




Cite this: *New J. Chem.*, 2023, 47, 9077

Fluorescent 7-azaindole *N*-linked 1,2,3-triazole: synthesis and study of antimicrobial, molecular docking, ADME and DFT properties†

Kanika Sharma,^a Ram Kumar Tittal,^a *^a Kashmiri Lal,^b ^b Ramling S. Mathpati^a and Ghule Vikas D. ^a

Two libraries of biologically essential and fluorescent 7-azaindole *N*-linked benzyl 1,2,3-triazole (**4a–4i**) and 7-azaindole *N*-linked phenyl 1,2,3-triazole (**4j–4q**) hybrids have been designed and synthesized from *N*-propargyl 7-azaindole, *i.e.*, 1-prop-2-ynyl-1*H*-pyrrolo[2,3-*b*]pyridine (**2**) with benzyl azides (**3a–3i**) and phenyl azides (**3j–3q**), respectively, *via* a Cu(I)-catalyzed alkyne azide cycloaddition (CuAAC) click reaction. The structures of the synthesized hybrids were confirmed with the help of the IR, ¹H-NMR, ¹³C-NMR, and HRMS data. However, the structure of one of the solid crystalline compounds, **4f**, was finally supported by its single-crystal X-ray data. *In vitro*, antimicrobial activity results revealed that most compounds exhibited superior efficacy towards *Aspergillus* compared to fluconazole. Compounds **4e**, **4f**, and **4g** showed high potency against *A. niger* with MIC values of 0.0193, 0.0170, and 0.0187 μmol mL⁻¹, respectively. Compound **4f** also showed increased activity against *P. aeruginosa* and *E. coli* (MIC: 0.0170 μmol mL⁻¹). The most promising compounds, **4d**, **4f**, and **4n**, showed excellent binding energies of -8.81, -8.76, and -8.79 kcal mol⁻¹ with a dynamic DNA site 14α-demethylase lanosterol enzyme (PDB ID:1EA1). Chemical properties such as chemical potential (μ), chemical hardness (η), and electrophilicity index (ω) were studied by DFT at the B3LYP level with a 6-311G(d,p) basis set. All compounds excellently follow Lipinski's rule of five.

Received 14th January 2023,
Accepted 3rd April 2023

DOI: 10.1039/d3nj00223c

rsc.li/njc

Introduction

Post-corona (COVID-19) effects have significantly threatened immune-compromised people and corona-recovered patients. Different complications are observed in COVID-19 patients, such as severe respiratory syndromes due to invasive pulmonary aspergillosis, also called COVID-19-associated pulmonary aspergillosis (CAPA), and other *Aspergillus* genus-based infections. The rapid growth of *Aspergillus* infections leads to critical conditions in infected patients with respiratory disorders. Unfortunately, in many cases, diseases due to the *Aspergillus* genus are not diagnosed at the early stage due to its rapid growth, which leads to severe respiratory impairment.¹ Sometimes, prolonged use of steroids and other heavy antibiotic drugs during the treatment of COVID-19 and other diseases causes loss of the immune system. It weakens the defense mechanism of the body to fight

pathogenic attacks, especially fungal attacks, which increases the risk for similar and other severe fungal infections.² The most commonly reported fungal infections in patients with COVID-19 include aspergillosis, candidiasis, and mucormycosis, also called “black fungus”.³ The intensity of various infectious pathogenic diseases, especially viral and fungal diseases, is increasing daily. It was observed that these microbial infections are asymptomatic, which slowly progress to active disease. The situation becomes more complicated when some microbial strains resist any drug(s). It alarms an immediate concern for developing some new class of antimicrobial drugs to eliminate resistant microbial strains with a different mode of action.⁴ Our lab employed a molecular hybridization approach for the development of new chemicals with improved properties.⁵ Molecular assimilation involves a combination of two or more biologically significant compounds to develop a new hybrid molecule with augmented properties compared to the parent fragments.⁶

7-Azaindole (7-AI) is well-known for its chromophoric properties and is a basic 7-azatryptophan (7-ATph) unit. 7-ATph is a non-natural amino acid. It has been used for the development of synthetic peptides and bacterial proteins.⁷ 7-AI is a better substitute for tryptophan (Tph) and is used as an optical probe for protein structure over most of the physiological pH range.⁸

^a Department of Chemistry, National Institute of Technology Kurukshetra, Haryana 136119, India. E-mail: rktittalid@nitkkr.ac.in

^b Department of Chemistry, GJUS&T Hisar, Haryana 12500, India

† Electronic supplementary information (ESI) available. CCDC 2192737 (**4f**). For ESI and crystallographic data in CIF or other electronic format see DOI: <https://doi.org/10.1039/d3nj00223c>

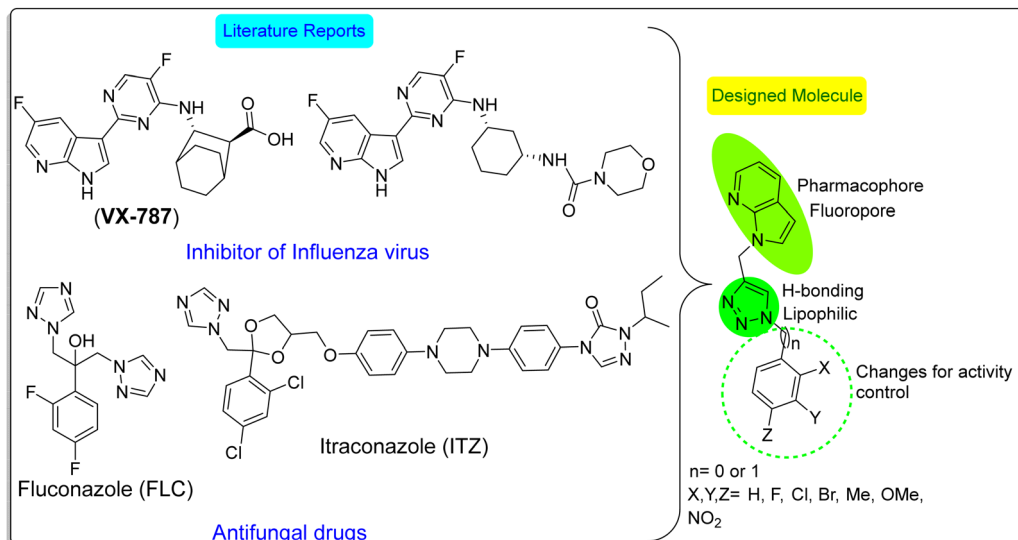


Fig. 1 Designed 7-azaindole *N*-linked aryl 1,2,3-triazoles.

Although the charge transfer (π - π^*) excited state of the 7-AI dimer has traditionally been considered as a model for base pairing in DNA⁹ molecules, the double proton transfer properties of 7-AI in water and other protic solvents such as methanol made it a suitable candidate for biological applications.¹⁰ Surprisingly, 7-AI moiety has not been given proper attention for developing advanced antimicrobial drugs except for one report for synthesizing only some derivatives with 4-Cl or 4-Br substituted 7-AI, e.g., 1-((1-(4-substituted benzyl)-1*H*-1,2,3-triazol-4-yl)methyl)-4-halo-1*H*-pyrrolo[2,3-*b*]pyridine.¹¹ However, Michael P. Clark and his group reported the development of a novel AI-based compound, VX-787 (Fig. 1), which was a first-in-class, orally bioavailable inhibitor of human influenza virus replication. It offers treatment for seasonal and pandemic influenza with essential advantages in care treatments, such as potency and efficacy.^{12–15} Benhamou *et al.* have utilized the fluorescence properties of azoles for constructing a molecular tool as a fluorescent probe to access detailed information about the site of drug action *via* real-time imaging. For the last three decades, azole-based antifungal drugs, especially fluconazole (FLC), as shown in Fig. 1, have extensively been explored as a first-line treatment drug for fatal fungal diseases. However, resistance to azole drugs, especially for *Candida* and *Aspergillus* species, has become a significant clinical concern.¹⁶ The much interesting and fascinating properties of the 7-AI moiety motivated us to utilize this moiety to construct some hybrid lead molecules.

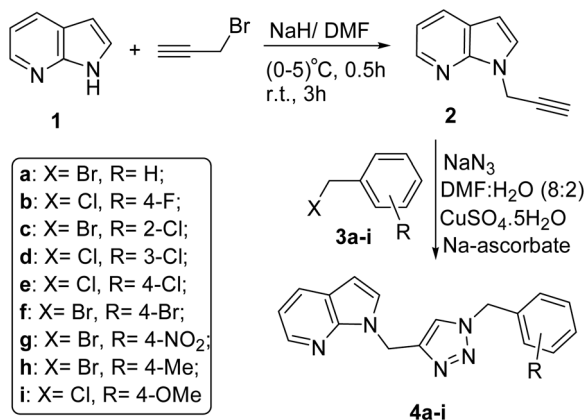
The pharmacophoric properties of the five-membered 1,2,3-triazole, such as its lipophilic character, aromatic stability, fair dipole moment, and resistance to both acid and base hydrolysis, made it a suitable pharmacophoric candidate for the development of medicinally important compounds.^{17–19} Also, the lipophilicity and H-bonding features of triazoles help them associate and bind with biological targets such as DNA to improve solubility and metabolic stability under physiological conditions.²⁰ The property of azoles to inhibit the antifungal target lanosterol 14 α -demethylase¹⁶ and their broad spectrum

of applications in medicine and materials science further add new dimensions to their importance.^{21–24} The important biological applications of 1,4-disubstituted 1,2,3-triazoles include their antifungal,²⁵ antibacterial,^{26,27} anticancer,^{28,29} antiviral,³⁰ anti-tubercular,²² anti-HIV,^{31,32} antidiabetic,³³ anti-inflammatory,^{22,34} antimalarial,³⁵ antiallergic,³⁶ and antioxidant properties.^{37,38} The regioselectivity compromised thermal synthesis of 1,2,3-triazole initially proposed by Huisgen^{39,40} was effortlessly modified and improved by Sharpless and Meldal using a Cu(I)-catalyst under the name of “click chemistry,” which is also called the copper-catalyzed alkyne-azide cycloaddition (CuAAC) reaction, and it got tremendous recognition.^{41–45} The wide use of CuAAC led to its use up to the biological level under “Bioorthogonal Chemistry” by Carolyn Bertozzi, who recently shared a Nobel Prize in the field.^{46–48}

In the current scenario, using fluorescent bioactive molecules for fluorescent labeling methods has become a vital tool in medical and biological applications.¹⁶ The abovementioned observations and our group’s research interest^{49–51} motivated us to synthesize 7-AI linked 1,2,3-triazole hybrid molecules as fluorescent and antimicrobial agents. Herewith, we report the synthesis of two libraries of biologically significant 7-azaindole *N*-linked benzyl/phenyl appended 1,2,3-triazole hybrids *via* a CuAAC-based click reaction as potential fluorescent antifungal agents.

Results and discussion

For the synthesis of 7-azaindole *N*-linked benzyl 1,2,3-triazole (**4a–4i**) hybrids as presented in Scheme 1, commercially available 7-azaindole (**1**) was subjected to *N*-propargylation as per the reported procedure,⁵² with the help of propargyl bromide in the presence of sodium hydride in dimethylformamide. The as-obtained *N*-propargyl 7-azaindole, *i.e.*, 1-prop-2-ynyl-1*H*-pyrrolo[2,3-*b*]pyridine (**2**), was purified by column chromatography using an ethylacetate: *n*-hexane (1:9 v/v) solution for eluting the pure compound in 80%



Scheme 1 Synthesis of 7-azaindole linked benzyl 1,2,3-triazoles (**4a–4i**).

yield. Compound **2** was used as a dipolarophile to synthesize the 7-azaindole *N*-linked 1,2,3-triazole hybrids (**4a–4q**).

First, 7-azaindole *N*-linked benzyl 1,2,3-triazole (**4a–4i**) derivatives were prepared using the CuAAC mediated click reactions from *in situ* generated organic azides with *N*-propargyl 7-azaindole (**2**) using CuSO₄·5H₂O (5 mol%) and sodium ascorbate (10 mol%) as a catalyst at room temperature. Organic azides were prepared as per the reported method from benzyl halides (**3a–3i**).⁵³ The desired representative 1,2,3-triazole compound **4a** was isolated in 97% yield through a short silica gel band of column chromatography using an ethyl acetate and *n*-hexane (3 : 7 v/v) solvent system for elution. The synthesized compound **4a** was thoroughly characterized with FTIR, ¹H-NMR, ¹³C-NMR, and ESI-MS data. The formation of 1,2,3-triazole was ascertained by the disappearance of the alkyne ≡C–H and C≡C stretching bands at 3292 cm⁻¹ and 2121 cm⁻¹, respectively, and the appearance of a new characteristic stretching band of 1,2,3-triazole due to =C–H stretching at ν_{max} 3119 cm⁻¹ in the FTIR spectrum of compound **4a**. The absorption bands at 1508 and 1306 cm⁻¹ due to the stretching vibrations of –C=N and –C–N of the 7-azaindole moiety were retained in compound **4a**, which appeared initially in 7-azaindole linked alkyne **2** at the exact wavenumber, *i.e.*, 1508 and 1306 cm⁻¹.

The ¹H-NMR spectrum of **4a** showed all the proton signals for eleven C–H signals of extended aromatic protons from δ 6.45 to 8.30 ppm and the rest of four aliphatic proton signals as singlets at δ 5.43 along with δ 5.57 ppm for two CH₂ protons. A sharp singlet signal at δ 7.41 ppm, which could be attributed to the triazolyl proton, showed confirmation of the triazole ring in the compound. The ¹³C-NMR spectrum of **4a** showed all the required seventeen carbon signals at δ 147.40, 144.58, 143.05, 136.58, 129.41, 129.31, 129.18, 128.70, 128.53, 124.00, 120.60, 116.38, 100.22, 53.33, and 39.41 including two of the overlapped phenyl carbon signals at δ 129.319 (2C) and 128.533 (2C) which were counted for two carbons.

The ESI mass spectrum of compound **4a** exhibited a peak at *m/z* 290.14 (M + H)⁺, which is precisely similar to the calculated value of (M + H)⁺ of 290.14 and thus showed good agreement with the molecular formula (C₁₇H₁₅N₅H⁺).

The final confirmation about the formation of 7-azaindole *N*-linked aryl 1,2,3-triazole derivatives was authenticated after

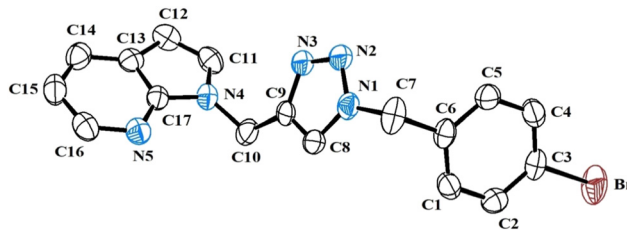


Fig. 2 ORTEP diagram of compound **4f** (H-atoms omitted for more clarity).

revealing the correct structural parameters of the anticipated structure about one of the solid crystalline compounds with the help of X-ray crystallographic data of **4f** (recrystallized from a solution of ethanol and chloroform). The crystal structure of compound **4f** and other details are given in Fig. 2 and Table SI-1 (ESI[†]). The X-ray crystallographic structure of compound **4f** (CCDC no. 2192737) crystallizes in a triclinic crystal system in the P $\bar{1}$ (2) space group. It was observed that two crystallographically independent molecular units were present in an asymmetric unit of compound **4f**, as shown in Fig. SI-1 (ESI[†]).

Before moving to various derivatization of 7-azaindole *N*-linked benzyl 1,2,3-triazoles, efforts were made to improve the yield of the reaction products by changing different solvent systems such as *t*-BuOH, H₂O, DMSO, THF, and DMF. The reaction temperature was also optimised, as detailed in Table 1. It was observed that the reaction did not complete in a single solvent such as *t*BuOH, DMSO, THF, or H₂O due to the solubility issue of inorganic catalysts and an organic compound. Thus, a minimum amount of H₂O for the catalyst solubility in a suitable solvent such as DMF for the solubility of the organic compound was the best choice for the completion of the reaction. All other derivatives of 7-azaindole *N*-linked benzyl 1,2,3-triazoles, *i.e.*, **4b–4i**, were synthesized and characterized with the help of the same spectral pattern of bands and peaks in the FTIR, ¹H-NMR, ¹³C-NMR, and ESI-MS data. The product reproducibility and

Table 1 Standardized reaction conditions for the reaction of 7-azaindole *N*-linked alkyne **2** and *in situ* generated aryl azides with CuSO₄·5H₂O/Na-ascorbate in some suitably selected solvent systems to form 1,2,3-triazole **4a**^a

S. no.	Solvent(s) solution (v/v)	Temperature (°C)	Time (h)	Yield ^b (%)
1	<i>t</i> -BuOH : H ₂ O (1 : 1)	35	7	74
2	<i>t</i> -BuOH : H ₂ O (8 : 2)	35	5.5	78
3	<i>t</i> -BuOH	35	24	^c
4	DMSO : H ₂ O (1 : 1)	35	5	82
5	DMSO : H ₂ O (8 : 2)	35	4	86
6	DMSO	35, 60	24	^c
7	THF : H ₂ O (1 : 1)	35	6	75
8	THF : H ₂ O (8 : 2)	35	4.5	79
9	THF	35, 60	24	^c
10	DMF : H ₂ O (1 : 1)	35	5	88
11	DMF : H ₂ O (8 : 2)	35	3	97
12	DMF	35	24	68
13	H ₂ O	35, 60	24	^c

^a Reaction conditions: aryl azide and 7-azaindole *N*-linked alkyne **2** (1 mmol); CuSO₄·5H₂O (10 mol%) and Na-ascorbate (20 mol%), rt, and 3–24 h. ^b Yield refers to purification by recrystallization. ^c No reaction.

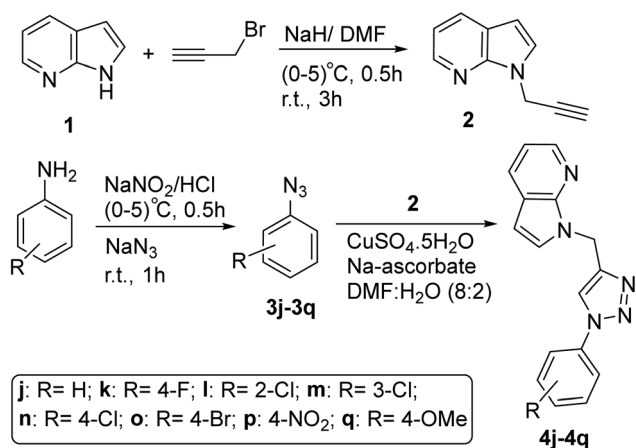
Table 2 Synthesis of 7-azaindole *N*-linked aryl 1,2,3-triazoles **4a–4q** from 7-azaindole linked alkyne **2**^a

S. no.	Compound	PhCH ₂ /Ph	R	Time (h)	Yield ^b (%)
1	4a	PhCH ₂	H	3	94
2	4b	PhCH ₂	4-F	5	87
3	4c	PhCH ₂	2-Cl	6	83
4	4d	PhCH ₂	3-Cl	7	92
5	4e	PhCH ₂	4-Cl	5	90
6	4f	PhCH ₂	4-Br	6	87
7	4g	PhCH ₂	4-NO ₂	4	80
8	4h	PhCH ₂	4-Me	7	92
9	4i	PhCH ₂	4-OMe	4	82
10	4j	Ph	H	3	84
11	4k	Ph	4-F	6	85
12	4l	Ph	2-Cl	5	92
13	4m	Ph	3-Cl	6	85
14	4n	Ph	4-Cl	4	84
15	4o	Ph	4-Br	5	84
16	4p	Ph	4-NO ₂	1	83
17	4q	Ph	4-OMe	4	80

^a **2** (1 mmol), **3a–q** (1 mmol), sodium azide (1.2 mmol), CuSO₄·5H₂O (10 mol%), Na-ascorbate (20 mol%), DMF:H₂O (8:2 v/v ratio), rt (35 °C), and 3–7 h. ^b Yields refer to purification *via* column chromatography using ethyl acetate:hexane with increased polarity gradient.

generalization of the reaction conditions were tested by extending the scheme for the synthesis of other derivatives of 7-azaindole *N*-linked benzyl 1,2,3-triazoles (**4a–4i**) in good to excellent yields of 80–94%, as detailed in Table 2.

To extend the library of novel 7-azaindole *N*-linked aryl 1,2,3-triazoles, we thought to synthesize phenyl azides from different aniline derivatives using the reported diazotization and azidation methods.⁵⁴ Thus prepared phenyl azides (**3j–q**), as shown in Scheme 2, were subjected to react with *N*-propargyl 7-azaindole (**2**) using CuSO₄·5H₂O (5 mol%) and sodium ascorbate (10 mol%) as a catalyst at room temperature for 1 to 6 h stirring to yield pure 7-azaindole *N*-linked phenyl 1,2,3-triazole (**4j–4q**) in 80–92% isolated yield after column chromatography. The library of 7-azaindole *N*-linked phenyl 1,2,3-triazoles (as detailed in Table 2, entries 10–17) were efficiently synthesized and characterized by following the similar spectral patterns of bands and signals in the FTIR, ¹H-NMR, ¹³C-NMR, and ESI-MS data.

**Scheme 2** Synthesis of 7-azaindole *N*-linked phenyl 1,2,3-triazoles (**4j–4q**).

The representative compound of this series, 7-azaindole *N*-linked phenyl 1,2,3-triazole **4j**, obtained in 84% yield, was successfully characterized using the FTIR, ¹H-NMR, ¹³C-NMR, and ESI-MS data. The disappearance of the alkyne $\text{C}\equiv\text{C-H}$ and $\text{C}\equiv\text{C}$ stretching bands at 3292 cm⁻¹ and 2121 cm⁻¹, respectively, and the appearance of a new characteristic stretching band of C-H at ν_{max} 3144 cm⁻¹ in the FTIR spectrum confirmed the formation of the 1,2,3-triazole ring. The absorption bands at 1506 and 1300 cm⁻¹ due to the stretching vibrations of -C=N and -C-N of the 7-azaindole moiety were observed nearly in the same region as in 7-azaindole linked alkyne (**2**) at 1508 and 1306 cm⁻¹.

The ¹H-NMR spectrum of compound **4j** confirmed all eleven aromatic protons from δ 6.48 to 8.35 ppm integrating and showing a sharp singlet signal at δ 5.68 ppm integrating two aliphatic protons. A sharp singlet that appeared at δ 7.92 ppm was attributed to the triazolyl proton, which confirmed triazole ring formation.

The ¹³C-NMR spectrum of **4j** showed all the indispensable sixteen carbon signals at δ 147.08, 142.75, 136.87, 129.63, 129.05, 128.74, 128.05, 125.22, 120.81, 120.50, 116.01, 115.59, 100.35 and 39.54 including two of the overlapped phenyl carbon signals at δ 129.638 (2C) and 120.506 (2C).

The ESI mass spectrum of compound **4j** exhibited a peak at m/z 276.12 (M + H)⁺, which is precisely similar to the calculated value of (M + H)⁺ as per the molecular formula (C₁₆H₁₃N₅H⁺): 276.12. Both theoretical and experimental m/z peaks due to (M + H)⁺ are in good agreement with each other.

Pharmacological study

Antibacterial activity study

All 7-azaindole *N*-linked aryl 1,2,3-triazoles **4a–q** including the precursor alkyne **2**, as shown in Table 3, were screened for *in vitro* antibacterial activity against gram (+) strains *B. subtilis* and *S. aureus* and gram (–) strains *E. coli* and *P. aeruginosa* using the standard serial dilution method. In this method, a solution of different triazoles was prepared for stock using DMSO.⁵⁵ The triazole's minimum inhibitory concentration (MIC) was recorded in $\mu\text{mol mL}^{-1}$. From the activity data, it was observed that compound **4f** and **4o**, containing a Br-substituted benzene ring with MIC values of 0.0170 $\mu\text{mol mL}^{-1}$ (*P. aeruginosa* and *E. coli*) and 0.0177 $\mu\text{mol mL}^{-1}$ (*B. subtilis* and *E. coli*), respectively, showed significant results as compared to 7-azaindole linked alkyne **2** with an MIC value of 0.0800 $\mu\text{mol mL}^{-1}$. The bioactivity results revealed that the antibacterial activity of 1,2,3-triazoles is nearly five times better than that of the precursor alkyne. Further modifications can improve the activity of triazoles in the moiety compared to the reference drug ciprofloxacin. The SAR (structure–activity relationship) study showed that all 7-azaindole linked 1,4 disubstituted 1,2,3-triazole moieties reflected better potency than the alkyne, which confirmed that the incorporation of the 1,2,3-triazole unit into 7-azaindole linked alkyne **2** augmented the antibacterial activity to a better extent.

Evaluation of the antifungal activity

All the newly synthesized 7-azaindole linked 1,4 disubstituted 1,2,3-triazole moieties were subjected to *in vitro* antifungal

Table 3 *In vitro* biological data of alkyne **2** and triazole hybrids **4a–4q**, MIC in $\mu\text{mol mL}^{-1}$ ^a

S. no	Compounds	M. Wt.	A	B	C	D	E	F
1.	2	156.18	0.0800	0.0800	0.0800	0.0800	0.0800	0.0400
2.	4a	289.33	0.0432	0.0432	0.0432	0.0216	0.0216	0.0432
3.	4b	307.32	0.0407	0.0407	0.0407	0.0407	0.0814	0.0204
4.	4c	323.78	0.0386	0.0772	0.0193	0.0386	0.0772	0.0386
5.	4d	323.78	0.0386	0.0193	0.0386	0.0386	0.0193	0.0386
6.	4e	323.78	0.0386	0.0772	0.0193	0.0386	0.0772	0.0193
7.	4f	368.23	0.0170	0.0340	0.0170	0.0679	0.0679	0.0170
8.	4g	334.33	0.0374	0.0374	0.0187	0.0374	0.0748	0.0187
9.	4h	303.36	0.0412	0.0412	0.0206	0.0412	0.0412	0.0824
10.	4i	319.36	0.0391	0.0391	0.0196	0.0391	0.0783	0.0783
11.	4j	275.31	0.0454	0.0454	0.0227	0.0908	0.0908	0.0454
12.	4k	293.30	0.0426	0.0426	0.0426	0.0852	0.0426	0.0426
13.	4l	309.75	0.0202	0.0404	0.0404	0.0404	0.0807	0.0404
14.	4m	309.75	0.0807	0.0202	0.0404	0.0807	0.0807	0.0404
15.	4n	309.75	0.0202	0.0404	0.0404	0.0404	0.0404	0.0202
16.	4o	354.20	0.0353	0.0177	0.0177	0.0706	0.0706	0.0353
17.	4p	320.31	0.0390	0.0781	0.0195	0.0781	0.0781	0.0390
18.	4q	305.33	0.0409	0.0409	0.0205	0.0409	0.0819	0.0409
19.	Ciprofloxacin	0.0094	0.0094	0.0094	0.0094	—	—	—
20.	Fluconazole	—	—	—	—	0.0408	0.0408	—

A: *P. aeruginosa*; B: *B. subtilis*; C: *E. coli*; D: *S. aureus*; E: *C. albicans*; and F: *A. niger*.

activities using *C. albicans* and *A. niger* as fungal strains with the help of the standard dilution method.⁵⁶ The antifungal data were recorded in terms of $\mu\text{mol mL}^{-1}$. As per the tested results, compounds **4d** and **4n** having a Cl-moiety as an EWG on the benzene ring with minimum inhibitory concentration (MIC) values of 0.0193 and 0.0404 $\mu\text{mol mL}^{-1}$, respectively, reflected better activity for the *C. albicans* fungal strain as compared to the reference drug fluconazole with an MIC value of 0.0408 $\mu\text{mol mL}^{-1}$. The observations from the results indicated that compound **4d** (MIC: 0.0193 $\mu\text{mol mL}^{-1}$) reflected more than two-fold superior antifungal activity compared to the reference drug fluconazole (MIC: 0.0408 $\mu\text{mol mL}^{-1}$). However, for the *A. niger* fungal strain, almost all 1,2,3-triazole compounds (**4b–4g** and **4l–4q**) except for **4a** and **4h–4k** compounds showed better antifungal activity as compared to the standard reference drug fluconazole (MIC: 0.0408 $\mu\text{mol mL}^{-1}$). However, compound **4f** with an MIC value of 0.0170 $\mu\text{mol mL}^{-1}$ was found to be more influential among all 7-AI linked 1,2,3-triazoles against *P. aeruginosa* and *E. coli* bacterial strains along with the *A. niger* fungal strain. In addition, compounds **4d** and **4n** containing a Cl-moiety on the benzene ring also exhibited better activity with MIC values of 0.0193, 0.0193, and 0.0386 and 0.0202, 0.0404, and 0.0202 $\mu\text{mol mL}^{-1}$, for *B. subtilis*, *C. albicans*, and *A. niger* and *P. aeruginosa*, *C. albicans*, and *A. niger*, respectively, as compared to the reference antibacterial drug ciprofloxacin (MIC: 0.0094 $\mu\text{mol mL}^{-1}$) and antifungal drug fluconazole

(MIC: 0.0408 $\mu\text{mol mL}^{-1}$). After reviewing the antifungal data and their structure–activity relationship, an augmentation in the biological activity was observed due to the incorporation or linkage of the 7-azaindole moiety with different 1,2,3-triazole units. Substantially, the test results revealed that compounds **4d** and **4n** are most potent against both fungal strains and can be a better substitute for the pre-existing antifungal drug fluconazole.

Fluorescence activity

The UV-Vis spectrum of 7-AI (**1**), 7-AI-linked alkyne (**2**), and 7-azaindole-linked aryl 1,2,3-triazoles (**4a–4i**), as shown in Fig. SI-2 (ESI[†]), showed an absorbance peak from 265 to 285 nm which could be due to the presence of the 7-azaindole moiety in MeOH (concentration 2.7–8.4 mmol). The fluorescence change in compounds 7-AI (**1**), 7-AI-linked alkyne (**2**), and 7-azaindole-linked aryl 1,2,3-triazoles (**4a–4q**) was checked under UV light as shown in Fig. 3, which revealed that most of the synthesized 1,2,3-triazoles appeared fluorescent under UV light through naked eyes except for the 4-NO₂ (a well-known fluorescence quencher)⁵⁷ benzyl substituted 1,2,3-triazole **4g**. However, 7-AI-linked alkyne **2** seemed to be the most robust fluorescent compound through naked eyes under UV light. Also, a relative fluorescence intensity change in compounds 7-AI (**1**), 7-AI-linked alkyne (**2**), and 7-azaindole-linked aryl 1,2,3-triazoles (**4a–4q**) in MeOH (concentration 2.7–8.4 mmol) with $\lambda_{\text{ex}} = 285$ nm was monitored, as shown in Fig. 4. The relative decreasing order in the fluorescence intensity change in compounds 7-AI (**1**), 7-AI-linked alkyne (**2**), and 7-azaindole-linked aryl 1,2,3-triazoles (**4a–4q**) as shown below revealed that compound **4d** having a 3-Cl benzyl-substituted 1,2,3-triazole with 799.1815 a.u. intensity is highly fluorescent with the least intensity shared by compound **4p** having a 4-NO₂ (a well-known fluorescence quencher) phenyl-substituted 1,2,3-triazole moiety.

Decreasing order of the fluorescence intensity of compounds (**1**, **2**, and **4a–4q**): **4d** (799.1815) > **4c** (781.8753) > **4f** (774.9273) > **4h** (763.4579) > **4i** (754.3394) > **4b** (750.0651) > **4a** (737.9546) > **4e** (619.8882) > **4l** (592.2746) > **4k** (528.8380) > **2** (424.1054) > **4j** (311.8661) > **4n** (306.0840) > **4o** (298.9996) > **4q** (284.8308) > **4m** (279.5175) > **1** (08.3395) > **4g** (07.3845) > **4p** (05.8384).

Molecular docking

The studies of antimicrobial activity results revealed that the incorporation of the 1,2,3-triazole moiety improved the antifungal activities of almost all the synthesized triazoles more

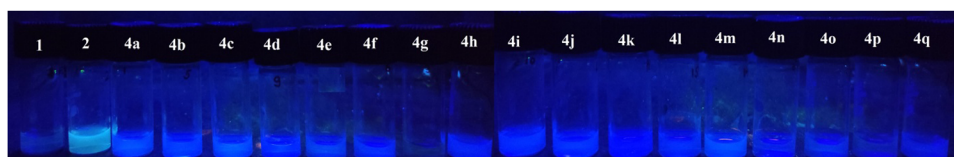


Fig. 3 A naked eye-detectable fluorescence change in 7-AI (**1**), 7-AI-linked alkyne (**2**), and 7-azaindole-linked aryl 1,2,3-triazoles (**4a–4q**) under UV light.

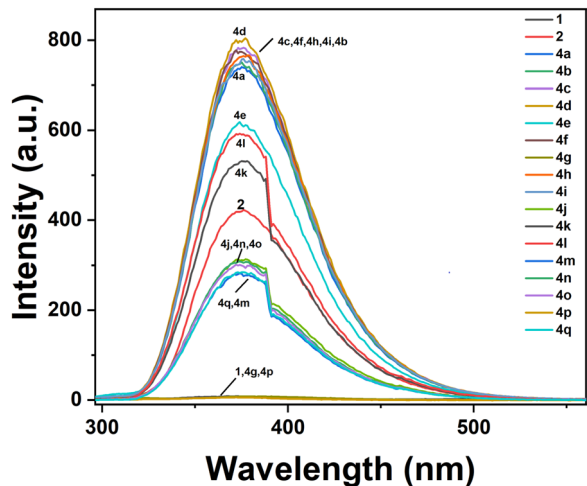


Fig. 4 A relative fluorescence intensity change in compounds 7-AI (**1**), 7-AI-linked alkyne (**2**), and 7-azaindole-linked aryl 1,2,3-triazoles (**4a–4q**) in MeOH (concentration 2.8–8.4 mmol), with $\lambda_{\text{ex}} = 285$ nm.

efficiently for both fungal strains than the antibacterial activities. The docking simulations of the predecessor alkyne **2** and 7-azaindole *N*-linked triazoles (**4a–4q**) were studied by performing molecular docking with an active site of the fungal enzyme DNA gyrase lanosterol 14α -demethylase (PDB ID: 1EA1) by using software Auto dock Vina open-source^{58–61} as shown in Table SI-2 (ESI[†]). The fungal enzyme 14α -demethylase (PDB ID: 1EA1) was obtained from the RCSB protein data bank (<https://www.rcsb.org>) in PDB format. CHEMSKETCH was used to draw 3D-structures of the ligands at www.acdlabs.com, and SPDBV software was used to perform their energy minimization. AutoDockTools-1.5.7 software was used to prepare the ligands and receptors in pdbqt format. During the preparation, water molecules were removed, Gasteiger charges were included, and only polar H-atoms were added. The results were visualized using a Discovery studio visualizer.⁶²

In docking simulations, different conformations of the hybrid molecules were obtained, and the energetically most favorable docked structure was studied. It was evident that the incorporation of a triazole unit into the alkyne **2** (with a docking score of -6.11 kcal mol⁻¹) showed improvement in the docking score (-8.38 to -9.28 kcal mol⁻¹) as apparent from 7-azaindole *N*-linked triazoles (**4a–4q**). For reference, we have considered the hybrid molecule **4d** and its alkyne **2** possessing binding energies of -8.81 and -6.11 kcal mol⁻¹, respectively. The alkyne **2** showed only one H-bond between the N-atom of the pyridine ring and THR260 (Fig. 5). The pi-electron density of the 7-AI ring system interacted with ALA400 and PRO320. Additionally, pi-Sigma interactions were observed with THR260. However, in the case of triazole **4d**, the N-atom present in the triazole ring is involved in three donor hydrogen bond interactions with CYS394, VAL395, and GLY396, denoted by a green-colored dotted line. The pi-orbitals of the pyridine ring formed pi-alkyl interactions with LEU105, LEU152, and ALA256. The chlorine-linked phenyl ring has interacted with LEU100 and ALA256 through pi-alkyl and Sigma alkyl bonds, respectively. The pi-orbitals of the triazole ring system also formed pi-alkyl interactions with CYS394, VAL395, and ALA256. The chlorine atom, present on the benzene ring, is involved in hydrophobic interactions with LEU100 and MET99. The pyrrole ring system is also involved in pi-alkyl-type interactions with LEU100. However, pi-Sigma interactions are formed between the pi-orbital of the pyrrole ring and LEU105 and ALA256. All triazoles exhibited various interactions; however, the dominant interactions for the binding process are conventional hydrogen bonding with THR260, CYS394, SER261, GLY72, ARG96, ARG326, VAL395, HIS259, HIS392, and GLN72, as presented in Fig. SI-3, ESI[†] and listed in Table SI-2, ESI[†]. The smaller value of bond length and the more significant negative value of the docking score are the leading indicators of the stability of hybrid molecules. Hence, theoretical data obtained through molecular docking seemed to agree with the experimentally obtained data, *i.e.*, biological activity testing. For reference, the cartoon images of dipolarophile **2** and the more

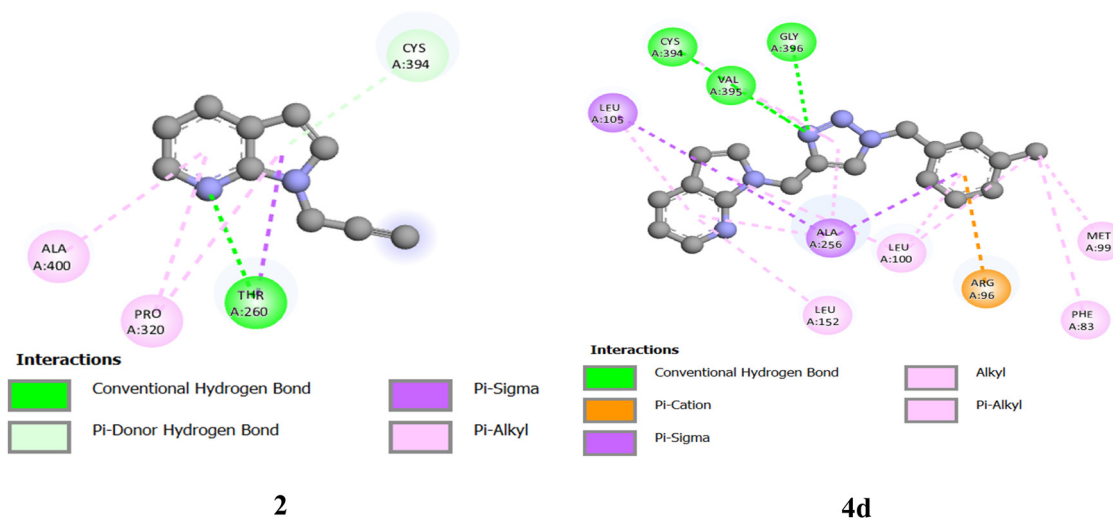


Fig. 5 2D images of alkyne **2** and 1,2,3-triazole hybrid **4d** showing conventional H-bonding and other interactions with an active site of DNA 14α -demethylase lanosterol fungal enzyme.

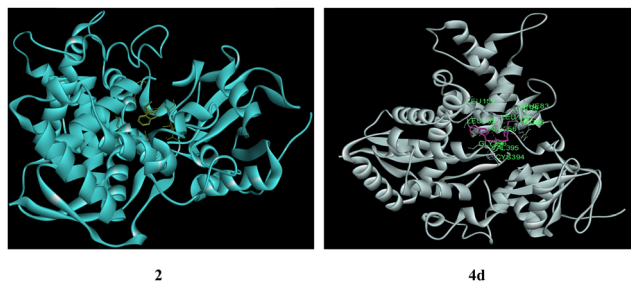


Fig. 6 Cartoon images of **2** and **4d** docked in the active site of DNA gyrase lanosterol 14- α demethylase enzyme (PDBID:1EA1).

promising potent 1,2,3-triazole **4d** docked in the active site of DNA gyrase lanosterol 14- α demethylase enzyme (PDBID:1EA1) are shown in Fig. 6.

ADME prediction

The *in silico* ADME (Absorption, Distribution, Metabolism, and Excretion) properties of all the newly synthesized 7-azaindole linked 1,4 disubstituted 1,2,3 triazole hybrid molecules (**4a–q**) were predicted by using the Molinspiration online software (<https://www.molinspiration.com/cgi-bin/properties>) as shown in Table SI-3 (ESI[†]). Here, we have studied various parameters such as %Absorbance (%ABS), lipophilicity (miLogP), Total Polar Surface Area (TPSA), Molecular Weight (MW), number of H-bond acceptors, number of H-bond donors, *etc.* In addition to them, the number of violations to Lipinski's *rule of five* (N_{viol})⁶³ and the drug-likeness score were also ascertained, which are helpful for the selection of a particular bio-active compound from the library of synthesized compounds.^{64–66} The drug-likeness property also helps to study various pharmacokinetic properties such as absorption, distribution, metabolism, and excretion of the synthesized drug/compound in human beings. All the synthesized compounds **2** and **4a–q** exhibited a TPSA (Total Polar Surface Area) value of $<160 \text{ \AA}^2$ (*i.e.*, 18–95 \AA^2) and also the absorption value in the favorable range, *i.e.* %ABS = 76.44 to 102.84, which was calculated using the mathematical expression %ABS = $109 - (0.345 \times \text{TPSA})$.⁶⁷ It is observed from the data that an orally active drug should follow at least four or more points in Lipinski's *rule of five*.^{68,69}

According to Lipinski's *rule of five*, an orally active drug should satisfy these points:

- (1) Molecular weight should be ≤ 500
- (2) Octane–water partition coefficient ≤ 5
- (3) No. of H-bond acceptors ≤ 10 and
- (4) No. of H-bond donors ≤ 5 .

The synthesized alkyne **2** and triazole compounds (**4a–4q**) strictly adhere to the Lipinski's rule with zero violation as is observed from Table SI-3 (ESI[†]), wherein the molecular weight of all the compounds lies in the range 156.17–368.24, *i.e.*, less than 500, which showed their easy absorption, diffusion, and transportation from the human body. The miLogP value of all compounds is also less than 5 (*i.e.*, 1.49–3.48). The number of hydrogen bond acceptor atoms, *i.e.*, O and N atoms, lies in the

range 2 to 8, *i.e.*, less than 10. Additionally, the number of rotatable bonds is 1 to 5, *i.e.*, less than 10, indicating their easily binding flexibility. Hence, all these parameters confirmed effective drug-like properties.

In general, if the bioactivity score (G protein-coupled receptor (GPCR) ligand, a kinase inhibitor, ion channel modulator, nuclear receptor ligand, protease inhibitor, and enzyme inhibitor) of the synthesized compounds is > -0.5 , then the drug is biologically active, but if the score is < -0.5 , then the drug is not active. The bioactivity scores, as provided in Table SI-4 (ESI[†]), showed that alkyne **2** and triazole compounds **4a–4q** are active and confirmed their binding flexibilities.

DFT studies

The Density Functional Theory (DFT) method acts as a potent tool for the interpretation of reactivity parameters of the synthesized drug compounds,⁷⁰ invaluable information, complementary to the experimental data, about molecular systems and processes. Thus, to confirm the stability and the geometry optimization of the proposed 7-azaindole linked 1,2,3-triazole hybrid molecule, DFT calculations were carried out in the gas phase at the B3LYP 6-311G(d,p) level by using the Gaussian09 program package.⁷¹

According to the Frontier Molecular Orbital (FMO) theory, the higher energy orbital HOMO (Highest Occupied Molecular Orbital) and the lower energy orbital LUMO (Lowest Unoccupied Molecular Orbital) participate in the bond formation between the precursors (alkyne and azides). The energy gap between these orbitals contributes to the bioactivity of the synthesized hybrid molecules (**4a–4q**). With the help of this method, various energy parameters such as energy gap, chemical potential, and chemical hardness and chemical softness as represented by symbols $E_{\text{LUMO-HOMO}}$, μ , and η , respectively, were calculated using Koopman's theorem.⁷² Additionally, the electrophilicity index represented by ω was calculated using Robert G. Parr approximation.^{73–75}

The formulation is represented below to show the relationships among these parameters as–

$$\begin{aligned} \bullet \mu &= (E_{\text{HOMO}} + E_{\text{LUMO}})/2 \\ \bullet \eta &= (E_{\text{LUMO}} - E_{\text{HOMO}})/2 \\ \bullet \omega &= \mu^2/2\eta \end{aligned}$$

It is well known that the higher the energy band gap, the harder the compound, and correspondingly, the lower the band gap, the softer, towards the chemical reactivity, and less stable the compound.^{76–78} Hence, from our calculation data table, as shown in Table SI-5 (ESI[†]), it was observed that the incorporation of the triazole unit into the alkyne considerably decreases the bandgap energy ($\Delta E_{\text{LUMO-HOMO}}$) of the resultant triazoles **4a–4q** (3.35–5.44 eV) except for compound **4f** (5.44 eV) as compared to the initial alkyne (5.11 eV). Also, compounds **4g** and **4p** having $-\text{NO}_2$ as an EWG on the benzyl and phenyl rings showed the least energy band gap, *i.e.*, 3.35 eV. This indicates that nitro as a functional group decreased the band gap and favored the easy movement of electrons. Thus, it leads to the softness and more reactivity of the compound. The electrophilicity index (ω) indicates the stabilization energy of the compound.

Here, the computed data revealed that drugs **4g** and **4p** showed high electrophilicity index values of 5.97 eV and 6.27 eV, respectively, thus, supporting better results in correlation with the experimental results.

Conclusions

Two new libraries of fluorescence-active 7-azaindole *N*-linked benzyl 1,2,3-triazole (**4a–4i**) and 7-azaindole *N*-linked phenyl 1,2,3-triazole (**4j–4q**) hybrids have been designed and synthesized *via* a Cu(I)-catalyzed alkyne-azide cycloaddition (CuAAC) click reaction and successfully characterized using FT-IR, ¹H-NMR, ¹³C-NMR, and HRMS data. The structure of compound **4f** was finally supported by its single-crystal X-ray data. Compounds **4d** and **4n** showed better antifungal activity against *C. albicans* and *A. niger* (MIC: 0.0193, 0.0386, 0.0404, and 0.0202 μmol mL⁻¹, respectively) with reference to the existing drug fluconazole (MIC: 0.0408 μmol mL⁻¹). Compound **4f** was found to be a suitable alternative to the existing antifungal drug fluconazole for *A. niger* with the least MIC: 0.0170 μmol mL⁻¹ along with considerable antibacterial activity against *P. aeruginosa* and *E. coli* (MIC: 0.0170 μmol mL⁻¹, each) for the reference drug ciprofloxacin (MIC: 0.0094 μmol mL⁻¹). The most promising compounds **4d**, **4f**, and **4n** showed good binding energies of -8.81, -8.76, and -8.79 kcal mol⁻¹ with DNA 14α-demethylase lanosterol enzyme (PDB ID:1EA1) and showed good agreement with the experimental and theoretical results. The results of the studies on the DFT and ADME properties reasonably supported the experimental results.

Conflicts of interest

The authors declare no conflict of interest.

Acknowledgements

We acknowledge the Department of Chemistry, NIT Kurukshetra for the research facilities and financial assistance to KS. We also acknowledge the Central Instrumentation Laboratory GJUS&T, Hisar, Haryana 125001 for NMR analysis and IIT Ropar for ESI-MS facilities.

References

- L. Trovato, M. Calvo, G. Migliorisi, M. Astuto, F. Oliveri and S. Oliveri, *Respir. Med. Case Rep.*, 2021, **32**, 101367.
- S. Narayanan, J. V. Chua and J. W. Baddley, *Clin. Infect. Dis.*, 2022, **74**, 1279.
- P. Koehler, O. A. Cornely, B. W. Böttiger, F. Dusse, D. A. Eichenauer and F. Fuchs, *Mycoses*, 2020, **63**, 528.
- N. T. Raveendran, A. Mohandas, R. R. Menon, A. S. Menon, R. Biswas and R. Jayakumar, *ACS Appl. Bio Mater.*, 2019, **2**, 243.
- N. Nehra, R. K. Tittal, V. D. Ghule, N. Naveen and K. Lal, *J. Mol. Struct.*, 2021, **1245**, 131013.
- N. Nehra, R. K. Tittal and V. D. Ghule, *ACS Omega*, 2021, **6**, 27089.
- A. V. Smirnov, D. S. English, R. L. Rich, J. Lane, L. Teyton, A. W. Schwabacher, S. Luo, R. W. Thornburg and J. W. Petrich, *J. Phys. Chem. B*, 1997, **101**, 2758.
- M. Négrerie, F. Gai, S. M. Bellefeuille and J. W. Petrich, *J. Phys. Chem.*, 1991, **95**, 8663.
- R. Gelabert, M. Moreno and J. M. Lluch, *J. Phys. Chem. A*, 2006, **110**, 1145.
- O. H. Kwon and D. J. Jang, *J. Phys. Chem. B*, 2005, **109**, 8049.
- K. Harkala, L. Eppakayala, S. Sripelly and T. C. Maringanti, *Indian J. Heterocycl. Chem.*, 2015, **25**, 165.
- M. P. Clark, M. W. Ledebor, I. Davies, R. A. Byrn, S. M. Jones, E. Perola, A. Tsai, M. Jacobs, K. Nti-Addae, U. K. Bandarage, M. J. Boyd, R. S. Bethiel, J. J. Court, H. Deng, J. P. Duffy, W. A. Dorsch, L. J. Farmer, H. Gao, W. Gu, K. Jackson, D. H. Jacobs, J. M. Kennedy, B. Ledford, J. Liang, F. Maltais, M. Murcko, T. Wang, M. W. Wannamaker, H. B. Bennett, J. R. Leeman, C. McNeil, W. P. Taylor, C. Memmott, M. Jiang, R. Rijnbrand, C. Bral, U. Germann, A. Nezami, Y. Zhang, F. G. Salituro, Y. L. Bennani and P. S. Charifson, *J. Med. Chem.*, 2014, **57**, 6668.
- M. J. Boyd, U. K. Bandarage, H. Bennett, R. R. Byrn, I. Davies, W. Gu, M. Jacobs, M. W. Ledebor, B. Ledford, J. R. Leeman, E. Perola, T. Wang, Y. Bennani, M. P. Clark and P. S. Charifson, *Bioorg. Med. Chem. Lett.*, 2015, **25**, 1990.
- R. A. Byrn, S. M. Jones, H. B. Bennett, C. Bral, M. P. Clark, M. D. Jacobs, A. D. Kwong, M. W. Ledebor, J. R. Leeman, C. F. McNeil, M. A. Murcko, A. Nezami, E. Perola, R. Rijnbrand, K. Saxena, A. W. Tsai, Y. Zhou and P. S. Charifson, *Antimicrob. Agents Chemother.*, 2015, **59**, 1569.
- J. Liang, J. E. Cochran, W. A. Dorsch, L. Davies and M. P. Clark, *Org. Process Res. Dev.*, 2016, **20**, 965.
- R. I. Benhamou, M. Bibi, K. B. Steinbuch, H. Engel, M. Levin, Y. Roichman, J. Berman and M. F. Orcid, *ACS Chem. Biol.*, 2017, **12**, 1769.
- K. Bozorov, J. Zhao and H. A. Aisa, *Bioorg. Med. Chem.*, 2019, **27**, 3511.
- B. Schulze and U. S. Schubert, *Chem. Soc. Rev.*, 2014, **43**, 2522.
- D. Ashok, P. Chiranjeevi, A. V. Kumar, M. Sarasija, V. S. Krishna, D. Sriram and S. Balasubramanian, *RSC Adv.*, 2018, **8**, 16997.
- W. S. Horne, M. K. Yadav, C. D. Stout and M. R. Ghadiri, *J. Am. Chem. Soc.*, 2004, **126**, 15366.
- P. D. Jarowski, Y. L. Wu, W. B. Schweizer and F. Diederich, *Org. Lett.*, 2008, **10**, 3347.
- D. Dheer, V. Singh and R. Shankar, *Bioorg. Chem.*, 2017, **71**, 30.
- S. Shafia, M. M. Alama, N. Mulakayala, C. Mulakayala, G. Vanaja, A. M. Kalle, R. Pallud and M. S. Alam, *Eur. J. Med. Chem.*, 2012, **49**, 324.
- N. S. Vatmurge, B. G. Hazra, V. S. Pore, F. Shirazi, M. V. Deshpande, S. Kadreppa and S. Chattopadhyay, *Org. Biomol. Chem.*, 2008, **6**, 3823.
- N. Nehra, R. K. Tittal, V. D. Ghule, N. Kumar, A. K. Paul, K. Lal and A. Kumar, *ChemistrySelect*, 2021, **6**, 685.

- 26 N. Naveen, R. K. Tittal, V. D. Ghule, P. Rani, K. Lal and A. Kumar, *ChemistrySelect*, 2020, **5**, 6723.
- 27 K. Lal, N. Poonia, P. Rani, A. Kumar and A. Kumar, *J. Mol. Struct.*, 2020, **1215**, 128234.
- 28 P. Yadav, K. Lal, A. Kumar, S. Bhushan, S. K. Guru and S. Jaglan, *Eur. J. Med. Chem.*, 2017, **126**, 944.
- 29 V. D. Da Silva, B. M. de Faria, E. Colombo, L. Ascari, G. P. A. Freitas, L. S. Flores, Y. Cordeiro, L. Romao and C. D. Buarque, *Bioorg. Chem.*, 2019, **83**, 87.
- 30 Y. W. He, C. Z. Dong, J. Y. Zhao, L. L. Ma, Y. H. Li and H. A. Aisa, *Eur. J. Med. Chem.*, 2014, **76**, 245.
- 31 F. Silva, M. C. B. V. Souza, I. I. P. Frugulhetti, H. C. Castro, S. L. Souza, T. M. Souza, D. Q. Rodrigues, A. M. T. Souza, P. A. Abreu, F. Passamani, C. R. Rodrigues and V. F. Ferreira, *Eur. J. Med. Chem.*, 2009, **44**, 373.
- 32 M. Whiting, J. Muldoon, J. C. Lin, S. M. Silverman, W. Lindstorm, A. J. Olson, H. C. Kolb, M. G. Finn, K. B. Sharpless, J. H. Elder and V. V. Fokin, *Angew. Chem., Int. Ed.*, 2006, **45**, 1435.
- 33 F. Jabeen, S. A. Shehzadi, M. Q. Fatmi, S. Shaheen, L. Iqbal, N. Afza, S. S. Pandad and F. L. Ansari, *Bioorg. Med. Chem. Lett.*, 2015, **26**, 1029.
- 34 K. K. Angajala, S. Vianala, R. Macha, M. Raghavender, M. K. Thupurani and P. J. Pathi, *SpringerPlus*, 2016, **5**, 423.
- 35 R. Raj, P. Singh, P. Singh, J. Gut, R. J. Rosenthal and V. Kumar, *Eur. J. Med. Chem.*, 2013, **62**, 590.
- 36 D. R. Buckle, D. J. Outred, C. J. M. Rockell, H. Smith and B. A. Spicer, *J. Med. Chem.*, 1983, **26**, 251.
- 37 M. A. Bonache, S. M. Fernandez, M. Miguel, B. Sabater-Munoz and R. Gonzalez-Muniz, *ACS Comb. Sci.*, 2018, **20**, 694.
- 38 J. Brahmi, S. Bakari, S. Nasri, H. Nasri, A. Kadri and K. Aouadi, *Mol. Biol. Rep.*, 2019, **46**, 679.
- 39 R. Huisgen, *Angew. Chem., Int. Ed. Engl.*, 1963, **2**, 565.
- 40 V. V. Rostovtsev, L. G. Green, V. V. Fokin and K. B. Sharpless, *Angew. Chem., Int. Ed.*, 2002, **41**, 2596.
- 41 T. R. Chan, R. Hilgraf, K. B. Sharpless and V. V. Fokin, *Org. Lett.*, 2004, **6**, 2853.
- 42 P. Wu, A. K. Feldman, A. K. Nugent, C. J. Hawker, A. Scheel, B. Voit, J. Pyun, J. M. Frchet, K. B. Sharpless and V. V. Fokin, *Angew. Chem., Int. Ed.*, 2004, **43**, 3928.
- 43 M. Twiting, J. C. Tripp, Y. C. Lin, W. Lindstrom, A. J. Olson, J. H. Elder, K. B. Sharpless and V. V. Fokin, *J. Med. Chem.*, 2006, **49**, 7697.
- 44 E. J. Yoo, M. Ahlquist, S. H. Kim, I. Bae, V. V. Fokin, K. B. Sharpless and S. Chang, *Angew. Chem., Int. Ed.*, 2007, **46**, 1730.
- 45 W. Christian, T. C. Christensen and M. Meldal, *J. Org. Chem.*, 2002, **67**, 3057.
- 46 N. J. Agard, J. A. Prescher and C. R. Bertozzi, *J. Am. Chem. Soc.*, 2004, **126**, 15046.
- 47 J. C. Jewett, E. M. Sletten and C. R. Bertozzi, *J. Am. Chem. Soc.*, 2010, **132**, 3688.
- 48 J. C. Jewett and C. R. Bertozzi, *Chem. Soc. Rev.*, 2010, **39**, 1272.
- 49 S. Deswal, N. Naveen, R. K. Tittal, V. D. Ghule, K. Lal and A. Kumar, *J. Mol. Struct.*, 2020, **1209**, 127982.
- 50 N. Naveen, R. K. Tittal, V. D. Ghule, N. Kumar, L. Kumar, K. Lal and A. Kumar, *J. Mol. Struct.*, 2020, **1209**, 127951.
- 51 N. Naveen, R. K. Tittal, V. D. Ghule, P. Yadav, K. Lal and A. Kumar, *Steroids*, 2020, **161**, 108675.
- 52 P. K. Naikawadi, L. Mucherla, R. Dandela, M. Sambari and K. S. Kumar, *Adv. Synth. Catal.*, 2021, **363**, 3796.
- 53 L. Jing, Wu, X. Hao, F. A. Olotu, D. Kang, C. H. Chen, K. H. Lee, M. E. S. Solima, X. Liu, Y. Song and P. Zhan, *Eur. J. Med. Chem.*, 2019, **183**, 111696.
- 54 M. I. Mangione, R. A. Spanevello and M. B. Anzardi, *RSC Adv.*, 2017, **7**, 47681.
- 55 N. Naveen, R. K. Tittal, P. Yadav, K. Lal and A. Kumar, *New J. Chem.*, 2019, **43**, 8052.
- 56 S. Deswal, R. K. Tittal, P. Yadav, K. Lal, V. D. Ghule and N. Kumar, *ChemistrySelect*, 2019, **4**, 759–764.
- 57 C. X. Zhao, T. Liua, M. Xua, H. Lina and C. Zhang, *J. Chin Chem Lett.*, 2021, **32**, 1925.
- 58 N. Ö. Can, U. Acar Çevik, B. N. Sağlık, S. Levent, B. Korkut, Y. Özkay, Z. A. Kaplancıklı and A. S. Kopalal, *J. Chem.*, 2017, 9387102.
- 59 L. M. Podust, T. L. Poulos and M. R. Waterman, *Proc. Natl. Acad. Sci. U. S. A.*, 2001, **98**(6), 3068.
- 60 J. Eberhardt, D. S. Martins, A. F. Tillack and S. Forli, *J. Chem. Inf. Model.*, 2021, **61**(8), 3891.
- 61 O. Trott and A. J. Olson, *J. Comp. Chem.*, 2010, **31**(2), 455.
- 62 Dassault System BIOVIA, *Discovery Studio Visualizer v17.2.0.16349*, San Diego, 2016.
- 63 C. A. Lipinski, F. Lombardo, B. W. Dominy and P. J. Feeney, *Adv. Drug Delivery Rev.*, 2001, **46**(1), 3.
- 64 B. Shah, P. Modi and S. R. Sagar, *Life Sci.*, 2020, **252**, 117652.
- 65 T. Kampmann, R. Yennamalli, P. Campbell, M. J. Stoermer, D. P. Fairlie, B. Kobe and P. R. Young, *Antiviral Res.*, 2009, **84**(3), 234.
- 66 F. Shaikh, Y. Zhao, L. Alvarez, M. Iliopoulou, C. Lohans, C. J. Schofield, S. Padilla-Parra, S. W. I. Siu, E. E. Fry, J. Ren and D. I. Stuart, *J. Med. Chem.*, 2019, **62**(6), 2928.
- 67 MolinspirationChemoinformaticsBratislava, Slovak Republic, available from: <http://www.Molinspiration.com/cgibin/properties>, 2014.
- 68 A. Kumar, K. Lal, L. Kumar, A. Kumar, N. Naveen and R. K. Tittal, *Res. Chem. Intermed.*, 2022, **48**, 5089.
- 69 V. Kumar, K. Lal, A. Kumar, R. K. Tittal, M. B. Singh and P. Singh, *Res. Chem. Intermed.*, 2023, **49**, 917–937.
- 70 (a) L. H. Mendoza-Huizar and C. H. Rios-Reyes, *J. Mex. Chem. Soc.*, 2011, **55**, 142; (b) I. Piyanzina, B. Minisini, D. Tayurskii and J. F. Bardeau, *J. Mol. Model.*, 2015, **21**, 34; (c) M. Elango, R. Parthasarathi, V. Subramanian, U. Sarkar and P. K. Chattaraj, *J. Mol. Struct.*, 2005, **723**, 43.
- 71 M. J. Frisch, G. W. Trucks, H. B. Schlegel, G. E. Scuseria, M. A. Robb, J. R. Cheeseman, G. Scalmani, V. Barone, B. Mennucci, G. A. Petersson, H. Nakatsuji, M. Caricato, X. Li, H. P. Hratchian, A. F. Izmaylov, J. Bloino, G. Zheng, J. L. Sonnenberg, M. Hada, M. Ehara, K. Toyota, R. Fukuda, J. Hasegawa, M. Ishida, T. Nakajima, Y. Honda, O. Kitao, H. Nakai, T. Vreven, J. A. Montgomery Jr., J. E. Peralta,

- F. Ogliaro, M. Bearpark, J. J. Heyd, E. Brothers, K. N. Kudin, V. N. Staroverov, R. Kobayashi, J. Normand, K. Raghavachari, A. Rendell, J. C. Burant, S. S. Iyengar, J. Tomasi, M. Cossi, N. Rega, J. M. Millam, M. Klene, J. E. Knox, J. B. Cross, V. Bakken, C. Adamo, J. Jaramillo, R. Gomperts, R. E. Stratmann, O. Yazyev, A. J. Austin, R. Cammi, C. Pomelli, J. W. Ochterski, R. L. Martin, K. Morokuma, V. G. Zakrzewski, G. A. Voth, P. Salvador, J. J. Dannenberg, S. Dapprich, A. D. Daniels, O. Farkas, J. B. Foresman, J. V. Ortiz, J. Cioslowski and D. J. Fox, *Gaussian 09, Revision B.01*, Gaussian Inc., Wallingford, 2010.
- 72 T. Koopmans, *Physica*, 1934, **1**(1), 104.
- 73 R. G. Parr, L. V. Szentpály and S. Liu, *J. Am. Chem. Soc.*, 1999, **121**(9), 1922.
- 74 C. Lee, W. Yang and R. G. Parr, *Phys. Rev. B: Condens. Matter Mater. Phys.*, 1988, **37**(2), 785.
- 75 A. D. Becke, *J. Chem. Phys.*, 1993, **98**(7), 5648.
- 76 Richa, N. Kushwaha, S. Negi, A. Kumar, E. Zangrando, R. Kataria and V. Sain, *Dalton Trans.*, 2021, **50**, 13699.
- 77 Richa, S. Kumar, J. Sindhu, P. Choudhary, S. Jaglan, E. Zangrando, R. Kumar, S. C. Sahoo, V. Kumar, S. K. Mehta and R. Kataria, *J. Mol. Str.*, 2021, **1228**, 129460.
- 78 S. Kumar, V. Saini, I. K. Maurya, J. Sindhu, M. Kumari, R. Kataria and V. Kumar, *PLoS ONE*, 2018, **13**(4), 0196016.