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# Size and density adjustment of nanostructures in nanochannels for screening performance improvement†

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Biomimetic solid-state nanochannel/nanopore with flexible geometric structures, mechanical robustness and multifunctional surfaces have attracted extensive attention in separation, catalysis, drug delivery and other fields. Nanostructures have been introduced in nanoconfinement to compress substances pass through for high-efficient screening. However, precise controls of the nanostructure's growth in nanoconfinement is rare. Herein, we developed a method to control size and number density of nanoparticles in nanochannels by adjusting polydopamine reducing conditions, achieving (1) particle size increasing, density increasing; (2) particle size increasing, density decreasing; (3) particle size increasing, density invariant; (4) particle size invariant, density increasing. The nanoparticles compressed the space of functional molecules decorated on them. Increasing size and density of nanoparticle enhanced the steric hindrance of functional molecules decorated on them and improved the wetting and chirality screening through nanochannels.

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Biological pores are widespread in the membrane structure of organisms, which play a vital role in substances transport, energy transmission, signal exchange and other life processes.<sup>1–6</sup>

It is well-known that the high-efficient screening properties of biological pores stem from the numerous protein synaptophysin with fine structures and the precise functions on the surface of biological pores.<sup>7–10</sup> Inspired by biological pores, solid-state nanochannels (SSNs) have been developed rapidly and prosperously, which show the superior flexibility in terms of geometry, robustness, surface chemical diversity.<sup>11–14</sup> Functional elements (FEs) have been further introduced to improve the sensibility and selectivity of SSNs due to their strong effects on steric hindrance, electrostatic field and wettability in the confined channels at nanometric,<sup>15,16</sup> which are widely applied in sensing,<sup>17–19</sup> catalysis,<sup>20</sup> drug delivery,<sup>21</sup> and energy conversion.<sup>22–24</sup> Simultaneously, the geometry structure of SSNs affects

the performance of FEs. Kang *et al.* indicated that an improvement in the selectivity of K<sup>+</sup> and Na<sup>+</sup> by tuning pore diameter of graphite in combination with applying voltage.<sup>25</sup> Zhao *et al.* designed a Cu<sup>2+</sup> detection by using an asymmetric polyglutamic acid-modified nanochannel with the smaller pore diameter approximate to hydrate Cu<sup>2+</sup> radius.<sup>26</sup> More recently, researches focused on the performance improvement of SSNs by introducing nanostructures for loading FEs and further compressing the space in between FEs and the inner wall of SSNs.<sup>27–30</sup> For instance, Omar *et al.* developed a method to grow Au nanoparticles (AuNPs) reduced by polydopamine (PDA) in the nanopores which improved the rectification of a nanofluidic diode in a pH-dependent manner.<sup>31</sup> Tang *et al.* reported a novel strategy to modify AuNPs with magnetic Fe<sub>3</sub>O<sub>4</sub> before decorating aptamer on SSNs for sensing carcinoembryonic antigen.<sup>32</sup> We further used the PDA-reduced AuNPs to decrease the pore diameter down to 10 nm locally. The compressed space enhances the interaction between DNA probes on AuNPs and DNA targets pass-by, that the single-base mismatch recognition of linear DNA with high specificity.<sup>33–35</sup> It can be expected that the regulation of nanostructures in SSNs, such as size, geometry, surface chemistry and number density, will lead to more efficient regulation on substances transport by FEs. However, till now, the related studies have been rare.

Herein, we reported a method to control size and number density of PDA-reduced AuNPs in anodic aluminum oxide (AAO) SSNs by adjusting PDA reducing conditions. We obtained the SSNs modified with higher number density of AuNPs with

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bigger size by increasing PDA concentration in precursor and HCl concentration for the pre-etching of AAO surface. In the SSNs, the interactions between FEs and foreign substances on the AuNPs was further enhanced through the increase of AuNPs size and number density, which discriminated the transport of hydrophobic and hydrophilic molecules using hydrophobic FEs. Meanwhile, the transport of bull serum albumin (BSA) in a D-Cys and L-CyS modified SSNs was distinguished.

We investigated the reducing condition of PDA on the size and number density of AuNPs in SSNs, including (1) PDA concentration, (2) ambient temperature, (3) hydrochloric acid concentration and (4) filtration duration (Fig. 1a, Table S1, ESI†). The size of AuNPs varying monotonically or synchronically, are precisely regulated by adjusting the reaction conditions within the AAO channels with the pore diameter of  $85 \pm 15$  nm (Fig. S1–S6, ESI†). Specifically, on the one hand, we can synchronically control the particle size and the number density of AuNPs on the inner wall of AAO membrane. Specifically, by increasing the PDA concentration, the size of AuNPs increases slightly, then decreases, while the number density of AuNPs increases as shown in Fig. 1b. Or, by raising the ambient temperature, the particle size and the number density of the AuNPs increases (Fig. 1c). On the other hand, we also can decorate the AAO membrane with the particle size or number density monotonically. By prolonging the filtration duration, the particles increase, while the number density keeps no change (Fig. 1d). The PDA in nanochannels serve as reductant

and reduce  $\text{HAuCl}_4$  into AuNPs. By prolong the filtration duration, the size of AuNPs changes from 44.8 nm to 74.8 nm. The possible reason is that the long-time filtration will offer more enough PDA in nanochannel and realize sufficient reduction reaction of AuNPs. Thus, the large AuNPs emerge in nanochannels. Furthermore, by increasing the concentration of HCl, the number density of AuNPs increases, while the size of AuNPs has nearly no change as shown in Fig. 1e. In the present work, the variation of the AuNPs' size and number density would be realized by coordinating adjustment these four experimental conditions, from 30 nm to 80 nm and  $10^9$  to  $10^{12} \text{ m}^{-2}$ , respectively. According to the results, we found the appropriate preparation conditions for larger and more AuNPs that were  $2 \text{ mg mL}^{-1}$  PDA concentration,  $V_{\text{HCl}} : V_{\text{H}_2\text{O}} = 1 : 2$  and  $25^\circ\text{C}$  ambient temperature (this method is used below).

We next investigated the impact from the AuNPs on the distribution of FEs using laser confocal scanning microscopy (LCSM). The DNA was used as FEs due to their programmable structures and modification by numerous functional groups. The SSNs were firstly immersed in the buffer containing  $1 \mu\text{M}$  Cy5-modified DNA for overnight (Sequence 1 in Table S2, ESI†). Then, the as-prepared SSNs were rinsed thoroughly. As shown in Fig. 2a, in a SSN without any AuNPs, nearly all SSNs were dark. When using Cy5-modified DNA without the thiol ( $-\text{SH}$ ), shown in the fluorescence 2a, exists only at the outer surface of SSNs.

In contract, when using Cy5-modified DNA with thiol, the most of SSNs were lighted by Cy5 fluorescence, which indicated the immobilization of FEs at the AuNPs through specific Au–S interactions. In the SSN with higher number density and larger AuNPs, red domain covers nearly the whole region of the SSNs. It demonstrated that the thiol-modified FEs increased with the number density of AuNPs in SSNs.

The interactions between FEs and foreign substances taken place on the AuNPs were investigated taking the hybridization between thiol-modified DNA and their perfectly matched DNA chains as foreign substances as an example. Electrochemical Impedance Spectroscopy (EIS) is applied to characterize the variation of steric hindrance, in virtue of the higher sensitivity to the DNA grafting and hybridization than the  $I$ - $V$  characterization.<sup>33</sup> A typical two-electrode system was carried out and the equivalent circuit to simulate the impedance spectra was listed (Fig. S7, ESI†). Here, we set the charge-transfer resistances ( $R_{\text{ct}}$ ) of AAO–AuPNs (AAO–A) immobilized with single-stranded DNA (AAO–A–ss) as 100%. The real values are shown in Fig. S7 (ESI†). Interestingly, after the hybridization between ss-DNA and their perfectly matched DNA chains (AAO–A–ds (PM)), the increment of  $R_{\text{ct}}$  is 101% for the AAO membrane with higher density of bigger AuNPs, which is greater than the increment of  $R_{\text{ct}}$  as 69% for the AAO membrane with lower density of smaller AuNPs. Obviously, the larger size and higher density of AuNPs enhanced the interactions between FEs and foreign substances taken place on the AuNPs in SSNs.

SSNs are an important platform for substances screening, on account of that their narrow spaces enhance the interaction with the foreign substances. We investigated the screening property of the SSNs with larger size and higher number density

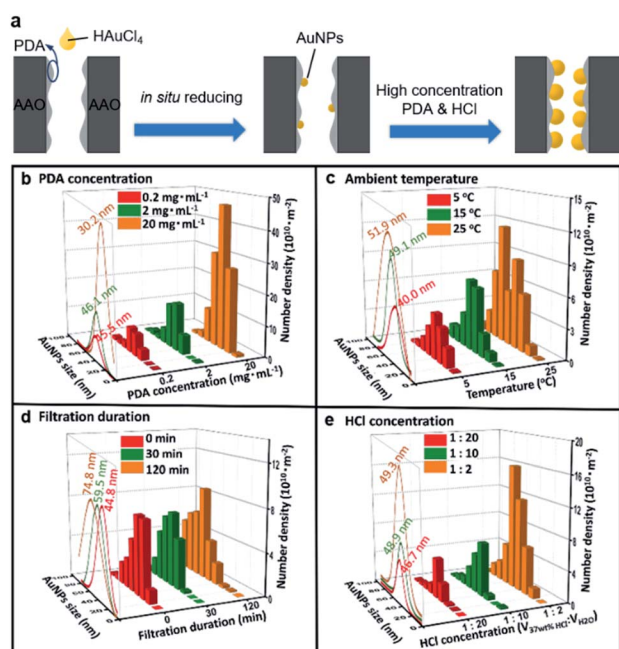


Fig. 1 (a) Schematic illustration of controlling the particle size and number distributions of AuNPs. Three-dimensional charts exhibiting the variation of number density and particle size by four kinds of reaction conditions as (b) PDA concentration, (c) ambient temperature, (d) filtration duration and (e) HCl concentration. The curve of the AuNPs size distribution along the number density projected on the Y–Z plane. The annotation inset as the average particle sizes of each sample under different reaction conditions.



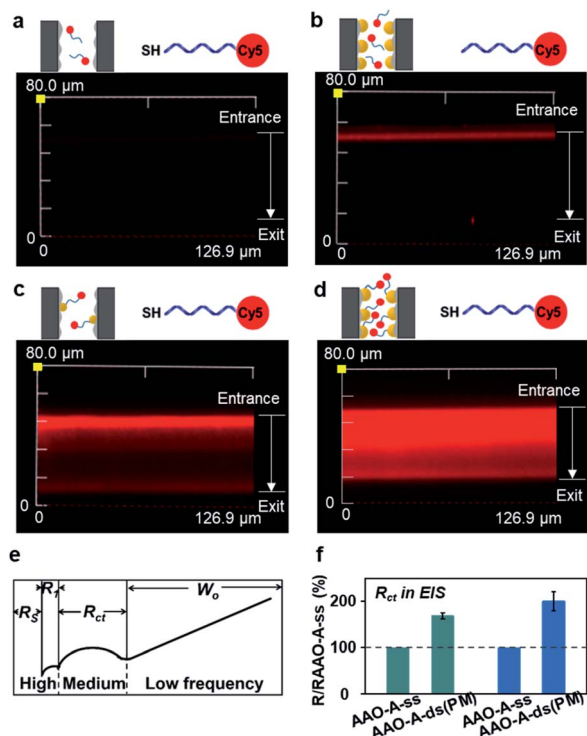


Fig. 2 The fluorescence images of AuNPs-decorated AAO membrane characterized by the LCSM. (a) AAO membrane without AuNPs decoration impregnated by Cy5-modified DNA probe. (b) AuNPs-decorated AAO membrane impregnated by Cy5-modified DNA probe without  $-SH$  modification. (c) Low AuNP density decorated AAO membrane impregnated by Cy5-modified DNA probe. (d) High AuNP number density decorated AAO membrane impregnated by Cy5-modified DNA probe. (e) The EIS sketch labels the impedance elements in different frequency. (f) Fractional comparison of impedances ( $R_{ct}$ ) in the EIS of the AAO membrane tested after sequent modification. Green columns are low AuNPs density and blue columns are high AuNPs density.  $R_{AAO-A-ss}$  correspond to the impedance of AAO membrane with AuNP decoration grafting single sequence DNA (set as 100%). Error bars are added according to the EIS characterizations of nine different AAO membranes.

AuNPs. The AuNPs were modified with hydrophobic alkane. We investigated the concentration of hydrophilic methylene blue (MB) and hydrophobic amitriptyline ( $C_{22}H_{46}S$ ). Schematic of our permeation experiment is shown in Fig. S8.† We applied the +2 V drive voltage to the permeate and the feed compartment, and the directionality drive lasted 40 min. We tested the ultra-violet (UV) absorption value of MB and  $C_{20}H_{23}N$  in permeate compartment, using the curve of standard concentration to calculate the concentrations (Fig. S9 and S10, ESI†). As can be seen in Fig. 3a, MB diffuses through AAO-A faster than AAO-A- $C_{22}H_{46}S$ , and  $C_{20}H_{23}N$  in reverse. It illustrated that the transmission of hydrophilic molecules in AAO was inhibited, and the transmission of hydrophobic molecules was promoted after the secondary modification of hydrophobic molecules, which improved the screening properties.

To verify the screening property, we did another experiment that fluorescein isothiocyanate-bovine serum albumin (FITC-BSA) diffused through AuNP-decorated AAO membrane with

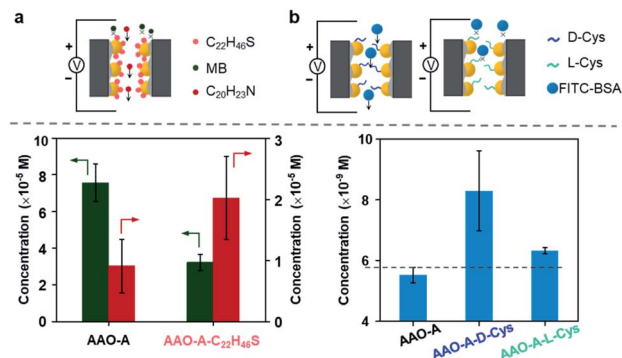


Fig. 3 The transport performance of AuNPs-decorated AAO membrane with secondary modification of FEs. (a) The permeate compartment concentration of MB (green) and  $C_{20}H_{23}N$  (red) through AAO-A membrane and AAO-A- $C_{22}H_{46}S$  membrane. (b) The permeate compartment concentration of FITC-BSA (blue) through AAO-A membrane, AAO-A-D-Cys membrane and AAO-A-L-Cys membrane respectively. Error bars are added according to the MB,  $C_{20}H_{23}N$  and FITC-BSA concentration of three different membranes, respectively.

secondary modification of L/D-cysteine (L/D-Cys).<sup>36</sup> We also tested the UV absorption value of FITC-BSA in permeate compartment, using the curve of standard concentration to calculate the concentrations (Fig. S11, ESI†). FITC-BSA diffuses through AAO ( $\approx 6.5$  nM) faster than AAO-A ( $\approx 5.5$  nM) in Fig. 3. The increasing steric hindrance of AAO membranes after AuNPs modification blocked FITC-BSA diffusing directionally. However, FITC-BSA diffuses through AAO-A-L-Cys ( $\approx 8.0$  nM) and AAO-A-D-Cys ( $\approx 6.4$  nM) faster than AAO-A. Interestingly, we also found that FITC-BSA diffusion showed the selection of chirality in AAO-A-L/D-Cys.

In conclusion, we reported a strategy for controlling size and number density of AuNPs decorated on the inner walls of nanochannels. The appropriate preparation conditions are  $2 \text{ mg mL}^{-1}$  PDA concentration,  $V_{HCl} : V_{H_2O} = 1 : 2$  and  $25^\circ \text{C}$  ambient temperature, which can get higher number density AuNPs with larger size. It can impact on the distribution of FEs and the interaction between FEs and foreign substances taken place on the AuNPs. By introducing larger and more AuNPs into nanochannels, it can increase the steric hindrance of channels, in order to increase the probability of the interactions between FEs and the foreign substances. It achieves the purpose of screening performance improvement. Through further modification of hydrophobic and chiral molecule on the AuNPs, the SSNs show the selection of wetting and chirality in SSNs, respectively. Such a strategy offers a platform for high-efficient interactions in a nanoconfine.

## Conflicts of interest

There are no conflicts to declare.

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