



 Cite this: *RSC Adv.*, 2021, 11, 4805

 Received 7th December 2020  
 Accepted 26th December 2020

DOI: 10.1039/d0ra10273c

[rsc.li/rsc-advances](http://rsc.li/rsc-advances)

# Two methods for the preparation of sitagliptin phosphate *via* chemical resolution and asymmetric hydrogenation†

 Fei Ye, \* Zhifeng Zhang, Wenxia Zhao, Jianhai Ding, Yali Wang and Xueyan Dang

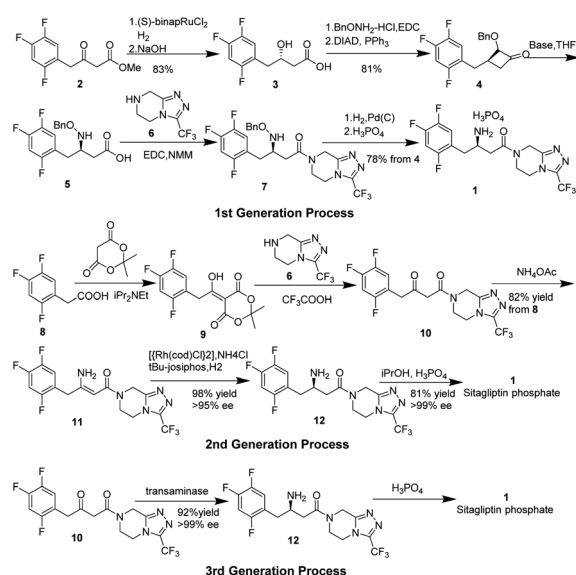
Two effective processes have been developed for the preparation of sitagliptin phosphate. The approach of chemical resolution obtained *R*-sitagliptin in five steps from commercially available starting materials using the inexpensive NaBH<sub>4</sub> to reduce the enamine and then using (–)-di-*p*-toluoyl-*L*-tartaric acid to resolve racemates in 11% yield overall. The route successfully avoids the use of expensive noble metal as catalysts compared with traditional synthesis methods, resulting in greatly reduced costs and simplified synthetic routes. Other alternative asymmetric hydrogenation of β-ketomide routes for the synthesis of sitagliptin were found, two of the intermediates were synthesized for the first time.

## Introduction

Type 2 diabetes<sup>1–4</sup> mellitus is a commonly and rapidly growing disease, with millions of new cases reported annually worldwide. Diabetes has become a serious threat to human health and brings enormously high economic costs, not only to individuals and families, but also to the global health system.<sup>5</sup> It is estimated that approximately 5 million people died from diabetes and its complications in 2015, representing one of the leading causes of death globally.<sup>6–8</sup> Dipeptidyl peptidase-4 (DPP-4) inhibitors, enzymes that act to degrade and inactivate glucagon-like peptide-1 (GLP-1), are a novel class of anti-hyperglycemic agents for the treatment of Type 2 diabetes mellitus. GLP-1 is an incretin hormone that stimulates insulin secretion and biosynthesis, inhibits glucagon release in a glucose-dependent manner, and also aids gastric emptying and reducing appetite.<sup>9–11</sup> Sitagliptin,<sup>12–15</sup> which was originally known as MK-0431 under the trade name Januvia, is a chiral beta-amino acid derivative that contains both a trifluorophenyl moiety and a trifluoromethylated triazole,<sup>16</sup> and also is the first agent developed in the class of DPP-4 inhibitors approved by the FDA in 2006. Because of its safety, efficiency, and being well-tolerated in various treatment regimens, it has tremendous commercial value (more than 11.4 billion dollars annually since 2018)<sup>17</sup> and thus has captured the interest of many pharmaceutical companies and research groups.

Although researchers from Merck have reported three generations of processes for the preparation of sitagliptin on a large scale, it does not diminish the interest of other research

groups in sitagliptin. The key problem with the synthesis of sitagliptin is how to install optically pure chiral central, β-amino acid. In Merck's first-generation synthesis process (Scheme 1),<sup>18</sup> starting from achiral β-keto ester **2**, chirality was introduced in form of a hydroxyl group in β-hydroxy acid **3** through a ruthenium-catalyzed asymmetric hydrogenation.<sup>19–21</sup> The hydroxyl group of **3** was transformed into protected amino acid **5** through several steps, followed by direct coupling to the triazolopiperazine **6** afforded sitagliptin **1** after cleavage of the *N*-benzyloxy group and salt formation. Despite the overall yield being quite high at 52%, multiple steps and isolations resulted in large amounts of waste and poor atom-economy.



Scheme 1 Merck's three generations synthesis processes.

School of Chemistry and Chemical Engineering, Ningxia Normal University, Guyuan, 756000, China. E-mail: yefei0310@outlook.com; yefeiqq@163.com

† Electronic supplementary information (ESI) available. See DOI: 10.1039/d0ra10273c

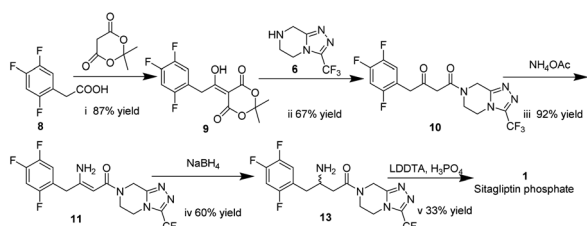


In their second-generation process,<sup>22</sup> dehydrostagliptin **11**<sup>23</sup> was firstly synthesized by a three-step one-pot process starting from trifluorophenyl acetic acid **8**, which contained within its structure the entire carbon skeleton of **1**. Crystallization and a simple filtration could give **11** in 82% overall yield with 99 wt% purity. The researchers then used the rhodium complex of *t*-Bu JOSIPHOS to asymmetrically hydrogenate the unprotected enamine amide **11** to install the chiral amine center into **12** in 98% conversion along with up to 95% ee.<sup>23,24</sup> Finally, sitagliptin **1** was recrystallized and isolated as its phosphate monohydrate salt in 84% yield with an upgrade in enantiomeric excess up to 99% ee.<sup>25</sup> Although this 2nd generation process for the sitagliptin manufacture has led to a significant reduction in waste as compared to the 1st generation process, its endgame leftover room for improvement because of the inherent drawbacks of utilizing an expensive transition metal mediated hydrogenation step. This process needs the specialized high-pressure equipment and a process for the complete removal of the precious metal from the product stream, both of which were significant cost drivers.

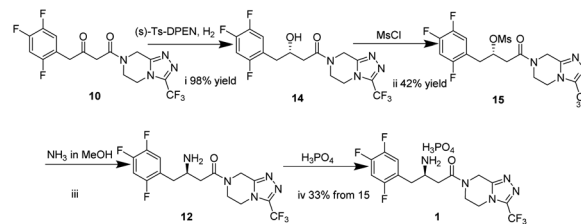
To circumvent these drawbacks, Merck and Codexis researchers have recently developed a highly efficient biocatalytic process (the 3rd generation process)<sup>26</sup> to prepare sitagliptin **1**. In the presence of the R-selective transaminase enzyme, direct amination of prostagliptin ketone to enantiomeric pure sitagliptin **1** (up to 99% ee) was carried out in multipurpose under mild conditions, and thus eliminated all the drawbacks of the 2nd generation process. Not only does the 3rd generation process lead to a reduction of the total waste produced (19%) as compared to the 2nd generation process, but also increase the overall yield (13%) and productivity (53%). This biocatalytic route should become an important auxiliary asymmetric synthesis in a pilot-scale production.

Besides, recently, several synthesis methods of sitagliptin have been reported on laboratory scale, including biocatalytic asymmetric reductive amination of  $\beta$ -ketamide,<sup>26,27</sup> enzymatic resolution of racemates,<sup>28</sup> chemical kinetic resolution,<sup>16,29</sup> and stereoselective reduction of an enamine derivative using NaBH<sub>4</sub>.<sup>30,31</sup> Although some of these synthetic routes are very efficient, given the enormous industrial interest, an inexpensive, convenient, industrially viable, and greener synthetic route is still in urgent need.

In this work, we report for the first time the process of chemical resolution of racemates **13** to sitagliptin **1** followed by reduction of enamine **11** using NaBH<sub>4</sub> (Scheme 2) and the



Scheme 2 The process for the preparation of sitagliptin phosphate via chemical resolution.



Scheme 3 The process for the preparation of sitagliptin phosphate via asymmetric hydrogenation.

process of directly asymmetric hydrogenation of  $\beta$ -ketamide **10** to **14**, a chiral intermediate of sitagliptin (Scheme 3).

## Results and discussion

### Chemical resolution preparation of sitagliptin 1

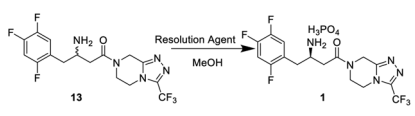
Meldrum's adduct **9** was firstly prepared from 2,4,5-trifluorophenyl acetic acid **8** utilizing the ability of Meldrum's acid to act as an acyl anion equivalent at 50 °C for 5 h in 87% yield. This process involves the activation of 2,4,5-trifluorophenyl acetic acid **8**, and we chose *N,N*-carbonyl di-imidazole (CDI)<sup>32,33</sup> as active agent instead of pivaloyl chloride<sup>34</sup> to simplify the experiment. **9** was then coupled to the 3-(trifluoromethyl)-1,2,4-triazolo-[4,3-*a*]piperazine hydrochloride **6** using *i*-Pr<sub>2</sub>NET, to provide  $\beta$ -ketamide **10** in 67% yield. Treatment of **10** with AcONH<sub>4</sub> and ammonia water results in formation of enamine **11**. As the reaction temperature was raised to 58 °C, the desired product **11** began to crystallize from the reaction mixture. Upon to the mild temperature, **11** was isolated as a white crystalline solid through simple filtration in 92% yield with 99 wt% purity. Next the reduction of enamine **11** was performed with NaBH<sub>4</sub> at -15 °C in 4.5 h, monitored by HPLC until enamine **11** disappeared. It was found that the reductive reaction of **11** and NaBH<sub>4</sub> could not be performed very well. Therefore, several

Table 1 Reductive reaction of enamine **11** to racemates **13**<sup>a</sup>

Entry	Reduction	Additive	Conversion (%)
1	NaBH <sub>4</sub>	None	0
2	NaBH <sub>4</sub>	BF <sub>3</sub> diethyl etherate	95
3	NaBH <sub>4</sub>	MsOH (methanesulfonic acid)	93
4	NaBH <sub>4</sub>	Acetic acid	10
5	NaBH <sub>4</sub>	TFA (trifluoroacetic acid)	2

<sup>a</sup> All reactions were carried out as follows: THF (50 mL) was firstly cooled to -10 °C. NaBH<sub>4</sub> (2.33 g, 61.68 mmol) and an additive such as MsOH (5.9 g, 61.68 mmol) were added dropwise at -10 to -5 °C, and then enamine **11** (5.0 g, 12.34 mmol) and isopropanol (30 mL) were added. The reaction mixture was aged at -15 °C for 4.5 h, monitored by HPLC. Isolated yield after ethyl acetate extraction.



Table 2 Effects of resolution agents on the configuration of (*R*)-**1**<sup>a</sup>


Entry	Resolution agent	Solvent	Configuration (ee%)
1	(-)-L-Tartaric acid	MeOH	4%
2	(-)-Di- <i>p</i> -toluoyl-L-tartaric acid	MeOH	96%

<sup>a</sup> All reactions were carried out using racemates **13** (0.5 g, 1.23 mmol) and resolution agents such as (-)-di-*p*-toluoyl-L-tartaric acid (0.24 g, 0.62 mmol) in solvent (15 mL) at 65 °C for 1 h. White sitagliptin tartrate was obtained after filtration. And tartrate was hydrolyzed by using ammonia water (10 mL), and extracted twice by ethyl acetate (30 mL × 2). The organic layer was concentrated and isopropanol (15 mL) was added. Then 85 wt% H<sub>3</sub>PO<sub>4</sub> was added dropwise and the reaction mixture was aged at 78 °C for 1 h to afford sitagliptin **1**.

additional chemicals were screened to improve the conversion of the reduction reaction.

As indicated in Table 1, these additives play a catalyst role in reaction mixtures, and significantly improve the conversion of the reductive reaction. BF<sub>3</sub> diethyl etherate yields the highest conversion (entry 2). However, because of the extreme toxicity of BF<sub>3</sub> diethyl etherate, we chose MsOH as the additive of the reductive reaction.

After the reductive reaction, the resolution of racemates **13** was carried out by treatment with di-*p*-toluoyl-L-tartaric acid in MeOH to form *R*-sitagliptin **1** in 33% yield with 99 wt% purity and 96% ee. At the same time, we compared the effects of the two L-tartaric acids on the resolution, as shown in Table 2. Lastly, the separated filtrate was treated to obtain *S*-sitagliptin phosphate with 90% ee.

### Asymmetric hydrogenation of β-ketoester **10** to sitagliptin **1** via three steps

The difficulty of this route (Scheme 3) lies in the asymmetric reduction of **10**. The carbonyl could be reduced by the solution of formic acid and Et<sub>3</sub>N in the presence of a chiral catalyst at room temperature in the literature.<sup>35,36</sup> Although the reaction mixture of **10** and formic acid were heated to 80 °C, the reaction did not proceed. The β-ketoester **10** was hydrogenated by hydrogen under 435 psi at 65 °C for 12 h to afford (*S*)-3-hydroxy-1-(3-(trifluoromethyl)-5,6-dihydro[1,2,4]triazolo[4,3-*a*]pyrazin-7(8*H*)-yl)-1-(2,4,5-trifluorophenyl)butan-1-one **14**. Treatment of **14** with MsCl resulted in (*S*)-4-oxo-4-(3-(trifluoromethyl)-5,6-dihydro-[1,2,4]triazolo[4,3-*a*]pyrazin-7(8*H*)-yl)-1-(2,4,5-trifluorophenyl)butan-2-yl methanesulfonate **15** in 42% yield with 98 wt% purity and 7% ee. And then methanesulfonate **15** reacted with NH<sub>3</sub> in MeOH under 145 psi to form *R*-sitagliptin **1** in 33% yield with 99 wt% purity and 7% ee. Although this approach failed to afford adequate results, an alternative route of the synthesis of sitagliptin was developed, **14** and **15** were firstly synthesized as new substrates.

## Conclusion

In this study, we established two new efficient routes of the synthesis of sitagliptin phosphate. The approach of chemical resolution of racemates **13** to sitagliptin **1** not only successfully obtained *R*-sitagliptin with up to 96% ee but also afforded *S*-sitagliptin with 90% ee by using chiral resolution agent, (-)-di-*p*-toluoyl-L-tartaric acid. Reduction of enamine **11** and chemical resolution of racemates **13** as two key steps, makes the synthetic route simple and practical and greatly reduce the cost.

The approach of asymmetric hydrogenation of β-ketoester **10** to **1** using [Ru(cymene)Cl]<sub>2</sub> and (*s*)-Ts-DPEN as a chiral catalyst to directly hydrogenate **10** and reacts with MsCl to form the methanesulfonate **15**, a key intermediate, and finally successfully obtained *R*-sitagliptin **1**. Although this approach failed to afford idealized optical purity results, an alternative route of the synthesis of sitagliptin was found, compound **14** and **15** were firstly synthesized as new substrates.

## Experimental section

### General

All of the reagents and solvents were purchased from commercial sources and used without purification. Melting points were determined on a Reichert micro-hot-stage apparatus. The IR spectra were recorded on an FSM-2201 spectrometer with Fourier transform from samples dispersed in mineral oil. High resolution mass spectra were recorded in ESI mode using magnetic sector mass analyzer and TOF mass spectrometer. <sup>1</sup>H and <sup>13</sup>C spectra were recorded with a 400 MHz NMR instrument at 295 K. Chemical shifts are reported relative to the residual signals of tetramethylsilane in CDCl<sub>3</sub> or deuterated solvent CDCl<sub>3</sub>/DMSO-*d*<sub>6</sub> for <sup>1</sup>H and <sup>13</sup>C NMR spectroscopy, respectively. Multiplicities are reported as follows: singlet (s), doublet (d), broad singlet (br s), doublet of doublets (dd), doublet of triplets (dt), triplet (t), quartet (q), and multiplet (m). All of the reactions were monitored by HPLC or TLC. Tetrahydrofuran was distilled over sodium before use.

**5-[1-Hydroxy-2-(2,4,5-trifluorophenyl)ethylidene]-2,2-dimethyl-1,3-dioxane-4,6-dione 9.** 2,4,5-Trifluorophenyl acetic acid **8** (10.0 g, 52.60 mmol) was dissolved in dry THF (80 mL), activated by *N,N'*-carbonyl di-imidazole (CDI) (10.0 g, 57.86 mmol), and then Meldrum's acid (8.4 g, 57.86 mmol) was added at 50 °C and aged for 5 h. The reaction mixture changed from light yellow to orange, and was monitored by TLC. The reaction mixture was cooled to room temperature, and the prepared solution of isopropyl acetate/water in 1/1 was added and stirred for 0.5 h. And the pH of the mixture was adjusted to 2–3 with concentrated hydrochloric acid. The aqueous layer was separated, and the organic layer was washed with 0.1 M HCl. The organic layer was concentrated, flushed with IPAc, and the residue was slurried in 2 : 1 heptane/IPAc. The mixture was cooled in an ice-bath and filtered, and then the solid is rinsed with 2 : 1 heptane/IPAc. After drying, the Meldrum's acid adduct **9** (14.6 g, 99 wt%) was obtained as a white solid in 87% yield. Mp 121–123 °C; ESI-MS (*m/z*): (317, M<sup>+</sup> + H); IR (KBr), ν, cm<sup>-1</sup>: 3448, 3001, 1734, 1653, 1581, 1525, 1428, 1338, 1307, 1203, 1153,



1024, 922, 789.  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  15.49 (s, 1H), 7.15 (m, 1H), 6.95 (m, 1H), 4.45 (s, 2H), 1.76 (s, 6H).

**4-Oxo-4-[3-(trifluoromethyl)-5,6-dihydro[1,2,4]triazolo[4,3-*a*]pyrazin-7(8*H*)-yl]-1-(2,4,5-trifluorophenyl)butan-2-one 10.** 5-[1-Hydroxy-2-(2,4,5-trifluorophenyl)ethylidene]-2,2-dimethyl-1,3-dioxane-4,6-dione **9** (5.0 g, 15.81 mmol), 3-(trifluoromethyl)-1,2,4-triazolo-[4,3-*a*]piperazine hydrochloride **6** (3.6 g, 15.81 mmol), and isopropyl acetate (50 mL) were charged into a 250 mL three-neck flask. *i*-Pr<sub>2</sub>NEt (3 mL) was added dropwise at room temperature, and the reaction mixture was aged at 75–80 °C for 5 h. After gradually cooled to 30 °C, water (50 mL) was added and the mixture was transferred to a separatory funnel. The aqueous layer was separated and organic layer was washed three times with water (150 mL). The organic layer was concentrated under diminished pressure to 10 mL and *n*-heptane (50 mL) was added dropwise. The mixture was stirred for 2 h and filtered, the ketoamide **10** (9.2 g, 90 wt%) was obtained in 67% yield. ESI-MS (*m/z*): (407,  $\text{M}^+$  + H). IR (KBr),  $\nu$ ,  $\text{cm}^{-1}$ : 3060, 2925, 1725, 1653, 1519, 1435, 1330, 1270, 1207, 1154, 1061, 1014, 935, 882, 852, 771, 680.  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.13–6.99 (m, 1H), 6.95–6.91 (m, 1H), 5.21–4.90 (m, 2H), 4.24–4.11 (m, 4H), 3.94–3.83 (m, 4H).

**(Z)-3-Amino-1-(3-(trifluoromethyl)-5,6-dihydro-8*H*-[1,2,4]triazolo[4,3-*a*]pyrazin-7-yl)-4-(2,4,5-trifluorophenyl)but-2-en-1-one 11.** Ketoamide **10** (10.0 g, 24.63 mmol) was dissolved in MeOH (100 mL), and  $\text{NH}_4\text{OAc}$  (9.5 g, 123.13 mmol) and ammonia water (10 mL) was added. The reaction mixture was heated to 58 °C and stirred for 30 min, and white solids began to precipitate. After an additional 2 h age, the mixture was cooled to room temperature and filtered. The wet cake was washed with methanol (100 mL) and dried at ambient temperature in a vacuum oven to afford white crystalline **11** (9.2 g, 99 wt%) in 92% yield. Mp 196–198 °C; ESI-MS (*m/z*): (406,  $\text{M}^+$  + H); IR (KBr),  $\nu$ ,  $\text{cm}^{-1}$ : 3408, 3043, 2044, 1626, 1516, 1425, 1179, 1013, 886, 750.

### Racemates 13

THF (50 mL) was charged into a 250 mL three-neck flask and cooled to –10 °C.  $\text{NaBH}_4$  (2.33 g, 61.68 mmol) and MsOH (5.9 g, 61.68 mmol) were added dropwise at –10 to –5 °C, and then enamine **11** (5.0 g, 12.34 mmol) and isopropanol (30 mL) were added. The reaction mixture was aged at –15 °C for 4.5 h, monitored by HPLC. When the reaction was finished, water was added and ethyl acetate (60 mL  $\times$  2) was extracted twice. The organic layer was dried by  $\text{MgSO}_4$  and concentrated under diminished pressure. Oil racemates **13** (3.0 g, 78 wt%) was obtained in 60% yield.

### Sitagliptin 1

Racemates **13** (0.5 g, 1.23 mmol) was dissolved in methanol (15 mL) and solution of (–)-*di*-*p*-toluoyl-L-tartaric acid (0.24 g, 0.62 mmol) in isopropanol (15 mL) was added. Then the mixture was heated to 65 °C and stirred for 1 h, and white solids began to precipitate. The reaction mixture was cooled to ambient temperature and aged for an additional 2 h before filtration. The cake was recrystallized by methanol solution in water (1 : 1). Sitagliptin tartrate was obtained in 96% ee.

Sitagliptin tartrate was directly dissolved in water (30 mL) and the pH of the mixture was adjusted to 10–12 with 28 wt% ammonia water (10 mL), and extracted twice by ethyl acetate (30 mL  $\times$  2). The organic layer was concentrated and isopropanol (15 mL) was added. Then 85 wt%  $\text{H}_3\text{PO}_4$  was added dropwise and the reaction mixture was aged at 78 °C for 1 h. The slurry was then cooled to room temperature and aged for an additional 0.5 h before filtration. The wet cake was washed with 15 mL cold ethanol and dried at 40 °C in a vacuum oven to afford sitagliptin **1** (0.21 g, 99 wt%) in 33% yield and 96% ee. Mp 218–220 °C; ESI-MS (*m/z*): (408,  $\text{M}^+$  + H); IR (KBr),  $\nu$ ,  $\text{cm}^{-1}$ : 3196, 3027, 2861, 1649, 1521, 1453, 1167, 1093, 850.  $^1\text{H}$  NMR (400 MHz,  $\text{DMSO}-d_6$ )  $\delta$  7.29 (m, 1H), 7.16 (m, 1H), 4.94 (s, 2H), 4.30 (d,  $J = 17.6$  Hz, 1H), 4.27 (d,  $J = 17.6$  Hz, 1H), 4.00 (m, 2H), 3.12 (m, 2H), 2.99–3.08 (m, 1H), 2.92 (dd,  $J = 17.1$  Hz, 1H), 2.87 (m, 1H);  $^{13}\text{C}$  NMR ( $\text{CDCl}_3$ )  $\delta$  36.2, 38.0, 39.1, 40.0, 40.1, 41.6, 42.4, 43.2, 43.6, 48.5, 105.6, 118.2, 1119, 121.7, 143.7, 146.6, 148.8, 149.7, 150.4, 156.1, 170.2, 170.58.

**(S)-3-Hydroxy-1-(3-(trifluoromethyl)-5,6-dihydro[1,2,4]triazolo[4,3-*a*]pyrazin-7(8*H*)-yl)-1-(2,4,5-trifluorophenyl)butan-1-one 14.**  $[\text{Ru}(\text{cymene})\text{Cl}]_2$  (0.4 g, 0.58 mmol) and (S)-Ts-DPEN (0.6 g, 1.64 mmol) was charged into 500 mL flask under a nitrogen atmosphere. Degassed methanol was added and the mixture was stirred at room temperature for 1 h. The ketoamide **10** (10.0 g, 24.63 mmol) along with MeOH (100 mL) was charged into the mixture. The ketoamide **10** was hydrogenated under 435 psi at 65 °C for 12 h after the slurry was degassed three times, and monitored by HPLC. When the reaction finished, the reaction mixture was concentrated and Brown oily (S)-3-hydroxy-1-(3-(trifluoromethyl)-5,6-dihydro[1,2,4]triazolo[4,3-*a*]pyrazin-7(8*H*)-yl)-1-(2,4,5-trifluorophenyl)butan-1-one **14** (9.9 g) was obtained in 98% assay yield. ESI-MS (*m/z*): (409,  $\text{M}^+$ ).

**(S)-4-Oxo-4-(3-(trifluoromethyl)-5,6-dihydro[1,2,4]triazolo[4,3-*a*]pyrazin-7(8*H*)-yl)-1-(2,4,5-trifluorophenyl)butan-2-yl methanesulfonate 15.** The above (S)-3-hydroxy-1-(3-(trifluoromethyl)-5,6-dihydro[1,2,4]triazolo[4,3-*a*]pyrazin-7(8*H*)-yl)-1-(2,4,5-trifluorophenyl)butan-1-one **14** crude solution (10.0 g, 24.63 mmol) and ethyl acetate (100 mL) were charged into 250 mL three-neck flask and cooled to –10 °C. The trimethylamine (6 mL 50 mmol) was added and MsCl (18 mL 50 mmol) was added dropwise at –10 to –5 °C. The reaction mixture was aged at –10 °C for 2 h, and monitored by HPLC. When the reaction was finished, the mixture was filtered and the filtrate was concentrated to dryness to give a residue which was chromatographed on silica gel with the solution of ethyl acetate-petroleum ether (1 : 1) (150 mL). The eluant was cooled to –10 °C and stirred for 2 h, and white solids precipitated. After an additional 2 h age, the mixture was filtered. The wet cake was washed with ethyl acetate (50 mL) and dried at ambient temperature in a vacuum oven to obtain white crystalline **15** (5.1 g, 98 wt%) in 42% yield and 7% ee. Mp 157–159 °C; ESI-MS (*m/z*): (487,  $\text{M}^+$ ); IR (KBr),  $\nu$ ,  $\text{cm}^{-1}$ : 3044, 2934, 1658, 1511, 1443, 1338, 1172, 1015, 528.  $^1\text{H}$  NMR (400 MHz,  $\text{DMSO}-d_6$ )  $\delta$  7.54–7.44 (m, 2H), 5.19 (m, 1H), 4.98 (t, 1H), 4.92–4.84 (m, 1H), 4.24 (m, 1H), 4.12–4.08 (m, 1H), 3.96–3.94 (m, 2H), 3.09–3.04 (m, 3H), 3.02 (s, 3H), 2.94–2.83 (m, 1H).



## Sitagliptin 1

(S)-4-Oxo-4-(3-(trifluoromethyl)-5,6-dihydro-[1,2,4]triazolo[4,3-a]pyrazin-7(8H)-yl)-1-(2,4,5-trifluorophenyl)butan-2-yl methanesulfonate 15 (3.0 g, 6.17 mmol) and the prepared solution of NH<sub>3</sub> in MeOH (13 wt% 60 mL) were charged into the flask. The reaction mixture was heated to 65 °C and stirred for 8 h under 145 psi. MTBE (methyl *tert*-butyl ether) (50 mL) and 0.5 M H<sub>3</sub>PO<sub>4</sub> (50 mL) were added after concentrated. The pH of the mixture was adjusted to 8–9 with the solution of NaOH. The aqueous layer was separated, and the organic layer was concentrated and isopropanol (40 mL) and water (40 mL) were added. Then 0.5 M H<sub>3</sub>PO<sub>4</sub> (1 mL) was added dropwise and the mixture was heated to 70 °C and stirred for 2 h. The slurry was filtered after cooled to the room temperature. The wet cake was washed with 15 mL isopropanol and dried at 40 °C in a vacuum oven to obtain sitagliptin 1 (1.2 g, 99 wt%) in 33% yield and 7% ee. ESI-MS (*m/z*): (408, M<sup>+</sup> + H); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.00–7.25 (m, 5H), 4.61–4.91 (m, 2H), 4.31–4.36 (m, 1H), 3.84–3.99 (s, 2H), 2.78–3.82 (m, 2H), 2.66–2.72 (m, 1H), 2.59–2.64 (m, 1H), 2.51 (s, 2H); <sup>13</sup>C NMR (D<sub>2</sub>O) δ: 30.9, 33.7, 38.0, 41.8, 43.1, 48.2, 105.8, 115.4, 118.5, 120.9, 145.5, 148.7, 157.2, 170.2.

## Conflicts of interest

The authors declare that there are no conflicts of interest regarding the publication of this article.

## Acknowledgements

This work was partially supported by the Natural Science Foundation for Ningxia Province (Grants no. 2018AAC03235), Liupanshan Resources Engineering Technology Central (Grants no. HGZD19-10), the Research Award Fund for First-class Discipline Construction (Education Discipline) in Higher Education Institutions of Ningxia (Grants no. NXYLXK2017B11), Development of Science and Technology of Ningxia (Grants no. 2019BEB04019).

## References

- 1 For leading references, see: N. A. Thornberry and A. E. Weber, *Curr. Top. Med. Chem.*, 2007, 7, 557–568.
- 2 S. L. Gwaltney II and J. A. Stafford, *Annu. Rep. Med. Chem.*, 2005, 40, 149–165.
- 3 B. Ahren, M. Landin-Olsson, P. A. Jansson, M. Svenson, D. Holmes and A. Schweizer, *J. Clin. Endocrinol. Metab.*, 2004, 89, 2078–2084.
- 4 International Diabetes Federation (IDF), *Diabetes Atlas*, 2019, 9th edn, <http://www.idf.org/diabetesatlas>.
- 5 See: (a) [www.diabetes.org](http://www.diabetes.org); (b) <http://www.who.int>.
- 6 E. Adeghate, H. Kalasz, G. Veress and K. Tekes, *Curr. Med. Chem.*, 2010, 17, 517–551.
- 7 E. Adeghate, E. Feher and H. Kalasz, *Expert Opin. Invest. Drugs*, 2015, 24, 1–15.
- 8 F. Peng, Y. Chen and C. Chen, *J. Org. Chem.*, 2017, 82(17), 9023–9029.
- 9 A. Weber, *J. Med. Chem.*, 2004, 48, 4135–4141.
- 10 D. Drucker, *Expert Opin. Invest. Drugs*, 2003, 12, 87–100.
- 11 S. A. Miller and E. L. St. Onge, *Ann. Pharmacother.*, 2006, 40, 1336.
- 12 D. Kim, L. Wang, M. Beconi and G. J. Eiermann, *J. Med. Chem.*, 2005, 48, 141–151.
- 13 C. S. Shultz and S. W. Krska, *Acc. Chem. Res.*, 2007, 40, 1320–1326.
- 14 L. H. Xia, *Chin. J. New Drugs*, 2007, 16, 979–981.
- 15 R. Pathak and M. B. Bridgeman, *Pharmacol. Ther.*, 2010, 35, 509–513.
- 16 H. Mei, J. Han, K. D. Klika, K. Izawa, T. Sato, N. A. Meanwell and V. A. Soloshonok, *Eur. J. Med. Chem.*, 2020, 186, 111826.
- 17 S. Matej, F. Rok and G. Stanislav, *ACS Omega*, 2020, 5(10), 5356–5364.
- 18 K. B. Hansen, J. Balsells, S. Dreher, Y. Hsiao, M. Kubryk, M. Palucki, N. R. Rivera, D. Steinhuebel, J. D. Armstrong III, D. Askin and E. J. Grabowski, *Org. Process Res. Dev.*, 2005, 9, 634–639.
- 19 J. Xu, H. O. Ok and E. J. Gonzalez, *Bioorg. Med. Chem. Lett.*, 2004, 14, 4759–4762.
- 20 U. Schollkopf, U. Groth and C. Deng, *Angew. Chem., Int. Ed. Engl.*, 1981, 20, 798–799.
- 21 D. Kim, L. Wang, M. Beconi, G. J. Eiermann and M. H. Fisher, *J. Med. Chem.*, 2005, 48, 141–151.
- 22 K. B. Hansen, H. Yi, F. Xu, N. Rivera, A. Clausen and M. Kubryk, *J. Am. Chem. Soc.*, 2009, 131, 8798–8804.
- 23 F. Xu, J. D. Armstrong, G. X. Zhou and B. Simmons, *J. Am. Chem. Soc.*, 2004, 126, 13002–13009.
- 24 Y. Hsiao, N. R. Rivera, T. Rosner and S. W. Krska, *J. Am. Chem. Soc.*, 2004, 126, 9918–9919.
- 25 A. M. Clausen, B. Dziadul, K. L. Cappuccio and M. Kaba, *Org. Process Res. Dev.*, 2006, 10, 723–726.
- 26 C. K. Savile, J. M. Janey, E. C. Mundorff, J. C. Moore, S. Tam, W. R. Jarvis, J. C. Colbeck, A. Krebber, F. J. Fleitz and J. Brands, *Science*, 2010, 329, 305–309.
- 27 Y. Wei, S. Xia and C. He, *Biotechnol. Lett.*, 2016, 38, 841–846.
- 28 L. L. Zeng, Y. J. Ding, G. C. Zhang, H. R. Song and W. H. Hu, *Chin. Chem. Lett.*, 2009, 20, 1397.
- 29 S. Zhou, J. Wang, X. Chen, J. L. Aceña, V. A. Soloshonok and H. Liu, *Angew. Chem., Int. Ed.*, 2014, 53, 7883–7886.
- 30 K. Lin, Z. Cai and W. Zhou, *Synth. Commun.*, 2013, 43, 3281.
- 31 O. Gutierrez, D. Metil and N. Dwivedi, *Org. Lett.*, 2015, 17, 1742–1745.
- 32 A. J. Blacker, *Process for the Preparation of aromatic Amines*, WO2004110976, 2004, 09, 06.
- 33 D. Spencer, I. Norihiro and N. Eugenia, *Process to Chiral Beta-amino acid Derivatives*, WO2004085661, 2004, 03, 19.
- 34 Y. Xiao, D. Joseph and W. Shane, *Process for the Preparation of Chiral Beta amino Acid Derivatives by Asymmetric Hydrogenation*, WO2004085378, 2004, 03, 15.
- 35 B. H. Lipshutz, K. Noson and W. Chrisman, *J. Am. Chem. Soc.*, 2001, 123, 12917–12918.
- 36 H. Shimizu, I. Nagasaki and K. Matsumura, *Acc. Chem. Res.*, 2007, 40, 1385–1393.

