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Light-responsive Pickering emulsions based on azobenzene-modified particles†

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Light-responsive particle-stabilised (Pickering) emulsions can in principle be selectively emulsified/demulsified on-demand through the remote application of light. However, despite their wide-ranging potential in applications such as drug delivery and biphasic catalysis, their rational design is extremely challenging and there are very few examples to date. Herein, we investigate a model system based on silica particles functionalised with azobenzene photoswitches to understand the key factors that determine the characteristics of light-responsive Pickering emulsions. The particle hydrophobicity is tuned through judicious variation of the spacer length used to graft the chromophores to the surface, the grafting density, and irradiation to induce *trans*–*cis* photoisomerisation. For select emulsions, and for the first time, a reversible transition between emulsified water-in-oil droplets and demulsified water and oil phases is observed with the application of either UV or blue light, which can be repeatedly cycled. A combination of surface energy analysis and optical microscopy is shown to be useful in predicting the stability, and expected light-response, of a given emulsion. Using the observed trends, a set of design rules are presented which will help facilitate the rational design, and therefore, more widespread application of light-responsive Pickering emulsions.

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Introduction

Pickering or particle-stabilised emulsions have recently seen a resurgence in interest^{1–3} as a method of encapsulation for the key areas of food, pharmaceutical technologies and biphasic catalysis.^{4,5} In a Pickering emulsion, one liquid is dispersed as droplets throughout a second immiscible liquid, and the interface is decorated with solid ‘stabilising’ particles. Stimuli-responsive Pickering emulsions are of particular interest as they permit the selective stabilisation and destabilisation of emulsion droplets, or inversion between oil-in-water (O/W) and water-in-oil (W/O) emulsions, in response to external factors such as pH,^{6,7} magnetic fields,⁸ temperature⁹ and light.^{3,10} Of all the stimuli, light may be considered the most advantageous, offering superior remote, spatial and temporal control, compared to pH or temperature.^{3,11} For biphasic catalysis this permits facile product separation and catalyst recovery,¹² while for medicinal therapeutics it offers targeted, non-invasive treatments that avoid premature delivery of the droplet cargo.²

Despite this potential, only a handful of light-responsive Pickering emulsions have previously been reported. Bai *et al.*

used inherently light-responsive TiO₂ particles as stabilisers, which caused emulsion destabilisation upon UV irradiation.¹³ Chen *et al.* demonstrated light responsivity using silica particles whose surface was grafted with photoswitchable donor-acceptor Stenhouse adducts (DASAs).¹⁰ Likewise, Chen *et al.* used spiropyran-type photoswitches to accomplish a similar result.¹⁴ In both cases, inversion between O/W and W/O emulsions was achieved upon irradiation. Recently Li *et al.* used ionic liquid surfactants based on azobenzene (Azo) photoswitches adsorbed to silica, to induce reversible emulsification and demulsification for application in biphasic catalysis.¹² It was shown that the system was fully recyclable, allowing separation of the ionic liquid and catalyst.

The design of light-responsive particles for Pickering emulsions is non-trivial. As shown in Fig. 1 (regions I and IV), particles that lie at the extremes of hydrophobic or hydrophilic character are completely wetted by the oil or water phase, respectively, and form aggregates. For particles of intermediate hydrophobicity, it is energetically favourable for them to sit at the interface as they reduce the overall interfacial tension between the oil and water. Particles that are more hydrophobic and therefore better wetted by the oil phase, will favour the formation of W/O emulsions (Fig. 1, region III) and will have an angle >90°, as measured through the oil–water interface. Conversely, particles that are more hydrophilic will favour O/W emulsions (Fig. 1, region II), with an angle <90°. It is therefore important to tune the hydrophobicity/hydrophilicity

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† Electronic supplementary information (ESI) available: DRUV data, elemental analysis, DLS, surface energy estimation, optical microscopy processing and surface coverage calculation details. See DOI: <https://doi.org/10.1039/d2sm00697a>



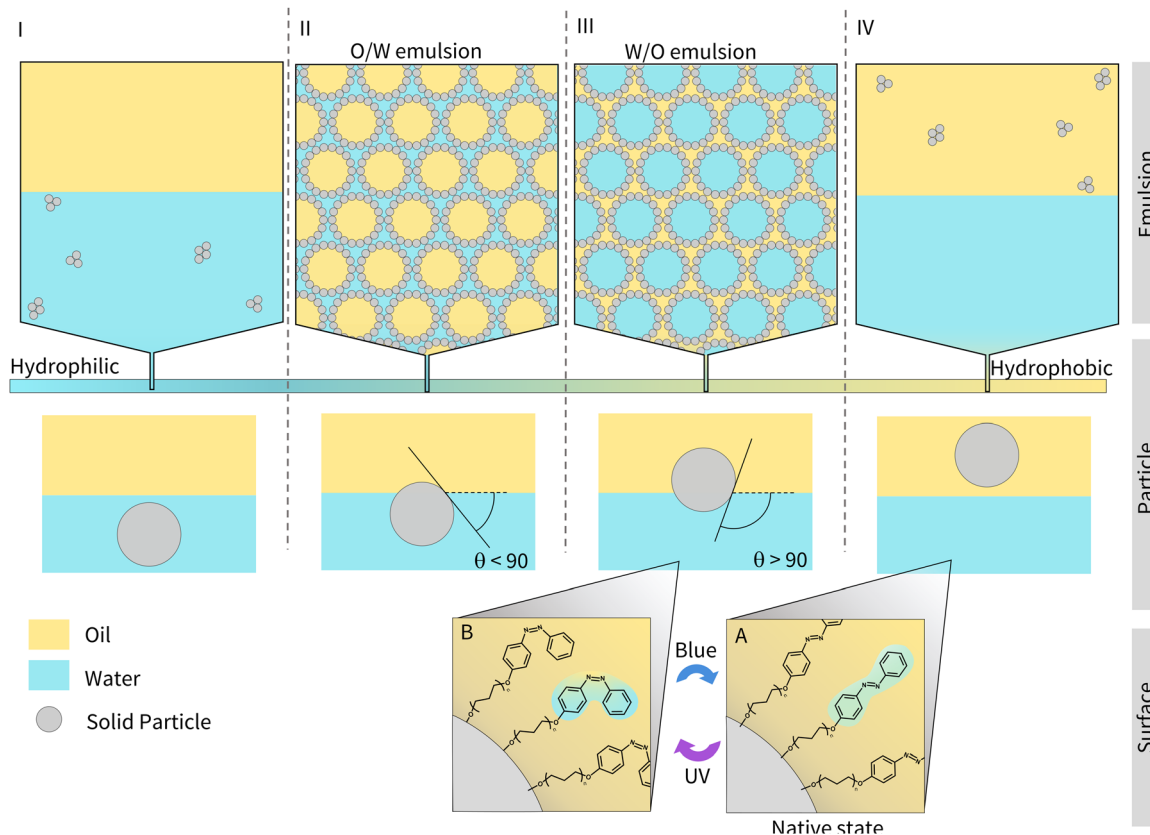


Fig. 1 The relationship between the hydrophobicity of the stabilising particle and the type of Pickering emulsion produced. I–IV show the different morphologies that can be accessed (I – aggregates in water, II oil-in-water (O/W) emulsion, III water-in-oil (W/O) emulsion, IV aggregates in oil). A and B show the reversible photoisomerisation of azobenzene molecules grafted to the particle surface and the resulting changes in hydrophobicity. The oil–water contact angle, θ , which increases with increasing hydrophobicity is indicated.

of the particles to form a stable emulsion.¹⁵ For the emulsion to be light-responsive, two key criteria must be met. Firstly, the particles should exist in two different states of hydrophobicity upon irradiation (*e.g.*, Fig. 1 A and B). Secondly, the two states must alter the interfacial energy sufficiently, such that different morphological states can be accessed (Fig. 1 I to IV).¹⁶

Despite these observations, to the best of our knowledge, there has been no detailed discussion about how to design or choose an appropriate lead for a light-responsive Pickering system (*i.e.*, choice of photoswitch, chain-length, grafting density, and oil selection). With this information, properties such as the morphologies that light-responsive emulsions invert between, or the oil that they are compatible in could be tailored to a given purpose, *i.e.*, biphasic catalysis or drug delivery, promoting their more widespread use. Herein, we address this omission, by identifying a set of key principles that will facilitate the rational design of light-responsive Pickering emulsions. To achieve this, we examine a model system based on fumed silica particles whose surface is functionalised with three different azobenzene derivatives (Fig. 2 – AzoOH, AzoO(CH₂)₃OH and AzoO(CH₂)₁₁OH). Azo chromophores have been widely studied^{17–19} due to their high and reversible polarisability upon *trans-cis* photoisomerisation (*ca.* 0 D to 3 D, respectively for native, unmodified azobenzene²⁰). This property

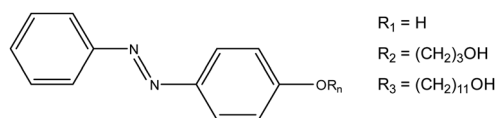


Fig. 2 Azobenzene derivatives studied in this work. The naming convention is as follows: **R₁** corresponds to **AzoOH**, **R₂** is **AzoO(CH₂)₃OH** and **R₃** is **AzoO(CH₂)₁₁OH**.

has previously been exploited to alter the wettability of solid surfaces, with some examples showing an ability to reversibly cycle between superhydrophobic and superhydrophilic states upon irradiation with different wavelengths of light.²¹ Moreover, there has been extensive work on functionalised azobenzenes affixed to gold nanoparticles, which reversibly aggregate in single-phase media.^{22–24} Dimethyl amino substitution of the Azo molecules,²² or the incorporation of different thiolate linkers, has been shown to modify both the nanoparticle solubility and the wavelength at which self-assembly occurs.²⁵ Similar results have also been observed in related systems using spiropyran photoswitches.^{26,27}

This work aims to apply similar principles to larger (200 nm) fumed-silica particles in biphasic systems, inducing morphological changes on the tens of micron scale. The hydrophobicity



of Azo-functionalised particles in the native *trans*-state is modulated by using alkyl spacers of different lengths and by using three different grafting densities. Pickering emulsions are prepared using diethyl adipate as a polar oil, selected as it is capable of stabilising emulsions with fumed silica and shows limited solubility in water.²⁸ Moreover, adipate oils are capable of stabilising, typically unstable, hydrophilic particles *in situ*.²⁹

Whereas previous work has taken an application-led approach to creating light-responsive emulsions,^{10,12–14} here we demonstrate that through detailed characterisation of the light-responsive particle stabilisers, the morphology of the emulsion morphology can be predicted and the particle/oil composition tailored to deliver the desired characteristics. A combination of optical microscopy and contact angle tensiometry are used to investigate how the particle hydrophobicity, surface morphology of the droplets and surface energy of the particles determine the stability of the resultant emulsion. Through rationalisation of these key structure–property relationships we propose key selection criteria for the design of light-responsive Pickering emulsions based on the judicious choice of photoswitch, its grafting density and an appropriate oil.

Experimental

Materials

Three azobenzene derivatives were used in this study (Fig. 2). 4-(Phenyldiazenyl)phenol (AzoOH) was used as received from the supplier (Sigma-Aldrich, 99%) and the other derivatives were prepared *via* a Williamson ether synthesis as outlined below. 11-Bromo-1-undecanol (99%), *n*-hexadecane (99%), fumed silica (0.2–0.3 μm diameter, $200 \pm 25 \text{ m}^2 \text{ g}^{-1}$ surface area, 5 free hydroxyl groups per nm^2) and silicone oil (100 mPa S (20 °C), $[-\text{Si}(\text{CH}_3)_2\text{O}-]_n$, molecular weight *ca.* 5970 g mol^{-1} , $\geq 99\%$)³⁰ were purchased from Sigma-Aldrich. 3-Bromo-1-propanol (97%) dicyclohexano-18-crown-6 (97%) and potassium carbonate (99%) were purchased from Alfa Aesar. Acetone (analytical grade), dichloromethane (DCM, HPLC grade), diethyl adipate (99%), ethanol (≥ 99.8), ethyl acetate (analytical grade), *n*-hexane (95%), dimethyl formamide (DMF, $\geq 99\%$), methanol ($\geq 99\%$) and Nile Red ($\geq 99\%$) were purchased from Fisher Scientific. Deuterated chloroform (99.8%) was purchased from Eurisotop. Water was obtained from a Millipore Simak 2 water purification system. All reagents were used as received.

Synthesis of azobenzene derivatives with varying spacer lengths

4-(Phenyldiazenyl)phenol (1.02 g, 5.04 mmol) was dissolved in DMF (50 mL). To this mixture was added either 3-bromo-propan-1-ol (0.715 g, 5.04 mmol) or 11-bromo-1-undecanol (1.29 g, 5.04 mmol), potassium carbonate (0.47 g, 3.4 mmol) and dicyclohexano-18-crown-6 (0.067 g, 0.25 mmol). The mixture was heated at 100 °C for 3 h. The dark orange solution that formed was extracted in ethyl acetate (100 mL) and distilled water (100 mL). The organic layer was then dried and recrystallised from a minimum volume of hot ethanol. For the 3-bromo-1-undecanol reaction, additional purification *via* column

chromatography was required (DCM–MeOH 99 : 1). The reaction products were obtained as solids and characterised by NMR spectroscopy (ESI,† Fig. S1 and S2) and mass spectrometry (ESI,† Fig. S3 and S4). (i) 3-(4-(Phenyldiazenyl)phenoxy)propan-1-ol (AzoO(CH₂)₃OH, 0.74 g, 57%). ¹H NMR (400 MHz; CDCl₃) δ 7.94–7.87 (m, 4H, CH_{aromatic}), 7.50 (t, 2H, CH_{aromatic}, $J = 7.4$ Hz), 7.01 (d, 2H, CH_{aromatic}, $J = 8.97$ Hz), 4.19 (t, 2H, CH₂, $J = 6.00$ Hz), 3.87 (dd, 2H, CH₂, $J = 10.6, 5.5$ Hz), 2.07 (p, 2H, CH₂, $J = 6.0$ Hz), 1.95 (t, 1H, OH, $J = 5.0$ Hz). ¹³C NMR (100 MHz; CDCl₃) 61.40 (C_{ar}), 152.82 (C_{ar}), 147.11 (C_{ar}), 130.51 (C_{ar}), 129.14 (C_{ar}), 124.87 (C_{ar}), 122.66 (C_{ar}), 114.80 (C_{ar}), 65.81 (CH₂), 60.09 (CH₂), 32.03 (CH₂). m/z [M+H⁺] C₁₅H₁₇N₂O: calc.: 257.1245; found: 257.1300. (ii) 11-(4-(Phenyldiazenyl)phenoxy)undecane-1-ol (AzoO(CH₂)₁₁OH, 1.18 g, 65%). ¹H NMR(CDCl₃) δ 7.93–7.87 (m, 4H, CH_{ar}), 7.52–7.41 (m, 3H, CH_{ar}), 7.00 (d, 2H, CH_{ar}, $J = 8.91$ Hz), 4.03 (t, 2H, CH₂, $J = 6.55$ Hz), 3.63 (t, 2H, CH₂, $J = 6.52$ Hz), 1.82 (p, 2H, CH₂, $J = 7.20$ Hz), 1.56 (p, 2H, CH₂, $J = 2.80$ Hz), 1.477 (p, 2H, CH₂, $J = 2.80$ Hz), 1.38–1.29 (m, 13H, CH₂). ¹³C NMR (100 MHz; CDCl₃) δ 161.71 (C_{ar}), 152.78 (C_{ar}), 146.83 (C_{ar}), 130.30 (CH_{ar}), 129.02 (CH_{ar}), 124.75 (CH_{ar}), 122.53 (CH_{ar}), 114.69 (CH_{ar}), 68.35 (CH₂), 63.06 (CH₂), 32.81 (CH₂), 29.60 (CH₂), 29.55 (CH₂), 29.52 (CH₂), 29.44 (CH₂), 29.38 (CH₂), 29.19 (CH₂), 26.01 (CH₂), 25.76 (CH₂). m/z [M + H⁺] C₂₃H₃₃N₂O₂: calc.: 369.2497; found: 369.2551. *ar = aromatic.

Surface grafting of azobenzene derivatives to fumed silica

Ethanol (20 mL) was added to fumed silica (0.5 g) and the sample was sonicated for 2 min at room temperature. To this mixture, either AzoOH, AzoO(CH₂)₃OH or AzoO(CH₂)₁₁OH was added at a concentration corresponding to 100, 60 or 20% of the maximum available surface hydroxyl groups. The solution was refluxed for 3 h and dried *in vacuo* before being transferred to an oven and heated at 200 °C for 18 h. The resulting powder was Soxhlet-extracted for 48 h in acetone and dried. Nine different solid samples were produced and are referred to as AzoOH-silica, AzoO(CH₂)₃OH-silica and AzoO(CH₂)₁₁OH-silica with a nominal surface grafting density of either 1, 3 or 5 molecules per nm^2 .

Characterisation of modified fumed silica particles

Diffuse-reflectance UV-Vis (DRUV) measurements were carried out using a PerkinElmer Lambda 750 spectrophotometer equipped with a Labsphere 60 mm RSA ASSY integrating sphere. Samples were measured in a powder sample holder with a fused quartz disc. Spectra were then transformed using the Kubelka–Munk transformation.³¹ Dynamic light scattering (DLS) was conducted using a Malvern Zetasizer Nano equipped with a He–Ne laser emitting at 633 nm with a 4 mW power source. The scattering intensity was detected at a backscatter angle of 173°. Samples (0.2 mg mL^{-1}) were prepared by dispersing azo-grafted silica particles in triple-filtered (0.2 μm) H₂O and sonicating for 2 min. For samples that were too hydrophobic to be dispersed in H₂O, propan-2-ol was used. The hydrodynamic diameter (D_h) was calculated by assuming that the diffusing particles were monodispersed spheres. The D_h was averaged from 3 runs of approximately 20 measurements each. Cumulants analysis, which



is a fit of the logarithm of the correlation function, gave a mean value for the diameter (Z -average) and width parameter of the distribution, the polydispersity index (PDI). Elemental analysis was conducted using an Exeter Analytical CE-440 elemental analyser and the percentage of nitrogen present used to calculate the number of azobenzene molecules that are present at the surface of the silica (see ESI,† Section 2). Dispersion tests were carried out by adding 1 mg of the functionalised silica powders to 1 mL of either silicone oil (100 mPa S), diethyl adipate, n -hexadecane or water and sonicating for 2 min using a Fisherbrand S-series sonication bath. Samples were considered dispersed if no aggregates were visible to the naked eye.

Determination of surface energy from contact angle measurements

The difficulties that exist in determining the surface energy of irregular solid powders have been discussed previously.^{32,33} This work follows a similar procedure to Binks *et al.*^{32,33} Briefly, azobenzene-modified silica powders were adhered to glass slides using cotton-paper double-sided tape and tested with 4 different liquids: water, glycerol, ethylene glycol and α -bromonaphthalene. An FTA1000 B contact tensiometer was used to measure the contact angles between test liquids and the solid. Static contact angles of a drop of each test liquid (*ca.* 10 μ L) were measured and the average of at least 3 droplets was taken. For UV-irradiated samples the process was carried out in the dark and a 575 nm longpass filter applied to the front of the instrument's illumination source (a diffuse white light-emitting diode (LED)). Further details regarding data processing can be found in the ESI† (Section 7).

Preparation of Pickering emulsions

Pickering emulsions were prepared by adding as-synthesised Azo-silica particles to equal volumes (1 mL) of H₂O (Millipore) and diethyl adipate or n -hexadecane. Particles were initially dispersed in the oil by sonicating at room temperature for 2 min using a Fisherbrand S-series sonication bath and water added. Homogenisation was then carried out using a Sciquip basic homogeniser (max. 30 000 rpm) operating at 50% speed for 2 min while cooling the samples in an ice bath. Samples were contained in a vial during homogenisation (inner diameter 16.8 mm, outer diameter 18.6 mm) and the homogeniser tip placed 8 mm from the bottom of the vial.

Characterisation of Pickering emulsions

Pickering emulsions were characterised *via* optical microscopy by pipetting a droplet of the emulsion layer onto a glass slide. Samples were analysed both 15 min and 24 h after homogenisation. Images were recorded using a Canon 400D camera mounted to a Swift 350T microscope using an $\times 4$ objective at a resolution of 5184 \times 3456 px. To acquire the droplet sizes, a computer vision technique that uses the circle Hough transformation was first used to extract a series of 'circles' from the microscopy images.³⁴ A programme utilising the circle Hough transform, provided as part of OpenCV,³⁵ was developed, which is able to scan across the acquired microscope images.³⁶

More information on this technique can be found in the ESI† (Section 3). To ascertain whether oil-in-water or water-in-oil emulsions formed, samples were dyed with a minimum quantity of Nile red (a lipophilic dye)³⁷ and the stained component of the optical micrographs was determined to be the oil phase. The emulsion stability was determined by assessing the fraction of oil/water phase resolved over the course of 21 days for n -hexadecane emulsions and 14 days for adipate emulsions.

Irradiation with light

Sample irradiation was carried out using high-power UV and blue LEDs (365 nm, 6.00 mW cm⁻² irradiance and 455 nm, 5.16 mW cm⁻² irradiance, respectively) for 20 min. Solid powders were irradiated directly with regular agitation of the samples. Coated slides were irradiated similarly. For samples that showed photoswitching (as determined by DRUV), a colour change from yellow to dark orange was observed, corresponding to photoisomerisation of the azobenzene chromophore from the *trans* to *cis* form.

Results and discussion

The hydrophobicity of the particle stabilisers in their native state (*i.e.*, before photoswitching) is important in determining which type of emulsion forms (see Fig. 1, regions I–IV). In this work, the hydrophobicity is modulated in two ways: (i) the length of alkyl spacer separating the azobenzene photoswitch from the particle surface and (ii) the grafting density on the particle surface (n Azo molecules per nm²). For clarity, the azobenzene-modified particles are denoted as "Azo-silica (number of molecules of azo per nm² surface area)."

Grafting of azobenzene derivatives to fumed silica

AzoOH, AzoO(CH₂)₃OH and AzoO(CH₂)₁₁OH were grafted to fumed silica particles *via* alkoxylation of the surface silanol groups using a heat treatment method.³⁸ Grafting was confirmed *via* DRUV measurements following a solvent extraction step which removed non-bound azobenzene. The DRUV spectra of the particles show characteristic peaks at approximately 340 nm and 440 nm corresponding to the π - π^* and n - π^* transitions of the azobenzene species, respectively (ESI,† Fig. S5).¹⁷ No significant changes in the absorption maxima were observed for the different azobenzene derivatives, suggesting that the length of the carbon spacer has negligible effect on the electronic character of the chromophore. We note that DRUV spectra are comparable to UV/Vis absorption spectra for the Azo ligands in solution (ESI,† Fig. S6).

The degree of surface functionalisation was synthetically controlled by varying the quantity of azobenzene species relative to the surface hydroxyl groups in the reaction mixture. The resulting grafting density was determined *via* elemental analysis (EA, see Table 1; further information available in the ESI† Section 2 and Table S3). As anticipated, a direct correlation between the azobenzene concentration in the reaction mixture and final grafting density is observed. The surface of the fumed



Table 1 Grafting density and dispersibility of Azo-silica particles. Columns 2 and 3 show the relationship between the number of molecules used in grafting synthesis and the calculated grafting density of the powders determined from elemental analysis (EA). Columns 4–5 show the results of liquid dispersion tests in water, diethyl adipate and silicone oil. Key: ● sample dispersed; ○ sample did not disperse; ⊙ sample dispersed after the application of UV light. Literature values for the polarity of the test liquids can be found in the ESI, Table S7

System	No. of molecules per nm ² (synthesis)	Grafting density molecules per nm ² (EA)	Water	Diethyl adipate	<i>n</i> -Hexadecane	Silicone oil
AzoOH-silica	1	0.5	●	●	●	○
	3	1.0	●	●	●	○
	5	1.5	●	●	●	○
AzoO(CH ₂) ₃ OH-silica	1	0.6	●	●	●	○
	3	1.4	○	●	●	●
	5	1.7	○	●	●	●
AzoO(CH ₂) ₁₁ OH-silica	1	0.4	●	●	●	○
	3	1.5	○	⊙	●	●
	5	1.7	○	⊙	●	●

silica particles has 5 free hydroxyl groups per nm² (as according to supplier information³⁹). As calculated from EA data, between 0.4 and 1.7 of the available hydroxyl groups (per nm² surface area) undergo alkoxidation with the Azo-species. This result is consistent with existing literature for silanol–alcohol condensation^{38,40,41} and the density obtained does not show a significant dependence on the choice of Azo chromophore. Notably, the relationship between the reaction concentration and grafting density is non-linear, and never reaches the maximum theoretical grafting density, suggesting that there is reduced access to the silanol sites at higher grafting densities, or that some of the reactive silanol sites may have been removed during the heating step (200 °C) of the grafting reaction.^{38,42}

DLS confirmed that no significant change in the size or polydispersity of the particle occurred during the grafting process. All particles showed a mean hydrodynamic diameter of $D_H \sim 200 \pm 50$ nm, which is in good agreement with the unmodified fumed silica samples (ESI,† Table S4 and Fig. S8).

Dispersion tests with liquids of varying polarity (Table 1) showed a marked change in hydrophobicity for AzoO(CH₂)₃OH- and AzoO(CH₂)₁₁OH-silica particles with surface coverages >20%. These samples no longer dispersed in water but could be dispersed in silicone oil. This was not observed for AzoOH, which was insoluble in silicone oil at all grafting densities consistent with previous studies.³⁸

Light-response of Azo-silica powders

We have seen that the hydrophobicity of fumed silica can be tuned by varying the spacer length and grafting density of the Azo modifiers. The next step was to assess the change in hydrophobicity that resulted from photoisomerisation of the grafted chromophores under UV light. This could be observed visually by the sample colour change from yellow to orange (ESI,† Fig. S7). Conversion to a *cis*-dominant photostationary state (PSS) is further indicated in the DRUV spectra by a decrease in intensity of the π - π^* transition at 340 nm, accompanied by an increase of the *cis*-centred n - π^* transition at 440 nm.^{17,43,44}

Comparable DRUV spectra were obtained for both AzoO(CH₂)₃OH- and AzoO(CH₂)₁₁OH-silica samples at all grafting densities (Fig. S5, ESI†). The PSS composition was

determined to be ~70–85% *cis*-isomer for both AzoO(CH₂)₃OH- and AzoO(CH₂)₁₁OH-silica, depending on the grafting density (see Tables S1 and S2 and Section 2, ESI†). In contrast, the *cis*-isomer absorption peak was essentially absent for AzoOH-silica at all grafting densities. This is due to the enhanced rate at which AzoOH thermally isomerises back to the *trans*-form, particularly in polar solvents to which it can hydrogen bond,¹⁷ making measurement of the *cis*-isomer difficult without the use of ultra-fast spectroscopic techniques.

Dispersion tests were also performed using UV-irradiated Azo-silica particles. From Table 1 it can be seen that irradiation had a negligible effect on the dispersion results for all samples except AzoO(CH₂)₁₁OH-silica-(1.5 and 1.7 molecules per nm²). In the *trans* form, these samples aggregated in diethyl adipate but upon UV irradiation their hydrophobicity decreased sufficiently for them to disperse. For all other samples it is expected that there is a change in hydrophobicity, but this is not sufficient to change the compatibility with the dispersants used here.

Emulsion stability

So far, we have demonstrated a facile method to control the surface grafting and thus hydrophobicity of fumed silica particles using Azo derivatives with different spacer lengths. These particles also exhibit different photoswitchable responses. A series of emulsions was then made in order to evaluate how these particle characteristics affect the emulsion stability and their ability to undergo morphological changes in response to light.

Emulsions were prepared containing an equal volume of H₂O and diethyl adipate and 10 mg mL⁻¹ particle loading. Fig. 3 shows representative images of the emulsions obtained. Average particle sizes and dispersity for all samples can be found in the in the ESI,† Table S5. For all particles, characteristic droplets of a W/O emulsion are seen, where water droplets are dispersed throughout the continuous diethyl adipate phase. Emulsion stability was assessed by measuring the fraction of oil or water phase that was resolved over the course of >14 days (ESI,† Fig. S9 and S10). No significant change in the droplet size or dispersity was observed over this period.

Interestingly, AzoO(CH₂)₁₁OH-silica (1.5 and 1.7 molecules per nm²) did not form stable emulsions in the *trans*-state



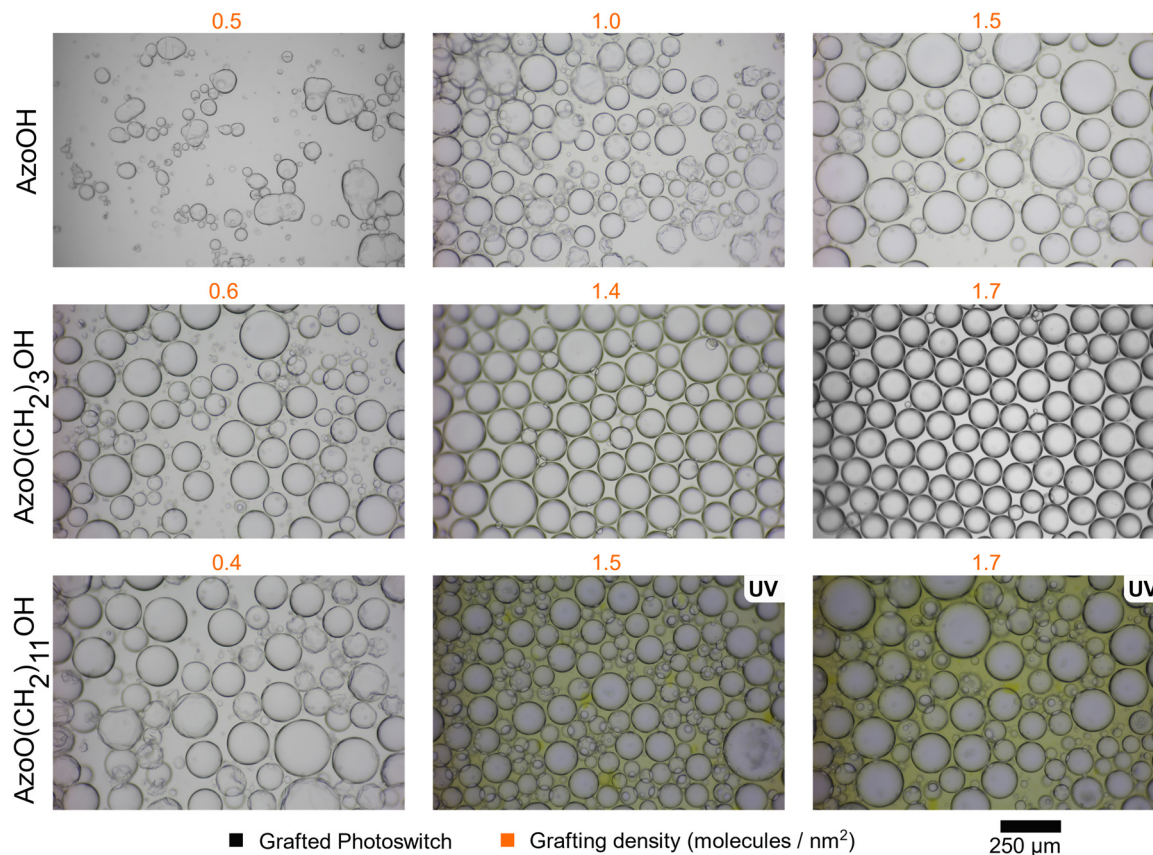


Fig. 3 Optical microscopy images of W/O particle-stabilised emulsions (10 mg mL^{-1} relative to H_2O) with (i) AzoOH-, (ii) AzoO(CH₂)₃OH- and (iii) AzoO(CH₂)₁₁OH-silica at different grafting densities (values shown in orange text). Morphological differences arising from changes in surface energy of the particles are apparent. AzoO(CH₂)₁₁OH-silica (1.5 and 1.7 molecules per nm^2) were irradiated with UV light before emulsion preparation as the non-irradiated counterparts did not form a stable emulsion. All images were taken 24 h after emulsion formation.

(ESI,† Fig. S9c), instead favouring formation of oil-phase aggregates (Fig. 1 – IV). However, when irradiated with UV light, the emulsion was stabilised (Fig. 1 – III). From Fig. 3, we observed that these stable emulsions are more polydisperse than *trans*-stable emulsions for the other systems and the oil phase appears yellow, indicating the presence of aggregated particles. This suggests that not all of particles reach the interface during homogenisation. This could be due to the higher hydrophobicity of both isomers of AzoO(CH₂)₁₁OH-silica (compared to the other Azos) or alternatively a result of the reduced penetration depth of the light into the particle aggregates, which results in incomplete photoswitching. These UV-stabilised emulsions have a similar stability to coalescence/sedimentation (see ESI,† Fig. S9) as similar grafting density, but more hydrophilic particles (*i.e.*, AzoO(CH₂)₃OH). For all other samples, UV irradiation did not affect the emulsion morphology or stability, with similar emulsion droplet radii and polydispersity observed. The stability of the UV-stabilised emulsions was also assessed at different water fractions to see if catastrophic inversion could occur,³² but no such process was observed (ESI,† Section 5 and Fig. S11).

In situ irradiation experiments were performed on the AzoO(CH₂)₁₁OH-silica emulsion ($1.5 \text{ molecules per nm}^2$, 10 mg mL^{-1}) to demonstrate the reversibility of the emulsification/demulsification

process and to show that switching could occur while the particles were adsorbed at the droplet interface (Fig. 4 and Fig. S12, ESI†). The emulsion mixture was irradiated with either UV (365 nm) or blue (450 nm) light and homogenised between images. In the *trans* form, the particles are too hydrophobic to form a stable emulsion, and favour aggregation. Following UV irradiation, an emulsion containing densely packed, spherical droplets is formed. When this sample is then irradiated with blue light under stirring to induce reverse *cis-trans* isomerisation, the droplets disperse and particle aggregates form. The irradiation-homogenisation process can be cycled with similar morphologies achieved each time. However, the mean droplet radius decreases with cycling, and the sample becomes less polydisperse (Fig. 5). This is attributed to better dispersion of the Azo-silica aggregates through repeated homogenisation processes, which effectively increases the available surface area on the stabilising particles. The ability to cycle these materials between stable and unstable states suggests that they are suitable candidates for light-responsive encapsulant applications such as biphasic catalysis.

We note that diethyl adipate oil is quite polar and partially soluble in water.²⁹ It was chosen in this study as it is likely to form unstable emulsions with hydrophobic particles (Table 1).



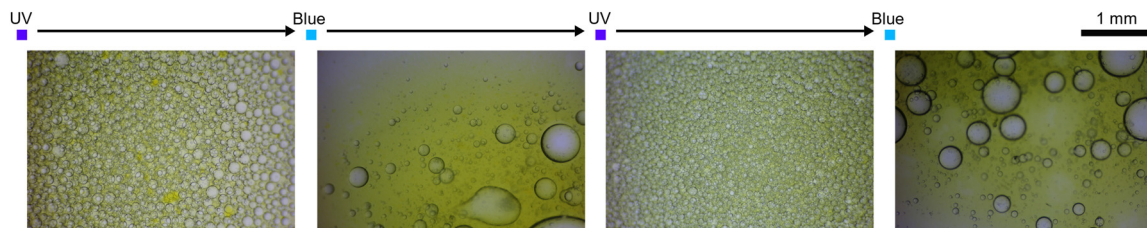


Fig. 4 Optical microscopy images of W/O AzoO(CH₂)₁₁OH-silica (1.5 molecules per nm²) stabilised emulsion (particle conc. = 10 mg mL⁻¹ relative to H₂O) that is light-responsive. The oil–water mixture was irradiated with either UV (365 nm) or blue (450 nm) light and homogenised prior to image capture. The images represent 2 cycles of emulsification and demulsification.

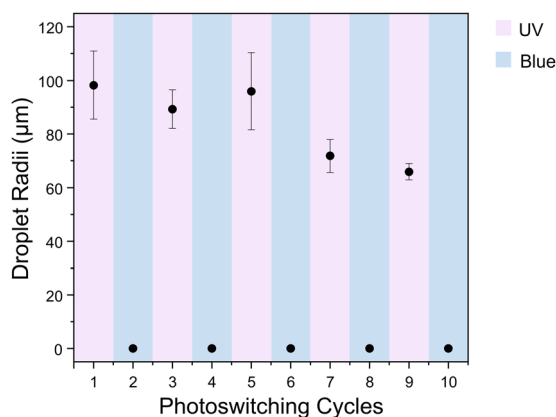


Fig. 5 Change in mean droplet radii after successive irradiation and homogenisation cycles with UV and blue light, alternating between ~60–120 µm droplets and 0 µm, indicating aggregation and loss of droplets. Blue bars indicates that the emulsion has been irradiated with blue light and pink bars indicates irradiation with UV light. Error bars show polydispersity of droplets formed.

To show parity with existing literature, hexadecane, a less polar oil was also investigated. The analogous hexadecane systems follow broadly similar trends (ESI†, Fig. S10).

The main difference is that all emulsions remain stable and O/W emulsions are now accessed by the most hydrophilic particles (see ESI†, Table S6). None of the emulsions were found to show significant changes upon UV irradiation suggesting that in hexadecane the hydrophobicity of the particles is not near enough to any of the boundary points identified in Fig. 1.

Estimation of particle surface energy

We have shown qualitatively that the particle hydrophobicity can be tuned and increased to a point where an emulsion is no longer stable (for a given oil) and that UV-irradiation increases the hydrophilicity and allows emulsion stabilisation for some systems. However, to rationally design a light-responsive emulsion, we now need to quantify the particle hydrophobicity and use this to predict the emulsion stability for each photoisomer.

The contact angle of the solid particle at the oil–water interface (θ_{ow}) dictates whether it sits preferentially in the oil or water phases. This gives an indication of the preference of the solid particles for the oil phase in both the *cis* and *trans* states and allows us to predict whether a light-response would be seen with oils other than diethyl adipate.²⁸ While θ_{ow} is

difficult to measure experimentally, it can be estimated from the interfacial energies according to:

$$\cos \theta_{ow} = \frac{\gamma_{so} - \gamma_{sw}}{\gamma_{ow}} \quad (1)$$

where γ is the interfacial energy between the various solid (s), oil (o), and water (w) phases. Several models have been proposed for the calculation of γ , which split the surface energy into a number of components, including Zisman,⁴⁵ Owens–Wendt,⁴⁶ Fowkes,⁴⁷ and van Oss theory.⁴⁸ Here we follow the approach used by Binks *et al.* expressing the interfacial energy for a given component (γ) as the sum of the polar (γ^p) and dispersive (γ^d) components:^{32,33}

$$\gamma = \gamma^d + \gamma^p \quad (2)$$

The interfacial energy between two different phases can then be calculated as follows, shown for the oil–water interface as an example:

$$\gamma_{ow} = \gamma_o + \gamma_w - 2\sqrt{\gamma_o^d \gamma_w^d} - 2\sqrt{\gamma_o^p \gamma_w^p} \quad (3)$$

Values for the various γ components for common liquids are readily available in the literature.^{32,33,49} Estimation of γ_{so} or γ_{sw} is more complicated as γ_s^d and γ_s^p for the Azo-silica particles are unknowns. One solution is to determine these values indirectly from the solid–liquid contact angle, obtained from combination of the relevant forms of eqn (1) and (3):

$$\cos \theta_{sl} = -1 + \frac{2\sqrt{\gamma_s^d \gamma_l^d}}{\gamma_l} + \frac{2\sqrt{\gamma_s^p \gamma_l^p}}{\gamma_l} \quad (4)$$

Using contact angle measurements between a particle-coated surface and a series of test liquids (l) of known polarity enables values for γ_s^d and γ_s^p to be estimated using a weighted least-squares analysis to find the values that best fit all the liquids simultaneously, represented as a 3D surface energy plot (see Fig. 6 and Fig. S13, ESI† for examples).³³ The test liquids chosen in this work are water, glycerol, ethylene glycol and α -bromonaphthalene (see Table S7, ESI† for literature data of interfacial tensions). Further information on these calculations can be found in the ESI† (Section 7).

Table 2 gives the polar and dispersive components for each Azo-silica sample determined from the surface energy plots, and the estimated total surface energy, γ_s , at the solid–air interface (calculated from eqn (2)). The overall trend shows a



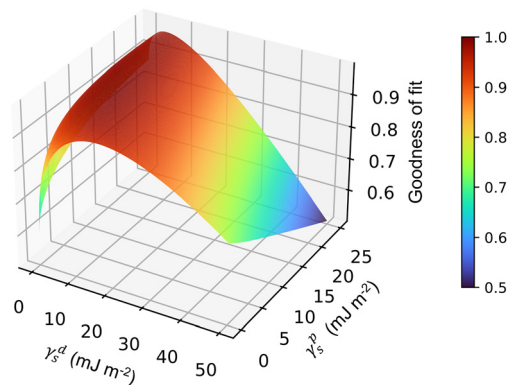


Fig. 6 Example 3D surface energy plot for AzoO(CH₂)₁₁OH-silica (0.6 molecules per nm²) giving the 'goodness of fit' for different values of γ_s^p and γ_s^d . The highest point represents the best fit values of γ_s^p and γ_s^d .

decrease in the surface energy with increasing length of the carbon spacer, and by extension, an increase in the hydrophobicity. A similar surface energy is obtained for all grafting densities of AzoOH, due to its relatively hydrophilic nature compared to AzoO(CH₂)₃OH and AzoO(CH₂)₁₁OH. This is consistent with studies on the hydrophobisation of fumed silica, where partial or substantial fluorination, respectively, led to similar changes in surface energy.³³

These results also confirm observations from the dispersion tests, where samples containing the longer carbon spacers showed a marked increase in hydrophobicity at approximately 20% coverage. For AzoO(CH₂)₁₁OH-silica, γ_s drops from 27.5 mJ m⁻² to 10.0 mJ m⁻² upon increasing the surface coverage from 0.4 to 1.5 molecules per nm², rendering it unable to disperse in the moderately polar diethyl adipate ($\gamma_1 = 31.7$ mJ m⁻²).²⁸ Moreover, comparison of the different Azo-silica samples shows that the choice of adsorbed molecule is more critical than the grafting density, when modulating the surface hydrophobicity is the aim. Furthermore, there is a critical grafting density (*ca.* 20%) at which there is a stark increase in hydrophobicity. This may be attributed to both the bulkiness and degree of coverage for molecules with a carbon spacer, which prevent the adsorption of a layer of water.³⁸ It should also be noted that AzoO(CH₂)₃OH-silica and AzoO(CH₂)₁₁OH-silica show a higher surface energy at

1.7 than 1.5 molecules per nm², which we believe is due to the imprecision of contact angle measurements at high angles (>100°).

Now that we have estimates for the polar and dispersive components of the solid, we can in principle use eqn (3) to calculate the various interfacial energies required in eqn (1) to obtain the contact angle. However, this approach led to over-predicted values of θ_{ow} (>180°). It has previously been shown that a number of systematic errors inherent to the measurement of finely divided solids make it difficult to accurately predict θ_{ow} for a given emulsion.²⁸ Moreover, the emulsions investigated here are stable in polar oils. These have a tendency to be overpredicted using this simplistic model, which does not account for the complex structuring of polar molecules at the interface.^{33,49} Despite this, we can obtain a comparative understanding of the emulsion stability using values of $\cos \theta_{ow}$.

Fig. 7 shows the relationship between the surface energy, γ_s , of the particle and $\cos \theta_{ow}$, for diethyl adipate oil. Alternate oils (silicone oil, *n*-hexadecane and ethylene glycol) can be found in the ESI,† Fig. S14. The closer $\cos \theta_{ow}$ is to zero, the greater the preference of the solid particles for the aqueous phase. Emulsions that appear below the lower boundary line were experimentally determined to be unstable and formed aggregates. In diethyl adipate all emulsions favoured the oil phase, forming W/O emulsions, so the $\cos \theta_{ow}$ values are negative. Overall, increasing the grafting density and using longer carbon spacers led to a shift to more negative values of $\cos \theta_{ow}$, which is attributed to an increasing surface energy with the water phase. Interestingly, the hydrophilic AzoOH-silica has strong preference for the oil phase, greater than AzoO(CH₂)₃OH-silica (at the same grafting density), arising from its low surface energy with diethyl adipate. It can also be seen that as the Azo grafting density is increased, this preference for the oil phase decreases. This agrees with previous literature where a strong affinity between unmodified fumed silica and adipate oils was reported.²⁹

Comparing these results to the stability study and optical microscopy data helps to explain the previously qualitative observations. For example, in diethyl adipate why does AzoO(CH₂)₁₁OH-silica form aggregates at 1.5 and 1.7 molecules per nm² whereas the other solid particles do not? AzoO(CH₂)₁₁OH-silica has the greatest preference for the oil phase, particularly for

Table 2 Calculated surface energies for Azo-silica powders with different grafting densities. A decrease in surface energy with increased grafting density and longer carbon spacers is observed. Surface energy is given with its respective polar and dispersive components for samples before and after irradiation with UV light

System	Azo grafting density (molecules per nm ² (EA))	γ_s^d /mJ m ⁻²		γ_s^p /mJ m ⁻²		γ_s /mJ m ⁻²	
		No UV	UV	No UV	UV	No UV	UV
AzoOH-silica	0.50*	28.3 ± 1.7	24.3 ± 7.8	5.6 ± 0.1	8.5 ± 3.3	33.9 ± 1.7	32.8 ± 6.2
	1.0	25.5 ± 2.3	16.1 ± 5.1	7.8 ± 0.6	18.3 ± 1.8	33.3 ± 2.0	34.3 ± 6.0
	1.5	20.3 ± 1.6	15.5 ± 1.1	15.2 ± 3.0	12.9 ± 1.8	35.4 ± 3.1	28.3 ± 2.0
AzoO(CH ₂) ₃ OH-silica	0.6	6.2 ± 0.8	11.5 ± 1.2	25.0 ± 1.7	24.8 ± 0.3	31.2 ± 1.2	36.2 ± 0.9
	1.4	1.4 ± 0.3	2.4 ± 0.2	10.9 ± 1.4	15.5 ± 0.7	12.3 ± 0.6	17.9 ± 1.2
	1.7	1.61 ± 0.4	3.6 ± 0.4	13.7 ± 1.8	17.0 ± 0.6	15.3 ± 1.1	20.6 ± 1.3
AzoO(CH ₂) ₁₁ OH-silica	0.4	9.2 ± 2.4	5.2 ± 2.2	11.1 ± 2.6	22.2 ± 2.5	20.4 ± 1.7	27.5 ± 1.0
	1.5	1.2 ± 0.2	0.6 ± 1.5	2.9 ± 1.5	9.4 ± 0.6	4.1 ± 1.0	10.0 ± 0.6
	1.7	1.8 ± 0.5	1.4 ± 0.4	5.1 ± 0.7	16.2 ± 0.5	6.9 ± 1.3	17.6 ± 0.7



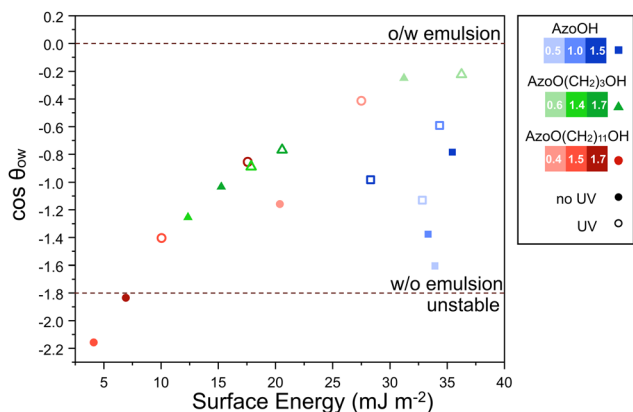


Fig. 7 Relationship between the surface energy, γ_{sw} , and predicted $\cos \theta_{ow}$ (from eqn (1)) for Azo-silica particles in diethyl adipate/water emulsions. A general decrease in surface energy with increasing grafting density/length of carbon spacer and a resulting decrease in $\cos \theta_{ow}$ is observed, indicating that emulsion formation is less favourable. Note that values are over-predicted and therefore exceed -1 and are used for comparison only. Darker shades of each colour represent higher grafting densities for a given azo derivative. Hollow shapes indicate that the surface energy was calculated using samples irradiated with UV light. The crossover between W/O and unstable emulsion was determined experimentally using optical microscopy on emulsions (particle conc. 5 mg mL^{-1} relative to H_2O) 24 h after preparation and is shown by the dashed lines.

grafting densities of 1.5 and 1.7 molecules per nm^2 ($\cos \theta_{ow} = -2.2$ and -1.8 , respectively). This is driven by the high interfacial energy with the aqueous phase ($\gamma_{sw} = 35$ and 42 mJ m^{-2} respectively). By comparison, the most hydrophobic AzoO(CH₂)₃OH-silica (1.4 molecules per nm^2) has a lower preference for the oil phase ($\cos \theta_{ow} = -1.2$) and so remains stable even at this higher grafting density. This $\cos \theta_{ow}$ value is similar to that for AzoO(CH₂)₁₁OH (0.4 molecules per nm^2) which also forms a stable emulsion. From this it is evident that, in diethyl adipate oil, there is a boundary point at *ca.* 8 mJ m^{-2} , below which particles aggregate in the oil phase.

For each of the light-responsive systems (AzoO(CH₂)₃OH- and AzoO(CH₂)₁₁OH-silica), irradiation with UV light (Fig. 7, hollow shapes) leads to an increase in the surface energy and a shift in $\cos \theta_{ow}$ to more positive values, reducing the preference for the oil phase. This effect is more pronounced at higher grafting densities and may be attributed to a higher surface concentration of Azo moieties. Consistent with optical microscopy, AzoO(CH₂)₁₁OH-silica (1.5 and 1.7 molecules per nm^2)-stabilised systems uniquely formed W/O emulsions in the *cis* state, but not in the *trans*. For the other emulsions, UV irradiation had negligible effect on the droplet morphology, as neither the *cis* nor *trans* states had a strong enough preference for the adipate oil phase to induce aggregation.

Predictions were also made for non-UV responsive *n*-hexadecane emulsions (ESI,† Fig. S14). Overall, results remain over-predicted, especially for polar AzoOH-silica emulsions. However, it can be seen that the most hydrophobic samples (*i.e.*, AzoO(CH₂)₁₁OH-silica, 1.4 and 1.7 molecules per nm^2) lie further from $\cos \theta_{ow} = 0$, which is in agreement with the observed transition from O/W to W/O morphology. We note

that, for some azo-silica samples, despite *cis* and *trans* isomers lying at different morphologies (*i.e.*, $\cos \theta_{ow}$ above or below 0), no change in emulsion inversion was observed. We attribute this to the imprecision of surface energy analysis noted earlier.

When designing light-responsive emulsions it is clear that both photoisomers need to have a sufficiently different affinities for the oil and water phases to enable different morphologies to form. It should also be noted that the results here are dependent on the choice of oil used in the emulsion. For example, calculations suggest that the use of a more polar oil (*e.g.*, ethylene glycol) would lower the adsorption energy of the particles at the oil–water interface ($\cos \theta_{ow}$ shifted closer to 0 – Fig. S10, ESI†) and as such, fewer of the emulsions would be expected to be stable in their native *trans* state.

Effect of particle concentration

Emulsions containing different particle concentrations were also prepared (2.5, 5, 10, 20 and 50 mg mL^{-1} with respect to the aqueous phase) to evaluate the surface coverage at the droplet interface and to see if UV irradiation had an effect on the composition of the droplets or the mechanism of stabilisation (ESI,† Table S5). Overall, there is an increase in droplet diameter with decreasing particle concentration, similar to previous studies.^{50,51} For the more hydrophobic emulsions it can be seen that a minimum particle concentration of 5 mg mL^{-1} is required to stabilise the emulsion, which supports the lower interfacial stability predicted from surface energy measurements.

Under ideal conditions, the droplet diameter is directly related to the number of stabiliser particles that are able to sit at the interface between the two phases:^{32,51}

$$\frac{1}{D} = \frac{m_p}{4CV_d\rho_p d_p} \quad (5)$$

where D is the droplet diameter (μm), m_p is the mass of particles (g), V_d is the volume of the dispersed phase (cm^3) and d_p is the particle diameter (μm). C is the surface coverage, which may be defined by the ratio of the interfacial surface area covered by the stabilising particles (approximating them to spheres) to the total surface area of the droplets. This parameter may be used to identify the process by which droplets are stabilised. ρ_p is the particle density (g cm^{-3}).

Fumed silica has a fractal aggregate structure and so its density is best described by:⁵⁰

$$\rho_p = \rho_{\text{SiO}_2} \left(\frac{d_p}{d_{\text{SiO}_2}} \right)^{D_f-3} \quad (6)$$

where D_f is the fractal dimension of silica (2.3)⁵⁰ and d_{SiO_2} is the size of the elementary particles of silica (20 nm).⁵⁰

Fig. 8(a) gives an example histogram of the droplet sizes obtained from optical microscopy for an AzoO(CH₂)₃OH-silica (1.4 molecules per nm^2) stabilised emulsion (20 mg mL^{-1}).

Fig. 8(b) shows a typical plot of mass *versus* $1/D$, where the y -axis values are given as a histogram to better represent the particle size distribution. At low particle loadings, eqn (5) holds, indicating that the emulsions form *via* limited coalescence.¹⁶



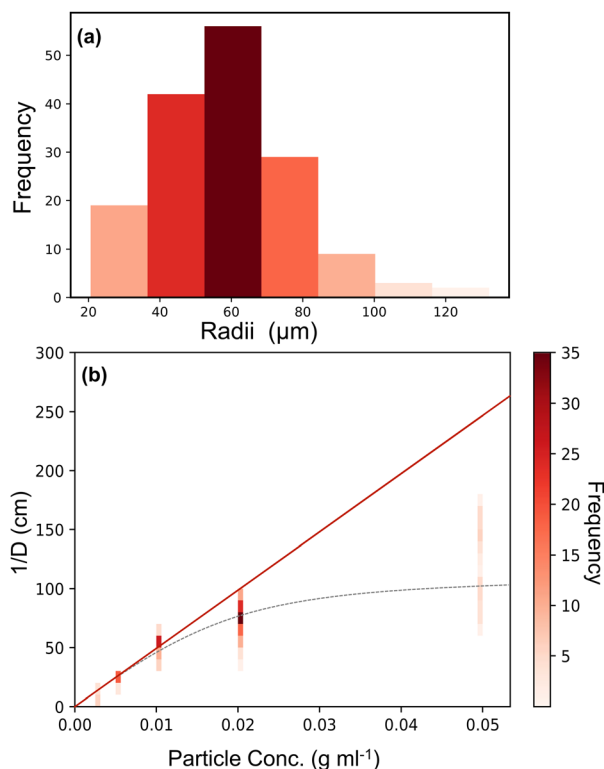


Fig. 8 Example plots used for the determination of surface coverage of AzoO(CH₂)₃OH-silica (1.4 molecules per nm²) particle emulsions. (a) Representative histogram of droplet sizes obtained from optical microscopy for a particle loading of 20 mg mL⁻¹. (b) Plot of inverse mean droplet diameter (1/D) against particle concentration (relative to the oil phase). The red line shows linearity at low concentration and regression is performed using the masses in this region (< 0.01 g mL⁻¹). The grey dashed line (---) is only to guide the eye and shows the non-linearity at higher particle concentrations. Coloured vertical bars are analogous to histogram (a) for each particle concentration. More examples can be found in the ESI,† Fig. S15.

However, at higher particle concentrations the values deviate from linearity, suggesting that other stabilisation mechanisms dominate, such as aggregation and network formation.⁵⁰

Using eqn (5) in conjunction with eqn (6), it can be seen that in general the samples show a surface coverage of roughly 20%, as reported in Table 3. This is true for all samples which are stable in their native *trans* state. This result is consistent with other studies that use fumed silica and the low coverage supports the idea that the interface is stabilised by networks of the branched fumed silica particles.⁵² The particles, which have a fractal aggregate morphology, are attached to the interface through a reduced number of anchoring points and protrude into the continuous phase.⁵¹

Notably, the *cis*-stabilised (UV irradiated) emulsions (AzoO(CH₂)₁₁OH-silica, 1.5 and 1.7 molecules per nm²) and the AzoO(CH₂)₃OH-silica (1.4 molecules per nm²) emulsion formed with a significantly higher surface coverage > 0.4. These are also three of the samples with the lowest surface energy. This suggests that the emulsions formed from the most hydrophobic particles are not well described by eqn (5) and are

Table 3 Surface coverage of particle-stabilised emulsion droplets calculated from the gradient of inverse mean droplet diameter against mass of stabilising particle and showing a difference in coverage between stable and UV-only stable emulsions. *Samples did not form spherical droplets and so were omitted from surface coverage calculations. **Samples did not form a stable emulsion

System	Grafting density (molecules per nm ²)	Surface coverage (± 0.01)	
		No UV	UV
AzoOH-silica	0.50*	N/A*	N/A*
	1.0	0.18	0.15
	1.5	0.20	0.18
AzoO(CH ₂) ₃ OH-silica	0.6	0.21	0.18
	1.4	0.58	0.20
	1.7	0.20	0.20
AzoO(CH ₂) ₁₁ OH-silica	0.4	0.18	0.21
	1.5	N/A**	0.42
	1.7	N/A**	0.39

not stabilised by the same mechanism. As shown by optical microscopy, greater particle aggregation occurs in the oil phase for these samples, decreasing the effective mass of particles available at the interface.

Design rules for light-responsive Pickering emulsions

Despite the limited range of morphologies encountered in this system, *i.e.*, W/O emulsions and unstable oil aggregates (Fig. 1 regions III and IV respectively), this work highlights several design considerations when creating light-responsive Pickering emulsions. (i) Foremost, the choice of photoswitch. Photoswitchable particles such as AzoO(CH₂)₁₁OH-silica remain isomerised for > 1 h and showed a decrease in their surface energy when switched. As a result, a shift from demulsified to emulsified state was seen. Azo-OH-silica, meanwhile, does not switch in a suitable time frame and so no change was seen in the emulsion morphology.

(ii) The length of carbon chain used to anchor the photoswitch to the particle surface is also important. Upon increasing from a zero-carbon spacer for Azo-OH-silica to a three-carbon chain for AzoO(CH₂)₃OH-silica, a photoswitch response was observed, which persisted long enough (> 10 min) to be measurable by DRUV spectroscopy. Moreover, longer spacers increase the hydrophobicity of the particle surface. Optical microscopy of particle-stabilised emulsions (10 mg mL⁻¹) showed AzoOH-silica (0.5 molecules per nm²) systems form non-spherical droplets due to their hydrophilic character, while AzoO(CH₂)₁₁-silica (1.5 and 1.7 molecules per nm²) do not form stable emulsions in their native *trans*-state due to their hydrophobic character.

(iii) The grafting density directly influences the particle hydrophobicity, but the relationship is non-linear. For example, a dramatic decrease in surface energy at above 1 molecule per nm² was observed for both AzoO(CH₂)₃OH-silica and AzoO(CH₂)₁₁OH-silica (*ca.* 15 mJ m⁻²). The effect that this has on the resulting emulsion is best seen for AzoO(CH₂)₁₁OH-silica, which forms W/O emulsions at 0.4 molecules per nm² and aggregates in the oil phase at 1.5 molecules per nm² for the *trans* form (Fig. 1 – III to IV). The *cis* isomer does not show the same behaviour.



(iv) Finally, the choice of oil is also important in determining which morphology is the most stable. Using the calculated surface energy, it is possible to predict the particle's preference for a given phase. For example, in a polar oil a hydrophobic particle (e.g., AzoO(CH₂)₁₁OH-silica, 1.5 molecules per nm²) will have a much greater preference for the oil phase and form aggregates. Therefore, by choosing an appropriate oil we can modulate the solid–oil interfacial energy so that in the *trans* state the particle forms aggregates, while in the *cis* state it forms a stable emulsion (see ESI,† Fig. S14).

In theory, these general principles can be used to access and to transition between any of the morphological regions shown in Fig. 1. For instance, if targeting a transition between aqueous aggregates to O/W emulsions upon irradiation (i.e., Fig. 1 – region I to II), a polar photoswitch should be chosen (short chain, modification with a hydrophilic group), which becomes more hydrophobic upon irradiation (decrease in surface energy) and a hydrophobic oil used. However, there are several limiting factors, such as oil–water miscibility which restricts the maximum polar character of the oil, and the minimum requirement for the number of photoswitch moieties on the surface required to induce changes in wettability upon irradiation. For the present example, increasing the grafting density of the hydrophilic photoswitch is likely to only have a minor effect on the hydrophobicity as bare fumed silica is already hydrophilic in character.

Conclusions

In summary, we have demonstrated that water-in-oil Pickering emulsions (water–diethyl adipate, 1 : 1 vol) can be stabilised by fumed silica particles that are functionalised with Azo photoswitches. Alkoxidation of the surface using alcohol-terminated azobenzenes proved to be a facile method for varying the surface density of photoswitches on fumed silica particles. Moreover, the surface hydrophobicity of the particles could be tuned by varying the length of the carbon spacer used to attach Azo to the particle surface. Notably, the most hydrophobic particles (AzoO(CH₂)₁₁OH-silica, 1.5 and 1.7 molecules per nm²), did not form stable emulsions in the native *trans*-state. However, irradiation of these systems with UV light resulted in emulsification, due to conversion of the grafted Azo-molecules to their more hydrophilic *cis*-state. These systems could be cycled between emulsified and demulsified states through the application of UV or blue light under shear, respectively. To our knowledge, these are the first examples of light-responsive Pickering emulsions that switch between W/O emulsions to aggregates in the oil phase.

A second emulsions system using hexadecane oil was chosen for comparison and a (> 14 day) stability study carried out. A mixture of O/W and W/O hexadecane emulsions of varying stability were formed, depending on the hydrophobicity of the particles, and they did not show the same light-response character. These trends were as expected based on the degree of surface modification and choice of photoswitch used on the particles.

Surface energy estimations and interfacial energy predictions combined with optical microscopy analysis on this model system have been harnessed as a powerful tool to understand emulsion stability. This approach has allowed us to identify a set of design principles that should enable light-responsive Pickering emulsions to be accessed more readily. These rules show that through consideration of the key components (particle, photoswitch and oil), it should be possible to tailor the system and its light-response to a given application. Furthermore, by adding electron-donating or withdrawing substituents, the azobenzene chromophore can be tailored to switch at different wavelengths,^{53,54} overcoming limitations such as poor penetration depth or UV exposure which could hinder use in biological systems.^{2,53} While we have demonstrated this for a model system based on the Azo chromophore, it is general and can be applied to other photoswitches, e.g., DASA, spiropyran. This work represents an important advance in the rational design of light-responsive Pickering emulsions and the methodologies employed can be applied anywhere that transient encapsulation is required.

Conflicts of interest

There are no conflicts of interest to declare.

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Notes and references

- 1 C. Albert, M. Beladjine, N. Tsapis, E. Fattal, F. Agnely and N. Huang, *J. Controlled Release*, 2019, **309**, 302–332.
- 2 C. L. G. Harman, M. A. Patel, S. Guldin and G.-L. Davies, *Curr. Opin. Colloid Interface Sci.*, 2019, **39**, 173–189.
- 3 J. Tang, P. J. Quinlan and K. C. Tam, *Soft Matter*, 2015, **11**, 3512–3529.
- 4 Y. Zhang, M. Zhang and H. Yang, *ChemCatChem*, 2018, **10**, 5224–5230.
- 5 M. Pera-Titus, L. Leclercq, J.-M. Clacens, F. De Campo and V. Nardello-Rataj, *Angew. Chem., Int. Ed.*, 2015, **54**, 2006–2021.
- 6 H. Yang, T. Zhou and W. Zhang, *Angew. Chem., Int. Ed.*, 2013, **52**, 7455–7459.
- 7 K. Liu, J. Jiang, Z. Cui and B. P. Binks, *Langmuir*, 2017, **33**, 2296–2305.
- 8 K. Y. A. Lin, H. Yang, C. Petit and W. der Lee, *J. Colloid Interface Sci.*, 2015, **438**, 296–305.
- 9 S. Tsuji and H. Kawaguchi, *Langmuir*, 2008, **24**, 3300–3305.
- 10 Y. Chen, Z. Li, H. Wang, Y. Pei, Y. Shi and J. Wang, *Langmuir*, 2018, **34**, 2784–2790.
- 11 E. A. Kelly, J. E. Houston and R. C. Evans, *Soft Matter*, 2019, **15**, 1253–1259.



- 12 Z. Li, Y. Shi, A. Zhu, Y. Zhao, H. Wang, B. P. Binks and J. Wang, *Angew. Chem., Int. Ed.*, 2021, **60**, 3928–3933.
- 13 R.-X. Bai, L.-H. Xue, R.-K. Dou, S.-X. Meng, C.-Y. Xie, Q. Zhang, T. Guo and T. Meng, *Langmuir*, 2016, **32**, 9254–9264.
- 14 Z. Chen, L. Zhou, W. Bing, Z. Zhang, Z. Li, J. Ren and X. Qu, *J. Am. Chem. Soc.*, 2014, **136**, 7498–7504.
- 15 Y. Chevalier and M.-A. Bolzinger, *Colloids Surf., A*, 2013, **439**, 23–34.
- 16 C. Whitby and E. Wanless, *Materials*, 2016, **9**, 626.
- 17 H. M. D. Bandara and S. C. Burdette, *Chem. Soc. Rev.*, 2012, **41**, 1809–1825.
- 18 A. A. Beharry and G. A. Woolley, *Chem. Soc. Rev.*, 2011, **40**, 4422–4437.
- 19 C. J. Barrett, J. Mamiya, K. G. Yager and T. Ikeda, *Soft Matter*, 2007, **3**, 1249.
- 20 G. S. Hartley and R. J. W. Le Fèvre, *J. Chem. Soc.*, 1939, 531–535.
- 21 C. Zong, M. Hu, U. Azhar, X. Chen, Y. Zhang, S. Zhang and C. Lu, *ACS Appl. Mater. Interfaces*, 2019, **11**, 25436–25444.
- 22 D. Manna, T. Udayabhaskararao, H. Zhao and R. Klajn, *Angew. Chem., Int. Ed.*, 2015, **127**, 12571–12574.
- 23 H. Zhao, S. Sen, T. Udayabhaskararao, M. Sawczyk, K. Kučanda, D. Manna, P. K. Kundu, J.-W. Lee, P. Král and R. Klajn, *Nat. Nanotechnol.*, 2016, **11**, 82–88.
- 24 R. Klajn, *Pure Appl. Chem.*, 2010, **82**, 2247–2279.
- 25 Z. Chu, Y. Han, T. Bian, S. De, P. Král and R. Klajn, *J. Am. Chem. Soc.*, 2019, **141**, 1949–1960.
- 26 P. K. Kundu, D. Samanta, R. Leizrowice, B. Margulis, H. Zhao, M. Börner, T. Udayabhaskararao, D. Manna and R. Klajn, *Nat. Chem.*, 2015, **7**, 646–652.
- 27 R. Klajn, *Chem. Soc. Rev.*, 2014, **43**, 148–184.
- 28 J. Frelichowska, M.-A. Bolzinger and Y. Chevalier, *Colloids Surf., A*, 2009, **343**, 70–74.
- 29 B. P. Binks and D. Yin, *Soft Matter*, 2016, **12**, 6858–6867.
- 30 Sigma Aldrich Certificate of Analysis, <https://www.sigmaaldrich.com/certificates/Graphics/COFAInfo/fluka/pdf/PDF640814.pdf>, (accessed 23 February 2022).
- 31 F. M. Mirabella, *Modern Techniques in Applied Molecular Spectroscopy*, John Wiley & Sons, New York, 1998.
- 32 B. P. Binks and S. O. Olusanya, *Chem. Sci.*, 2017, **8**, 708–723.
- 33 B. P. Binks and A. T. Tyowua, *Soft Matter*, 2013, **9**, 834–845.
- 34 V. F. Leavers, *Shape Detection in Computer Vision Using the Hough Transform*, Springer, London, 1992.
- 35 OpenCV, <https://opencv.org/>, (accessed 29 September 2021).
- 36 Hough-scan, <https://github.com/KRichardsF/Hough-Scan>, (accessed 28 May 2021).
- 37 R. Plenderleith, T. Swift and S. Rimmer, *RSC Adv.*, 2014, **4**, 50932–50937.
- 38 M. Fuji, T. Takei, T. Watanabe and M. Chikazawa, *Colloids Surf., A*, 1999, **154**, 13–24.
- 39 Sigma-Aldrich Product Information S5505, <https://www.sigmaaldrich.com/deepweb/assets/sigmaaldrich/product/documents/246/034/s5505pis.pdf>, (accessed 8 November 2021).
- 40 H. J. H. Kang, R. F. Ali, M. T. Y. Paul, M. J. Radford, I. Andreu, A. W. H. Lee and B. D. Gates, *Chem. Commun.*, 2019, **55**, 10452–10455.
- 41 L. T. Zhuravlev, *Colloids Surf., A*, 2000, **173**, 1–38.
- 42 A. Kawamura, S. Ueno, C. Takai, T. Takei, H. Razavi-Khosroshahi and M. Fuji, *Adv. Powder Technol.*, 2017, **28**, 2488–2495.
- 43 P. Sierocki, H. Maas, P. Dragut, G. Richardt, F. Vögtle, L. De Cola, F. Brouwer and J. I. Zink, *J. Phys. Chem. B*, 2006, **110**, 24390–24398.
- 44 Z. Wu, X. Li, X. Jiang, T. Xie, H. Li, G. Zhang and J. Jiang, *Phys. Chem. Chem. Phys.*, 2019, **21**, 21030–21037.
- 45 W. A. Zisman, *Adv. Chem.*, 1964, **43**, 1–51.
- 46 D. K. Owens and R. C. Wendt, *J. Appl. Polym. Sci.*, 1969, **13**, 1741–1747.
- 47 F. M. Fowkes, *Ind. Eng. Chem.*, 1964, **56**, 40–52.
- 48 C. J. Van Oss, R. J. Good and M. K. Chaudhury, *J. Colloid Interface Sci.*, 1986, **111**, 378–390.
- 49 B. P. Binks and J. H. Clint, *Langmuir*, 2002, **18**, 1270–1273.
- 50 J. Frelichowska, M.-A. Bolzinger and Y. Chevalier, *J. Colloid Interface Sci.*, 2010, **351**, 348–356.
- 51 S. Arditty, C. P. Whitby, B. P. Binks, V. Schmitt and F. Leal-Calderon, *Eur. Phys. J. E: Soft Matter Biol. Phys.*, 2003, **11**, 273–281.
- 52 H. Katepalli, V. T. John, A. Tripathi and A. Bose, *J. Colloid Interface Sci.*, 2017, **485**, 11–17.
- 53 D. Wang, W. Zhao, Q. Wei, C. Zhao and Y. Zheng, *Chem-PhotoChem*, 2018, **2**, 403–415.
- 54 J. Calbo, A. R. Thawani, R. S. L. Gibson, A. J. P. White and M. J. Fuchter, *Beilstein J. Org. Chem.*, 2019, **15**, 2753–2764.

