

Cite this: *J. Mater. Chem. B*, 2023,
11, 2334

Surface modification of TiO₂ nanoparticles with organic molecules and their biological applications

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In recent years, titanium(IV) dioxide nanoparticles (TiO₂NPs) have shown promising potential in various biological applications such as antimicrobials, drug delivery, photodynamic therapy, biosensors, and tissue engineering. For employing TiO₂NPs in these fields, their nanosurface must be coated or conjugated with organic and/or inorganic agents. This modification can improve their stability, photochemical properties, biocompatibility, and even surface area for further conjugation with other molecules such as drugs, targeting molecules, polymers, etc. This review describes the organic-based modification of TiO₂NPs and their potential applications in the mentioned biological fields. In the first part of this review, around 75 recent publications (2017–2022) are mentioned on the common TiO₂NP modifiers including organosilanes, polymers, small molecules, and hydrogels, which improve the photochemical features of TiO₂NPs. In the second part of this review, we presented 149 recent papers (2020–2022) about the use of modified TiO₂NPs in biological applications, in which specific bioactive modifiers are introduced in this part with their advantages. In this review, the following information is presented: (1) the common organic modifiers for TiO₂NPs, (2) biologically important modifiers and their benefits, and (3) recent publications on biological studies on the modified TiO₂NPs with their achievements. This review shows the paramount significance of the organic-based modification of TiO₂NPs to enhance their biological effectiveness, paving the way toward the development of advanced TiO₂-based nanomaterials in nanomedicine.

Received 25th November 2022,
Accepted 24th January 2023

DOI: 10.1039/d2tb02576k

rsc.li/materials-b

1. Introduction

With the advent of nanotechnology, numerous nanomaterials have been synthesized and applied for various applications and among them, titanium dioxide nanoparticles (TiO₂NPs) are commonly used in the fields of biomedicine,¹ food industry,² wastewater purification,³ and cosmetics⁴ owing to their unique physicochemical properties such as high chemical stability and photodynamic effects. Regarding their industrial importance, the global market size of TiO₂NPs was estimated to be \$1.1 billion in 2021, and forecasted to have a compound annual growth rate of 6.5% until 2026, with annual production

predicted to reach 2.5 million tons by 2025.^{5,6} In the biomedical fields, TiO₂NPs are frequently studied for photodynamic therapy,^{1,7} drug delivery,⁸ antimicrobial applications,^{3,9–12} biosensors,¹³ and tissue engineering.¹⁴ For these applications, an appropriate surface modification of TiO₂NPs is required to improve their physicochemical properties and biological effectiveness and, more importantly, decrease their potential toxicity in mammalian cells.⁵ The surface modification not only prevents the agglomeration of TiO₂NPs but also provides the possibility for further functionalization/conjugation. The modification of TiO₂NPs can be achieved using two different approaches: non-covalent and covalent conjugation of organic (or inorganic) species with TiO₂NPs. The non-covalent strategy is based on physical interactions (electrostatic, hydrogen bond, van der Waals, and hydrophobic interactions), having benefits of being relatively simple and not changing the structure of the modifiers. However, this type of modification can be easily influenced by different external stimuli, such as temperature, pH, and ionic strength.¹⁵ On the other side, the covalent modification (or chemical modification) occurs *via* covalent

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bonding of modifiers to the TiO_2NP surface which can be performed using various coupling agents such as polymers, organophosphorus molecules, carboxylic acids, and organosilanes, and among them, silane compounds are more common for the surface modification of TiO_2NPs .¹⁶ In recent years, there has been increasing interest for these two types of modification strategies and this review presents recent publications of organic-based TiO_2NP modification, followed by the presentation of those organic modifiers which were used for the biomedical applications with their advantages.

2. Surface chemistry of TiO_2NPs

Like other types of nanoparticles, TiO_2NPs have two distinct atoms: (1) internal and (2) surface atoms. The internal part of TiO_2NPs is chemically inert (each Ti atom has four chemical bonds with four neighboring oxygens) and so, the remaining surface atoms are mainly responsible for the interaction of nanoparticles with the environment. The surface atoms are not chemically saturated (they are only attached to the internal atoms), so they should complete their coordination number to gain a stable electronic configuration. Ti and O are considered as hard atoms based on the HSAB concept (hard and soft (Lewis) acids and bases)⁸ and they tend to interact with the hard atoms. Due to the adsorption of water molecules from the environment, the surface of TiO_2NPs has two main OH groups: (1) Ti-OH and (2) Ti-O_{br}H-Ti (br: bridging) (Fig. 1).^{17–21} The interaction of TiO_2NPs with the surroundings occurs through these two types of OH groups.

3. Surface modification of TiO_2NPs

Organic stabilizers can bind to the TiO_2NP surface by either physical or chemical interactions. The chemical stabilization (or covalent stabilization) is much stronger than the physical one and, in most cases, it results in long-term stability for the modified TiO_2NPs .²² In the following sections, the common surface stabilizers of TiO_2NPs will be presented into two groups of (1) organofunctional silanes and (2) polymers, small molecules, and hydrogels. In the structure of silane modifiers, there are active functional groups (for example hydrolysable alkoxy groups) which are suitable for the chemical interaction with TiO_2NPs .²³ On the other hand, physical modification can be

carried out by using these organic molecules which are adsorbed on the TiO_2NP surface by electrostatic, hydrogen bond, van der Waals, and hydrophobic interactions.²⁴

3.1. Organofunctional silanes

3.1.1. Tetraethoxysilane (TEOS). Silica coating is a common chemical method for the surface modification of TiO_2NPs , providing several benefits such as long-term stability, biocompatibility, and hydrophilicity of silane-modified TiO_2NPs ($\text{TiO}_2\text{NPs}@SiO_2$). There are standard procedures to control the thickness of the silica layer and four main approaches are commonly used to prepare $\text{TiO}_2\text{NPs}@SiO_2$ (Table 1).²⁵

The Stöber method is the most employed method for the silica modification of bare TiO_2NPs . In the standard Stöber approach, TiO_2NPs are uniformly dispersed in an ethanol solution, followed by the addition of tetraethoxysilane (TEOS) and aqueous ammonia solution ($\text{NH}_3(\text{aq})$), respectively.²⁶ Ammonia acts as a basic catalyst to control TEOS hydrolysis and silica thickness to form particles with a regular morphology (see Fig. 2). In this hydrolysis reaction, the $-\text{Si}-\text{OC}_2\text{H}_5$ groups of TEOS convert to silanol groups ($-\text{Si}-\text{OH}$), and then, the condensation reaction occurs between these $-\text{Si}-\text{OH}$ groups and the surface $-\text{OH}$ groups of TiO_2NPs to form chemical Ti-O-Si bonds. This sol-gel reaction results in a 3D silica network around the TiO_2NP core. Chen *et al.* applied the Stöber process to have varying thicknesses of SiO_2 onto the surface of anatase and rutile TiO_2NPs .^{27,28} They reported an enhancement of the photocatalytic activity of the modified TiO_2NPs when the SiO_2 loading weight was lower than 3.25 wt%, while with higher loading percentages, lower photocatalytic activity was observed. Regarding the rutile phase, the complete coverage of TiO_2NPs with SiO_2 resulted in an enhancement of the photocatalytic activity.

The second approach of silica modification is the micro-emulsion method, having two different types: (1) water-in-oil (W/O, normal micelles) and (2) oil-in-water (O/W, reverse micelles). Using this method, Xie *et al.* prepared monodisperse $\text{TiO}_2\text{NPs}@SiO_2$ core-shell particles and showed that the contents of the anatase and rutile crystalline phases of these TiO_2NPs were decreased and increased, respectively, when the temperature was increased from 550 °C to 650 °C. In the temperature range of 600–800 °C, the $\text{TiO}_2\text{NPs}@SiO_2$ particles were mainly anatase.²⁹

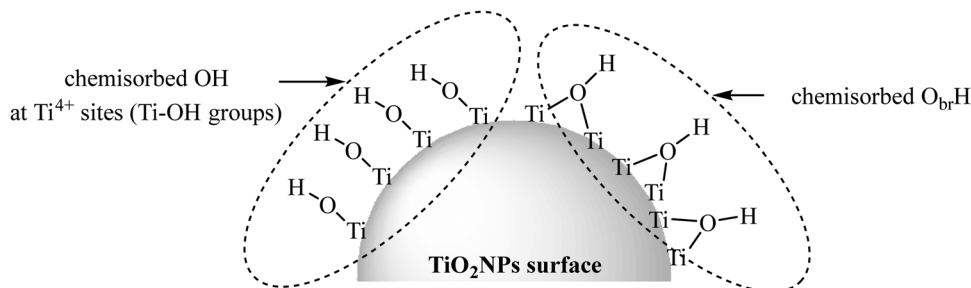


Fig. 1 Two different OH groups of the TiO_2NP surface. Adapted with permission from ref. 20. Copyright 2014 American Chemical Society.



Table 1 Four main methods for the silica coating of TiO₂NPs.²⁵

| Synthesis methods | Advantages | Disadvantages |
|---|---|--|
| Stöber method | Controllable silica shell and TiO ₂ monodispersity | Lack of understanding of its kinetics and mechanism |
| Microemulsion | Control of the TiO ₂ NP size and high homogeneity | Poor yield, time consuming and large amounts of solvent required |
| Aerosol pyrolysis | Highly productive and suitable for large-scale production | Complex experimental conditions |
| Methods based on sodium silicate solution | Control of the crystallinity and surface area | Depends on the preparation method |

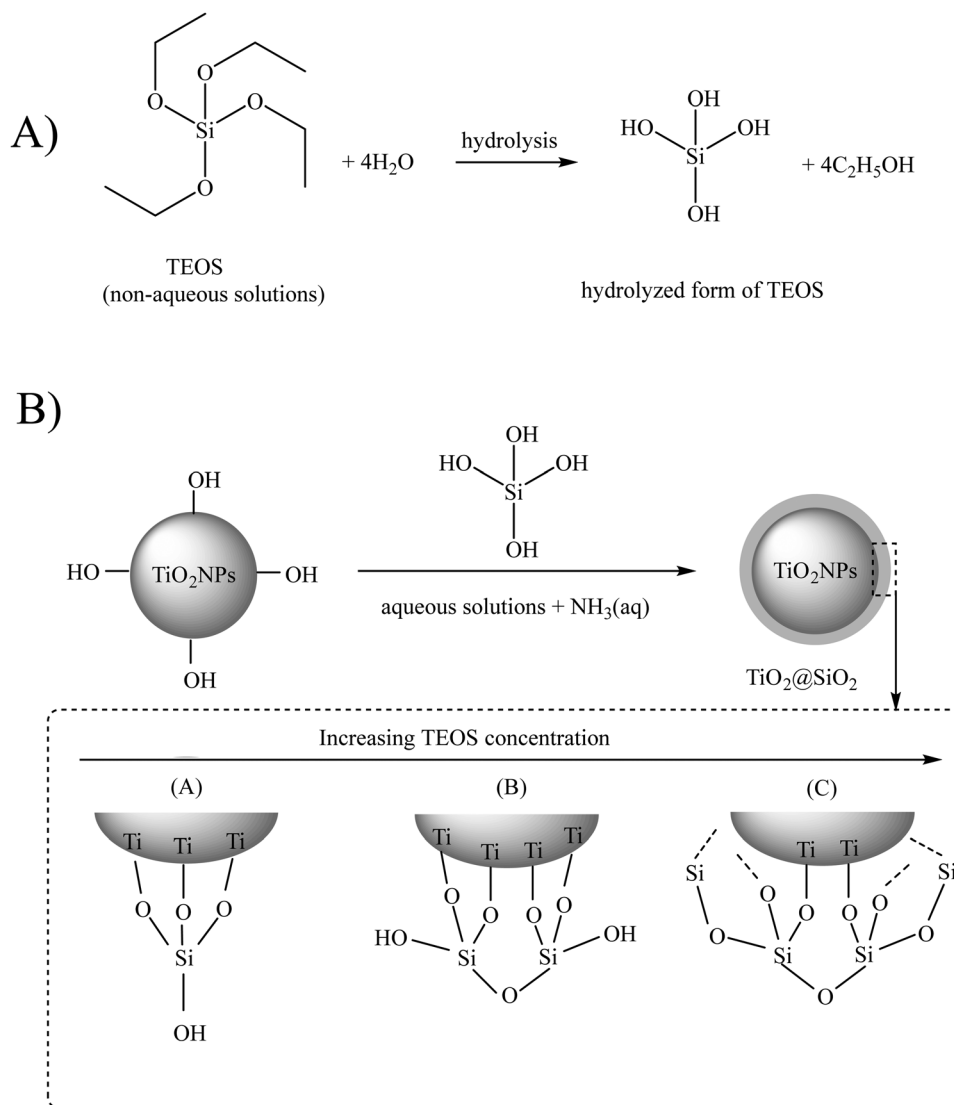


Fig. 2 (A) Hydrolysis and (B) condensation reactions of tetraethoxysilane (TEOS). Adapted with permission from ref. 22. Royal Society of Chemistry.

The third strategy to synthesize TiO₂NPs@SiO₂ particles is aerosol pyrolysis, considered as an innovative and productive approach, which is usually carried out in a flame environment and can be used for the large-scale production of modified TiO₂NP powders. In 2021, Temerov *et al.* synthesized TiO₂NPs@SiO₂ (50–70 nm) using a liquid flame spray (LFS) deposition method in a single flame environment.³⁰ They

studied the photocatalytic activity of deposited TiO₂NPs@SiO₂ for oxidation of acetylene into carbon dioxide and they investigated the effect of the silica shell on the photocatalytic activity of these modified TiO₂NPs. They reported that the catalytic activity was significantly suppressed when the SiO₂ content was increased to 0.5%, 1.0%, 3.0% and 5.0% (33%, 44%, 70% and 100% of suppression, respectively). They mentioned that this



suppression might be due to the thick passivating silica layer around the TiO₂NP core. Maskrot *et al.* synthesized a core-shell TiO₂NPs@SiO₂ composite with different Ti/Si ratios, by the laser pyrolysis of a gas-spray mixture of TEOS and titanium tetra-isopropoxide.³¹ By increasing the Ti/Si ratio, the color of these modified TiO₂NPs@SiO₂ composite changes from dark to light blue. Their results showed the correlation between the chemical composition and the size of these TiO₂NPs@SiO₂ nanoparticles as a function of the Ti/Si ratio.

The fourth route is based on sodium silicate solution as a cheap silica precursor. For instance, Shao *et al.* used sodium silicate to prepare the TiO₂-SiO₂ composites using controllable and reproducible approaches to improve the textural properties of the nanostructures.³² The practical photocatalytic application of these TiO₂-SiO₂ composites was successfully tested for decolorization of methylene blue, as a model pollutant in textile industries.

3.1.2. Bifunctional silane coupling agents (trialkoxysilanes). In the previous section, it was mentioned that the TEOS silica coating provides modified TiO₂NPs@SiO₂ having -OH

groups on the outer part of the nanosurface (see Fig. 2), which come from the silica shell. However, for some specific biological applications, it is necessary to introduce new functional groups on the surface of modified TiO₂NPs for retaining the benefits of the silica coating as well. For this goal, trialkoxysilanes are one of the most important surface modifiers which can provide both biocompatibility and long-term colloidal stability for the particles. More importantly, these bifunctional silanes supply suitable functional groups on the surface of silane-modified TiO₂NPs for the further attachment of other species to the nanoparticles. The commercially available trialkoxysilanes, (RO)₃Si-(CH₂)_n-X (X = -NH₂, -SH, -C=C, epoxy, *etc.*; n = typically 3), are considered as effective bifunctional silane linkers having two different functional groups in their structures including: (1) -OR moiety (attached to the -Si) and (2) -SH/or -NH₂ (or other functional groups) attached to the end of a carbon chain.²² In the general formulation of (RO)₃Si-(CH₂)_n-X, R can be an alkyl, aryl or generally organofunctional group. As shown in Fig. 3, for the surface modification of TiO₂NPs, the

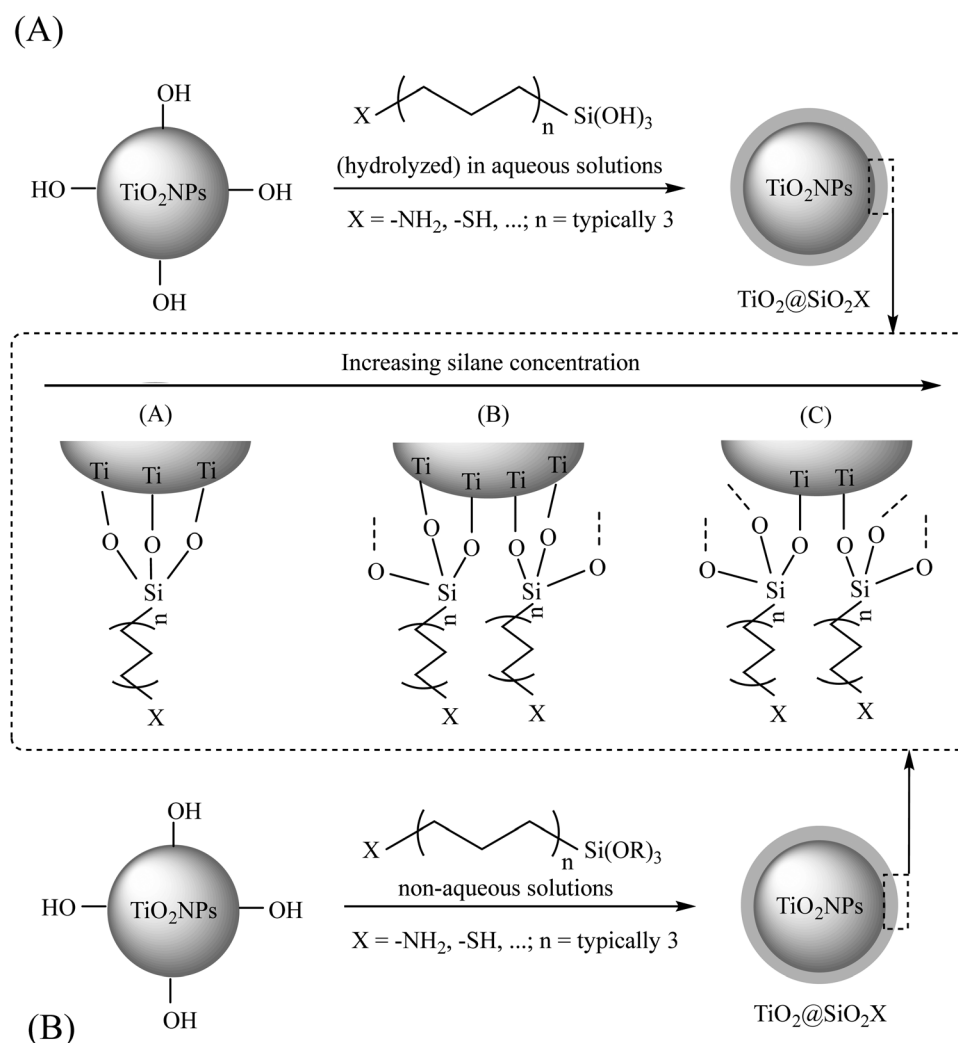


Fig. 3 The interaction between a general silane coupling agent and TiO₂NPs in aqueous (A) and non-aqueous (B) solutions. Adapted with permission from ref. 22. Royal Society of Chemistry.



–OR groups should be hydrolyzed to form silanol groups (–SiOH), followed by the condensation of these silanols with the –OH groups of the bare TiO₂ nanosurface, resulting in the formation of a silica network around the TiO₂NP core and providing the suitable X functional groups onto the surface. As common organosilane coupling agents, 3-methacryloxypropyltrimethoxysilane (MPS),³³ 3-aminopropyltriethoxysilane (APTES),^{34–38} 3-glycidioxypropyltrimethoxysilane (GPS),³⁹ and *n*-propyltriethoxysilane²² are used for different applications. The other recommendations for this type of modification are shown in Table 2.

Because of the importance of such bifunctional silane linkers, recent publications on some of these triorganosilanes will be presented as follows:

3.1.3. (3-Aminopropyl)triethoxysilane (APTES). As one of the most important trialkoxysilanes, APTES can provide –NH₂ groups onto the outer surface of modified TiO₂NPs. In 2022, Mokhtari Aghdami *et al.* functionalized TiO₂NPs with APTES to improve the thermal, mechanical, and antibacterial properties of poly(lactic acid)/silicone rubber (PLA/SR) blends, reinforced with APTES-functionalized TiO₂NPs.⁵³ In another recent study, Massoumi *et al.* immobilized *S*-nitroso-*N*-acetylpenicillamine (SNAP) on TiO₂NPs to form a NO-releasing TiO₂NP/SNAP system, for antibacterial applications. For this goal, TiO₂NPs were first silanized with APTES, and then *N*-acetyl-*D*-penicillamine was grafted to them *via* an amide bond.⁵⁴ TiO₂NPs@APTES-SNAP showed antimicrobial properties (5 mg mL^{–1}) on both Gram-positive *Staphylococcus aureus* (*S. aureus*) and Gram-negative *Escherichia coli* (*E. coli*) bacteria and the biocompatibility on 3T3 mouse fibroblast cells, at all tested concentrations. Dymerska *et al.* proposed a nanocomposite made from ultrasmall TiO₂NPs embedded in a silica/carbon matrix.⁵⁵ This composite was fabricated first by the surface modification of TiO₂NPs with APTES, followed by the encapsulation of the TiO₂NPs@APTES in a carbon sphere-like structure formed with biocompatible glucose as the carbon precursor. The resultant system exhibited high photocatalytic activity for degradation of methylene blue as a model hazardous organic dye. Yoo *et al.* synthesized highly pure TiO₂NPs through thermal plasma and deposited them directly on an interdigitated electrode.⁵⁶ The nanosurface of the TiO₂-deposited electrode was modified with APTES, followed by attaching a single-stranded probe deoxyribonucleic acid (DNA) to construct a DNA biosensor for the selective detection of *E. coli* O157:H7, as a well-known pernicious pathogenic bacterial species. Zhang *et al.* developed a novel electrode material for supercapacitors, in which they used APTES to link polyaniline (PANI) to TiO₂ nanowires (TNWs).⁵⁷ Compared to PANI–TNW, the better capacitive properties of PANI–APTES–TNW were observed due to the anchoring effect of APTES, which was highly interactive and showed compact structures between the TNW nanoparticles and PANI, resulting in a stable structure during the rapid charge–discharge process. Wanag *et al.* reported TiO₂NP modification with APTES utilizing a solvothermal method, with different concentrations of APTES (10–3000 mM).⁵⁸ The photocatalytic activity of these TiO₂NPs@APTES was successfully tested for methylene blue decomposition under UV light irradiation.

Their results showed that the photocatalytic properties enhanced with the increase of the APTES concentration. Mallakpour and Barati reported the APTES modification of TiO₂NPs and showed the improvement of the heat resistance of these modified TiO₂NPs.⁵⁹ Shakeri *et al.* synthesized a stable TiO₂NPs@APTES nanohybrid for the photocatalytic degradation of a pink food dye used as an organic pollutant.⁶⁰ Klaysri *et al.* reported a one-step synthesis method of APTES-functionalized TiO₂NPs for the photocatalytic decolonization of methylene blue.⁶¹

3.1.4. (3-Mercaptopropyl)trimethoxysilane (MPTMS). MPTMS is another interesting silane coupling agent that can provide free –SH groups on the surface of modified TiO₂NPs. This trimethoxysilane is specifically suitable for the chemical attachment of nanoparticles such as Au- and AgNPs to TiO₂NPs. For instance, Meng *et al.* employed MPTMS to functionalize commercial TiO₂NPs with the silica layer and, more importantly, provide the surface –SH groups to decorate AgNPs on the modified TiO₂NPs.⁴⁷ They synthesized TiO₂NPs@MPTMS-AgNPs for the catalytic reduction of 4-nitrophenol, as a chemical pollutant. Lee *et al.* reported the surface modification of the photocatalyst TiO₂ using MPTMS, on a nylon fabric surface.⁶² They showed that the antimicrobial and photodegradation properties of this nylon fabric were improved for the treatment of *S. aureus* and *E. coli* strains on the treated nylon containing the modified TiO₂@MPTMS.

3.1.5. Vinyltriethoxysilanes (VTES) and other non-silane coupling agents. In some applications it is necessary to introduce –C=C on the TiO₂NP surface, and vinyltriethoxysilane (VTES) can be an excellent recommendation in this regard. Aqeel Ashraf *et al.* successfully used VTES and TEOS to modify TiO₂NPs (*via* a hybrid sol–gel coating) to protect the commercial AZ91 magnesium alloy against corrosion in a 0.05 M NaCl solution.⁶³ Tangchantra *et al.* modified TiO₂NPs with three different silane coupling agents, such as VTES, hexadecyltrimethoxysilane (HTMS), and aminopropyltrimethoxysilane (APTMS).⁶⁴ Their results demonstrated that the VETS modification could improve the dispersibility and mechanical properties of the TiO₂NPs. Yang *et al.* reported the VTES silanization of TiO₂ particles (*via* a sol–gel method) and their results exhibited the improvement of colloidal stability in tetrachloroethylene solvent.⁶⁵

Regarding the other types of silanes and non-silane coupling agents, there are several worthwhile publications which are briefly mentioned here. For example, Caris *et al.* utilized conventional emulsion polymerization to encapsulate TiO₂ in poly(methyl methacrylate) (PMMA).⁶⁶ Weng and Wei studied the radical polymerization of styrene and methyl methacrylate (MMA), initiated at the surface of TiO₂ particles by adsorbed hydroperoxide macroinitiators.⁶⁷ Erdem *et al.* modified TiO₂NPs by the miniemulsion polymerization of styrene and polybutene–succinimide pentamine being used as the stabilizer at the oil/water interface.⁶⁸ Rong *et al.* reported the modification of TiO₂NPs by (3-trimethoxysilyl)propylmethacrylate, followed by the free-radical copolymerization of styrene with the methacrylate group of 3-methacryloxypropyltrimethoxysilane (MPS).⁶⁹



Table 2 Some trialkoxysilane agents and other organic functionalizing molecules containing the active functional groups

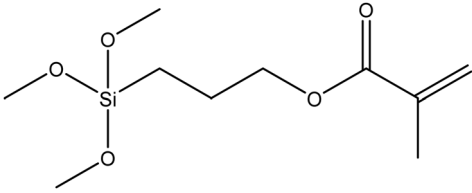
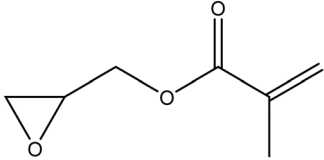
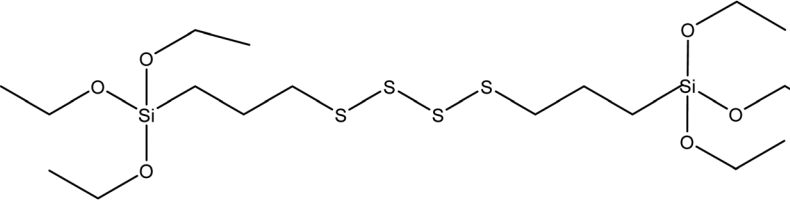
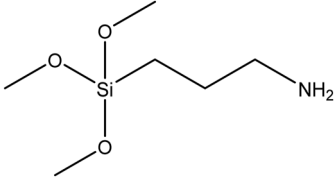
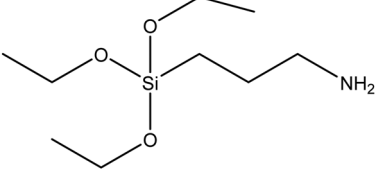
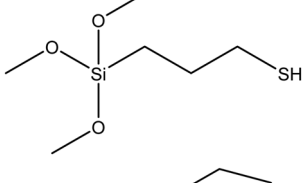
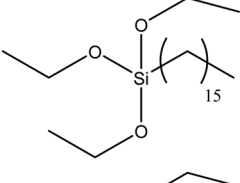
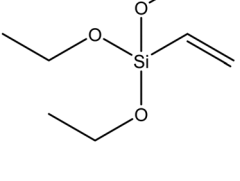
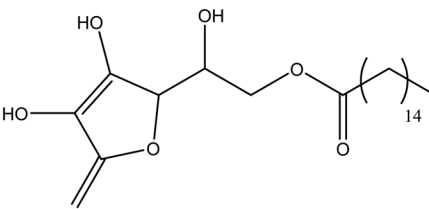
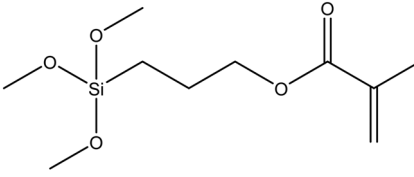
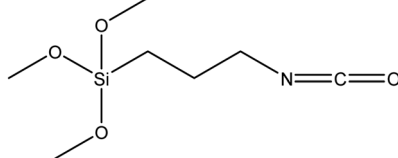
| Modifying agent | Structure | Ref. |
|--|--|-----------|
| (3-Trimethoxysilyl)propyl methacrylate, KH-570 |  | 40 |
| Fluorosilane | $\text{H}_3\text{Si-F}$ | 41 |
| Glycidyl methacrylate |  | 42 |
| Bis-(3-triethoxysilylpropyl)tetrasulfide (TESPT) |  | 43 |
| (3-Aminopropyl)trimethoxysilane |  | 44 and 45 |
| (3-Aminopropyl)triethoxysilane (APTES) |  | 46 |
| (3-Mercaptopropyl)triethoxysilane (MPTES) |  | 47 |
| Hexadecyltrimethoxysilane |  | 48 |
| Vinyltrimethoxysilane (VTMS) |  | 49 |



Table 2 (continued)

| Modifying agent | Structure | Ref. |
|--|--|------|
| Ascorbic acid 6-palmitate |  | 50 |
| (3-Methacryloxypropyl)trimethoxysilane |  | 51 |
| 3-Isocyanato propyl trimethoxysilane |  | 52 |

Yang and Dan used a similar approach to attach poly(methyl methacrylate) on the modified surface of TiO₂NPs.⁷⁰ Milanesi *et al.* employed a mixture of isomeric octyltriethoxysilanes (OTESs) to form a hydrophobic layer around TiO₂NPs.⁷¹ They reported the formation of cross-linked and chemical bonded Ti–O–Si onto the modified TiO₂NPs. Xiang *et al.* used MPS to modify the TiO₂NP surface and enhance their compatibility with the poly(butyl acrylate) (PBA) matrix.⁷² In another study, Qi *et al.* synthesized hydrophobic TiO₂NPs using the acrylonitrile–styrene–acrylate (ASA) terpolymer for cool materials.⁵¹ Wang *et al.* functionalized commercial TiO₂NPs with MPS *via* ultrasonic treatment at room temperature.⁷³ Godnjavec *et al.* coated TiO₂NPs by 3-glycidyloxypropyltrimethoxysilane (GLYMO) as an additive in a clear polyacrylic coating and reported that the modified TiO₂NPs improved dispersion, transparency, and UV protection of the clear acrylic coating.⁷⁴ Dalod *et al.* modified TiO₂NPs with APTES, 3-(2-aminoethylamino)propyldimethoxymethylsilane (AEAPS), and *n*-decyltriethoxysilane (DTES) using a hydrothermal method and reported that the shape and structure of these nanoparticles depend on the type of silane coupling groups.⁷⁵

3.2. Polymers, small molecules, and hydrogels

3.2.1. Polymers. In recent years, the polymeric modification of TiO₂NPs has attracted growing attention owing to their widespread application in various nanomedicinal fields. *In situ* coating and post (*ex situ*) surface coating are two common methods for the polymeric modification of TiO₂NPs. For *in situ* coating, the polymer is used simultaneously with the TiO₂ precursor during the synthesis of TiO₂NPs, and both synthesis and modification occur simultaneously in a single step. However, for the post (*ex situ*) modification of TiO₂NPs, the polymers are added to pre-synthesized TiO₂NPs, which is a separate step (next step) from the synthesis. To date, polyethylene glycol

(PEG), dextran, chitosan, alginate, polyvinyl alcohol (PVA), polydopamine (PDA), polysaccharides, polyethyleneimine (PEI), polyvinylpyrrolidone (PVP), polyetherimide, and polyamidoamine (PAMAM) have been applied for the surface modification of TiO₂NPs (Table 3).

PEG is a commonly used water-soluble polymer for the surface modification of TiO₂NPs, which can enhance the biocompatibility and hydrophilicity of the nanoparticles for biological applications. Recently, several excellent research studies have been reported on the PEG-coated TiO₂NPs; for example, in 2022, Connolly *et al.* compared the bioaccumulation, biodistribution and depuration profile of uncoated TiO₂NPs and PEG-modified TiO₂NPs in rainbow trout, after 10 days dietary exposure and a 42 day depuration phase.⁷⁶ Their results showed that PEG modification had an influence on levels of uptake and distributions of the modified TiO₂NPs, and a higher uptake of PEG-coated TiO₂NPs was observed, compared to the fish exposed to the uncoated TiO₂NPs. Tsotetsi *et al.* synthesized TiO₂NPs and then modified their surface with PEG, polyvinylpyrrolidone (PVP), and Pluronic F127 as pore forming agents, to investigate the effects of surface modification on the pore size, morphology, specific surface area, and optical properties of the TiO₂NPs.⁷⁷ All these three modified samples showed porous morphologies with spherical shapes and specific surface areas of ~69.82, 37.80 and 57.08 m² g⁻¹ for TiO₂-F127, TiO₂-PVP and TiO₂-PEG, respectively (after calcination at 550 °C). The pore sizes were estimated to be ~13.01, 10.10 and 8.53 nm for TiO₂-F127, TiO₂-PVP and TiO₂-PEG, respectively. Their results indicated that the surface modification of bare TiO₂NPs can improve their photophysical properties to act as an efficient electron transporting layer in solar cell applications. Koushali *et al.* studied the effects of synthesized TiO₂-PEG on the morphological, thermal, and mechanical properties of unsaturated polyester (UPE) nanocomposites.⁷⁸



Table 3 Main polymers used for TiO₂NP modification

| Polymers | Source/production/preparation ²⁵ | Ref. |
|----------------------------|---|-------------|
| Polyethylene glycol (PEG) | Produced by the interaction of ethylene oxide with water, ethylene glycol, or ethylene glycol oligomers | 76–85 |
| Polyvinylpyrrolidone (PVP) | Made from the monomer <i>N</i> -vinylpyrrolidone | 86 |
| Polyethyleneimine (PEI) | Branched PEI: by the ring opening polymerization of aziridine Linear PEI: by the post-modification of other polymers like poly(2-oxazolines) or <i>N</i> -substituted polyaziridines | 87 |
| Polyacrylic acids (PAA) | Polymerization of acrylic acid | 88 and 89 |
| Polyvinyl alcohol (PVA) | Polymerization of vinyl acetate and then the saponification of polyvinyl acetate | 90 and 91 |
| Polydopamine (PDA) | Formed from dopamine at slightly basic pH | 92 |
| Dextran | Produced by lactic acid bacteria | 93 |
| Chitosan | Extracted from shellfish or the fungal cell wall | 94–97 |
| Starch | Produced by green plants | 98 and 99 |
| Alginate | Extracted from brown algae | 100 and 101 |
| Polyphenol | Found in some common plant foods like cocoa, beans, tea, and vegetables | 102 |
| Amino acids | In nature | 103 |
| Flavonoids | Found in some common plant foods like fruits, vegetables, beans, and tea | 104 and 105 |

The UPE/PEG/TiO₂ nanocomposites were prepared by direct mechanical mixing of these three components at different weight ratios of both TiO₂NPs and PEG. Their results showed an improvement in the mechanical and thermal characteristics of the nanocomposites containing 0.5 wt% of synthesized TiO₂NPs and 10 wt% of PEG, compared to the pristine polyester. Abasifard Dehkordi *et al.* studied the addition of TiO₂/ZnO nanoparticles and PEG (with different molar ratios) to Portland cement to improve the photocatalytic and antibacterial activities of the cement.⁷⁹ They evaluated the potential ability of this composite for decolorization of an azo dye (as an organic pollutant) and inactivation of *E. coli* and *S. aureus* mutants. Their results showed a concentration-dependence antibacterial effect of the modified cement and effective photocatalytic degradation of the dye, which showed promising potential of this modified cement to be used as a self-cleaning and antibacterial coating for urban constructions. Iqbal *et al.* deposited gold nanoparticles onto TiO₂NPs, followed by the PEG modification of the TiO₂-Au nanohybrid.¹⁰⁶ The authors used this nanohybrid as a novel photosensitizing agent after biodistribution toward the targeted site (cancerous cell injury in the MCF-7 cell line), and showed that the different morphologies of PEG-modified Au-doped TiO₂NPs provided various therapeutic effects. Landolsi *et al.* synthesized TiO₂NPs and then modified them with PEG, followed by decoration of Fe₂O₃NPs onto the surface of modified TiO₂NPs.⁸¹ The photocatalytic activity of the TiO₂NPs-PEG-Fe₂O₃NPs was studied for the degradation of methylene blue and the significant degradability performance was observed under visible light irradiation (up to 70% after 150 min of irradiation). Bai *et al.* synthesized ultrasmall iron-doped TiO₂ nanodots (Fe-TiO₂ NDs) as a type of sonosensitizers to study their potential applications in sonodynamic therapy.⁸² After PEG modification, Fe-TiO₂-PEG NDs showed improved physiological stability and biocompatibility, with efficient tumor retention. Compared to commercial TiO₂NPs, this modified TiO₂ nanostructure demonstrated higher *in vivo* therapeutic performance with no long-term toxicity to the treated mice

after one month. Birinci *et al.* used PEG-modified TiO₂NPs in the formulation of a novel nano-antioxidant that utilized quercetin-conjugated TiO₂NPs (QTiO₂, quercetin is a potent antioxidant) for fortifying skin defense against oxidative toxicity.⁸³ The PEG-modified TiO₂NPs exhibited better colloidal stability and biocompatibility (compared to the unmodified TiO₂NPs), which causes an easy adhesion of these nanosurfaces onto living cells. This nano-antioxidant QTiO₂ showed an efficient delivery of Q molecules into mouse fibroblast cells and improved the cellular antioxidant defense system against oxidative toxicity. Wang *et al.* synthesized TiO₂NPs and studied their surface modification with single and mixed stabilizers, such as PEG, cetyltrimethylammonium bromide (CTAB) and carboxamide.¹⁰⁷ Their results showed that the surfactants strongly affect the morphology of these TiO₂NPs and, for the PEG, they obtained ellipse modified TiO₂NPs having an improved photocatalytic activity for the degradation of methyl orange under UV irradiation. Wang *et al.* synthesized ultrafine titanium monoxide nanorods (TiO_{1+x} NRs) and then modified them with PEG.⁸⁴ TiO_{1+x} NRs-PEG was used as a new sonodynamic agent and showed much more efficiency for the ultrasound-induced generation of reactive oxygen species (ROS), compared to the conventional sonosensitizer. Interestingly, TiO_{1+x} NRs-PEG could also generate hydroxyl radicals (OH[•]) from endogenous H₂O₂ in the tumor to enable chemodynamic therapy (CDT). For the treated mice, TiO_{1+x} NRs-PEG showed efficient passive retention in tumors post-intravenous injection, with no significant long-term toxicity, indicating the potential ability of this modified TiO₂ nanostructure to be used as a sonosensitizer and a CDT agent.

Chitosan (CS) is another frequent hydrophilic polymer for the surface modification of TiO₂NPs, which has low toxicity, good biocompatibility and biodegradability.⁹⁴ Chitosan-modified TiO₂NPs (TiO₂NPs-CS) have potential applications in various technologies such as photocatalytic nanostructures,¹⁰⁸ antibacterial package materials,¹⁰⁹ wound healing materials,^{110,111} wastewater treatment,¹¹² and sensors.¹¹³



From a structural point of view, CS has reactive amino and hydroxyl side groups which can interact with TiO₂NPs by hydrogen bonding and form stable nanoco-hybrids.⁹⁴ For instance, in 2022, Moulahou *et al.* synthesized TiO₂-chitosan nanocomposites (TiO₂NPs-CS) and combined them with various metal ions (silver, zinc, copper, and iron) to prepare novel chitosan-based films and improve the physicochemical and biological activities of the individual components.⁹⁵ The antimicrobial properties of these films were studied against three pathogens (*P. aeruginosa*, *C. albicans*, and *S. aureus*) under different light conditions. Among these nanocomposites, TiO₂NPs-CS-Ag showed the best antibacterial activity. Besides, the nanocomposites were tested for the photocatalytic degradation of methylene blue (MB), and the best data were observed for TiO₂NPs-CS-Cu with a higher specificity towards MB than the other two tested dyes (methyl orange and bromophenol blue). These results showed that the TiO₂NPs-CS-metal ions have great potential as ambient light packaging materials, coating materials, and photocatalysts. In another recent study, Castillo *et al.* used TiO₂NPs-chitosan for the electroanalytical detection of imidacloprid, (a neonicotinoid) that is a systemic insecticide and can accumulate in agricultural products and negatively affect human health.⁹⁴ The authors showed that the TiO₂NPs-chitosan, with a high surface area, served as molecular recognition sites for the imidacloprid detection, with an optimum concentration of 40 wt% of TiO₂NPs. Elmehbad *et al.* prepared two new chitosan derivatives by incorporating salicylhydrazide into a chitosan Schiff base (SCsSB) and chitosan (SCs) to make two nanohybrids, SCs/TiO₂-1% and SCs/TiO₂-3%.⁹⁶ The anti-biofilm and antimicrobial activities of these nanostructures were ranked as SCs/TiO₂-3% > SCs/TiO₂-1% > SCs > SCsSB. Their results showed that the modification of TiO₂NPs with the chitosan polymer enhanced the antibacterial performance of the components. SCs/TiO₂-3% was biocompatible with normal human cells, indicating the potential ability of this nanocomposite for antimicrobial agents. Majnis *et al.* integrated chitosan (CS) and ZrO₂ into TiO₂NPs through a sol-gel fabrication method, to make the CS-ZrO₂/TiO₂ photocatalyst having a smaller band gap, compared to the unmodified TiO₂NPs.⁹⁷ The photodegradation efficiency of CS-ZrO₂/TiO₂ was tested (under solar irradiation) using a model dye, malachite green (MG), and the results showed an increase efficiency from 11.87% (for TiO₂NPs alone) to 30.72% (with ZrO₂ and CS addition in TiO₂NPs).

Also, other polymers such as polydopamine (PDA), polysaccharide, polylactic acid (PLA), polyacrylic acid (PAA), alginate (Al), polyvinylidene fluoride, PEI, PVP, and PAMAM (polyamidoamine)^{114,115} are used for the surface modification of TiO₂NPs (see Table 4). For example, Dong *et al.* used polydopamine (PDA) with excellent hydrophilicity for the surface modification of TiO₂NPs.⁹² Then, these PDA-modified TiO₂NPs were combined with hydrophobic graphene (Gr) *via* the π - π non-covalent interaction to enhance the water dispersion stability of Gr. Regarding practical applications, this nanocomposite was tested for its fire resistance ability inside the intumescent waterborne epoxy coating. The results showed

that introducing PDA and TiO₂NPs effectively improved the oxidation resistance and stability of the composite. Zhang *et al.* prepared TiO₂NP-PDA hybrid nanoparticles to reduce oxygen vacancies in TiO₂NPs and provide better stability for the polymer matrix. TiO₂NP-PDA was composited with polylactic acid (PLA) to prepare PLA/TiO₂NP-PDA nanocomposites.¹¹⁶ These nanocomposite films showed excellent UV-shielding performance without sacrificing transparency (the transmittance at 550 nm was 85.7%). PLA is very sensitive to UV light; however, due to the shielding effect of TiO₂NPs-PDA in this film, the performance of PLA was improved, and it can broaden its application for environmental materials having a wider range of uses with a longer service life.

TiO₂NPs are also used as food whitening in candies, chocolates, and cakes with a high carbohydrate content. However, there is a little knowledge about the potential interaction between the food carbohydrate and food grade-TiO₂NPs. In the case of polysaccharides, Qiaorun *et al.* studied the interaction between TiO₂NPs and seven common carbohydrates (including monosaccharides, disaccharides, and polysaccharides).¹¹⁷ Their results showed that the TiO₂NPs can interact with all these tested carbohydrates and enter the body as a food additive, and interact with the food matrix for a series of reactions. The food polysaccharides showed stronger adsorption onto the TiO₂NPs than monosaccharides and disaccharides.

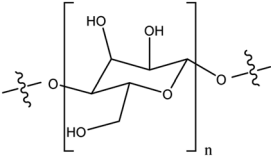
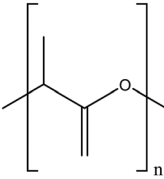
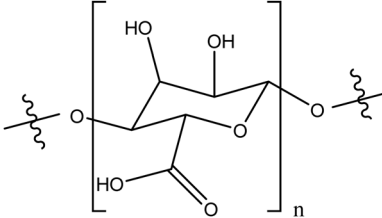
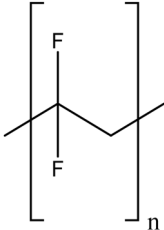
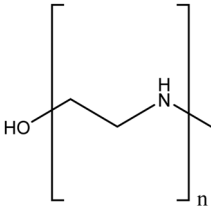
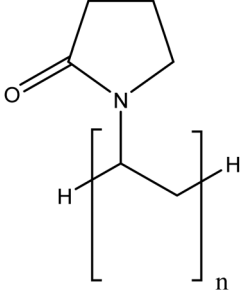
Regarding polylactic acid (PLA), Tajdari *et al.* prepared ZnO-PLA, TiO₂-PLA and ZnO/TiO₂-PLA nanocomposites with different percentages of nanoparticles and two different types of ZnO morphologies.¹¹⁸ Their results showed the enhanced mechanical and optical properties of PLA when it was combined with the nanoparticles. Also, the antibacterial activity of PLA was improved against Gram-positive *L. monocytogenes* and Gram-negative bacteria *E. coli* by incorporating nanoparticles.

Dextran (Dex) is a polysaccharide with excellent biocompatibility and good water solubility, so the modification of TiO₂NPs with dextran can improve the physicochemical properties of the nanoparticle. Naghibi *et al.* compared the *in situ* and *ex situ* surface modifications of TiO₂NPs using dextran (Dex) and Dex/poly ethylene glycol (PEG), by comparing the colloidal stability of the modified TiO₂NPs prepared by these two methods.⁹³ Their results showed that the *in situ* additions of Dex and PEG, during the synthesis of TiO₂NPs (hydrothermal synthesis), resulted in a highly stable colloidal solution (more than 60 days), whereas the *ex situ* additions of Dex and PEG (after the TiO₂NP synthesis) did not significantly impact on the colloidal stability of TiO₂NPs.

3.2.2. Small molecules. Another important group of stabilizing agents are small molecules which can provide either lipophilic or hydrophilic character for the modified TiO₂NPs. The lipophilic small molecules such as oleic acid are considered as “fat-loving” or “fat-liking” and have great importance for the preparation of lipophilic TiO₂NPs with good dispersity in non-aqueous solutions. More importantly, oleic acid can form a dense protective monolayer around the TiO₂NPs and can strongly attach to the nanosurface.²⁵ The chemical structure of oleic acid and two recent publications on oleic acid-modified TiO₂NPs are summarized in Table 5.



Table 4 Some characteristics of polymers used for the surface modification of TiO₂NPs

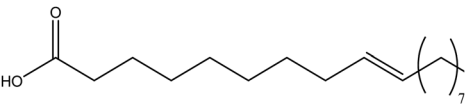
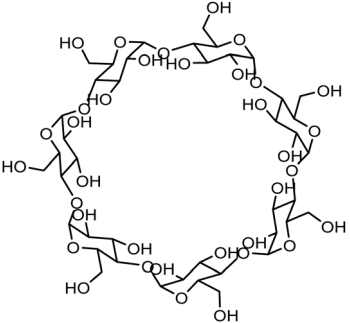
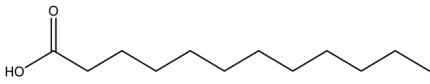
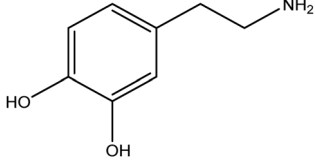
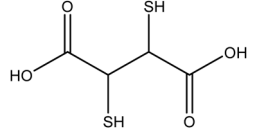
| Name | Structure | Stability | Modification mechanism (covalent/non-covalent) | Applications | Ref. |
|----------------------------|---|--|--|---|-----------------|
| Polysaccharide |  | Strongly modified the TiO ₂ NP stability by inducing their partial and rapid disagglomeration, by steric effects and electrostatic interactions | N/A | Water purification | 119 |
| Poly lactic acid |  | Highly dispersible by steric effects | Hydrogen bonding interactions | Antimicrobial against <i>S. aureus</i> , <i>Salmonella</i> and <i>E. coli</i> | 90, 120 and 121 |
| Alginate acid |  | Enhanced stability in water, compared to unmodified TiO ₂ NPs through combination of electrostatic repulsion and steric effects | Electrostatic interactions | Environmental applications | 100 and 101 |
| Polyvinylidene fluoride |  | Excellent stability of modified TiO ₂ NPs by steric effects | N/A | Increased anti-fouling properties | 122 and 123 |
| Polyethyleneimine (PEI) |  | Enhanced stability in water, compared to unmodified TiO ₂ NPs by electrostatic repulsion | Physical interactions | Photodegradation of methylene blue | 87 |
| Polyvinylpyrrolidone (PVP) |  | Considerable improvement in terms of stability compared to unmodified TiO ₂ NPs via steric hindrance | N/A | Enhancing the TiO ₂ NP dispersion in blood and urine | 86 |

However, in some biomedical applications, the use of lipophilic-modified TiO₂NPs is greatly limited due to low dispersity of the nanoparticles in biological aqueous solutions. For the medical applications of modified TiO₂NPs, the research is more focused on the synthesis of hydrophilic or water dispersible TiO₂NPs. In this regard, several small organic molecules such as amino acids, citric acid, cyclodextrin, dopamine, lauric

acid, and dimercaptosuccinic acid (DMSA) are often used for the surface modification of TiO₂NPs to enhance the hydrophilicity of modified nanoparticles for the biological applications. In the case of citrate (or citric acid), Connolly *et al.* studied the bioaccumulation of uncoated TiO₂NPs and TiO₂NPs–citrate in fish to investigate the relationship between surface coating and uptake (biokinetics) *in vivo*.⁷⁶ Rainbow trout (*Oncorhynchus mykiss*) were



Table 5 Small organic molecules for the surface modification of TiO₂NPs

| Name | Structure | Stability | Modification mechanism | Applications | Ref. |
|--------------------------------|--|--|---|--|------------|
| Oleic acid |  | Steric hindrance | Chemically bonded with the surface titanium ion (by bidentate linkages) | Healing excision wounds were studied in the rat animal model | 126–128 |
| Cyclodextrin |  | Improvement of the colloidal stability, steric hindrance | N/A | Degradation of wastewater pollutants, antibacterials | 129 |
| Lauric acid |  | The functionalized TiO ₂ NPs exhibited significantly reduced agglomeration, both in dry and in dispersed states (in oily media) | Chemical interaction with silane-functionalized TiO ₂ NPs | UV filtering ability | 130 |
| Dopamine |  | Steric stabilization of TiO ₂ NPs when it is polymerized to polydopamine | Probable chemical interactions (determined by computational chemistry) | Drug discovery, diagnostics, environmental applications, and food safety | 131 |
| Dimercaptosuccinic acid (DMSA) |  | Improve dispersivity of TiO ₂ NPs in solutions and increase electrostatic repulsion between nanoparticles | N/A | Cytotoxicity on human aortic and endothelial cells | 132 133 |

fed diets spiked with the uncoated and citrate-coated TiO₂NPs (100 mg NPs per kg feed) for 10 days and thereafter, fish were allowed to depurate for 42 days. Their results showed that the surface modification affected the uptake and, in some cases, caused slower depuration and distinct distributions. In another research, Liu *et al.* synthesized citrate-coated Gd-doped TiO₂ ellipsoidal nanoparticles (GdTi-SC NPs) to improve the efficiency of gadolinium-based T1 contrast agents (CAs) for magnetic resonance imaging (MRI).¹²⁴ The *in vivo* MRI tests on rats demonstrated that the modified TiO₂NPs have a high potential ability as high-performance T1 contrast agents for the sensitive imaging of blood vessels and the accurate diagnosis of vascular lesions. Peper *et al.* studied redox reactions of aqueous colloidal solutions of both citrate capped- and uncapped-TiO₂NPs (c-TiO₂ and uc-TiO₂) and reported the different redox behaviors of these two systems.¹²⁵

3.2.3. Hydrogels. Hydrogels are gel-like materials processing a three-dimensional (3D) network composed of hydrophilic polymer building blocks. The polymers link to each other forming an insoluble 3D matrix which can absorb and trap a significant amount of water. The hydrogels have excellent biocompatibility, reversible swelling/deswelling behavior, and high potential adsorption capacity, making them suitable

materials for biological applications including tissue engineering, drug delivery, cosmetics, personal hygiene, and diapers.^{134,135} They have suitable functional groups for the interaction with the TiO₂NPs and therefore, they can be used for the modification/stabilization and encapsulation of TiO₂NPs.¹³⁶ In all these systems, the hydrogel matrix provides a protecting and modifying environment for the TiO₂NPs and stabilize them in a gel-like media, preventing their agglomeration. Moreover, hybridization of TiO₂NPs with hydrogels can have a significant positive effect on the structural, mechanical, and thermal properties of the hydrogels. For example, in 2022, Makhado *et al.* prepared a ghatti gum/poly(acrylic acid)/TiO₂NPs (GG/poly(AA)/TiO₂) hydrogel nanocomposite and studied the structure, morphology, and thermomechanical characteristics of the synthesized hydrogel nanocomposite.¹³⁷ The incorporation of TiO₂NPs with the hydrogel matrix improved the hydrogel thermal stability and mechanical strength. Zhao *et al.* synthesized highly dispersible TiO₂NPs modified with a 3D graphene hydrogel composite (TiO₂NPs-rGH) using a hydrothermal method.¹³⁸ The combination of 2D graphene sheets with hybrid TiO₂NPs enhanced the specific surface area of the TiO₂NPs-rGH 3D composite. Their results showed that the TiO₂NPs-rGH composites have higher photocatalytic



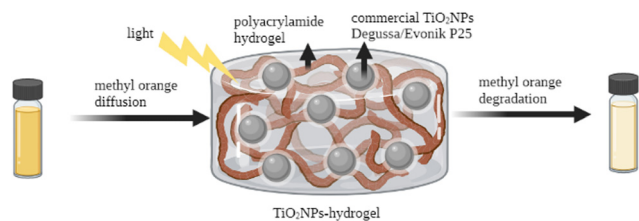


Fig. 4 TiO₂NP–hydrogel system for the photocatalytic degradation of methyl orange. Adapted with permission from ref. 139. Copyright 2022 Elsevier.

performance than unmodified TiO₂NPs. Mansurov *et al.* reported the improvement of the photocatalytic degradation of commercial TiO₂NPs Degussa/Evonik P25 after modification with the composite polyacrylamide hydrogel (Fig. 4).¹³⁹

Ulu *et al.* reported the preparation and characterization of chitosan/PEG/TiO₂NP (CH/PEG/TiO₂NP) composite hydrogels for antibacterial applications.¹⁴⁰ Their results showed that the CH/PEG/TiO₂NPs improved the mechanical and thermal properties of the hydrogel due to the presence of TiO₂NPs. More importantly, this TiO₂ nanocomposite showed potential antimicrobial activity. Yue *et al.* prepared a novel photocatalytic hydrogel by loading TiO₂NPs onto the surface of 2,2,6,6-tetramethylpiperidine-1-oxyl (TEMPO)-oxidized chitin nanofibers (TOCNs), which were further incorporated into the polyacrylamide (PAM) matrix.¹⁴¹ The presence of TiO₂NPs enhanced the compressive strength of this hydrogel with excellent stretchability and photocatalytic activity.

In the first part of this review, it can be concluded that the organic-based surface modification of TiO₂NPs can result in a significant improvement of their physicochemical properties for preparing much more effective TiO₂NP-based nanostructures and decreasing the potential toxicity of TiO₂NPs as well. For the following part of this review, the recent biological applications of TiO₂-based nanomaterials will be presented. The main aim of this section is to present some specific organic modifiers and polymers for TiO₂NPs which have been recently used (2020–2022) in eight main fields of drug delivery, photodynamic therapy, antibacterial, biosensors, antiviral, antifungal, cancer therapy, and tissue engineering. Also, the advantages of these organic modifiers will be discussed.

4. Biological applications of modified TiO₂NPs

4.1. Photo-, thermo-, and sonodynamic effects of modified TiO₂NPs on cancer cells

According to the World Health Organization, there is an increase in annual cancer cases from 14 to 22 million in 2012–2030 period.¹⁴² During recent years, modified TiO₂NPs have been studied as promising alternatives for the cancer therapy *via* different methodologies including photodynamic, photothermal, and sonodynamic therapies.^{1,7,143–145}

In the photodynamic therapy, they are regarded as inorganic photosensitizers for anti-cancerous photodynamic therapy

(PDT) owing to their unique phototoxic effect upon UV light irradiation. The UV light absorption can excite valence electrons of TiO₂NPs to generate electrons and holes on the nanosurface, and consequently, a series of redox reactions are initiated which produce anti-cancerous reactive oxygen species (ROS) such as hydroxyl radicals (HO•⁻), superoxide anions (O₂•⁻), hydrogen peroxide (H₂O₂), *etc.*^{1,7} In spite of their advantages, UV irradiation is not always suitable for PDT due to its limited penetration depth, a lower light content and, more importantly, its harmful side effects for the patients exposed to UV light.¹⁴⁶ On this basis, much research has been dedicated to extending the photoresponse of TiO₂NPs to the visible light region. In this field, the surface modification of TiO₂NPs is focused on the use of biologically active species on the surface of TiO₂NPs to enhance the selectivity and therapeutic efficiency of TiO₂NPs. Besides, this type of surface modification can reduce the potential toxicity of unmodified TiO₂NPs, which is reported in recent publications.⁵ In the following, some recent publications will be presented in which the TiO₂NPs have been modified with such types of organic modifiers to prepare the efficient therapeutic TiO₂NPs.

The surface modification of TiO₂NPs with organic dyes, especially porphyrins, has attracted growing interest as it can broaden the absorption range of TiO₂NPs from the UV region to the visible region.¹⁴⁷ Chlorin e6 (Ce6) is a porphyrin-based photosensitizer (PS) with a high sensitizing efficiency,¹⁴⁸ which can be conjugated with the TiO₂NP surface either by non-covalent or covalent modalities.¹⁴⁶ In general, the physical conjugation of PS might suffer from desorption which limit the nanosystem efficiency. Conversely, the covalent attachment of the PS to TiO₂NPs can guarantee the stability of the nanosystem, especially when the silane linkers are used which hold a high affinity towards the hydroxyl groups of TiO₂NPs. For example, Youssef *et al.* studied the attachment of Ce6 to TiO₂NPs by two approaches: (1) TiO₂NPs were encapsulated with silanes (APTES and TEOS) and Ce6 (as PS), followed by polyethylene glycol (PEG) grafting on this shell to obtain TiO₂NPs@4Si-Ce6-PEG and (2) for the second approach, the TiO₂NPs were first modified only by APTES (as the silane linker) and then Ce6 was covalently attached onto the modified TiO₂NPs@APTES *via* an amide bond to construct TiO₂NPs@APTES-Ce6.¹⁴⁶ *In vitro* tests on glioblastoma U87 cells were performed to study the cellular uptake, phototoxicity, and dark cytotoxicity of the modified and unmodified TiO₂NPs. In contrast to the PEGylated TiO₂NPs, the APTES-modified ones showed more PDT efficiency, in which a %89 decrease of U87 viability was observed for 200 μg mL⁻¹ of TiO₂NPs@APTES-Ce6, which corresponds to 0.22 μM of Ce6. This surface modification resulted in a change of the absorption profile of the hybridized TiO₂NPs from the UV region (for unmodified TiO₂NPs) to the visible region. Also, it can enhance the biocompatibility of TiO₂NPs and their stability, due to the presence of silane coupling agents on the surface of TiO₂NPs.

The modification of TiO₂NPs with targeting molecules (such as folic acid (FA)) significantly enhances the selectivity of TiO₂NPs to some types of cancer, which is an alternative way



to improve the therapeutic efficiency of PDT. The folic acid-modified TiO₂NPs can be accumulated in the target sites by increasing the affinity of folic acid-modified NPs to the pathological tissue. Also, this modification can improve cell membrane penetration through folate receptors, which are overexpressed on the surface of some types of cancer cells. For example, Liang *et al.* synthesized a novel TiO₂NPs–folic acid–Al(III) phthalocyanine chloride tetrasulfonic acid (TiO₂NPs–FA–Pc) targeting nanosystem for therapy of the folate receptor-positive cancer cells.¹⁴⁹ In this system, folic acid (FA) was conjugated with the TiO₂NPs as a tumor-targeting agent which enhanced the selectivity of TiO₂NPs toward the cancer cells. It should be mentioned that the conventional photosensitizer (Pc) exhibited low selectivity for tumor targeting and low two-photon absorption. The modification of TiO₂NPs using this photosensitizer enhanced its two-photon absorption of TiO₂NPs–FA–Pc. The *in vitro* studies of these modified TiO₂NPs showed a high PDT efficiency and biocompatibility. Also, it exhibited tumor growth suppression in mice bearing HeLa xenograft tumors with minimal side effects, using low dose of this nanocomposite under low light irradiation.

In another study, Salama *et al.* investigated the attachment of the epidermal growth factor receptor (EGFR) on PEG-modified TiO₂NPs for increasing the PDT effect for epithelial cell carcinoma (A431 cell line).¹⁵⁰ The EGFR is vital for cell proliferation and it is highly expressed on many cancer cells, so for this reason, the modification of TiO₂NPs with EGF could increase the efficiency and selectivity of TiO₂-based PDT. Their results showed that the EGF modification of TiO₂NPs–PEG diminished the cell viability of the cancer cells *via* interrupting DNA synthesis. Also, the PEG modification of the TiO₂ core could enhance the stability and bioavailability of this nanosystem.

In spite of the advantage of PDT, the *in vivo* production of toxic ROS and high photosensitivity of treated patients could limit the PDT technique.⁷ The development of modified TiO₂NPs with a high photothermal conversion efficiency has recently gained much attention, as an efficient and non-invasive method, to destroy target tumor tissues.¹⁵¹ The heat generated by the vibrational relaxation of stimulated TiO₂NPs (> 42 °C) could trigger several photothermal effects in tumors such as causes necrosis, apoptosis, and necroptosis. The limitation of PTT (photothermal therapy) is the low NIR absorption of cancer cells located far from the tissue surface which can be overcome by modifying TiO₂NPs with the organic molecules absorbing long-wavelength visible light or NIR. Behnam *et al.* used PEG-modified TiO₂NPs as PTT agents to increase the water dispersibility and biocompatibility of TiO₂NPs.¹⁵² Besides, these PEGylated TiO₂NPs could escape the reticuloendothelial system (RES) and reach to their target tumors. The *in vivo* results showed a relatively high PTT efficacy of these TiO₂NP–PEG nanosystems on reducing the melanoma tumor size without any symptom of cancer cells in treated cases. Therefore, TiO₂NPs–PEG can be utilized as a potent agent with low toxicity in the hyperthermia cancer therapy.

As another advanced technique, combined PDT/PTT approaches have much stronger effects than expected; for

instance, Gao *et al.* synthesized polydopamine-modified TiO₂NPs (TiO₂-b-P25@PDA NPs) forming a high core-shell structure, as an improved PTT nanosystem.¹⁵³ They then prepared synergistic nanoprobes (TiO₂-b-P25@PDA-Ce6 (Mn)) by combining chlorine e6 (Ce6) and chelating Mn²⁺ for use in combined PDT/PTT. These modified-TiO₂NPs showed high ROS generation and high photothermal conversion efficiency (32.12%). Their *in vivo* tests on a 4T1 tumor-bearing nude mouse model illustrated a synergistic significant antitumor effect of the nanosystem (under the combination of PDT/PTT with a low-dose laser), compared to the partial tumor inhibition by single PDT and single PTT. So, the co-modification of TiO₂NPs with the PTT and PDT agents can dramatically enhance the therapeutic efficacy of modified TiO₂NPs, compared to the unmodified structures.

In 2022, Dai *et al.* modified TiO₂NPs with hyaluronan and porphine for the simultaneous PTT/PDT therapies.¹⁵⁴ They used these two surface modifiers (hyaluronan and porphine) to mildly reduce the lipid level of RAW 264.7 cells without triggering the harsh cell apoptosis, which is an important strategy for the treatment of chronic cardiovascular diseases. For both PTT alone and PTT + PDT therapies, their result demonstrated a considerable decrease of intracellular lipid load without triggering apoptotic cell death or necrosis, below the 45 °C. Conversely, the PDT modality showed a small decrease in lipid levels and a significant apoptosis or necrosis. These results indicated that the surface modification of TiO₂NPs could increase the PTT efficiency and enhance the local temperature to relatively moderate levels (44 °C) after NIR irradiation, which prevented excessive cell apoptosis or necrosis, while PDT resulted in harsh cell death.

Regarding the sonodynamic effect of TiO₂NPs, there have been an admirable effort for developing sonodynamic TiO₂NPs, as a non-invasive method, having high tissue penetration and spatiotemporal selectivity. In SDT (sonodynamic therapy), the ROS generation is triggered under ultrasound (US) stimulation, resulting in selective tumor targeting with minimal damage to nearby healthy cells.¹⁵⁵

Pancreatic cancer is considered as the third-leading cause of death in 2022 because of its increasing cases and mortality rates.¹⁵⁶ In the advanced-stage of this cancer, surgical resection is the primary method but only 20–15% of patients can survive and the other types of therapeutic modalities, such as chemotherapy and immunotherapy, show poor response to the majority of clinical treatments.¹⁵⁷ Sonodynamic therapy (SDT) has shown to be a promising alternative in this case. However, pancreatic tumors are surrounded by the interstitial fluid pressure (IFP) and hypoxia tumor microenvironment (TME) which decreases the sonosensitizer penetration into the tumor, resulting in low SDT efficiency.^{158,159} Collagen is the most abundant protein in the ECM (extracellular matrix) of pancreatic cancer¹⁶⁰ and so, the modification of the TiO₂NP sonosensitizer with collagenase is a promising strategy to improve the SDT efficiency in pancreatic cancer. Recently, Luo *et al.* synthesized collagenase-modified hollow TiO₂NPs (H-TiO₂NPs-Co) capable of degrading stromal barriers and producing sufficient



ROS.¹⁶¹ The *in vivo* tests in a patient-derived xenograft (PDX) model showed an enhanced penetration and retention of the TiO₂NPs within tumor tissues, due to the presence of Co on the TiO₂NP surface. The ultrasonic irradiation caused the controlled release of collagenase which degraded tumor matrix fibers. The attached collagenase (Co) resulted in accumulation of modified TiO₂NPs within the tumor which generate abundant ROS under the ultrasound (US) irradiation and dramatically increase the selectivity and therapeutic efficiency of SDT.

In 2021, Wei *et al.* synthesized newly modified TiO₂NPs, functionalized with a malignant melanoma cell membrane (B16F10M) and a targeting aPD-L1 antibody for enhanced sonodynamic tumor therapy.¹⁶² Under ultrasound irradiation, these modified TiO₂NPs showed a high efficiency to generate ROS (¹O₂) along with precise targeting effects, high tumor uptake, and intracellular sonocatalytic killing of the B16F10 cells. In this study, the modification of TiO₂NPs with the mentioned biomolecules resulted in a dramatic enhancement of biocompatibility, selectivity and therapeutic yield of the modified TiO₂NPs.

Lin *et al.* reported the synthesis of a multifunctional modified TiO₂NP sonosensitizer (TiO₂NPs-Ce6-CpG, CpG: a targeting oligonucleotide) for highly efficient cancer immunotherapy.¹⁶³ To improve the biocompatibility and sonotherapeutic ability of these TiO₂NPs, they were modified with chlorin e6 (Ce6) and a CpG oligonucleotide (CpG ODN) to enhance the immune response. Ce6 is a hydrophilic porphyrin-type sonosensitizer, which accumulates effectively in tumors, and can generate ROS under the ultrasound activation to induce apoptosis and necrosis of the tumor cells. The CpG ODN oligonucleotide is an immunological adjuvant that can trigger cellular immune responses to enhance the anticancer properties of a variety of cancer treatments. The injected TiO₂NPs-Ce6-CpG could induce the release of tumor-associated antigens and demonstrated vaccine-like functions together with the CpG adjuvant, which activated dendritic cells (DCs) and enhanced tumor-infiltrating CD8⁺ T cells to the tumor tissues, inducing a robust antitumor immunological response.

Lee *et al.* studied the potential application of SDT against glioblastoma cells using TiO₂NPs modified with a targeting molecule, anti-EGFR antibody.¹⁶⁴ Their results showed a dramatic enhancement of the selectivity and internalization of modified TiO₂NPs toward the target cells, due to the presence of the anti-EGFR antibody on the TiO₂NP surface. Under the ultrasound irradiation of modified TiO₂NPs, cell viabilities were reduced because of the ROS generation with minimal effects on apoptosis.

In 2021, Yousefi, *et al.* used porphyrin-loaded TiO₂NPs and studied their sonotoxicity on MDA-MB-231 cells.¹⁶⁵ To increase the biocompatibility and ultrasound absorption efficiency, the surface of TiO₂NPs was modified first with the polyvinyl alcohol (PVA) polymer and then with porphyrin. The *in vitro* results indicated that these modified TiO₂NPs are non-toxic and under the ultrasound radiation they could damage the breast cancer cells.

Pariante *et al.* synthesized sono-responsive TiO₂NPs modified with poly(ethylene oxide)-poly(propylene oxide) (PEO-PPO) copolymers for their potential in sonodynamic applications.¹⁶⁶ Their results showed an enhanced biocompatibility of these modified TiO₂NPs, due to the modifying copolymer. Upon irradiation with the therapeutic ultrasound, the nanoparticles generated ROS and induced the apoptosis of Rh30 cells. The compatibility and cellular uptake of these modified TiO₂NPs were confirmed on the Rh30 cell line, as a model of rhabdomyosarcoma without any significant hemolysis over 24 h treatment.

The other recent publications on the therapeutic effect of TiO₂NP-based nanostructures are summarized in Table 6.

4.2. Drug delivery

Chemotherapy is limited by the uncontrolled distribution of chemodrugs towards both cancerous and healthy cells which results in adverse side effects for the treated patients. TiO₂NP-based drug delivery systems have attracted much more interest in recent years to enhance the target specificity of chemotherapy and reduce the systemic side effects. These advanced drug delivery systems benefit various controlled-release mechanisms including pH- and thermo-sensitive, photo-induced, and enzyme-responsive techniques which result in an enhanced specificity of the drugs toward the cancer cells and subsequently, the drug dosage can be significantly minimized while still maintaining the pharmacological effects. Recently, the modified TiO₂NPs have been used for the delivery of various anticancer drugs, such as temozolomide, cisplatin, doxorubicin, and daunorubicin.^{8,182} For instance, Han *et al.* reported the synthesis of poly(acrylic acid)-calcium phosphate modified TiO₂NPs (TiO₂NPs@PAA-CaP) for the efficient drug delivery of doxorubicin (DOX).¹⁸³ This surface modification of TiO₂NPs resulted in a significant enhancement of DOX loading and encapsulation up to eight times, compared to that of unmodified TiO₂NPs. Due to the pH-responsive surface properties of the PAA-CaP modifying layer, DOX-loaded TiO₂NPs@PAA-CaP exhibited much faster cumulative DOX release at acidic pH = 5.2 than at neutral pH = 7.4. TiO₂NPs@PAA-CaP(DOX) illustrated an enhanced cellular uptake and a higher cytotoxicity towards MCF-7 tumor cells, compared to that of free DOX. More importantly, this modified nanosystem demonstrated synergistic chemo- and photodynamic therapeutic effects on the target cells.

Neuroblastoma is considered as one of the leading causes of cancer-related deaths in children worldwide¹⁸⁴ and temozolomide (TMZ) has been widely used to treat neuroblastoma.^{185,186} The synthesis of TiO₂NP-TMZ was studied previously for its potential to treat neuroblastoma;¹⁸⁷ however, the clinical application of TiO₂NPs is strictly limited by its serious cytotoxicity, inflammation, and brain damage.¹⁸⁸ For this reason, alginate was used as a TiO₂NP modifier due to the biological advantages of alginate such as biocompatibility, anti-inflammatory effects, antioxidant properties, and easy degradation with little toxicity.¹⁸⁹ Zhao *et al.* reported the modification of TiO₂NPs-temozolomide (TiO₂NPs-TMZ) with alginate and studied their anti-oxidant, anti-inflammatory, and anti-tumor effects on



Table 6 Other recent publications (2021–2022) on the therapeutic effect of TiO₂NP-based nanostructures

| Nanosystem | Applications | Ref. |
|---|--|------|
| Fe ₂ O ₃ -TiO ₂ nanocomposites, using polyvinylpyrrolidone-polyethylene glycol (PVP-PEG) | Showed remarkable PDT activity in HeLa cell lines <i>via</i> the generation of intracellular ROS | 167 |
| New nanocomposite, TiO ₂ NPs@Ru@siRNA | Remarkable PDT activity on patient-derived xenograft (PDX) and rat oral experimental carcinogenesis models | 168 |
| Folic acid-functionalized TiO ₂ NPs | Modeling of active targeting of tumor cells | 169 |
| Chelate-free gadolinium loaded TiO ₂ NPs coated with transferrin (Tf) | Coating of this TiO ₂ -Gd NPs with Tf stabilized the nanoconstruct and minimized aggregation, showing a dramatic selectivity for the photodynamic targeting of studied cancer cells | 170 |
| TiO ₂ NP-Ag nanohybrid modified with the Pluronic [®] F-127 polymer, which is permitted by the Food and Drug Administration (FDA) | This polymer improved the biocompatibility of TiO ₂ NP-Ag, tested in 4T1 breast cancer cells and the nanohybrid showed endocytosed by cancer cells produced high intracellular ROS under UV conditions (5.6 mW cm ⁻²), resulting in cancer cell apoptosis | 171 |
| TiO ₂ /Cur@ZIF-8 nano-composite (Cur: chemotherapeutic agent curcumin) | Synergistic photodynamic-chemotherapy and pH-/and NIR-stimulated drug release | 172 |
| N-doped graphene quantum dots (QDs)/titanium dioxide nanocomposites (N-GQDs/TiO ₂ NPs) modified with citric acid | Upon the photo-activation of N-GQDs/TiO ₂ NPs with near-infrared (NIR) light, the nanocomposites generated reactive oxygen species (ROS), mainly singlet oxygen (¹ O ₂), which caused more significant cell death in MDA-MB-231 (an epithelial, human breast cancer cells) than in HS27 (human foreskin fibroblast) | 173 |
| Tc-99m-labeled lupulone-conjugated Fe ₃ O ₄ @TiO ₂ nanocomposite | Lupulone-conjugated Fe ₃ O ₄ @TiO ₂ nanocomposites showed suitable dispersion and the photodynamic effect on prostate cancer without visible aggregation | 174 |
| Doped TiO ₂ rhombic nanocomposites modified with Pluronic [®] F-68 | Mn-TiO ₂ -PF-68 RNCs demonstrated negligible toxicity with physiological stability. Mn ³⁺ doped with photosensitizers (TiO ₂) also exhibited a great synergistic effect of photo-killing <i>in vitro</i> by developing hydroxyl radicals | 175 |
| Titanium-oxo nanoclusters modified with dopamine and PEG | The introduced dopamine (DA) ligands not only facilitated the water solubility and the photocatalytic properties of the NPs but also involved the tumor-targeting behavior through the binding affinity with DA receptors on cancer cells. Under Cerenkov irradiation, these nanocomposites enable efficient hydroxyl radical generation | 176 |
| C-doped TiO ₂ NPs | They were prepared and tested as a photosensitizer for visible-light-driven photodynamic therapy against cervical cancer cells (HeLa) | 177 |
| Tablet-like TiO ₂ /c nanocomposite with a metal-organic-framework (MOF)-derived carbon structure | This nanocomposite continued to generate ROS in response to repeated ultrasound irradiation and was able to induce tumor cell apoptosis <i>via</i> SDT-induced DNA damage <i>in vitro</i> and <i>in vivo</i> . This TiO ₂ /C nanocomposite also exhibited good biocompatibility and did not induce any apparent toxicity <i>in vitro</i> and <i>in vivo</i> . | 178 |
| Hypoxia-tolerant MOF@TiO ₂ (MOF, metal-organic framework) | Hypoxia-tolerant type I photodynamic therapy against hypoxic cancer | 179 |
| MnCO@TPP@C-TiO ₂ NPs | MnCO@TPP@C-TiO ₂ NPs selectively localized in the mitochondria of HeLa cells where the overexpressed-H ₂ O ₂ triggered CO released, resulting in mitochondrial damage | 180 |
| Semiconductor quantum dots (CdX, X = S, Te, Se)-TiO ₂ NPs modified with folic acid | Prepared FA-CdX-TiO ₂ NPs (X = S, Se) exhibited excellent cancer-targeting ability during PDT treatment. The optimum PDT efficiency of FA-CdSe-TiO ₂ NPs indicated that the photocatalytic and targeting abilities were much higher than those of the pure TiO ₂ NPs and CdSe-TiO ₂ NPs | 181 |

neuroblastoma.¹⁹⁰ Their *in vivo* results showed that the alginate modification enhanced the cytotoxicity toward neuroblastoma cells and decreased inhibitory activity toward normal neuronal cells. This modification increased the antioxidant, anti-inflammatory, and antitumor activities of TiO₂NPs-TMZ and prolonged the survival time of the neuroblastoma model ($P < 0.05$). The results showed that the alginate modification controlled the TMZ release from the TiO₂NPs-TMZ-alginate nanoparticles.

In 2020, Kelin *et al.* reported a novel drug delivery vehicle of TiO₂NPs, encapsulated by bilayer shells that allow the reversible incorporation of hydrophobic drugs.¹⁹¹ In these systems, TiO₂NPs were chemically encapsulated by the covalent binding of hydrophobic phosphonic acid, followed by the second surface modification by amphiphilic sodium dodecylbenzenesulfonate *via* hydrophobic interactions between the

dodecylbenzene moiety and the hydrophobic first shell. This two-layer modification makes the hydrophobic surface suitable for the loading of hydrophobic drugs. These modified TiO₂NPs were loaded with hydrophobic anticancer drugs 7-amino-4-methylcoumarin and quercetin. The results showed a sustained release of these anticancer drugs into the cytoplasm due to the presence of these modifying layers around the TiO₂NP core and induce apoptosis in MCF-7 cancer cells.

Zheng *et al.* synthesized a novel TiO_x (TiO_x: oxidized TiO₂) nanocomposite modified with PEG, targeting peptide YSA, and an anticancer drug cantharidin (CTD).¹⁹² In this nanosystem, PEG could enhance the stability of the nanoplatform and blood circulation time, which increased the tumor accumulation after systemic administration. The YSA peptide, with the YSAYPDSVPMMSK sequence, has been proven to be a targeting motif that mediates drug delivery to tumor cells expressing



EphA2. The anticancer drug cantharidin (CTD), as one of the active components of mylabris, was loaded into TiO_x for the combination of chemotherapy and PDT. The results showed that this nanosystem could significantly increase ROS production under X-ray exposure and provided a new drug delivery nanocarrier for CTD in combination with PDT to achieve more effective treatment.

In 2020, Kim *et al.* synthesized modified TiO₂NPs using a tumor targeting polymer phenylboronic acid (pPBA) to encapsulate the anticancer drug doxorubicin (DOX) as a sonodynamic chemotherapeutic agent.¹⁹³ In this system, the self-assembled TiO₂NP-DOX was encapsulated into polymeric phenylboronic acid (pPBA) *via* the formation of phenylboronic esters bonds, which are cleavable by ROS. Also, in this modified TiO₂NP nanocomposite, the phenylboronic acid (PBA) moiety acts as a tumor-targeting moiety because of its high affinity toward sialylated epitopes overexpressed on the membrane of various cancers. This polymeric modification also provided an enhanced loading of DOX by the interaction of PBA with 1,3-*cis* diol of DOX, and it was reported that DOX can bind on the surface of TiO₂NPs *via* the coordination of hydroquinone and the quinone moiety of the DOX structure. The loaded DOX was readily released by the sonodynamic ROS generation of TiO₂NPs due to the ROS-cleavable characteristics of the phenylboronic ester bond. Their results confirmed the tumor targeting by the PBA moiety, intracellular ultrasound-generated ROS, and high tumor accumulation of this nanosystem and its efficient anti-tumor effect on tumor-bearing mice.

Yu *et al.* modified Au@TiO₂NPs with poly(lactic-*co*-glycolic acid) (PLGA) followed by loading of the CPT-11 (irinotecan) drug as the targeting moiety.¹⁹⁴ The PLGA modification showed that these Au@TiO₂NPs-CPT-11-PLGA have an enhanced anti-metastatic activity and reduced cell invasion effects in B-CPAP and FTC-133 thyroid cancer cell lines, with and without NIR irradiation. In this nanosystem, the Au moiety could enhance the NIR absorption of Au@TiO₂NPs-CPT-11-PLGA, increasing the anticancer effect.

In 2021, Chen *et al.* prepared nanocomposites based on polypyrrole-coated mesoporous TiO₂NPs with a suitable size distribution for the co-delivery of doxorubicin (DOX) and aspirin prodrugs, with a superior drug loading capacity, due to the presence of the modifier, polypyrrole.¹⁹⁵ Also, these modified TiO₂NPs showed sonodynamic therapeutic properties and an excellent photothermal conversion efficiency (over 50.8%), with a simultaneous prodrug activation and sustained drug release, under near-infrared (NIR) and ultrasound (US) irradiation. The results showed an enhanced synergistic effect of chemotherapy and photo/sonodynamic effect to suppress the tumor.

As another biologically active polymeric modifier, polypyrrole (PPY) is an ideal photothermal conversion polymer with high photostability which has been successfully used in PTT. He *et al.* modified TiO₂NPs with polypyrrole (PPY), (mTiO₂NPs@PPY) to have the synergistic effect of TiO₂NPs and this polymer enhanced the photothermal effect of TiO₂NPs.¹⁹⁶ They used this modified mTiO₂NPs@PPY as a drug

carrier, a photothermal agent and a sonosensitizer, in a single nanoplatform. They loaded honokiol (HNK), as the model antitumor drug, which demonstrated antitumor efficacy in several cancer types such as breast cancer, pancreatic cancer, prostate cancer, lung cancer, hepatoma, and bladder cancer. The modified mTiO₂NPs@PPY showed the suitable size distribution and good biosafety, due to the presence of PPY on the surface of TiO₂NPs. The *in vitro* and *in vivo* animal experiments demonstrated that mTiO₂NPs@PPY-HNK could simultaneously have chemotherapeutic, photothermal, and sonodynamic effects under the laser and ultrasound irradiation. Other recent publications on the drug delivery application of TiO₂NPs are summarized in Table 7.

4.3. Antibacterials

The widespread overuse of traditional antibiotics has resulted in the emergence of multidrug-resistant bacterial strains and causes serious concerns in different aspects of life such as food safety and human health. In recent years, the research on new antimicrobial substances has focused on metal oxide nanoparticles. Specifically, TiO₂ nanostructures are one of the most attractive antimicrobial compounds, mainly due to their photocatalytic effect and chemical stability, low toxicity, and cost-effectiveness.⁹ Different research studies have shown that modified TiO₂NPs can demonstrate excellent antibacterial properties against a broad range of both Gram-positive and Gram-negative bacteria.^{10,204} This section presents the latest advancements and publications in the antibacterial activity of the modified TiO₂NPs. It is worth mentioning that the antibacterial effect of TiO₂NPs is due to their ability to absorb light (UV-Vis) to generate reactive oxygen species (ROS) which can be used to damage the chemical structure of microbes. Recently, Diana and Mathew reported the surface modification of TiO₂NPs with the alpha-lipoic acid (ALA) functionalized bovine serum albumin (BSA) conjugate, as a biocompatible antibacterial (and anticancer) system. The antibacterial ability of this nanosystem was studied against *S. aureus*, *E. coli*, *Streptococcus pneumoniae* (*S. pneumoniae*), *Candida albicans* (*C. albicans*), and *Aspergillus niger* (*A. niger*). The results proved the antimicrobial properties of the developed system and the *in vitro* cytotoxicity of these modified TiO₂NPs showed that the cytotoxicity was selective for cancer cells and negligible for normal cells.²⁰⁵ In another recent study, Maheswari *et al.* studied the antibacterial and anticancer properties of six TiO₂NP systems modified with three plant extracts including: *Withania somnifera* (Ashwagandha), *Eclipta prostrata* (Karisalankanni) and *Glycyrrhiza glabra* (Athimathuram), known as medicinal plants with pharmacological applications.²⁰⁶ The antibacterial features of these six samples were studied against three Gram-negative bacterial strains (*E. coli*, *Klebsiella pneumoniae* (*K. pneumoniae*), and *Pseudomonas aeruginosa* (*P. aeruginosa*)) and two Gram-positive bacterial strains (*S. aureus* and *Streptococcus mutans* (*S. mutans*)). Among the modified and unmodified TiO₂NP samples, *Withania somnifera*-*Eclipta prostrata* modified TiO₂NPs showed the good antibacterial nature against the studied bacteria. Also, these modified TiO₂NPs exhibited excellent



Table 7 Other research published in 2022 on the drug delivery application of TiO₂NP-based nanostructures

| Nanosystem | Applications | Ref. |
|---|---|------|
| Iron-supplement coated anatase TiO ₂ NPs modified with folic acid | Fe@TiO ₂ NPs showed a controlled pH-sensitive delivery of the loaded imatinib molecules | 197 |
| PLGA-TiO ₂ NPs (PLGA: poly(lactic-co-glycolic acid)) | Controlled release of a natural extract (international patent No. PCT/IB2020/061916) | 198 |
| Caffeic acid-mediated synthesis of TiO ₂ NPs (CA-TiO ₂ NPs) | The results indicated that CA-TiO ₂ NPs, as a promising compound with excellent biocompatibility, can be used in healthcare products and clinical and medicinal applications | 199 |
| GO-FA-PEG-TiO ₂ -Avi/Bio | Enhanced water solubility and potential anti-tumor activity and targeted co-delivery of anticancer drug, SN-38 | 200 |
| TiO ₂ NPs-polydopamine | Delivery of loaded icariin (Ica) for the improvement of the osseointegration process | 201 |
| TiO ₂ and mSiO ₂ drug delivery systems modified with folic acid | <i>In vitro</i> drug release of DOX experiments, hemolysis experiments, and cytotoxicity experiments on HeLa cell lines confirmed that the drug delivery system has good biocompatibility and GSH concentration-dependent drug release behavior | 202 |
| pH-Sensitive mesoporous bisphosphonate-based TiO ₂ NPs (modified with alendronate sodium trihydrate (AST)) | They were used as nanocarriers for dexamethasone (DEX) drug delivery. The pH-sensitive behavior of the NPs can be attributed to the presence of AST's amine groups in the hybrid nanoparticles | 203 |

anticancer activities against KB oral cancer cells, among the other bio modified and unmodified TiO₂NP samples. These results indicated the improved biological activities of TiO₂NPs after surface modification. In 2022, PV *et al.* synthesized a TiO₂/ZnO nanostructure by decorating ZnO nanoparticles over a commercial TiO₂ nanosurface.²⁰⁷ The ZnO formed over the anatase TiO₂ layer showed excellent antibacterial activity against both *S. aureus* and *E. coli* and was found to be non-toxic towards MG-63 osteosarcoma cells. In 2022, Goñi-Ciaurriz and Vélaz prepared polylactic acid (PLA) and cellulose acetate

(CA) composite films with β-cyclodextrin-modified TiO₂NPs (Fig. 5).¹²⁹

Benzoic acid (BA) and ascorbic acid (SA) were incorporated into β-cyclodextrin-modified TiO₂NPs, and the antibacterial activities of the PLA and CA composite films were successfully tested against *E. coli* and *S. aureus*. The highest antibacterial activity was observed with the film containing 5% modified TiO₂NPs achieving 71% inhibition of *E. coli* and 88% inhibition of *S. aureus*. The modification of the TiO₂NP surface with β-cyclodextrin provided an efficient carrier nanosystem and

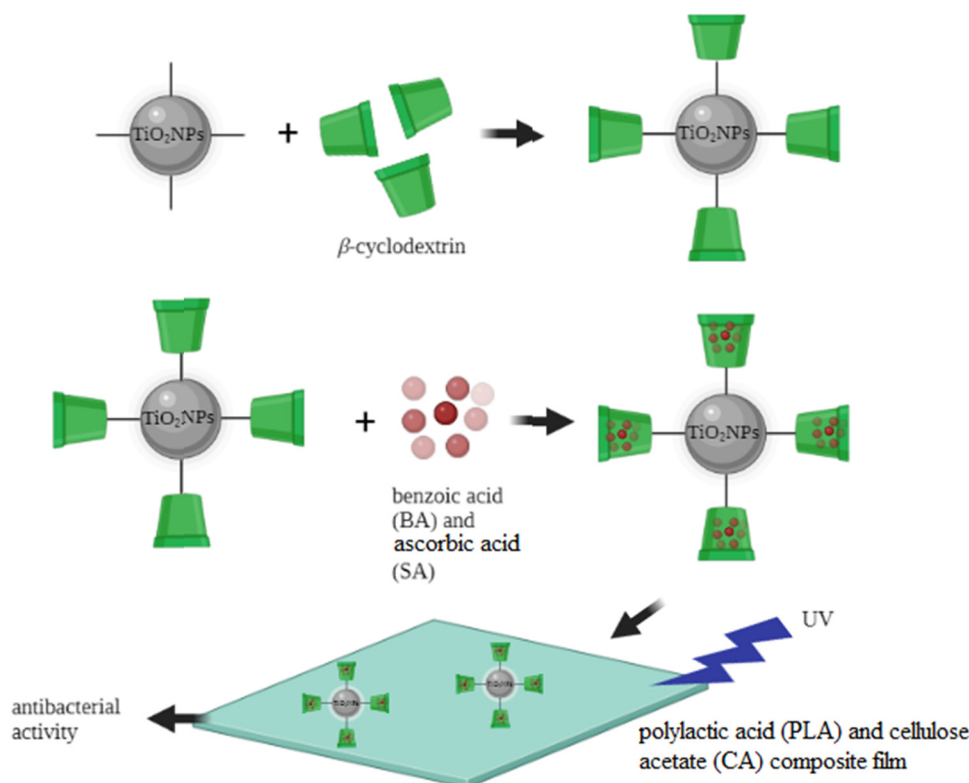


Fig. 5 β-Cyclodextrin-modified TiO₂NPs for antibacterial applications. Adapted with permission from ref. 129. Copyright 2022 Elsevier.



enhanced the therapeutic ability of the TiO₂NP core. Previously, these modified TiO₂NPs were successfully tested to load and release different food preservatives from the β-cyclodextrin grafted TiO₂NPs. The controlled release of therapeutic molecules from the cavity of β-cyclodextrin may extend the antimicrobial effect of the TiO₂NPs. Also, it can enhance the thermal stability of the film against volatilization or thermal conversion, when high temperature is applied in the food packing applications. Benzoic acid (BA) and sorbic acid (SA) are known as antimicrobial preservatives in food industry, due to their stable antimicrobial effectiveness against a broad range of microorganisms, including some bacteria, yeasts, and fungi. The authors used the polylactic acid (PLA)/cellulose acetate (CA) film, as a biodegradable polymer matrix, for fixation and stabilization of the modified TiO₂NPs, as potential active food packaging. The controlled release of benzoic acid (BA) and sorbic acid (SA) from the β-cyclodextrin-modified TiO₂NPs could dramatically improve the antimicrobial characters of the TiO₂NPs. These results indicated the great potential of this TiO₂NP-based nanocomposite to be used as antimicrobial food packaging.

Sathiyaseelan *et al.* synthesized modified TiO₂NPs using an aqueous extract of the endophytic fungus *Paraconiothyrium brasiliense* (Pb) to improve the antibacterial activity of common standard antibiotics at a minimum concentration.²⁰⁸ The modification of TiO₂NPs with the Pb fungus significantly enhanced the antimicrobial and antioxidant properties, biocompatibility, and stability of the TiO₂NPs. The authors used these modified TiO₂NPs with standard antibiotics (erythromycin, ampicillin, gentamicin, vancomycin, and tetracycline) to improve the antibacterial properties of these antibiotics without significant adverse effects. Antibacterial studies showed low activity of modified TiO₂NPs–Pb at a concentration of 20 μg mL⁻¹. However, a combination of tetracycline hydrochloride (TCH) with TiO₂NPs–Pb significantly enhanced the inhibition of the *E. coli* biofilm. The authors reported the moderate toxicity of TiO₂NPs–Pb (100 μg mL⁻¹) on the cell line NIH3T3, red blood cells (RBC), and egg embryos. This research revealed that the antibiotics could be mixed with the modified TiO₂NPs–Pb to improve the antibacterial efficiency and minimize antimicrobial resistance and environmental toxicity.

Özdemir *et al.* prepared modified TiO₂NPs using cotton fabric by hydrolysis of the TiCl₄ precursor solution over cotton fabric.²⁰⁹ The modified TiO₂NPs enhanced the photodegradation of rhodamine B, compared to the cotton fabric alone. The antibacterial activity of this modified TiO₂NP was successfully tested against *S. aureus* (ATCC 6538) and *E. coli* (ATCC 25922) as representative strains of Gram-positive and Gram-negative bacteria, respectively. In this modified TiO₂NP system, the presence of cotton fabric acted as a template to provide active sites for the adsorption of pollutant molecules and microorganisms and more importantly, this template facilitated the transfer of produced ROS to the target molecules for their degradations which resulted in a significant increase of the photocatalytic effect of the TiO₂NPs.

Metanawin and Metanawin used the mini-emulsion polymerization of the TiO₂NP–polystyrene (TiO₂NP–PS) hybrid

antibacterial material.²¹⁰ Triethylene glycol dimethacrylate (TEGDMA) was employed, as a crosslinking agent, to improve the stability/modification efficiency of TiO₂NPs and photocatalytic activity. Their results showed an excellent antibacterial effect of these TiO₂NPs–PS against both Gram-positive (*S. aureus*) and Gram-negative (*K. pneumoniae*) bacteria. Also, the photocatalytic efficiency of TiO₂NPs–PS was tested for the photodegradation of methylene blue under UV irradiation. The photocatalytic effect of TiO₂NPs–PS was increased in the presence of the crosslinking agent TEGDMA due to the self-organized structure of this hybrid system. These surface modifiers could enhance the surface area of TiO₂NPs which has paramount importance to enhance photocatalytic activity during photocatalysis.

Elbarbary *et al.* synthesized biodegradable poly(PVA/PLA/TiO₂NPs) nanocomposite films by combining polyvinyl alcohol (PVA) and polylactic acid (PLA) doped with TiO₂NPs.⁹⁰ The addition of 0.8 wt% of TiO₂NPs showed significant enhancement of the thermal stability of the films and the water resistance properties were obtained using a 2 : 1 PVA : PLA ratio. This poly(PVA/PLA/TiO₂NPs) nanocomposite displayed an improved antibacterial activity against *S. aureus* and *E. coli* strains. The biodegradation tests were performed in soil burial and the results showed a rapid increase of the degradation of the film in the initial 12 weeks with a significant change of morphology. These results suggested the potential application of this biodegradable poly(PVA/PLA/TiO₂NP) nanocomposite for developing packaging materials with low environmental impact. The authors used polylactic acid (PLA) because of its thermoplasticity and biodegradability with a wide range of potential industrial applications, such as packaging materials for fresh fruit and vegetables. However, due to the ester group in PLA, it has low mechanical/thermal stability, high rigidity, and poor hydrophilicity. To use PLA in the antibacterial films, it could be combined with synthetic polymers such as polyvinyl alcohol (PVA) to improve the properties of PLA. PVA is considered as a hydrophilic, biodegradable, biocompatible and cost-effective polymer, which is commonly studied in different biological applications such as drug delivery and food packaging. This PLA/PVA film was applied as a template for fixing TiO₂NPs, enhance their stability and antibacterial effectiveness.

Singh *et al.* conducted the green and cost-effective synthesis of TiO₂NPs using waste leaves of water hyacinth (WH) (*Eichhornia crassipes*), an aquatic weed, under ambient conditions.²¹¹ The antibacterial efficiency of these modified TiO₂NPs was tested on a commonly known toilet bacteria, *Serratia marcescens*. They reported a ~3.0 cm diameter of the inhibition zone at a 150 μg mL⁻¹ concentration of the nanocomposite which is superior to commercial TiO₂NPs and the WH leaf extract. This research showed great potential of the modified TiO₂NPs in healthcare industries. In this study, the authors used water hyacinth (WH) as a natural antimicrobial reagent for the TiO₂NP modification to improve the stability, biocompatibility, and antimicrobial effect of the TiO₂NPs.



Table 8 Other publications on the antibacterial applications of TiO₂NP-based nanostructures

| Nanosystem | Applications | Ref. |
|--|---|------|
| TiO ₂ NPs modified with bio agents: <i>Syzygium aromaticum</i> , <i>Eleteria cardamomum</i> , and <i>Cinnamomum verum</i> | Antibacterial and anticancer (KB oral cancer cell line) | 215 |
| Pure TiO ₂ nanoparticles and turmeric-, ginger-, garlic-modified TiO ₂ NPs | Antibacterial against five bacterial strains, anticancer activity against the KB oral cancer cell line | 216 |
| Pure TiO ₂ NPs, Aqua Rosa-modified TiO ₂ NPs and protein powder-modified TiO ₂ NPs | Antibacterial against five bacterial strains, anticancer activity against the KB oral cancer cell line | 217 |
| TiO ₂ NP incorporation into the heparin-polyvinyl alcohol nanocomposite | Enhanced <i>in vitro</i> antibacterial activity and care of <i>in vivo</i> burn injury | 218 |
| Polylactic acid/halloysite nanotubes-TiO ₂ NPs | High efficiency to both Gram-positive and Gram-negative bacteria | 219 |
| Nano-natural antimicrobial agent@polymeric microgels-TiO ₂ hybrid films | Antibacterial on the touch screen panel | 220 |
| Bio-nanocomposite film (polyvinyl alcohol)/TiO ₂ /chitosan/chlorophyll | Inhibits the growth of both <i>S. aureus</i> and <i>E. coli</i> bacteria under LED light irradiation | 221 |
| <i>In situ</i> coating of the TiO ₂ surface by plant-inspired tannic acid for the fabrication of thin film nano-composite nano-filtration membranes | Enhanced antibacterial performance | 222 |
| Functionalization of a polyvinylidene fluoride membrane by the biocidal oxine/TiO ₂ nanocomposite | Anti-biofouling properties | 223 |
| Cellulose acetate/TiO ₂ nanoparticles | Exhibited good antibacterial activity against <i>E. coli</i> with 55.6% sterilization in 12 h | 224 |
| Preparation of <i>monsonia burkeana</i> plant extracts | The material was found to be selective against <i>E. coli</i> . In real water samples, this material demonstrated remarkable activity | 225 |
| Polylactic acid (PLA)/TiO ₂ nanocomposite | Antibacterial activities with optimal inhibition zones against <i>S. aureus</i> followed by <i>E. coli</i> | 120 |

Mallakpour and Mohammadi prepared sodium alginate-pectin composite (ALG-PEC CS) and nanocomposite (NC) films, containing different concentrations of TiO₂NPs (0.5, 1, and 2 wt%).²¹² They used CaCl₂ and glutaraldehyde (Glu) as cross-linkers, which produce rigid scaffolds for hydroxyapatite (HA) sedimentation. The film containing 2 wt% TiO₂NPs exhibited the best bioactivity and biocompatibility on the MG-63 cell line, as well as the best antibacterial effect against *S. aureus*. The authors modified TiO₂NPs with the polymeric matrix to enhance the bioactivity and mechanical properties of implants. Generally, the Ti-OH surface groups facilitated the formation of HA on the composite solid surface. To facilitate the interaction between the implant and surrounding bone tissues, TiO₂NPs should be modified/incorporated with a polymeric substrate.

Youssef *et al.* synthesized TiO₂NPs and incorporated them into pure low-density polyethylene (TiO₂NPs-LDPE) at different concentrations (0.5, 1, and 2% weight of the polymer) for potential applications in the packaging materials industry.²¹³ Their antibacterial tests revealed the high ability of TiO₂NPs to generate the reactive radical species (ROS) which induced the antibacterial activity of LDPE against Gram-negative and Gram-positive bacteria. Among synthetic polymers, low-density polyethylene (LDPE) is commonly used in food packaging due to its thermal stability and flexibility, cost-effectiveness, transparency, ease of processability, and biocompatibility. Most of the antimicrobial food packaging studies have focused on LDPE polymers and for these reasons this polymer can be a good candidate for the surface modification of TiO₂NPs, as a biologically active agent to enhance the effectiveness of TiO₂NPs and their biocompatibility in the food packing industry.

Makableh *et al.* investigated the addition of TiO₂NPs and ciprofloxacin (CIPRO) to polydimethylsiloxane (PDMS) to

enhance the antibacterial activity and hydrophobicity.²¹⁴ The nanocomposite of PDMS was prepared by combining the TiO₂NPs and/or CIPRO with PDMS before the crosslinking step. Various loading concentrations of TiO₂NPs (1–5 wt%) were used while the CIPRO concentration was fixed at 0.5 wt%. The antibacterial results revealed the synergistic effect of both TiO₂NPs and ciprofloxacin which led to an enhanced antibacterial activity against *S. aureus* and *E. coli*. The PDMS polymer has self-healing properties, biocompatibility, cost-effectiveness, high flexibility, and antimicrobial activity. Ciprofloxacin (CIPRO) is also known as a fluoroquinolone drug and considered as one of the most bactericidal agents used widely. So, the modification of TiO₂NPs with the PDMS polymer and CIPRO could be a promising strategy to enhance the biological effectiveness of TiO₂NPs and decrease their potential toxicity.

Other recent studies on the antibacterial applications of TiO₂NP-based nanostructures are summarized in Table 8.

4.4. Biosensors

In recent years, there has been an admirable effort to develop TiO₂NP-based biosensors,²²⁶ specifically for the development of novel biomolecule-TiO₂NP systems leading to a dramatic success in the fabrication of bio-nanohybrid devices, such as biomolecule-sensitized solar cells (BSSCs) and photoelectrochemical cells (PECs).²²⁷ The high sensitivity of such biosensors can provide opportunities to improve clinical methods in monitoring the patient's response to medical or surgical therapy. A biosensor should have several essential characteristics for the practical applications, including biocompatibility, cost-effectiveness, user-friendliness, low sensitive detection limit/high accuracy, rapid response, and easy manufacturing for the large-scale production.²²⁸ In this regard, TiO₂-based nanostructures can fulfill the mentioned properties to be used in



biosensing applications and there has been an extensive scientific work in this area due to their unique electron-transfer properties.^{229,230} For instance, in 2022, Feng *et al.* introduced a new fluorescence method to detect the tyrosine phosphatase 1B protein (PTP1B) using modified TiO₂NPs/single-wall carbon nanohorns (TiO₂NPs-SWCNHs) (Fig. 6).¹³ Single-walled carbon nanohorns (SWCNHs) are a new type of carbon nanomaterials which have a large specific surface area, internal space, and fluorescence-quenching ability to construct optical systems, such as SWCNH-based detection systems for biological molecules. In this study, the TiO₂NPs were decorated with SCWNHs (TiO₂-SWCNHs) for providing the on/off quencher moiety on the TiO₂NP surface which enhanced the discrimination difference in SWCNHs between the phosphorylated and nonphosphorylated peptides. This work was reported as the first TiO₂NPs-SWCNHs.

The resultant TiO₂-SWCNH nanocomposite could effectively quench the fluorescence of the phosphorylated-peptide substrate labeled by the fluorophore with a low fluorescence background. In the presence of the target PTP1B protein, dephosphorylation of the attached peptide occurred (due to the PTP1B/peptide reaction), resulting in a detachment of the dye-labeled peptide from the TiO₂-SWCNH surface and fluorescence enhancement was observed in the system. This demonstrated a simple and fast approach to detect PTP1B activity, having an ultra-low detection limit of 0.01 ng mL⁻¹ with a linear range of 0–10 ng mL⁻¹. The biosensor can be used in the serum medium using the standard addition method and showed the possibility for screening PTP1B inhibitors.

Tao *et al.* prepared TiO₂NPs modified with graphitized carbon nanofibers (TiO₂NPs/GNFs) for the sensitive detection

of organophosphorus pesticide residues (OPs).²³¹ The modification of TiO₂NPs with GNF resulted in enhanced biocompatibility, catalytic properties, and conductivity, and provided a hydrophilic surface for the effective immobilization of acetylcholinesterase (AChE), as the recognizing moiety. In more detail, the Ti atoms of this nanosurface coordinated with AChE to enhance its stability, and TiO₂ has a high tendency for adsorption on OPs. The AChE/TiO₂/GNFs/GCE biosensor exhibited a high affinity to acetylthiocholine chloride (ATCh) and demonstrated a low detection limit (3.3 fM) with a wide detection linear range (1.0 × 10⁻¹³–1.0 × 10⁻⁸ M), for paraoxon detection (a model of OPs). It was successfully tested for the determination of OPs in lake water, showing high anti-interference, long-term stability, and acceptable reproducibility, with great potential for the analysis of OPs in ecological environments.

Hong *et al.* developed a label-free electrochemical immunosensor for the ultra-sensitive determination of β-lactoglobulin (β-LG), in which they used TiO₂NPs, carbon nanochips, and AuNPs on chitosan (as a conducting polymer).²³² This biosensor demonstrated a linear relationship between the log β-LG concentration and the square wave voltammetry (SWV) response, with a detection limit of 0.01 pg mL⁻¹. Due to its high stability, reproducibility, and sensitivity, this approach can be applied for the detection of β-LG in real food samples. In this study, TiO₂NPs incorporated with chitosan (CS), as a biopolymer having high adhesion ability, biocompatibility and mechanical strength and improve the stability of the electrode surface for the biosensing applications. Due to the low conductivity of CS, they used carbon nanochips and AuNPs to facilitate electron transfer and increase the conductivity of this TiO₂NP-based nanosensor.

Shi *et al.* proposed a novel biosensor for a highly sensitive detection of H9N2 AIV. In fact, the H9N2 subtype avian

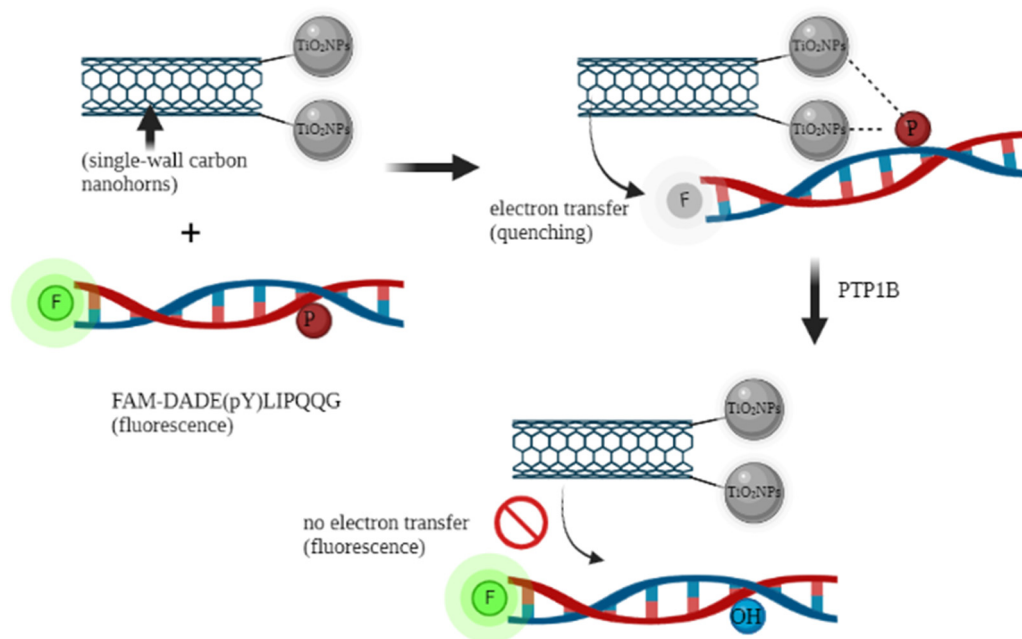


Fig. 6 Modified TiO₂NP single-wall carbon nanohorns (TiO₂-SWCNHs) for detecting the tyrosine phosphatase 1B protein (PTP1B). Adapted with permission from ref. 13. Copyright 2022 Elsevier.



Table 9 Recent publications on modified TiO₂NPs applied as biosensors

| Nanosystem | Applications | Ref. |
|--|--|------|
| TiO ₂ NRs and graphene oxide | For detecting dichlorvos (DDVP) | 237 |
| TiO ₂ nanotubes and AgNPs | Heat shock protein 70 (HSP70) as a potential tumor marker with high diagnostic sensitivity | 238 |
| Gallic acid–TiO ₂ nano-composites | Detection of DNA | 239 |
| TiO ₂ nanotube (NT) arrays | Tumor cell detection | 240 |
| 11-Mercaptoundecanoic acid self-assembly and the amidated nano-TiO ₂ film | For the selective and ultrafast detection of phosphoproteins in food | 241 |
| Nanocomposite graphene/TiO ₂ | Glucose biosensor | 242 |
| TiO ₂ -graphene composite modified carbon paste electrode | Determination of sufentanil in human plasma and urine | 243 |

influenza virus (AIV) is a low-pathogenicity AIV that seriously threatens the healthy development of the poultry industry and public health systems.²³³ The sensor was constructed by employing a dual-resonance long-period fiber grating (DR-LPFG) modified with TiO₂NPs, followed by the chemical attachment of the anti-H9N2 monoclonal antibody (anti-H9N2 MAbs) to the TiO₂NPs on the surface of DR-LPFG. The detection limit of this biosensor was estimated to be $\sim 2.7 \text{ ng mL}^{-1}$ with a high specificity and rapid detection of 96.1%, which is higher than that of a DR-LPFG-based biosensor modified with the Eudragit L100 copolymer. In this system, DR-LPFG provided a stabilizing medium for the TiO₂NPs which increased biocompatibility under biological conditions. The attachment of the monoclonal antibody (anti-H9N2 MAbs) to the TiO₂NPs resulted in high selectivity of this biosensor to detect H9N2 AIV.

Rajeshwari *et al.* combined poly(*p*-phenylenediamine) with TiO₂ and a multiwalled carbon nanotube to make a biosensor nanocomposite for the *in vivo* detection of dopamine, as a biomarker of many mental illnesses.²³⁴ The biosensor demonstrated a considerable sensitivity with a linear range of 3.81×10^{-11} – $4.76 \times 10^{-6} \text{ M}$ with a low detection limit of $9.45 \times 10^{-12} \text{ M}$. The incorporation of TiO₂NPs with poly(*p*-phenylenediamine) and carbon nanotubes significantly enhanced the conductivity of the TiO₂NPs, along with its stability and biocompatibility.

Zheng *et al.* used HKUST-1 MOFs (molecular organic frameworks) and its derivative, HKUST-CuO, for incorporation with TiO₂NPs to form two resultant composites of HKUST-1/TiO₂ and HKUST-CuO/TiO₂ to modify the electronic properties of TiO₂NPs and make a well-suitable band gap energies (E_g).²³⁵ Compared with mono-component HKUST-1 or HKUST-CuO, both TiO₂-based composites showed a synergistic photoelectrochemical (PEC) response due to their heterogeneous structure. The HKUST-CuO/TiO₂-modified electrode showed a higher photocurrent response which may be due to its hollow structure, greatly enhancing visible light harvesting. Then the authors successfully attached the targeting moiety to this nano-hybrid and fabricated the S1 (probe DNA)/HKUST-CuO/TiO₂/ITO PEC platform for colitoxin DNA detection without using ascorbic acid (AA) as an electron donor. Compared with S1/HKUST-1/TiO₂/ITO, the S1/HKUST-CuO/TiO₂/ITO electrode demonstrated a wider linear response range (1.0×10^{-6} – $4.0 \times 10^{-1} \text{ nM}$) with a lower detection limit of $3.73 \times 10^{-7} \text{ nM}$ (S/N = 3). Due to its good specificity and stability, this biosensor exhibited a promising strategy for molecular diagnosis in the bio-analysis field.

Singh *et al.* developed an electrochemical biosensor for the detection of organophosphorus (OP) pesticides based upon AChE-inhibition, operating in the pM concentration range.²³⁶ The synthesized TiO₂NPs and molybdenum disulfide nanomaterials were deposited on a screen-printed electrode, followed by modification with chitosan and immobilization of AChE on the modified electrode. The AChE modification of this TiO₂-based electrode provided a selectivity for this biosensor. Also, the chitosan modification resulted in an enhanced stability and biocompatibility of the TiO₂NPs on the surface of this electrode. The biosensor was successfully tested for the low OP pesticide concentration detection in forensic visceral samples demonstrating a low detection limit of 50 pM. Other recent works on the biosensing applications of modified TiO₂NPs are summarized in Table 9.

4.5. Antifungal

More than 300 million people worldwide are being threatened by severe fungal infections which have caused 1.6 million deaths every year.²⁴⁴ To overcome this issue, many nano-based approaches have been developed, and among them, the photocatalytic deactivation of fungi has become a promising strategy for disinfection of aqueous media to have microbial control.^{245–248} As a semiconductor material, TiO₂NPs have been introduced as a great candidate for the development of advanced antifungal agents.

For instance, in 2022, Wang fabricated chitosan/alginate–TiO₂NP based bilayer films incorporated with different concentrations of cinnamon essential oil (CEO) to study the effect of this bilayer film on improving the postharvest quality of mangoes.²⁴⁹ In this study, chitosan was used as a packaging material for food preservation due to its biodegradable, antibacterial, and good film-forming properties. The cinnamon essential oil (CEO) is also considered as a natural antioxidant and antibacterial agent and has attracted increasing attention in the field of packaging material. In this study, the film of chitosan/CEO was formed by the interaction of the aldehyde group of CEO with the amino group of the chitosan matrix to improve the hydrophobicity, antibacterial, and antioxidant properties of the chitosan films. To improve the photostability of this film, the modified TiO₂NPs–alginate was used as anti-ultraviolet packaging materials, in which the alginate contains –COO[−] functional groups which can electrostatically interact with cations of the inner chitosan layer to form a bilayer to



prevent the volatilization of CEO in the inner layer. The modification of TiO₂NPs with alginate improved the antimicrobial performance of the outer part of this film due to their synergistic advantages of TiO₂NPs and alginate. This TiO₂NP-based film demonstrated an improved mechanical and antimicrobial properties for the film which could be a promising candidate as a multifunctional packaging material to maintain the freshness of harvested mangoes.

Siddiqui *et al.* fabricated TiO₂NPs using 37 strains of cyanobacteria and evaluated their antifungal, antioxidant, antibacterial, and hemolytic activities.²⁵⁰ *Synechocystis* NCCU-370 was introduced as the best strain for the synthesis of TiO₂NPs in terms of size (73.39 nm), followed by optimization of the synthesis to obtain smaller nanoparticles (an average grain size of 16 nm). The antifungal activity was studied against *Candida albicans* (MIC = 125 μg mL⁻¹), *Candida glabrata* (MIC = 500 μg mL⁻¹), and *Candida tropicalis* (MIC = 250 μg mL⁻¹), and the modified TiO₂NPs demonstrated the partial synergistic effect and excellent biocompatibility. The biocompatible nature of these biomodified TiO₂NPs is an advantage for their potential in biomedical purposes.

Sultan *et al.* fabricated gelatin active packaging films based on nano-sized droplets of coconut oil emulsified by pickering emulsion (PE) and stabilized by chitosan/Arabic gum (CH/AG) nanoparticles, in the presence of TiO₂NPs.²⁵¹ The films showed a significant antifungal activity for all tested microorganisms, such as *Bacillus cereus* and *C. albicans*. The antimicrobial and antioxidant packaging materials are frequently produced by embedding natural antimicrobial and antioxidant additives into the natural polymer matrices providing new functionalities to the film and extend shelf life of packaged food. As a natural biopolymer, gelatin shows excellent biodegradability, biocompatibility, and film-forming ability. The coconut oil is another biocompatible candidate for using the packaging biofilm due to its potential antimicrobial and antioxidant characteristics. Chitosan is also considered as an excellent biocompatible, non-toxic, and biodegradable material, showing some important functions such as antibacterial and antifungal properties. Arabic gum (AG) is the last organic component of this film, having amphiphilic polysaccharides with emulsifying properties. These organic polymeric matrices provided a stabilizing biocompatible medium for the TiO₂NPs to synergistically enhance the antifungal properties of the individual components of this film.

Mohammad Taghizadeh Kashania *et al.* synthesized TiO₂NPs modified with *C. arabicus* and studied their effect on improving the biological properties of the dichloromethane fraction (DF) of *C. arabicus* root smoke (the largest species in the Costaceae plant family).²⁵² The synthesized DF/TiO₂NPs (200 mg L⁻¹) showed the maximum radical scavenging level up to the IC₅₀ = 8.31 μg mL⁻¹. In this study, the TiO₂NPs were modified with the biologically active biomolecule, *C. arabicus* which is known as a good candidate for the treatment of infectious diseases. The modification of TiO₂NPs with this biomolecule can provide the synergistic effect of antifungal for this modified system and can decrease the potential toxicity of TiO₂NPs.

Duan *et al.* prepared a nanocomposite film made by *K-carrageenan* (KC), *Konjac glucomannan* (KGM) and TiO₂NPs. The TiO₂NPs improved the mechanical and thermal properties of the KC/KGM films.²⁵³ Specifically, the film containing 7 wt% of TiO₂NPs showed effective photocatalytic antifungal activity (79%) against *Penicillium viridicatum* after irradiation for 6 h and revealed a protective effect on strawberry storage. The results demonstrated that the nanocomposite films have a broad potential for food preservation and packaging applications. For the food packaging applications, *K-carrageenan* (KC) is a hydrophilic biomolecule, obtained from red seaweed, with gelling and film-forming properties which allow biodegradable packing films to be produced. Also, *Konjac glucomannan* (KGM) is a natural polysaccharide which has been widely used to prepare film materials due to its good film-forming ability. Thus, when TiO₂NPs are incorporated into these polymer matrices, it will effectively inhibit bacterial growth and food spoilage due to the synergistic effects of these three components to improve the properties of each other. Other studies on the antifungal applications of modified TiO₂NPs are summarized in Table 10.

4.6. Antiviral

Since 2019, with the emergence of pathogenic human coronavirus pandemic, SARS-CoV-2 (COVID-19) has caused serious issues in many aspects of life such as public health and economy, all around the world. Based on the 2019 World Health Organization prediction, the mortality of infection-related diseases will be similar to that of cancer by 2050.^{11,12,261–263} Modified TiO₂NPs have provided some promising candidates for controlling virus-type infections, supported by recent worthwhile publications in this frontier area of nanomedicine.

Because of the paramount importance of antiviral modified TiO₂NP systems, recent publications in this field are presented; for instance, in 2022, Elsayed *et al.* studied the condensation of 3-acetylindol, thiophene-2-carbaldehyde and malononitrile in the presence of TiO₂NPs, yielded 2-amino-6-(1*H*-indol-3-yl)-4-(thiophen-2-yl)-4*H*-pyran-3-carbonitrile derivatives.²⁶⁴ Then, they were reacted with formic acid, formamide, ethyl chloroacetate, chloroacetyl chloride, thiourea and sodium nitrite to form several three-combination systems which were safe, ecologically friendly, and non-toxic. The synthesized compounds were successfully tested for antiviral activity against Vero cells (HAV) and showed an effective activity. In this research, new indoles were synthesized and used for the TiO₂NP modification and tested for their antioxidant's outcome. The indole nominees verified their strength as antioxidants, antimicrobial, and anticancer. Specifically, the authors used synthesized contestants containing pyrimidine, pyrazole, pyrane, and pyridine rings. As a crucial nucleobase, pyrimidine derivatives are considered as antioxidant agents in contrast to ROS and reactive nitrogen species (RNS). Similarly, pyridine, pyrazole, and pyrane rings displayed antioxidants properties in their derivatives. The modification of the TiO₂NP surface with these compounds



Table 10 Recent studies of antifungal applications of the TiO₂NPs

| Nanosystem | Applications | Ref. |
|--|--|------|
| TiO ₂ NPs were produced by <i>Bacillus sp.</i> bacteria | Significant antifungal activities against the oral <i>C. albicans</i> pathogen | 254 |
| Green synthesis of S-doped TiO ₂ NPs using <i>Malva parviflora</i> plant extract | Antimicrobial and antioxidant activities under sunlight illumination | 255 |
| TiO ₂ /Ag nanoparticles | For its activity as an antifungal material for the inhibition of <i>C. albicans</i> in water under visible light irradiation | 256 |
| TiO ₂ -SiO ₂ /chitosan | Enhancement of antifungal capability | 257 |
| TiO ₂ NPs were synthesized by using <i>trianthema portulacastrum</i> , <i>chenopodium quinoa</i> leaf extracts and sol-gel method | Antifungal activities against wheat rust | 258 |
| Cyclodextrin-grafted TiO ₂ NPs | As food preservative carriers | 259 |
| PVA/TiO ₂ -based nanocomposites | Antifungal activity study | 260 |

could provide opportunity to increase the antiviral effect of TiO₂NPs.

Souza *et al.* developed TiO₂-based antiviral hydrophobic cellulose cotton or non-woven fabrics for their potential virucidal effect on Murine Coronavirus (MHV-3) and Human Adenovirus (HAdV-5), under indoor light irradiation.²⁶⁵ In the non-woven fabric, the results demonstrated 90% and 99% reduction of HAdV-5 and MHV-3, respectively, with no reduction of HAdV-5 in cotton fabric. The antiviral activity was assigned to the photocatalytic effects of the modified TiO₂ powders, and the hydrophobic properties of fabrics and high surface of the TiO₂ particles facilitated their interaction with the viruses, especially MHV-3. These results showed the potential ability of these composite materials as highly effective virucidal agents against MHV-3 and HAdV-5 viruses, particularly for applications in healthcare indoor contaminated environments. In fact, the fabrics used in this study acted as a support for TiO₂NPs to significantly promote the interaction between viruses and TiO₂NPs. In this regard, cellulose-based fabrics such as cotton and non-woven fabrics have been commonly studied for the application of TiO₂ hydrosols to prepare fabrics with photocatalytic and self-cleaning properties. The flexible, porous, and layered surface structures of cotton contributed to the incorporation of TiO₂NPs in its structure, providing enhanced antiviral properties.

Regarding SARS-CoV-2 infection, Da Silva *et al.* developed antimicrobial cotton fabrics based on the Ag/TiO₂ nanohybrid and they showed that more than 50% of infectious SARS-CoV-2 survived after direct contact with the nanohybrid under the tested conditions, which indicated that more studies are required on using silver and TiO₂ nanostructures as self-disinfecting agents for the prevention of coronavirus transmission.²⁶⁶ In this case, the cotton provided a matrix for the immobilization of Ag/TiO₂NPs, which protected the nanosystem against aggregation.

Wang *et al.* successfully tested the antiviral efficacy of TiO₂-chitosan (CS)-AgNP filter for viral aerosols and reported the infection risk reduction and long-term antiviral efficacy of this nanosystem.²⁶⁷ In this study, the TiO₂-CS-AgNP filter was synthesized for the removal and deactivation of airborne MS2 bacteriophage particles. In the air purification system, their results showed a 93% removal of the airborne MS2 particles, and more than 95% of MS2 can be efficiently deactivated on the

surface of this nanosystem within 20 minutes. The filter could maintain 50% of its original antiviral efficiency after continuous operating for 1 week. The surface modification of TiO₂ with the chitosan polymer provided possibility for the AgNP attachment to the surface of TiO₂.

Levina *et al.* used a combination of biocompatible TiO₂NPs and immobilized polylysine-containing oligonucleotides (PL) with native (ODN) and partially modified (ODNm) internucleotide to form a new delivery system for the effective attack of oligonucleotides on the viral genome of highly pathogenic H5N1 influenza A virus (IAV) *in vivo*.²⁶⁸ The intraperitoneal injection of this TiO₂-PL-ODN nanocomposite exhibited 65–70% survival of mice, while the intraperitoneal or oral administration of TiO₂-PL-ODN was more efficient (~80% survival). The nanocomposites showed no toxicity on mice under the tested conditions. Interestingly, the TiO₂NPs, unbound ODN, and the nanocomposite bearing the random oligonucleotide demonstrated a low protective effect, demonstrating the key role of targeting oligonucleotides in this nanocomposite for the site-specific interaction with complementary RNAs of the target virus. In this antiviral system, the ODN loaded polylysine was non-covalently immobilized onto the TiO₂NPs, which enhanced the biocompatibility and enabled the ODN delivery *via* the cell membrane. More importantly, this modification of TiO₂NPs could stabilize and protect ODN against intracellular enzymes, and the carried oligonucleotides were released from the nanocomposites in the cytoplasm or penetrated into the nuclei and bind to the RNA molecules.

León-Gutiérrez *et al.* studied the modification of TiO₂NPs with secondary metabolites implanted to prepare antiviral TiO₂NPs (SMNP) and tested them on SARS-CoV-2 infectivity and healthy cells as well. Surprisingly, SMNP showed a considerable reduction of viral infectivity *in vitro* with minimal toxicity to healthy cells when compared to other commercially available antiseptics (glutaraldehyde, chlorine, chlorhexidine, ethanol, and Lysol™), which indicated this SMNP nanosystem as a safe and effective antiviral against SARS-CoV-2.²⁶⁹ Citrus-derived compounds have shown the clear clinical benefit for viral infections, inducing stimulate immunity. Natural secondary metabolites are considered as antiviral agents due to their inhibitory effect on key metabolic enzymes that influence signaling pathways, cellular function, and gene expression. Conjugation of these therapeutic agents with the TiO₂NP



surface could provide the synergistic effect for the antiviral ability of the modified TiO₂NPs.

4.7. Tissue engineering

Tissue engineering is a multidisciplinary field which includes the fabrication of these biocompatible materials suitable for repairs or replacement of abnormal tissues/organs. It is worth mentioning that the scaffolds, used for different applications in tissue engineering, need to be highly compatible with the cellular matrix inside the body and they should not cause cytotoxic, immunogenic, inflammatory, or any other host reaction. Also, they should exhibit suitable mechanical and physical properties suitable for the biological conditions. Despite many advantages of TiO₂NPs in tissue engineering, one of their restrictions is their agglomeration which diminishes their biological effectiveness. Therefore, there is a need for a polymeric matrix to fix and stabilize these TiO₂NPs and prevent their leaching out to different parts of the body. In recent years, modified TiO₂NPs have attracted significant attention for using in different platforms/scaffolds in the tissue engineering field to promote biological and physicochemical processes of cell/tissue culturing.^{270,271} In this field, the TiO₂NPs are often used as a part of the biocompatible polymeric matrix which can fix and stabilize TiO₂NPs, enhancing the biocompatibility and effectiveness of these nanoparticles. Simultaneously, these incorporated TiO₂NPs can improve the mechanical and physicochemical properties of the scaffolds.

For bone regeneration scaffolds, in 2022, Karbowniczek *et al.* reported the additions of hydroxyapatite (HA) and TiO₂NPs on poly(3-hydroxybutyrate-co-3-hydroxyvalerate) (PHBV) based fibers and studied the tensile strength, elongation, and toughness of the fibers after this addition.¹⁴ It should be mentioned that the biodegradable poly(3-hydroxybutyrate-co-3-hydroxyvalerate) (PHBV) is a biopolymer synthesized by bacteria, considered as a good alternative for many non-biodegradable synthetic polymers. For tissue engineering purposes, this biopolymer can be processed *via* electrospinning for the construction of scaffolds. To improve its *in vitro* cell growth and mechanical properties, PHBV can be combined with ceramic particles such as hydroxyapatite (HA) and antibacterial TiO₂NPs. Regarding the effect of HA on the surface properties of TiO₂NPs, the authors reported that the presence of HA could decrease the agglomeration of TiO₂NPs in the PHBV + HA + TiO₂ composite, compared to that in PHBV + TiO₂, which is very important for increasing the effectiveness of TiO₂NPs. Also, they observed a dramatic improvement in the mechanical strength of the PHBV fibers containing HA nanoparticles, compared with the fibers alone. The homogenous distribution of HA nanoparticles resulted in a 3-time improved tensile strength and a 16-time higher toughness. The authors showed a strategy for tuning mechanical properties by controlling the size and distribution of ceramic fillers (TiO₂ and HA) in hybrid scaffolds.

As another surface modifier, poly-*ortho*-toluidine (POT) has been used as a conductive polymer to stimulate a multitude of cell functions such as attachment, proliferation, migration, and

differentiation *via* the modulation of transferred electrical stimuli from the external support to cell. So, the modification of TiO₂NPs with the POT polymer brings several benefits to the TiO₂NPs, including promoted cell-material interaction, better antibacterial activity, and excellent biocompatibility. In this regard, Balan *et al.* synthesized an organic/inorganic nanocomposite poly-*ortho*-toluidine-TiO₂ to construct the POT + TiO₂/PCL nanocomposite scaffolds.²⁷² The surface roughness of this nanocomposite provided a great influence on the viability of different cells. Besides, the *in vitro* antibacterial activities of POT + TiO₂ and POT + TiO₂/PCL composite scaffolds were tested on *S. aureus* and *E. coli* and the POT + TiO₂/PCL scaffold demonstrated an improved surface roughness, cell viability and antibacterial activity, indicating economical and effective TiO₂NP-based nanocomposites for tissue engineering applications.

Sharaf Saeed *et al.* prepared a series of poly(ethylene-co-vinyl alcohol)/TiO₂NPs (PEVAL/TiO₂) nanocomposites containing 1, 2, 3, 4 and 5 weight ratios of TiO₂NPs.²⁷³ The cell culture tests of these nanohybrids were evaluated on human gingival fibroblast cells (HGFs) in accordance with ISO 10993-5 and ISO 10993-12 standards, with studying the cell viability after 1, 4, and 7 days. The results showed a time-dependent improvement in the cell activity for all systems, and the cell survival for all samples was higher than that of the virgin PEVAL on day 7 ($p < 0.002$). The bio-SEM results also demonstrated the successful cell adhesion and growth of HGFs on all types of scaffolds (PEVAL/TiO₂). In this system, the incorporation of TiO₂NPs into the PEVAL polymer matrix exhibited a uniform dispersion of these TiO₂ fillers, which are well covered by the copolymer. This could be due to the presence of good affinity between these two components in which the expansion PEVAL macromolecule chain in the solvent promotes the dislocation of the aggregated TiO₂, leading to their uniform dispersion in the polymer matrix. This homogeneity and stability could positively affect the biocompatibility of TiO₂NPs for the cell culturing purpose.

Alginate is considered as one of the most promising surface modifiers for TiO₂NPs in the tissue engineering field, as a marine-based polysaccharide found in brown algae. Compared to synthetic polymers, this biopolymer provides several benefits such as biocompatibility, gel-forming ability at biological pH and temperature, non-toxicity, water solubility, and cost-effectiveness. Alginate-based scaffolds can enhance alkaline phosphatase activity and expressing osteocalcin and mineralization resulting in promoted osteogenesis. In this case, Mallakpour and Naghdi applied a TiO₂NPs-alginate nanohybrid to fabricate a bone scaffold to take the benefits of both TiO₂NPs and alginate.²⁷⁴ In this study, the alginate provided suitable conditions for the biomineralization process, and it can help better dispersion and fixation of TiO₂NPs and decrease their aggregations. This TiO₂NP-alginate was incubated in a simulated body fluid at 37 °C for 28 days to evaluate its bioactivity. Cytotoxicity tests were performed on the MG-63 cell line and the scaffold showed no toxicity effects. Besides, this scaffold demonstrated a potent antibacterial effect against the Gram positive strain *S. aureus*, indicating the great properties of this scaffold to be used for bone tissue engineering applications.



As another well-known polymer in tissue engineering, polyvinyl alcohol (PVA) shows great film-forming properties, high tensile strength, biodegradability, emulsifying characteristics, and water solubility. However, for using scaffolds, it requires to blend this polymer with other polymers and inorganic fillers to improve the mechanical and biological properties.²⁷⁵ Currently, nano-cellulose has been attracting considerable interest due to its strengthening effect, biodegradability, high surface area, non-toxicity, and great mechanical/optical properties. Cellulose can also support cell attachment and facilitate apatite growth. Because of these typical properties, they have been incorporated into nanocomposites and the polymer matrix as reinforcing materials improving the desired properties, which makes it a good choice for biomedical applications.²⁷⁵ TiO₂NPs are ideal illustration of inorganic fillers due to their cell adhesion and anti-corrosive properties. Fatima *et al.* prepared a TiO₂NP-doped ternary nanocomposite film (PVA/NC-TiO₂) by the fusion of polyvinyl alcohol (PVA), nano-cellulose (NC), and TiO₂NPs (0.01 wt%), as well as its binary nanocomposite film (PVA/NC).²⁷⁵ The presence of TiO₂NPs in the structure of this film significantly improved the thermo-mechanical stability of the nanocomposite film, which can be due to an enhancement of intermolecular interactions in the PVA/NC-TiO₂ film, compared to the PVA/NC binary film. Furthermore, their results showed a higher glass transition temperature and a storage modulus of the PVA/NC-TiO₂ film (9131.26 MPa, 74.3 °C) than those of the PVA/NC film (7588 MPa, 64.3 °C). The ternary PVA/NC-TiO₂ film showed a more uniform surface layer of the Ca-P mineral than the PVA/NC film, upon incubation in simulated buffer saline. The low percentage of TiO₂NPs improved the strength of this PVA/NC-TiO₂ film, with a cell viability more than 80% and hemolysis less than 1.7%, demonstrating high potential of this nanocomposite to develop better bone templates for bone regeneration applications. The interaction of PVA and NC with the TiO₂NPs resulted in more homogeneity for the particles in this film, compared to the PVA/NC film. Besides, this surface modification improves the biocompatibility of these TiO₂NPs.

Nanofiber (NF) scaffolds hold great promise for utilization in bone regeneration purposes and they are interesting stabilizing and modifying matrices for enhancing the efficiency of TiO₂NPs used in the scaffolds. Among various biomaterials, poly-ε-caprolactone (PCL), an FDA approved biodegradable polyester, has attracted much attention in this field due to its appropriate biocompatibility and mechanical properties.²⁷⁶ Several works demonstrated that a blend of PCL and gelatin (GEL) can generate a highly biomimetic nanofibrous scaffold with appropriate biodegradability and mechanical stability, and most importantly excellent cytocompatibility. However, PCL/GEL NFs alone are not appropriate for bone regeneration due to the absence of inherent osteoinductive capability. The incorporation of osteoinductive agents or components to these scaffolds can lead to higher mechanical properties and improved biofunctionality of PCL/GEL NFs for bone regeneration. In this regard, nanocomposites based on ceramic nanoparticles such as TiO₂NPs have recently drawn considerable attention as bone substitute materials and injectable pastes to

fill defects due to their excellent mechanical properties, high chemical stability, and bioactivity.²⁷⁶ In this regard, Ahmadi *et al.* employed TiO₂NPs and metformin (MET)-co-embedded composite electrospun poly-ε-caprolactone/gelatin nanofibers (PCL/GEL NFs) to form scaffolds for the improved osteoblastic differentiation of human adipose-derived stem cells (hADSCs).²⁷⁶ The results showed an improvement of the mechanical properties of this composite due to the presence of TiO₂NPs, as well as an enhanced adhesion and growth rate of hADSCs on the nTiO₂/MET-loaded NFs. The nanosurface of TiO₂NPs and the released metformin could result in a high differentiation capacity toward osteoblasts due to the promoted mineralization content and enhanced expression of osteoblastic markers. This work again demonstrated the high capacity of TiO₂NP-based scaffolds for bone tissue regeneration and engineering. The incorporation of TiO₂NPs in the polymer/drug matrix can enhance the cell-differentiating abilities of TiO₂NPs and prevents their aggregation.

As a soft tissue, skin has potential to repair itself so the key challenge for researchers and clinicians is to find ways to harness this skin regenerative potential to treat cutaneous injuries and diseases. At present, the only gold standard treatment is autologous skin graft for full-thickness skin wounds; however, this treatment also has shortcomings like restricted availability and morbidity of the donor site. Therefore, one of the alternative therapeutic options to treat the skin injuries is tissue-engineered skin (TES) which makes use of natural polymers including proteins which can resolve the autologous donor graft shortcomings and can provide protection against water-electrolyte imbalance and microbial infection as well.²⁷⁷ Silk fibroin (SF) is a protein (FDA approved), mechanically strong, biocompatible, biodegradable, and permeable material which has been used in a wide range of applications of tissue engineering and wound healing.²⁷⁷ Similarly, collagen (CG) is biocompatible and biodegradable protein-based natural polymer. These two biocompatible polymers can be applied as the matrix scaffold useful for the incorporation of TiO₂NPs for the tissue engineering studies. Khalid *et al.* designed silk fibroin/collagen (SF)/(CG) membranes combined with TiO₂NPs for skin tissue regeneration applications.²⁷⁷ The membranes exhibited good biocompatibility and antibacterial activity and can be introduced as potential candidates for skin tissue regeneration and wound healing applications.

The natural polymer of chitosan has been used extensively in the field of tissue engineering due to its biocompatibility, biodegradability, non-toxicity and antibacterial properties. Also, bredigite is one of the most well-known silicone biochemicals that has a high potential for the release of silicon ions, which results in the growth of osteoblasts and cellular differentiation. Ghasemi and Ghomi prepared the composite scaffolds of bredigite/TiO₂, followed by coating with the chitosan polymer to enhance the mechanical, biological, and antibacterial properties of the scaffold.²⁷⁸ The results showed that the addition of TiO₂ to the scaffold of bredigite resulted in reduced porosity and enhancement of the compressive strength of scaffolds from 0.299 to 0.687 MPa. In addition, the chitosan



coating reduced porosity from 83% to 63% and strongly improved the compressive strength from 0.585 to 2.339 MPa. The scaffolds showed antibacterial effects against *E. coli* (an inhibition zone of 22 mm) and *S. aureus* (an inhibition zone of 29 mm). Besides, these scaffolds exhibited no toxicity on the MG63 bone cells adjacent to the scaffolds. Regarding the modification effect of chitosan, the results showed that the presence of this modifying polymer could enhance the stability, antibacterial, and biocompatibility of the scaffold containing TiO₂NPs.

Polyurethane is a synthetic, thermoplastic, elastomeric, biodegradable, and biocompatible polymer having increasing applications in tissue engineering.³² However, the nanofibers made from polyurethane polymers have hydrophobic nature, which makes them undesirable when cultured in the presence of cells and/or implanted *in vivo* using animal models.^{33,34} These fibers are efficiently utilized as a bone mineral and modifying the matrix for the TiO₂NPs and other antibacterial NPs. This stabilizing matrix could enhance the biocompatibility of the used nanoparticles. Ashraf *et al.* fabricated nanofibers containing TiO₂NPs (as the osteoconductive component) and AgNP (as self-healing) nanostructures.²⁷⁹ The incubation of the nanofibers in the simulated body fluid at 37 °C triggered mineralization on nanofiber scaffolds and resulted in Ca and P crystals' formation. Also, the scaffolds showed antibacterial activity against *E. coli* (8.3 ± 0.9 mm) than *S. aureus* (1.2 ± 0.1 mm), and the MTT assay on the pre-osteoblasts demonstrated the biocompatibility of both TiO₂NPs and AgNPs with the bone-like cells. However, at higher contents of AgNPs (*i.e.*, 0.07 M), the scaffold showed cytotoxicity.

Recently, novel hybrid fillers such as TiO₂NPs/graphene oxide have been constantly emerging to be used in the structure of polymeric scaffolds to enhance the properties of composites. Since TiO₂NPs are antibacterial components, they have been widely used as nanofillers to enhance the performance of the composite scaffold. In this case, recently, Zhang *et al.* synthesized the reduced graphene oxide/TiO₂NP (RGO@TiO₂NPs) nanohybrid as a filler to investigate its synergistic effects on electrospun regenerated silk fibroin (RSF) mats (Fig. 7).²⁸⁰ The hybrid filler resulted in an increase in the average diameter of RSF fibers and decrease the content of β-sheet conformation. Interestingly, a 220% increase of the strength of the RSF/RGO@TiO₂NP composite mat was detected. Moreover, the

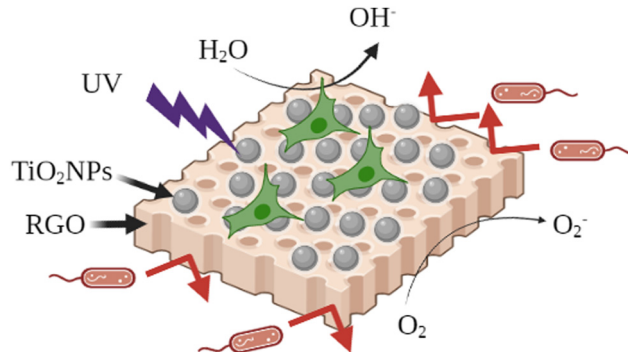


Fig. 7 The graphene oxide/titanium dioxide (RGO@TiO₂) nanohybrid as a filler to investigate their synergistic effects on electrospun regenerated silk fibroin (RSF) mats. Adapted with permission from ref. 280. Copyright 2021 Elsevier.

growth and expansion of the cells were reported on the composite mat due to the good antibacterial properties of TiO₂NPs. In this system, the RGO could bring a stabilizing matrix for the TiO₂NPs, protecting them against agglomeration which could result in well dispersion of TiO₂NPs in the scaffold and enhanced their antibacterial activity.

For bone tissue engineering, a scaffold with the optimized pore size and interconnected porosity is desirable to establish the tissue response and cell growth. It is essential for cell penetration and migration to effectively vascularize the growth of new tissues to have a maximum porosity (90%) and a suitable pore diameter (minimum 100 μm and maximum 450 μm). The TiO₂NPs play a significant role in controlling the pore size and porosity. For bone regeneration, a combination of TiO₂NPs with bioceramics (such as hydroxyapatite (HA)) has gained attention, due to the chemical similarity of HA to the mineral phase of natural bone and its extreme biocompatibility. HA can gain optimized microstructures for defected bone by combining with biodegradable polymers (including poly(glycolic acid), chitosan, arabinosylan, and guar gum). These biodegradable polymers also act as binders that reduce brittleness of HA. Aslam Khan *et al.* synthesized a TiO₂NP-based nanocomposite containing nano-hydroxyapatite (HA NPs), acrylic acid (AA)/guar gum (GG), and optimum graphene oxide (GO) to construct porous scaffolds coated with silver sulfadiazine (as a drug).²⁸¹ Their results showed that the TiO₂NPs and optimized

Table 11 Recent publications of TiO₂NPs for the tissue engineering applications

| Nanosystem | Applications | Ref. |
|---|--|------|
| Poly(lactate-co-glycolate) (PLGA)/TiO ₂ scaffolds | Scaffolds | 283 |
| TiO ₂ /poly(vinyl alcohol) nanocomposite | Bone tissue engineering | 284 |
| Silk fibroin/TiO ₂ nanocomposite | Bone tissue engineering | 285 |
| Nanostructured chitosan/PLA/HA scaffolds doped with TiO ₂ /Au/Pt NPs | Bone tissue engineering | 286 |
| TiO ₂ -chitosan/sodium alginate blended nanocomposite | Bone tissue engineering | 287 |
| Nanotubular TiO ₂ with gold nanoparticles | | 288 |
| Poly(lactic acid)/TiO ₂ nanocomposite | Potential ability of PLA/TiO ₂ nanocomposites to reduce cutaneous scarring in scaffolds | 289 |
| Alginate/chitosan multilayer films coated on IL-4-loaded TiO ₂ nanotubes | Alginate/chitosan multilayer films coated on IL-4-loaded TiO ₂ nanotubes for modulation of the macrophage phenotype | 290 |



GO improved the physicochemical and microstructural properties of this scaffold and the promising results obtained with mouse pre-osteoblast (MC3T3-E1) cell lines. Also, the organic part of this scaffold provided a stabilizing matrix for the TiO₂NPs, increasing their effectiveness.²⁸⁰

Poly(lactic acid) (PLA) is a well-known biocompatible and biodegradable polymer and it is a promising candidate to be used in the bone tissue engineering as the scaffold. It biodegrades to lactic acid without the aid of any enzymes which is innocuous to the body and avoids inflammatory responses. Modification of TiO₂NPs with this kind of biodegradable polymer provides a golden opportunity to prepare advanced bone scaffolds; for instance, Mota *et al.* incorporated TiO₂NPs inside the PLA matrix to prepare a modified nanohybrid for its potential ability in bone tissue engineering.²⁸² The cell viability tests on L929 fibroblast confirmed the biocompatibility of this TiO₂NP-PLA nanohybrid and its potential for use in the scaffolds. In this system, PLA could provide a biodegradable matrix for the well dispersion and fixation of TiO₂NPs against aggregation, enhancing the biocompatibility of the TiO₂NPs. Also, other recent articles in this field are summarized in Table 11.

5. Conclusion and perspectives

This review demonstrates the paramount importance of the surface modification of TiO₂NPs to improve their physicochemical properties and biocompatibility. In this regard, a broad range of organic and organosilane molecules were presented in this review paper which have been used as surface modifiers, according to the final applications of the modified TiO₂NPs. The potential biomedical applications of these modified TiO₂NPs have been studied in different fields, *e.g.*, photodynamic therapy, drug delivery, antibacterial, tissue engineering, anticancer, antifungals, biosensors, and antivirals. Furthermore, the anti-COVID-19 performance of modified TiO₂NPs has attracted growing attention of multidisciplinary groups to develop this field and improve current therapeutic methods. In these fields, surface modifiers were selected from biocompatible and bioactive materials which can improve the therapeutic effectiveness of the TiO₂NPs. It is worth mentioning that the clinical trials of the modified TiO₂NPs require much more research to fulfill all the requirements of clinical agents in terms of physicochemical and biological properties. For example, much more *in vivo* studies should be performed for evaluating the toxicity of modified TiO₂NPs on mammals. Thus, it is still a frontier research area with worthwhile background knowledge provided by previous research. So, future studies can be more focused on the development of safe, cost-effective, and scalable modified TiO₂NPs for the clinical fields, specifically antimicrobial agents. We believe that the modified TiO₂NPs will be an important research topic with great vitality and practical potential in biomedical applications.

Conflicts of interest

There are no conflicts to declare.

References

- N. Rodríguez-Barajas, M. L. Anaya-Esparza, Z. Villagrán-de la Mora, A. J. Sánchez-Burgos and A. Pérez-Larios, *Adv. Anticancer Agents Med. Chem.*, 2022, **22**, 2241–2254.
- X. He, H. Deng and H. Hwang, *J. Food Drug Anal.*, 2019, **27**, 1–21.
- L. M. Margarucci, V. Romano Spica, G. Gianfranceschi and F. Valeriani, *Environ. Int.*, 2019, **133**, 105095.
- A. Morlando, M. Chaki Borrás, Y. Rehman, S. Bakand, P. Barker, R. Sluyter and K. Konstantinov, *J. Mater. Chem. B*, 2020, **8**, 4016–4028.
- H. Chang, Q. Wang, X. Meng, X. Chen, Y. Deng, L. Li, Y. Yang, G. Song and H. Jia, *Chem. Res. Toxicol.*, 2022, **35**, 1435–1456.
- C. J. Dedman, A. M. King, J. A. Christie-Oleza and G.-L. Davies, *Environ. Sci.: Nano*, 2021, **8**, 1236–1255.
- S. Sargazi, S. ER, S. Sacide Gelen, A. Rahdar, M. Bilal, R. Arshad, N. Ajalli, M. Farhan Ali Khan and S. Pandey, *J. Drug Delivery Sci. Technol.*, 2022, **75**, 103605.
- R. Javed, N. Ul Ain, A. Gul, M. Arslan Ahmad, W. Guo, Q. Ao and S. Tian, *IET Nanobiotechnol.*, 2022, **16**, 171–189.
- F. Valeriani, L. M. Margarucci and V. Romano Spica, *Int. J. Environ. Res. Public Health*, 2018, **15**, 2675.
- G. Lofrano, F. Ubaldi, L. Albarano, M. Carotenuto, V. Vaiano, F. Valeriani, G. Libralato, G. Gianfranceschi, I. Fratoddi, S. Meric, M. Guida and V. Romano Spica, *Nanomaterials*, 2022, **12**, 2831.
- L. M. Margarucci, V. Romano Spica, C. Protano, G. Gianfranceschi, M. Giuliano, V. Di Onofrio, N. Mucci, F. Valeriani, M. Vitali and F. Romano, *Ann. Ig.*, 2019, **31**, 461–473.
- L. M. Margarucci, G. Gianfranceschi, V. Romano Spica, G. D'Ermo, C. Refi, M. Podico, M. Vitali, F. Romano and F. Valeriani, *Int. J. Environ. Res. Public Health*, 2021, **18**, 8662.
- T. Feng, S. Yan, S. Hou and X. Fan, *Spectrochim. Acta, Part A*, 2022, **280**, 121548.
- J. E. Karbowniczek, D. P. Ura and U. Stachewicz, *Composites, Part B*, 2022, **241**, 110011.
- G. Sanità, B. Carrese and A. Lamberti, *Front. Mol. Biosci.*, 2020, **7**, 587012.
- K. V. Korpany, C. Mottillo, J. Bachelder, S. N. Cross, P. Dong, S. Trudel, T. Frišćić and A. S. Blum, *Chem. Commun.*, 2016, **52**, 3054–3057.
- S. Benkoula, O. Sublemontier, M. Patanen, C. Nicolas, F. Sirotti, A. Naitabdi, F. Gaie-Levrel, E. Antonsson, D. Aureau, F.-X. Ouf, S.-I. Wada, A. Etcheberry, K. Ueda and C. Miron, *Sci. Rep.*, 2015, **5**, 15088.
- S. Wendt, R. Schaub, J. Matthiesen, E. K. Vestergaard, E. Wahlström, M. D. Rasmussen, P. Thostrup, L. M. Molina, E. Lægsgaard, I. Stensgaard, B. Hammer and F. Besenbacher, *Surf. Sci.*, 2005, **598**, 226–245.
- M.-I. Baraton and L. Merhari, *J. Eur. Ceram. Soc.*, 2004, **24**, 1399–1404.
- C. E. Nanayakkara, W. A. Larish and V. H. Grassian, *J. Phys. Chem. C*, 2014, **118**, 23011–23021.



- 21 L. T. Zhuravlev, *Colloids Surf., A*, 2000, **173**, 1–38.
- 22 P. Pallavicini, E. Cabrini, A. Casu, G. Dacarro, Y. Antonio Diaz-Fernandez, A. Falqui, C. Milanese and F. Vita, *Dalton Trans.*, 2015, **44**, 21088.
- 23 E. P. Plueddemann, *Nature of Adhesion Through Silane Coupling Agents, Silane Coupling Agents*, Springer US, Boston, MA, 1991, p. 115.
- 24 F. Ahangaran and A. H. Navarchian, *Adv. Colloid Interface Sci.*, 2020, **286**, 102298.
- 25 N. Zhu, H. Ji, P. Yu, J. Niu, M. U. Farooq, M. W. Akram, I. O. Udego, H. Li and X. Niu, *Nanomaterials*, 2018, **8**, 810.
- 26 R. S. Fernandes, I. M. Raimundo and M. F. Pimentel, *Colloids Surf., A*, 2019, 577, 1–7.
- 27 C. Chen, W. Wu, W. Z. Xu and P. A. Charpentier, *Nanotechnology*, 2017, **28**, 115709.
- 28 N. D. Bansod, B. P. Kapgate, C. Das, A. Das, D. Basu and S. C. Debnath, *J. Sol-Gel Sci. Technol.*, 2016, **80**, 548–559.
- 29 J. Xie, L. F. Mei, L. B. Liao, G. C. Lv, Z. G. Xia and G. X. Du, *Key Eng. Mater.*, 2014, **602–603**, 59–62.
- 30 F. Temerov, J. Haapanen, J. M. Mäkelä and J. J. Saarinen, *Inorganics*, 2021, **9**, 21.
- 31 H. Maskrot, N. Herlin-Boime, Y. Leconte, K. Jursikova, C. Reynaud and J. Vicens, *J. Nanopart. Res.*, 2006, **8**, 351–360.
- 32 G. N. Shao, Y. Kim, S. M. Imran, S. J. Jeon, P. B. Sarawade, A. Hilonga, J.-K. Kim and H. T. Kim, *Microporous Mesoporous Mater.*, 2013, **179**, 111–121.
- 33 Y. Zhang, F. Fang, C. Wang, L. Wang, X. Wang, X. Chu, J. Li, X. Fang, Z. Wei and X. Wang, *Polym. Compos.*, 2014, **35**, 1204–1211.
- 34 S. Mallakpour and M. Madani, *J. Mater. Sci.*, 2014, **49**, 5112–5118.
- 35 S. W. Chong, C. W. Lai, J. C. Juan and B. F. Leo, *Sol. Energy*, 2019, **191**, 663–671.
- 36 H. P. Duong, C.-H. Hung, H. C. Dao, M. D. Le and C.-Y. Chen, *New J. Chem.*, 2018, **42**, 8745–8751.
- 37 D. Meroni, L. Lo Presti, G. Liberto, M. Ceotto, R. G. Acres, K. C. Prince, R. Bellani, G. Soliveri and S. Ardizzone, *J. Phys. Chem. C*, 2017, **121**, 430–440.
- 38 S. Raqema, U. Hashim, N. Azizah, S. Nadzirah, M. K. M. Arshad, A. R. Ruslinda and S. C. B. Gopinath, *AIP Conf. Proc.*, 2017, **1808**, 20007.
- 39 Y.-L. Liu, Y.-H. Su and J.-Y. Lai, *Polymer*, 2004, **45**, 6831–6837.
- 40 Z.-M. Dang, Y.-J. Xia, J.-W. Zha, J.-K. Yuan and J. Bai, *Mater. Lett.*, 2011, **65**, 3430–3432.
- 41 Q. F. Xu, Y. Liu, F.-J. Lin, B. Mondal and A. M. Lyons, *ACS Appl. Mater. Interfaces*, 2013, **5**, 8915–8924.
- 42 M. M. Rahim-Abadi, A. R. Mahdavian, A. Gharieh and H. Salehi-Mobarakeh, *Prog. Org. Coat.*, 2015, **88**, 310–315.
- 43 P. Toh-Ae, B. Junhasavasdiikul, N. Lopattananon and K. Sahakaro, *Adv. Mater. Res.*, 2014, **844**, 276–279.
- 44 J. D. Ambrósio, C. V. M. Balarim and G. B. de Carvalho, *Polym. Compos.*, 2016, **37**, 1415–1424.
- 45 M. Sabzi, S. M. Mirabedini, J. Zohuriaan-Mehr and M. Atai, *Prog. Org. Coat.*, 2009, **65**, 222–228.
- 46 T. P. Selvin, J. Kuruvilla and T. Sabu, *Mater. Lett.*, 2004, **58**, 281–289.
- 47 L. Meng, Z. Liu, C. Lan and N. Xu, *Catal. Lett.*, 2022, **152**, 912–920.
- 48 P. A. Zapata, H. Palza, K. Delgado and F. M. Rabagliati, *J. Polym. Sci., Part A: Polym. Chem.*, 2012, **50**, 4055–4062.
- 49 V. G. Nguyen, H. Thai, D. H. Mai, H. T. Tran, D. L. Tran and M. T. Vu, *Composites, Part B*, 2013, **45**, 1192–1198.
- 50 E. Džunuzović, M. Marinović-Cincović, J. Vuković, K. Jeremić and J. M. Nedeljković, *Polym. Compos.*, 2009, **30**, 737–742.
- 51 Y. Qi, B. Xiang, W. Tan and J. Zhang, *Appl. Surf. Sci.*, 2017, **419**, 213–223.
- 52 J. Zhao, M. Milanova, M. M. C. G. Warmoeskerken and V. Dutschk, *Colloids Surf., A*, 2012, **413**, 273–279.
- 53 R. Mokhtari Aghdami, S. R. Mousavi, S. Estaji, R. K. Dermeni, H. A. Khonakdar and A. Shakeri, *Polym. Compos.*, 2022, **43**, 4165–4178.
- 54 H. Massoumi, R. Kumar, M. K. Chug, Y. Qian and E. J. Brisbois, *ACS Appl. Bio Mater.*, 2022, **5**, 2285–2295.
- 55 A. Dymerska, B. Zielińska, K. Sielicki, X. Chen and E. Mijowska, *Diamond Relat. Mater.*, 2022, **125**, 109027.
- 56 J. Yoo, H. Jeong, S. K. Park, S. Park and J. S. Lee, *Biosensors*, 2021, **11**, 2012.
- 57 P. Zhang, L. Cao, X. Wang, J. Cui, Z. Lin, S. Ngai, F. Vogel, H. Wang, W. Li, S. Li and Q. Wang, *Ceram. Int.*, 2022, **48**, 1731–1739.
- 58 A. Wanag, A. Sienkiewicz, P. Rokicka-Konieczna, E. Kusiak-Nejman and A. W. Morawski, *J. Environ. Chem. Eng.*, 2020, **8**, 103917.
- 59 S. Mallakpour and A. Barati, *Prog. Org. Coat.*, 2011, **71**, 391–398.
- 60 A. Shakeri, D. Yip, M. Badv, S. M. Imani, M. Sanjari and T. F. Didar, *Materials*, 2018, **11**, 1003.
- 61 R. Klaysri, T. Tubchareon and P. Praserttham, *J. Ind. Eng. Chem.*, 2017, **45**, 229–236.
- 62 G. Lee, J. Lee and C. Kang, *J. Coat. Technol. Res.*, 2019, **16**, 1399–1409.
- 63 M. A. Ashraf, Z. Liu, W.-X. Peng and N. Yoysefi, *Prog. Org. Coat.*, 2019, **136**, 105296.
- 64 N. Tangchantra, J. Krueenate, C. Aumnate and T. Sooksom-song, *Adv. Mater. Res.*, 2010, **93–94**, 300–303.
- 65 C. Yang and C. Yang, *J. Mater. Sci.: Mater. Electron.*, 2014, **25**, 3285–3289.
- 66 C. H. M. Caris, R. P. M. Kuijpers, A. M. van Herk and A. L. German, *Makromol. Chem., Macromol. Symp.*, 1990, **35–36**, 535–548.
- 67 C.-C. Weng and K.-H. Wei, *Chem. Mater.*, 2003, **15**, 2936–2941.
- 68 B. Erdem, E. D. Sudol, V. L. Dimonie and M. S. El-Aasser, *J. Polym. Sci., Part A: Polym. Chem.*, 2000, **38**, 4431–4440.
- 69 M. Z. Rong, M. Q. Zhang, H. B. Wang and H. M. Zeng, *Appl. Surf. Sci.*, 2002, **200**, 76–93.
- 70 M. Yang and Y. Dan, *Colloid Polym. Sci.*, 2005, **284**, 243–250.
- 71 F. Milanese, G. Cappelletti, R. Annunziata, C. L. Bianchi, D. Meroni and S. Ardizzone, *J. Phys. Chem. C*, 2010, **114**, 8287–8293.



- 72 J. Z. B. Xiang and G. Jiang, *Plast., Rubber Compos.*, 2015, **44**, 148–154.
- 73 C. Wang, H. Mao, C. Wang and S. Fu, *Ind. Eng. Chem. Res.*, 2011, **50**, 11930–11934.
- 74 J. Godnjavec, B. Znoj, J. Vince, M. Steinbacher, A. Žnidaršič and P. Venturini, *Mater. Technol.*, 2012, **46**, 19–24.
- 75 M.-A. E. Antoine, R. M. Dalod, L. Henriksen and T. Grande, *Beilstein J. Nanotechnol.*, 2017, **8**, 304–312.
- 76 M. Connolly, D. Hernández-Moreno, E. Conde, A. Garnica, J. M. Navas, F. Torrent, I. Rucandio and M. L. Fernandez-Cruz, *Environ. Sci. Eur.*, 2022, **34**, 1.
- 77 D. Tsotetsi, M. Dhlamini and P. Mbule, *Results Mater.*, 2022, **14**, 100266.
- 78 S. Katebi Koushali, M. Hamadian, A. R. Ghasemi and M. Ashrafi, *J. Nanostruct.*, 2021, **11**, 38–47.
- 79 B. A. Dehkordi, M. R. Nilforoushan, N. Talebian and M. Tayebi, *Mater. Res. Express*, 2021, **8**, 35403.
- 80 H. M. Yadav, N. D. Thorat, M. M. Yallapu, S. A. M. Tofail and J.-S. Kim, *J. Mater. Chem. B*, 2017, **5**, 1461–1470.
- 81 Z. Landolsi, I. Ben Assaker, D. Nunes, E. Fortunato, R. Martins, R. Chtourou and S. Ammar, *J. Mater. Sci.: Mater. Electron.*, 2020, **31**, 20753–20773.
- 82 S. Bai, N. Yang, X. Wang, F. Gong, Z. Dong, Y. Gong, Z. Liu and L. Cheng, *ACS Nano*, 2020, **14**, 15119–15130.
- 83 Y. Birinci, J. H. Niazi, O. Aktay-Çetin and H. Basaga, *Enzyme Microb. Technol.*, 2020, **138**, 109559.
- 84 X. Wang, X. Zhong, L. Bai, J. Xu, F. Gong, Z. Dong, Z. Yang, Z. Zeng, Z. Liu and L. Cheng, *J. Am. Chem. Soc.*, 2020, **142**, 6527–6537.
- 85 H. M. Yadav, N. D. Thorat, M. M. Yallapu, S. A. M. Tofail and J.-S. Kim, *J. Mater. Chem. B*, 2017, **5**, 1461–1470.
- 86 S. Salou, C.-M. Cirtiu, D. Larivière and N. Fleury, *Anal. Bioanal. Chem.*, 2020, **412**, 1469–1481.
- 87 Y. Li, Z. Qin, H. Guo, H. Yang, G. Zhang, S. Ji and T. Zeng, *PLoS One*, 2014, **9**, e114638.
- 88 V. A. Ortega, D. Boyle, J. W. Hodgkinson, D. B. D. Simmons, M. Belosevic, J. L. Stafford and G. G. Goss, *Environ. Sci.: Nano*, 2021, **8**, 1910–1926.
- 89 V. A. Ortega, M. S. Bahniuk, S. Memon, L. D. Unsworth, J. L. Stafford and G. G. Goss, *Biointerphases*, 2020, **15**, 51003.
- 90 H. E. Ali, A. M. Elbarbary, A. M. Abdel-Ghaffar and N. A. Maziad, *J. Appl. Polym. Sci.*, 2022, **139**, 52344.
- 91 Z. He, H. Wu, Z. Shi, Z. Kong, S. Ma, Y. Sun and X. Liu, *ACS Omega*, 2022, **7**, 7084–7095.
- 92 S. Dong, G. Xiao, C. Chen, Z. Yang, C. Chen, Q. Wang and L. Lin, *Prog. Org. Coat.*, 2021, **157**, 106291.
- 93 S. Naghibi, H. R. Madaah Hosseini and M. A. Faghihi Sani, *Ceram. Int.*, 2013, **39**, 8377–8384.
- 94 B. E. Castillo, E. Prokhorov, G. Luna-Bárcenas and Y. Kovalenko, *Polymers*, 2022, **14**, 1686.
- 95 H. Moulahoum, F. Ghorbanizamani, S. Sakarya and S. Timur, *Prog. Org. Coat.*, 2022, **169**, 106923.
- 96 N. Y. Elmehbad, N. A. Mohamed and N. A. Abd El-Ghany, *Int. J. Biol. Macromol.*, 2022, **205**, 719–730.
- 97 M. F. Majnis, O. C. Yee, M. A. Mohd Adnan, M. R. Yusof Hamid, K. Z. Ku Shaari and N. Muhd Julkapli, *Opt. Mater.*, 2022, **124**, 111967.
- 98 O. D. Saliu, M. Mamo, P. Ndungu and J. Ramontja, *J. Energy Storage*, 2022, **49**, 104155.
- 99 F. L. Gomes de Menezes, R. H. de Lima Leite, F. K. Gomes dos Santos, A. I. Aria and E. M. M. Aroucha, *Colloids Surf., A*, 2021, **630**, 127661.
- 100 M. Ren, H. Horn and F. H. Frimmel, *Water Res.*, 2017, **123**, 678–686.
- 101 F. Loosli, L. Vitorazi, J.-F. Berret and S. Stoll, *Water Res.*, 2015, **80**, 139–148.
- 102 M. N. Alomary and M. A. Ansari, *Chemistry*, 2021, **27**, 5817–5829.
- 103 G. Sarigul, I. Chamorro-Mena, N. Linares, J. García-Martínez and E. Serrano, *Adv. Sustainable Syst.*, 2021, **5**, 2100076.
- 104 S. Dessai, M. Ayyanar, S. Amalraj, P. Khanal, S. Vijayakumar, N. Gurav, N. Rarokar, M. Kalaskar, S. Nadaf and S. Gurav, *Mater. Lett.*, 2022, **311**, 131639.
- 105 S. Gulla, V. C. Reddy, P. B. Araveti, D. Lomada, A. Srivastava, M. C. Reddy and K. R. Reddy, *J. Mol. Struct.*, 2022, **1249**, 131556.
- 106 S. Iqbal, M. Fakhar-e-Alam, K. S. Alimgeer, M. Atif, A. Hanif, N. Yaqub, W. A. Farooq, S. Ahmad, Y.-M. Chu, M. Suleman Rana, A. Fatehmulla and H. Ahmad, *Saudi J. Biol. Sci.*, 2021, **28**, 1226–1232.
- 107 H.-X. Wang, X.-X. Li and L. Tang, *Appl. Phys. A: Mater. Sci. Process.*, 2020, **126**, 448.
- 108 A. Razzaz, S. Ghorban, L. Hosayni, M. Irani and M. Aliabadi, *J. Taiwan Inst. Chem. Eng.*, 2016, **58**, 333–343.
- 109 A. Babaei-Ghazvini, B. Acharya and D. R. Korber, *Polymers*, 2021, **13**, 2790.
- 110 V. K. Bui, D. Park and Y.-C. Lee, *Polymers*, 2017, **9**, 21.
- 111 R. Gobi, P. Ravichandiran, R. S. Babu and D. J. Yoo, *Polymers*, 2021, **13**, 1962.
- 112 X. Wang, Z. Li, Y. Wu, H. Guo, X. Zhang, Y. Yang, H. Mu and J. Duan, *ACS Appl. Mater. Interfaces*, 2021, **13**, 10902–10915.
- 113 R. Khan and M. Dhayal, *Electrochem. Commun.*, 2008, **10**, 492–495.
- 114 S. A. Matboo, S. Nazari, A. Niapour, M. V. Niri, E. Asgari and S. A. Mokhtari, *Water Sci. Technol.*, 2022, **85**, 605–616.
- 115 A. Maleki, B. Hayati, F. Najafi, F. Gharibi and S. W. Joo, *J. Mol. Liq.*, 2016, **224**, 95–104.
- 116 Z. Zhang, Y. Wang, T. Li, P. Ma, X. Zhang, B. Xia, M. Chen, M. Du and W. Dong, *Ind. Eng. Chem. Res.*, 2021, **60**, 3999–4008.
- 117 Z. Qiaorun, S. Honghong, L. Yao, J. Bing, X. Xiao, D. Julian McClements, C. Chongjiang and Y. Biao, *Food Res. Int.*, 2022, **159**, 111574.
- 118 A. Tajdari, A. Babaei, A. Goudarzi and R. Partovi, *J. Plast. Film Sheeting*, 2020, **36**, 285–311.
- 119 F. Loosli, P. Le Coustumer and S. Stoll, *Sci. Total Environ.*, 2015, **535**, 28–34.
- 120 S. Li, G. Chen, S. Qiang, Z. Yin, Z. Zhang and Y. Chen, *Int. J. Food Microbiol.*, 2020, **331**, 108763.
- 121 A. Deghiche, N. Haddaoui, A. Zerriouh, S. E. Fenni, D. Cavallo, A. Erto and Y. Benguerba, *J. Environ. Chem. Eng.*, 2021, **9**, 106541.



- 122 J. Zhang, M. Zheng, Y. Zhou, L. Yang, Y. Zhang, Z. Wu, G. Liu and J. Zheng, *Membranes*, 2022, **12**, 386.
- 123 J.-H. Li, Y.-Y. Xu, L.-P. Zhu, J.-H. Wang and C.-H. Du, *J. Membr. Sci.*, 2009, **326**, 659–666.
- 124 K. Liu, Z. Cai, X. Chi, B. Kang, S. Fu, X. Luo, Z.-W. Lin, H. Ai, J. Gao and H. Lin, *Nano Lett.*, 2022, **22**, 3219–3227.
- 125 J. L. Peper, N. E. Gentry, B. Boudy and J. M. Mayer, *Inorg. Chem.*, 2022, **61**, 767–777.
- 126 Y.-Z. Lü, S.-N. Zhang, Y.-F. Du, M.-T. Chen and C.-R. Li, *J. Inorg. Mater.*, 2013, **28**, 594–598.
- 127 Y. Lv, C. Li, Q. Sun, M. Huang, C. Li and B. Qi, *Nanoscale Res. Lett.*, 2016, **11**, 515.
- 128 M. S. Hanafy, W. M. Desoky, E. M. Hussein, N. H. El-Shaer, M. Gomaa, A. A. Gamal, M. A. Esawy and O. W. Guirguis, *J. Biomed. Mater. Res., Part A*, 2021, **109**, 232–247.
- 129 L. Goñi-Ciaurriz and I. Vélaz, *Int. J. Biol. Macromol.*, 2022, **216**, 347–360.
- 130 T. Ukmar, A. Godec, U. Maver, O. Planinšek, M. Bele, J. Jamnik and M. Gaberšček, *J. Mater. Chem.*, 2009, **19**, 8176–8183.
- 131 C. Ronchi, D. Selli, W. Pipornpong and C. Di Valentin, *J. Phys. Chem. C*, 2019, **123**, 7682–7695.
- 132 M. Qi, C. Li, Z. Song and L. Wang, *Drug Delivery*, 2021, **28**, 1785–1794.
- 133 R. Mohan, J. Drbohlavova and J. Hubalek, *Nanoscale Res. Lett.*, 2013, **8**, 503.
- 134 R. Binaymotlagh, L. Chronopoulou, F. Hajareh Haghghi, I. Fratoddi and C. Palocci, *Materials*, 2022, **15**, 5871.
- 135 R. Binaymotlagh, A. Del Giudice, S. Mignardi, F. Amato, A. G. Marrani, F. Sivori, I. Cavallo, E. G. Di Domenico, C. Palocci and L. Chronopoulou, *Gels*, 2022, **8**, 700.
- 136 X. Yang, Y. Wang, W. Qi, R. Xing, X. Yang, Q. Xing, R. Su and Z. He, *J. Mater. Chem. B*, 2019, **7**, 2981–2988.
- 137 E. Makhado, B. R. Motshabi, D. Allouss, K. E. Ramohlola, K. D. Modibane, M. J. Hato, R. M. Jugade, F. Shaik and S. Pandey, *Chemosphere*, 2022, **306**, 135524.
- 138 S. Zhao, C. Hou, L. Shao, W. An and W. Cui, *Appl. Surf. Sci.*, 2022, **590**, 153088.
- 139 R. R. Mansurov, V. S. Zverev and A. P. Safronov, *J. Catal.*, 2022, **406**, 9–18.
- 140 A. Ulu, E. Birhanlı, S. Köytepe and B. Ateş, *Int. J. Biol. Macromol.*, 2020, **163**, 529–540.
- 141 Y. Yue, X. Wang, Q. Wu, J. Han and J. Jiang, *J. Colloid Interface Sci.*, 2020, **564**, 99–112.
- 142 R. F. Bonan, M. F. Mota, R. M. da Costa Farias, S. D. da Silva, P. R. F. Bonan, L. Diesel, R. R. Menezes and D. E. da Cruz Perez, *Mater. Sci. Eng., C*, 2019, **104**, 109876.
- 143 S. Shiva Samhitha, G. Raghavendra, C. Quezada and P. Hima Bindu, *Mater. Today: Proc.*, 2022, **54**, 765–770.
- 144 N. Lagopati, K. Evangelou, P. Falaras, E.-P. C. Tsilibary, P. V. S. Vasileiou, S. Havaki, A. Angelopoulou, E. A. Pavlatou and V. G. Gorgoulis, *Pharmacol. Ther.*, 2021, **222**, 107795.
- 145 S. Çeşmeli and C. Biray Avci, *J. Drug Targeting*, 2019, **27**, 762–766.
- 146 Z. Youssef, V. Jouan-Hureau, L. Colombeau, P. Arnoux, A. Moussaron, F. Baros, J. Toufaily, T. Hamieh, T. Roques-Carmes and C. Frochot, *Photodiagn. Photodyn. Ther.*, 2018, **22**, 115–126.
- 147 Z. Youssef, P. Arnoux, L. Colombeau, J. Toufaily, T. Hamieh, C. Frochot and T. Roques-Carmes, *J. Photochem. Photobiol., A*, 2018, **356**, 177–192.
- 148 P. Huang, C. Xu, J. Lin, C. Wang, X. Wang, C. Zhang, X. Zhou, S. Guo and D. Cui, *Theranostics*, 2011, **1**, 240–250.
- 149 X. Liang, Y. Xie, J. Wu, J. Wang, M. Petković, M. Stepić, J. Zhao, J. Ma and L. Mi, *J. Photochem. Photobiol., B*, 2021, **215**, 112122.
- 150 B. Salama, C.-J. Chang, K. Kanehira, E.-S. El-Sherbini, G. El-Sayed, M. El-Adl and A. Taniguchi, *Molecules*, 2020, **25**, 4467.
- 151 S. Gai, G. Yang, P. Yang, F. He, J. Lin, D. Jin and B. Xing, *Nano Today*, 2018, **19**, 146–187.
- 152 M. A. Behnam, F. Emami, Z. Sobhani and A. R. Dehghanian, *Iran. J. Basic Med. Sci.*, 2018, **21**, 1133–1139.
- 153 Y. Gao, L. Zhang, Y. Liu, S. Sun, Z. Yin, L. Zhang, A. Li, G. Lu, A. Wu and L. Zeng, *Nanoscale*, 2020, **12**, 1801–1810.
- 154 T. Dai, W. He, S. Tu, J. Han, B. Yuan, C. Yao, W. Ren and A. Wu, *Bioact. Mater.*, 2022, **17**, 18–28.
- 155 Y. Zhang, X. Zhang, H. Yang, L. Yu, Y. Xu, A. Sharma, P. Yin, X. Li, J. S. Kim and Y. Sun, *Chem. Soc. Rev.*, 2021, **50**, 11227–11248.
- 156 R. L. Siegel, K. D. Miller, H. E. Fuchs and A. Jemal, *Ca-Cancer J. Clin.*, 2021, **71**, 7–33.
- 157 A. Vincent, J. Herman, R. Schulick, R. H. Hruban and M. Goggins, *Lancet*, 2011, **378**, 607–620.
- 158 A. Zinger, L. Koren, O. Adir, M. Poley, M. Alyan, Z. Yaari, N. Noor, N. Krinsky, A. Simon and H. Gibori, *ACS Nano*, 2019, **13**, 11008–11021.
- 159 K. P. Olive, M. A. Jacobetz, C. J. Davidson, A. Gopinathan, D. McIntyre, D. Honess, B. Madhu, M. A. Goldgraben, M. E. Caldwell and D. Allard, *Science*, 2009, **324**, 1457–1461.
- 160 A. Neesse, P. Michl, K. K. Frese, C. Feig, N. Cook, M. A. Jacobetz, M. P. Lolkema, M. Buchholz, K. P. Olive and T. M. Gress, *Gut*, 2011, **60**, 861–868.
- 161 J. Luo, J. Cao, G. Ma, X. Wang, Y. Sun, C. Zhang, Z. Shi, Y. Zeng, T. Zhang and P. Huang, *ACS Appl. Mater. Interfaces*, 2022, **14**, 40535–40545.
- 162 X. Wei, Z. Feng, J. Huang, X. Xiang, F. Du, C. He, M. Zhou, L. Ma, C. Cheng and L. Qiu, *ACS Appl. Mater. Interfaces*, 2021, **13**, 32810–32822.
- 163 X. Lin, R. Huang, Y. Huang, K. Wang, H. Li, Y. Bao, C. Wu, Y. Zhang, X. Tian and X. Wang, *Int. J. Nanomed.*, 2021, **16**, 1889–1899.
- 164 G. P. Lee, A. Willis, S. Pernal, A. Phakatkar, T. Shokuhfar, V. Blot and H. H. Engelhard, *Nanomedicine*, 2021, **16**, 523–534.
- 165 E. Yousefi, S. Javadpour, M. Ansari and H. Eslami, *Mater. Technol.*, 2021, **36**, 521–528.
- 166 A. Pariente, E. Peled, I. Zlotver and A. Sosnik, *Mater. Today Chem.*, 2021, **22**, 100613.
- 167 P. Magesan, K. I. Dhanalekshmi, J. Prabha, M. J. Umamathy, X. Zhang, N. Punitha, K. Kadambary and



- K. Sangeetha, *Photodiagn. Photodyn. Ther.*, 2022, **40**, 103064.
- 168 J.-Y. Zhou, W.-J. Wang, C.-Y. Zhang, Y.-Y. Ling, X.-J. Hong, Q. Su, W.-G. Li, Z.-W. Mao, B. Cheng, C.-P. Tan and T. Wu, *Biomaterials*, 2022, **289**, 121757.
- 169 E. Donadoni, P. Siani, G. Frigerio and C. Di Valentin, *Nanoscale*, 2022, **14**, 12099–12116.
- 170 L. Fang, H. Huang, J. D. Quirk, J. Zheng, D. Shen, B. Manion, M. Mixdorf, P. Karmakar, G. P. Sudlow, R. Tang and S. Achilefu, *Curr. Anal. Chem.*, 2022, **18**, 826–835.
- 171 Y. Hou, A. Mushtaq, Z. Tang, E. Dempsey, Y. Wu, Y. Lu, C. Tian, J. Farheen, X. Kong and M. Z. Iqbal, *J. Sci.: Adv. Mater. Devices*, 2022, **7**, 100417.
- 172 X. Wen, N. Liu, J. Ren, X. Jiao, J. Lv, M. H. Akhtar, H. Qi, J. Zhu, C. Yu and Y. Li, *New J. Chem.*, 2022, **46**, 6966–6970.
- 173 P. Ramachandran, B.-K. Khor, C. Y. Lee, R.-A. Doong, C. E. Oon, N. T. K. Thanh and H. L. Lee, *Biomedicines*, 2022, **10**, 421.
- 174 E. Tutun, V. Tekin, V. Yasakci, Ö. Aras and P. Ünak, *Appl. Organomet. Chem.*, 2021, **35**, e6435.
- 175 A. Mushtaq, Y. Hou, C. Tian, T. Deng, C. Xu, Z. Sun, X. Kong and M. Zubair Iqbal, *Mater. Res. Bull.*, 2021, **144**, 111481.
- 176 J. Li, S. Dai, R. Qin, C. Shi, J. Ming, X. Zeng, X. Wen, R. Zhuang, X. Chen, Z. Guo and X. Zhang, *ACS Appl. Mater. Interfaces*, 2021, **13**, 54727–54738.
- 177 M. Matijević, J. Žakula, L. Korićanac, M. Radoičić, X. Liang, L. Mi, J. F. Tričković, A. V. Šobot, M. N. Stanković, Đ. Nakarada, M. Mojović, M. Petković, M. Stepić and M. D. Nešić, *Photochem. Photobiol. Sci.*, 2021, **20**, 1087–1098.
- 178 J. Cao, Y. Sun, C. Zhang, X. Wang, Y. Zeng, T. Zhang and P. Huang, *Acta Biomater.*, 2021, **129**, 269–279.
- 179 Z. Shi, X. Meng, K. Zhang, S. Tang, C. Zhang, Z. Yang, H. Dong and X. Zhang, *ACS Mater. Lett.*, 2021, **3**, 781–789.
- 180 Q. Tang, H.-L. Zhang, Y. Wang, J. Liu and S.-P. Yang, *J. Mater. Chem. B*, 2021, **9**, 4241–4248.
- 181 Q. Pan, M. Li, M. Xiao, Y. He, G. Sun, T. Xue, Y. Luo, L. Chen, B. Ai and J. Xiong, *J. Nanomater.*, 2021, **2021**, 4125350.
- 182 A. Mansoor, Z. Khurshid, M. T. Khan, E. Mansoor, F. A. Butt, A. Jamal and P. J. Palma, *Nanomaterials*, 2022, **12**, 3670.
- 183 J. Han, E.-K. Jang, M.-R. Ki, R. G. Son, S. Kim, Y. Choe, S. P. Pack and S. Chung, *J. Ind. Eng. Chem.*, 2022, **112**, 258–270.
- 184 J. Cano-Mejia, R. A. Burga, E. E. Sweeney, J. P. Fisher, C. M. Bollard, A. D. Sandler, C. R. Y. Cruz and R. Fernandes, *Nanomedicine*, 2017, **13**, 771–781.
- 185 L. Chen, F. Pastorino, P. Berry, J. Bonner, C. Kirk, K. M. Wood, H. D. Thomas, Y. Zhao, A. Daga and G. J. Veal, *Int. J. Cancer*, 2019, **144**, 3146–3159.
- 186 S. G. DuBois, Y. P. Mosse, E. Fox, R. A. Kudgus, J. M. Reid, R. McGovern, S. Groshen, R. Bagatell, J. M. Maris and C. J. Twist, *Clin. Cancer Res.*, 2018, **24**, 6142–6149.
- 187 T. Lopez, J. Sotelo, J. Navarrete and J. A. Ascencio, *Opt. Mater.*, 2006, **29**, 88–94.
- 188 F. Grande and P. Tucci, *Mini-Rev. Med. Chem.*, 2016, **16**, 762–769.
- 189 M. Tian, X. Chen, Z. Gu, H. Li, L. Ma, X. Qi, H. Tan and C. You, *Carbohydr. Polym.*, 2016, **144**, 522–530.
- 190 J. Zhao, L. Yao, S. Nie and Y. Xu, *Int. J. Biol. Macromol.*, 2021, **167**, 921–933.
- 191 S. Klein, T. Luchs, A. Leng, L. V. R. Distel, W. Neuhuber and A. Hirsch, *Bioengineering*, 2020, **7**, 1–22.
- 192 K. Zheng, R. Chen, Y. Sun, Z. Tan, Y. Liu, X. Cheng, J. Leng, Z. Guo and P. Xu, *Thorac. Cancer*, 2020, **11**, 1476–1486.
- 193 S. Kim, S. Im, E.-Y. Park, J. Lee, C. Kim, T.-I. Kim and W. J. Kim, *Nanomedicine*, 2020, **24**, 102110.
- 194 T. Yu, L. Tong, Y. Ao, G. Zhang, Y. Liu and H. Zhang, *Drug Delivery*, 2020, **27**, 855–863.
- 195 W. Chen, J. Wang, L. Cheng, W. Du, J. Wang, W. Pan, S. Qiu, L. Song, X. Ma and Y. Hu, *ACS Appl. Bio Mater.*, 2021, **4**, 1483–1492.
- 196 Y. He, J. Wan, Y. Yang, P. Yuan, C. Yang, Z. Wang and L. Zhang, *Adv. Healthcare Mater.*, 2019, **8**, 1801254.
- 197 S. Bhullar, N. Goyal and S. Gupta, *Sci. Rep.*, 2022, **12**, 4600.
- 198 M. I. Torres-Ramos, M. F. Martín-Marquez, M. D. C. Leal-Moya, S. Ghotekar, J. A. Sánchez-Burgos and A. Pérez-Larios, *Int. J. Mol. Sci.*, 2022, **23**, 10755.
- 199 A. Chahardoli, F. Qalekhani, Y. Shokoohinia and A. Fattahi, *J. Mol. Liq.*, 2022, **361**, 119674.
- 200 N. Karki, H. Tiwari, M. Matiyani, R. Bal, M. Pal and N. G. Sahoo, *J. Vinyl Addit. Technol.*, 2022, **28**, 474–486.
- 201 A.-M. Negrescu, V. Mitran, W. Draghicescu, S. Popescu, C. Pirvu, I. Ionascu, T. Soare, S. Uzun, S. M. Croitoru and A. Cimpean, *J. Funct. Biomater.*, 2022, **13**, 43.
- 202 X. Zeng, W. Yang, F. X. Song, H. X. Wang and Y. Li, *J. Drug Delivery Sci. Technol.*, 2022, **68**, 103120.
- 203 M. Motiei Pour, M. R. Moghbeli, B. Larijani and H. Akbari Javar, *Chem. Pap.*, 2022, **76**, 439–451.
- 204 Y. Feng, L. Liu, J. Zhang, H. Aslan and M. Dong, *J. Mater. Chem. B*, 2017, **5**, 8631–8652.
- 205 E. J. Diana and T. V. Mathew, *Colloids Surf., B*, 2022, **220**, 112949.
- 206 P. Maheswari, S. Harish, M. Navaneethan, C. Muthamizhchelvan, S. Ponnusamy and Y. Hayakawa, *Mater. Sci. Eng., C*, 2020, **108**, 110457.
- 207 A. M. Mathew, V. I. Chukwuike, K. Venkatesan, S. Raveendran, R. C. Barik and D. K. Pattanayak, *Surf. Interfaces*, 2022, **33**, 102275.
- 208 A. Sathiyaseelan, K. Saravanakumar, K. V. Naveen, K.-S. Han, X. Zhang, M. S. Jeong and M.-H. Wang, *Environ. Res.*, 2022, **212**, 113237.
- 209 A. O. Özdemir, B. Caglar, O. Çubuk, F. Coldur, M. Kuzucu, E. K. Guner, B. Doğan, S. Caglar and K. V. Özdoğur, *Mater. Chem. Phys.*, 2022, **287**, 126342.
- 210 S. Metanawin and T. Metanawin, *Polym. Int.*, 2022, **71**, 777–789.
- 211 T. Singh, D. B. Pal, A. H. Almalki, Y. S. Althobaiti, M. F. Alkhanani, S. Haque, S. Sharma and N. Srivastava, *Mater. Lett.*, 2022, **316**, 132012.
- 212 S. Mallakpour and N. Mohammadi, *Carbohydr. Polym.*, 2022, **285**, 119226.



- 213 A. M. Youssef, M. E. Abd El-Aziz and S. M. M. Morsi, *Polym. Bull.*, 2022, **79**, 1–15.
- 214 Y. F. Makableh, N. F. Momani, T. Athamneh, R. Al-Abed and I. Alshorman, *Polym. Bull.*, 2022, **79**, 1–13.
- 215 P. Maheswari, S. Ponnusamy, S. Harish, C. Muthamizhchelvan and Y. Hayakawa, *Mater. Sci. Semicond. Process.*, 2020, **105**, 104724.
- 216 P. Maheswari, S. Ponnusamy, S. Harish, M. R. Ganesh and Y. Hayakawa, *Arabian J. Chem.*, 2020, **13**, 3484–3497.
- 217 P. Maheswari, S. Ponnusamy, S. Harish, C. Muthamizhchelvan, M. R. Ganesh and Y. Hayakawa, *Appl. Surf. Sci.*, 2019, **494**, 989–999.
- 218 S. Li, J. Zeng, D. Yin, P. Liao, S. Ding, P. Mao and Y. Liu, *Mater. Res. Express*, 2021, **8**, 85012.
- 219 A. M. Alakrach, A. A. Al-Rashdi, T. Alqadi, M. A. Al Saadi, S. S. Ting, O. S. Dahham and N. N. Zulkepli, *Mater. Sci. Forum*, 2021, **1021**, 270–279.
- 220 X. Wang, X. Li, X. Yang, K. Lei and L. Wang, *Colloids Surf., B*, 2021, **197**, 111410.
- 221 V. Soltaninejad and A. Maleki, *J. Photochem. Photobiol., A*, 2021, **404**, 112906.
- 222 T. Li, Y. Xiao, D. Guo, L. Shen, R. Li, Y. Jiao, Y. Xu and H. Lin, *J. Colloid Interface Sci.*, 2020, **572**, 114–121.
- 223 R. K. Manoharan, S. Ayyaru and Y.-H. Ahn, *New J. Chem.*, 2020, **44**, 807–816.
- 224 Y. Gao, X. Wang, X. Li and H. Dai, *New J. Chem.*, 2020, **44**, 20751–20758.
- 225 N. M. Ngoepe, M. M. Mathipa and N. C. Hintsho-Mbita, *Optik*, 2020, **224**, 165728.
- 226 S. Janfaza, M. Banan Nojavani, M. Nikkhah, T. Alizadeh, A. Esfandiar and M. R. Ganjali, *Microchim. Acta*, 2019, **186**, 137.
- 227 B. Mahyad, S. Janfaza and E. S. Hosseini, *Adv. Colloid Interface Sci.*, 2015, **225**, 194–202.
- 228 M.-C. Estevez, M. A. Otte, B. Sepulveda and L. M. Lechuga, *Anal. Chim. Acta*, 2014, **806**, 55–73.
- 229 Y. Xu, J. Lin, X. Wu, X. Xu, D. Zhang, Y. Xie, T. Pan, Y. He, A. Wu and G. Shao, *J. Mater. Chem. B*, 2022, **10**, 3808–3816.
- 230 Q. Y. Siew, S. Y. Tham, H.-S. Loh, P. S. Khiew, W. S. Chiu and M. T. T. Tan, *J. Mater. Chem. B*, 2018, **6**, 1195–1206.
- 231 S. Tao, Y. Guo, S. Wang, F. Xu, X. Zhou and Q. Guo, *Anal. Methods*, 2022, **14**, 2396–2404.
- 232 S. P. Hong, N. F. Mohd-Naim, N. A. Keasberry and M. U. Ahmed, *Electroanalysis*, 2022, **34**, 684–691.
- 233 S. Shi, Q. Nie, S. Jiang, S. Wu, B. Tang and M. Zhao, *Acta Opt. Sin.*, 2022, **42**, 0106001.
- 234 V. Rajeshwari, C. Vedhi and J. Fernando, *Mater. Today: Proc.*, 2022, **68**, 287–293.
- 235 D. Zheng, M. Chen, J. Peng, J. Chen, T. Chen, Y. Chen, L. Huang and W. Gao, *Microchim. Acta*, 2021, **188**, 328.
- 236 A. P. Singh, S. Balayan, S. Gupta, U. Jain, R. K. Sarin and N. Chauhan, *Process Biochem.*, 2021, **108**, 185–193.
- 237 J. Zhang, H. Hu and L. Yang, *Microchem. J.*, 2021, **168**, 106435.
- 238 M. Nycz, K. Arkusz and D. G. Pijanowska, *Materials*, 2021, **14**, 3767.
- 239 B. Baykal, G. Kadikoylu, H. Senturk, Y. O. Donar, A. Sinağ and A. Erdem, *J. Electroanal. Chem.*, 2021, **892**, 115262.
- 240 N. Gao, B. Fan, L. Li, X. Sun, X. Wang, H. Ma, Q. Wei and H. Ju, *ACS Appl. Bio Mater.*, 2021, **4**, 4479–4485.
- 241 J. Guo, G. Fang, S. Wang and J. Wang, *Food Chem.*, 2021, **344**, 128656.
- 242 R. H. Sakban, S. M. Abdulmohsin and M. D. Noori, *J. Phys. Conf. Ser.*, 2021, **1818**, 12038.
- 243 M. K. Choinńska, I. Šestáková, V. Hrdlička, J. Skopalová, J. Langmaier, V. Maier and T. Navrátil, *Biosensors*, 2022, **12**, 26.
- 244 J. Geddes-McAlister and R. S. Shapiro, *Ann. N. Y. Acad. Sci.*, 2019, **1435**, 57–78.
- 245 J. You, Y. Guo, R. Guo and X. Liu, *Chem. Eng. J.*, 2019, **373**, 624–641.
- 246 X. Zhao, G. Zhang and Z. Zhang, *Environ. Int.*, 2020, **136**, 105453; Y. Rilda, D. Dwiyantri, S. Syukri, A. Agustien and H. Pard, *J. Dispers. Sci. Technol.*, 2021, **42**, 784–790.
- 247 X. He, P. Wu, S. Wang, A. Wang, C. Wang and P. Ding, *J. Clean. Prod.*, 2021, **289**, 125755.
- 248 P. A. K. Reddy, P. V. L. Reddy, E. Kwon, K.-H. Kim, T. Akter and S. Kalagara, *Environ. Int.*, 2016, **91**, 94–103.
- 249 T. Wang, Z. Yang, C. Zhang, X. Zhai, X. Zhang, X. Huang, Z. Li, X. Zhang, X. Zou and J. Shi, *Int. J. Biol. Macromol.*, 2022, **222**, 2843–2854.
- 250 T. Siddiqui, N. J. Khan, N. Asif, I. Ahamad, D. Yasin and T. Fatma, *Environ. Sci. Pollut. Res.*, 2022, **29**, 39052–39066.
- 251 M. Sultan, H. Elsayed, A. E. F. Abdelhakim and G. Taha, *J. Appl. Polym. Sci.*, 2022, **139**, 51442.
- 252 L. Mohammad Taghizadeh Kashani, Shiva Masoudi and M. M. Ahmadian-Attari, *Inorg. Nano-Metal Chem.*, 2022, **52**, 297–307.
- 253 N. Duan, Q. Li, X. Meng, Z. Wang and S. Wu, *Food Chem.*, 2021, **364**, 130441.
- 254 H. Moradpoor, M. Safaei, A. Golshah, H. R. Mozaffari, R. Sharifi, M. M. Imani and M. S. Mobarakeh, *Inorg. Chem. Commun.*, 2021, **130**, 108748.
- 255 E. T. Helmy, E. M. Abouellef, U. A. Soliman and J. H. Pan, *Chemosphere*, 2021, **271**, 129524.
- 256 N. Rahmat, E. T. Wahyuni and A. Suratman, *Indones. J. Chem.*, 2021, **21**, 14–23.
- 257 H. P. Yetria Rilda, Dita Dwiyantri, Syukri Syukri and Anthoni Agustien, *J. Dispers. Sci. Technol.*, 2021, **42**, 784–790.
- 258 M. A. Irshad, R. Nawaz, M. Z. U. Rehman, M. Imran, J. Ahmad, S. Ahmad, A. Inam, A. Razzaq, M. Rizwan and S. Ali, *Chemosphere*, 2020, **258**, 127352.
- 259 L. Goñi-Ciaurriz, M. Senosiain-Nicolay and I. Vélaz, *Int. J. Mol. Sci.*, 2021, **22**, 2257.
- 260 R. Chougale, D. Kasai, S. Nayak, S. Masti, A. Nasalapure and A. V. Raghu, *Green Mater.*, 2020, **8**, 40–48.
- 261 S. Krishnan, A. Dusane, R. Morajkar, A. Venkat and A. A. Vernekar, *J. Mater. Chem. B*, 2021, **9**, 5967–5981.
- 262 M. A. Sadique, S. Yadav, P. Ranjan, S. Verma, S. T. Salammal, M. A. Khan, A. Kaushik and R. Khan, *J. Mater. Chem. B*, 2021, **9**, 4620–4642.



- 263 H. Liu, W. Zhong, X. Zhang, D. Lin and J. Wu, *J. Mater. Chem. B*, 2021, **9**, 7878–7908.
- 264 D. A. Elsayed, M. G. Assy, S. M. Mousa, G. T. El-Bassyouni, S. M. Mouneir and W. S. Shehab, *Bioorg. Chem.*, 2022, **124**, 105805.
- 265 D. C. S. Souza, S. M. Amorim, R. D. Cadamuro, G. Fongaro, R. A. Peralta, R. M. Peralta, G. L. Puma and R. F. P. M. Moreira, *Carbohydr. Polym. Technol. Appl.*, 2022, **3**, 100182.
- 266 D. J. da Silva, A. G. Souza, G. S. Ferreira, A. Duran, A. D. Cabral, F. L. A. Fonseca, R. F. Bueno and D. S. Rosa, *ACS Appl. Nano Mater.*, 2021, **4**, 12949–12956.
- 267 I.-J. Wang, Y.-C. Chen, C. Su, M.-H. Tsai, W.-T. Shen, C.-H. Bai and K.-P. Yu, *J. Aerosol Med. Pulm. Drug Delivery*, 2021, **34**, 293–302.
- 268 A. Levina, M. Repkova, N. Shikina, Z. Ismagilov, M. Kupryushkin, A. Pavlova, N. Mazurkova, D. Pyshnyi and V. Zarytova, *Eur. J. Pharm. Biopharm.*, 2021, **162**, 92–98.
- 269 G. León-Gutiérrez, C. Cabello-Gutiérrez, M. H. Martínez-Gómez, P. Azuara, B. Madden, J. Shalkow and A. Mejía, *J. Nano Res.*, 2021, **70**, 137–145.
- 270 L. Zhang, H. Forgham, A. Shen, J. Wang, J. Zhu, X. Huang, S.-Y. Tang, C. Xu, T. P. Davis and R. Qiao, *J. Mater. Chem. B*, 2022, **10**, 7473–7490.
- 271 Z.-Y. Chen, S. Gao, Y.-W. Zhang, R.-B. Zhou and F. Zhou, *J. Mater. Chem. B*, 2021, **9**, 2594–2612.
- 272 R. Balan and V. Gayathri, *Polym. Bull.*, 2022, **79**, 4269–4286.
- 273 W. S. Saeed, D. H. Alotaibi, A.-B. Al-Odayni, A. S. Haidyrah, A. A. Al-Owais, R. Khan, M. A. De Vera, A. Alrahlah and T. Aouak, *Int. J. Mol. Sci.*, 2022, **23**, 3449.
- 274 S. Mallakpour and M. Naghdi, *Ceram. Int.*, 2022, **48**, 2045–2057.
- 275 T. Fatima, R. Jolly, M. R. Wani, G. G. H. A. Shadab and M. Shakir, *Results Mater.*, 2021, **12**, 100240.
- 276 S. Ahmadi, Y. Pilehvar, N. Zarghami and A. Abri, *J. Drug Delivery Sci. Technol.*, 2021, **66**, 102798.
- 277 H. Khalid, H. Iqbal, R. Zeeshan, M. Nasir, F. Sharif, M. Akram, M. Irfan, F. A. Khan, A. A. Chaudhry and A. F. Khan, *Polym. Bull.*, 2021, **78**, 7199–7218.
- 278 S. Ghasemi and H. Ghomi, *J. Biomater. Appl.*, 2021, **36**, 406–418.
- 279 R. Ashraf, T. Maqbool, M. A. Beigh, A. H. Jadhav, H. S. Sofi and F. A. Sheikh, *J. Appl. Polym. Sci.*, 2021, **138**, 50594.
- 280 C. Zhang, X. Wang, A. Liu, C. Pan, H. Ding and W. Ye, *Mater. Lett.*, 2021, **291**, 129563.
- 281 M. U. Aslam Khan, W. S. Al-Arjan, M. S. Binkadem, H. Mehboob, A. Haider, M. A. Raza, S. I. Abd Razak, A. Hasan and R. Amin, *Nanomaterials*, 2021, **11**, 1319.
- 282 R. C. de Azevedo Gonçalves Mota, L. R. de Menezes and E. O. da Silva, *J. Mater. Res.*, 2021, **36**, 406–419.
- 283 S. S. Pelaseyed, H. R. Madaah Hosseini and A. Samadikuchak-saraei, *J. Biomed. Mater. Res., Part A*, 2020, **108**, 1390–1407.
- 284 N. A. Pattanashetti, C. Hiremath, S. R. Naik, G. B. Heggannavar and M. Y. Kariduraganavar, *New J. Chem.*, 2020, **44**, 2111–2121.
- 285 N. Johari, H. R. Madaah Hosseini and A. Samadikuchak-saraei, *Iran. Polym. J.*, 2020, **29**, 219–224.
- 286 J. Radwan-Pragłowska, Ł. Janus, M. Piątkowski, D. Bogdał and D. Matysek, *Polymers*, 2020, **12**, 792.
- 287 B. K. Shanmugam, S. Rangaraj, K. Subramani, S. Srinivasan, W. K. Aicher and R. Venkatachalam, *Mater. Sci. Eng., C*, 2020, **110**, 110710.
- 288 X. Zheng, J. Sun, W. Li, B. Dong, Y. Song, W. Xu, Y. Zhou and L. Wang, *J. Electroanal. Chem.*, 2020, **871**, 114362.
- 289 M. Kaseem, K. Hamad and Z. U. Rehman, *Materials*, 2019, **12**, 3659.
- 290 X. Yin, Y. Li, C. Yang, J. Weng, J. Wang, J. Zhou and B. Feng, *Int. J. Biol. Macromol.*, 2019, **132**, 495–505.

