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Recognition, detection and host—guest chemistry of hydrogen peroxide in a fluorescent metal—organic framework with chiral helical channels†

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Combining supramolecular chemistry and bioscience, metal–organic frameworks (MOFs) built with biomolecules as ligands possess highly ordered structures and unique pores with accessible metal binding sites and rich interactions to drive molecular recognition processes. In this work, a novel purine-based MOF with chiral helical channels has been successfully constructed, named HSTC-1, which identifies H_2O_2 with a fluorescence quenching effect. Notably, the successful encapsulation of H_2O_2 could be observed by single-crystal X-ray diffraction. Metadynamics and density functional theory calculations reveal that the adsorption of H_2O_2 molecules through the chiral channels induces chiral distribution of subsequent guests to form a host–guest recognition system of H_2O_2 @HSTC-1, which slows down the oxidation of the host framework. This work provides unique insights for the construction of intelligent protection matrix materials and is of great significance in the fields of pharmacochemistry and biomedicine.

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Introduction

Hydrogen peroxide (H_2O_2) is the most symbolic reactive oxygen species and plays an important physiological role in medicine and the human body.^{1,2} However, H_2O_2 is referred to as a 'Jekyll and Hyde' molecule in the scientific literature.³ Excessive amounts of H_2O_2 in the body will attack cells or biomolecules, including protein structures, liposomes, and DNA, leading to oxidative stress and toxic and lethal chain reactions, and is even associated with severe diseases.^{4,5} But a major issue for the application of H_2O_2 probes in biological cells is the lack of reaction retention due to the highly oxidizing nature of H_2O_2 , which triggers a reaction once it comes into

Metal-organic frameworks (MOFs) have been emerging as one of the most promising materials due to their tunable pore sizes, diverse structures, and exposed metal sites, 9-12 which capture molecules by means of solid-liquid phases without changing their activity and functional groups, providing advantageous conditions for studying their interactions at the atomic level in host-guest chemistry. The utilization of biomolecular linkers¹³⁻¹⁷ can enhance the recognition capability of MOFs, and the diversity in the size and shape of channels within MOFs endows them with unique properties. 18-23 Typical examples are adenine-based ZnBTCA, which adaptively recognizes thymidine using sinusoid-like channels to achieve A-T base pairing, 18 and a hierarchical pore Zn-MOF based on 6-benzylaminopurine, which exclusively recognizes acetone through multiple hydrogen bond interactions between hosts and guests.23

Purine, an alkaloid in the body, can produce a variety of tautomeric forms influenced by electron transfer.²⁴ Its rigid

contact with reactants. The time from the addition of H_2O_2 to the actual experiment usually spans hours, resulting in a constantly changing "background" signal, and it is difficult to guarantee and demonstrate uniform probe distribution in samples, which is also one of the reasons why the identification and quantitative detection of H_2O_2 in cells remains challenging.^{6–8}

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structure facilitates the formation of robust MOFs, with the four N atoms of purines having more coordination modes and supramolecular interactions, such as hydrogen bonding. The unique non-covalent interactions between metal ions and purine make their MOFs highly dynamic²⁵ and provide a prerequisite for the formation of special shaped channels. Herein, a novel three-dimensional chiral MOF, named HSTC-1, was successfully constructed using the purine biomolecule. Under mild conditions, HSTC-1 could capture and identify H2O2 using chiral helical channels and successfully decelerate oxidation by strong oxidants. This provides alternative technical support for the identification and detection of H₂O₂.

Results and discussion

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Single-crystal X-ray diffraction (SCXRD) analyses revealed that HSTC-1 crystallizes in an orthorhombic crystal system with a chiral space group of $P2_12_12_1$ (Table S1†). The asymmetric unit contains one crystallographically independent Zn center with tetrahedral coordination geometry. Zn2+ binds to N of three different purine ligands and an acetate oxygen atom, forming a mononuclear Zn(purine)3(acetate) secondary building unit (SBU). The polymer chain consists of Zn²⁺ and purine arranged along the 21 screw axis in the form of a helix. These 1D helical chains are further connected at the Zn²⁺ centers, giving rise to a three-dimensional framework with one-dimensional helical chiral channels (Fig. 1a-c). It contains an irregular rectangular channel (ca. 6.52 Å \times 7.56 Å) running along the [100] direction with a porosity of 27.2% per unit cell volume calculated after

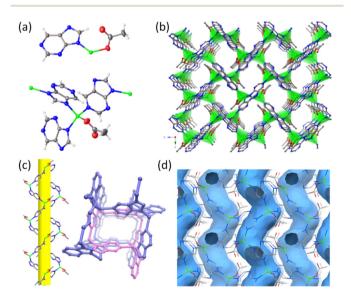
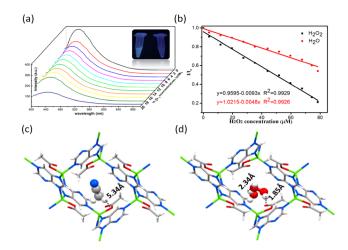


Fig. 1 (a) The asymmetric unit and the coordination environment of HSTC-1. (b) The 3D structure with 1D channels viewed along the a-axis. (c) Representation of the 2_1 screw axis along the a-axis of HSTC-1, shown as a yellow cylinder, and the perspective view of the 1D helical chiral channel shown in two different colours, magenta and blue. (d) The chiral helical pores along the b-axis are connected with the a-axis pore with the contact surface, probe radius: 1.2 Å. Colour codes: C, grey; N, blue; O, red; Zn, green (tetrahedra); H, white or omitted.

the removal of the guest molecule acetonitrile. The sinusoidlike chiral helical pores along the b-axis connect with the a-axis channel (Fig. 1d), while uncoordinated oxycarboxylate oriented toward the channel provides a potential active site for molecular recognition. The spiral chirality of the framework was further determined by means of solid-state circular dichroism (CD) measurements. Bulk samples tend to be uniform mixtures of crystals with opposing chirality, and thus have no Cotton-effect signals, whereas individual crystals with homochirality can be formed by spontaneous resolution. CD spectra exhibit positive and negative Cotton effects at the same positions, confirming that they are enantiotropic (Fig. S10†). A topological description of HSTC-1 can be achieved by denoting the SBU as a three-connected (3-c) node and purine as a (3-c) node, respectively. Then, it is simplified as a uninodal 3-c net with the point symbol $\{10^3\}$, which has been registered as an srs topological type in the Topos Topological Database (TTD) collection (Fig. S8 and topological analysis in the ESI†).

N₂ (77 K) and CO₂ (273 K) adsorption experiments were performed on HSTC-1 to assess its permanent porosity. HSTC-1 shows normal adsorption capacity and the mode pore-size is 6.86 Å; this accounted for the dominant distribution, which is consistent with the pore size predicted by the single-crystal structure (Fig. S1†). After soaking the synthesized samples in different organic solvents for 24 h, their powder X-ray diffraction (PXRD) patterns still match well with the simulated PXRD patterns from single-crystal data indicating their structural integrity and good chemical stability (Fig. Thermogravimetric analysis (TGA) of HSTC-1 shows that the weight loss of 15.3% below 300 °C is due to the loss of acetonitrile guest molecules, while the host framework starts to collapse above 500 °C, manifesting the excellent thermostability of HSTC-1 (Fig. S3a†).

Solid HSTC-1 emitted blue light under ultraviolet light irradiation at room temperature. The broad emission band of **HSTC-1** is located at around 455 nm ($\lambda_{\text{max}} = 380 \text{ nm}$) (Fig. S5†). Interestingly, after the addition of 20 µL of 30% H₂O₂, the luminescence of HSTC-1 showed a quenching effect, which prompted us to explore this phenomenon further (Fig. 2a, S6†). To quantitatively describe the fluorescence signal response of HSTC-1 to H₂O₂, concentration-dependent fluorescence titration experiments were performed. As shown in Fig. 2a, the fluorescence intensity of the HSTC-1 acetonitrile suspension decreased distinctively with the addition of H₂O₂. Meanwhile, a linear function correlation between the emission intensity of HSTC-1 and H2O2 concentration was observed, with the Stern-Volmer equation $I_0/I = 0.9983 + 0.0065[M]$ ($R^2 = 0.0065[M]$ 0.9718) in the range of 0–80 μ M (I_0 = luminescence intensity of pristine HSTC-1, I = luminescence intensity of HSTC-1 afterthe addition of H2O2, and [M] is the molar concentration of H₂O₂) (Fig. S7†). Therefore, the sensitivity value could reach 6540 L mol⁻¹, which is comparable to those values of previously reported^{2,7,8} materials with moderate sensitivity properties. By calculation using the $3\sigma/k$ formula (σ is the standard deviation of 5 replicating luminescence measurements of blank solutions, and k is the slope of the calibration curve),



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Fig. 2 (a) Emission spectra of HSTC-1 as a suspension in acetonitrile excited at 380 nm upon the incremental addition of H₂O₂. The inset is a photograph of vials under 365 nm UV light. (b) Corresponding I/I₀ plot after the addition of H_2O_2 and H_2O in the range of 0-80 μM . (c) Pore structures of CH₃CN@HSTC-1 and (d) H₂O₂@HSTC-1. Colour codes: C, grey; N, blue; O, red; Zn, green; H, white.

the detection limit for H₂O₂ was found to be 10.05 ppm. Water is also well-known as a strong fluorescence quencher. Therefore, we compared the fluorescence titration experiment with H2O under the same conditions and obtained the equation as $I/I_0 = 1.0215 - 0.0048x$ ($R^2 = 0.9926$). From Fig. 2b, it can be found that the decrease of fluorescence intensity (I/I_0) on dropping H_2O is smaller than that on dropping H_2O_2 , indicating that H2O2 plays a role in fluorescence quenching with the synergistic effect of H₂O. The absorption spectra of HSTC-1 acetonitrile solution feature two absorption peaks at 215 nm and 280 nm. Upon H₂O and H₂O₂ being added into the solution, absorbance increases and blue shifts occur at 280 nm. With the addition of H₂O₂, the increase of the relative absorbance ratio of the two peaks (A_{215}/A_{280}) is higher than that with the addition of H₂O (Fig. S11†). Thus, competitive absorption of excitation light by H2O2 may reduce the luminescence intensity of HSTC-1.26

To further clarify the luminescence quenching mechanism, the samples were soaked in 3% H₂O₂ for 2 h and then filtered out for PXRD measurements. No obvious difference in the PXRD pattern was observed compared with the simulated one (Fig. S2c†). Interestingly, it can be seen from Fig. S2c† that the positions of the PXRD peaks of HSTC-1 changed significantly after being immersed in H2O2 for more than 1 day (named HSTC-1a). SCXRD analyses revealed that Zn²⁺ coordinates with two imidazolate N atoms of two different purines and two bridging O atoms to form a two-dimensional non-porous framework (Fig. S12†). To ascertain the process of the transformation of HSTC-1, we performed in situ PXRD experiments by dropping H₂O₂ (or H₂O) on ground HSTC-1 powder and monitoring results in real-time. The results showed that the PXRD patterns did not change in 48 hours by dropping either H₂O₂ or H_2O (Fig. S2c-f†). We concluded that **HSTC-1** undergoes a

crystal transformation in recognition of H2O2, which may be attributed to the tautomerism of purine ligands. In solution, the gradual decomposition of H₂O₂ causes electron transfer, and H₂O is a strong ligand, which will cause purine tautomerization and thus change the coordination mode with Zn2+ (Fig. S12a†). However, from in situ solid-phase PXRD experiments, H₂O₂ guests are protected by the channels of HSTC-1 from decomposition, resulting in a lack of opportunity for electron transfer, and the structure is unaltered.

To gain further structural information, single-crystal to single-crystal (SC-SC) transformation was performed to in situ load H₂O₂ into the framework. Fortunately, the H₂O₂ molecule became locked in the chiral pore of HSTC-1 and could be verified by SCXRD (denoted as H₂O₂@HSTC-1). In addition, H₂O molecules in a 30% H₂O₂ solution are also adsorbed into the pore. The distances between the H of the terminal H₂O₂ and uncoordinated carboxylate O, the H of the terminal H2O and O of H₂O₂, and the H of the terminal H₂O₂ and the O atom of H_2O are 1.85 Å, 2.34 Å, and 1.84 Å, respectively, which can permit formation of effective hydrogen bonding (Fig. 2d). The formation of H-bond chains between H2O2 molecules and between H₂O₂ and H₂O molecules reduces the oxidation effect of H₂O₂ on the host framework. In contrast, the aprotic solvent acetonitrile in pristine HSTC-1 has no hydrogen bonding interaction over the long distances of the guest and host (4.8-6.3 Å) (Fig. 2c). The above experimental results indicate that the mechanism of HSTC-1 luminescence quenching in the presence of H₂O₂ may be ascribed to the synergistic effect of the transformation of the framework, competitive absorption and the host-guest and guest-guest interactions.²³

Inspired by a previously reported purine-based MOF, 21,27 we optimized the synthesis conditions and obtained another Zn (pur)(OAc) MOF, named HSTC-2. HSTC-2 crystallizes in a tetragonal crystal system with a space group of I4 and possesses a square one-dimensional channel running along the c-axis with opening dimensions of ca. 7.608 Å \times 7.608 Å, while its topology is of the lig type (Fig. S8 and ESI† topological analysis). HSTC-2 has the same asymmetric units and SBUs as HSTC-1, with the main difference being the shape and size of the channel. The side views highlighting the channel shape (tube in blue) and pore-metrics analyses 28,29 of HSTC-1 (Fig. 3a and c) and HSTC-2 (Fig. 3b and d) allow visualization of the spatial effect. Under the same conditions, the SC-SC transformation of HSTC-2 loaded with H2O2 showed that partially coordinated acetate was oxidized to peroxyacetate, and only H2O molecules were found in the square channel (Fig. 3e and f). Unfortunately, after soaking for more than 12 h, it was difficult to find good quality crystals for SCXRD data collection, and the frameworks had collapsed using PXRD after filtration. We therefore speculated that the shape and dimensions of the chiral channels play a key role in the recognition of chiral H_2O_2 .

We performed metadynamics (MTD) and density functional theory (DFT) calculations to compare the magnitude of energy generated by HSTC-1, HSTC-2 and H2O2 during the identification process. One to six H₂O₂ molecules were added into the

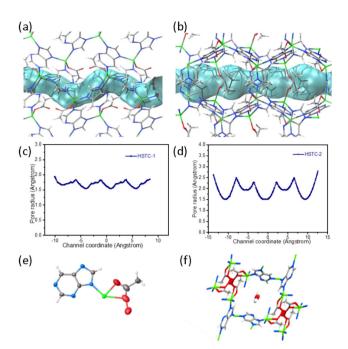


Fig. 3 (a) and (c) Side view of HSTC-1 highlighting the shape of helical channels (tube in blue) and the pore-metrics analysis (curve in navy), and (b) and (d) the side view of HSTC-2 highlighting the shape of square channels and the pore-metrics analysis. (e) Asymmetric unit of H₂O₂ oxidized-HSTC-2. (f) Crystal structure of oxidized-HSTC-2 contains only the H₂O guest molecule. Colour codes: C, grey; N, blue; O, red; Zn, green (tetrahedra); H, white.

channel to locate the binding site and find equilibrium structures (see the computation details, Experimental section). The results show that when the first H₂O₂ molecule was added, the average host-guest binding energy of HSTC-1 was about $-6.7 \pm$ 2.05 kcal mol^{-1} , while that of HSTC-2 was about -7.96 \pm $1.15 \text{ kcal mol}^{-1}$. With an increase of the number of H_2O_2 molecules, although an overall slightly larger host-guest binding energy was found in HSTC-1, the difference between two isomers was insignificant to distinguish the above experimental results (Fig. 4a and Table S2†). So, the effect of guestguest interaction is further considered by mapping H₂O₂ molecules in the channel in sequence. As the number of H₂O₂ molecules entering the channel increased, the average guestguest binding energy of HSTC-1 was about -12 kcal mol⁻¹, while that of HSTC-2 was about -5 to -6 kcal mol⁻¹. The difference in the energy of the guest-guest interaction between HSTC-1 and HSTC-2 became more significant (Fig. 4b and Table S3†). Meanwhile, the theoretical calculation results indicate that H₂O₂ molecules are distributed more dispersedly in each square channel of HSTC-2 and tend to only interact with the framework, resulting in weak guest-guest interactions. In this manner, H₂O₂ has a higher chance to oxidize HSTC-2 to form oxidized-HSTC-2 or undergo structural transformation. However, on account of the chiral pore of HSTC-1, the adsorption of the primary chiral H₂O₂ molecule through the chiral channel induced a series of guests to form a chiral distribution, slowing down the effect of oxidation tendency on the

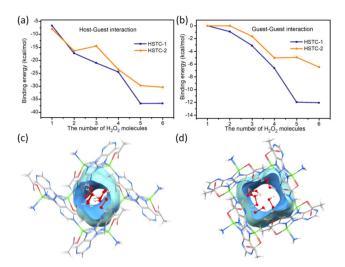


Fig. 4 Binding energies for host-guest interactions (a) and guestguest interactions (b) in HSTC-1 and HSTC-2; distribution of 6 H2O2 molecules in a chiral channel of HSTC-1 (c) and in a square channel of HSTC-2 (d)

host frameworks. This is consistent with behavior predicted from analysis of the single-crystal structure. The initial incoming H₂O₂ and the host framework exhibit a synergistic effect on the subsequent guest molecules to stack along the chiral rotation axis (Fig. 4c and d).

demonstrate the encapsulation of H₂O₂ in H₂O₂@HSTC-1, the crystals were rinsed with catalase three times (5 ml each) to remove H₂O₂ from the surface, and then treated with 3,3',5,5'-tetramethyl-benzidine (TMB), a commonly used chromogenic agent for H₂O₂ detection.^{30,31} The results showed that HSTC-1 and H2O2@HSTC-1 barely changed in 30 minutes after the addition of TMB ethanol solution (400 µL, 0.2 M). By adjusting the pH value with buffer, the colour gradually changed from colourless to blue, and an obvious absorption peak appeared at 652 nm, while for HSTC-1 alone it did not. This is due to the gradual release of H₂O₂ from H₂O₂@HSTC-1, oxidizing TMB to ox-TMB after the addition of buffer solution (Fig. 5a). From the time-dependent absorbance plots and the photographs of the TMB chromogenic reaction, it can be seen that oxidized-HSTC-2 without buffer rapidly changes from colourless to blue, and the absorbance increases continuously until it exceeds the detection range after 1400 s. This is caused by the certain oxidation from the peroxy bond of oxidized-HSTC-2. In contrast, the colour of H₂O₂@HSTC-1 remains unchanged in the absence of buffer solution (Fig. 5b). The above experiments indicate that the chiral channels of HSTC-1 can temporarily protect H₂O₂, providing a suitable reaction retention time for practical applications.

To quantify the energies of the host-guest interactions between H₂O₂ and HSTC-1, isothermal titration calorimetry (ITC) experiments were performed. 32,33 The thermograms obtained by titration of HSTC-1 suspensions with H₂O₂ showed negative peaks, implying the host-guest interaction

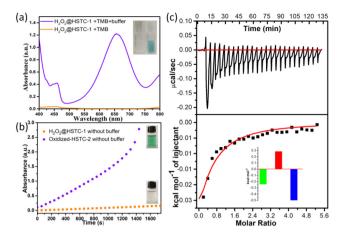


Fig. 5 (a) Comparison of the chromogenic reactions of H₂O₂@HSTC-1 and HSTC-1 with TMB. The inset is a photograph of their TMB chromogenic reaction, respectively. (b) The time-dependent absorbance plots of H₂O₂@HSTC-1 and oxidized-HSTC-2 after the addition of TMB solution without buffer. The inset is a photographic comparison of their TMB chromogenic reactions. (c) ITC thermogram of the titration of the HSTC-1 suspension (4 mM, 1.8 mL) with H₂O₂ (99 mM, 15 μL each injection) and fitting with a single-site model. The inset depicts the magnitude of the calculated thermodynamic parameters.

through a favorable exothermic pathway (Fig. 5c). Then, the Gibbs free energy change (ΔG) was calculated to be -528.8 cal mol^{-1} . The $-\Delta G$ and $+\Delta S$ values suggest that the spontaneous processes are mainly due to an enthalpic driving force with a compensating favorable entropic factor. On comparing the effects of H2O, the ITC experiment involving titration of HSTC-1 suspensions with H₂O was performed under the same conditions, and no significant thermal changes were observed in the thermogram (Fig. S9†). We speculated that the thermal changes in the system mainly came from the interaction between H₂O₂ and HSTC-1 frameworks.

Conclusions

In summary, utilizing purine biomolecules as ligands, we successfully synthesized fluorescent HSTC-1 with chiral pores, which spontaneously identify and capture H_2O_2 . The H_2O_2 and H₂O guests stack along the chiral rotation axis in chiral helical channels due to a synergistic effect between initial incoming H₂O₂ and the host framework, decreasing the oxidation of the framework. The resulting unique MOF, HSTC-1, can successfully use the chiral channel to slow down the attack of the strong oxidizing agent H2O2, thus exhibiting higher reaction retention, which provides a new perspective for the construction of intelligent protective matrix materials, and has potential applications in biopharmacology and cutting-edge medicine. Although there is still a long way to go until reliable optical H₂O₂ sensors can be used for quantitative and continuous measurements, we propose that in the future, bioinspired luminescent MOFs will become feasible platforms for simple,

rapid and accurate determination of H2O2 concentrations in biological systems.

Experimental

Materials and methods

All chemicals were purchased from commercial sources and used without further purification. The organic solutions were purchased from Xilong Science Co., Ltd and Guangdong Guanghua Technology Co., Ltd. Single-crystal X-ray diffraction (SCXRD) analysis was performed using a Rigaku XtaLAB PRO MM007-DW diffractometer system, an RA-Micro7HF-MR-DW (Cu/Mo) X-ray generator and a Pilatus 3R-200K-A detector (Cu $K\alpha$, $\lambda = 1.54178$ Å). PXRD patterns of the bulk samples were measured on a Rigaku MiniFlex600 X-ray diffractometer (Rigaku, Kyoto, Japan). ITC experiments were conducted on a Malvern MicroCal VP-ITC System (Malvern, Europe). Elemental analyses of C, H, and N were carried out on a PerkinElmer PE 2400II. pH was confirmed using a PHS-3C pH meter. Thermogravimetric analyses (TGA) were performed using a Shimadzu synchronous differential thermal-thermogravimetric analyzer (DTG-60) in the temperature range of room temperature to 800 °C under nitrogen flow (40 mL min⁻¹) at a typical heating rate of 10 °C min⁻¹. Infrared spectra (IR) were collected in KBr disks using a Shimadzu IRAffinity-1 in the range of 4000-400 cm⁻¹. Solid-state and solution luminescence spectra were measured on a PF-5301PC. UV-Vis spectra and results of ultraviolet titrating experiments were recorded on a TU-1950 ultraviolet and visible spectrophotometer (Persee, Beijing, China). Circular dichroism (CD) spectra were recorded on a Chirascan V100 (Applied Photophysics Ltd, UK) in the range of 200-500 nm (equipped with a PMT detector).

Synthesis of HSTC-1. A mixture of Zn(CH₃COO)₂·2H₂O (0.12 mmol), purine (0.04 mmol), and acetonitrile (5 mL) was sealed in a Pyrex glass tube with the pH adjusted using 50 µL of 28% NH₃ aqueous solution, and heated in an oven at 120 °C for 72 h. After cooling to room temperature at a rate of 5 °C h⁻¹, colourless crystals were obtained by filtration and washed with acetonitrile 3 times. Selected suitable crystals were immediately mounted on the single-crystal X-ray diffractometer for structural determination. IR spectrum (see Fig. S4a†): 3427(s), 3103(w), 1907(w), 1617(s), 1418(s), 1407(s), 1319(s), 1211(s), 1099(m), 1018(m), 958(s), 800(s), 646(s). Elemental analysis (CHN), C₇H₆N₄O₂Zn(H₂O)_{1.2}, calculated (%): C, 31.7076; H, 3.1931; N, 21.1298. Found (%): C, 31.19; H, 3.116; N, 21.47.

Synthesis of HSTC-2. A mixture of Zn(CH₃COO)₂·2H₂O (0.12 mmol), purine (0.04 mmol), and acetonitrile (5 ml) was sealed in a Pyrex glass tube with the pH adjusted using 400 μ L of 1 M HNO3, and heated in an oven at 120 °C for 72 h. After cooling to room temperature at a rate of 5 °C h⁻¹, yellowish crystals were obtained by filtration and washing with acetonitrile 3 times. Selected suitable crystals were immediately mounted on the single-crystal X-ray diffractometer for structural determination. IR spectrum (see Fig. S4b†). 3416(s),

3114(w), 1907(w), 1619(s), 1480(m), 1408(s), 1314(s), 1209(s), 1096(m), 953(s), 799(s), 657(s).

Fluorescence titration experiments

10 mg of the HSTC-1 solid sample was fully ground into powders and then transferred to a 10 mL volumetric flask. Acetonitrile was added and sonicated for 30 min to form a uniformly dispersed MOF suspension (1 mg mL⁻¹). The suspension was further diluted to the corresponding concentrations with acetonitrile for concentration-dependent fluorescence titration experiments. To 2.50 mL of the above suspensions, 10 μL of H₂O₂ solution (1 M, 0.1 M, 0.01 M, 1 mM, 0.1 mM) was incrementally added to monitor the fluorescence emission intensity (λ_{max} = 380 nm) until the endpoint.

TMB oxidation experiments

In a typical assay, 200 µL of TMB ethanol solution (0.2 M, 0.1 M), 1.5 mL of ethanol, 1.5 mL of 1 mg mL⁻¹ HSTC-1 (or H₂O₂(a)HSTC-1) suspension, and 200 μL of H₂O, were sequentially placed into a 5 mL vial with (or without) sodium acetate buffer solution (pH 5.0). The mixture (with a total volume of 3.4 mL) was stirred and then filtered through a syringe filter (PTFE, hydrophobic, 0.24 µm). The content of oxTMB in the filtrate was assessed through UV-Vis absorbance.

The steady-state kinetic assays were performed through the oxidation of TMB in the presence of H₂O₂ to produce blue products. The substrate solution was prepared by dissolving TMB in 2 mL of 30% ethanol solution. The formation of oxTMB was determined by UV-Vis spectrophotometry at 652 nm after adding 1 mg of MOFs (H2O2@HSTC-1 or oxidized-HSTC-2) to the substrate solution.

Isothermal titration calorimetry (ITC) experiments

In a typical titration, H₂O₂ solution was incrementally added to the ITC sample cell containing an aqueous suspension of HSTC-1. All titration experiments were performed in aqueous solution without adjusting pH under the following conditions: reference power (10–15 μcal s⁻¹), initial injection delay (300 s), stirring speed (850 rpm), feedback mode gain (high feedback), the spacing between injections (300 s), and filter period (10 s). The thermodynamic profile of each binding process was calculated by fitting the data with a single site interaction model.

Computation details

The computational models are extracted from the singlecrystal structures of HSTC-1 and HSTC-2, containing sixteen Zn²⁺ cations, sixteen adenine ligands, and sixteen acetate anions. All Zn2+ cations are retained in a tetrahedral fourcoordination mode by completing coordinated N atoms as NH₃; hydrogen atoms are added to adenine ligands to maintain the neutral framework. All hydrogen atoms are optimized with Gaussian 16 at the BLYP level of theory and 6-31G* basis sets. 34-37 To study the host-guest and/or guest-guest interactions, one to six hydrogen peroxide molecules are added into the channel to locate the binding site and find equilibrium structures by adopting Crest 2.11.2 software and

GFN-FF force field. 38,39 Metadynamics (MTD) algorithms are used to acquire all possible structures at 298.15 K within a 5 kcal mol⁻¹ window from the lowest-energy structure.⁴⁰ The five lowest energy structures of $(H_2O_2)_{1-6}$ (a) HSTC-1/2 are screened to be optimized by GFN2-xTB methods in order to obtain the defined binding sites with xtb 6.4.0 software, in which an ALPB water solvation model is implicitly used. 41,42 Binding energies are calculated with the ORCA 4.2.1 software package at the BLYP level of theory. 43,44 Def2-SVP basis sets, along with auxiliary Def2/J and Def2-SVP/C, are adopted for Zn, O, N, C, and H atoms. 45,46 Solvation is treated by the SMD water solvation model.⁴⁷ Grimme's D4 method and geometrical counterpoise (gCP) are considered for dispersion interaction corrections and to overcome the basis set superposition error (BSSE) when using small basis sets, respectively. 48,49

Author contributions

H. Cai conceived and wrote the manuscript and supervised the research; D. Li supervised the research and polished the manuscript; J.-W. Wu, X.-J. Cai, Y.-Y. Huang, Y.-H. Lu, J.-W. Cai, and W. Lu conducted the synthesis and fluorescence, UV-Vis, CD and TMB testing experiments; J.-X. Sun and Z.-L. Yuan contributed to SCXRD and PXRD data experiments and analysis; H.-Y. Zhang conducted TG and IR experiments; Y.-L. Lai conducted ITC experiments and analysis. Z. Lu conducted the theoretical calculations. All authors read and commented on the manuscript.

Conflicts of interest

There are no conflicts to declare.

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