinolin-2(1*H*)-ones and 2-oxindoles for anti-hepatic fibrosis†

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(*Z*)-4-(Iodomethylene)-3-(2,2,2-trifluoroethyl)-3,4-dihydroquinolin-2(1*H*)-ones and fluorinated 3,3-disubstituted 2-oxindoles are synthesized and evaluated for anti-hepatic fibrosis. CCK-8 assay indicates that most of the compounds have no obvious cytotoxicity on the human hepatic stellate cells (HSC) cell line. Collagen I and fibrosin expression levels are tested by ELISA, and the results show that several compounds can inhibit the expression of collagen I and fibrosin. Additionally, results from real time-PCR reveal that only one compound can inhibit the expression level of α -SMA, suggesting that this compound can inhibit the activation of the HSC cell line. These studies demonstrate that this compound may be a potential novel drug candidate for anti-hepatic fibrosis (approximately 5–6 lines).

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Hepatic fibrosis is an important pathophysiological process of chronic liver diseases involving excessive deposition of collagen-based extracellular matrix (ECM), which is extremely rich in collagen I and III, since it can lead to cirrhosis and portal hypertension.¹ Hepatic stellate cells (HSC) (lipocytes, Ito cells) are identified as the main collagen-producing cells in the liver, which are the main cells involved in liver fibrogenesis.² Many factors have been shown to affect the activity of HSCs, including interleukin-1, tumor necrosis factor, transforming growth factor (TGF), and basic fibroblast growth factors. TGF- β is confirmed to be the most potent stimulus for the activation of HSC.³ Additionally, fibrosin (FBRs) is one new fibrogenic cytokine that was first identified as a product of inflammatory cells and was cloned as “fibrosin”.⁴ Our previous studies showed that serum FBRs levels in patients with chronic hepatitis B were positively correlated with the degree of hepatic fibrosis, which could be used to evaluate hepatic fibrosis.⁵

Many anti-fibrotic treatments have been tried, but none is recognized worldwide. Thus, it is urgent to explore the effective medicine or drug candidate to treat hepatic fibrosis. Meanwhile, nitrogen-containing heterocycles are important in drug-discovery process due to their unique biological activities.⁶ Therefore, significant efforts have been given to the methodologies development and library construction for the rapid preparation of heterocycles and their derivatives. On the other hand, the incorporation of fluorinated groups into privileged scaffolds has received considerable attention in both synthetic and medicinal chemistry community, with the aim to improve their biological properties.⁷ Recently, we accomplished the synthesis of (*Z*)-4-(iodomethylene)-3-(2,2,2-trifluoroethyl)-3,4-dihydroquinolin-2(1*H*)-ones through a photoinduced radical cyclization of benzene-tethered 1,7-enynes with Togni reagent in the presence of sodium iodide under ultraviolet irradiation.⁸ Additionally, fluorinated 3,3-disubstituted 2-oxindoles could be generated as well under ultraviolet irradiation through a photoinduced radical cyclization of *N*-arylacrylamides with fluorinated alkyl iodides.⁹ Herein, we describe the identification and biological evaluation of (*Z*)-4-(iodomethylene)-3-(2,2,2-trifluoroethyl)-3,4-dihydroquinolin-2(1*H*)-ones including **3a–f** and fluorinated 3,3-disubstituted 2-oxindoles including **5a–h**. The results show that compounds **3b** and **5a** can inhibit the expression of collagen I and FBRs. Moreover, real-time-PCR shows that only compound **5a** can inhibit the expression level of α -SMA, suggesting that compound **5a** can inhibit the activation of LX-2 cells. These studies demonstrate that compound **5a** may be a potential novel drug candidate for anti-hepatic fibrosis.

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Results and discussion

Chemistry: synthesis of (Z)-4-(iodomethylene)-3-(2,2,2-trifluoroethyl)-3,4-dihydroquinolin-2(1H)-ones and fluorinated 3,3-disubstituted 2-oxindoles

The general synthetic route to the target (Z)-4-(iodomethylene)-3-(2,2,2-trifluoroethyl)-3,4-dihydroquinolin-2(1H)-ones and fluorinated 3,3-disubstituted 2-oxindoles is described in Scheme 1. Accordingly, a series of (Z)-4-(iodomethylene)-3-(2,2,2-trifluoroethyl)-3,4-dihydroquinolin-2(1H)-ones were prepared through a three-component cyclization of benzene-tethered 1,7-enynes **1** with Togni reagent **2** in the presence of sodium iodide under ultraviolet irradiation (Scheme 1).⁸ Additionally, by treatment of *N*-arylacrylamides **4** and fluorinated alkyl iodides under ultraviolet irradiation at room temperature, the corresponding fluorinated 3,3-disubstituted 2-oxindoles **5** could be generated in moderate to good yields (Scheme 1).⁹ As a result, a small library of (Z)-4-(iodomethylene)-3-(2,2,2-trifluoroethyl)-3,4-dihydroquinolin-2(1H)-ones and fluorinated 3,3-disubstituted 2-oxindoles was constructed for the subsequent biological evaluation.

Chemical structure of chemical compounds

(Z)-4-(Iodomethylene)-3-(2,2,2-trifluoroethyl)-3,4-dihydroquinolin-2(1H)-ones **3** included compounds **3a–f** and fluorinated 3,3-disubstituted 2-oxindoles included compounds **5a–h**. Their chemical structures are presented in Fig. 1. General experimental procedure for the synthesis of (Z)-4-(iodomethylene)-3-(2,2,2-trifluoroethyl)-3,4-dihydroquinolin-2(1H)-ones **3** and fluorinated 3,3-disubstituted 2-oxindoles **5** is presented in the ESI.†

Effects of chemical compounds on LX-2 viability

The data showed that all compounds at 0.1 μ M, 1 μ M, 10 μ M and 100 μ M had no significant inhibition or cytotoxicity on LX-2 cells after 24 h, 48 h or 72 h treatment, and some compounds contributed to the proliferation of LX-2 cells (Fig. 2).

Collagen I expression levels were reduced by some compounds

Liver fibrosis is the excessive accumulation of extracellular matrix proteins including collagen that occurs in most types of chronic liver diseases. HSC is responsible for initiating and

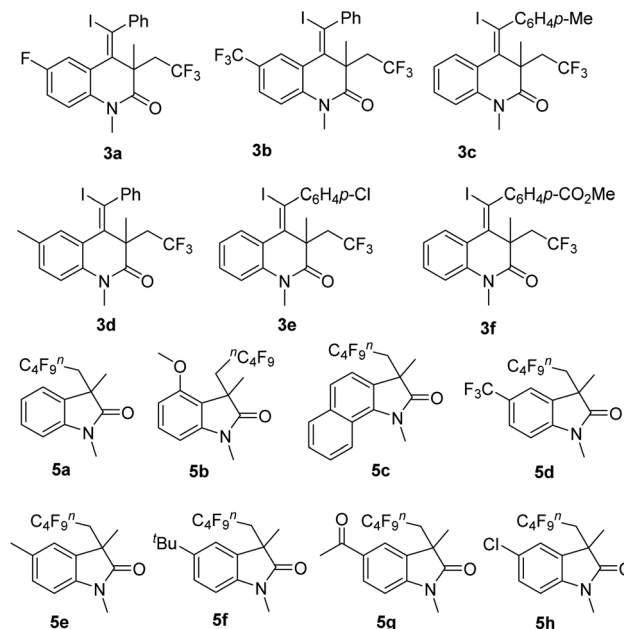


Fig. 1 Chemical structures of fluorinated 3,4-dihydroquinolin-2(1H)-ones and 2-oxindoles.

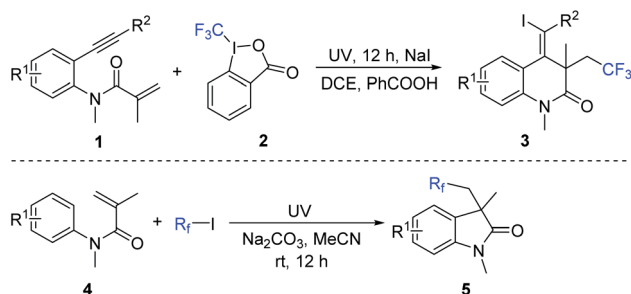
spreading liver fibrosis, secreting fibrogenic factors that encourage portal fibrocytes, fibroblasts, and bone marrow-derived myofibroblasts.¹⁰ Collagen I was markedly expressed by activated HSCs. In this study, LX-2 cells were activated by TGF- β 1, and then compounds were individually added into the culture medium. After 72 h, collagen I levels expressed by LX-2 cells were tested by ELISA. The results showed that collagen I level was up-regulated by TGF- β 1, and the up-regulation of collagen I was inhibited by compounds **3b**, **3c**, **5a** and **5c**, which suggested that the above components could inhibit the expression of collagen I in LX-2 cells (Fig. 3).

Elevated expression levels of FBRS on LX-2 were inhibited by compounds

Fibrosin is one fibrogenic cytokine, and our previous studies showed that FBRS levels were closely related to the degree of hepatic fibrosis.⁵ In this study, some effective components including compounds **3b**, **3c**, **5a** and **5c** were further explored in the effects on FBRS expression levels. Our results showed that FBRS levels were up-regulated by TGF- β 1, and the up-regulation of FBRS was inhibited by compounds **3b** and **5a**, which suggested that compounds **3b** and **5a** might inhibit the progress of fibrosis (Fig. 4).

Up-regulation of α -SMA level in LX-2 was decreased by compound **5a**

Upon the liver injury, quiescent HSCs become activated and trans-differentiate into myofibroblast-like cells, which are characterized by enhanced cell growth and undergo profound phenotypic changes, including *de novo* expression of α -smooth muscle actin (α -SMA) and production of large amounts of collagens. We further screened the effects of all compounds on



Scheme 1 Synthesis of (Z)-4-(iodomethylene)-3-(2,2,2-trifluoroethyl)-3,4-dihydroquinolin-2(1H)-ones and fluorinated 3,3-disubstituted 2-oxindoles.



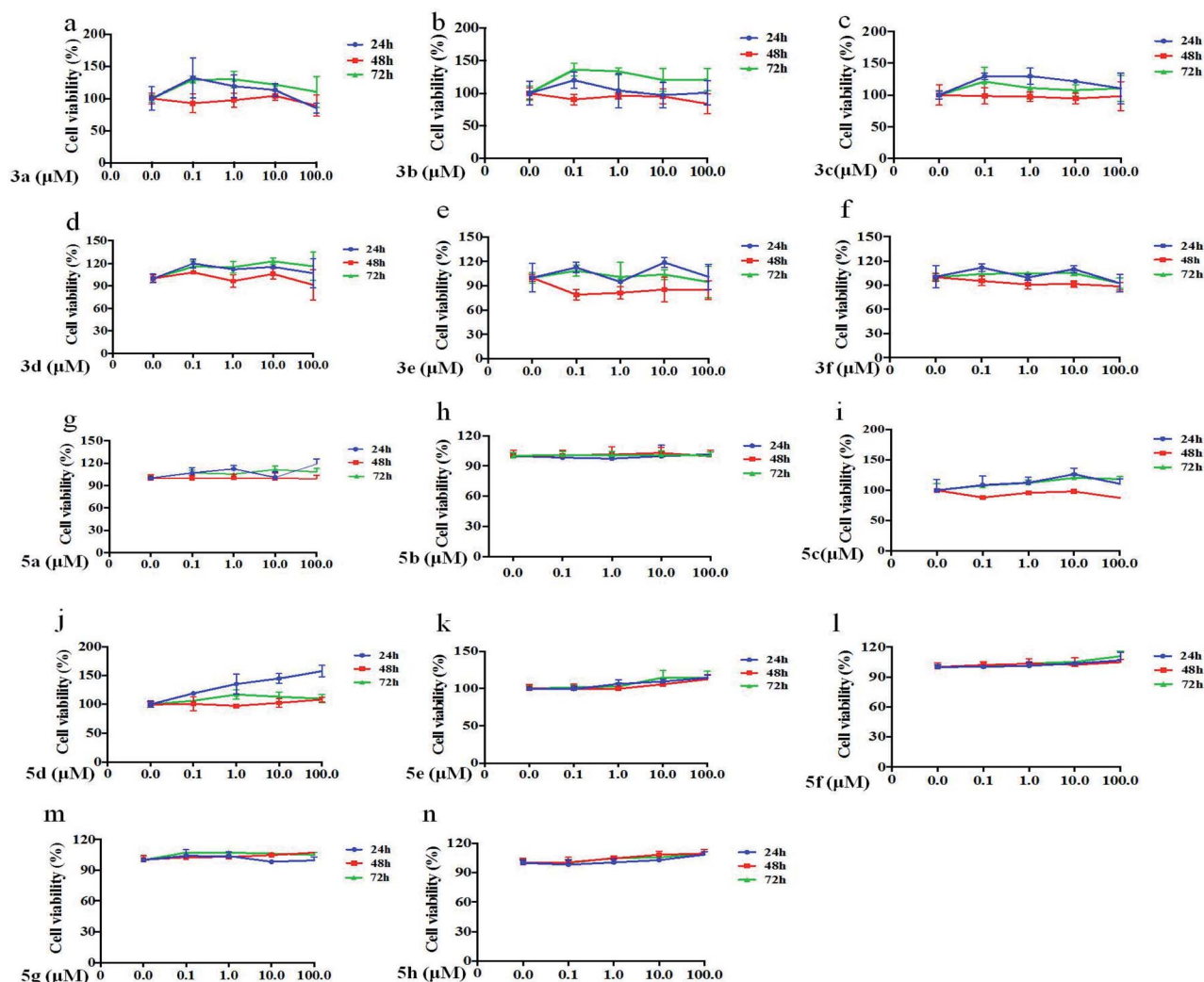


Fig. 2 Effects of chemical compounds on LX-2 viability. (a–n) LX-2 cells were incubated with chemical compounds at different concentrations and times, and then cell viabilities were tested by CCK-8 assay.

TGF- β 1/ α -SMA signaling. In this study, LX-2 cells were treated by the compounds including **3b** and **5a**, and then α -SMA mRNA levels in LX-2 cells induced by TGF- β 1 were detected. The result showed that α -SMA mRNA levels was up-regulated by TGF- β 1,

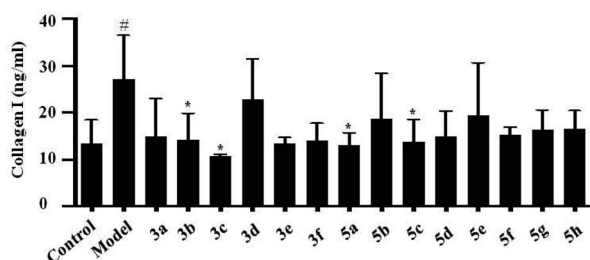


Fig. 3 Changes of collagen I expression levels induced by some compounds. LX-2 cells were incubated with TGF- β 1 and chemical compounds, and then collagen I levels in culture supernatant were verified by ELISA. Note: #, compared with control group, $P < 0.05$; *, compared with model group, $P < 0.05$.

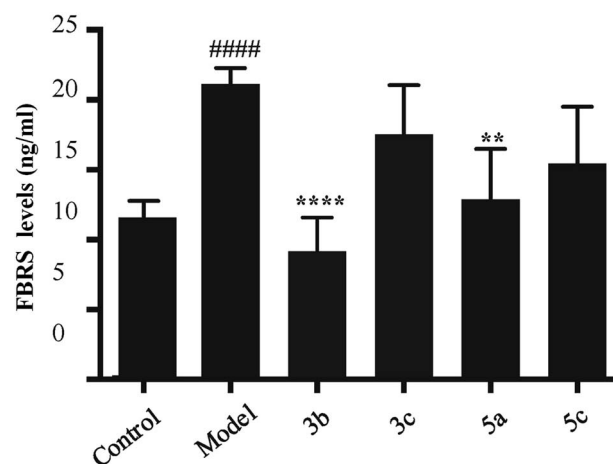


Fig. 4 Expression levels of FBS were inhibited by chemical compounds. LX-2 cells were incubated with TGF- β 1 and chemical compounds, and then FBS levels in culture supernatant were verified by ELISA. Note: ####, compared with control group, $P < 0.001$; *, compared with model group, $P < 0.05$; ****, compared with model group, $P < 0.001$.



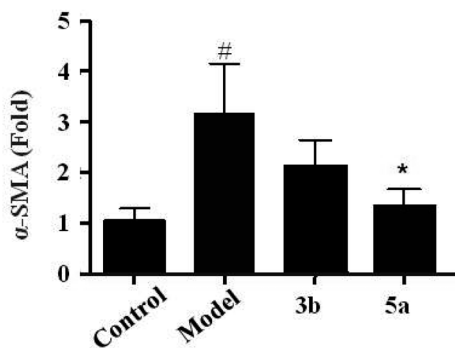


Fig. 5 α -SMA expression levels were inhibited by chemical compounds. LX-2 cells were incubated with TGF- β 1 and chemical compounds, and then α -SMA levels in culture supernatant were verified by ELISA. Note: #, compared with control group, $P < 0.05$; *, compared with model group, $P < 0.05$.

and the up-regulation of α -SMA was inhibited by compound 5a, which suggested that compound 5a could obviously inhibit the activation of LX-2 cells (Fig. 5).

Conclusions

In summary, liver fibrosis is a serious health problem worldwide, which is the final common pathway of chronic or iterative liver damage. The approaches for reversing fibrosis are the major unmet medical need. In this study, (Z)-4-(iodomethylene)-3-(2,2,2-trifluoroethyl)-3,4-dihydroquinolin-2(1H)-ones and fluorinated 3,3-disubstituted 2-oxindoles are synthesized and evaluated for anti-hepatic fibrosis. CCK-8 assay indicates that most of the compounds have no obvious cytotoxicity on human HSC cell line. Collagen I and FBRs expression levels are tested by ELISA, and the results show that several compounds can inhibit the expression of collagen I and FBRs. Additionally, results from real-time-PCR reveal that only compound 5a can inhibit the expression level of α -SMA, suggesting that this compound can inhibit the activation of HSC cell line. These studies demonstrate that compound 5a may be a potential novel drug candidate for anti-hepatic fibrosis (approximately 5–6 lines), which decreases the expression levels of collagen I and FBRs, and reduced the activation of HSC.

Experimental

General experimental procedure for the synthesis of (Z)-4-(iodomethylene)-3-(2,2,2-trifluoroethyl)-3,4-dihydroquinolin-2(1H)-ones

Benzene-tethered 1,7-enynes **1** (0.2 mmol) was added to a mixture of Togni reagent **2** (0.3 mmol), NaI (0.2 mmol) and PhCOOH (0.2 mmol) in DCE (4.0 mL). The mixture, placed around the mercury lamp (purchased from Yuming, Shanghai) with a distance of 10 centimeters, was stirred under UV irradiation (0.67 W cm⁻¹) for 12 hours at room temperature. After completion of reaction as indicated by TLC, the solvent was evaporated, and the residue was purified directly by flash column chromatograph (EtOAc/*n*-hexane, 1 : 8) to give the desired products **3**.

General experimental procedure for the synthesis of fluorinated 3,3-disubstituted 2-oxindoles

Fluorinated alkyl iodide (0.36 mmol) was added to a solution of *N*-arylacrylamides **4** (0.3 mmol), Na₂CO₃ (0.3 mmol) in CH₃CN (4.0 mL) under N₂ atmosphere. The mixture was stirred under ultraviolet irradiation for 12 hours. After completion of reaction as indicated by TLC, the solvent was evaporated, and the residue was purified directly by flash column chromatograph (EtOAc/*n*-hexane, 1 : 8) to give the desired products **5**.

Cell culture and experimental design

The human LX-2 cell line was a well characterized human HSC cell line, that could transdifferentiate *in vitro* from a quiescent-like phenotype to a more proliferative and activated behavior, and it provided a useful platform to assess anti-fibrotic drugs. LX-2 cells were donated by Institute of Liver Diseases Affiliated to Shanghai University of Traditional Chinese Medicine. LX-2 cells were cultured in Dulbecco's Modified Eagle's Medium (DMEM, Thermo Fisher Scientific), containing penicillin (100 U mL⁻¹), and streptomycin (100 μ g mL⁻¹) and fetal bovine serum at 10% final concentration. Cells were maintained at 37 °C in a humidified atmosphere of 5% CO₂, and the medium was changed once per day. When the cell density reached 80–90%, the cells were passaged. To evaluate the effects of chemicals on the activation of LX-2 cells, cells were cultivated in media containing 10% fetal bovine serum in the presence of TGF- β and chemicals.

Drug concentration

All chemicals were dissolved in DMSO at 0.05% (v/v). The concentrations of all chemicals were tested for the measurement of cell viability at 0.1 μ M, 1 μ M, 10 μ M and 100 μ M.

Cytotoxicity assay

The cytotoxicity analyses were performed by a Cell Counting Kit-8 (CCK-8; Dojindo Molecular Technologies, Gaithersburg, MD, USA). For the assay, 3×10^3 LX-2 cells were cultured in wells of 96-well plates for 12 h, followed by the incubation of different concentrations of chemicals (0.1 μ M, 1 μ M, 10 μ M and 100 μ M) for extra 72 h. Next, the cultured cell medium was collected, and CCK-8 solution (10 μ L) was added to the culture medium, and the cultures were incubated for 2 h at 37 °C. The absorbance was detected by Microplate Reader (Bio-Rad; Hercules, CA, USA) under the optical density (OD) at 450 nm.

Activation of LX-2 cells

To activate LX-2 cells, TGF- β was added to the culture medium at the final concentration of 10 μ g mL⁻¹.

ELISA

Collagen I and fibrosin levels in culture medium were tested by ELISA. Human collagen I ELISA kit was purchased from Abcam (ab210966). Human fibrosin ELISA kit was purchased from Shanghai Ji Ning Technology Co., Ltd. In brief, 100 μ L of sample



was added into every well. After incubating 2 h at room temperature, the plate was washed and 100 μ L antibody was added to each well. After incubating 1 h at room temperature, the plate was washed and 200 μ L substrate solution was added to each well. After incubating for 20 minutes at room temperature, 50 μ L of stop solution was added to each well. The absorbance was detected by Microplate Reader (Bio-Rad; Hercules, CA, USA) under the optical density (OD) at 450 nm.

RNA extraction and real-time-PCR analyses

Total RNA was isolated from LX-2 cells using Trizol Reagent (Invitrogen) and quantified. RNA was synthesized from 1 μ g of total RNA using cDNA Reverse Transcription kit (Invitrogen). Real-time PCR was performed using the Mastercycler ep real-plex 4 real-time PCR system (Eppendorf) with an SYBR Green qPCR Master Mix (Fermentas) according to the manufacturer's protocol. The sequences of primers for human α -SMA (NC_000010.11) were 5'-CCACCGCAAATGCTTCTAAGT-3' (forward) and 5'-GGCAGGAATGATTGGAAAGG-3' (reverse). The primers for human GAPDH (NM_002046.3) were 5'-TGCAC-CACCAACTGCTT AGC-3' (forward) and 5'-GGCATG-GACTGTGGTCATGAG-3' (reverse). The amplification conditions were as follows: 95°C for 10 min, and 40 cycles of 95°C for 15 s, 64°C for 30 s and 72°C 20 s. The amount of α -SMA transcripts of individual samples was normalized to GAPDH.

Conflicts of interest

There are no conflicts to declare.

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Notes and references

- (a) T. Tsuchida and S. L. Friedman, *Nat. Rev. Gastroenterol. Hepatol.*, 2017, **14**, 397; (b) T. Tsuchida and S. L. Friedman, *Nat. Rev. Gastroenterol. Hepatol.*, 2017, **14**, 397; (c) M. Giannandrea and W. C. Parks, *Dis. Models Mech.*, 2014, **7**, 193.
- (a) R. Lichtinghagen, D. Pietsch, H. Bantel, M. P. Manns, K. Brand and M. J. Bahr, *J. Hepatol.*, 2013, **59**, 1365; (b) H. Sekiguchi, N. Hemmi, T. Maki, A. Ozawa, E. Kadowaki, J. Kamiie, M. Yamamoto, K. Arishima and M. Sakaue, *J. Vet. Med. Sci.*, 2013, **75**, 553; (c) M. J. Yum, S. Koppula, J. S. Kim, G. M. Shin, Y. J. Chae, T. Yoon, C. S. Chun, J. D. Lee and M. D. Song, *Pharm. Biol.*, 2017, **55**, 1577; (d) S. L. Friedman, F. J. Roll, J. Boyles and D. M. Bissell, *Proc. Natl. Acad. Sci. U. S. A.*, 1985, **82**, 8681; (e) J. J. Maher and R. F. McGuire, *J. Clin. Invest.*, 1990, **86**, 1641.
- (a) J. J. Maher, D. M. Bissell, S. L. Friedman and F. J. Roll, *J. Clin. Invest.*, 1988, **82**, 450; (b) S. L. Friedman, *J. Biol. Chem.*, 2000, **275**, 2247.
- (a) D. J. Wyler, *Int. Arch. Allergy Immunol.*, 1996, **111**, 326; (b) S. Prakash, P. W. Robbins and D. J. Wyler, *Proc. Natl. Acad. Sci. U. S. A.*, 1995, **92**, 2154.
- M. Li, Z. J. Zeng, X. H. Sun, X. J. Zhu, Z. H. Zhou and Y. Q. Gao, *J. Clin. Hepatol.*, 2011, **27**, 1312.
- (a) A. Armstrong and J. C. Collins, *Angew. Chem., Int. Ed.*, 2010, **49**, 2282; (b) A. R. Katritzky and S. Rachwal, *Chem. Rev.*, 2010, **110**, 1564; (c) M. A. P. Martins, C. P. Frizzo, D. N. Moreira, L. Buriol and P. Machado, *Chem. Rev.*, 2009, **109**, 4140; (d) E. Ruijter, R. Scheffelaar and R. V. A. Orru, *Angew. Chem., Int. Ed.*, 2011, **50**, 6234.
- (a) S. Purser, P. R. Moore and S. Swallow, *Chem. Soc. Rev.*, 2008, **37**, 320; (b) D. O'Hagan, *Chem. Soc. Rev.*, 2008, **37**, 308; (c) K. Hagmann, *J. Med. Chem.*, 2008, **51**, 4359; (d) Y. Y. Zhou, J. Wang, Z. Gu, S. Wang, W. Zhu, J. L. Aceña, V. A. Soloshonok, K. Izawa and H. Liu, *Chem. Rev.*, 2016, **116**, 422.
- Y. An, Y. Kuang and J. Wu, *Org. Chem. Front.*, 2016, **3**, 994.
- Y. An, Y. Li and J. Wu, *Org. Chem. Front.*, 2016, **3**, 570.
- C. Y. Zhang, W. G. Yuan, P. He, J. H. Lei and C. X. Wang, *World J. Gastroenterol.*, 2016, **22**, 10512.

