RSC Advances



View Article Online

View Journal | View Issue

PAPER



Cite this: RSC Adv., 2023, 13, 33067

Received 8th August 2023 Accepted 26th October 2023

DOI: 10.1039/d3ra05381d

rsc.li/rsc-advances

1. Introduction

Surface-enhanced Raman spectroscopy (SERS) is one of the most sensitive analytic techniques, which has been widely investigated for application in various fields,^{1,2} especially in surface chemical analysis.^{3,4} SERS effects occur when the inelastic light scattering of a molecule is enhanced in the presence of a SERS substrate, resulting in an enhancement factor of 10⁶–10¹⁴. There are two mechanisms contributing to the SERS phenomenon: electromagnetic mechanism (EM) and chemical mechanism (CM), leading to enormous enhancements in the Raman signal. On the

† Electronic supplementary information (ESI) available. See https://doi.org/10.1039/d3ra05381d

Enhanced sensing performance of carbaryl pesticide by employing a MnO₂/GO/e-Ag-based nanoplatform: role of graphene oxide as an adsorbing agent in the SERS analytical performance⁺

Dao Thi Nguyet Nga, (1) ‡^a Quan Doan Mai, (1) ‡^a Ha Anh Nguyen, (1) ‡^a Nguyen Le Nhat Trang, (1) ^a Pham Minh Khanh, ^a Nguyen Quang Hoa, ^b Vu Dinh Lam, ^c Van-Tuan Hoang (1) *^a and Anh-Tuan Le (1) ^{ad}

A functional ternary substrate was developed for surface-enhanced Raman scattering (SERS) sensing systems. MnO_2 nanosheets were synthesized by a simple and controllable hydrothermal method, followed by the integration of graphene oxide (GO) nanosheets. Subsequently, MnO_2/GO nanostructures were decorated with plasmonic Ag nanoparticles (e-AgNPs). The $MnO_2/GO/e$ -Ag substrate could enhance the SERS sensing signal for organic chemicals without the assistance of chemical bonds between those analytes and the semiconductor within the ternary substrate, which have been proven to promote charge transfer and elevate the SERS enhancement in previous studies. Instead, GO nanosheets acted as a carpet also supporting the MnO_2 nanosheets and e-AgNPs to form a porous structure, allowing the analytes to be well-adsorbed onto the ternary substrate, which improved the sensing performance of the SERS platform, compared to pure e-AgNPs, MnO_2/e -Ag, and GO/e-Ag alone. The GO content in the nanocomposite was also considered to optimize the SERS substrate. With the most optimal GO content of 0.1 wt%, $MnO_2/GO/e$ -Ag-based SERS sensors could detect carbaryl, a pesticide, at concentrations as low as 1.11×10^{-8} M in standard solutions and 10^{-7} M in real tap water and cucumber extract.

one hand, the EM was reported to be the major contributor.5,6 Hot spots with intense electromagnetic fields have been detected on corrugated surfaces, especially at sharp tips/corners of noble metal nanostructures, such as silver and gold, resulting in a significant enhancement in the Raman signal under the EM.^{7,8} On the other hand, in terms of the CM, the improvements in Raman signal are also contributed to by charge-transfer (CT) transitions between the substrate and the molecule. In a 2008 study, Jensen et al. described three types of CT transitions: interfacial ground-state charge transfer (GSCT), photoinduced charge transfer (PICT), and resonance Raman scattering (RRS).9 First, GSCT is characterized by chemisorption interactions between the molecule and the substrate in the ground state without excitations. Second, PICT is a CT process depending on the excitation wavelength assigned to the charge transition between the substrate and the molecule. Third, RRS occurs within the molecule itself when the laser excitation frequency is close to its electronic transition frequency. Although the contribution of the CM in total SERS enhancement was reported to be much lower than that of the EM, many studies have demonstrated the remarkable role of substrate-to-molecule CT in the improvements of the Raman signal while utilizing nanocomposite SERS substrates.10-12

8

^aPhenikaa University Nano Institute (PHENA), Phenikaa University, Hanoi 12116, Vietnam. E-mail: tuan.hoangvan@phenikaa-uni.edu.vn

^bFaculty of Physics, VNU University of Science, Vietnam National University, Hanoi, 334 Nguyen Trai, Thanh Xuan, Hanoi, Vietnam

^cInstitute of Materials Science (IMS) and Graduate University of Science and Technology (GUST), Vietnam Academy of Science and Technology, 18 Hoang Quoc Viet, Hanoi 10000, Vietnam

^dFaculty of Materials Science and Engineering (MSE), Phenikaa University, Hanoi 12116, Vietnam

[‡] D. T. N. Nga, M. Q. Doan and N. H. Anh contributed equally to this work.

In a recent study, we improved the SERS sensing performance of a tricyclazole (TCZ) sensor by employing a MnO₂/e-Ag nanocomposite as an active substrate, in comparison to the use of pure electrochemically synthesized silver nanoparticles (e-AgNPs). With a sulfur atom within the structure, TCZ could be linked to MnO₂ nanostructures via chemical bonds. Therefore, the presence of MnO₂ in the nanocomposite promoted more hot electrons from the e-AgNPs to be transferred to TCZ via a metal-to-semiconductor PICT pathway and then, a semiconductor-to-molecule IGCT pathway. This resulted in an EF of 1.38×10^6 when using MnO₂/e-Ag as the SERS substrate, which was 10 times higher than that when using pure e-AgNPs.13 Similar enhancements were also described in the report of Zhou et al., who employed ZnO nanorods/AuNPs to detect dopamine in a SERS sensing system.¹⁴ Therefore, taking advantage of the specific interaction between the desired analyte and the inorganic semiconductor, we designed a SERS active material to improve the Raman signal by promoting charge transfer via the IGCT pathway. However, there are numerous other compounds that are chemically inactive in the interaction with MnO₂, thus, it would not be convenient for them to experience the enhancement in CT arising on the nanocomposites. Thus, in this study, we aimed to continue modifying the nanomaterial to fill in the gap.

GO is a semiconductor material with abundant oxygencontaining groups, such as epoxide, carbonyl, and hydroxyl groups, which can be obtained by graphene modification. It can provide a large surface for the adsorption of various organic compounds.15,16 Furthermore, GO can be an excellent supporter for the grafting of other materials, thus it has potential to be a component in hybrid materials.^{17,18} There have been numerous studies into the construction of GO, semiconductor, and noble metals composite nanostructures as potential SERS substrates. For instance, Wang and co-authors utilized a novel Ag/GO/TiO₂ nanorod array as a SERS substrate for organic detection.¹⁹ Zhang et al. successfully prepared a coating of GO on a TiO₂-AgNPs nanoarray for the rapid, accurate, and reliable detection of multiple organic pollutants, including crystal violet, rhodamine 6G, and malachite green. Hence, in this study, we chose GO nanosheets to modify our metal-semiconductor nanocomposite, MnO₂/e-Ag, and employed the as-prepared ternary structure as a SERS substrate for carbaryl (CBR), a pesticide that does not directly bind to MnO2 via chemical bonds.

In this work, we employed MnO₂ nanosheets, GO nanosheets, and e-AgNPs, which were successfully fabricated as described in our previous studies,13,20 to prepare ternary nanostructures via two simple steps: First, the MnO₂/GO structure was created using ultrasonication, followed by the grafting of e-AgNPs onto that structure by stirring. The detailed preparation is shown in Scheme 1. Subsequently, it was demonstrated that MnO₂/GO/e-Ag could improve the SERS signal of CBR, compared to the use of pure e-AgNPs, MnO₂/e-Ag, and GO/e-Ag substrates. In addition, by varying the GO content, the substrates were optimized. With the most optimal GO content of 0.1 wt%, MnO₂/GO/e-Ag could detect CBR at concentrations down to 1.11×10^{-8} M in standard solutions and 10^{-7} M in real samples of tap water and cucumbers.

2. Experimental

Materials and SERS substrates 2.1.

Manganese dioxide (MnO₂) nanosheets, graphene oxide (GO) nanosheets, and e-Ag nanoparticles were synthesized as per our group's previous research.13,20 Cabarryl (CBR), tricyclazole (TCZ), chloramphenicol (CAP), methylene blue (MB), and 4 nitrophenol (4-NP) were provided by Sigma-Aldrich. All chemicals were used directly without further purification. The utilized conical flasks and Teflon-lined hydrothermal autoclave were treated with aqua regia for 24 h before the experiments. In addition, the double-distilled water used in the whole process was purified through a Milli-O[®] system (18.2 M Ω cm at 25 °C).

The aluminum (Al) substrate was prepared with dimensions of 1 cm \times 1 cm \times 0.2 cm with a surface-active area of diameter 0.2 cm. The substrate was cleaned properly. First, the substrates were soaked into a dilute hydrochloric acid solution to remove the oxide/hydroxide on the surface. Second, the substrate samples were washed with ethanol and double-distilled water by ultrasound for 15 min. Finally, the samples were dried naturally at room temperature.

2.2. Preparation of MnO₂/e-Ag nanocomposites

MnO₂/e-Ag nanocomposites were prepared as described in our previous report using two components: MnO2 nanosheets and e-AgNPs with the most optimal ratio of 1:3.13 Concerning the MnO2 nanosheets, 30 mL of a mixed solution containing 0.67 M NaOH and 9.79 M H₂O₂ was divided into 20 aliquots and poured



Scheme 1 Schematic illustration of the synthesis process of MnO₂/GO/e-Ag nanocomposites

Paper

into 10 mL of 0.3 M Mn(NO₃)₂ solution, at a rate of an aliquot per minute under constant stirring. Then, the solution mixtures were stirred together for 30 min. The collected precipitate was washed 3 times with double-distilled water and then dried. The dried precipitate was dispersed in 2 M NaOH. This homogeneous solution was poured into a hydrothermal autoclave reactor and heated at a temperature *T* of 160 °C with a holding time of 16 h. The obtained product was centrifuged, washed three times with ethanol and distilled water, and then dried at 60 °C. On the other hand, e-AgNPs were prepared using an electrochemical method, which was fully described in another previous study of our group, resulting in the formation of citrate-coated e-AgNPs with an average diameter of 24 nm and ζ potential value of about – 25 mV.²¹ Subsequently, 50 mg of MnO₂ was dissolved in 50 mL of double-distilled water with the help of ultrasound to form a homogeneous suspension. Then, the MnO2 solution was mixed with a certain volume of e-AgNP solution in a ratio of 1:3 and subjected to ultrasonic waves for 15 min. As a result, MnO₂/e-Ag was formed within the mixture. Full characterization of this nanocomposite was reported in our recent study.22

2.3. Preparation MnO₂/GO/e-Ag nanocomposites for SERS measurement

The MnO₂/GO/e-Ag nanocomposite was fabricated using the procedure described in Scheme 1. First, 50 mg of MnO₂ was completely dissolved in 50 mL double-distilled water under the aid of ultrasound treatment to form a homogeneous suspension. Then, the MnO₂ solution (1 mg mL^{-1}) was slowly added to a set of prepared mixtures containing different amounts of GO (0.05-0.5 wt%). GO was synthesized based on previous research.20 The above-mixed solutions were ultrasonicated for 30 min. Next, a defined volume of e-AgNPs solution (1 mg mL^{-1}) was added to the above mixture solution and mixed well with magnetic stirring for 30 min. Thus, the MnO₂/GO/e-Ag nanocomposites were synthesized.

2.4. Characterization and SERS measurement

Scanning electron microscopy (SEM) was used to observe the morphology, and size of the prepared samples. The composition and chemical properties of MnO2/GO/e-Ag were investigated by Raman spectroscopy (Horiba Macro-RAMt) with 785 nm laser excitation.

SERS active materials (*i.e.*, AgNPs, MnO₂/e-Ag, and MnO₂/ GO/e-Ag) were loaded onto the surface-active area of the Al substrates via evaporation. In detail, for each measuring substrate, 5 µL SERS active material solution was drop-cast directly on to the surface-active area of the Al substrate. Then it was dried at 50 °C before being used for SERS measurement.²¹ CBR solutions with different concentrations $(10^{-3}-10^{-8} \text{ M})$, TCZ (10^{-5} M) , MB (10^{-5} M) CAP (10^{-3} M) , and 4-NP (10^{-3} M) were prepared in distilled water before being dropped directly onto the prepared substrates. After that, they were dried naturally at room temperature and taken to perform the SERS measurements. The SERS spectra were monitored by a MacroRaman[™] Raman spectrometer (Horiba). Raman measurements were acquired by means of a 100×

objective with a numerical aperture of 0.90. The laser power was set to 45 mW at a 45° contact angle, with a focal length of 115 nm. An exposure time of 10 s was used with three accumulations. At every point of the SERS measurement, the experiment was repeated three times.

Cucumbers were purchased from a local supermarket and washed thoroughly using deionized water. Then the cucumbers were minced and mashed. Next, 25 g of the obtained cucumber was mixed with 30 mL of double-distilled water by a vortex mixer. The mixture was filtered with Whatman filter paper. Finally, the cucumber extract was collected. CBR standard solution at different concentrations was added into the cucumber extract samples. Subsequently, 5 µL spiked sample was drop-cast directly on to the prepared SERS substrates for SERS measurements under ambient room conditions.

To test the CBR detection in the tap-water sample, the experiments were conducted as follows. Different concentrations of CBR in the linear range $(10^{-4}-10^{-7} \text{ M})$ were prepared directly utilizing tap water, which was taken from the local area. Next, 5 µL CBR solution was drop-cast directly on to the prepared SERS substrates for the SERS measurements under ambient room conditions.

3. Results and discussion

3.1. Importance of the chemical bonds in the SERS effect on MnO₂ nanosheets

To confirm the importance of chemical bonds in the SERS effect on the MnO₂/e-Ag substrate, we evaluated the SERS enhancement of two groups of organic compounds. Group 1 consisted of chemicals that were reported to be degraded by MnO₂, thus, there would be chemical interactions between the semiconductor and the organic compounds, such as tricyclazole (TCZ) and methylene blue (MB). The chemicals in Group 2, in contrast, have not been reported to be degraded in the presence of MnO2. Moreover, they possessed no specific functional group to bind directly to MnO₂ nanosheets via chemical bonds. The molecular structures of the chemicals in Group 1 and Group 2 are shown in Table S1.†

Concerning Group 1, the interaction between TCZ and MnO₂ via the S atom of TCZ was fully described in our previous study.13 This chemical bond allows hot electrons to be transferred from MnO2 to TCZ via an interfacial ground-state chargetransfer (IGCT) pathway. Thus, using MnO₂/e-Ag nanocomposite as a SERS substrate, MnO2 would act as a bridge for CT from the laser-excited e-AgNPs to TCZ. As a result, more hot electrons can be transferred from the plasmonic nanomaterial to the analyte, leading to enhancements in the SERS signal of TCZ on MnO₂/Ag, compared to the use of pure e-AgNPs alone. In detail, at the concentration of TCZ (10^{-3} M), the SERS intensity of TCZ on MnO_2/e -Ag, measured by the peak of 431 cm⁻¹, was estimated to be 6.66 times higher than that on AgNPs. This result was presented in our previous study.13 Similar to TCZ, MB was reported to be degraded by MnO2. In a 2021 study, Yan et al. employed XPS characterization to confirm that the N atom within the benzene ring of MB could be directly linked to the MnO2 surface.23 Thus, it was expected that MnO2/e-Ag could



Fig. 1 SERS intensities of (a) MB (10⁻⁵ M), (b) CAP (10⁻³ M), (c) CBR (10⁻³ M), and (d) 4-NP (10⁻³ M) at the characteristic peaks on e-Ag, and MnO₂/ e-Ag nanocomposites.

enhance the Raman signal of MB better than pure e-AgNPs. This hypothesis was confirmed by the SERS measurements of MB (10^{-5} M) on MnO₂/e-Ag and pure e-AgNPs, as shown in Fig. 1a. On the MnO₂/e-Ag substrate, the SERS intensity of MB was obviously higher than that on pure e-AgNPs. For the characteristic peaks of 453, 1393, and 1622 cm⁻¹, the SERS intensities of MB on MnO₂/Ag were 2.83, 1.61, and 1.69 times higher than those on pure AgNPs. In contrast, the organic compounds in Group 2 were not reported to be degraded or linked to MnO2 via chemical bonds. The SERS spectra of chloramphenicol-CAP (10^{-3} M) , carbaryl-CBR (10^{-3} M) , and 4 nitrophenol-4-NP (10^{-3} M) on MnO₂/Ag were compared to those on pure AgNPs, as shown in Fig. 3b-d, respectively. It was clear that on the MnO₂/Ag substrate, the Raman signals of the organic compounds in Group 2 were less enhanced, compared to those on pure AgNPs, at their characteristic peaks. The assignments for the characteristic peaks of MB, CAP, CBR, and NP are listed in Table S1.† This result confirmed the importance of the chemical bonds between the substrate and the analyte in SERS enhancement. Furthermore, it raised a question of how to improve the sensing performance of SERS sensors for analytes that do not bind directly to the active substrates via chemical bonds. GO, with its excellent adsorbing ability, would be a solution for further material modification to overcome this obstacle.

3.2. Characterization of MnO₂/GO/e-Ag nanocomposite

A set of MnO₂/GO/e-Ag composites was prepared with different GO contents ranging from 0.05-0.5 wt%. Meanwhile, the Ag content of 75% was proven to be effective for the SERS phenomenon arising in our previous study,13 thus, this was fixed as the Ag content within every composite in this study. The contents of the components were calculated based on their weight added during preparation. Fig. 2a shows the Raman spectrum of MnO₂/GO/e-Ag, indicating the presence of all three components. Two characteristic peaks of GO could be observed at \sim 1313 cm⁻¹ (D band) and \sim 1605 cm⁻¹ (G band), representing the breathing modes of the rings (K-point phonons) with the A_{1g} symmetry and the in-plane bond-stretching E_{2g} mode of C sp² atoms, respectively.²⁴ In addition, the spectrum exhibited two bands at 585 and 645 cm⁻¹, which were assigned to the symmetric Mn–O vibrations and are characteristic of α-MnO₂ with a (2×2) tunnel.²⁵ Besides, the band at 240 cm⁻¹ was associated with the stretching vibrations of Ag-O bonds.26 Additionally, the presence of MnO₂, Ag, and GO in the nanocomposite was also confirmed by the XRD pattern, as shown in Fig. 2b. The XRD result revealed the tetragonal crystal system of α -MnO₂ with intense and sharp peaks for the (110), (200), (310), (400), (301), (600), and (521) crystal phase corresponding to JCPDS card no. 44-0141. The main diffraction peaks of Ag for the (111), (200), and (220) planes also existed in the XRD pattern

8

Paper



Fig. 2 Raman spectrum of (a) $MnO_2/GO/e$ -Ag; XRD pattern of (b) $MnO_2/GO/e$ -Ag (with 0.1 wt% of GO); SEM images of (c) GO and (d and e) $MnO_2/GO/e$ -Ag (with 0.1 wt% of GO) at different magnitudes.

(JCPDS no. 01-087-0719). The characteristic peak of GO was obtained at $2\theta = 11.5^{\circ}$, revealing the reflection plane (001).²⁷ Moreover, the formation of the nanocomposite was also confirmed by the SEM images. Due to the exceptional brightness of e-AgNPs compared to MnO2 and GO nanosheets under the electron microscope and low content of GO in the nanocomposite (0.1 wt%), unfortunately we could not observe the presence of GO in Fig. 2d and e. However, Fig. 2d shows that MnO2 nanosheets and e-AgNPs had gathered together to create micro-sized structures, corresponding to the structure displayed in Fig. 2c, which was formed by layers of GO nanosheets. Therefore, MnO2 could have covered GO sheets during the preparation of MnO₂/GO via ultrasonication, followed by the grafting of e-AgNPs onto this material. The assembly of MnO₂ on GO sheets could be observed more clearly in Fig. S6a and b.† Without the bright e-AgNPs, the MnO₂ nanostructures can be detected as small flakes on the large GO surface. With a higher content of GO, namely 2.0 wt% in MnO₂/GO/e-Ag nanocomposites, which would be referred to as 0.5 wt% in $MnO_2/GO/e$ -Ag nanocomposites, the surface of the GO sheet could be better observed in Fig. S6b.† In contrast, when the GO content was five-times lower, MnO_2 flakes covered it more properly (Fig. S6a†), forming a nest for harboring the e-AgNPs (Fig. 2e). Thus, the GO content influenced the structure of MnO_2/GO . Its effects on $MnO_2/GO/e$ -Ag then its SERS performance will be further discussed in the following sections.

3.3. Effects of GO content on SERS analytical performance of the MnO₂/GO/e-Ag nanocomposites-based sensing platform

To investigate the effect of GO and its content on the SERS performance of $MnO_2/GO/e$ -Ag nanocomposites-based SERS sensors, we measured and compared the SERS signals of CBR (10^{-3} M) on pure e-AgNPs and four types of $MnO_2/GO/e$ -Ag nanocomposites with different contents of GO (0.01, 0.1, 0.3, and 0.5 wt%) (Fig. 3a). On all the substrates, the SERS spectra exhibited characteristic peaks of CBR at 480, 723, and



Fig. 3 (a) SERS spectra of CBR (10^{-3} M) on e-AgNPs, and MnO₂/GO/e-Ag nanocomposites with different GO contents. (b) SERS intensities of CBR (10^{-3} M) at 480, 723, and 1440 cm⁻¹ on e-AgNPs, and MnO₂/GO/e-Ag nanocomposites with different GO contents.

1440 cm⁻¹. However, they showed significant differences in SERS intensity, revealing the levels of their sensing performance. Comparisons of the SERS intensities at 480, 723, and 1440 cm^{-1} are presented in Fig. 3b. With the addition of only 0.05 wt% of GO, the SERS intensity of CBR obviously increased, compared to the use of pure e-AgNPs. It is worth remembering that without GO, the SERS intensities at 480, 723, and 1440 cm^{-1} of CBR on MnO₂/e-Ag were 3.38, 3.02, and 2.83 times lower those on pure e-AgNPs, respectively (Fig. 1c). Moreover, they increased with the increase in GO content from 0.05 to 0.1 wt%. However, when the GO content reached 0.3 wt%, the SERS intensity of CBR dropped, and it was even lower than when using pure e-AgNPs. This downward trend continued when the GO content was raised to 0.5 wt%. Therefore, GO and its content showed noticeable effects on the SERS enhancement of CBR on these Ag-based substrates. In addition, using a GO content of 0.1 wt% in MnO2/GO/e-Ag nanocomposite as a SERS substrate for CBR detection resulted in an enhancement factor (EF) of 1.10×10^6 , which was 5.88 times higher than that when using e-AgNPs (EF = 1.87×10^5). The EF values were calculated as described in the ESI.† GO has been reported to be an excellent adsorbent for organic compounds. Many studies have also claimed that GO exhibited favorable adsorption sites for aromatic compounds,²⁸⁻³⁰ thanks to π - π and hydrophobic interactions. Although these interactions are not as strong as chemical bonds, which highly promote charge transfer, this adsorbent could allow more analytes, especially aromatic ones, to be adsorbed onto the substrate. As a result, more analytes could experience a SERS effect originating from e-AgNPs. With a naphthalene ring, which is a fusion of two aromatic rings, in the structure, CBR would have been well-adsorbed onto MnO₂/ GO/e-Ag thanks to the presence of GO nanosheets. However, it was not only GO that was responsible for the improvement of the SERS signal of CBR on MnO₂/GO/e-Ag in comparison to the use of pure e-AgNPs. Fig. S1[†] shows that also with a GO content of 0.1 wt%, the use of the GO/Ag nanocomposite did not result in better SERS enhancement than when using pure e-AgNPs alone. This emphasizes the role of the integration between MnO2 and GO within MnO2/GO/e-Ag for improving the SERS performance of the sensing platform. As mentioned in the previous section, this GO content led to the formation of nestlike MnO₂/GO structures (Fig. S6a[†]) for the anchoring of e-AgNPs. Besides, this porous structure of the composite could provide a larger surface area for the adsorption of CBR. When increasing the GO content to three- and five-time higher (i.e., 0.3 wt% and 0.5 wt%), the GO nanosheets could have veiled MnO₂ nanostructures during MnO₂/GO preparation, preventing them forming such a porous structure, and preventing CBR being well adsorbed onto the active substrate, leading to a remarkable decrease in SERS intensity, as shown in Fig. 3b. Fig. S6b[†] shows that with a high GO content, MnO₂/GO did not exhibit the nest-like morphology. Instead, a large surface of GO was revealed. Although in this sample, the GO content was fivetimes higher than that in Fig. S6a,† it was still extremely smaller than the MnO₂ content. Therefore, there was no such significant difference in MnO₂ content within those two structures, and the slightly lower content of MnO₂ should not be responsible for the fact that the MnO₂ nanosheets did not properly cover the GO surface. Instead, with a closer look at the GO surface in Fig. S6b,[†] we could observe flake-like structures covered by the GO sheet, confirming our hypothesis. Therefore, the GO content of 0.1 wt% was the most optimal for MnO₂/GO/ e-Ag as an active SERS substrate for CBR detection. On this substrate, the SERS signals of other compounds in Group 2, including CAP and 4-NP, were also enhanced, as shown in Fig. S2.[†] Hence MnO₂/GO/e-Ag could be employed as the SERS substrate to sense other organic molecules. However, in this study, we selected $MnO_2/GO/e$ -Ag nanocomposite with the most optimal GO content of 0.1 wt% to develop SERS sensors for CBR.

To confirm that the MnO₂/GO/e-Ag nanostructure with a GO content could improve the adsorption capacity of CBR, we performed an electrochemical model. Related to the electroactive surface area (EASA) of the modified electrodes using e-AgNPs, MnO₂/e-Ag, and MnO₂/GO/e-Ag (with 0.1 wt% of GO), cyclic voltammetric (CV) measurements were carried out in 0.1 M KCl containing 5 mM [Fe(CN)₆]^{3-/4} and at various scan rates ranging from 10–60 mV s⁻¹, as shown in Fig. 4. Changes in the response current intensity and the reversible redox peak potential could be detected while using every modified electrode. Corresponding linear plots were recorded between the peak current ($I_{\rm p}$, μ A) and the sqrt. of the scan rate ($\nu^{1/2}$). Therefore, the redox reactions on the electrode surface exhibited a diffusion process.

The EASA value was calculated based on the Randles–Sevick equation:³¹⁻³³

$$I_{\rm p} = 2.69 \times 10^5 \times n^{3/2} D^{1/2} A C v^{1/2} \tag{1}$$

where I_p is the peak current intensity, *n* is the number of electrons transferred (n = 1), *D* is the diffusion coefficient of $[Fe(CN)_6]^{3-/4-}$ ($D = 6.5 \times 10^{-6}$ cm s⁻¹), *A* is the EASA value, *v* is the scan rate, and *C* refers to the bulk concentration of the redox probe. The value of EASA was estimated to be about 0.137 cm² for Ag/SPE, 0.212 cm² for MnO₂/e-Ag_SPE, and 0.223 cm² for MnO₂/GO/e-Ag_SPE. Thus, the MnO₂/GO/e-Ag electrode had the largest active surface area.

CV measurements in pH 7 PBS buffer containing 50 μM CBR were carried out in the scan rate range from 20 to 70 mV s^{-1} to

investigate the response kinetics of CBR on the modified electrodes. Fig. 5 presents the impact of varying the scan rate (20–70 mV s⁻¹) on the electrochemical behaviors of CBR. The above results showed that as the scanning speed increased, all electrodes' anode/cathode maximum current gradually increased. Images 2a', b', and c' showed a highly linear relationship between $I_{\rm pa}$ and ν , with the linear regression equations as follows:

$$I_{\rm p} (\mu A) = 0.132 \ \nu \ (\text{mV s}^{-1}) + 0.494 \ (R^2 = 0.99) \text{ for e-Ag_SPE}$$
$$I_{\rm p} (\mu A) = 0.162 \ \nu \ (\text{mV s}^{-1}) + 0.79 \ (R^2 = 0.99) \text{ for MnO}_2/\text{e-Ag_SPE}$$
$$I_{\rm p} (\mu A) = 0.23 \ \nu \ (\text{mV s}^{-1}) + 1.07 \ (R^2 = 0.99) \text{ for MnO}_2/\text{GO/e-Ag_SPE}$$

Based on these linear relationships, the adsorption capacity (Γ) of CBR on the modified electrode surfaces was also determined through the equation:³⁴ $I_p = n^2 F^2 A \Gamma \nu / 4RT$. According to that, the Γ values of CBR were estimated to be around 19.4 \times 10^{-12} , 20 \times 10⁻¹², and 25.7 \times 10⁻¹² mol cm⁻² for e-Ag_SPE, MnO₂/e-Ag_SPE, and MnO₂/GO/e-Ag_SPE, respectively. Clearly, the MnO₂/GO/e-Ag modified electrode possessed the largest Γ value of CBR, followed by MnO2/e-Ag and then e-Ag. The increase in electrode active surface area would be related to the increase in adsorption capacity of the analyte. The improvement in adsorption capacity of CBR on MnO₂/GO/e-Ag indicated that the nanocomposite could adsorb the largest number of CBR molecules compared to the other materials, allowing them to experience the SERS effect by a chemical enhancement mechanism, resulting in the best enhanced Raman signal. This result confirms the importance of GO in the nanocomposite as a SERS substrate for the effective detection of CBR.



Fig. 4 (a-c): CV curves of the proposed electrodes at various scan rates ($10-60 \text{ mV s}^{-1}$) in 0.1 M KCl containing 5 mM [Fe(CN)₆]^{3-/4-}; and (a'-c'): corresponding linear plots of the reduction in peak current against the square root (sqrt.) of the scan rate.

8



Fig. 5 (a-c): CVs responses of 50 μ M CBR at the modified electrodes in 0.1 M PBS buffer at different scan rates, and (a'-c'): plots of the peak current (*I*) versus scan rate (ν).

3.4. SERS performance of MnO₂/GO/e-Ag nanocompositesbased carbary sensors

Fig. 6a displays the SERS spectra of CBR at various concentrations $(10^{-3}-10^{-8} \text{ M})$ on MnO₂/GO/e-Ag (with 0.1 wt% of GO). As CBR is a naphthalene-derivatized carbamate, the characteristic peaks of CBR were dominated by ones associated with naphthalene and the carbamate vibrational mode. The band at 453 cm⁻¹ was assigned to the low-frequency C–C bending modes. The band at 723 cm⁻¹ was associated with NCOC bending with some ring breathing character. The band at 1374 cm⁻¹ represented the symmetric ring vibration modes. The band at 1437 cm⁻¹ was related to the C–H wagging mode, while that at 1576 cm⁻¹ was related to the C–H wagging mode in the naphthalene ring.³⁵⁻³⁸ The SERS spectra of CBR on the MnO₂/GO/e-Ag substrate were consistent with the Raman spectrum of CBR (Fig. S3†).

With the decrease in the CBR concentration from 10^{-3} M to 10^{-8} M, the typical peak intensity of CBR pesticide demonstrated a sharp decline. When the concentration was the lowest at 10^{-7} M, the characteristic absorption bands of CBR at 480 cm⁻¹ could be still distinctly observed, showing that the MnO₂/GO/e-Ag material exhibited the effective sensitivity of the sensor to CBR pesticide detection, which was similar to some SERS sensors for CBR (Table 1). Fig. 6b shows a plot of the logarithmic SERS intensity at 480 cm⁻¹ against the logarithmic concentration of CBR with a linear relationship in the region from 10^{-4} – 10^{-7} M ($R^2 = 0.93$). The detection limit of the sensor was 1.11×10^{-8} M (~0.003 ppm), which was much lower than the maximum residue limit (MRL) for farm

products (10 ppm).³⁹ The quantification range from 10^{-4} - 10^{-7} M (~0.03 ppm to ~30 ppm) of the sensor also covered MRLs for food, such as cucumber (3 ppm), corn (1 ppm), and milk (0.05 ppm), established by Food and Agriculture Organization of the United Nations (FAO). Thus, it has the potential for food safety monitoring. Moreover, this MnO₂/GO/e-Ag substrate also presented relatively good uniformity in the detection of CBR on 30 different points on a MnO₂/GO/e-Ag substrate containing CBR (10^{-5} M) (Fig. 6c). The RSD was calculated to be 14.03%. In addition, the reproducibility of the substrate was determined by measuring the SERS signal of CBR (10^{-5} M) on five MnO₂/GO/e-Ag substrates, which were prepared similarly to each other, as shown in Fig. 6d. The RSD was determined to be 10.65%. The LOD and RSD values were calculated as described in the ESI.[†]

3.5. Practicability of the MnO₂/GO/e-Ag nanocompositesbased CBR sensors with real samples of tap water and cucumber

In order to evaluate the efficiency of the SERS sensor and their applications in real samples, CBR at four concentrations from 10^{-4} – 10^{-7} M were detected in tap water and cucumber extract samples. Fig. 7a and c display the SERS spectra of CBR in tap water and cucumber extract, respectively. They all exhibited characteristic peaks of CBR, which were similar to the standard solutions (Fig. 6a). However, the spectra of CBR in the cucumber extract sample showed several minor shifts and a few new peaks, which might be related to the interaction of the available components in the real sample, such as



Fig. 6 (a) SERS spectra of CBR ($10^{-3}-10^{-8}$ M) on MnO₂/GO/e-Ag. (b) Plot of the log of SERS intensity at 480 cm⁻¹ against CBR concentration. (c) Uniformity and (d) reproducibility of SERS sensors for CBR (10^{-5} M) using the MnO₂/GO/e-Ag substrate (with 0.1 wt% of GO).

Table 1	Comparison	with	other	CBR	sensors

Material	LOD	Linear range	Ref.
Ag nanoparticles-coated Si nanowire-SiNW	$1.00 imes 10^{-7}~{ m M}$	10^{-2} - 10^{-7} M	39
Poly(ethyleneterephthalate) PET/ITO/Ag	${\sim}4.00 imes10^{-7}~{ m M}$	_	40
Gold nanorod arrays	${\sim}2.5 imes10^{-6}~{ m M}$	0–1.5 $ imes$ 10 $^{-4}$ M	41
Silver nanoparticles on cotton swabs	$5 imes 10^{-7}~{ m M}$	$5 imes 10^{-4}$ – $5 imes 10^{-7}~{ m M}$	42
MnO ₂ /GO/e-Ag	$1.11\times 10^{-8}~\text{M}$	10^{-3} - 10^{-7} M	This work

carbohydrate, pectin, and minerals. In tap water, inorganic ions could also be absorbed onto the SERS substrate. These constituents could inhibit the connection between the target analytes and the nanoparticles surface, thus reducing the enhancement signals of the SERS substrate.⁴³ However, the ternary material could detect CBR in real samples at concentration down to 10^{-7} M (~0.03 ppm) (Fig. 7b and d), which is lower than the maximum residue limit (MRL) for cucumbers and other melons established by the FAO (3 ppm). The recovery rates ranged from 84–107% with RSDs of 9.70– 13.46% in tap water and from 87–111% with RSDs of 7.42– 13.57% in cucumber extract (Table S2†), indicating the high accuracy of the sensing system when employed for the detection of CBR in real samples. However, real samples are usually complicated with interferences from other chemical compounds in addition to the targeted analyte. For example, a vegetable sample may contain more than one pesticide. Therefore, in this study, we designed an experiment to determine the presence of CBR in cucumber samples containing two other pesticides: 4-nitrophenol and glyphosate. As previously mentioned, CBR did not form any specific bonds with MnO₂/GO/e-Ag; therefore, the sensing platform did not exhibit high selectivity as the SERS spectra showed characteristic peaks of all those pesticides with much noise (Fig. S4†). However, it is worth stressing that in the interferences of the other two, the characteristic peak at 480 cm⁻¹ of



Fig. 7 SERS spectra of TCZ $(10^{-4}-10^{-7} \text{ M})$ in real samples of (a) tap water and (c) cucumber on the MnO₂/GO/e-Ag (with 0.1 wt% of GO) SERS substrate; SERS spectra of CBR (10^{-7} M) on MnO₂/GO/e-Ag (with 0.1 wt% of GO) in (b) tap water, and (d) cucumber samples.

CBR could still be detected at concentrations down to 10^{-6} M. Based on the intensity of the peak, the CBR concentration could be calculated with recoveries of 94–110% and RSDs of 11.54–14.52% (Table S3[†]).

4. Conclusions

In this work, we performed a modification of the MnO₂/e-Ag substrate reported in our recent research due to its limited ability to form links between the substrate and organic molecules. In summary, we successfully fabricated a sensitive and reliable MnO₂/GO/e-Ag nanocomposite SERS substrate for the detection of carbaryl pesticide through some simple mechanical methods, including ultrasound and stirring. The influence of GO on the SERS sensing performance of the MnO₂/e-Ag nanocomposite was systematically investigated. The results displayed that the synergistic effects of MnO2 and GO nanosheets endowed the sensing substrate with improved sensitivity, repeatability, uniformity, and stability. With the GO content of 0.1 wt%, the SERS sensor exhibited low detection limits for CBR (1.11 \times 10^{-8} M in standard solutions and 10^{-7} M in real samples of tap water and cucumber extract). Moreover, the composite nanomaterial could also enhance the SERS signals of other organic compounds that do not bind directly to MnO₂, such as CAP and 4-NP. Hence, this upgraded MnO₂/GO/ e-Ag nanocomposite will be an ideal SERS substrate with broad applications for the rapid detection of organic residues.

Author contributions

D. T. N. Nga: conceptualization, validation, investigation, Q. D. Mai: validation, investigation; H.A. Nguyen: conceptualization, methodology, formal analysis, writing-review & editing; writingoriginal draft; N. L. N. Trang: validation, investigation; M. K. Pham: validation, investigation; H. V. Tuan: methodology, investigation, supervision; A. T. Le: conceptualization, methodology, supervision, project administration, writing-review & editing.

Conflicts of interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

Acknowledgements

This research was supported by the Phenikaa University under grant number 2-04.2021.03 and was also acknowledged to the Phenikaa Group through Financial Supports for Key Research Group (NEB Lab). The authors would like to acknowledge the supports for Raman, Electrochemical & UV-vis measurements from NEB Lab (Phenikaa University), XRD measurements from VNU-Hanoi and SEM characterization from GUST-VAST.

Paper

References

- 1 A. Huefner, W. L. Kuan, K. H. Müller, J. N. Skepper, R. A. Barker and S. Mahajan, Characterization and visualization of vesicles in the endo-lysosomal pathway with surface-enhanced raman spectroscopy and chemometrics, ACS Nano, 2016, 10(1), 307-316, DOI: 10.1021/acsnano.5b04456.
- 2 J. Su, et al., Multicolor Gold-Silver Nano-Mushrooms as Ready-to-Use SERS Probes for Ultrasensitive and Multiplex DNA/miRNA Detection, Anal. Chem., 2017, 89(4), 2531-2538, DOI: 10.1021/acs.analchem.6b04729.
- 3 K. Sivashanmugan, J. Der Liao, B. H. Liu, C. K. Yao and S. C. Luo, Ag nanoclusters on ZnO nanodome array as hybrid SERS-active substrate for trace detection of malachite green, Sens. Actuators, B, 2015, 207(Part A), 430-436, DOI: 10.1016/j.snb.2014.10.088.
- 4 J. Huang, D. Ma, F. Chen, M. Bai, K. Xu and Y. Zhao, Ag Nanoparticles Decorated Cactus-Like Ag Dendrites/Si Nanoneedles as Highly Efficient 3D Surface-Enhanced Raman Scattering Substrates toward Sensitive Sensing, Anal. Chem., 2015, 87(20), 10527-10534, DOI: 10.1021/ acs.analchem.5b02788
- 5 V. Kiran and S. Sampath, Enhanced raman spectroscopy of molecules adsorbed on carbon-doped TiO 2 obtained from titanium carbide: A visible-light-assisted renewable substrate, ACS Appl. Mater. Interfaces, 2012, 4(8), 3818-3828, DOI: 10.1021/am300349k.
- 6 H. Zhang, S. Yang, Q. Zhou, L. Yang, P. Wang and Y. Fang, The Suitable Condition of Using LSPR Model in SERS: LSPR Effect Versus Chemical Effect on Microparticles Surface-Modified with Nanostructures, Plasmonics, 2017, 12(1), 77-81, DOI: 10.1007/s11468-016-0231-4.
- 7 G. McNay, D. Eustace, W. E. Smith, K. Faulds and D. Graham, Surface-enhanced Raman scattering (SERS) and surface-enhanced resonance raman scattering (SERRS): A review of applications, Appl. Spectrosc., 2011, 65(8), 825-837, DOI: 10.1366/11-06365.
- 8 T. Tan, et al., LSPR-dependent SERS performance of silver nanoplates with highly stable and broad tunable LSPRs prepared through an improved seed-mediated strategy, Phys. Chem. Chem. Phys., 2013, 15(48), 21034-21042, DOI: 10.1039/c3cp52236a.
- 9 L. Jensen, C. M. Aikens and G. C. Schatz, Electronic structure methods for studying surface-enhanced Raman scattering, Chem. Soc. Rev., 2008, 37(5), 1061-1073, DOI: 10.1039/ b706023h.
- 10 A. Musumeci, et al., SERS of semiconducting nanoparticles (TiO 2 hybrid composites), J. Am. Chem. Soc., 2009, 131(17), 6040-6041, DOI: 10.1021/ja808277u.
- 11 L. Zhou, et al., Irreversible accumulated SERS behavior of the molecule-linked silver and silver-doped titanium dioxide hybrid system, Nat. Commun., 2020, 11(1), 1-10, DOI: 10.1038/s41467-020-15484-6.
- 12 L. Yang, X. Jiang, W. Ruan, B. Zhao, W. Xu and J. R. Lombardi, Observation of enhanced Raman scattering

for molecules adsorbed on TiO 2 nanoparticles: chargetransfer contribution, J. Phys. Chem. C, 2008, 112(50), 20095-20098, DOI: 10.1021/jp8074145.

- 13 D. T. N. Nga, et al., A manganese dioxide-silver nanostructure-based SERS nanoplatform for ultrasensitive tricyclazole detection in rice samples: effects of semiconductor morphology on charge transfer efficiency and SERS analytical performance, Phys. Chem. Chem. Phys., 2023, 25, 17496-17507.
- 14 J. Zhou, et al., Plasmon-induced hot electron transfer in Au-ZnO heterogeneous nanorods for enhanced SERS, Nanoscale, 2019, 11(24), 11782-11788, DOI: 10.1039/ c9nr02969a.
- 15 A. Peyravi, F. Ahmadijokani, M. Arjmand and Z. Hashisho, Graphene oxide enhances thermal stability and microwave absorption/regeneration of a porous polymer, J. Hazard. Mater., 2022, 433, 128792, DOI: 10.1016/ j.jhazmat.2022.128792.
- 16 F. Khan, M. S. Khan, S. Kamal, M. Arshad, S. I. Ahmad and S. A. A. Nami, Recent advances in graphene oxide and reduced graphene oxide based nanocomposites for the photodegradation of dyes, J. Mater. Chem. C, 2020, 8(45), 15940-15955, DOI: 10.1039/d0tc03684f.
- 17 M. Kim, M. Yoo, Y. Yoo and J. Kim, Capacitance behavior of composites for supercapacitor applications prepared with different durations of graphene/nanoneedle MnO2 reduction, Microelectron. Reliab., 2014, 54(3), 587-594, DOI: 10.1016/j.microrel.2013.11.005.
- 18 M. Zhang, et al., The influences of graphene oxide (GO) and plasmonic Ag nanoparticles modification on the SERS sensing performance of TiO2 nanosheet arrays, J. Alloys Compd., 2021, 864, 158189, DOI: 10.1016/ j.jallcom.2020.158189.
- 19 Y. Wang, et al., Facile fabrication of Ag/graphene oxide/TiO2 nanorod array as a powerful substrate for photocatalytic degradation and surface-enhanced Raman scattering detection, Appl. Catal., B, 2019, 252(April), 174-186, DOI: 10.1016/j.apcatb.2019.03.084.
- 20 D. T. Nguyet Nga, et al., Elucidating the roles of oxygen functional groups and defect density of electrochemically exfoliated GO on the kinetic parameters towards furazolidone detection, RSC Adv., 2022, 12(43), 27855-27867, DOI: 10.1039/d2ra04147b.
- 21 M. Q. Doan, et al., Ultrasensitive Detection of Methylene Blue Using an Electrochemically Synthesized SERS Sensor Based on Gold and Silver Nanoparticles: Roles of Composition and Purity on Sensing Performance and Reliability, J. Electron. Mater., 2022, 51(1), 150-162, DOI: 10.1007/s11664-021-09228-5.
- 22 D. T. Nguyet Nga, et al., A manganese dioxide-silver nanostructure-based SERS nanoplatform for ultrasensitive tricyclazole detection in rice samples: effects of semiconductor morphology on charge transfer efficiency and SERS analytical performance, Phys. Chem. Chem. Phys., 2023, 25(26), 17496-17507, DOI: 10.1039/D3CP01592K.

- 23 G. Yan, et al., Adsorption-Oxidation Mechanism of δ-MnO₂ to Remove Methylene Blue, Adsorpt. Sci. Technol., 2021, 2021, 3069392, DOI: 10.1155/2021/3069392.
- 24 L. G. Cançado, M. A. Pimenta, B. R. A. Neves, M. S. S. Dantas and A. Jorio, Influence of the atomic structure on the Raman spectra of graphite edges, *Phys. Rev. Lett.*, 2004, 93(24), 5–8, DOI: 10.1103/PhysRevLett.93.247401.
- 25 Q. Q. Li, *et al.*, Rapid and sensitive detection of pesticides by surface-enhanced Raman spectroscopy technique based on glycidyl methacrylate-ethylene dimethacrylate (GMA-EDMA) porous material, *Chin. Chem. Lett.*, 2013, 24(4), 332–334, DOI: 10.1016/j.cclet.2013.02.002.
- 26 A. J. Kora and J. Arunachalam, Green fabrication of silver nanoparticles by gum tragacanth (astragalus gummifer): A dual functional reductant and stabilizer, *J. Nanomater.*, 2012, 2012, 869765, DOI: 10.1155/2012/869765.
- 27 X. Qiao, S. Liao, C. You and R. Chen, Phosphorus and Nitrogen Dual Doped and Simultaneously Reduced Graphene Oxide with High Surface Area as Efficient Metal-Free Electrocatalyst for Oxygen Reduction, *Catalysts*, 2015, 5, 981–991, DOI: 10.3390/catal5020981.
- 28 S. Zhang, W. Li, H. Tang, T. Huang and B. Xing, Revisit the adsorption of aromatic compounds on graphene oxide: Roles of oxidized debris, *Chem. Eng. J.*, 2022, 450, 137996, DOI: 10.1016/j.cej.2022.137996.
- 29 H. Tang, *et al.*, Wrinkle- and Edge-Adsorption of Aromatic Compounds on Graphene Oxide as Revealed by Atomic Force Microscopy, Molecular Dynamics Simulation, and Density Functional Theory, *Environ. Sci. Technol.*, 2018, 52(14), 7689–7697, DOI: 10.1021/acs.est.8b00585.
- 30 H. Tang, et al., Theoretical insight into the adsorption of aromatic compounds on graphene oxide, Environ. Sci.: Nano, 2018, 5(10), 2357–2367, DOI: 10.1039/c8en00384j.
- 31 T. N. Pham, *et al.*, Roles of Phase Purity and Crystallinity on Chloramphenicol Sensing Performance of CuCo2O4/ CuFe2O4-based Electrochemical Nanosensors, *J. Electrochem. Soc.*, 2021, 168(2), 26506, DOI: 10.1149/1945-7111/abde80.
- 32 T. N. Pham, X. N. Pham, H. Thi Nguyen, T. Pham, Q. H. Tran and A.-T. Le, Exploiting the Balance Between Conductivity and Adsorption Capacity/Redox Electrocatalytic Ability In MIL-Based Porous Crystalline Materials for the Electrochemical Response, *J. Electrochem. Soc.*, 2022, **169**(5), 56521, DOI: **10.1149/1945-7111/ac707b**.
- 33 S. K. Ponnaiah, P. Periakaruppan and B. Vellaichamy, New Electrochemical Sensor Based on a Silver-Doped Iron Oxide Nanocomposite Coupled with Polyaniline and Its Sensing Application for Picomolar-Level Detection of Uric Acid in

Human Blood and Urine Samples, *J. Phys. Chem. B*, 2018, **122**(12), 3037–3046, DOI: **10.1021/acs.jpcb.7b11504**.

- 34 T. N. Pham, *et al.*, An insight of light-enhanced electrochemical kinetic behaviors and interfacial charge transfer of CuInS(2)/MoS(2)-based sensing nanoplatform for ultra-sensitive detection of chloramphenicol, *Anal. Chim. Acta*, 2023, **1270**, 341475, DOI: **10.1016**/ **j.aca.2023.341475**.
- 35 R. L. McCreery, *Raman spectroscopy for chemical analysis*, John Wiley & Sons, 2005.
- 36 B. Liu, P. Zhou, X. Liu, X. Sun, H. Li and M. Lin, Detection of Pesticides in Fruits by Surface-Enhanced Raman Spectroscopy Coupled with Gold Nanostructures, *Food Bioprocess Technol.*, 2013, 6(3), 710–718, DOI: 10.1007/ s11947-011-0774-5.
- 37 Y. Fan, K. Lai, B. A. Rasco and Y. Huang, Determination of carbaryl pesticide in Fuji apples using surface-enhanced Raman spectroscopy coupled with multivariate analysis, *Lwt*, 2015, **60**(1), 352–357, DOI: **10.1016/j.lwt.2014.08.011**.
- 38 C. S. Shende, F. Inscore, A. Gift, P. Maksymiuk and S. Farquharson, Analysis of pesticides on or in fruit by surface-enhanced Raman spectroscopy, *Nondestruct. Sens. Food Safety, Qual. Nat. Resour.*, 2004, 5587, 170, DOI: 10.1117/12.569595.
- 39 X. T. Wang, W. S. Shi, G. W. She, L. X. Mu and S. T. Lee, Highperformance surface-enhanced Raman scattering sensors based on Ag nanoparticles-coated Si nanowire arrays for quantitative detection of pesticides, *Appl. Phys. Lett.*, 2010, 96(5), 2008–2011, DOI: 10.1063/1.3300837.
- 40 A. B. Nowicka, M. Czaplicka, A. A. Kowalska, T. Szymborski and A. Kamińska, Flexible PET/ITO/Ag SERS Platform for Label-Free Detection of Pesticides, *Biosensors*, 2019, **9**(3), 111, DOI: **10.3390/bios9030111**.
- 41 Z. Zhang, Q. Yu, H. Li, A. Mustapha and M. Lin, Standing Gold Nanorod Arrays as Reproducible SERS Substrates for Measurement of Pesticides in Apple Juice and Vegetables, *J. Food Sci.*, 2015, 80(2), N450–N458, DOI: 10.1111/1750-3841.12759.
- 42 L. L. Qu, Y. Y. Geng, Z. N. Bao, S. Riaz and H. Li, Silver nanoparticles on cotton swabs for improved surfaceenhanced Raman scattering, and its application to the detection of carbaryl, *Microchim. Acta*, 2016, **183**(4), 1307– 1313, DOI: **10.1007/s00604-016-1760-4**.
- 43 B. Hu, D. W. Sun, H. Pu and Q. Wei, Rapid nondestructive detection of mixed pesticides residues on fruit surface using SERS combined with self-modeling mixture analysis method, *Talanta*, 2020, 217(March), 120998, DOI: 10.1016/ j.talanta.2020.120998.