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In vivo visualization of enantioselective targeting of amyloid and improvement of cognitive function by clickable chiral metallohelices†

Zhi Du,^a Chun Liu,^{ab} Zhenqi Liu,^{ab} Hualong Song,^{ID c} Peter Scott,^{ID c} Xiubo Du,^d Jinsong Ren^{ID ab} and Xiaogang Qu^{ID *ab}

The pathogenesis of Alzheimer's disease (AD) is closely related to several contributing factors, especially amyloid- β (A β) aggregation. Bioorthogonal reactions provide a general, facile, and robust route for the localization and derivatization of A β -targeted agents. Herein, a pair of chiral alkyne-containing metallohelices ($\Delta\Delta$ and $\Delta\Lambda$) were demonstrated to enantioselectively target and modulate A β aggregation, which has been monitored in triple-transgenic AD model mice and proved to improve cognitive function. Compared with its enantiomer $\Delta\Lambda$, $\Delta\Delta$ performed better in blocking A β fibrillation, relieving A β -triggered toxicity, and recovering memory deficits *in vivo*. Moreover, clickable $\Delta\Delta$ could act as a functional module for subsequent visualization and versatile modification of amyloid *via* bioorthogonal reaction. As a proof-of-concept, thioflavin T, tacrine, and magnetic nanoparticles were conjugated with $\Delta\Delta$ to realize A β photo-oxygenation, acetylcholinesterase inhibition, and A β clearance, respectively. This proof-of-principle work provided new insights into the biolabeling and bioconjugation of multifunctional metallosupramolecules through click reactions for AD therapy.

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Introduction

Alzheimer's disease (AD) is a stubborn neurodegenerative disorder that plagues millions of people worldwide.^{1,2} The accumulation and aggregation of amyloid- β (A β) peptides are implicated as the initial and prominent pathophysiology features of AD and further trigger a series of cascade reactions, such as oxidative stress, neurotransmitter deficiencies, and neuronal injury.³⁻⁵ A β -binding agents have been actively developed to modulate the amyloidogenic behavior and alleviate concomitant cytotoxicity.⁶ Tracking A β ligands in real-time is of great significance for precision medicine, which conduces to identify the distribution of the ligands and evaluate treatment effect. To tackle poly-pathological features in AD, the rational design of multifunctional agents by integrating two or more target-interacting moieties into one single molecule has gained particular attention.⁷⁻⁹

As a quintessential bioorthogonal reaction, Cu(I)-mediated azide/alkyne cycloaddition (CuAAC) is characterized by high

selectivity, mild conditions, and fast kinetics.¹⁰⁻¹² Thus, it has been extensively investigated in the biochemical research, such as bioconjugation,^{13,14} biolabeling,^{15,16} and prodrug activation.^{17,18} From this point of view, bioorthogonal catalysis has a great potential to endow A β binders with additional capabilities for their localization and versatile modification to address the multifactorial nature of AD.

Owing to the distinctive physicochemical properties, various metal complexes display a fascinating affinity with A β and thus are exploited for the theranostics of AD.¹⁹⁻²² Given that A β aggregates carry the chirality at different-length scales, the incorporation of enantioselective recognition is a bonus for AD therapeutics.²³⁻²⁵ Chiral metallohelices, a type of three-dimensional metal complexes assembled by multidentate organic ligands wrapping around two or more metal centers, are analogous to α -helical peptides in terms of size, amphiphilicity, and stereochemistry.²⁶⁻³⁰ As potential non-peptide mimetics, a pair of alkyne-bearing metallohelices enantiomers ($\Delta\Delta$ and $\Delta\Lambda$, Scheme 1)³¹ are chosen, as an example to employ bioorthogonal activity for enantioselective targeting and visualization of amyloid without the need of *de novo* synthesis.

Results and discussion

Enantioselective modulation of A β aggregation by $\Delta\Delta$

The influence of the chiral metallohelices $\Delta\Delta$ and $\Delta\Lambda$ on the course of A β 40 fibril formation was investigated using thioflavin T (ThT) assays, circular dichroism (CD), and transmission

^aLaboratory of Chemical Biology, State Key Laboratory of Rare Earth Resource Utilization, Changchun Institute of Applied Chemistry, Chinese Academy of Sciences, Changchun, Jilin 130022, China. E-mail: xqu@ciac.ac.cn

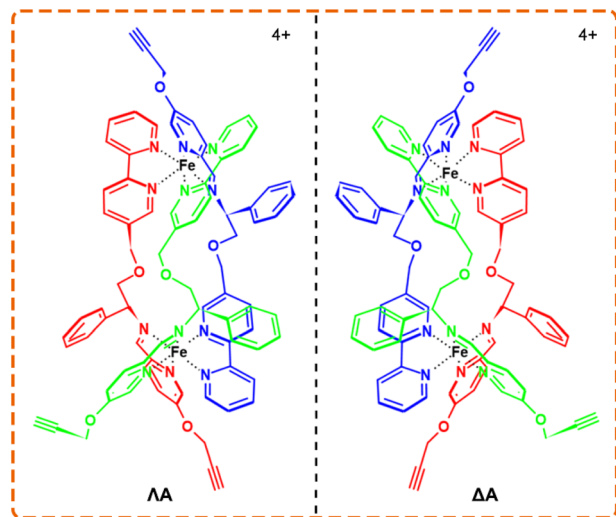
^bUniversity of Science and Technology of China, Hefei, Anhui 230029, China

^cDepartment of Chemistry, University of Warwick, Coventry CV4 7AL, UK

^dCollege of Life Sciences and Oceanography, Shenzhen University, Shenzhen 518060, P. R. China

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Scheme 1 The structures of Δ (left-handed twist) and $\Delta\Delta$ (right-handed twist) enantiomers of the chiral metallohelices.

electron microscopy (TEM) (Fig. 1). ThT, whose fluorescence is augmented considerably upon binding to β -sheet-rich aggregates, is commonly exploited for monitoring the amyloid aggregation kinetics. ThT assays demonstrated that $\Delta\Delta$ exhibited potent inhibition of A β 40 aggregation while $\Delta\Delta$

displayed a weak effect (Fig. 1b). The control experiment indicated the chiral metallohelices had a small impact on the binding of ThT to A β fibrils (Fig. S1b[†]). In addition, the free chiral ligands of $\Delta\Delta$ and $\Delta\Delta$ had very feeble regulation ability on A β 40 aggregation (Fig. S1d[†]), indicating that stereoscopic structures of the chiral metallohelices played a vital role on A β discrimination.

CD spectra, secondary structure analysis, and TEM images further verified that $\Delta\Delta$ prohibited A β 40 from aggregating into β -sheet-rich fibrils more powerfully than its enantiomer (Fig. 1c–e). Besides, $\Delta\Delta$ could also block A β 42 fibrillation, as depicted in Fig. S2.[†] The performance of the chiral metallohelices on A β 42 oligomerization was evaluated with a dot blotting immunoassay. As shown in Fig. S2d,[†] the sample treated with $\Delta\Delta$ displayed a weaker reactivity to A β oligomer-specific antibody A11, indicating that $\Delta\Delta$ decreased the formation of toxic oligomeric intermediates. UV-Vis absorption spectra and high-resolution mass spectra (HRMS) demonstrated the chiral metallohelices were stable in the phosphate-buffered saline (PBS) (Fig. S3[†]).

The complexation between $\Delta\Delta$ and A β

Isothermal titration calorimetry (ITC), a distinguished method for evaluating molecular interactions, was employed to estimate the apparent binding constant (K_a), enthalpy change (ΔH), and binding stoichiometry (n) of A β 40 with chiral metallohelices. According to Fig. 2a and b, an exothermic ITC isotherm was recorded and the binding stoichiometry was close to 1 : 1. $\Delta\Delta$ showed a higher binding affinity to A β 40 ($K_a = 8.05 \times 10^6 \text{ M}^{-1}$) than the Δ enantiomer ($K_a = 9.38 \times 10^5 \text{ M}^{-1}$), and the corresponding Gibbs free energy changes (ΔG) were -39.4 and $-34.1 \text{ kJ mol}^{-1}$, respectively. Competition dialysis experiment (Fig. S4[†]) further confirmed the higher affinity between A β 40 and $\Delta\Delta$.

The fragments 12–28 and 25–35, which are the critical regions responsible for A β aggregation,³² were utilized to explore the binding sites of A β to the chiral metallohelices. As shown in Fig. S5,[†] $\Delta\Delta$ efficiently blocked the aggregation of A β 12–28 while it displayed modest modulation ability to A β 25–35, implying that $\Delta\Delta$ preferentially binds to A β 12–28 rather than A β 25–35. Remarkably, A β 15–20 is considered the self-recognition region and often exploited as an A β -target peptide.³³ Table S1[†] revealed that $\Delta\Delta$ possessed comparative regulation capability towards the fibrillation of A β 15–20 and A β 1–40. It is inferred from the above results that $\Delta\Delta$ might bind to A β 15–20 as the mechanism of altering A β aggregation.

To further dissect the structural features of the chiral metallohelices and A β 40, we performed docking studies with AutoDock Vina. A β 1–40 monomer (PDB ID 2LFM), in which the core hydrophobic region adopts a helical conformation,³⁴ was used as a working model for molecular docking. Interestingly, $\Delta\Delta$ was nearly parallel to the α -helix of the A β monomer while Δ was sloping (Fig. 2c–f). As a result, there were four sets of cation– π and σ – π interactions together between the aromatic nuclei of $\Delta\Delta$ and the K16 residue of A β (Fig. 2d); this is in accord with aforementioned observation from inhibiting aggregation

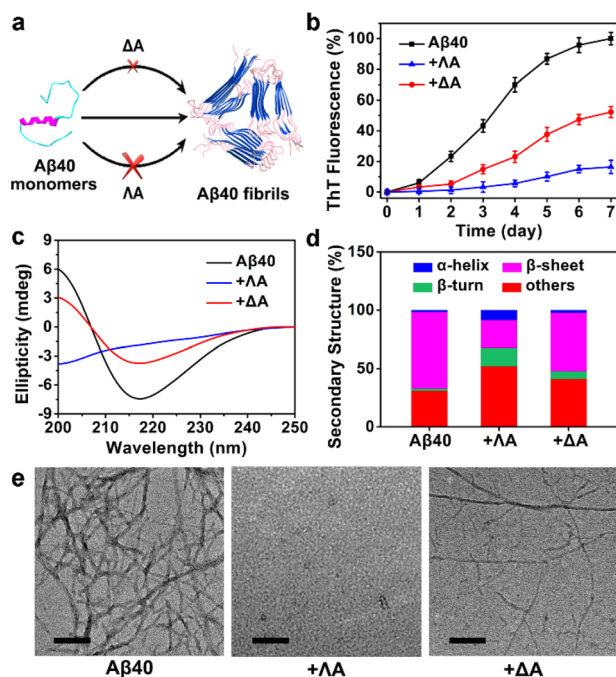


Fig. 1 Enantioselective regulation on A β 40 fibrillation with the chiral metallohelices. (a) Overview of interrupting A β 40 aggregation with $\Delta\Delta$ and $\Delta\Delta$. (b) ThT assays for monitoring A β 40 fibrillogenesis. Error bars represented \pm standard deviation (s.d.) of three independent experiments. (c) CD spectra of A β 40. For the spectra of A β in the presence of metallohelices, the CD spectra of the metallohelices were subtracted to reduce interference. (d) Secondary structure content estimations. (e) TEM images of A β 40 aggregates. Scale bars are 50 nm.



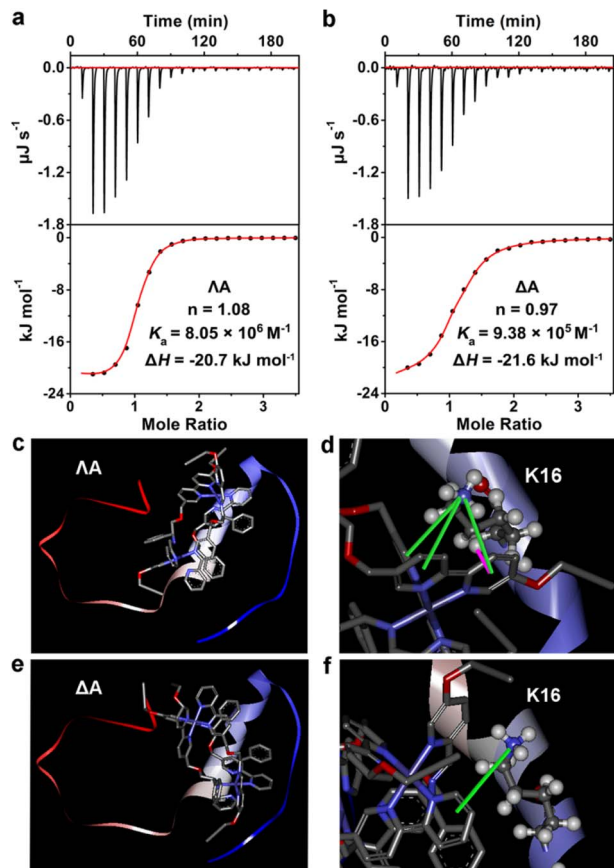


Fig. 2 The interaction between A β 40 and Δ A. (a) and (b) Presented ITC data of A β 40 binding to Δ A and Δ A, respectively. The top panels are raw calorimetric data, and the bottom panels are fitting results. (c) and (d) Showed the lowest-energy conformations of A β 40 with Δ A. (e) and (f) Displayed the lowest-energy conformations of A β 40 with Δ A. The cation- π and σ - π interactions were indicated by green and magenta lines, respectively.

assays of A β fragments. In contrast, only one set of cation- π interaction was found between Δ A and A β (Fig. 2f). This structural analysis provides a rationale for the stronger binding between Δ A and A β and more powerful modulation activity towards amyloid aggregation of Δ A.

Alleviation of A β -induced toxicity by Δ A in cells and the nematode model

A β -triggered oxidative stress is another feature in AD pathogenesis.³⁵ It has been proven that quenching overproduced reactive oxygen species (ROS) plays an active role in delaying AD progression.^{36,37} A series of Fe complexes have been reported to mimic catalase (CAT) or superoxide dismutase (SOD) and display promising anti-oxidation features.^{38–40} Hence we speculated that the chiral metallohelicenes might also scavenge deleterious ROS. As expected, Δ A and Δ A efficiently decomposed H₂O₂ (Fig. 3a and b) and slowed down pyrogallol auto-oxidation (Fig. 3c and d), confirming their CAT-mimic and SOD-mimic activities.

Based on the above encouraging results, we further investigated whether the metallohelicenes were able to protect PC12 cells

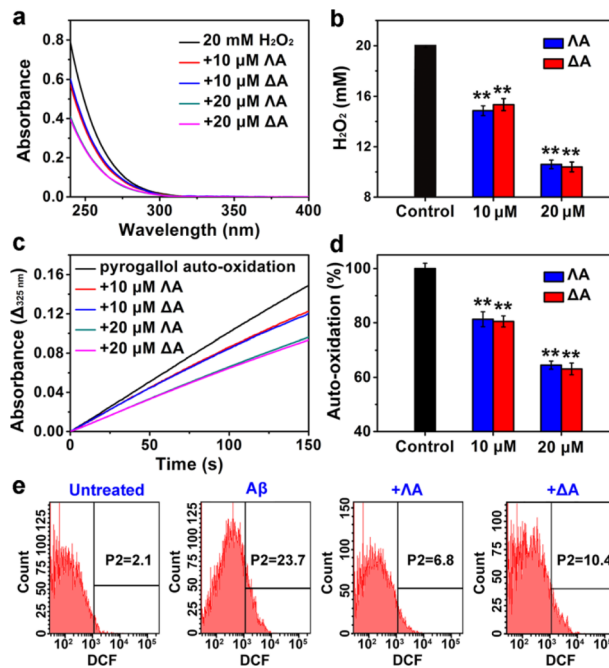


Fig. 3 ROS-quenching properties of Δ A and Δ A. (a) CAT-mimic ability measured by UV-Vis absorption of H₂O₂. (b) The concentration of remained H₂O₂ after incubating with chiral metallohelicenes for 30 min. (c) SOD-like activity monitored by the kinetic study of pyrogallol auto-oxidation. The absorbance change at 325 nm ($\Delta_{325\text{nm}}$) represented oxidated pyrogallol in 150 s. (d) Percent of pyrogallol auto-oxidation. Pyrogallol auto-oxidation without chiral metallohelicenes was set as the control. Each experiment was repeated three times. Error bars indicate \pm s.d. Significance was determined by one-sided analysis of variance: $***P < 0.01$. (e) Flow cytometric analysis. PC12 cells were stained by ROS probe DCFH-DA. Untreated cells acted as the control.

from A β -induced toxicity. Intracellular ROS was monitored by the probe 2',7'-dichlorofluorescein diacetate (DCFH-DA), which can be oxidized to fluorescent dichlorofluorescein (DCF). Flow cytometric analysis revealed that Δ A-treatment significantly decreased A β -mediated oxidative stress, owing to the synergistic effect of regulating A β aggregation as well as CAT-mimic and SOD-mimic activities (Fig. 3e). Moreover, microscopic images presented the shrunken, rounded, and aggregated morphologies of A β -treated cells, while Δ A substantially restored these abnormalities (Fig. S6a \dagger). 3-(4,5-Dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) assays further verified Δ A exhibited noticeable neuroprotection against A β -induced cytotoxicity (Fig. S6c \dagger).

With wild-type N2 strain as the control, we further evaluated the therapeutic potential of chiral metallohelicenes in the transgenic AD model *Caenorhabditis elegans* (*C. elegans*) CL2006, which constitutively expressed A β .²⁹ Thioflavin S (ThS) staining, ROS measurement, motility quantification, and survival curves demonstrated that A β aggregates led to paralysis phenotype, oxidative stress, behaviour defects, and shortened lifespan in the CL2006 model (Fig. 4 and S7 \dagger). Whereas the presence of Δ A reduced A β plaques, rescued A β -triggered paralysis, decreased the ROS level, and improved the motility of the CL2006 strain.



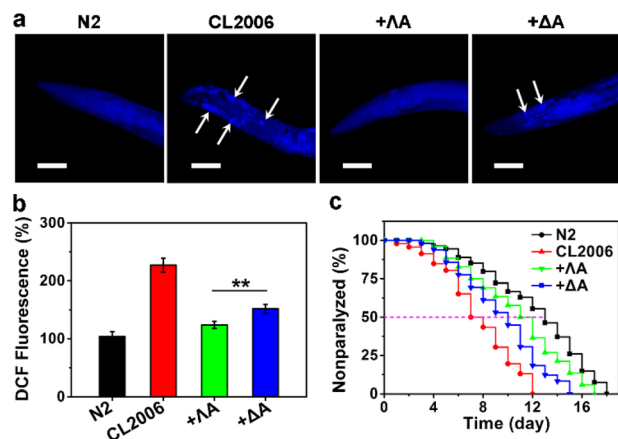


Fig. 4 Rescuing A β -associated paralysis in CL2006 strain with Δ A. (a) ThS-staining A β plaques in CL2006 strain treated with Δ A or Δ A. Arrows indicate A β plaques. Scale bars: 50 μ m. (b) ROS level in the worms monitored with DCFH-DA. All results were expressed as the mean \pm s.d. from three independent experiments. Significance was determined by one-sided analysis of variance: ** $P < 0.01$. (c) Survival curves of CL2006 strain treated with the chiral metallohelices.

Likewise, Δ A performed better than its enantiomer Δ A in relieving these A β -induced toxicities.

Improvement of cognitive function in AD model mice by Δ A

We next evaluated the performances of the metallohelices in the triple-transgenic mice model of AD (3 \times Tg-AD mice).⁴¹ The potential blood-brain barrier (BBB) permeability of the metallohelices was evaluated after the attachment of fluorescent Cyanine5 (Cy5). Fig. S8 \dagger indicated that Δ A successfully crossed the BBB and accumulated in the brain parenchyma. Besides, their potential toxicity was investigated through hematoxylin and eosin (H&E) staining and biochemistry analysis (Fig. S9 and S10 \dagger). Neither obvious tissue harm nor significant change in histopathology analysis was found, suggesting favorable biocompatibility of the metallohelices.

The effects of the metallohelices on amyloid deposition were determined *via* immunoassay. As depicted in Fig. 5a, Δ A significantly decreased the levels of A β species in the brain compared to the treatment with Δ A, further confirming the enantioselectivity towards A β . Morris water maze (MWM) assay was conducted to assess the spatial learning behavior of 3 \times Tg-AD mice after administration. Four evaluation indicators were analyzed, including the escape latency, crossing frequency, time spent in the target quadrant, and swimming path of the mice (Fig. 5b–e). After the treatment of Δ A, 3 \times Tg-AD mice took shorter paths and less time to reach the target platform and displayed a spatially oriented swimming behavior. Taken together, these results indicate that Δ A could recover impaired learning and memory in 3 \times Tg-AD mice.

Bioorthogonal PAM of Δ A to extend its biochemical applications

Inspired by protein post-translational modification (PTM), post-assembly modification (PAM) has been developed as

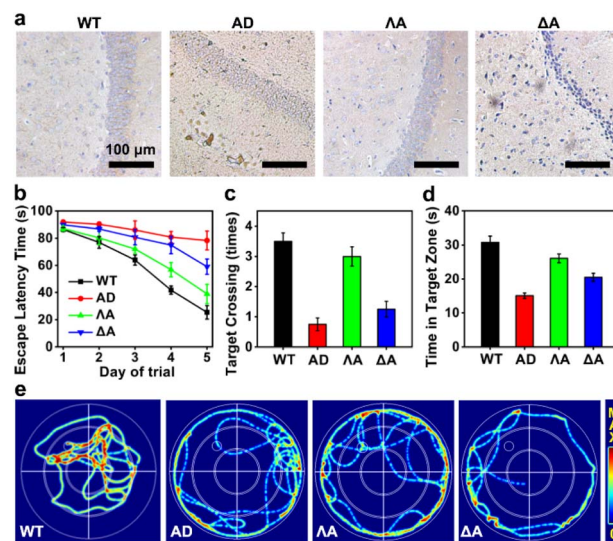


Fig. 5 Improvement of cognitive function in AD model mice with Δ A. (a) Representative images of A β staining in the hippocampus. (b) Escape latency time in the training trial. The escape latency of all groups was gradually shortened after successive training. (c) The frequency to cross the target platform. (d) Time spent in the target quadrant. (e) Representative swimming paths of mice to seek the target platform (the small circular region in the left upper corner).

a powerful tool to optimize metallosupramolecular performance by covalently introducing new functional groups, in which case the cautious pre-assembly ligand design is avoided while retaining the desired framework and properties.^{42,43} In this context, we investigated the chemoselective conjugation of bioactive moieties to the alkyne-bearing metallohelix Δ A *via* CuAAC reaction to broaden its biochemical applications. A commercial fluorescent dye Cyanine5-azide (Cy5-N₃) was used for labelling the metallohelices in both PC12 cells and *C. elegans* CL2006 strain (Fig. 6). Colocalization quantitative analysis was evaluated *via* Manders' colocalization coefficients (MCC), conducted using ImageJ software with the JACoP plugin.⁴⁴ Encouragingly, most of Δ A (about 90%) was found to be co-localized with A β antibodies in PC12 cells (Fig. 6a), and the MCC value of Δ A with A β was more significant than that of Δ A (Fig. 6c and d). This is well consistent with the above observation that Δ A dramatically outperforms Δ A in terms of specificity on targeting A β , inhibition of amyloid aggregation, and attenuation of A β -mediated toxicity. Fig. 6b and e further displayed the potential of click reaction for metallohelix visualization in the worms.

Next, we integrated various functional moieties with Δ A *via* bioorthogonal PAM to modulate multiple facets of AD, as illustrated in Fig. 7a. Previous studies have demonstrated that photoactivated ThT and its analogs can oxygenate A β and thereby decrease the aggregation property and pathogenicity of A β .^{45,46} The azide derivative of ThT (ThT-N₃) was grafted onto Δ A employing the CuAAC reaction. The obtained Δ A-ThT hybrid thus bore three copies of ThT moiety (Fig. S11a \dagger). UV-Vis absorption and CD spectra indicated that PAM did not disrupt the metallohelix architecture (Fig. S11b and c \dagger).



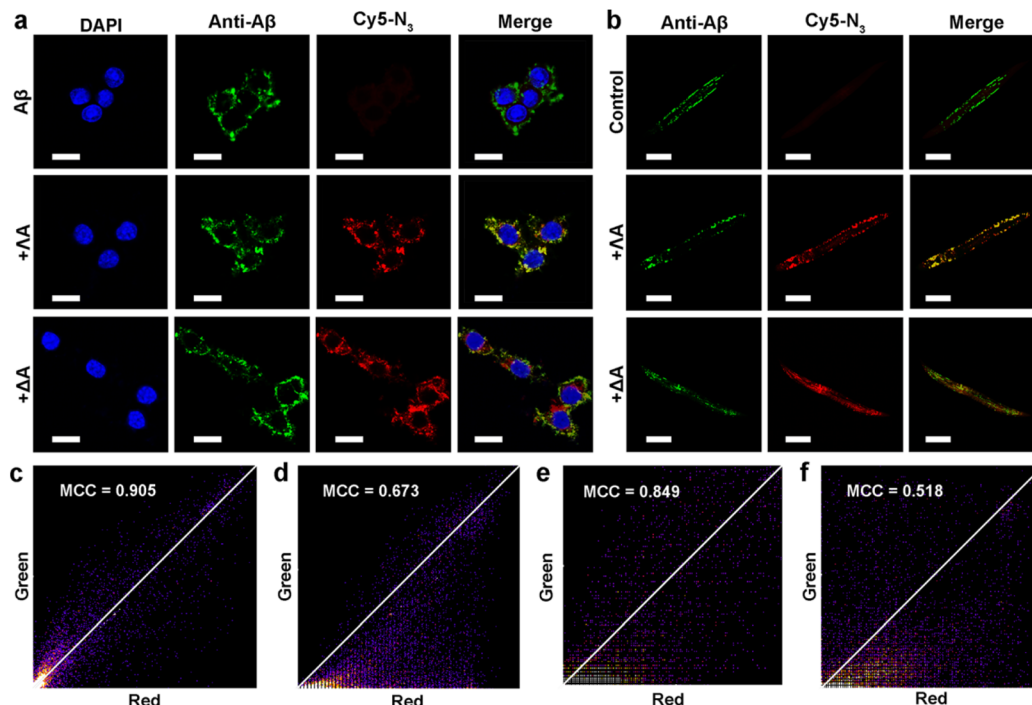


Fig. 6 Co-localization of ΔA and $A\beta$ in the PC12 cells and CL2006 strain. (a) Immunofluorescence staining of PC12 cells treated with $A\beta$ and the chiral metallohelices. $A\beta$ was labeled with $A\beta$ -specific antibody B4 (green channel), and the alkyne-bearing metallohelices ΔA and ΔA were visualized by $Cy5-N_3$ (red channel) via CuAAC reaction. The nucleus was dyed with DAPI. Scale bars: 10 μm . (b) Fluorescence images of the CL2006 strain, which were treated with the chiral metallohelices and then labeled with $Cy5-N_3$ and $A\beta$ antibody. Scale bars: 100 μm , (c) and (d) were the MCC values of $A\beta$ with $Cy5$ -labelled metallohelices in the PC12 cells, respectively, (e) and (f) were the MCC values of $A\beta$ with $Cy5$ -labelled metallohelices in the CL2006 strain, respectively.

Interestingly, photoactivated ΔA -ThT could selectively oxygenate $A\beta$ (Fig. 7b, c and S11e†), presumably because of the transient lifetime and restricted diffusion range of highly reactive singlet oxygen (1O_2) in biological systems.^{47,48} 8-Anilinoanthracene-1-sulfonate (ANS), which displays enhanced fluorescence upon binding to protein hydrophobic residues, was used for monitoring $A\beta$ fibrillation. Our observations manifested that photoactivated ΔA -ThT reduced $A\beta$ hydrophobicity (Fig. S11f†) and prohibited $A\beta$ aggregation (Fig. S11g†) through a synergistic mechanism.

Tacrine (1,2,3,4-tetrahydroacridine-9-amine, THA), the first clinically-used acetylcholinesterase (AChE) inhibitor for AD treatment, is hindered by severe adverse reactions, especially hepatic toxicity, which perhaps results from oxidative stress.^{49,50} Due to reliable anti-AChE potency, classical pharmacophore, and synthetic accessibility, THA-based scaffolds are still at the forefront of developing safer AChE inhibitors.⁵¹⁻⁵³ Recent reports revealed that antioxidants could repress THA-triggered hepatotoxicity by scavenging overproduced ROS during administration.^{54,55} In these contexts, the azide derivative of THA (THA- N_3) was also integrated with ΔA through click reaction to obtain ΔA -THA (Fig. S12†). The AChE inhibitory activity of ΔA -THA was evaluated by modifying Ellman's method.⁵¹ As shown in Fig. 7d and e and Table S2,† the hybrid metallohelix ΔA -THA exhibited a modest anti-AChE activity and diminished hepatotoxicity, which might be ascribed to a ROS-quenching

feature inherited from ΔA (Fig. S12d-f†). In addition, ΔA -THA retained the capacity to intervene $A\beta$ aggregation (Fig. S12g†).

To further assess the universality of bioorthogonal chemistry, ΔA was also conjugated with azide functionalized nanoparticles. Magnetic nanoparticles (MNPs) have been widely exploited as versatile nano-platforms for the therapeutics of neurodegenerative diseases, because of their excellent biocompatibility, BBB permeability, and attractive magnetic properties.⁵⁶⁻⁵⁹ Recently, Hyeon's group constructed multifunctional nano-assemblies (magnetite and ceria nanocomposites) for $A\beta$ clearance through selective binding and magnetic separation.⁶⁰ Inspired by that, ΔA was attached to the MNPs to form $MNP-\Delta A$ through the same click reaction (Fig. S13 and S14†). As a proof of concept, $MNP-\Delta A$ was harnessed for $A\beta_{40}$ capture, and the adsorbed $A\beta_{40}$ was measured by MS quantitative analysis with $A\beta_{42}$ as the internal standard. The amino acid sequences of $A\beta_{1-40}$ and $A\beta_{1-42}$ differ by only two amino acids, so $A\beta_{42}$ is an ideal internal standard for $A\beta_{40}$ in MS analysis (Fig. 7f). Fig. 7g showed that $MNP-\Delta A$ was able to remove $A\beta_{40}$ from the mice serum in a concentration-dependent manner. Taken together, these results indicate that, through bioorthogonal PAM, ΔA can act as a universal scaffold to integrate various bioactive moieties, thus conferring the superiority of multidirectional treatment for AD.



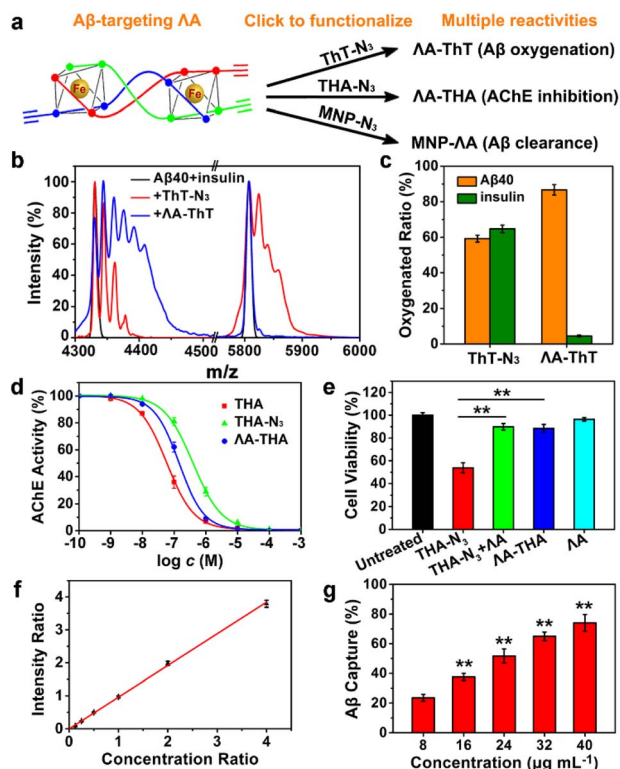


Fig. 7 Bioorthogonal PAM of ΔA to introduce multiple reactivities. (a) Scheme of PAM of clickable ΔA . (b) The mass spectra of A β and insulin oxidized by photo-activated ThT-N₃ or ΔA -ThT. Insulin, which could form amyloid fibrils under certain conditions, was used as the control to verify A β -specific oxygenation with ΔA -ThT. (c) The oxygenated ratio of A β 40 and insulin. Error bars represented \pm s.d. of three independent experiments. (d) AChE inhibition study. (e) *In vitro* hepatotoxicity evaluated with HepG2 cells. The cells were treated with THA-N₃ (120 μ M), ΔA -ThA (40 μ M), or ΔA (40 μ M) for 24 h. (f) MS intensity ratio of A β 40/A β 42 versus the concentration ratio. (g) The concentration-dependent A β 40 capture with MNP- ΔA . Each experiment was repeated three times. Error bars indicate \pm s.d. Significance was determined by one-sided analysis of variance: ** $P < 0.01$.

Conclusion

In summary, chiral metallohelix ΔA displayed enantioselectivity on targeting and regulating A β aggregation. Meanwhile, it mimicked CAT and SOD to quench overproduced ROS and maintain redox homeostasis. Such synergism enabled the impressive ability of ΔA to attenuate A β -mediated toxicity in cells and the *C. elegans* CL2006 strain, while the Δ enantiomer was far less efficient. Moreover, ΔA efficiently reduced the A β burden in the brain of 3 \times Tg-AD mice and recovered their impaired learning and memory. Beyond the inherent bioactivity, the alkyne-bearing ΔA served as a building block that allowed subsequent chemoselective conjugation *via* click reaction for extended functionalities, including biolabeling, bioconjugation, and nanopatform construction. Through fluorescent tagging, clickable ΔA was found to be well co-localized with A β *in vivo*. As representatives, several types of multi-target-directed hybrid agents against the interrelated pathological features in AD were constructed, where ΔA -ThT

reduced A β hydrophobicity and aggregation tendency; ΔA -ThA showed modest anti-AChE potential and attenuated hepatotoxicity; and MNP- ΔA conveniently captured and cleared A β . More importantly, incorporating new functional units did not interfere with the core metallohelix framework, and the final hybrid simultaneously inherited the biological activities from diverse parent frameworks. Hence, bioorthogonal chemistry promises a fresh and unique perspective for customized supramolecules with desirable functionality against multifaceted AD.

Data availability

We have included all data in the ESI Section.†

Author contributions

X. Q. and J. R. conceived the project. Z. D. performed the experimental study and drafted the manuscript. H. S. and P. S. synthesized the chiral triplex metallohelices, and P. S. assisted in revising the manuscript. C. L., and Z. L. carried out the experiments on the AD mice and analyzed the data. X. D. helped to analyze the data. All authors approved the final version. Z. D., C. L., and Z. L. contributed equally to this work.

Conflicts of interest

There are no conflicts to declare.

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References

- J. Gaugler, B. James, T. Johnson, J. Reimer, M. Solis, J. Weuve, R. F. Buckley and T. J. Hohman, *Alzheimer's Dementia*, 2022, **18**, 700–789.
- P. C. Ke, M. A. Sani, F. Ding, A. Kakinen, I. Javed, F. Separovic, T. P. Davis and R. Mezzenga, *Chem. Soc. Rev.*, 2017, **46**, 6492–6531.
- M. G. Savelieff, G. Nam, J. Kang, H. J. Lee, M. Lee and M. H. Lim, *Chem. Rev.*, 2019, **119**, 1221–1322.
- I. W. Hamley, *Chem. Rev.*, 2012, **112**, 5147–5192.
- K. P. Kepp, *Chem. Rev.*, 2012, **112**, 5193–5239.
- Z. Du, M. Li, J. Ren and X. Qu, *Acc. Chem. Res.*, 2021, **54**, 2172–2184.
- S. Lee, X. Zheng, J. Krishnamoorthy, M. G. Savelieff, H. M. Park, J. R. Brender, J. H. Kim, J. S. Derrick, A. Kochi, H. J. Lee, C. Kim, A. Ramamoorthy, M. T. Bowers and M. H. Lim, *J. Am. Chem. Soc.*, 2014, **136**, 299–310.
- M. R. Jones, E. Mathieu, C. Dyrager, S. Faissner, Z. Vaillancourt, K. J. Korshavn, M. H. Lim,



- A. Ramamoorthy, V. Wee Yong, S. Tsutsui, P. K. Stys and T. Storr, *Chem. Sci.*, 2017, **8**, 5636–5643.
- 9 T. Yang, Z. Zhu, E. Yin, Y. Wang, C. Zhang, H. Yuan, H. Zhang, S. Jin, Z. Guo and X. Wang, *Chem. Sci.*, 2019, **10**, 10149–10158.
- 10 L. Taiariol, C. Chaix, C. Farre and E. Moreau, *Chem. Rev.*, 2022, **122**, 340–384.
- 11 X. Ji, Z. Pan, B. Yu, L. K. De La Cruz, Y. Zheng, B. Ke and B. Wang, *Chem. Soc. Rev.*, 2019, **48**, 1077–1094.
- 12 M. G. Finn and V. V. Fokin, *Chem. Soc. Rev.*, 2010, **39**, 1231–1232.
- 13 J. Chen, K. Li, S. E. Bonson and S. C. Zimmerman, *J. Am. Chem. Soc.*, 2020, **142**, 13966–13973.
- 14 Y. Wang, J. Weng, J. Lin, D. Ye and Y. Zhang, *J. Am. Chem. Soc.*, 2020, **142**, 2787–2794.
- 15 M. Yang, J. Li and P. R. Chen, *Chem. Soc. Rev.*, 2014, **43**, 6511–6526.
- 16 L. Liang and D. Astruc, *Coord. Chem. Rev.*, 2011, **255**, 2933–2945.
- 17 Y. You, Q. Deng, Y. Wang, Y. Sang, G. Li, F. Pu, J. Ren and X. Qu, *Nat. Commun.*, 2022, **13**, 1459.
- 18 J. Clavadetscher, S. Hoffmann, A. Lilienkampf, L. Mackay, R. M. Yusop, S. A. Rider, J. J. Mullins and M. Bradley, *Angew. Chem., Int. Ed.*, 2016, **55**, 15662–15666.
- 19 H. Liu, Y. Qu and X. Wang, *Future Med. Chem.*, 2018, **10**, 679–701.
- 20 J. M. Suh, G. Kim, J. Kang and M. H. Lim, *Inorg. Chem.*, 2019, **58**, 8–17.
- 21 D. J. Hayne, S. Lim and P. S. Donnelly, *Chem. Soc. Rev.*, 2014, **43**, 6701–6715.
- 22 L. M. F. Gomes, J. C. Bataglioli and T. Storr, *Coord. Chem. Rev.*, 2020, **412**, 213255.
- 23 N. Rubin, E. Perugia, M. Goldschmidt, M. Fridkin and L. Addadi, *J. Am. Chem. Soc.*, 2008, **130**, 4602–4603.
- 24 J. A. Raskatov, A. R. Foley, J. M. Louis, W. M. Yau and R. Tycko, *J. Am. Chem. Soc.*, 2021, **143**, 13299–13313.
- 25 M. Tornquist and S. Linse, *Angew. Chem., Int. Ed.*, 2021, **60**, 24008–24011.
- 26 H. Song, M. Postings, P. Scott and N. J. Rogers, *Chem. Sci.*, 2021, **12**, 1620–1631.
- 27 A. D. Faulkner, R. A. Kaner, Q. M. Abdallah, G. Clarkson, D. J. Fox, P. Gurnani, S. E. Howson, R. M. Phillips, D. I. Roper, D. H. Simpson and P. Scott, *Nat. Chem.*, 2014, **6**, 797–803.
- 28 S. E. Howson, A. Bolhuis, V. Brabec, G. J. Clarkson, J. Malina, A. Rodger and P. Scott, *Nat. Chem.*, 2011, **4**, 31–36.
- 29 Y. Guan, Z. Du, N. Gao, Y. Cao, X. Wang, P. Scott, H. Song, J. Ren and X. Qu, *Sci. Adv.*, 2018, **4**, eaao6718.
- 30 M. Li, S. E. Howson, K. Dong, N. Gao, J. Ren, P. Scott and X. Qu, *J. Am. Chem. Soc.*, 2014, **136**, 11655–11663.
- 31 H. Song, N. J. Rogers, S. J. Allison, V. Brabec, H. Bridgewater, H. Kostrhunova, L. Markova, R. M. Phillips, E. C. Pinder, S. L. Shepherd, L. S. Young, J. Zajac and P. Scott, *Chem. Sci.*, 2019, **10**, 8547–8557.
- 32 J. Wang, C. Zhao, A. Zhao, M. Li, J. Ren and X. Qu, *J. Am. Chem. Soc.*, 2015, **137**, 1213–1219.
- 33 V. Armiento, A. Spanopoulou and A. Kapurniotu, *Angew. Chem., Int. Ed.*, 2020, **59**, 3372–3384.
- 34 S. Vivekanandan, J. R. Brender, S. Y. Lee and A. Ramamoorthy, *Biochem. Biophys. Res. Commun.*, 2011, **411**, 312–316.
- 35 R. Jakob-Roetne and H. Jacobsen, *Angew. Chem., Int. Ed.*, 2009, **48**, 3030–3059.
- 36 H. J. Kwon, M. Y. Cha, D. Kim, D. K. Kim, M. Soh, K. Shin, T. Hyeon and I. Mook-Jung, *ACS Nano*, 2016, **10**, 2860–2870.
- 37 Y. Zhang, Z. Wang, X. Li, L. Wang, M. Yin, L. Wang, N. Chen, C. Fan and H. Song, *Adv. Mater.*, 2016, **28**, 1387–1393.
- 38 L. M. F. Gomes, A. Mahammed, K. E. Prosser, J. R. Smith, M. A. Silverman, C. J. Walsby, Z. Gross and T. Storr, *Chem. Sci.*, 2019, **10**, 1634–1643.
- 39 X. M. Zhang, J. Tang, L. N. Wang, D. Yao, Q. Yu, F. P. Huang and H. D. Bian, *Polyhedron*, 2017, **133**, 433–440.
- 40 T. P. Ribeiro, F. L. Fonseca, M. D. de Carvalho, R. M. Godinho, F. P. de Almeida, T. D. Saint’Pierre, N. A. Rey, C. Fernandes, A. Horn, Jr. and M. D. Pereira, *Biochem. J.*, 2017, **474**, 301–315.
- 41 M. Ma, Z. Liu, N. Gao, Z. Pi, X. Du, J. Ren and X. Qu, *J. Am. Chem. Soc.*, 2020, **142**, 21702–21711.
- 42 D. A. Roberts, B. S. Pilgrim and J. R. Nitschke, *Chem. Soc. Rev.*, 2018, **47**, 626–644.
- 43 M. M. Gan, J. Q. Liu, L. Zhang, Y. Y. Wang, F. E. Hahn and Y. F. Han, *Chem. Rev.*, 2018, **118**, 9587–9641.
- 44 K. W. Dunn, M. M. Kamocka and J. H. McDonald, *Am. J. Physiol.: Cell Physiol.*, 2011, **300**, C723–C742.
- 45 Z. Du, D. Yu, X. Du, P. Scott, J. Ren and X. Qu, *Chem. Sci.*, 2019, **10**, 10343–10350.
- 46 A. Taniguchi, Y. Shimizu, K. Oisaki, Y. Sohma and M. Kanai, *Nat. Chem.*, 2016, **8**, 974–982.
- 47 P. Rajaputra, M. Bio, G. Nkepan, P. Thapa, S. Woo and Y. You, *Bioorg. Med. Chem.*, 2016, **24**, 1540–1549.
- 48 J. L. Pan and J. Shi, *Adv. Funct. Mater.*, 2014, **24**, 7318–7327.
- 49 B. Sameem, M. Saeedi, M. Mahdavi and A. Shafiee, *Eur. J. Med. Chem.*, 2017, **128**, 332–345.
- 50 D. Galimberti and E. Scarpini, *Expert Opin. Invest. Drugs*, 2016, **25**, 1181–1187.
- 51 A. Kochi, T. J. Eckroat, K. D. Green, A. S. Mayhoub, M. H. Lim and S. Garneau-Tsodikova, *Chem. Sci.*, 2013, **4**, 4137–4145.
- 52 W. G. Lewis, L. G. Green, F. Grynspan, Z. Radić, P. R. Carlier, P. Taylor, M. G. Finn and K. B. Sharpless, *Angew. Chem., Int. Ed.*, 2002, **41**, 1053–1057.
- 53 M. Ouberai, K. Brannstrom, M. Vestling, A. Olofsson, P. Dumy, S. Chierici and J. Garcia, *Org. Biomol. Chem.*, 2011, **9**, 1140–1147.
- 54 M. Benchekroun, A. Romero, J. Egea, R. Leon, P. Michalska, I. Buendia, M. L. Jimeno, D. Jun, J. Janockova, V. Sepsova, O. Soukup, O. M. Bautista-Aguilera, B. Refouvelet, O. Ouari, J. Marco-Contelles and L. Ismaili, *J. Med. Chem.*, 2016, **59**, 9967–9973.
- 55 E. Nepovimova, J. Korabecny, R. Dolezal, K. Babkova, A. Ondrejcek, D. Jun, V. Sepsova, A. Horova, M. Hrabnova, O. Soukup, N. Bukum, P. Jost, L. Muckova, J. Kassa, D. Malinak, M. Andrs and K. Kuca, *J. Med. Chem.*, 2015, **58**, 8985–9003.



- 56 Y. Li, Y. Li, W. Ji, Z. Lu, L. Liu, Y. Shi, G. Ma and X. Zhang, *J. Am. Chem. Soc.*, 2018, **140**, 4164–4171.
- 57 K. L. Viola, J. Sbarboro, R. Sureka, M. De, M. A. Bicca, J. Wang, S. Vasavada, S. Satpathy, S. Wu, H. Joshi, P. T. Velasco, K. MacRenaris, E. A. Waters, C. Lu, J. Phan, P. Lacor, P. Prasad, V. P. Dravid and W. L. Klein, *Nat. Nanotechnol.*, 2015, **10**, 91–98.
- 58 C. Wang, X. Wang, H. N. Chan, G. Liu, Z. Wang, H. W. Li and M. S. Wong, *Adv. Funct. Mater.*, 2020, **30**, 1909529.
- 59 J. Jang and C. B. Park, *Sci. Adv.*, 2022, **8**, eabn1675.
- 60 D. Kim, H. J. Kwon and T. Hyeon, *Adv. Mater.*, 2019, **31**, e1807965.

