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Naphthopyran molecular switches and their emergent mechanochemical reactivity

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Naphthopyran molecular switches undergo a ring-opening reaction upon external stimulation to generate intensely colored merocyanine dyes. Their unique modularity and synthetic accessibility afford exceptional control over their properties and stimuli-responsive behavior. Commercial applications of naphthopyrans as photoswitches in photochromic ophthalmic lenses have spurred an extensive body of work exploring naphthopyran–merocyanine structure–property relationships. The recently discovered mechanochromic behavior of naphthopyrans has led to their emergent application in the field of polymer mechanochemistry, enabling advances in the design of force-responsive materials as well as fundamental insights into mechanochemical reactivity. The structure–property relationships established in the photochemical literature serve as a convenient blueprint for the design of naphthopyran molecular force probes with precisely tuned properties. On the other hand, the mechanochemical reactivity of naphthopyran diverges in many cases from the conventional photochemical pathways, resulting in unexpected properties and opportunities for deeper understanding and innovation in polymer mechanochemistry. Here, we highlight the features of the naphthopyran scaffold that render it a powerful platform for the design of mechanochromic materials and review recent advances in naphthopyran mechanochemistry.

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1 Introduction

1.1 Polymer mechanochemistry

Since the seminal report of site-specific mechanochemical activation of covalent bonds in 2005,¹ polymer mechanochemistry has quickly grown into a burgeoning area of research.^{2,3} Mechanophores are force sensitive molecules or moieties that respond selectively to mechanical perturbation leading to a productive chemical transformation.⁴ Polymer chains are essential for transducing mechanical force to covalently linked mechanophores. On the one hand, the ability to control the force responsivity of molecules has revolutionized the design of smart materials capable of autonomously responding to mechanically dynamic environments.⁵ Recent research has also unveiled remarkable capabilities for tailoring the macroscopic mechanical properties of materials by controlling the molecular level force sensitivity of individual mechanophores within a polymer network.^{6,7} On the other hand, mechanical force can be leveraged as an external stimulus to effect desired chemical reactions with spatial and temporal precision.^{8–10} In an ever-growing collection of more than 100 different mechanophores developed to date,¹¹ judicious structural design has enabled a diverse range of force-responsive functions including catalyst

activation,^{12,13} conductivity switching,¹⁴ mechanically triggered chemiluminescence,¹⁵ and cargo release,^{16–18} among many others.^{19,20} Among these various categories, mechanochromic



Fig. 1 Mechanophores covalently incorporated into polymer chains undergo productive chemical reactions in response to mechanical force. The mechanochemical transformation of naphthopyran is illustrated.

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mechanophores, which change or produce color in response to force, have been widely developed as molecular force probes for enabling detection of stress and/or strain in polymeric materials (Fig. 1).^{20–24} Mechanical stress typically causes degradation of polymeric materials resulting, in part, from nonspecific chain scission.^{25–28} While this microscopic damage is often difficult to detect, mechanochromic mechanophores have enabled the visualization of single bond breaking events providing important mechanistic insights into fracture mechanics.²⁹ Additionally, because optical changes are straightforward to monitor and characterize, mechanochromic mechanophores have facilitated wide-ranging fundamental investigations of polymer mechanochemistry.^{30–33}

Recent research has showcased naphthopyran to be an effective and highly modular mechanophore for not only designing diverse mechanochromic materials, but also revealing unusual reactivity unique to mechanochemical activation pathways. Naphthopyrans undergo a ring-opening reaction in response to ultraviolet (UV) light or mechanical force to produce a highly colored merocyanine dye.^{34,35} Due to their synthetic accessibility and modularity coupled with the highly tunable properties of their merocyanine dyes, naphthopyrans have previously found widespread commercial application in photochromic ophthalmic lenses.^{36–38} These same qualities make naphthopyrans an ideal mechanophore scaffold for constructing mechanochromic materials, developing mechanochemical structure–property relationships, and studying fundamental aspects of mechanochemical reactivity. In this *Perspective*, we begin by briefly discussing mechanochromic mechanophores more broadly to provide context for the (mechano)chemistry of naphthopyran. Next, we describe the synthesis of naphthopyrans and review their photochemistry with an emphasis on the extensive and generalizable structure–property relationships of their merocyanine dyes. Finally, we discuss the mechanochemical reactivity of naphthopyrans, highlighting notable recent discoveries that demonstrate their unique potential as an emergent class of diverse molecular force probes.

1.2 Mechanochromic mechanophores

Mechanochromic mechanophores function as molecular force sensors, signaling critical stress and strain in materials through a convenient visual output. The color changes that accompany activation of a mechanochromic mechanophore identify locations in a material that have experienced a critical level of force and may serve as a warning for impending catastrophic failure.³⁹ Alternatively, these mechanophores have been leveraged to provide mechanistic insights into the chain scission events that typically characterize degradation of polymeric materials subjected to mechanical loads.^{29,40,41} The development of mechanophores that exhibit force dependent changes in color, or multicolor mechanochromism, represents a grand challenge for being able to not only locate, but remotely quantify the stress state of a material through simple visual analysis. We will revisit the concept of force dependent multicolor mechanochromism below, including progress that has been

made toward achieving this goal enabled by naphthopyran mechanochemistry. This section is not intended to be comprehensive, but rather provide context and highlight the breadth of structures and strategies that exist for covalent mechanophores that complement naphthopyran mechanochemistry. For further reading, we refer to several recently published reviews.^{20–24}

A wide range of mechanochromic mechanophores have been developed that rely on many different reaction mechanisms and produce a variety of dyes, each with their own unique properties and limitations. The Otsuka group has developed a series of scissile mechanophores that generate a pair of stable, colored free radical species following force-induced bond homolysis,^{42–47} as exemplified by diarylbibenzothiophenone (DABBT) illustrated in Scheme 1.⁴⁵ Mechanical dissociation is thermally reversible and the lifetime of the radical products depends strongly on the environment.^{42,44,47,48} In some cases, only partial recombination occurs.⁴⁹ Polymers that display colors spanning the visible spectrum can be accessed by blending a small library of structurally distinct radical-type mechanophores that individually produce pink, green, or blue colored radicals upon mechanochemical activation.⁴⁵ Mechanochemical retro-cycloaddition reactions, and in particular retro-[4 + 2] cycloadditions, have also been leveraged to design mechanochromic mechanophores useful for stress sensing. For example, Göstl and Sijbesma have developed mechanophores that produce π -extended anthracene derivatives through a mechanochemical retro-Diels–Alder reaction of anthracene–maleimide adducts (see Scheme 1).⁵⁰ The extended π system of the anthracene products was cleverly employed to both enhance the fluorescence quantum yield and shift the absorption into the visible range. The color of the anthracene products can be adjusted by substitution of the extended π system with various arenes,^{41,51} although like the radical-type mechanophores, this requires the synthesis of



Scheme 1 Examples of mechanochromic mechanophores that generate color upon mechanochemical activation *via* homolysis (top) or a retro-Diels–Alder reaction (bottom).





Scheme 2 Poly(ladderene) undergoes a force-induced unzipping reaction to produce conjugated polyacetylene.

discrete congeners and not simply peripheral substitution. Another notable and unique class of mechanophores has been pioneered by Xia and coworkers with the development of poly-ladderene mechanochemistry (Scheme 2).^{14,52,53} Upon mechanochemical activation, poly-ladderene unzips through a multistep radical process transforming the colorless insulating polymer into semiconducting polyacetylene with a deep purple color.¹⁴ While polyacetylene typically suffers from oxidative degradation, a fluorinated ladderene derivative was recently demonstrated to provide a significantly more stable conjugated product after mechanochemical activation.⁵³

Spiropyran is arguably the most ubiquitous mechanochromic mechanophore and also one of the most reactive (Scheme 3).^{54,55} One of the earliest papers in the field from Moore and coworkers in 2007 described the mechanochemical 6π electrocyclic ring-opening reaction of spiropyran leading to the generation of a highly colored merocyanine dye.⁵⁶ Mechanochemical activation was achieved in solution upon ultrasonication of linear polymers incorporating spiropyran near the chain center where cavitation-induced elongational forces are maximized.^{3,57} In 2009, Sottos and coworkers demonstrated the mechanochemical activation of spiropyran in polymeric materials upon uniaxial tension and compression, establishing a new modality for stress sensing in polymers and opening the door to an entire domain of research innovation.³⁹ Since these initial breakthroughs, many additional mechanochromic mechanophores—including naphthopyran—have been developed that rely on a mechanochemical ring-opening reaction to produce a highly colored conjugated dye such as spirothiopyran,^{58,59} rhodamine,^{60,61} and oxazine^{62,63} (Scheme 3). Unlike some of the other mechanophores described earlier, these mechanophores are typically non-scissile and thermally or photochemically reversible, enabling multiple cycles of mechanical activation in elastomeric materials. All of these mechanophores are also prominent photochromic compounds, establishing an interesting parallel between their photochemical and mechanochemical reactivity for which a mechanistic basis remains to be better understood. Unlike the quinoidal merocyanines typically derived from naphthopyrans, the mechanophores shown in Scheme 3 produce dyes that are usually zwitterionic. Furthermore, in addition to UV light and mechanical force, their ring-opening reactions are triggered by



Scheme 3 Examples of mechanochromic mechanophores that undergo ring-opening reactions upon mechanochemical activation to produce thermally reversible colored dyes.

various other stimuli including heat, metal ions, electrical potential, pH changes, or polar environments, reducing the specificity of their stimuli-responsivity that can present practical challenges.^{64–66} For spiropyran, while synthetic modification can often prove challenging, certain substitution patterns have been shown to enhance selectivity for mechanochemical activation, but at the same time rendering a merocyanine product that is exceptionally thermally transient.⁶⁷

Although structurally related, naphthopyrans offer excellent fatigue resistance, stimulus specificity, synthetic modularity, and an extensive structure–property range, distinguishing them from spiropyran and related spiroheterocyclic mechanophores.^{36,37} Naphthopyrans and their corresponding merocyanine dyes typically exhibit a high degree of chemical stability as well as selective reactivity to mechanical force in the absence of UV light, making it an excellent scaffold for force sensing applications. Dye formation from the molecular switches shown in Scheme 3 is facilitated by a conjugated amine, whereby the pair of electrons on the nitrogen atom assists the ring-opening reaction and extends the length of the resulting conjugated π system.^{64,68} In contrast, naphthopyran-



derived merocyanines are stabilized by extended conjugation afforded by the naphthalene core, which retains aromaticity in the quinoidal form.^{34,38,69} This feature significantly reduces the susceptibility of naphthopyran to spontaneous coloration, for example in polar environments, without sacrificing the stability of the merocyanine dye.⁷⁰ We discuss the properties of naphthopyrans in more detail in the following sections.

1.3 Naphthopyran molecular switches

Naphthopyrans, or benzo-annulated benzopyrans (chromenes), are a class of molecular switches that undergo a reversible 6π electrocyclic ring-opening reaction upon photoirradiation with UV light or under extrinsic force, resulting in conversion of the colorless naphthopyran into a colored merocyanine dye (Scheme 4). There are three constitutional isomers of naphthopyran with differing position and orientation of the pyran ring about the naphthalene core (Fig. 2).⁶⁹ However, only the angular *2H*-naphtho[1,2-*b*]pyran and *3H*-naphtho[2,1-*b*]pyran regioisomers display appreciable photochromic activity near room temperature. As a result, the linear *2H*-naphthopyran congener has received comparatively little attention in the literature. We will typically refer to the two angular variants by their simpler familial names of *2H*-naphthopyrans and *3H*-naphthopyrans. As mentioned above, naphthopyran photo-switches have been widely developed commercially for use as reversible coloring agents in plastic ophthalmic lenses,^{36,37,71} with other notable applications in textiles,⁷² anti-counterfeiting measures,⁷³ and cosmetics.⁷⁴ Naphthopyrans have also been studied as conductivity switches,^{75–81} logic gates,^{82–84} molecular actuators,⁸⁵ chemodosimeters,⁸⁶ solar cell sensitizers,⁸⁷ and artificial models of photo-excitable neurons.⁸⁸

The photochemistry of naphthopyran has been extensively studied since the photoinduced ring-opening reaction was first mentioned by Becker and Michl in 1966.³⁴ Exactly 50 years later in 2016,³⁵ it was discovered that the ring-opening reaction of naphthopyran can be accomplished with mechanical force applied across the labile C–O pyran bond. Naphthopyran has since emerged as a highly accessible and versatile mechanophore scaffold aided by the extensive structure–property relationships developed in the photochemical arena, which serve as a convenient and useful blueprint for the design of molecular force probes with an impressively broad range of properties. In a relatively short timeframe, research has illustrated that naphthopyran mechanophores can be scissile or non-scissile

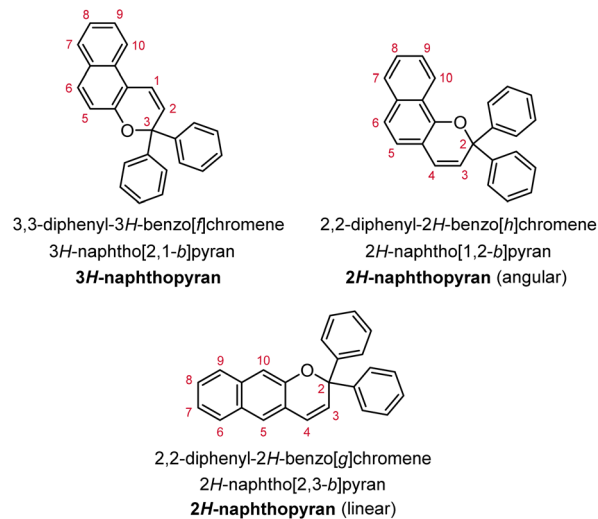


Fig. 2 Molecular scaffolds depicting the three diaryl naphthopyran regioisomers with atomic positions identified. The different naming conventions are illustrated for each structure.

and generate merocyanine dyes with precisely tunable colors and lifetimes ranging from highly transient to permanent. Beyond the development of materials with a binary mechanochromic response, studies of naphthopyran have deepened insights into fundamental aspects of mechanochemical reactivity, elucidated reaction pathways that diverge significantly from the photochemical transformations, and expanded the mechanochemical toolbox toward the design of materials with multicolor mechanochemical behavior.

2 Synthesis of naphthopyrans

Naphthopyrans benefit from modular construction through straightforward synthetic routes in which structural diversity can be built relatively quickly and easily. Facile modification of the naphthopyran scaffold enables access to merocyanine dyes that display a wide range of properties with highly tunable color and lifetimes. Unless mentioned otherwise, all of the synthetic methods described below are applicable to the formation of both *2H*-naphthopyrans and *3H*-naphthopyrans.

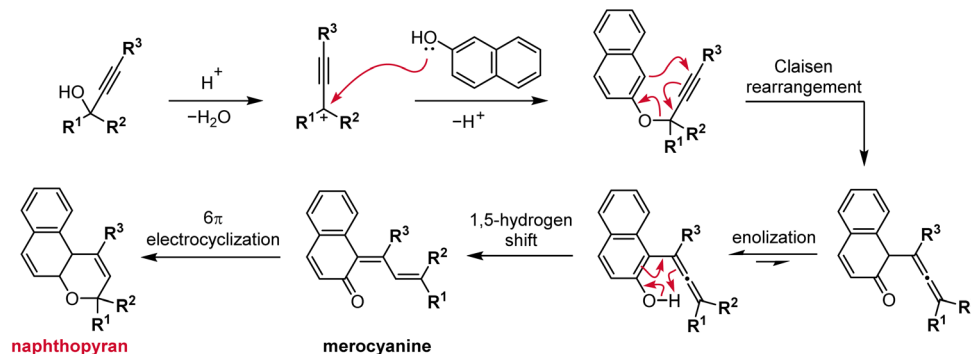
2.1 Acid-catalyzed reactions of propargyl alcohols and naphthols

The most straightforward and robust method of synthesizing naphthopyrans relies on the thermal rearrangement of naphthyl propargyl ethers (Scheme 5).⁸⁹ These substrates are typically generated *in situ* via the condensation of a naphthol with a propargyl alcohol under acidic conditions. An ideal catalyst for this transformation is *para*-toluenesulfonic acid (TsOH) due to its solubility in organic solvents.^{75,76,90–95} Heterogeneous catalysis using acidic alumina is also exceedingly common, enabling efficient syntheses on large scale.^{35,96–101} The mechanism follows initial protonation of the propargyl alcohol and loss of water to generate a propargyl cation that is attacked by the nucleophilic naphthol. Next, a Claisen rearrangement and



Scheme 4 Ring-opening reaction of a naphthopyran molecular switch based on the *3H*-naphtho[2,1-*b*]pyran skeleton to reversibly generate a colored merocyanine dye.



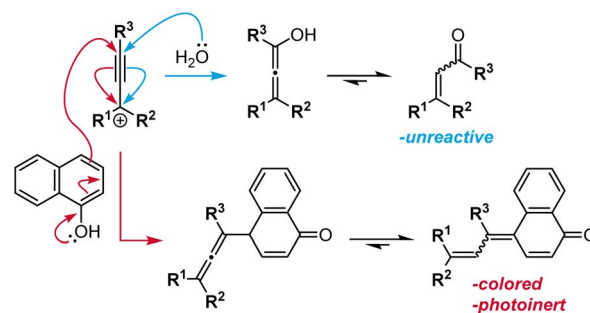


Scheme 5 Typical synthesis of naphthopyrans *via* acid-catalyzed condensation of naphthols and diaryl propargyl alcohols that proceeds through a multistep pathway. The synthesis of a 3*H*-naphtho[2,1-*b*]pyran is illustrated.

subsequent enolization generates the intermediate allenyl naphthol species, which undergoes a 1,5-hydrogen shift to produce the merocyanine form of the naphthopyran. Finally, a thermal 6π electrocyclicization closes the ring to form the colorless naphthopyran.¹⁰² Notably, a wide range of substituents on both the naphthol and propargyl alcohol coupling partners are tolerated due to the relatively mild conditions. However, because the reaction mechanism proceeds *via* the intermediacy of the merocyanine, substituents that strongly stabilize the merocyanine form can decrease the yield of the naphthopyran.¹⁰³ In our experience, photoirradiation with visible light helps to facilitate the final ring-closure in some cases.

The propargyl alcohol and naphthol building blocks are either commercially abundant, or readily prepared and/or modified using robust chemistry, which reinforces the utility of this synthetic approach. Many inexpensive 1-naphthols and 2-naphthols are commercially available. The propargyl alcohol coupling partners can often be acquired from commercial sources or efficiently synthesized by treating the appropriate benzophenone with an alkali metal acetylide.^{75,99,104,105} More specialized benzophenones can also frequently be prepared in one step by Friedel–Crafts acylation of an arene with an acyl halide. The modularity of this strategy has enabled the construction of diverse libraries of naphthopyran derivatives displaying a wide range of substitution patterns, both on the naphthalene core and on the peripheral aryl groups, from a relatively small collection of simple yet highly customizable building blocks.

While generally robust, this synthetic route can involve the formation of undesired side products. If naphthol condensation does not occur rapidly after acidic dehydration, interception of the propargyl carbocation by adventitious water results in Meyer–Schuster rearrangement to produce a terminal α,β -unsaturated carbonyl compound (Scheme 6, blue arrows).¹⁰⁶ Electron-rich diaryl propargyl alcohols form relatively stable carbocations and are particularly susceptible to this rearrangement side reaction.⁹⁹ Additionally, the synthesis of 2*H*-naphthopyrans is complicated by the nucleophilicity at the 4-position *para* to the hydroxyl group. Nucleophilic attack at this carbon center instead of the oxygen produces a deeply colored dye that exhibits no photoswitching activity (Scheme 6, red



Scheme 6 Side reactions encountered in the synthesis of naphthopyrans *via* the acid-catalyzed route including the Meyer–Schuster rearrangement (top pathway, blue arrows) and conjugate addition to the propargyl cation at the 4-position of the 1-naphthol (bottom pathway, red arrows).

arrows).^{99,107} The yield of this side product increases when particularly electron-rich propargyl alcohols are employed or if the nucleophilicity of the naphthol is diminished due to steric or electronic effects.

2.1.1 Improved acid-catalyzed synthesis of naphthopyrans.

An optimized one pot procedure that minimizes the side reactions described above was developed by Zhao and Carreira in 2003 that employs pyridinium *para*-toluenesulfonate (PPTS) as a milder acid alternative to TsOH (Scheme 7).¹⁰⁸ High-boiling, polar aprotic 1,2-dichloroethane was found to be an ideal solvent for facilitating formation of the carbocation intermediate. Importantly, the inclusion of trimethyl orthoformate as a dehydrating agent significantly increased the yield of the desired product while also shortening reaction times. Notably, the synthesis of bis-naphthopyrans that are traditionally



Scheme 7 Reaction conditions for the acid-catalyzed synthesis of naphthopyrans developed by Zhao and Carreira.¹⁰⁸

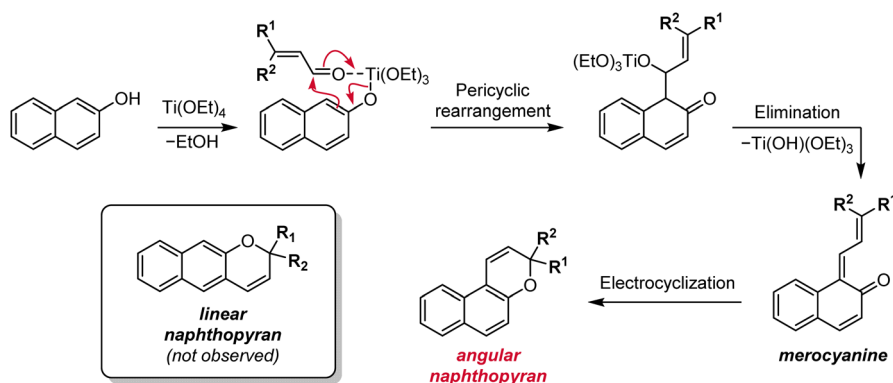


accessed in low yields¹⁰⁹ progressed cleanly enough that simple filtration provided pure product. Generally, these conditions produce naphthopyrans in excellent yields in 1–3 h and tolerate a variety of substitution patterns on both the aryl groups and naphthalene core. The high yields, clean conversion, and fast reaction times make this method superior to traditional methods in nearly every regard.^{97,103,110–120} We note, however, that the dehydrating conditions will not tolerate all functional groups, and in particular, primary alcohols are incompatible with this synthetic method.

2.2 Alternative synthetic routes

Other methods have historically been used to synthesize substituted naphthopyrans, including Grignard addition to unsubstituted benzocoumarins,^{121–124} reduction of substituted naphthopyranones,^{124,125} and addition of a lithiated naphthol to cinnamaldehyde derivatives.^{124,126,127} However, these routes involve the use of harsh reagents, which limit the scope of accessible naphthopyrans, or suffer from low yields and have largely been rendered obsolete by improved protocols like the ones discussed above. Additionally, the starting materials are often not commercially available and require multistep syntheses. Although there may be niche applications for which these alternative methods prove useful, they can generally be regarded as inferior to the more modular, functional group tolerant, and straightforward synthetic approaches discussed above.

One notable exception is a protocol involving the treatment of a naphthol with a substituted cinnamaldehyde derivative in the presence of titanium tetraethoxide, affording the naphthopyran in one operational step (Scheme 8).^{90,96,124,128} The mechanism involves displacement of one of the ethoxide ligands on titanium by the naphthol and coordination of the cinnamaldehyde, which is followed by a pericyclic rearrangement and elimination of the titanium alkoxide to produce the merocyanine. Thermal electrocyclicization then produces the naphthopyran similar to the mechanism in the acid-catalyzed route. This reaction is remarkably robust with broad functional group tolerance and has proven to be more efficient than the acid-catalyzed conditions for some heterocycle-fused naphthopyrans and related chromenes.^{90,129}



Scheme 8 Alternative titanium-mediated synthesis of angular naphthopyrans from naphthols and substituted cinnamaldehydes.



Scheme 9 Solvent-free reaction conditions for the synthesis of naphthopyrans *via* grinding or ball-milling.

Other methods for synthesizing naphthopyrans have also been developed including those that use environmentally benign solvents like water or solid-state reactions that eliminate solvents altogether. Naphthopyrans were attained in good yields by heating an aqueous mixture of propargyl alcohol and naphthol starting materials with catalytic β -cyclodextrin hydrate.¹³⁰ In a solid state protocol, grinding a mixture of propargyl alcohol, naphthol, TsOH, and silica gel at room temperature with a mortar and pestle produced the corresponding naphthopyrans, albeit in poor to moderate yields (Scheme 9).¹⁰⁹ Substantial improvements were achieved using ball-milling and catalytic InCl₃ leading to naphthopyran formation in excellent yields with minimal side products.¹³¹ Notably, the exclusion of a Brønsted acid catalyst lends this method to the construction of naphthopyrans containing acid-sensitive functional groups. The authors also demonstrated the solid state synthesis of a wide variety of propargyl alcohol starting materials in very good yields by ball-milling a terminal alkyne and a benzophenone in the presence of KOH and 18-crown-6, circumventing the use of potentially hazardous pyrophoric alkali metal acetylide species.¹³¹

2.3 Post-synthetic modification of naphthopyrans

Naphthopyrans generally tolerate a wide range of reaction conditions that include acids or bases,^{94,132,133} strong nucleophiles^{94,134} or electrophiles,^{134,135} organolithium species,¹³³ and reducing agents.¹³⁶ Thus, naphthopyran libraries can also be constructed *via* a divergent synthetic strategy in which a naphthopyran substrate is derivatized at a later stage in the synthesis. The most versatile of these strategies are those in



which halide or triflate groups on the naphthopyran core or peripheral aryl rings serve as functional handles for cross-coupling reactions. Notably, Suzuki,^{79,94,137,138} Sonogashira,¹³⁹ and Buchwald–Hartwig^{87,140} couplings have been used to install aryl/vinyl, alkynyl, or amine substituents, respectively. Further diversity is afforded by copper catalyzed Ullman-type¹³² or Grignard coupling.⁹⁴ Appropriate cross-coupling handles have also been installed *via* selective halogenation¹³⁷ or lithium–halogen exchange reactions.¹³³

The robustness of naphthopyran also enables its covalent incorporation into polymers necessary for the construction of mechanochromic materials using a wide variety of synthetic approaches. Naphthopyrans are generally tolerant of organic radicals, conveniently allowing naphthopyran-based initiators and monomers to be used in the synthesis of polymers *via* common radical polymerization techniques.^{33,134,135,138,141–144} Additionally, naphthopyrans have been covalently incorporated into siloxane networks *via* Pt-catalyzed hydrosilylation reactions,^{9,35,143–146} and in organic/inorganic composites *via* a sol–gel method using a Sn catalyst.¹³⁴ Cu-catalyzed azide–alkyne cycloaddition (CuAAC) reactions have also been successfully employed for the preparation of naphthopyran-containing polymers.^{134,147}

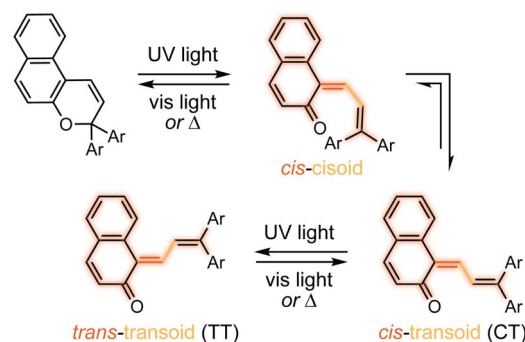
3 Structure–activity relationships for naphthopyran-derived merocyanine dyes

Becker and Michl first alluded to the photochromism of naphthopyrans in 1966 in a seminal report on photochromic benzopyrans (chromenes),³⁴ with formal structural and experimental details for naphthopyran provided in a subsequent patent.¹⁴⁸ Whereas the benzene ring of benzopyrans undergoes complete loss of aromaticity in its merocyanine form, the naphthalene core maintains partial aromaticity in the ring-opened state of the analogous naphthopyran. This additional stabilization enables naphthopyrans to produce comparatively intense coloration with significantly improved fatigue resistance.^{34,38,69} In contrast to closely related spiropyran,¹⁴⁹ naphthopyrans exhibit little to no thermochromism^{37,69} because the thermal barrier of the ring-opening reaction^{96,150} is significantly greater than that of the reverse ring-closing reaction.^{151,152} These properties, combined with its synthetic accessibility and modularity, have made naphthopyran a particularly robust and practical photoswitch for a wide range of applications. Given its industrial relevance, studies of naphthopyran photochemistry are abundant with numerous detailed investigations of merocyanine structure–property relationships toward the development of naphthopyrans as commercial photochromic switches.^{36,37,71} Since the same merocyanine dyes are typically produced upon mechanochemical activation of naphthopyran, the structure–property relationships established in the photochemical literature serve as an invaluable resource for designing mechanochromic mechanophores with predictable and highly tunable absorption and reversion properties.^{143,145} Furthermore, mechanistic studies of

photochemically and thermally mediated naphthopyran ring-opening reactions (*vide infra*) are convenient points of comparison to the closely related, but also often divergent mechanically mediated processes, deepening fundamental insights into the nature of mechanical force-driven chemical transformations. In the next section, we will highlight some key developments in naphthopyran photochemistry and prominent structure–property relationships that have proved useful in the design of mechanochromic mechanophores. More exhaustive reviews dedicated to the topic are available for the interested reader.^{36–38,69,153}

3.1 Photochemical ring-opening and alkene isomerization reactions

The photochemical 6π electrocyclic ring-opening reaction of naphthopyran and possible olefin photoisomerization reactions generate several different stereoisomers of the merocyanine dye (Scheme 10).^{69,151,152,154–156} Rotation about the C–C single bond connecting the two newly formed alkenes from a cisoid to transoid conformation occurs rapidly to reduce steric interactions between the aryl substituents and the naphthalenone core. While the *cis*-cisoid form is a frequently predicted intermediate in computational studies^{150,157,158} and has been observed by transient spectroscopy for benzopyran,¹⁵⁹ this species has not been experimentally observed for naphthopyrans.^{150,156} This experimental observation is consistent with the low barrier for bond rotation of ~ 5 kcal mol^{−1} predicted computationally.¹⁵⁰ The resulting *cis*-transoid (CT) species, wherein the exocyclic alkene is in the *cis* configuration, can revert back to the colorless naphthopyran thermally or with visible light irradiation *via* the *cis*-cisoid intermediate. Alternatively, *cis*-to-*trans* isomerization of this exocyclic alkene occurs with further UV irradiation, converting the CT form to the much more thermally stable *trans*-transoid (TT) merocyanine. While thermal *trans*-to-*cis* isomerization does occur, photochemical isomerization of the exocyclic alkene with visible light is significantly more efficient.^{93,150,152,155,160} We note that although the *cis*-transoid (CT) and *trans*-transoid (TT) or similar nomenclature is used broadly in the naphthopyran literature, later we will frequently refer to these merocyanine isomers



Scheme 10 Exemplary photochemical ring-opening reaction of a 3H-naphthopyran and alkene isomerization to generate principal *cis*-transoid (CT) and *trans*-transoid (TT) isomeric merocyanine products.



simply by the configuration of the exocyclic alkene as being either *cis* or *trans*, respectively, because the *cisoid* and *transoid* descriptors are of little practical relevance. Evidence that naphthopyran derived merocyanine dyes assume a charge-neutral quinoidal form in the ground state is derived from NMR data,^{103,154,161} X-ray crystallography (for especially thermally stable species),^{103,117,162} and solvatochromic behavior.^{90,151,162–164} These merocyanines exhibit positive solvatochromism, or bathochromic shifts in λ_{\max} with increasing solvent polarity, reflecting a weakly polar ground state with minimal charge delocalization that is characteristic of a quinoidal structure.^{160,163,165}

Merocyanine stereoisomers with different stereochemical configuration of the exocyclic alkene (*i.e.*, CT and TT forms) exhibit modest differences in absorption with values of λ_{\max} typically within ~ 10 nm.^{103,116,120,151,166} However, comparisons of the extinction coefficients between the CT and TT forms are often complicated by short lifetimes that render isolation difficult. In the case of 3,3-diphenyl-3*H*-naphtho[2,1-*b*]pyran, the extinction coefficients of the CT and TT isomers were determined to be 18 100 and 16 900 M⁻¹ cm⁻¹ in toluene, respectively, after careful isolation of each species by low temperature chromatography.¹¹⁶ Extinction coefficients have been determined in other cases based on structural isolation *via* low temperature chromatography^{120,167} or with especially stable merocyanines,^{103,117,162,167} or estimated by fitting experimental absorption profiles to kinetic models^{117,152,168–171} or with computational simulations.^{103,117,172} Notably, determination of extinction coefficients for the TT stereoisomer is more prone to error because this isomer typically comprises only a small fraction of the photochemically generated merocyanine mixture.¹⁵² Given the challenges in characterizing individual stereoisomers, these species are often assumed to have a similar extinction coefficient to simplify kinetic analysis.^{80,94,118,162,173} To our knowledge, the photophysical changes between stereoisomers with varying configuration at the distal olefin for merocyanines containing dissimilar peripheral aryl groups have not been determined, but these are likely even less pronounced than the relatively subtle differences in absorption arising from isomerization of the exocyclic alkene.

Careful analysis of reaction kinetics using UV-vis absorption spectroscopy has elucidated information about the photochemical reaction mechanism, as well as the quantum yields and extinction coefficients for naphthopyrans and the corresponding merocyanines. However, photochemical activation produces complex mixtures of several isomeric species,^{150,174} including several with overlapping absorption spectra. Analytical¹⁷⁵ or even numerical^{80,115,133,171,176} solutions to such kinetic problems are complicated, and often assumptions must be introduced to simplify analysis.^{115,171,176} Mechanistic studies of naphthopyran photochemistry are dramatically simplified by monitoring reaction progress using NMR spectroscopy, which allows the evolution of each isomeric species to be monitored separately. Particularly elegant studies have employed fluorinated naphthopyrans to track each individual species using ¹⁹F NMR spectroscopy with excellent resolution.^{115,133,154,174,177–180} These studies support that UV photoirradiation of typical

naphthopyrans results first in the nearly instantaneous formation of the CT merocyanine isomer; additional irradiation with UV light causes *cis*-to-*trans* isomerization of the exocyclic alkene to generate the TT merocyanine isomer.^{81,181}

Greater mechanistic detail for the photochemical ring-opening reaction of naphthopyran has been revealed using transient spectroscopy techniques and supported by computation. Here, we briefly discuss findings for the simple photo-switch 3,3-diphenyl-3*H*-naphtho[2,1-*b*]pyran (Ar = Ph in Scheme 10). It should be noted that while changes in solvent and temperature alter the lifetimes of the individual states,¹⁶⁰ the overall photochemical pathway is largely unaffected by solvent polarity,^{151,156} which is a notable distinction from closely related spiropyran.^{68,70,149} Photochemical ring-opening reactions of naphthopyran proceed *via* a singlet excited state that has a picosecond lifetime and is structurally similar to the ring-closed naphthopyran (Fig. 3).^{150,156,182–185} Transition to the ground state proceeds over an insignificant activation barrier (~ 1 kcal mol⁻¹) and through a conical intersection, at which point scission of the pyran C–O bond occurs to produce the CT merocyanine isomer in a concerted process.^{150,156} The essentially barrierless excited state reaction proceeds so rapidly that it competes even with vibrational relaxation,^{156,158,186–189} resulting in mode-dependent behavior wherein the reaction proceeds from any of several distinct vibronic states accessed from excitation sources of distinct wavelengths.¹⁹⁰ While the quantum yield of the ring-opening reaction is typically high (~ 0.8),^{150–152} a fraction of the excited naphthopyran species undergoes internal conversion back to the ground state.¹⁵⁶

The *cis*-to-*trans* isomerization of the exocyclic alkene to generate the TT isomer also occurs through a singlet excited state and transition to the ground state through a conical intersection, but only after a second photon absorption, rather than directly from the excited state of naphthopyran.^{150,156,166} Consistent with observations from NMR spectroscopy noted above, this assertion is further supported by theoretical modeling as well as flash photolysis experiments demonstrating minimal formation of the TT merocyanine isomer after brief exposure to UV light.^{80,155,191} Theoretical modeling suggests that isomerization proceeds through either a single twist about the



Fig. 3 Representative reaction pathway for the photochemical ring-opening reaction of naphthopyran.



exocyclic olefin, or through a “bicycle pedal” motion consisting of concerted rotation about both olefins of the merocyanine bridge.¹⁹² While the single twist pathway is favored for 3,3-diphenyl-3*H*-naphtho[2,1-*b*]pyran, the concerted dual-isomerization pathway is promoted in polar solvents or with certain electronic substituents. Excitation of the CT isomer can also lead to photoenolization to form the relatively unstable allenyl naphthol species.^{150,166,174,179} Notably, however, internal conversion to the ground state is extremely rapid for the excited CT merocyanine isomer, resulting in inefficient photoisomerization to either the TT isomer or the allene, consistent with the typically limited accumulation of these species observed experimentally.¹⁶⁶

Whereas spectroscopic and computational investigations^{150,156,158} support a rapid and concerted electrocyclic ring-opening pathway upon photochemical activation of naphthopyran, there is evidence that this mechanism is not conserved when other stimuli are used. While inefficient, the thermal ring-opening reaction has been experimentally investigated *via* the racemization of chiral naphthopyrans.^{96,193} Both solvent and substituent effects on the thermal activation barrier indicate a polar, heterolytic ring-opening mechanism in direct contrast to the transient absorption studies performed on the photochemical transformations of naphthopyran¹⁵⁶ and closely related benzopyran,¹⁵⁹ which again reveal no evidence of a zwitterionic intermediate.

3.2 Thermal reversion of naphthopyran-derived merocyanine dyes

Thermal transformations play an important role in naphthopyran–merocyanine equilibria. As noted above, the ring-opening reaction of naphthopyran is thermally accessible, but the process is extremely inefficient. Experimental⁹⁶ and computational^{150,157,194–196} studies indicate that the thermal activation barrier for ring opening is $\sim 25\text{--}30$ kcal mol⁻¹. By comparison, the barrier to ring closure from the CT merocyanine isomer has been estimated to be in the range 11–21 kcal mol⁻¹.^{150–152,195} Thus, thermal reversion of the merocyanine dye is comparatively rapid such that the accumulation of colored products (*i.e.* thermochromism) is not typically observed. Therefore, electrocyclization of the merocyanine¹⁵¹ is the most kinetically relevant thermally mediated process in the context of naphthopyran chemistry. The rate of thermal reversion depends strongly on the regiochemistry and geometry of the merocyanine, which will be the focus of this section.

As discussed earlier, the stereochemical configuration of the exocyclic alkene significantly impacts the kinetic stability of the merocyanine and therefore its tendency to revert to the ring-closed naphthopyran form. *Cis* and *trans* merocyanines exhibit similar thermodynamic stability, $\sim 10\text{--}14$ kcal mol⁻¹ higher in energy than the naphthopyran form.^{117,150} However, electrocyclization occurs exclusively from the *cis* merocyanine isomers, necessitating *trans-to-cis* isomerization of the exocyclic alkene prior to ring-closure.^{152,155,160,181} While *trans-to-cis* isomerization of the exocyclic alkene is possible thermally, this process is impeded by a substantial barrier of ~ 28 kcal mol⁻¹

(*ref.* 150) and thus proceeds significantly slower than the thermal ring-closing reaction of *cis* merocyanines.^{80,93,118,133,151,152,154,155,197} In some cases, *trans* merocyanines have even proven stable enough to isolate and characterize under ambient conditions.^{93,103,117,160,162,178} Isomerization of the exocyclic alkene occurs much more efficiently upon irradiation with visible light as discussed earlier. Therefore, while naphthopyrans are generally considered to be T-type (*i.e.* thermally reversible) photoswitches,^{36,37,71} P-type (*i.e.* photochemically reversible) behavior is observed for some species if extended exposure to UV light generates an appreciable amount of thermally persistent *trans* merocyanine. Thermal *cis-to-trans* isomerization of the exocyclic olefin is uncommon and has only been observed in exceptional cases.¹³³

Reversion kinetics for mixtures of merocyanine stereoisomers are reliably determined using multiple methods, but most often by fitting time-dependent absorption data to a biexponential decay function.^{94,112,114,115,118,152,154} The smaller rate constant obtained is attributed to conversion of the *trans* stereoisomer. Thus, if little to no *trans* merocyanine isomer is present, fitting with a monoexponential decay function is sufficient. The enthalpy and entropy of activation for isomerization and ring-closure for several merocyanine derivatives has also been determined by Eyring-type analysis of reversion kinetics using both UV-vis absorption and NMR spectroscopy methods.^{133,151,171,198,199} The rates of naphthopyran formation typically differ by several orders of magnitude from *cis* and *trans* merocyanine dyes with different stereochemical configurations of the exocyclic alkene.^{80,118,133,150,152,154,155,168,181,197} In contrast, the rates of thermal ring-closure have been shown to vary only modestly for merocyanine isomers with different configuration of the distal disubstituted olefin.^{81,115,200}

Naphthopyrans and their corresponding merocyanines are somewhat susceptible to environmental effects, although to a significantly lesser degree than other molecular switches like spiropyrans. Polar solvents destabilize the quinoidal merocyanine and increase the rate of thermal reversion,^{90,163,164,201,202} although the activation barrier for this process is typically invariant across solvents.^{90,151,202} Interestingly, the thermal ring-opening reaction of naphthopyran, which again is thought to proceed through a polar, heterolytic mechanism,⁹⁶ is also faster in polar solvents that may facilitate polarization of the pyran C–O bond.^{90,164} Nevertheless, thermochromism is extremely weak. Relatively viscous solvents have also been found to slow the rate of thermal merocyanine reversion *via* interactions with the alkylidene bridge.^{124,203,204} Similarly, the rate of thermal merocyanine reversion is slower in polymeric matrices^{36,38,112,114,145,150,161,199} and ceramic coatings^{205,206} in which conformational freedom is restricted. The photochromism of naphthopyran is more efficient in polymeric materials with low glass transition temperatures and flexural moduli compared to rigid or glassy matrices.³⁶ However, naphthopyrans do exhibit photochromism in the crystalline state, which is rare for photochromic compounds.^{162,199} The resulting merocyanines are thermally stable, but revert to the colorless ring-closed naphthopyran upon irradiation with visible light.





Scheme 11 The ring-opening reactions of 3*H*- and 2*H*-naphthopyrans generate merocyanine isomers with varying degrees of steric demand, which strongly influences absorption properties and reversion kinetics.

The thermal reversion behavior of merocyanine dyes derived from 2*H*-naphthopyrans and 3*H*-naphthopyrans is markedly different. Merocyanines produced from 2*H*-naphthopyrans are typically more stable and longer-lived than those arising from 3*H*-naphthopyrans due to reduced steric crowding in both the *cis* and *trans* stereoisomers (Scheme 11).²⁰⁷ As a result of the increased steric demand, merocyanines derived from 3*H*-naphthopyran typically recyclize at rates roughly two orders of magnitude faster than those derived from 2*H*-naphthopyrans.^{90,152,155} The reduced steric interactions in merocyanines derived from 2*H*-naphthopyrans also results in greater planarization of the diene portion of the chromophore resulting in better conjugation with the quinone and visible absorption features that are bathochromically shifted compared to merocyanine dyes generated from analogous 3*H*-naphthopyrans (Fig. 4).²⁰⁸ Merocyanines generated from 3*H*-naphthopyrans typically display only one broad absorption peak in the visible

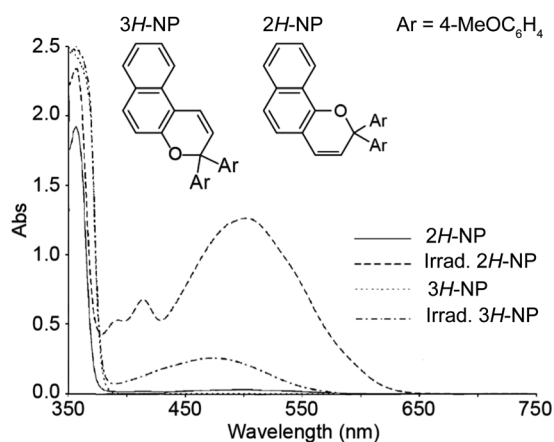


Fig. 4 UV-vis absorption spectra of 2,2-bis(4-methoxyphenyl)-2*H*-naphtho[1,2-*b*]pyran (2*H*-NP) and 3,3-bis(4-methoxyphenyl)-3*H*-naphtho[2,1-*b*]pyran (3*H*-NP) in toluene before and after irradiation with UV light. Adapted with permission from ref. 38. Copyright 2006, Elsevier.

region, whereas 2*H*-naphthopyrans produce merocyanine dyes that exhibit two distinct visible absorption features. The more intense absorption peak is commonly bathochromically shifted by approximately 45 nm compared to that of similarly substituted 3*H*-naphthopyrans.²⁰⁸ The merocyanine dyes generated from 3*H*-naphthopyrans range from red, orange, yellow, blue, or purple depending on substitution. While 2*H*-naphthopyran-derived merocyanines span a similar color palette, their unique absorption properties also enable access to commercially desirable neutral hues including brown and grey.^{37,209–212} We turn our attention to the relationships between merocyanine structure, absorption, and lifetime in the next section.

3.3 Structural features affecting merocyanine color, intensity, and lifetime

Commercial interest in the development of naphthopyran derivatives as photoswitches for use in ophthalmic lenses has culminated in a diverse and extensive library of structures producing merocyanine dyes that exhibit a wide range of colors and thermal reversion behavior (Fig. 5).^{36,37,71} In the following sections, we highlight some of the most important structure–property relationships for the naphthopyran–merocyanine system that serves as a convenient and useful reference in the context of designing mechanochromic materials.

3.3.1 Aryl substitution at the *para*-position. The absorption properties and reversion kinetics of merocyanine dyes are significantly affected by *para*-substitution of the peripheral aryl groups (Fig. 6).^{38,213} Various aryl substituents are readily introduced through modification of the benzophenone starting materials employed in the synthesis of naphthopyrans (refer to

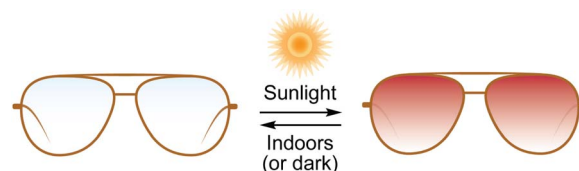


Fig. 5 Naphthopyrans have been developed extensively as UV light-responsive coloring (*i.e.*, photochromic) agents in plastic ophthalmic lenses.



Fig. 6 Electron-donating *para*-substituents result in bathochromically shifted (longer wavelength) merocyanine absorption and increase the rate of thermal reversion. Values shown were measured in toluene at 20 °C.³⁷





Fig. 7 Merocyanine color can be finely tuned by modulating the steric demand of the group adjacent to a *para*-pyrrolidine substituent. The λ_{\max} and half-life ($t_{1/2}$) are given for merocyanines with various *meta*-substituents (X). Values shown were measured in toluene at 20 °C.²¹⁴

Section 2.1). Electron-donating *para*-substituents significantly bathochromically shift the absorption of merocyanine dyes derived from both 2*H*-naphthopyrans and 3*H*-naphthopyrans with values of λ_{\max} varying by more than 100 nm. Control over the electron-donating character of these substituents enables precise modulation of merocyanine color. For example, incorporation of a *para*-pyrrolidine substituent transforms the color of the merocyanine from yellow to purple relative to the unsubstituted derivative.^{98,101} Notably, symmetrical *para*-dialkylamino substitution of both phenyl rings on a 3*H*-naphthopyran results in an additional shorter wavelength absorption feature with significant impact on the merocyanine color.³⁸ Incorporation of an additional *meta*-substituent adjacent to a pyrrolidine group at the *para*-position of the phenyl ring affords another handle for fine-tuning the absorption properties (Fig. 7).²¹⁴ Increasing steric demand by the *meta*-substituent forces the pyrrolidine heterocycle to twist out of plane, systematically reducing its degree of conjugation with the rest of the π system. Electronic factors can also be influential, however, as exemplified by the absorption behavior of the *meta*-methoxy derivative, which has a λ_{\max} that is bathochromically shifted by nearly 20 nm relative to the merocyanine without *meta*-substitution (X = H in Fig. 7). In addition to the effects on merocyanine color, *para*-substitution also dramatically alters the rate of thermal reversion. For both 2*H*- and 3*H*-naphthopyrans, the rate of the ring-closing reaction of the merocyanine dye increases with increasing electron-donating character of the *para*-substituent, with amine substituents typically eliciting the greatest effects.

3.3.2 The *ortho*-aryl substitution effect. Substitution at the *ortho*-position of the peripheral aryl groups in 3*H*-naphthopyrans significantly reduces the rate of merocyanine thermal reversion.^{98,101,213,214} This is primarily a steric effect with the degree of rate retardation scaling with the size of the *ortho*-substituent, irrespective of its electronic character (Table 1).^{98,101}

Table 1 Effect of remote *ortho*-aryl substituents on the color and lifetime of merocyanines with a *para*-pyrrolidine group. Values shown were measured in toluene at 20 °C.⁹⁸

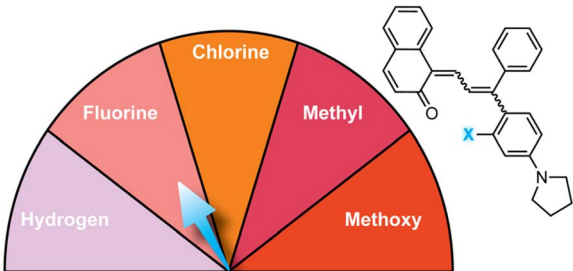
Entry	X	λ_{\max} (nm)	$t_{1/2}$ (s)
1	H	538	5
2	F	554	40
3	Cl	554	741
4	Br	554	1024
5	I	553	1167
6	OMe	555	351
7	Me	555	639

Larger *ortho*-substituents hinder formation of the *cis*-cisoid stereoisomer from which electrocyclization ultimately occurs. Notably, this is an instance in which the *cisoid* and *transoid* conformation of the merocyanine is consequential. The effect is compounded for merocyanines bearing substituents at both *ortho*-positions of a single aryl ring, resulting in even slower rates of thermal reversion.²¹³ The influence of *ortho*-substitution is effective enough to substantially mitigate the reversion-accelerating effect of an electron-rich *para*-substituent when both substituents are on the same aryl ring (Table 2). Importantly, this effectively decouples the impact of electron-donating *para*-substituents on merocyanine reversion kinetics from its influence on color, enabling access to purple merocyanines with relatively long lifetimes, for example. The systematic studies summarized in Table 1 were conducted on naphthopyrans with *para*-pyrrolidine substituents;⁹⁸ however, the *ortho*-substitution effect is general regardless of specific substitution patterns.^{37,115,213} For these same reasons, it is notable that the synthetic yields for naphthopyrans bearing significantly bulky *ortho*-aryl groups are often reduced because thermal reversion of the merocyanine is a requisite step in the mechanistic pathway, as discussed earlier.

Substitution at the *ortho*-aryl position has a moderate effect on the absorption properties of merocyanines. If the *ortho*-substituent is located on a different aryl ring than one bearing a *para*-pyrrolidine group, a moderate bathochromic shift in λ_{\max} of 15–17 nm is observed regardless of the size or electronic character (Table 1).⁹⁸ In this case, the pyrrolidine-substituted



Table 2 Effect of *ortho*-substituent steric demand on the color and lifetime of merocyanines with a *para*-pyrrolidine group on the same aryl ring. Values shown were measured in toluene at 21 °C¹⁰¹

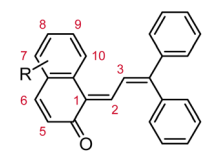


Entry	X	λ_{\max} (nm)	$t_{1/2}$ (s)
1	H	538	5
2	F	394, 501	21
3	Cl	403, 480	44
4	OMe	399, 519	73
5	Me	410, 507	104

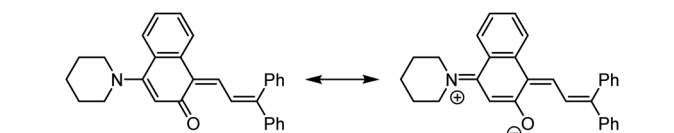
ring adopts a coplanar conformation with the merocyanine bridge to maximize conjugation with the rest of the π system. The *ortho*-substituted ring rotates out of the plane to minimize steric interactions, reducing its influence on merocyanine color. If the two substituents are on the same aryl ring, however, the absorption is hypsochromically shifted relative to the merocyanine without an *ortho*-aryl substituent due to less effective conjugation resulting from similar out-of-plane rotation (Table 2).¹⁰¹ An additional visible absorption peak around 400 nm is also observed for merocyanines with this substitution pattern. Increasing the size of the *ortho*-substituent generally leads to greater hypsochromic shifts, although electron-donating *ortho*-aryl substituents can counteract this effect. For example, the *ortho*-chlorinated merocyanine absorbs most strongly at 480 nm (Table 2, entry 3), while the merocyanine with a similarly sized *ortho*-methyl substituent has a λ_{\max} of 507 nm (Table 2, entry 5). The interplay between these steric and electronic factors has been harnessed to precisely modulate the color and reversion behavior of naphthopyran-derived merocyanines for photochromic applications.

3.3.3 Substitution of the naphthopyran core. In contrast to the effects of substitution on the peripheral aryl groups described above, the influence of substituents on the naphthopyran core is not generalizable between *2H*-naphthopyrans and *3H*-naphthopyrans.^{38,69} Typically, substituents in conjugation with the merocyanine diene bridge have the strongest influence on the photophysical properties and thermal stability of the merocyanine dyes. Electron donating alkoxy and amine substituents at the 5- and 8-positions of *3H*-naphthopyrans significantly slow thermal reversion by stabilizing the conjugated merocyanine.²¹³ Substituents at these positions also moderately affect the absorption properties, with 8-methoxy substitution resulting in a bathochromic shift in λ_{\max} of ~40 nm compared to the unsubstituted merocyanine (Table 3, entry 4).¹⁶¹ Substituents in conjugation with the ketone

Table 3 Effect of an electron-donating methoxy substituent on the absorption properties of merocyanine dyes derived from *3H*-naphthopyrans. The numbering system reflects that of the parent naphthopyran. Values of λ_{\max} were determined in an acrylic matrix¹⁶¹



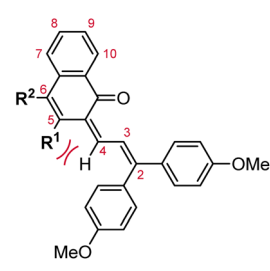
Entry	R	λ_{\max} (nm)
1	5-OMe	435
2	6-OMe	423
3	7-OMe	435
4	8-OMe	477
5	9-OMe	432
6	H	436



Scheme 12 Amine substituents can stabilize *3H*-naphthopyran-derived merocyanines through resonance.

influence λ_{\max} in the opposite way, producing minor hypsochromic shifts with electron donating groups at the 6- and 9-positions (Table 3, entries 2 and 5). More importantly, strongly electron donating substituents at the 6-position, and amine

Table 4 Effect of an electron-donating methoxy substituent on the absorption properties of merocyanine dyes derived from *2H*-naphthopyrans. Values shown were measured in toluene at 21 °C.³⁸ The numbering system reflects that of the parent naphthopyran



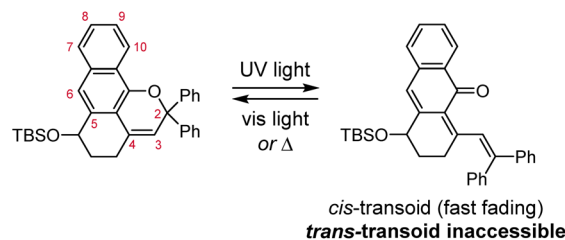
Entry	R ¹	R ²	λ_{\max} (nm)	$t_{1/2}$ (s)
1	H	H	412, 508	>1800
2	Me	H	416, 496	178
3	Me	Me	421, 494	66
4	Ph	Ph	418, 508	20
5	CO ₂ Me	H	492	2
6	CO ₂ Me	Me	416, 492	4
7	CO ₂ Me	Ph	416, 506	7



substituents in particular, can dramatically increase the optical density at the photostationary state by affording another major resonance contributor that stabilizes the merocyanine (Scheme 12).^{69,215} The effects of core substitution on the absorption properties of 2*H*-naphthopyrans are summarized in Table 4. Merocyanines derived from 2*H*-naphthopyrans exhibit absorption spectra that are bathochromically shifted relative to their 3*H*-naphthopyran analogues with significantly slower thermal reversion, as discussed earlier.^{38,69} For 2*H*-naphthopyrans, core substitution produces only slight changes in λ_{max} .³⁸ The rate of thermal reversion is primarily dictated by the steric interaction between the substituent at the 5-position and the hydrogen atom on the exocyclic alkene.

Commercial interests in the development of photochromic ophthalmic lenses have driven efforts to design naphthopyrans that generate fast-fading merocyanine dyes to achieve photo-switches with rapid switching behavior when transitioning between sunlight and indoor environments. For example, the persistence of merocyanines derived from 3*H*-naphthopyrans is significantly reduced by introducing bulky substituents at the 2- or 10-positions, which sterically impede the formation of the more thermally stable *trans* stereoisomer (Scheme 13a).^{137,185} These types of naphthopyrans produce merocyanine dyes with sub-millisecond half-lives ($t_{1/2}$). Alternatively, the presence of an alkoxy group at the 10-position of a 3*H*-naphthopyran enables a hydrogen bonding interaction in the merocyanine between the ether oxygen and the hydrogen atom on the exocyclic alkene that favors the *cis* stereoisomer and, in several cases, nearly completely suppresses photochemical accumulation of the *trans* merocyanine (Scheme 13b).^{94,185}

3.3.4 Other substitution patterns. Other structural features that are often more synthetically challenging to introduce than the ones discussed above have also been explored to control the thermal reversion behavior and absorption properties of naphthopyran-derived merocyanine dyes. These typically involve structural attributes that suppress or sometimes



Scheme 14 Ring fusion at the 4- and 5-positions of 2*H*-naphthopyrans prevents isomerization of the *cis* merocyanine isomer to the more thermally stable *trans* configuration.²²⁰

eliminate formation of the more thermally stable *trans* merocyanine stereoisomer. For example, fusing the 4- and 5-positions of a 2*H*-naphthopyran with an alkyl bridge prevents isomerization of the exocyclic olefin, resulting in the exclusive formation of the more transient *cis* merocyanine stereoisomer upon photochemical activation (Scheme 14).^{216–220} These types of merocyanines exhibit typical half-lives on the order of seconds to minutes even when *ortho*-aryl substituents are present. Conversely, some substitution patterns strongly stabilize the *trans* merocyanine dyes, resulting in thermal half-lives ranging from days to years.^{117,178,221} For example, naphthopyrans containing fused benzopyrrole or phenanthrene ring systems generate a merocyanine dye in which the *trans* stereoisomer is selectively stabilized through favorable electronic interactions between the core-extended π framework and the peripheral aryl groups.¹¹⁷ Consequently, thermal reversion to the *cis* merocyanine is strongly disfavored. Expansion of the core π framework can also lead to merocyanine dyes with colors not accessible using the strategies discussed earlier. For example, photochromic pyrans with an anthracene skeleton in place of naphthalene produce green merocyanine dyes resulting from two absorption bands at 400–480 and 580–620 nm.²²² Similarly colored merocyanines are also accessed by extending the conjugation length of the peripheral aryl groups.⁸⁷

Tables 5 and 6 summarize the privileged substitution patterns discussed in Section 3.3 and their impact on the thermal reversion rate and absorption properties of merocyanine dyes derived from 3*H*-naphthopyran and 2*H*-naphthopyrans, respectively. Beyond those discussed here, many other structural attributes of naphthopyrans have been studied and identified for controlling the properties of their merocyanine dyes. Combining substitution of the peripheral aryl groups with variations in the naphthopyran core structure vastly expands the scope of accessible thermal reversion and absorption properties. The immense range of properties available to naphthopyran-derived merocyanine dyes as showcased in this section, coupled with the synthetic modularity and accessibility of naphthopyrans, lead to their description as “the most industrially important type of photochromic colorant”.³⁷ These features not only make naphthopyran a privileged photoswitch, but in combination with their emergent mechanochemical activity, provide a highly useful platform for the design of



Scheme 13 (a) Substituents at the 2- and 10-positions of 3*H*-naphthopyrans result in severe steric repulsion in the *trans* merocyanine. (b) Hydrogen bond acceptors at the 10-position of 3*H*-naphthopyrans stabilize the *cis* merocyanine stereoisomer.



Table 5 Summary of notable substituent effects on the properties of merocyanines derived from 3*H*-naphthopyrans


Position	Substituent property	Effect on reversion	Shift in λ_{\max}	Section
A	More e ⁻ donating	Faster	Bathochromic	3.3.1
B	Larger size	—	Hypsochromic (if A = pyrrolidine)	3.3.1
C	Larger size	Slower	Hypsochromic (if A = pyrrolidine)	3.3.2
D	Larger size	Slower	Bathochromic	3.3.2
2	Larger size	Faster	—	3.3.4
5	More e ⁻ donating	Slower	—	3.3.3
6	More e ⁻ donating	Slower	Hypsochromic	3.3.3
8	More e ⁻ donating	Slower	Bathochromic	3.3.3
10	Larger size	Faster	—	3.3.4

Table 6 Summary of notable substituent effects on the properties of merocyanines derived from 2*H*-naphthopyrans


Position	Substituent property	Effect on reversion	Shift in λ_{\max}	Section
A	More e ⁻ donating	Faster	Bathochromic	3.3.1
B	Larger size	—	Hypsochromic (if A = pyrrolidine)	3.3.1
C	Larger size	Slower	Hypsochromic (if A = pyrrolidine)	3.3.2
D	Larger size	Slower	Bathochromic	3.3.2
5	More e ⁻ donating	Faster	—	3.3.4
6	More e ⁻ donating	Slower	—	3.3.3

mechanochromic polymers with an equally vast array of properties and force-responsive functions.

4 The emergent mechanochemistry of naphthopyran

4.1 Discovery of the mechanochemical ring-opening reaction of naphthopyran

The mechanically triggered ring-opening reaction of naphthopyran was first described by Moore and Sottos in 2016.³⁵ The regiochemistry of polymer attachment on the naphthopyran scaffold was found to be a critical parameter for achieving mechanochemical activation and merocyanine formation. Three different 3*H*-naphthopyran regioisomers were investigated, each with a polymer attached at the *para*-position of one of the peripheral aryl groups and another at either the 5-, 8-, or 9-position of the naphthopyran (Fig. 8a). Density functional theory (DFT) calculations performed using the constrained

geometries simulates external force (CoGEF) method,²²³ which is a reliable and straightforward tool for predicting mechanochemical reactions,¹¹ indicated that only the regioisomer with polymer attachment at the 5-position of the naphthopyran (NP5) would undergo a productive ring-opening reaction to generate the merocyanine dye. Regioisomers with pulling points at the 8- and 9-position of the naphthopyran were predicted to undergo nonproductive bond scission elsewhere in the structure upon mechanical elongation. These structure–mechanochemical activity predictions were validated by experiments performed on elastomeric poly(dimethyl siloxane) (PDMS) materials covalently incorporating each of the naphthopyran regioisomers as crosslinkers. While each material exhibited the expected photochromism upon irradiation with UV light, only the sample crosslinked with the naphthopyran regioisomer attached at the 5-position displayed mechanochemical behavior, producing an orange-yellow merocyanine dye under uniaxial tension consistent with the DFT predictions (Fig. 8b).





Fig. 8 (a) Regioisomer specific mechanochemical activity of 3H-naphthopyran. Three naphthopyran regioisomers with varying polymer attachment positions all exhibit photochromic behavior, but only regioisomer NP5 is mechanochemically active due to effective force distribution across the labile C–O pyran bond. (b) Photographs of polydimethylsiloxane samples covalently crosslinked with the three different naphthopyran regioisomers subjected to uniaxial tension or UV light irradiation. Reproduced with permission from ref. 35. Copyright 2016, American Chemical Society.

Furthermore, no mechanochromic response was observed in control experiments performed on an analogous sample incorporating a similar monofunctional naphthopyran, in which mechanical force is not transferred across the pyran C–O bond, supporting the mechanochemical nature of the transformation.

The regioisomer-specific mechanochemical reactivity of the three different naphthopyran congeners suggested that a pulling geometry providing sufficient alignment between the pyran C–O bond and the direction of externally applied mechanical force along the reaction coordinate is critical for achieving the mechanochemical ring-opening reaction of naphthopyran.

Indeed, the angle between the pyran C–O bond and the external force vector at maximum elongation prior to bond rupture was calculated to be relatively narrow for the NP5 regioisomer and significantly wider for the other two mechanochemically unreactive naphthopyran regioisomers studied. Such geometric factors have emerged as one of the most important determinants for mechanochemical reactivity. A similar geometrical analysis was extended to furan–maleimide Diels–Alder adducts, further establishing important molecular design rules based on regiochemistry and stereochemistry and their effects on the efficiency of mechanochemical coupling.²²⁴

4.2 Structure–mechanochemical activity relationships

The discovery of a mechanochemical reaction pathway for naphthopyran-to-merocyanine conversion opened up exciting new opportunities in polymer mechanochemistry and enabled the design of diverse mechanochromic polymers by leveraging the extensive structure–property relationships developed for naphthopyran photoswitches. In 2020, our group capitalized on the modularity of the naphthopyran scaffold to design a series of mechanochromic mechanophores that produce merocyanine dyes with highly tunable color and fading kinetics after mechanochemical activation in polymeric materials (Fig. 9).¹⁴⁵ Strategic structural modifications to the naphthopyran mechanophore produced dramatic differences in both the color and thermal reversion behavior of the corresponding merocyanine dyes generated upon tensile activation of PDMS elastomers. Similar to the original approach described above, 3H-naphthopyran crosslinkers bearing esters with terminal vinyl groups were covalently incorporated into PDMS materials using hydrosilylation chemistry.^{35,54} The color of the mechanically generated merocyanine dyes was varied from orange-yellow to purple upon the introduction of an electron donating pyrrolidine substituent at the *para*-position of one of the peripheral aryl groups. Likewise, the lifetime of the merocyanine dyes was modulated by varying the electronic character of the polymer attachment at the 5-position of the naphthopyran mechanophores using either a weakly electron donating alkyl linker or a more strongly electron donating alkoxy linker, which produced a faster and slower fading merocyanine, respectively. In addition, incorporation of an *ortho*-fluoro substituent on one of the peripheral aryl rings was found to significantly increase the lifetime of the merocyanine dyes produced upon mechanical activation according to the steric effects described previously in Section 3.3.2. Ultimately, this study demonstrated the generality of the design principles established for naphthopyran photoswitches, harnessing judicious yet straightforward structural modifications for the construction of naphthopyran-based mechanochromic polymers that display both fast-fading and persistent coloration upon mechanical activation with a range of accessible absorption properties. As discussed later in Section 4.4, the fine control of these features uniquely afforded by the naphthopyran framework was further exploited to design polymeric materials that exhibit desirable multicolor mechanochromic and complex stimuli-responsive behavior.





Fig. 9 (a) Different 3H-naphthopyran mechanophores with varying substitution affecting merocyanine color and reversion rate. (b) Photographs of polydimethylsiloxane samples covalently incorporating the naphthopyran crosslinkers before and after being subjected to uniaxial tension and subsequent stress relaxation, or UV light irradiation, demonstrating the modularity of the naphthopyran mechanophore for designing mechanochromic materials. Adapted from ref. 145.

In addition to the 3H-naphthopyran framework examined in earlier studies, our group has found that 2H-naphthopyrans are also competent mechanophores leading to the mechanically triggered generation of merocyanine dyes with comparatively longer lifetimes and bathochromically shifted absorption. For 2H-naphthopyran, polymer attachment at the 9-position was found to effectively couple external mechanical force with the

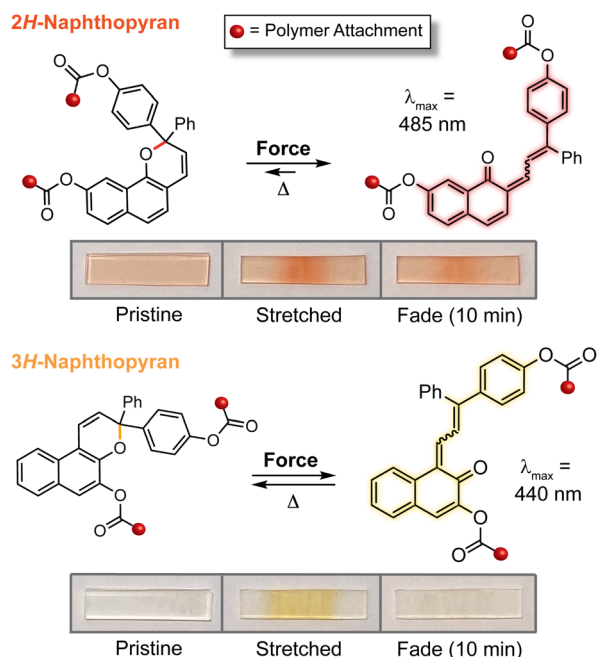


Fig. 10 Isomeric 2H- and 3H-naphthopyran mechanophores generate merocyanine dyes with distinct coloration and thermal stability. Photographs illustrate PDMS materials covalently incorporating the different naphthopyran crosslinkers with 2H-naphthopyran generating a red merocyanine that reverts relatively slowly compared to the more transient yellow merocyanine produced from a similarly substituted 3H-naphthopyran mechanophore.

selective scission of the labile pyran C–O bond leading to the desired ring-opening reaction (Fig. 10).¹⁴³ Notably, mechanochemical activation of both 2H-naphthopyran and 3H-naphthopyran mechanophores incorporated near the center of poly(methyl acrylate) polymers was demonstrated for the first time in solution using ultrasonication, which generates elongational forces on linear polymers maximized near the chain midpoint as a result of acoustic cavitation.^{3,57} While accumulation of the merocyanine dye derived from the 2H-naphthopyran mechanophore was observed at ~ 15 °C in solution, cooling the reaction to -13 °C was required to observe mechanochemical activation of the 3H-naphthopyran mechanophore, reflecting the significant differences in the thermal lifetimes of the resulting merocyanine dyes. UV-vis absorption spectra recorded after the ultrasound-induced mechanochemical activation of each mechanophore closely matched the spectra obtained after photochemical activation, indicating that similar merocyanine products are generated using mechanical force and UV light. It is interesting to note, however, that photochemical activation with UV light generates a fraction of merocyanine that is thermally persistent, in contrast to the product generated mechanochemically, which reverts completely back to the naphthopyran. These results are consistent with the photochemical generation of the more thermally stable *trans* merocyanine stereoisomer, whereas the less stable isomer with *cis* geometry of the exocyclic alkene generally appears to be produced exclusively using mechanical force. Indeed, the mechanochemical *cis* to *trans* isomerization of alkenes is uncommon,^{14,225,226} although an alternative mechanism for accessing *trans* merocyanine products from related 2H-naphthopyrans in force-coupled pathways has been postulated.¹⁴⁷ Mechanochemical activation of the 2H-naphthopyran mechanophore was also successfully demonstrated in cross-linked PDMS materials, generating a red merocyanine dye under tension that was significantly longer lived following stress relaxation than the yellow merocyanine dye produced



from the similarly substituted 3*H*-naphthopyran mechanophore (see Fig. 10). Building upon the earlier results on substituent effects, this study identified naphthopyran isomerism as an additional structural handle for modulating the mechanochromic properties of naphthopyran mechanophores, demonstrating that the diverse range of structure–property relationships developed for naphthopyran photoswitches is translatable to mechanochromic behavior.

4.3 Anomalous mechanochemical reactivity of naphthopyran

Mechanical force is a unique stimulus capable of promoting unusual reaction pathways and enabling the formation of unexpected products. For example, mechanical force has been shown to facilitate reactions that formally override conventional orbital symmetry rules including the disrotatory electrocyclic ring-opening reaction of *cis*-disubstituted benzocyclobutene²²⁷ and the conrotatory ring-opening reactions of *gem*-dihalocyclopropanes.^{228,229} Under sufficiently large forces, distortion of the potential energy surface fundamentally changes the reaction landscape.^{230–232} In the preceding sections, we described how the structure–activity relationships established for naphthopyran photoswitches translate to mechanochemical reactions of naphthopyran useful toward the design of mechanochromic polymers with a diverse set of highly tunable properties. In the following sections, we describe three exemplary cases in which the mechanochemical reactivity of naphthopyran diverges from its photochemical reactivity, leading to reaction pathways and products that are fundamentally unique. This anomalous reactivity has enabled the formation of an unusual permanent merocyanine dye and the design of multimodal mechanophores that exhibit complex mechanochromic behavior beyond the typical binary response including force-

dependent multicolor mechanochromism from a single mechanophore.

4.3.1 Mechanochemical generation of a persistent merocyanine dye. The vast majority of mechanochromic mechanophores, including most naphthopyrans, generate transiently colored products. While this feature of molecular force probes is advantageous for real-time assessment of stress and strain, the reversibility of the mechanochromic response precludes the ability to record the mechanical history of a material and also presents challenges for characterizing the kinetics of mechanochemical transformations.²² During the course of our investigation into the mechanochromic behavior of 2*H*-naphthopyran, our group discovered a naphthopyran mechanophore that produces a permanent merocyanine dye upon mechanochemical activation, in direct contrast to the reversible photochromism observed using light as the stimulus (Fig. 11).¹⁴⁴ When one of the polymer chains is attached *via* an ester linkage at the 10-position instead of the 9-position, as in the case of the 2*H*-naphthopyran mechanophore discussed earlier, scission of the C(O)–O ester bond accompanies the ring-opening reaction and establishes a β -hydroxy ketone motif that locks the merocyanine in the open state *via* an intramolecular hydrogen bonding interaction. This 2*H*-naphthopyran mechanophore was initially incorporated into poly(methyl acrylate) polymers near the chain midpoint by controlled radical polymerization from the naphthopyran bis-initiator following a commonly employed literature protocol.^{4,233} Photochemical activation of the polymer with UV light produced an orange merocyanine with λ_{max} at 470 nm that was thermally reversible. Ultrasound-induced mechanochemical activation, however, generated a distinct red merocyanine species with λ_{max} at 510 nm that persisted indefinitely in solution at room temperature. The structure of the unusual β -hydroxy ketone product was confirmed by comparisons to small molecule models supported by ¹H NMR and UV-vis absorption spectroscopy, and the selective cleavage of the C(O)–O ester bond was further corroborated by CoGEF calculations. Mechanochemical generation of the locked merocyanine was also successfully reproduced in bulk polymeric materials, which retained the characteristic red-orange coloration for several months under ambient conditions.

4.3.2 Force-dependent multicolor mechanochromism from a bis-naphthopyran mechanophore. The development of mechanochromic mechanophores capable of distinguishing between different stress states through unique visual cues represents a grand challenge in polymer mechanochemistry. In 2019, our group reported the first mechanophore that exhibits force-dependent multicolor mechanochromism,¹³⁵ which leveraged a bis-naphthopyran (BNP) structure capable of undergoing two distinct ring-opening reactions with each resulting in a characteristic change in color.^{116,234} This type of multimodal reactivity has since attracted significant research interest motivated by the potential to enable quantitative characterization of stress and/or strain in polymeric materials through straightforward visual analysis.²³⁵ Bis-naphthopyrans contain two naphthopyran subunits joined by a conjugated linker and are among the few classes of photochromic

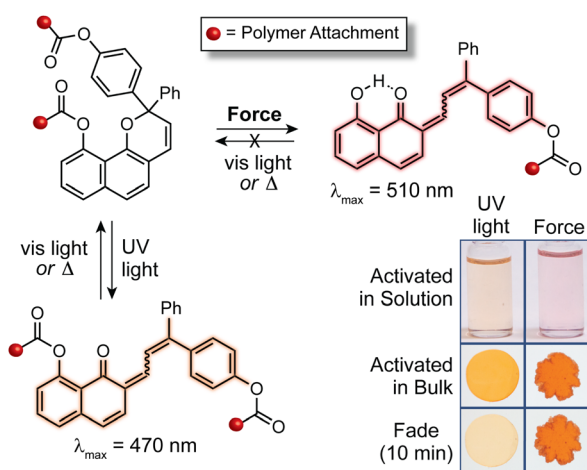


Fig. 11 In contrast to the reversible photochemical reaction, mechanochemical activation of a 2*H*-naphthopyran mechanophore with polymer attachment *via* an ester linkage at the 10-position generates a persistent merocyanine dye stabilized by an intramolecular hydrogen bonding interaction that is established upon scission of the C(O)–O ester bond.



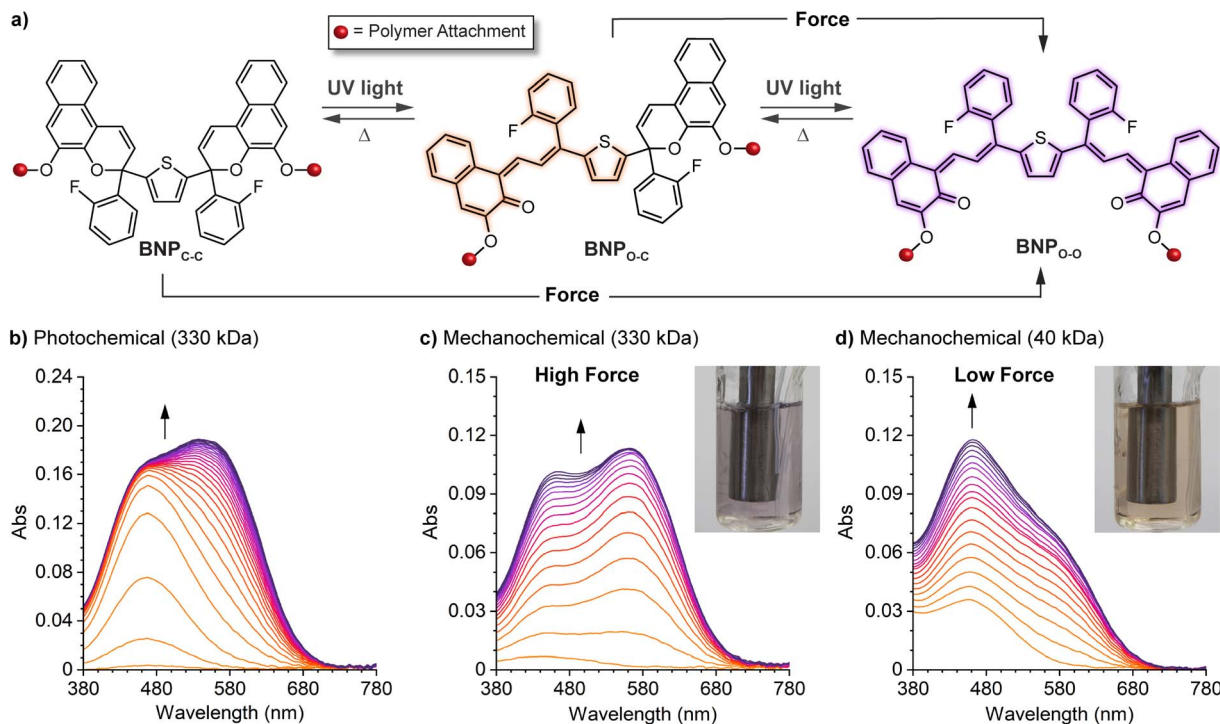


Fig. 12 Force-dependent multicolor mechanochromism from a bis-naphthopyran (BNP) mechanophore. (a) Unlike the sequential ring-opening reactions observed upon photochemical activation, mechanical force directly converts the BNP mechanophore to the purple bis-merocyanine species, which thermally reverts to the yellow monomerocyanine. Changing the amount of force delivered to the mechanophore biases the merocyanine distribution resulting in force-dependent coloration. (b) UV-vis absorption spectra of a 330 kDa poly(methyl acrylate) polymer with a chain-centered BNP unit illustrating sequential-ring opening behavior upon irradiation with UV light. Ultrasound-induced mechanochemical activation of (c) the same 330 kDa polymer, and (d) an analogous 40 kDa polymer that experiences lower force, illustrating non-sequential ring opening behavior. Insets show photographs of the final sonicated solutions. Adapted with permission from ref. 135. Copyright 2019, American Chemical Society.

compounds capable of accessing distinctly colored states based on the duration or intensity of light exposure.^{116,119,120,234,236–239} Encouraged by the photochromic properties of the bis-naphthopyran photoswitches originally reported by Zhao and Carreira,^{116,119,234} we designed a series of poly(methyl acrylate) polymers incorporating a bis-naphthopyran unit near the chain center for direct comparison of the photochemical and mechanochemical reactivity (Fig. 12a). Consistent with the expected sequential ring-opening behavior, photoirradiation of dilute polymer solutions with UV light first generated the yellow open-closed monomerocyanine species ($\text{BNP}_{\text{O-c}}$) with λ_{max} at 460 nm, followed by a second ring-opening reaction upon further photoirradiation to form the purple open-open bis-merocyanine product ($\text{BNP}_{\text{O-o}}$) with extended conjugation and significantly longer wavelength absorption (Fig. 12b). Remarkably, ultrasound induced mechanochemical activation of the same polymer resulted in substantially different kinetic behavior. Accumulation of the purple bis-merocyanine product occurred early in the mechanochemical reaction with the yellow monomerocyanine species being generated more slowly, in direct contrast to the photochemical transformation (Fig. 12c). Moreover, varying the magnitude of force delivered to the BNP mechanophore by modifying the length of the attached polymer chains²⁴⁰ resulted in a markedly different absorption spectrum

(Fig. 12d). These results suggested that the relative distribution of the two distinctly colored merocyanine products resulting from the mechanochemical reaction and the correlated visible absorption spectrum are force-dependent.

Further insights were provided by a detailed kinetic analysis, which confirmed that mechanochemical activation of the BNP mechanophore occurs *via* a mechanistically distinct pathway compared to the photochemical process. In contrast to the photochemical pathway that proceeds *via* two sequential ring-opening reactions, the data support a non-sequential mechanochemical ring-opening mechanism in which the colorless BNP mechanophore is effectively converted directly to the purple bis-merocyanine product and formation of the yellow mono-merocyanine occurs primarily, if not exclusively, from thermal electrocyclization, which is invariant with force (see Fig. 12a). Therefore, the relative distribution of the two distinctly colored merocyanine dyes is predictably modulated by varying the magnitude of force applied to the BNP mechanophore, resulting in a gradient change in the overall visible



absorption. The direct conversion of BNP to the bis-merocyanine species may reflect unusual dynamic effects observed in the ultrasound-induced mechanochemical transformations of other mechanophores whereby the momenta resulting from extrinsic force lead to flyby trajectories that completely bypass reactive intermediates.²⁴¹ We speculate that if the barriers associated with the two ring-opening reactions were sufficiently different, such as in the case of an asymmetric bis-naphthopyran mechanophore, then each ring-opening reaction may exhibit a discreet force threshold for activation leading to force-dependent multicolor mechanochromism that is not kinetically derived. Alternatively, the apparent mechanochemical reaction mechanism may be different in solid materials subjected to slower strain rates. In this case, it is reasonable to consider that strain could dominate the ring-opening reactivity and still lead to a similar multicolor mechanochromic response that more closely mirrors the sequential ring-opening behavior in the photochemical reaction pathway. Investigations of the solid state mechanochemical reactivity of bis-naphthopyran are currently underway in our laboratory that

should shed light on some of these questions and advance the development of multimodal mechanophores toward quantitative visual force sensing applications. We also recently described another bis-naphthopyran mechanophore based on the scissile *2H*-naphthopyran scaffold described above (refer to Section 4.3.3).¹⁴⁷ Experiments suggested that the mechanically unveiled β -hydroxy ketone subunit confers remarkable thermal stability to bis-merocyanine isomers with a *trans* exocyclic alkene on the other merocyanine subunit and implicated the formation of an unusual *trans* merocyanine isomer as the product of mechanochemical activation.

4.3.3 Dual ring-opening reaction of naphthodipyran with mechanical force. Naphthodipyrans are compounds that consist of two separate pyran rings fused to a single naphthalene core and represent a potentially promising platform for multimodal mechanochemical reactivity. However, in contrast to bis-naphthopyrans, the concurrent ring opening of both pyran moieties is inaccessible photochemically.^{38,124,207} Given the propensity for mechanical force to facilitate other unconventional transformations, we hypothesized that the dual ring-



Fig. 13 (a) Divergent photochemical and mechanochemical reactivity of a *3H*-naphthodipyran (NDP) moiety incorporated near the center of a poly(methyl acrylate) chain. (b) UV-vis-near-infrared absorption spectra before and after photoirradiation of the polymer with UV light or ultrasound-induced mechanochemical activation. Inset shows a photograph of the two solutions after activation. (c) Absorption spectra recorded during ultrasonication of the polymer and time-dependent absorption profiles at 625 and 820 nm characteristic of monomerocyanine $NDP_{O-C} \cdot nBF_3$ and dimerocyanine $NDP_{O-O} \cdot nBF_3$, respectively. Adapted with permission from ref. 146. Copyright 2022, American Chemical Society.



opening reaction of naphthodipyran to produce an elusive dimerocyanine species could be achieved mechanochemically. Critical to the investigation was the prior discovery that naphthopyran-derived merocyanines bearing electron rich amine substituents could be irreversibly trapped with boron trifluoride, which presumably coordinates to the ketone oxygen atom to prevent electrocyclization.¹¹⁰ Eliminating the thermal reversion of a hypothetical dimerocyanine species would rule out the possibility that it was generated but accumulation was not observed simply due to its transience. Promisingly, both pyran rings on a naphthodipyran model bearing *para*-pyrrolidine substituents on two of the pendant aryl groups were predicted to be mechanochemically active by CoGEF calculations, with mechanical elongation ultimately resulting in the formation of the dimerocyanine product.¹⁴⁶ Similar to the study of bis-naphthopyran described above, the naphthodipyran compound was incorporated near the center of a poly(methyl acrylate) polymer to compare its photochemical and mechanochemical reactivity (Fig. 13a). Photoirradiation of the polymer with UV light in the presence of BF₃ resulted in the formation of a persistent blue monomerocyanine dye with a single visible absorption peak at 625 nm (Fig. 13b). In direct contrast, ultrasound-induced mechanochemical activation of the polymer under otherwise similar conditions resulted in the generation of a persistent violet colored species with a unique absorption spectrum that extends into the near-infrared region, consistent with the formation of the dimerocyanine compound in which both pyran rings have opened. Further structural evidence was provided by UV-vis and ¹H NMR spectroscopy as well as high-resolution mass spectrometry performed on a small molecule model compound, which supported that the dimerocyanine species was trapped *via* the formation of a stable difluoroborocyclic heterocycle involving both of the oxygen atoms of the 1,2-diketone motif.

In a notable deviation from the kinetic behavior of the BNP mechanophore, the absorption spectra evolved during the ultrasound-induced mechanochemical activation of naphthodipyran (Fig. 13c). At early reaction times, only the shorter wavelength absorption features of the monomerocyanine were observed, with the longer wavelength absorption features characteristic of the dimerocyanine emerging later. The kinetic behavior observed for the naphthodipyran mechanophore reports colorimetrically on temporal exposure to ultrasound-induced mechanical force and is effectively indistinguishable from a sequential ring-opening mechanism. This behavior may, in fact, indicate substantial differences in the barriers of the first and second ring-opening reactions, which occur in sequence as opposed to directly as in the case of the BNP mechanophore discussed earlier. Alternatively, the temporal evolution of the absorption spectra may reflect kinetic limitations in the trapping reaction of BF₃ with the highly transient dimerocyanine species, requiring two separate chain-pulling events to form the thermally stable dimerocyanine adduct (NDP_{o-o}·nBF₃). Fundamental questions remain about how mechanical force is able to promote the dual ring-opening reaction of naphthodipyran in a process that diverges from the photochemical pathway. From a practical perspective, we

envision that the newly unveiled multimodal reactivity of naphthodipyran, in combination with the unusual mechanochemical activation kinetics, will enable the development of additional mechanophores and force-responsive polymers that exhibit multicolor mechanochromism.

4.4 Design of more complex stimuli-responsive materials

The design of materials with multicolor mechanochromic and more complex stimuli-responsive behavior has also been achieved by blending distinct mechanophores with complementary reactivity and/or properties. The Otsuka and Moore groups teamed up to develop a polymer/silica composite material capable of displaying different colors under varying degrees of mechanical stimulation as well as orthogonal responses to light and solvent swelling (Fig. 14a).¹³⁴ A naphthopyran mechanophore was incorporated into the soft polymer-rich domain while a radical-type diarylbibenzofuranone (DABBF) mechanophore was incorporated into a hard silica-rich domain. The design leverages the different colors of the merocyanine (orange-yellow) and stable free radical (blue) products that result from mechanochemical activation of naphthopyran and DABBF, respectively, as well as the disparate thermal reversion kinetics, which are also impacted by the substantial differences in mobility afforded by the hard and soft domains in the composite. The material turned blue upon gentle kneading in a mortar, which is characteristic of the mechanical activation of the more labile DABBF mechanophore at the polymer/silica interface. Stronger kneading caused the material to turn a greenish hue, reflecting the combined appearance of both the orange-yellow naphthopyran-derived merocyanine and the blue-colored radical species derived from the DABBF mechanophore. The authors also demonstrated that each colored product could be selectively bleached using either visible light to revert the merocyanine dye or upon the introduction of solvent to promote radical recombination, further modulating the appearance of the material with different stimuli following initial mechanical activation. This study illustrated that the combination of two different mechanophores with complementary reactivity and properties in a judiciously designed matrix could achieve more sophisticated stimuli-responsive behavior and ultimately increase the capacity of information encoded in a material *via* mechanical deformation.

The broad structure–property range accessible to naphthopyrans lends itself to a similar strategy of blending complementary mechanophores to achieve more complex force and stimuli-responsive function. By leveraging two different naphthopyran mechanophores that produce distinctly colored merocyanine dyes with substantially different thermal reversion kinetics, our group designed PDMS elastomers capable of distinguishing between different stress states with temporal resolution (Fig. 14b).¹⁴⁵ Specifically, elastomeric films were prepared that incorporated two different naphthopyran mechanophores, one that generates a fast-fading purple merocyanine dye and another that forms a more thermally persistent orange-yellow merocyanine dye (refer to Section 4.2). Mechanical activation of the material in tension caused the film to turn a red-orange



new insights into structure–mechanochemical activity relationships, elucidated anomalous reactivity that diverges from typical photochemical reaction pathways, and contributed broadly to the design and development of mechanochromic polymers and materials with diverse and often remarkable properties. The field of polymer mechanochemistry has exploded in research activity and interest in recent years because it represents a fundamentally different way to control and harness chemical reactivity. In the context of mechanochromism, mechanophores function as molecular force probes to enable the molecular-level visualization of critical stress and/or strain in polymers with applications in both basic and applied science. As new mechanophores are developed, focus is expanding to include the design of molecular force probes that exhibit complex, non-binary responses that enable the visual quantification of stress in materials. Naphthopyrans have proven to be an exceptional motif for constructing these types of multimodal mechanophores. As the field matures, naphthopyrans are privileged for their decades of industrial development as photoswitches that may prove fruitful in future translation of their mechanochromic function to commercial applications. Naphthopyran mechanophores have already played an important role in the development of the field and we look forward to seeing the impact of this unique and highly capable mechanophore platform on the bright and colorful future of polymer mechanochemistry.

Author contributions

M. E. M. wrote the first draft of the manuscript with contributions from R. W. B. and A. C. O. M. J. R. provided direction and edited the manuscript.

Conflicts of interest

There are no conflicts to declare.

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References

- 1 K. L. Berkowski, S. L. Potisek, C. R. Hickenboth and J. S. Moore, *Macromolecules*, 2005, **38**, 8975–8978.
- 2 M. K. Beyer and H. Clausen-Schaumann, *Chem. Rev.*, 2005, **105**, 2921–2948.
- 3 M. M. Caruso, D. A. Davis, Q. Shen, S. A. Odom, N. R. Sottos, S. R. White and J. S. Moore, *Chem. Rev.*, 2009, **109**, 5755–5798.
- 4 J. Li, C. Nagamani and J. S. Moore, *Acc. Chem. Res.*, 2015, **48**, 2181–2190.
- 5 J. F. Patrick, M. J. Robb, N. R. Sottos, J. S. Moore and S. R. White, *Nature*, 2016, **540**, 363–370.
- 6 S. Wang, H. K. Beech, B. H. Bowser, T. B. Kouznetsova, B. D. Olsen, M. Rubinstein and S. L. Craig, *J. Am. Chem. Soc.*, 2021, **143**, 3714–3718.
- 7 S. Wang, Y. Hu, T. B. Kouznetsova, L. Sapir, D. Chen, A. Herzog-Arbeitman, J. A. Johnson, M. Rubinstein and S. L. Craig, *Science*, 2023, **380**, 1248–1252.
- 8 Q. Wang, G. R. Gossweiler, S. L. Craig and X. Zhao, *Nat. Commun.*, 2014, **5**, 4899.
- 9 G. Kim, V. M. Lau, A. J. Halmes, M. L. Oelze, J. S. Moore and K. C. Li, *Proc. Natl. Acad. Sci. U. S. A.*, 2019, **116**, 10214–10222.
- 10 A. C. Overholts, W. Granados Razo and M. J. Robb, *Nat. Chem.*, 2023, **15**, 332–338.
- 11 I. M. Klein, C. C. Husic, D. P. Kovács, N. J. Choquette and M. J. Robb, *J. Am. Chem. Soc.*, 2020, **142**, 16364–16381.
- 12 A. Piermattei, S. Karthikeyan and R. P. Sijbesma, *Nat. Chem.*, 2009, **1**, 133–137.
- 13 P. Michael and W. H. Binder, *Angew. Chem., Int. Ed.*, 2015, **54**, 13918–13922.
- 14 Z. Chen, J. A. M. Mercer, X. Zhu, J. A. H. Romaniuk, R. Pfattner, L. Cegelski, T. J. Martinez, N. Z. Burns and Y. Xia, *Science*, 2017, **357**, 475–479.
- 15 Y. Chen, A. J. H. Spiering, S. Karthikeyan, G. W. M. Peters, E. W. Meijer and R. P. Sijbesma, *Nat. Chem.*, 2012, **4**, 559–562.
- 16 X. Hu, T. Zeng, C. C. Husic and M. J. Robb, *J. Am. Chem. Soc.*, 2019, **141**, 15018–15023.
- 17 X. Hu, T. Zeng, C. C. Husic and M. J. Robb, *ACS Cent. Sci.*, 2021, **7**, 1216–1224.
- 18 S. Huo, P. Zhao, Z. Shi, M. Zou, X. Yang, E. Warsawik, M. Loznik, R. Göstl and A. Herrmann, *Nat. Chem.*, 2021, **13**, 131–139.
- 19 M. A. Ghanem, A. Basu, R. Behrou, N. Boechler, A. J. Boydston, S. L. Craig, Y. Lin, B. E. Lynde, A. Nelson, H. Shen and D. W. Storti, *Nat. Rev. Mater.*, 2020, **6**, 84–98.
- 20 Y. Chen, G. Mellot, D. van Luijk, C. Creton and R. P. Sijbesma, *Chem. Soc. Rev.*, 2021, **50**, 4100–4140.
- 21 C. Calvino, L. Neumann, C. Weder and S. Schrettl, *J. Polym. Sci., Part A: Polym. Chem.*, 2017, **55**, 640–652.
- 22 R. W. Barber, M. E. McFadden, X. Hu and M. J. Robb, *Synlett*, 2019, **30**, 1725–1732.
- 23 M. Stratigaki and R. Göstl, *ChemPlusChem*, 2020, **85**, 1095–1103.
- 24 H. Traeger, D. J. Kiebal, C. Weder and S. Schrettl, *Macromol. Rapid Commun.*, 2021, **42**, 2000573.
- 25 H. Staudinger and H. F. Bondy, *Ber. Dtsch. Chem. Ges.*, 1930, **63**, 724–730.
- 26 H. Staudinger and H. F. Bondy, *Ber. Dtsch. Chem. Ges.*, 1930, **63**, 734–736.



- 27 H. Staudinger and W. Heuer, *Ber. Dtsch. Chem. Ges.*, 1934, **67**, 1159–1164.
- 28 J. Sohma, *Prog. Polym. Sci.*, 1989, **14**, 451–596.
- 29 E. Ducrot, Y. Chen, M. Bulters, R. P. Sijbesma and C. Creton, *Science*, 2014, **344**, 186–189.
- 30 P. A. May, N. F. Munaretto, M. B. Hamoy, M. J. Robb and J. S. Moore, *ACS Macro Lett.*, 2016, **5**, 177–180.
- 31 T. A. Kim, M. J. Robb, J. S. Moore, S. R. White and N. R. Sottos, *Macromolecules*, 2018, **51**, 9177–9183.
- 32 Y. Lin, M. H. Barbee, C. C. Chang and S. L. Craig, *J. Am. Chem. Soc.*, 2018, **140**, 15969–15975.
- 33 M. E. McFadden, A. C. Overholts, S. K. Osler and M. J. Robb, *ACS Macro Lett.*, 2023, **12**, 440–445.
- 34 R. S. Becker and J. Michl, *J. Am. Chem. Soc.*, 1966, **88**, 5931–5933.
- 35 M. J. Robb, T. A. Kim, A. J. Halmes, S. R. White, N. R. Sottos and J. S. Moore, *J. Am. Chem. Soc.*, 2016, **138**, 12328–12331.
- 36 S. N. Corns, S. M. Partington and A. D. Towns, *Color. Technol.*, 2009, **125**, 249–261.
- 37 A. Towns, *Phys. Sci. Rev.*, 2020, **5**, 20190085.
- 38 J. D. Hepworth and B. M. Heron, in *Functional Dyes*, Elsevier Science, 2006, pp. 85–135.
- 39 D. A. Davis, A. Hamilton, J. Yang, L. D. Cremar, D. Van Gough, S. L. Potisek, M. T. Ong, P. V. Braun, T. J. Martinez, S. R. White, J. S. Moore and N. R. Sottos, *Nature*, 2009, **459**, 68–72.
- 40 J. M. Clough, C. Creton, S. L. Craig and R. P. Sijbesma, *Adv. Funct. Mater.*, 2016, **26**, 9063–9074.
- 41 C. Baumann, M. Stratigaki, S. P. Centeno and R. Göstl, *Angew. Chem., Int. Ed.*, 2021, **60**, 13287–13293.
- 42 K. Imato, A. Irie, T. Kosuge, T. Ohishi, M. Nishihara, A. Takahara and H. Otsuka, *Angew. Chem., Int. Ed.*, 2015, **54**, 6168–6172.
- 43 F. Verstraeten, R. Göstl and R. P. Sijbesma, *Chem. Commun.*, 2016, **52**, 8608–8611.
- 44 K. Ishizuki, D. Aoki, R. Goseki and H. Otsuka, *ACS Macro Lett.*, 2018, **7**, 556–560.
- 45 K. Ishizuki, H. Oka, D. Aoki, R. Goseki and H. Otsuka, *Chem.–Eur. J.*, 2018, **24**, 3170–3173.
- 46 S. Kato, K. Ishizuki, D. Aoki, R. Goseki and H. Otsuka, *ACS Macro Lett.*, 2018, **12**, 28.
- 47 H. Sakai, T. Sumi, D. Aoki, R. Goseki and H. Otsuka, *ACS Macro Lett.*, 2018, **7**, 1359–1363.
- 48 S. Kato, S. Furukawa, D. Aoki, R. Goseki, K. Oikawa, K. Tsuchiya, N. Shimada, A. Maruyama, K. Numata and H. Otsuka, *Nat. Commun.*, 2021, **12**, 126.
- 49 T. Watabe, D. Aoki and H. Otsuka, *Macromolecules*, 2022, **55**, 5795–5802.
- 50 R. Göstl and R. P. Sijbesma, *Chem. Sci.*, 2016, **7**, 370–375.
- 51 C. Baumann and R. Göstl, *Synlett*, 2022, **33**, 875–878.
- 52 J. Yang, M. Horst, S. H. Werby, L. Cegelski, N. Z. Burns and Y. Xia, *J. Am. Chem. Soc.*, 2020, **142**, 14619–14626.
- 53 B. R. Boswell, C. M. F. Mansson, J. M. Cox, Z. Jin, J. A. H. Romaniuk, K. P. Lindquist, L. Cegelski, Y. Xia, S. A. Lopez and N. Z. Burns, *Nat. Chem.*, 2021, **13**, 41–46.
- 54 G. R. Gossweiler, G. B. Hewage, G. Soriano, Q. Wang, G. W. Welshofer, X. Zhao and S. L. Craig, *ACS Macro Lett.*, 2014, **3**, 216–219.
- 55 G. R. Gossweiler, T. B. Kouznetsova and S. L. Craig, *J. Am. Chem. Soc.*, 2015, **137**, 6148–6151.
- 56 S. L. Potisek, D. A. Davis, N. R. Sottos, S. R. White and J. S. Moore, *J. Am. Chem. Soc.*, 2007, **129**, 13808–13809.
- 57 P. A. May and J. S. Moore, *Chem. Soc. Rev.*, 2013, **42**, 7497–7506.
- 58 H. Zhang, F. Gao, X. Cao, Y. Li, Y. Xu, W. Weng and R. Boulatov, *Angew. Chem., Int. Ed.*, 2016, **55**, 3040–3044.
- 59 R. Yao, X. Li, N. Xiao, W. Weng and W. Zhang, *Nano Res.*, 2021, **14**, 2654–2658.
- 60 Z. Wang, Z. Ma, Y. Wang, Z. Xu, Y. Luo, Y. Wei and X. Jia, *Adv. Mater.*, 2015, **27**, 6469–6474.
- 61 M. Wu, Y. Li, W. Yuan, G. De Bo, Y. Cao and Y. Chen, *J. Am. Chem. Soc.*, 2022, **144**, 17120–17128.
- 62 H. Qian, N. S. Purwanto, D. G. Ivanoff, A. J. Halmes, N. R. Sottos and J. S. Moore, *Chem*, 2021, **7**, 1080–1091.
- 63 Q. Qi, G. Sekhon, R. Chandradat, N. M. Ofodum, T. Shen, J. Scrimgeour, M. Joy, M. Wriedt, M. Jayathirtha, C. C. Darie, D. A. Shipp, X. Liu and X. Lu, *J. Am. Chem. Soc.*, 2021, **143**, 17337–17343.
- 64 V. I. Minkin, *Chem. Rev.*, 2004, **104**, 2751–2776.
- 65 H. N. Kim, M. H. Lee, H. J. Kim, J. S. Kim and J. Yoon, *Chem. Soc. Rev.*, 2008, **37**, 1465–1472.
- 66 S. V. Paramonov, V. Lokshin and O. A. Fedorova, *J. Photochem. Photobiol., C*, 2011, **12**, 209–236.
- 67 G. I. Peterson, M. B. Larsen, M. A. Ganter, D. W. Storti and A. J. Boydston, *ACS Appl. Mater. Interfaces*, 2015, **7**, 577–583.
- 68 S. Swansburg, E. Buncel and R. P. Lemieux, *J. Am. Chem. Soc.*, 2000, **122**, 6594–6600.
- 69 B. Van Gemert, in *Organic Photochromic and Thermochromic Compounds*, Springer, Boston, MA, 2002, pp. 111–140.
- 70 J. Hoble, M. J. Lear and H. Fukumura, in *Photochemistry of Organic Molecules in Isotropic and Anisotropic Media*, CRC Press, 2003.
- 71 A. Towns, in *Applied Photochemistry*, Springer International Publishing, 2016, pp. 227–279.
- 72 S. Ghosh, J. Hall and V. Joshi, *J. Text. Inst.*, 2018, **109**, 723–729.
- 73 K. Jin, X. Ji, T. Yang, J. Zhang, W. Tian, J. Yu, X. Zhang, Z. Chen and J. Zhang, *Chem. Eng. J.*, 2021, **406**, 126794.
- 74 X. Blin and J. C. Simon, *Fr. Pat.*, FR2845910A1, 2004.
- 75 C. Moustrou, N. Rebière, A. Samat, R. Guglielmetti, A. E. Yassar, R. Dubest and J. Aubard, *Helv. Chim. Acta*, 1998, **81**, 1293–1302.
- 76 A. Yassar, N. Rebière-Galy, M. Frigoli, C. Moustrou, A. Samat, R. Guglielmetti and A. Jaafari, in *Synthetic Metals*, Elsevier, 2001, vol. 124, pp. 23–27.
- 77 A. Yassar, H. Jaafari, N. Rebière-Galy, M. Frigoli, C. Moustrou, A. Samat and R. Guglielmetti, *Eur. Phys. J.: Appl. Phys.*, 2002, **18**, 3–8.
- 78 A. Yassar, F. Garnier, H. Jaafari, N. Rebière-Galy, M. Frigoli, C. Moustrou, A. Samat and R. Guglielmetti, *Appl. Phys. Lett.*, 2002, **80**, 4297–4299.
- 79 M. Frigoli, C. Moustrou, A. Samat and R. Guglielmetti, *Eur. J. Org. Chem.*, 2003, **2003**, 2799–2812.



- 80 M. Frigoli, V. Pimienta, C. Moustrou, A. Samat, R. Guglielmetti, J. Aubard, F. Maurel and J.-C. Micheau, *Photochem. Photobiol. Sci.*, 2003, **2**, 888–892.
- 81 F. Ortica, P. Smimmo, G. Favaro, U. Mazzucato, S. Delbaere, D. Venec, G. Vermeersch, M. Frigoli, C. Moustrou and A. Samat, *Photochem. Photobiol. Sci.*, 2004, **3**, 878–885.
- 82 P. L. Gentili, A. L. Rightler, B. M. Heron and C. D. Gabbutt, *Dyes Pigm.*, 2016, **135**, 169–176.
- 83 P. L. Gentili, A. L. Rightler, B. M. Heron and C. D. Gabbutt, *Chem. Commun.*, 2016, **52**, 1474–1477.
- 84 P. L. Gentili, M. S. Giubila and B. M. Heron, *ChemPhysChem*, 2017, **18**, 1831–1841.
- 85 H. Wu, A. Huang, Q. Liao, P. Lin, W. Yao, Q. Li and Z. Li, *ACS Mater. Lett.*, 2023, **5**, 753–761.
- 86 Z. M. Dong, H. Ren, J. N. Wang and Y. Wang, *Microchem. J.*, 2020, **155**, 104676.
- 87 V. M. Mwalukuku, J. Liotier, A. J. Riquelme, Y. Kervella, Q. Hualmé, A. Haurez, S. Narbey, J. A. Anta and R. Demadrille, *Adv. Energy Mater.*, 2023, **13**, 2203651.
- 88 P. L. Gentili, M. S. Giubila, R. Germani and B. M. Heron, *Dyes Pigm.*, 2018, **156**, 149–159.
- 89 I. Iwai and J. Ide, *Chem. Pharm. Bull.*, 1962, **10**, 926–933.
- 90 J.-L. Pozzo, A. Samat, R. Guglielmetti, R. Dubest and J. Aubard, *Helv. Chim. Acta*, 1997, **80**, 725–738.
- 91 C. D. Gabbutt, T. Gelbrich, J. D. Hepworth, B. M. Heron, M. B. Hursthouse and S. M. Partington, *Dyes Pigm.*, 2002, **54**, 79–93.
- 92 E. N. Ushakov, V. B. Nazarov, O. A. Fedorova, S. P. Gromov, A. V. Chebun'kova, M. V. Alfimov and F. Barigelletti, *J. Phys. Org. Chem.*, 2003, **16**, 306–309.
- 93 X. Sallenave, S. Delbaere, G. Vermeersch, A. Saleh and J. L. Pozzo, *Tetrahedron Lett.*, 2005, **46**, 3257–3259.
- 94 Y. Inagaki, Y. Kobayashi, K. Mutoh and J. Abe, *J. Am. Chem. Soc.*, 2017, **139**, 13429–13441.
- 95 H. Kuroiwa, Y. Inagaki, K. Mutoh and J. Abe, *Adv. Mater.*, 2019, **31**, 1805661.
- 96 G. Harié, A. Samat, R. Guglielmetti, I. van Parys, W. Saeyens, D. de Keukeleire, K. Lorenz and A. Mannschreck, *Helv. Chim. Acta*, 1997, **80**, 1122–1132.
- 97 S. Aiken, O. D. C. C. de Azevedo, K. Chauhan, T. Driscoll, P. I. Elliott, C. D. Gabbutt and B. M. Heron, *J. Org. Chem.*, 2020, **85**, 952–966.
- 98 B. M. Heron, C. D. Gabbutt and A. C. Instone, *Heterocycles*, 2003, **60**, 843.
- 99 C. D. Gabbutt, B. M. Heron, A. C. Instone, D. A. Thomas, S. M. Partington, M. B. Hursthouse and T. Gelbrich, *Eur. J. Org. Chem.*, 2003, **2003**, 1220–1230.
- 100 C. D. Gabbutt, B. M. Heron, S. B. Kolla, C. Kilner, S. J. Coles, P. N. Horton and M. B. Hursthouse, *Org. Biomol. Chem.*, 2008, **6**, 3096–3104.
- 101 C. D. Gabbutt, B. M. Heron and A. C. Instone, *Tetrahedron*, 2006, **62**, 737–745.
- 102 G. Harié, A. Samat and R. Guglielmetti, *Mol. Cryst. Liq. Cryst.*, 1997, **297**, 263–268.
- 103 S. Aiken, K. Booth, C. D. Gabbutt, B. M. Heron, C. R. Rice, A. Charaf-Eddin and D. Jacquemin, *Chem. Commun.*, 2014, **50**, 7900–7903.
- 104 B. Van Gemert and M. P. Bergomi, *US Pat.*, US5066818A, 1991.
- 105 C. D. Gabbutt, J. D. Hepworth, B. M. Heron, S. M. Partington and D. A. Thomas, *Dyes Pigm.*, 2001, **49**, 65–74.
- 106 S. Swaminathan and K. V. Narayanan, *Chem. Rev.*, 1971, **71**, 429–438.
- 107 L. M. Carvalho, A. M. S. Silva, C. I. Martins, P. J. Coelho and A. M. F. Oliveira-Campos, *Tetrahedron Lett.*, 2003, **44**, 1903–1905.
- 108 W. Zhao and E. M. Carreira, *Org. Lett.*, 2003, **5**, 4153–4154.
- 109 K. Tanaka, H. Aoki, H. Hosomi and S. Ohba, *Org. Lett.*, 2000, **2**, 2133–2134.
- 110 K. Guo and Y. Chen, *J. Phys. Org. Chem.*, 2010, **23**, 207–210.
- 111 S. Aiken, B. Allsopp, K. Booth, C. D. Gabbutt, B. M. Heron and C. R. Rice, *Tetrahedron*, 2014, **70**, 9352–9358.
- 112 K. Guo and Y. Chen, *J. Mater. Chem.*, 2010, **20**, 4193–4197.
- 113 S. Delbaere, G. Vermeersch, M. Frigoli and G. H. Mehl, *Org. Lett.*, 2010, **12**, 4090–4093.
- 114 S. Han and Y. Chen, *J. Mater. Chem.*, 2011, **21**, 4961–4965.
- 115 S. Delbaere, J.-C. Micheau, Y. Teral, C. Bochu, M. Campredon and G. Vermeersch, *Photochem. Photobiol.*, 2001, **74**, 694.
- 116 W. Zhao and E. M. Carreira, *Chem.–Eur. J.*, 2007, **13**, 2671–2688.
- 117 M. Frigoli, J. Marrot, P. L. Gentili, D. Jacquemin, M. Vagnini, D. Pannacci and F. Ortica, *ChemPhysChem*, 2015, **16**, 2447–2458.
- 118 P. J. Coelho, M. A. Salvador, M. M. Oliveira and L. M. Carvalho, *J. Photochem. Photobiol., A*, 2005, **172**, 300–307.
- 119 W. Zhao and E. M. Carreira, *Org. Lett.*, 2006, **8**, 99–102.
- 120 X. Lu, Q. Dong, X. Dong and W. Zhao, *Tetrahedron*, 2015, **71**, 4061–4069.
- 121 W. D. Cotterill and R. Livingstone, *J. Chem. Soc.*, 1970, 1758–1764.
- 122 R. Livingstone, D. Miller and S. Morris, *J. Chem. Soc.*, 1960, 5148–5152.
- 123 J. Cottam and R. Livingstone, *J. Chem. Soc.*, 1964, 5228–5231.
- 124 J. D. Hepworth and B. M. Heron, in *Progress in Heterocyclic Chemistry*, Elsevier Ltd, 2005, vol. 17, pp. 33–62.
- 125 H. J. Kabbe, *Synthesis*, 1978, **1978**, 886–887.
- 126 J. J. Talley, *Synthesis*, 1983, **10**, 845–847.
- 127 H. G. Heller, J. R. Levell, D. E. Hibbs, D. S. Hughes and M. B. Hursthouse, *Mol. Cryst. Liq. Cryst.*, 1997, **297**, 123–130.
- 128 G. Sartori, G. Casiraghi, L. Bolzoni and G. Casnati, *J. Org. Chem.*, 1979, **44**, 803–805.
- 129 J.-L. Pozzo, V. A. Lokshin and R. Guglielmetti, *J. Chem. Soc., Perkin Trans. 1*, 1994, 2591–2595.
- 130 A. Ghatak, S. Khan and S. Bhar, *Adv. Synth. Catal.*, 2016, **358**, 435–443.
- 131 Y.-W. Dong, G.-W. Wang and L. Wang, *Tetrahedron*, 2008, **64**, 10148–10154.
- 132 M. Naga Venkata Sastry, S. Claessens, P. Habonimana and N. De Kimpe, *J. Org. Chem.*, 2010, **75**, 2274–2280.



- 133 S. Delbaere, J.-C. Micheau, M. Frigoli and G. Vermeersch, *J. Org. Chem.*, 2005, **70**, 5302–5304.
- 134 T. Kosuge, X. Zhu, V. M. Lau, D. Aoki, T. J. Martinez, J. S. Moore and H. Otsuka, *J. Am. Chem. Soc.*, 2019, **141**, 1898–1902.
- 135 M. E. McFadden and M. J. Robb, *J. Am. Chem. Soc.*, 2019, **141**, 11388–11392.
- 136 C. D. Gabbutt, B. M. Heron, D. A. Thomas, M. E. Light and M. B. Hursthouse, *Tetrahedron Lett.*, 2004, **45**, 6151–6154.
- 137 K. Arai, Y. Kobayashi and J. Abe, *Chem. Commun.*, 2015, **51**, 3057–3060.
- 138 C. J. Blackwell, C. D. Gabbutt, J. T. Guthrie and B. M. Heron, *Dyes Pigm.*, 2012, **95**, 408–420.
- 139 M. Frigoli, C. Moustrou, A. Samat and R. Guglielmetti, *Helv. Chim. Acta*, 2000, **83**, 3043–3052.
- 140 R. Demadrille, C. Moustrou, A. Samat and R. Guglielmetti, *Heterocycl. Commun.*, 1999, **5**, 123–126.
- 141 W. Sriprom, M. Néel, C. D. Gabbutt, B. M. Heron and S. Perrier, *J. Mater. Chem.*, 2007, **17**, 1885–1893.
- 142 N. Malic, J. A. Campbell and R. A. Evans, *Macromolecules*, 2008, **41**, 1206–1214.
- 143 S. K. Osler, M. E. McFadden and M. J. Robb, *J. Polym. Sci.*, 2021, **59**, 2537–2544.
- 144 M. E. McFadden and M. J. Robb, *J. Am. Chem. Soc.*, 2021, **143**, 7925–7929.
- 145 B. A. Versaw, M. E. McFadden, C. C. Husic and M. J. Robb, *Chem. Sci.*, 2020, **11**, 4525–4530.
- 146 M. E. McFadden, S. K. Osler, Y. Sun and M. J. Robb, *J. Am. Chem. Soc.*, 2022, **144**, 22391–22396.
- 147 S. K. Osler, M. E. McFadden, T. Zeng and M. J. Robb, *Polym. Chem.*, 2023, **14**, 2717–2723.
- 148 R. S. Becker, *US Pat.*, US3567605A, 1971.
- 149 L. Kortekaas and W. R. Browne, *Chem. Soc. Rev.*, 2019, **48**, 3406–3424.
- 150 S. Brazevic, S. Nizinski, R. Szabla, M. F. Rode and G. Burdzinski, *Phys. Chem. Chem. Phys.*, 2019, **21**, 11861–11870.
- 151 H. Görner and A. K. Chibisov, *J. Photochem. Photobiol., A*, 2002, **149**, 83–89.
- 152 G. Ottavi, G. Favaro and V. Malatesta, *J. Photochem. Photobiol., A*, 1998, **115**, 123–128.
- 153 S. Helmy and J. Read de Alaniz, *Adv. Heterocycl. Chem.*, 2015, **117**, 131–177.
- 154 S. Delbaere, B. Luccioni-Houze, C. Bochu, Y. Teral, M. Campredon and G. Vermeersch, *J. Chem. Soc., Perkin Trans. 2*, 1998, **2**, 1153–1157.
- 155 S. Jockusch, N. J. Turro and F. R. Blackburn, *J. Phys. Chem. A*, 2002, **106**, 9236–9241.
- 156 T. T. Herzog, G. Ryseck, E. Ploetz and T. Cordes, *Photochem. Photobiol. Sci.*, 2013, **12**, 1202–1209.
- 157 S. Brazevic, M. Sikorski, M. Sliwa, J. Abe, M. F. Rode and G. Burdzinski, *Dyes Pigm.*, 2022, **201**, 110249.
- 158 A. Migani, P. L. Gentili, F. Negri, M. Olivucci, A. Romani, G. Favaro and R. S. Becker, *J. Phys. Chem. A*, 2005, **109**, 8684–8692.
- 159 B. H. Strudwick, C. O'Bryen, H. J. Sanders, S. Woutersen and W. J. Buma, *Phys. Chem. Chem. Phys.*, 2019, **21**, 11689–11696.
- 160 J. Hobley, V. Malatesta, K. Hatanaka, S. Kajimoto, S. L. Williams and H. Fukumura, *Phys. Chem. Chem. Phys.*, 2002, **4**, 180–184.
- 161 J. C. Crano, T. Flood, D. Knowles, A. Kumar and B. Van Gemert, *Pure Appl. Chem.*, 1996, **68**, 1395–1398.
- 162 J. Hobley, V. Malatesta, R. Millini, W. Giroladini, L. Wis, M. Goto, M. Kishimoto and H. Fukumura, *Chem. Commun.*, 2000, 1339–1340.
- 163 F. Ortica, A. Romani, F. Blackburn and G. Favaro, *Photochem. Photobiol. Sci.*, 2002, **1**, 803–808.
- 164 G. Harié, A. Samat, R. Guglielmetti, D. De Keukeleire, W. Saeyens and I. Van Parys, *Tetrahedron Lett.*, 1997, **38**, 3075–3078.
- 165 G. Such, R. A. Evans, L. H. Yee and T. P. Davis, *J. Macromol. Sci., Part C: Polym. Rev.*, 2003, **43**, 547–579.
- 166 S. Brazevic, M. Baranowski, M. Sikorski, M. Rode and G. Burdzinski, *ChemPhysChem*, 2020, **21**, 1402–1407.
- 167 G. Favaro, A. Romani and R. S. Becker, *Photochem. Photobiol.*, 2000, **72**, 632–638.
- 168 M. Maafi and R. G. Brown, *Int. J. Chem. Kinet.*, 2005, **37**, 717–727.
- 169 M. Maafi and R. G. Brown, *Int. J. Chem. Kinet.*, 2006, **38**, 421–430.
- 170 M. Maafi, *Molecules*, 2008, **13**, 2260–2302.
- 171 S. Delbaere, G. Vermeersch and J. C. Micheau, *J. Photochem. Photobiol., C*, 2011, **12**, 74–105.
- 172 R. M. Christie, J. D. Hepworth, C. D. Gabbutt and S. Rae, *Dyes Pigm.*, 1997, **35**, 339–346.
- 173 C. Lenoble and R. S. Becker, *J. Photochem.*, 1986, **33**, 187–197.
- 174 S. Delbaere and G. Vermeersch, *Tetrahedron Lett.*, 2003, **44**, 259–262.
- 175 M. Maafi and R. G. Brown, *Int. J. Chem. Kinet.*, 2005, **37**, 162–174.
- 176 M. Maafi and R. G. Brown, *Int. J. Chem. Kinet.*, 2006, **38**, 431–438.
- 177 J. Berthet, S. Delbaere, D. Levi, P. Brun, R. Guglielmetti and G. Vermeersch, *J. Chem. Soc., Perkin Trans. 2*, 2002, 2118–2124.
- 178 J. Berthet, P. J. Coelho, L. M. Carvalho, G. Vermeersch and S. Delbaere, *J. Photochem. Photobiol., A*, 2009, **208**, 180–185.
- 179 S. Delbaere, J. C. Micheau and G. Vermeersch, *Org. Lett.*, 2002, **4**, 3143–3145.
- 180 S. Delbaere, J.-C. Micheau and G. Vermeersch, *J. Org. Chem.*, 2003, **68**, 8968–8973.
- 181 D. Venec, S. Delbaere, J. C. Micheau, M. Frigoli, C. Moustrou, A. Samat and G. Vermeersch, *J. Photochem. Photobiol., A*, 2006, **181**, 174–179.
- 182 R. G. Brown, M. Maafi, P. Foggi and L. Bussotti, in *Femtochemistry and Femtobiology: Ultrafast Events in Molecular Science*, 2004, pp. 283–286.
- 183 J. Aubard, F. Maurel, G. Buntinx, O. Poizat, G. Levi, R. Guglielmetti and A. Samat, *Mol. Cryst. Liq. Cryst.*, 2000, **345**, 215–220.



- 184 B. Moine, G. Buntinx, O. Poizat, J. Rehault, C. Moustrou and A. Samat, *J. Phys. Org. Chem.*, 2007, **20**, 936–943.
- 185 S. Brazevic, M. Sliwa, Y. Kobayashi, J. Abe and G. Burdzinski, *J. Phys. Chem. Lett.*, 2017, **8**, 909–914.
- 186 P. L. Gentili, A. Romani, R. S. Becker and G. Favaro, *Chem. Phys.*, 2005, **309**, 167–175.
- 187 P. L. Gentili, E. Danilov, F. Ortica, M. A. J. Rodgers and G. Favaro, *Photochem. Photobiol. Sci.*, 2004, **3**, 886–891.
- 188 G. Favaro, A. Romani and R. S. Becker, *Photochem. Photobiol.*, 2001, **74**, 378–384.
- 189 R. S. Becker, E. Dolan and D. E. Balke, *J. Chem. Phys.*, 1969, **50**, 239–245.
- 190 A. P. Demchenko, V. I. Tomin and P.-T. Chou, *Chem. Rev.*, 2017, **117**, 13353–13381.
- 191 M. Maafi and R. G. Brown, *Int. J. Chem. Kinet.*, 2007, **39**, 472–479.
- 192 S. Brazevic, S. Nizinski, M. Sliwa, J. Abe, M. F. Rode and G. Burdzinski, *Int. J. Mol. Sci.*, 2020, **21**, 7825.
- 193 A. Mannschreck, K. Lorenz and M. Schinabeck, in *Organic Photochromic and Thermochromic Compounds: Volume 2: Physicochemical Studies, Biological Applications, and Thermochromism*, ed. J. C. Crano and R. J. Guglielmetti, Springer US, Boston, MA, 2002, pp. 261–295.
- 194 B. Gierczyk, M. F. Rode and G. Burdzinski, *Sci. Rep.*, 2022, **12**, 10781.
- 195 F. Castet, R. Méreau and D. Liotard, *J. Chem. Educ.*, 2014, **91**, 924–928.
- 196 F. Maurel, S. Delbaere, S. L. Truong, T. Couesnon, R. Dubest, G. Vermeersch and J. Aubard, *J. Phys. Org. Chem.*, 2007, **20**, 944–952.
- 197 B. Luccioni-Houzé, M. Campredon, R. Guglielmetti and G. Giusti, *Mol. Cryst. Liq. Cryst.*, 1997, **297**, 161–165.
- 198 M. Rumi, T. J. White and T. J. Bunning, *J. Phys. Chem. B*, 2016, **120**, 12755–12767.
- 199 M. R. di Nunzio, P. L. Gentili, A. Romani and G. Favaro, *J. Phys. Chem. C*, 2010, **114**, 6123–6131.
- 200 G. Pan, W. Jingqiang, A. Zhu, M. Yangfu, F. Meigong and Y. Side, *Sci. China, Ser. B: Chem.*, 2001, **44**, 276–282.
- 201 P. Fan, G. Pan, J. Wei, Y. Ming, A. Zhu, M. G. Fan and W. M. Hung, *Mol. Cryst. Liq. Cryst.*, 2000, **344**, 283–288.
- 202 G. Favaro, F. Ortica, A. Romani and P. Smimmo, *J. Photochem. Photobiol., A*, 2008, **196**, 190–196.
- 203 Y. Goto, K. Sugita, T. Takahashi, Y. Ohga and T. Asano, *Chem. Lett.*, 2003, **32**, 618–619.
- 204 K. Sugita, Y. Goto, M. Ono, K. Yamashita, K. Hayase, T. Takahashi, Y. Ohga and T. Asano, *Bull. Chem. Soc. Jpn.*, 2004, **77**, 1803–1806.
- 205 M. Zayat and D. Levy, *J. Mater. Chem.*, 2003, **13**, 727–730.
- 206 R. Pardo, M. Zayat and D. Levy, *J. Mater. Chem.*, 2005, **15**, 703–708.
- 207 B. Van Gemert, A. Kumar and D. B. Knowles, *Mol. Cryst. Liq. Cryst.*, 1997, **297**, 131–138.
- 208 A. Kumar, B. Van Gemert and D. B. Knowles, *Mol. Cryst. Liq. Cryst.*, 2000, **344**, 217–222.
- 209 Y.-P. Chan and P. Jean, *US Pat.*, US6207084B1, 2001.
- 210 D. A. Clarke, B. M. Heron, C. D. Gabbutt, J. D. Hepworth, S. M. Partington and S. N. Corns, *WO Pat.*, WO2000018755A1, 2000.
- 211 D. A. Clarke, B. M. Heron, C. D. Gabbutt, J. D. Hepworth, S. M. Partington, and S. N. Corns, *WO Pat.*, WO1998042693A2, 1998.
- 212 B. Van Gemert, *WO Pat.*, WO1996014596A1, 1996.
- 213 B. Van Gemert, M. Bergomi and D. Knowles, *Mol. Cryst. Liq. Cryst.*, 1994, **246**, 67–73.
- 214 C. D. Gabbutt, B. M. Heron, A. C. Instone, P. N. Horton and M. B. Hursthouse, *Tetrahedron*, 2005, **61**, 463–471.
- 215 Z. Wang, Q. Meng, Z. Zhang, D. Fu and W. Zhang, *Tetrahedron*, 2011, **67**, 2246–2250.
- 216 C. M. Sousa, J. Berthet, S. Delbaere, A. Polónia and P. J. Coelho, *J. Org. Chem.*, 2017, **82**, 12028–12037.
- 217 C. M. Sousa, J. Berthet, S. Delbaere and P. J. Coelho, *Dyes Pigm.*, 2019, **169**, 118–124.
- 218 C. M. Sousa and P. J. Coelho, *Eur. J. Org. Chem.*, 2020, **2020**, 985–992.
- 219 P. J. Coelho, C. J. R. Silva, C. Sousa and S. D. F. C. Moreira, *J. Mater. Chem. C*, 2013, **1**, 5387–5394.
- 220 C. M. Sousa, J. Berthet, S. Delbaere and P. J. Coelho, *J. Org. Chem.*, 2012, **77**, 3959–3968.
- 221 C. M. Sousa, P. J. Coelho, G. Vermeersch, J. Berthet and S. Delbaere, *J. Photochem. Photobiol., A*, 2010, **216**, 73–78.
- 222 A. M. Gorelik, O. V. Venidiktova, O. I. Kobeleva, T. M. Valova and V. A. Barachevsky, *Dyes Pigm.*, 2021, **184**, 108833.
- 223 M. K. Beyer, *J. Chem. Phys.*, 2000, **112**, 7307–7312.
- 224 R. Stevenson and G. De Bo, *J. Am. Chem. Soc.*, 2017, **139**, 16768–16771.
- 225 M. Radiom, P. Kong, P. Maroni, M. Schäfer, A. F. M. Kilbinger and M. Borkovec, *Phys. Chem. Chem. Phys.*, 2016, **18**, 31202–31210.
- 226 W. Huang, Z. Zhu, J. Wen, X. Wang, M. Qin, Y. Cao, H. Ma and W. Wang, *ACS Nano*, 2017, **11**, 194–203.
- 227 C. R. Hickenboth, J. S. Moore, S. R. White, N. R. Sottos, J. Baudry and S. R. Wilson, *Nature*, 2007, **446**, 423–427.
- 228 J. M. Lenhardt, M. T. Ong, R. Choe, C. R. Evenhuis, T. J. Martinez and S. L. Craig, *Science*, 2010, **329**, 1057–1060.
- 229 J. Wang, T. B. Kouznetsova, Z. Niu, M. T. Ong, H. M. Klukovich, A. L. Rheingold, T. J. Martinez and S. L. Craig, *Nat. Chem.*, 2015, **7**, 323–327.
- 230 M. T. Ong, J. Leiding, H. Tao, A. M. Virshup and T. J. Martínez, *J. Am. Chem. Soc.*, 2009, **131**, 6377–6379.
- 231 J. Ribas-Arino, M. Shiga and D. Marx, *Angew. Chem., Int. Ed.*, 2009, **48**, 4190–4193.
- 232 A. G. Roessler and P. M. Zimmerman, *J. Phys. Chem. C*, 2018, **122**, 6996–7004.
- 233 N. H. Nguyen, B. M. Rosen, G. Lligadas and V. Percec, *Macromolecules*, 2009, **42**, 2379–2386.
- 234 W. Zhao and E. M. Carreira, *J. Am. Chem. Soc.*, 2002, **124**, 1582–1583.
- 235 D. Kim, M. S. Kwon and C. W. Lee, *Polym. Chem.*, 2022, **13**, 5177–5187.



- 236 F. Ortica, C. Moustrou, J. Berthet, G. Favaro, A. Samat, R. Guglielmetti, G. Vermeersch and U. Mazzucato, *Photochem. Photobiol.*, 2003, **78**, 558–566.
- 237 P. J. Coelho, M. A. Salvador, B. M. Heron and L. M. Carvalho, *Tetrahedron*, 2005, **61**, 11730–11743.
- 238 C. D. Gabbutt, B. M. Heron, A. C. Instone, S. B. Kolla, K. Mahajan, P. J. Coelho and L. M. Carvalho, *Dyes Pigm.*, 2008, **76**, 24–34.
- 239 S. Aiken, C. D. Gabbutt, B. M. Heron and S. B. Kolla, *Dyes Pigm.*, 2015, **113**, 239–250.
- 240 J. A. Odell and A. Keller, *J. Polym. Sci., Part B: Polym. Phys.*, 1986, **24**, 1889–1916.
- 241 Y. Liu, S. Holm, J. Meisner, Y. Jia, Q. Wu, T. J. Woods, T. J. Martinez and J. S. Moore, *Science*, 2021, **373**, 208–212.

