RSC Advances



View Article Online

REVIEW

Check for updates

Cite this: RSC Adv., 2024, 14, 6557

Received 27th November 2023 Accepted 1st February 2024

DOI: 10.1039/d3ra08025k

rsc.li/rsc-advances

Introduction

Natural products play a crucial role in the exploration of new drugs as they possess broad-spectrum activity against bacteria, fungi, viruses, cancer, and other diseases, and they exhibit a vast array of chemically diverse structures, which hold the potential to serve as lead compounds in drug discovery. In particular, numerous compounds derived from natural

"Hainan Provincial Key Laboratory for Research and Development of Tropical Herbs, Key Laboratory of Tropical Translational Medicine of Ministry of Education, School of Pharmacy Hainan Medical University, No. 3, XueYuan Road, LongHua District, Haikou City, Hainan Province, 571199, China. E-mail: wang.shuojin@hainmc.edu.cn ^bDepartment of Thoracic Surgery, Shanghai Pulmonary Hospital, School of Medicine, Tongji University, Shanghai 200433, China. E-mail: xuyong@tongji.edu.cn

A review of typical biological activities of glycyrrhetinic acid and its derivatives

Liang Chen, 🗅 a Jingwen Gong, a Xu Yong, *b Youbin Lia and Shuojin Wang 🕒 *a

Glycyrrhetinic acid, a triterpenoid compound primarily sourced from licorice root, exhibits noteworthy biological attributes, including anti-inflammatory, anti-tumor, antibacterial, antiviral, and antioxidant effects. Despite these commendable effects, its further advancement and application, especially in clinical use, have been hindered by its limited druggability, including challenges such as low solubility and bioavailability. To enhance its biological activity and pharmaceutical efficacy, numerous research studies focus on the structural modification, associated biological activity data, and underlying mechanisms of glycyrrhetinic acid and its derivatives. This review endeavors to systematically compile and organize glycyrrhetinic acid derivatives that have demonstrated outstanding biological activities over the preceding decade, delineating their molecular structures, biological effects, underlying mechanisms, and future prospects for assisting researchers in finding and designing novel glycyrrhetinic acid derivatives, foster the exploration of structure-activity relationships, and aid in the screening of potential candidate compounds.

> products have already exhibited substantial therapeutic potential in the treatment of specific ailments.¹⁻⁵ Among these natural products, glycyrrhetinic acid is the triterpenoid aglycone constituent of glycyrrhizinic acid (Fig. 1), derived from the roots of the licorice plant (Glycyrrhiza glabra).6,7 There are two isomers of glycyrrhetinic acid (GA), one is (3β,18β)-3-hydroxy-11oxoolean-12-en-30-oic acid, often called 18β-glycyrrhetinic acid or enoxolone, denoted by 18 β -GA. Another one is (3 β ,18 α)-3hydroxy-11-oxoolean-12-en-29-oic acid, known as 18a-glycyrrhetinic acid, denoted by 18α-GA, as shown in Fig. 2. 18β-GA is the major bioactive constituent of Glycyrrhiza glabra and has been investigated to possess a wide range of biological activities, including anti-inflammatory, antitumor, antibacterial, antiviral, and antioxidant. Apart from these characteristic



Liang Chen

Liang Chen received his B.S. degree from Xuzhou Medical University. He is a graduate student at Hainan Medical University. He is a graduate student working in associate Professor Shuojin Wang's group at Hainan Medical University.



Jingwen Gong

Jingwen Gong received her

Master's degree from Southwest

University and is currently an

assistant researcher at Hainan

Medical University. She is interested in the pharmacology of

traditional Chinese medicine.



Fig. 1 Structure of glycyrrhizic acid.

activities, glycyrrhetinic acid has been observed to exhibit additional properties, such as anti-diabetic, anticoagulant, immunoregulatory, anti-cholinesterase, antiarrhythmic, and anti-tetanus toxin actions.⁸

However, 18 β -GA's poor druggability, including low solubility and bioavailability, limits its clinical use.⁹⁻¹² To improve the pharmacokinetic properties and enhance the bioactivity, various structural modifications of glycyrrhetinic acid have been carried out to develop novel derivatives for making them attractive candidates for further development as potential drug leads; in the process, extensive studies on the structure–activity relationship (SAR) of 18 β -GA and its derivatives have been extensively investigated.¹³ Furthermore, these modifications focused on altering the chemical structure, including the



Xu Yong

Yong Xu received his B.S. and PhD degrees from the Tongji University. He is a dedicated researcher with expertise in the application of tissue engineering technologies for cartilage regeneration and a commitment to advancing foundational research and clinical translation in tracheal functional reconstruction.



Fig. 2 Structure of glycyrrhetinic acid



Fig. 3 Modification of C-3 sites are labeled in pink, modification of C-2 sites are labeled in red, and modification of C-11 to C-13 sites modification are labeled in fluorescent green. The C-20 carboxyl sites are labeled in blue, while the other sites are labeled in fluorescent blue.

introduction of functional groups, changes in stereochemistry, and modifications of the aglycone skeleton. Studies on the pharmacological activities of 18β -GA derivatives have shown their potential as therapeutics for various diseases, such as inflammatory diseases, cancer, bacterial and viral infections, diabetes, and liver diseases, especially in the past two years.

The references incorporated in this review were exclusively sourced from the databases of Google Scholar, PubMed, and Web of Science. The compilation focusing on 18β -GA and its derivatives was based in works published within the temporal span of 2000 to 2023. Significantly, the majority of these citations were published within the most recent half-decade, highlighting the contemporaneity of our curated selection. In



Youbin Li

Youbin Li received his PhD degree from China Pharmaceutical University. He is a research fellow in the School of Pharmacy at the Hainan Medical University. His group is interested in the active ingredients and mechanism of action of natural medicines.

Shuojin Wang product medicinal chemistry.

Shuojin Wang received his B.S. and PhD degrees from Huazhong University of Science & Technology. He was senior researcher worked for WuXi AppTec, and was visiting scholar in Professor Weiping Tang's group at the University of Wisconsin-Madison. He is currently Associate Professor in the School of Pharтасу at Hainan Medical University. His group is interested in developing new synthetic methods and natural

addition, we meticulously scrutinized 266 compounds with significant biological activity from a pool of over 500 derivatives sourced from these cited references. To provide a more comprehensive and organized overview, we have compiled tables summarizing the chemical structures and effects or mechanisms of the typical biological activities of 18 β -GA and its derivatives, including anti-inflammatory, anti-tumor, antibacterial, antiviral and antioxidant effects. The labeling scheme for the modification sites of all 18 β -GA derivatives is described in the form of a diagram. Please refer to Fig. 3 for a visual representation of the labeling scheme.

Anti-inflammatory activity

Inflammation is considered to be a driver of many diseases, including arteriosclerosis, cancer, autoimmunity, and chronic infections.14 The inflammatory process involves multiple cell types, signaling pathways, and molecular mechanisms, leading to adverse reactions such as immunosuppression and gastrointestinal problems.15-21 Therefore, the design and optimization of drugs become more complicated. The presence of active ingredients in natural products opens up new opportunities for the development of anti-inflammatory drugs. Extensive research has shown that 18β-GA demonstrates antiinflammatory effects and holds significant potential as a therapeutic agent for various ailments.²² For instance, 18β-GA inhibits the expression of various inflammatory mediators, such as intercellular adhesion molecule-1 (ICAM-1), tumor necrosis factor-alpha (TNF-a), cyclooxygenase-2 (Cox-2), and inducible nitric oxide synthase (iNOS), by inhibiting the activity of the nuclear factor-κB (NF-κB) pathway.23 Additionally, 18β-GA has been found to reduce the production of inflammatory cytokines by inhibiting the activity of NF-kB and phosphoinositide 3-kinase (PI3K) and inhibiting the production of NO, prostaglandin E_2 (PGE₂), and reactive oxygen species (ROS) under lipopolysaccharide (LPS) stimulation.24 However, in an Ana-1 mouse macrophage model, 18β-GA induced the expression of Toll-like receptor 4 and activated the TLR-4 signaling pathway via the myeloid differentiation primary response 88 (MYD88) pathway.25

In recent years, the research of 18β -GA on anti-inflammation has been deepened. 18β -GA (40 mg kg⁻¹ day⁻¹) has been found to effectively improve lung function in ovalbumin (OVA)induced asthma mouse model, reduce lung inflammation and inflammatory cell infiltration, and inhibit the phosphorylation of NF-kB in the treatment of airway allergic inflammation. These effects are achieved through a decrease in the levels of interleukin-5 (IL-5) by approximately 40%, interleukin-13 (IL-13) by approximately 30%, and TNF- α by approximately 70%. Additionally, there is an increase in the levels of nuclear factor erythroid 2-related factor2 (Nrf2) by approximately 50% and heme oxygenase 1 (HO-1) by approximately 50%.26 Gupta et al. found that 18β-GA has potential therapeutic effects in treating depression. Specifically, it can improve symptoms caused by chronic unpredictable mild stress by activating the brainderived neurotrophic factor (BDNF)/Tropomyosin receptor kinase B (TrkB) signaling pathway in the prefrontal cortex (PFC)

and hippocampus. This activation leads to a reduction in neuroinflammation, liver biomarkers, and stress hormones while increasing the body weight and brain neurotransmitter concentrations.²⁷

Additionally, the complex of 18β-GA also exhibits remarkable anti-inflammatory activity. Ishida et al. demonstrated that the complex of 18β-GA and hydroxypropyl-β-cyclodextrin can mitigate indomethacin-induced small intestinal injury by reducing TNF-a expression by 27.5%, interleukin-6 (IL-6) by 16.2%, and interleukin-1ß (IL-1ß) by 17.9% compared to indomethacintreated tissue.28 The salt of 18β-GA and 1-arginine can be formed through a co-solvent evaporation reaction, and a solid dispersion called 18β-GA-SD can be created by adding a polymer solvent, Soluplus®, with a hydrophilic-hydrophobic chemical structure. 18B-GA-SD has higher solubility, cell utilization rate, and bioavailability than 18β-GA itself. Following treatment with 18B-GA-SD, enzyme-linked immunosorbent assay (ELISA) analysis revealed an increase in LPS-induced secretion levels of cytokines such as IL-1β, IL-6, macrophage inflammatory protein-1 (MCP-1), TNF-a, interleukin-23 (IL-23), and interleukin-17A (IL-17A) in RAW 264.7 cells; meanwhile, there was a decrease in the levels of interleukins-4 (IL-4) and -10 (IL-10).11

In the context of COVID-19, 18β-GA has been found to affect the disease by inhibiting the interleukin-17 (IL-17), IL-6, and TNF-α signaling pathways, thereby holding potential as a treatment strategy.²⁹ Another study found that a combination of 18β-GA and vitamin C (VC) treatment for COVID-19 was associated with an increase in immunity and a decrease in inflammatory stress, as well as activation of the T cell receptor signaling pathway, regulation of Fc gamma R-mediated phagocytosis, ErbB signaling pathway, and vascular endothelial growth factor signaling pathway.30 Furthermore, highly biocompatible 18β-GA nanoparticles have been synthesized and have shown promise as a treatment strategy for severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) infections.31 Zhou et al. demonstrated that 18β-GA inhibited the expression of intercellular adhesion molecule-1 (ICAM-1), TNF-a, COX-2, and iNOS, which was attributed to the inhibition of NF-KB expression and the attenuation of NF-kB nuclear translocation.32

Moreover, another study discovered that 18\alpha-GA suppressed the invasion on Matrigel-coated transwells of DU145 prostate cancer cells by regulating the expression of nu NF- κ B (p65), vascular endothelial growth factor (VEGF), and metalloproteinase-9 (MMP-9). 18a-GA also augmented the expression of non-steroidal anti-inflammatory gene-1 (NAG-1) in DU-145 cells, thereby indicating its capacity for antiinflammatory activity against prostate cancer cells.33 The mechanisms underlying the anti-inflammatory effects of GA discussed above are graphically depicted in Fig. 4. In the realm of hepatoprotective activity, 18β-GA has been shown to mitigate hepatic inflammatory injury caused by hepatitis virus infection by blocking the release of the high mobility group box 1 (HMGB1) cytokine and inhibiting its activity.34,35 Furthermore, 18β-GA has potential as a hepatoprotective agent through activating of Nuclear factor erythroid 2-related factor 2 (Nrf2) and peroxisome proliferator-activated receptor gamma (PPAR-y), and subsequent suppression of NF-KB, and 18β-GA has been



IL-7: interleukin-7, TNF-α: factor-alpha, ICAM-1: intercellular adhesion molecule-1, COX-2: cyclooxygenase-2, iNOS: nitric oxide synthase, NF-κB: nuclear factor-κB, IkB: IkappaB, IKK: IκB kinase, LPS: lipopolysaccharide, MyD88: myeloid differentiation primary response 88, TLR4: Toll-like receptor 4, TLR2: Toll-like receptor 2, BDNF: brain-derived neurotrophic facto, TrKB: Tropomyosin receptor kinase B, FXR: farnesoid X receptor, MAPK: mitogen-activated protein kinases, cAMP- REBP: cAMP-Responsive Element Binding Protein, IL-1β: interleukin-1β, IL-6: interleukin-6, mRNA: messenger RNA, IL-8: interleukin-8, HMGB1:

Fig. 4 Anti-inflammatory mechanisms of glycyrrhetinic acid and its derivatives.

shown to protect the liver from cholestatic liver injury induced by lithocholic acid (LCA) by inhibiting the TLR2/NF-κB pathway and upregulating hepatic farnesoid X receptor (FXR) expression, while reducing inflammation and promoting bile excretion. 18β-GA significantly increased the protein levels of the tubular bile acid (BA) efflux transporter bile salt export pump (BSEP) and the basolateral BA efflux transporters multidrug resistanceassociated proteins 3 and 4 (MRP3 and MRP4) but decreased the expression of the BA uptake transporter OATP2A1.^{23,36-39} Since the hepatic protection effect of 18β-GA is not only realized through the anti-inflammatory mechanism but could also through the antioxidant mechanism, the review about hepatic protection discussion is in the antioxidant part; Fig. 6 depicts all relevant studies.

In other investigations, various compounds derived from 18 β -GA, such as 1–15 (Table 1), have exhibited antiinflammatory effects. For instance, Ma *et al.* identified three major metabolites (compounds 1–3) produced by the microbial transformation of 18 β -GA. These metabolites exhibited potent anti-inflammatory activity by inhibiting LPS-induced NO production in mouse microglia BV2 cells.⁴⁰ The structure and inhibitory activity are shown in Table 1. Another investigation found that compound 4 showed improved pharmacokinetic properties and reduced toxicity in a similar way to fungal metabolism and LPS-induced mouse models.⁴¹ Li *et al.* found that compound 5 decreased the expression of iNOS, COX-2, and mitogen-activated protein kinases (MAPKs) as well as the activation of NF-KB in LPS-stimulated RAW 264.7 cells.42 More recently, Yang et al. investigated the anti-inflammatory effects of compound 6 on ear edema in mice and LPS-stimulated RAW 264.7 macrophages, respectively.43 Compound 6 was shown to decrease approximately 59.69% of 12-O-tetradecanoylphorbol-13-acetate (TPA)-induced ear edema with a gavage treatment of 40.0 mg mL $^{-1}$, and immunohistochemistry results revealed that this effect was related to the inhibition of TPA-induced upregulation of TNF-α. Compound 7 effectively inhibited the protein and mRNA expression of iNOS and the mRNA expression of TNF-α, IL-6, and IL-1β in LPS-stimulated RAW 264.7 macrophages. Bian et al. investigated the anti-inflammatory effects of compound 8 on LPS-induced RAW 264.7 cells and found that it suppressed the expression of pro-inflammatory cytokines including IL-6, TNF-a, and NO.44 Compounds 9-12 showed significant inhibition activity against NO and IL-6.45-47 Among these compounds, compound 12 was identified as the most potent anti-inflammatory agent, exhibiting a significant reduction in inflammatory cytokine levels in the mouse model of AKI by inhibiting TNF- α and IL-6 in a dose-dependent manner. Compound 13 also has anti-inflammatory activity, and studies have shown that it interacts with proteins in the inflammatory process, such as matrix metalloproteinase MMP9, neutrophil elastase, and thrombin.48 Tu et al. focus on the antiinflammatory activity of novel 18β-GA derivatives. The study evaluated the derivatives' activity in mouse models of acute inflammation induced by carrageenan. The results showed that

This article is licensed under a Creative Commons Attribution 3.0 Unported Licence.

(cc)) BY

Open Access Article. Published on 22 February 2024. Downloaded on 7/16/2025 11:59:10 PM.





This article is licensed under a Creative Commons Attribution 3.0 Unported Licence.

(cc) BY

Open Access Article. Published on 22 February 2024. Downloaded on 7/16/2025 11:59:10 PM.









several compounds demonstrated significant inhibition of paw edema and leukocyte infiltration.49 The results obtained from both in vitro and in vivo experiments indicate that compound 14 and compound 15 exhibit anti-inflammatory effects by reducing the expression of NO, pro-inflammatory cytokines, and chemokines, such as IL-1β, IL-6, IL-12, TNF-α, MCP-1, and macrophage inflammatory protein-1 alpha (MIP-1a) while increasing the expression of anti-inflammatory cytokine IL-10. Wang et al. introduced Soluplus®-glycyrrhetinic acid solid dispersion, which significantly improves the bioavailability and antiinflammatory activity of 18β-GA. The solubility of 18β-GA increased with the addition of Soluplus®, and the bioavailability was enhanced 2.61-fold. The anti-inflammatory activity of 18β-GA was also improved by 32.3%.11 Compounds 16-21 have been structurally modified at the C-2 and C-30 carboxyl positions of 18β-GA. These derivatives of 18β-GA have previously demonstrated outstanding anti-inflammatory activity, as seen in Table 1.50-52

In conclusion, 18β -GA has potential therapeutic applications for various conditions due to its anti-inflammatory effects. Although more research is required, the use of 18β -GA and its derivatives may provide new avenues for treating inflammationrelated diseases.

Antitumor activity

Cancer ravages and cripples the earth's inhabitants, ranking among the foremost destroyers of life.55 For countless years, scholars have been devoting themselves to the quest for a cure for tumors. Presently, the globe is awash with more than 80 conventional anti-tumor medications, ranging from cytotoxic drugs and hormones, to biological response modifiers (BRMs) and monoclonal antibodies.56 The majority of anticancer medications exhibit notable toxicity and necessitate administration in periodic cycles to mitigate adverse effects and impede the emergence of drug resistance. However, the excellent vitality of natural compounds adds new impetus to the research and development of anticancer drugs.57 And within this pantheon of treatment options stands the 18β-GA compound—a veritable powerhouse in its ability to vanquish cancerous cells from any part of the human body with unrivaled efficacy. Scores of meticulous studies attest to the fact that this drug is a gamechanger in the fight against various forms of cancer. The sterling performance against malignant cells has been proven time and time again, and it holds immense potential as an agent in the battle against cancer. Wang et al. demonstrated that 18β-GA has potent inhibitory effects on colorectal cancer cell proliferation in vitro and in vivo. This study showed that 18β-GA treatment resulted in a significant reduction in cell migration, invasion, and wound healing capability, accompanied by the downregulation of matrix metalloproteinase (MMP) expression. Moreover, 18β-GA decreased the protein levels of phosphorylated PI3K, protein kinase B (AKT), Signal Transducer and Activator of Transcription 3 (STAT3), c-Jun N-terminal Kinase (JNK), p38 mitogen-activated protein kinase (p38), and NF-κB p65, where the phosphorylation of PI3K and STAT3 decreased as early as 2 h after 18β-GA treatment.⁵⁸ Luo et al. found that 18βGA-induced apoptosis and G2/M cell cycle arrest and inhibited migration *via* the ROS/MAPK/STAT3/NF- κ B signaling pathways in A549 lung cancer cells. They also found that 18 β -GA could reduce tumor growth in a mouse xenograft model. In breast cancer treatment,⁵⁹ Shi *et al.* found that a combination of 18 β -GA and doxorubicin enhanced cytotoxicity, apoptosis, and loss of mitochondrial membrane potential *via* the upregulation of a mitochondrial-dependent apoptosis pathway against MCF-7 (breast adenocarcinoma cell line) cells.⁶⁰ In recent years, 18 β -GA has also been found to have potential in liver cancertargeted therapy. Speciale *et al.* provided a comprehensive review of the topic.⁶¹

The derivatives of 18β-GA have been unearthed to harbor even more potent cancer properties in comparison to the progenitor compound. One of the most remarkable advantages of 18β-GA lies in its all-encompassing efficacy in targeting a myriad of cancer types. It has conspicuously showcased outstanding effectiveness against cancers of the digestive tract, liver, nervous system, reproductive system, immune system, thyroid, and other organ-related cancers. This renders it an invaluable weapon in the war against cancer.^{62,63} The 18β-GA's anti-cancer effects are believed to stem from its capacity to incite apoptosis, a process of purposeful cell death, in cancer cells. Additionally, it also exhibits anti-inflammatory and antioxidant properties that can shield cells from harm and amplify the growth of healthy cells. As demonstrated in Table 2, we have amassed an extensive collection of 18β-GA derivatives with extraordinary anticancer activity.

In the realm of liver cancer treatment, researchers have discovered that 18β-GA holds significant potential due to its ability to exhibit toxicity against multiple liver cancer cell lines. A study conducted by Lai *et al.* found that 18β-GA derivatives **46–60** demonstrated selective cell toxicity against human hepatocellular carcinoma, hepG2 (hepatocellular carcinoma cell line) cells, and BEL-7402 (hepatocellular carcinoma cell line) cells, and BEL-7402 (hepatocellular carcinoma cell line) cells, and BEL-7402 (hepatocellular carcinoma cell line) cells, ⁶⁴ Similarly, derivatives **34**, **101–102**, **109–115**, **123–127**, and **147** displayed excellent cell toxicity against hepG2.^{65–72} Moreover, derivatives **73** and **74**, which were modified at position C30, exhibited noteworthy cell toxicity against SMMC-7721 (hepatocellular carcinoma cell line).^{73–76} Researchers also discovered the complex of 18β-GA-conjugated-β-cyclodextrin and emodin's superior cell toxicity against hep3B (hepatocellular carcinoma cell line) cells when compared to emodin alone.⁷⁷

In the domain of gastrointestinal cancers, encompassing those that affect the mouth, esophagus, colon, and stomach, the extraordinary cytotoxicity of 18β-GA and its derivatives has been strikingly demonstrated, particularly against colon cancer cell lines. The literature is replete with evidence of 18β-GA's potent effects on HCT-116 (colorectal carcinoma cell line), HCT-8 (colorectal adenocarcinoma cell line), DLD-1 (colorectal adenocarcinoma cell line), DLD-1 (colorectal adenocarcinoma cell line), and HT-29 (colorectal adenocarcinoma cell line) cells. For instance, derivatives **152–154** and **45** exhibit toxicity towards HCT-116, with derivative **45** also affecting HCT-8 cells and DLD-1. Likewise, derivatives **109–125** display remarkable cytotoxicity towards HT-29 cells.⁷⁸⁻⁸⁰ Moreover, Seribian *et al.*'s study unveiled the high cytotoxicity of 18β-GA 1,9-peroxide on numerous human tumor cell lines, Open Access Article. Published on 22 February 2024. Downloaded on 7/16/2025 11:59:10 PM.



Compounds	18β-GA	22-25	26-27	28-33
Structure	H H H H H H H H H H H H H H H H H H H	GH3 CH3	đ , , , , , , , , , , , , , , , , , , ,	
Effects or	но 218А2: IC ₅₀ = 83.92 µМ	$22: R = SO_2 CH_3$	26: R = I	28: R = OCH ₃
mechanisms	8505C: $IC_{50} = 86.50 \mu M$ A253: $IC_{50} = 80.78 \mu M$ A2780: $IC_{50} = 74.57 \mu M$ A471: $IC_{50} = 79.58 \mu M$	KU7: $IC_{50} = 3.3 \ \mu M$ Panc-1: $IC_{50} = 7.6 \ \mu M$ Panc-28: $IC_{50} = 9.7 \ \mu M$ 23: $R = I$	253JB-V: $C_{50} = 3.6 \mu M$ KU7I: $C_{50} = 2.6 \mu M$ Panc-1: $1C_{50} = 4.4 \mu M$ Panc-28: $1C_{50} = 3.6 \mu M$	253 JB-V: IC ₅₀ = 0.25 μ M KU7: IC ₅₀ = 1.59 μ M Panc-1: IC ₅₀ = 1.22 μ M Panc-28: IC ₅₀ = 1.80 μ M
	P_{22} : $P_{23} = a_{2.70} \mu m$ $DLD-1: IC_{50} = 81.21 \mu M$ $FDU: IC_{50} = 84.55 \mu M$ $HCT-8: IC_{50} = 78.85 \mu M$ $HT-29: IC_{50} = 80.09 \mu M$	2.33) $P_{V:I}$: $\Gamma_{S0} = 2.50 \mu M$ KU7: $\Gamma_{S0} = 3.0 \mu M$ Panc-1: $\Gamma_{S0} = 4.0 \mu M$ 2.4: $R = P=O(OCH_3)_2$ 2.53] $B_{V:I}$: $\Gamma_{S0} = 7.9 \mu M$	2.7: $N = Cr_3$ 253]B-V: $IC_{50} = 0.3 \ \mu M$ $KU7: IC_{50} = 1.3 \ \mu M$ Panc-1: $IC_{50} = 0.68 \ \mu M$ Panc-28: $IC_{50} = 1.1 \ \mu M$	253 R = Π 253 JB-V: $IC_{50} = 6.10 \ \mu M$ $KU7: IC_{50} = 5.88 \ \mu M$ Panc-1: $IC_{50} = 3.81 \ \mu M$ Panc-28: $IC_{50} = 7.32 \ \mu M$
	LIPO: $IC_{50} = 81.44 \ \mu M$ MCF-7: $IC_{50} = 84.70 \ \mu M$ SW480: $IC_{50} = 86.80 \ \mu M$ SW1736: $IC_{50} = 76.93 \ \mu M$ NIH 3T3: $IC_{50} = 18.52 \ \mu M$	KU7: $IC_{50} = 3.7 \mu M$ Panc-1: $IC_{50} = 6.1 \mu M$ Panc-28: $IC_{50} = 8.1 \mu M$ 25: $R = CF_3$ 253]B-V: $IC_{50} = 0.67 \mu M$		30: $R = piperidinyl$ HL-60: $IC_{50} = 1.4 \mu M$ 31: $R = 1,4$ -bipiperidinyl HL-60: $IC_{50} = 0.8 \mu M$ 32: $R = 4$ - methylpiperazinyl
Reference Compounds	HCT-11: $IC_{50} = 78.83 \mu M$ HCT-116: $IC_{50} = 78.83 \mu M$ 59-61 and 88 34	KU7: $IC_{50} = 0.38 \ \mu M$ Panc-1: $IC_{50} = 0.82 \ \mu M$ Panc-28: $IC_{50} = 1.1 \ \mu M$ 82 and 83 35-37	82 and 83 38-41	HL-60: $IC_{50} = 1.2 \ \mu M$ 33: $R = piperazinylHL-60: IC_{50} = 1.7 \ \mu M82 and 8342-43$
Structure	F O U U U U U U U U U U U U U U U U U U			NC N
Effects or mechanisms	но Хң 34: HepG-2: IC ₅₀ = 0.22 µМ	HO \overrightarrow{H} 3.5: R = piperidinyl $HL-60: IC_{50} = 5.5 \mu M$ $3.6: R = 1, 4^{-}bipiperidinyl$ $HL-60: IC_{50} = 3.3 \mu M$ 3.7: R = 4-methylpiperazinyl $HI-60: IC_{50} = 6.1 \mu M$	38: R = piperidinyl HL-60: $IC_{50} = 1.7 \mu M$ 39: R = 1,4'-bipiperidinyl HL-60: $IC_{50} = 7.7 \mu M$ 40: R = 4- methylpiperazinyl HL-60: $IC_{50} = 7.9 \mu M$	42: $R = piperidinyl$ HL-60: $IC_{50} = 8.6 \mu M$ 43: $R = 1,4'-bipiperidinyl$ HL-60: $IC_{50} = 7.5 \mu M$
			41: $R = piperazinyl HL-60: IC_{50} = 8.2 \mu M$	



Table 2 (Contd.)		
Reference Compounds	65 44	83 45
Structure		
Effects or mechanisms	44: R ₁ = O-i-Pr or OEt or OCH ₃ or OBn R ₂ = O-i-Pr or OEt or O-1-alanine or O-glycine 8505C: IC ₅₀ = $1.9-7.4 \mu$ M, A253: IC ₅₀ = $2.2-6.2 \mu$ M, A2780: IC ₅₀ = $1.3-5.9 \mu$ M A549: IC ₅₀ = $1.7-6.4 \mu$ M, DLD-1: IC ₅₀ = $2.5-8.5 \mu$ M, LIPO: IC ₅₀ = $2.3-7.5 \mu$ M	45: 518A2: $IC_{50} = 1.0 \ \mu\text{M}$, 8505C: $IC_{50} = 1.6 \ \mu\text{M}$, A253: IC_{50} $= 1.1 \ \mu\text{M}$ A2780: $IC_{50} = 1.3 \ \mu\text{M}$, A549: $IC_{50} = 1.5 \ \mu\text{M}$, DLD-1: IC_{50} $= 0.91 \ \mu\text{M}$ FADU: $IC_{50} = 1.7 \ \mu\text{M}$, HCT-116: $IC_{50} = 1.1 \ \mu\text{M}$, HCT-8: $IC_{50} = 0.6 \ \mu\text{M}$ HT-29: $IC_{50} = 0.5 \ \mu\text{M}$, LIPO: $IC_{50} = 1.5 \ \mu\text{M}$, MCF-7: IC_{50}
Reference Compounds	Average: IC ₅₀ = 2.3-7.0 μM 89 46-51 Q CH ₃	SW1736: IC ₅₀ = 1.6 μM, SW480: IC ₅₀ = 2.2 μM 80 5 2-60 solution solutita solutita solution solutita soluta
Structure	Development of the second seco	
Effects or mechanisms	46: $R = (CH_2)_2O$ BEL7402: $IC_{50} = 7.8 \mu M$ 47: $R = (CH_2)_3O$ BEL7402: $IC_{50} = 9.2 \mu M$ 48: $R = (CH_2)_2CH(CH_3)O$ BEL7402: $IC_{50} = 6.0 \mu M$ 49: $R = (CH_2)_4O$ BEL7402: $IC_{50} = 8.2 \mu M$ 50: $R = CH_2CH=CHCH_2O$ HepG2: $IC_{50} = 7.9 \mu M$, BEL7402: $IC_{50} = 7.3 \mu M$ 51: $R = CH_2CH_2NH$ HepG2: $IC_{50} = 2.9 \mu M$, BEL7402: $IC_{50} = 2.9 \mu M$	52: $R_1 = (CH_2)_2$, $R_2 = H$ HepG2: $IC_{50} = 9.0 \ \mu$ M, BEL7402: $IC_{50} = 1.3 \ \mu$ M 53: $R_1 = (CH_2)_3$, $R_2 = H$ HepG2: $IC_{50} = 3.7 \ \mu$ M, BEL7402: $IC_{50} = 0.43 \ \mu$ M 54: $R_1 = (CH_2)_2 CH(CH_3)$, $R_2 = H$ HepG2: $IC_{50} = 3.0 \ \mu$ M, BEL7402: $IC_{50} = 0.43 \ \mu$ M HepG2: $IC_{50} = 3.0 \ \mu$ M, BEL7402: $IC_{50} = 1.1 \ \mu$ M 56: $R_1 = (CH_2)_4$, $R_2 = H$ HepG2: $IC_{50} = 5.1 \ \mu$ M, BEL7402: $IC_{50} = 0.25 \ \mu$ M 56: $R_1 = (CH_2)_2 O(CH_2)_2$, $R_2 = H$ HepG2: $IC_{50} = 5.1 \ \mu$ M, BEL7402: $IC_{50} = 0.25 \ \mu$ M 56: $R_1 = (CH_2)_2 O(CH_3)_2$, $R_2 = H$ HepG2: $IC_{50} = 5.1 \ \mu$ M, BEL7402: $IC_{50} = 3.7 \ \mu$ M 58: $R_1 = CH_2CH=CHCH_3$, $R_2 = H$ 58: $R_1 = CH_2CH=CHCH_3$, $R_2 = H$

83

6566 | RSC Adv., 2024, 14, 6557-6597



MCF-7: $IC_{50} = 1.2 \ \mu M$, SW1736: $IC_{50} = 2.3 \ \mu M$



(Contd.)
Table 2

91 72 Rohit Hold Has	72: R = L-2,4-diaminobutanoyl or D-alanyl or sacrosyl or L- prolyl or L-phenylalanyl or L-methionyl or L-ornithyl or L-lysyl 8505C: $\Gamma_{50} = 2.4-9.6 \mu$ M, A253: $\Gamma_{50} = 2.2-7.4 \mu$ M, A2780: $\Gamma_{50} = 2.1-9.9 \mu$ M, DLD-1: $\Gamma_{50} = 1.4-8.7 \mu$ M, LIPO: $\Gamma_{50} = 0.8-7.9 \mu$ M MCF-7: $\Gamma_{50} = 0.8-7.9 \mu$ M MCF-7: $\Gamma_{50} = 2.2-6.0 \mu$ M
$\begin{array}{c} 00\\ 67-71\\ \end{array}$	67: $R_1 = CH_3$, $R_2 = H$ 518A2: $\Gamma C_{50} = 71.49 \ \mu M$, 8505C: $\Gamma C_{50} = 78.52 \ \mu M$, A2780: $\Gamma C_{50} = 62.78 \ \mu M$ A31: $\Gamma C_{50} = 86.13 \ \mu M$, A549: $\Gamma C_{50} = 79.13 \ \mu M$, DLD-1: $\Gamma C_{50} = 90.50 \ \mu M$ HTT-116: $\Gamma C_{50} = 90.30 \ \mu M$ HTT-29: $\Gamma C_{50} = 90.30 \ \mu M$ NIH 373: $\Gamma C_{50} = 90.30 \ \mu M$ SW1738: $\Gamma C_{50} = 72.47 \ \mu M$ NIH 373: $\Gamma C_{50} = 72.47 \ \mu M$ SW1738: $\Gamma C_{50} = 27.54 \ \mu M$, 8505C: $\Gamma C_{50} = 26.07 \ \mu M$, A2780: $\Gamma C_{50} = 27.54 \ \mu M$, 8505C: $\Gamma C_{50} = 24.36 \ \mu M$, A2780: $\Gamma C_{50} = 25.54 \ \mu M$, A549: $\Gamma C_{50} = 24.36 \ \mu M$, A2780: $\Gamma C_{50} = 25.54 \ \mu M$ A31: $\Gamma C_{50} = 25.74 \ \mu M$, 8505C: $\Gamma C_{50} = 24.36 \ \mu M$, A2780: $\Gamma C_{50} = 25.74 \ \mu M$, MCF-7: $\Gamma C_{50} = 24.36 \ \mu M$, A131: $\Gamma C_{50} = 22.10 \ \mu M$, HCT-8: $\Gamma C_{50} = 24.36 \ \mu M$, A131: $\Gamma C_{50} = 22.14 \ \mu M$, MCF-7: $\Gamma C_{50} = 24.36 \ \mu M$, A131: $\Gamma C_{50} = 22.31 \ \mu M$, MCF-7: $\Gamma C_{50} = 31.34 \ \mu M$, A131: $\Gamma C_{50} = 31.32 \ \mu M$, A549: $\Gamma C_{50} = 31.34 \ \mu M$, A131: $\Gamma C_{50} = 31.32 \ \mu M$, A549: $\Gamma C_{50} = 31.34 \ \mu M$, A131: $\Gamma C_{50} = 31.32 \ \mu M$, A549: $\Gamma C_{50} = 31.34 \ \mu M$, A131: $\Gamma C_{50} = 33.35 \ \mu M$, A549: $\Gamma C_{50} = 31.34 \ \mu M$, A131: $\Gamma C_{50} = 33.32 \ \mu M$, A549: $\Gamma C_{50} = 31.34 \ \mu M$, A131: $\Gamma C_{50} = 33.132 \ \mu M$, A549: $\Gamma C_{50} = 31.34 \ \mu M$, A131: $\Gamma C_{50} = 33.132 \ \mu M$, A549: $\Gamma C_{50} = 31.34 \ \mu M$, A131: $\Gamma C_{50} = 33.132 \ \mu M$, A549: $\Gamma C_{50} = 34.37 \ \mu M$, A131: $\Gamma C_{50} = 33.132 \ \mu M$, A549: $\Gamma C_{50} = 34.37 \ \mu M$, A131: $\Gamma C_{50} = 23.23 \ \mu M$, A549: $\Gamma C_{50} = 24.58 \ \mu M$, A131: $\Gamma C_{50} = 23.45 \ \mu M$, A549: $\Gamma C_{50} = 22.74 \ \mu M$, A131: $\Gamma C_{50} = 28.14 \ \mu M$,
Reference Compounds Structure	Effects or mechanisms

Table 2 (Contd.)



Table 2 (Contd.)





6572 | RSC Adv., 2024, 14, 6557-6597



(Contd. Table 2

HepG2: $IC_{50} = 2.439 \ \mu M$ MDCK: $IC_{50} = 4.645 \ \mu M$ MCF-7: $IC_{50} = 2.135 \ \mu M$ A549: $IC_{50}=2.109\ \mu M$ HeLa: $IC_{50} = 2.39 \ \mu M$ **123:** R = L-ala **124:** R = L-gly123-127 67 I HT-29: $IC_{50}=9.4~\mu\text{M}, A375; IC_{50}=7.1~\mu\text{M}, MCF7; IC_{50}$ HeLa: $IC_{50} = 2.6 \ \mu M$, A375: $IC_{50} = 2.3 \ \mu M$, MCF7: $IC_{50} =$ HepG2: $IC_{50} = 3.5 \ \mu\text{M}$, SH-SY5Y: $IC_{50} = 2.2 \ \mu\text{M}$, Jurkat: **114**: $R_1 = CH_2$, $R_2 = 1,2,3$ -triazolyl-4-methyl carboxylate HT-29: $IC_{50}=$ 8.9 $\mu M,$ A549: $IC_{50}=$ 7.9 $\mu M,$ MIAPaca2: HeLa: $IC_{50} = 5.4 \ \mu\text{M}$, A375: $IC_{50} = 4.9 \ \mu\text{M}$, MCF7: $IC_{50} =$ HepG2: $IC_{50} = 9.0 \ \mu\text{M}$, SH-SY5Y: $IC_{50} = 3.2 \ \mu\text{M}$, Jurkat: HepG2: $IC_{50} = 3.1 \mu M$, SH-SY5Y: $IC_{50} = 1.7 \mu M$, Jurkat: HT-29: $IC_{50} = 3.6 \mu M$, A549: $IC_{50} = 3.1 \mu M$, MIAPaca2: A375: $IC_{50}=7.2~\mu\text{M},$ MCF7: $IC_{50}=6.0~\mu\text{M},$ SH-SY5Y: HeLa: $IC_{50} = 2.2 \ \mu$ M, A375: $IC_{50} = 2.0 \ \mu$ M, MCF7: IC_{50} **119:** $R_1 = CO_2H$, $R_2 = C=0$, $R_3 = NHCH(CH_3)_2$ SH-SY5Y: IC₅₀ = 5.6 μ M, Jurkat: IC₅₀ = 2.4 μ M **113:** $R_1 = C=0$, $R_2 = 1,2,3$ -triazolyl-4-methyl **111:** $R_1 = C=0$, $R_2 = 2$ -methyl-1-imidazolyl **118:** $R_1 = CO_2H$, $R_2 = C=0$, $R_3 = NHC_6H_5$ **112:** $R_1 = CH_2$, $R_2 = 2$ -methyl-1-imidazolyl 117: $R_1 = CO_2 CH_3$, $R_2 = C=0$, $R_3 = OBn$ **116:** $R_1 = CO_2H$, $R_2 = C=0$, $R_3 = OBn$ NTUB1: $IC_{50} = 3.3 \ \mu M$ NTUB1: $IC_{50} = 2.3 \ \mu M$ NTUB1: $IC_{50} = 9.4 \ \mu M$ Jurkat: IC $_{50}=1.7\ \mu M$ BJ: $IC_{50} = 6.9 \ \mu M$ $IC_{50} = 3.3 \ \mu M$ $IC_{50}=1.3\ \mu M$ $IC_{50} = 3.7 \ \mu M$ $IC_{50} = 6.9 \ \mu M$ $IC_{50} = 1.5 \ \mu M$ $C_{50} = 1.1 \ \mu M$ carboxylate $= 5.6 \, \mu M$ 116-122 5.2 μM 3.0 µM 3.2 µM 99 mechanisms Compounds Reference Effects or Structure

128-143



130: $R_1 = NHCH_3$, $R_2 = Bn$ MCF-7: $IC_{50}=1.1~\mu M,$ PC-MCF-7: $IC_{50} = 3.8 \ \mu M$, PC-MCF-7: $IC_{50} = 1.1 \ \mu M$, PC-**129:** $R_1 = OCH_3$, $R_2 = Bn$ 3: $IC_{50} = 0.40 \ \mu M$ 131: $R_1 = NHEt, R_2 = Bn$ **128:** $R_1 = OH$, $R_2 = Bn$ $3{\rm : \ IC}_{50}=1.2\ \mu M$ 3: IC $_{50}=1.6\ \mu M$

Table 2 (Contd.)

120: $R_1 = CO_2CH_3$, $R_2 = H_2$, $R_3 = NHCH(CH_3)CO_2Me$ Jurkat: $IC_{50} = 9.6 \ \mu M$ 121: $R_1 = CO_2 \ CH_3$, $R_2 = CH_2$, $R_3 = NHCH(CH_3)$ F CO_2CH_3 Jurkat: $IC_{50} = 6.1 \ \mu M$ 122: $R_1 = CO_2 Et$, $R_2 = CH_2$, $R_3 = OEt$ 1 122: $R_1 = CO_2 Et$, $R_2 = C=O$, $R_3 = OEt$ 1 518A2: $IC_{50} = 9.2 \ \mu M$, A2780: $IC_{50} = 5.8 \ \mu M$ A 518A2: $IC_{50} = 9.2 \ \mu M$, A2780: $IC_{50} = 5.8 \ \mu M$ A 6 7	MCF-7: $IC_{50} = 2.853 \mu M$ HepG2: $IC_{50} = 3.472 \mu M$ HeLa: $IC_{50} = 3.01 \mu M$	132: $R_1 = NH$ -nPr, $R_2 = Bn$ MCF-7: $IG_{50} = 1.4 \mu M$, PC- $3 \cdot IC_{-0} - 0.46 \mu M$
: $CO_2 CH_3, R_2 = CH_2, R_3 = NHCH(CH_3)$ $I_{50} = 6.1 \mu M$: $CO_2Et, R_2 = C=O, R_3 = OEt$ $I_{50} = 9.2 \mu M, A2780: IC_{50} = 5.8 \mu M$	HeLa: $IC_{50} = 3.01 \ \mu M$	
₅₀ = 6.1 μM : CO ₂ Et, R ₂ = C=O, R ₃ = OEt ⁵⁰ = 9.2 μM, A2780: IC ₅₀ = 5.8 μM		$3.1050 - 0.10$ pure 133; $R_1 = pyrrolidinyl, R_2 = 0.00$
M s	MDCK: $IC_{50} = 3.749 \mu M$	MCF-7: $IC_{50} = 3.0 \mu M$, PC-
	125: $R = L-Boc-gly$	3: $10_{50} = 3.4 \mu M$ 134: $R_1 = morpholinyl, R_2 = 0.5$
	A549: $IC_{50} = 2.751 \ \mu M$	= BIL MCF-7: IC ₅₀ = 4.9 μ M, PC-
Υ Υ Υ	MCF-7: $IC_{50} = 3.811 \ \mu M$	3: $IC_{50} = 5.2 \ \mu M$ 135: $R_1 = 1,4$ -bipiperidinyl, $P_1 = P_2$
H V	HepG2: $IC_{50} = 3.306 \ \mu M$	$M_2 = DH$ MCF-7: $IC_{50} = 2.1 \mu M$, PC- 2: $IC = -2.0 \mu M$
4	HeLa: $IC_{50} = 3.296 \mu M$	3: $10_{50} = 3.0 \mu M$ 136: $R_1 = piperazinyl, R_2 = Rn$
	MDCK: $IC_{50} = 4.431 \ \mu M$	MCF-7: $IC_{50} = 3.1 \mu M$, PC- 2. IC $-2.7 \mu M$
1	126 : R = L-phe	$3.1050 - 2.7 \mu M$ 137: $R_1 = 1$ -
A	A549: $IC_{50} = 3.006 \ \mu M$	MCF-7: $IC_{50} = 3.3 \mu M$, PC-
4	MCF-7: $IC_{50} = 3.281 \mu M$	3: $\mathrm{IC}_{50}=3.1~\mu\mathrm{M}$ 138: $\mathrm{R_1}=1 ext{-Boc-}$
Ŧ	HepG2: $IC_{50} = 5.048 \mu M$	piperazinyl, $R_2 = Bn$ MCF-7: IC ₅₀ = 0.44 µM, PC-
		3: $IC_{50} = 0.23 \ \mu M$
F	HeLa: $IC_{50} = 3.296 \mu M$ MDCK: $IC_{50} = 5.024 \mu M$	139: $R_1 = anilinyl$, $R_2 = Bn$ MCF-7: $IC_{50} = 0.73 \mu$ M, PC-
-	107. D D	3: $IC_{50} = 0.45 \mu M$ 140. $D = -4. mitroomilinul$
-		$R_2 = Bn$
4	A549: $IC_{50} = 3.261 \ \mu M$	MCF-7: $IC_{50} = 5.8 \mu M$, PC- 3: $IC_{50} = 2.0 \mu M$
D	MCF-7: $IC_{50} = 7.623 \mu M$	141: $\mathbf{R}_1 = 4$ -chloroanilinyl,
4	HepG2: $\mathrm{IC}_{50}=2.143~\mu\mathrm{M}$	$M_{2} = DII$ MCF-7: $IC_{50} = 8.9 \mu M$, PC-
-		3: $IC_{50} = 0.85 \ \mu M$
H	HeLa: $IC_{50} = 2.209 \mu M$	142: $R_1 = 4$ - aminoperidinyl, $R_2 = Bn$
A	MDCK: $IC_{50} = 2.528 \ \mu M$	MCF-7: $IC_{50} = 0.98 \mu M$, PC-3: $IC_{50} = 0.69 \mu M$
		143: $K_1 = 1-BOC$ - piperazinyl, $R_2 = CH_3$ MCF-7· $IC_{22} = 1.0 \text{ mM} PC$ -
		$3: IC_{50} = 0.68 \mu M$



© 2024 The Author(s). Published by the Royal Society of Chemistry



66

99 151

Compounds

Reference

72 **150**





ss Article. Published on 22 February 2024. Downloaded on 7/16/2025 11:59:10 PM.	This article is licensed under a Creative Commons Attribution 3.0 Unported Licence.
Open Access A	(cc)) BY

Table 2 (Contd.)

Abbreviation

Reference

including HT-29 cells.⁸¹ Compounds **22–29** manifest substantial activity against Panc-1 (pancreatic carcinoma-1 cell line) and Panc-28 (pancreatic carcinoma-28 cell line) cells, and compounds **109–114** have been established as inhibitors of MIAPaca2 (pancreatic carcinoma cell line) cells.^{66,67,82,83} As for human oral epidermoid cancer cell lines, such as KB-3-1, KB-8-5, KB, and KB-VIN, compounds **85–90** and **148–150** have displayed their significant prowess.^{74,76,84–87}

In the context of prostate cancer cell lines such as PC-3 (androgen-independent) and LN-Cap, compounds **61–62**, **86–90**, and **128–143** have demonstrated significant inhibitory effects.^{75,76,97} In ovarian cancer cell lines like A2780, compounds **64–71** exhibited inhibitory activity up to 1.5 μ M.^{90,91} Notably, compounds **109–114**, **103**, **106,102**, **144**, and **146** displayed notable inhibitory activity against HeLa cells (cervical cancer cell line).^{70,71,81,98} Additionally, compounds **152–156** showed strong inhibitory activity against MCF-9 breast cancer cell line.^{79,101}

Beyond these realms, GA and its derivatives have also exhibited their anticancer activity in other areas. Prior research has established that GA and its derivatives have the ability to inhibit Neurosystem-associated cancer cell lines, such as SH-SY5Y (human neuroblastoma cell line) and SK-N-MC (human neuroblastoma cell line).^{66,84} In the investigation conducted by Csuk *et al.* conducted an investigation, which found that GA and its derivatives displayed robust activity against thyroid cancer.⁹¹ Li *et al.* found that 18β-GA exert anticancer effects as pin1 inhibitors.⁹⁵ Furthermore, GA and its derivatives have demonstrated significant inhibitory activity against various types of cancer cells including those associated with lung cancer, lymphoma, melanoma, and breast cancer.^{66-68,74-76,80,82,83,89,91-94,96}

In conclusion, 18 β -GA and its derivatives have shown promising anti-tumor properties in various types of cancer, including colorectal, breast, lung, and liver. The cytotoxic effects of 18 β -GA have been attributed to its ability to induce apoptosis, cell cycle arrest, inhibit migration, and downregulate various signaling pathways involved in cancer progression. In addition, 18 β -GA has been shown to enhance the cytotoxicity of conventional chemotherapeutic agents, making it a potential adjuvant therapy for cancer treatment. Although 18 β -GA and its derivatives have shown potential as anti-tumor agents, further studies are needed to fully understand their mechanisms of action and to optimize their pharmacological properties for clinical applications.

Antibacterial activity

The emergence and spread of drug-resistant bacteria pose a significant threat to global health. Conventional antibiotics are often rendered ineffective against these resistant strains, leading to prolonged and complicated treatment regimens, as well as increased morbidity and mortality rates. Consequently, there is a critical need to identify novel antibiotics that can effectively target and eliminate these drug-resistant bacteria.¹⁰² Researchers have turned their attention to natural compounds as potential sources of new antibiotics. Natural compounds have long been recognized for their diverse chemical structures



Table 3 Chemical structure and antibacterial activity of glycyrrhetinic acid and its derivatives 155–223

Compounds	18β-GA	157-163	164-166
	HO	o - Or	
Structure		ö	$\langle \langle \rangle$
Effects or mechanisms	Bacillus subtilis: MIC = 7.6 μ mL ⁻¹ Staphylococcus: MIC = 12.5 μ mL ⁻¹	157: $R = CH_2CH_3$ B. subtilis: MIC = 16.9 µg mL ⁻¹	164: R =
	A. actinomycetemcomitans: MIC = 8 $\mu g m L^{-1}$ E. corrodens: MIC = 16 $\mu g m L^{-1}$	S. scabies: MIC = 2.1 μ g mL ⁻¹ S. aureus: MIC = 4.2 μ g mL ⁻¹ MRSA: MIC = 4.0 μ g mL ⁻¹	Xoo: $EC_{50} = 2.28 \ \mu g \ mL^{-1}$ Xac: $EC_{50} = 1.42 \ \mu g \ mL^{-1}$ 165: $R =$
	C. sputigena: MIC = $8 \ \mu g \ mL^{-1}$	158 : $R = (CH_2)_2 CH_3$	
	Edwardsiella ictaluri: MIC > 470.7 μ g mL ⁻¹ H. pylori: MIC = 20.8 μ g mL ⁻¹	B. subtilis: MIC = >34.8 µg mL ⁻¹ S. scabies: MIC = 4.3 µg mL ⁻¹ S. aureus: MIC = 4.3 µg mL ⁻¹	Xoo: $EC_{50} = 3.57 \ \mu g \ mL^{-1}$ Xac: $EC_{50} = 0.93 \ \mu g \ mL^{-1}$ 166: $R =$
	P. aeruginosa: MIC = 160 $\mu g m L^{-1}$	MRSA: MIC = 2.0 $\mu g m L^{-1}$	- NH
	<i>P. gingivalis</i> ATCC 33277: MIC = 64 µg mL^{-1} <i>S. gordonii:</i> MIC = 64 µg mL^{-1} <i>N. gonorrhoeae:</i>	159: $R = (CH_2)_3 CH_3$ <i>B. subtilis:</i> MIC = >34.8 µg mL ⁻¹ <i>S. scabies:</i> MIC = 4.3 µg mL ⁻¹ <i>S. aureus:</i> MIC = 4.3 µg mL ⁻¹ MRSA: MIC	X00: $EC_{50} = 2.63 \ \mu g \ mL^{-1}$ Xac: $EC_{50} = 2.31 \ \mu g \ mL^{-1}$
	MIC = $3.9-62.5 \ \mu g \ m L^{-1}$	= 2.0 μ g mL ⁻¹ 160: R = CH ₃ <i>B. subtilis:</i> MIC = 4.0 μ g mL ⁻¹ <i>S. scabies:</i> MIC = 1.0 μ g mL ⁻¹ <i>S. aureus:</i> MIC = 2.0 μ g mL ⁻¹	
		161: $R = CH_2CH_3$ B. subtilis: MIC = 2.0 µg mL ⁻¹ S. scabies: MIC = 4.1 µg mL ⁻¹ S. aureus: MIC = 1.0 µg mL ⁻¹ MRSA: MIC = 1.0 µg mL ⁻¹ 162: $R = CH(CH_3)_2$ $P = CH(CH_3)_2$	
		S. scabies: MIC = -3.52 , pg mL ⁻¹ S. scabies: MIC = 4.2 µg mL ⁻¹ S. aureus: MIC = 2.0 µg mL ⁻¹ MRSA MIC = 2.0 µg mL ⁻¹ 163: R = $(CH_2)_3CH_3$ B. subtilis: MIC = >34.8 µg mL ⁻¹ S. scabies: MIC = $+3.3$ µg mL ⁻¹ S. arabies: MIC = -33.4 g µg mL ⁻¹ MUSA MIC = -22.0 µc mJ ⁻¹	



Table 3 (Contd.)

© 2024 The Author(s). Published by the Royal Society of Chemistry



НО

Staphylococcus epidermidis (ATCC 12228): MIC = $27.34 \text{ }\mu\text{g} \text{ }\text{mL}^{-1}$

Staphylococcus aureus (ATCC 29213): MIC = 54.68 μ g mL⁻¹

Staphylococcus aureus (ATCC 6538): MIC = $27.34 \text{ }\mu\text{g mL}^{-1}$



Table 3 (Contd.)		
Reference Compounds	43 173	123 174
Structure		NH NH NH NH NH NH NH NH NH NH NH NH NH N
Effects or mechanisms	173:Streptococcus pneumonia RCMB 010010:Diameter of inhibition zone = 15 mmStaphylococcus aureus ATCC25923:Diameter of inhibition zone = 15 mmMicrococcus luteus:Diameter of inhibition zone = 30 mmEscherichia coli ATCC25922:Diameter of inhibition zone = 20 mm	174: Streptococcus pneumonia RCMB 010010: Diameter of inhibition zone = 12 mm Staphylococcus aureus ATCC25923: Diameter of inhibition zone = 17 mm Micrococcus luteus: Diameter of inhibition zone = 30 mm Escherichia coli ATCC25922: Diameter of inhibition zone = 18 mm
Reference Compounds	Pseudomonas aeruginosa ATCC7853: Diameter of inhibition zone = 18 mm 79 175	Pseudomonas aeruginosa ATCC7853: Diameter of inhibition zone = 15 mm 79 176
Structure		
Effects or mechanisms	175:Streptococcus pneumonia RCMB 010010:Diameter of inhibition zone = 17 mmStaphylococcus aureus ATCC25923:Diameter of inhibition zone = 17 mmMicrococus luteus:Diameter of inhibition zone = 30 mmEscherichia coli ATCC25922:Diameter of inhibition zone = 16 mmPseudomonas aeruginosa ATCC7853:Diameter of inhibition zone = 15 mm	176: Streptococcus pneumonia RCMB 010010: Diameter of inhibition zone = 11 mm Staphylococcus aureus ATCC25923: Diameter of inhibition zone = 10 mm Micrococcus luteus: Diameter of inhibition zone = 29 mm Escherichia coli ATCC25922: Diameter of inhibition zone = 13 mm Pseudomonas aeruginosa ATCC7853: Diameter of inhibition zone = 13 mm

 Open Access Article. Published on 22 February 2024. Downloaded on 7/16/2025 11:59:10 PM.

 (cc) EX
 This article is licensed under a Creative Commons Attribution 3.0 Unported Licence.

Table 3 (Contd.)



Table 3 (Contd.)

	$\begin{split} R &= CH_2 CH_2 CH_3 \\ Theileria annulata (T339); GI_{50} &= 5.549 \ \mu mol \ L^{-1} \\ Theileria annulata (T3315); GI_{50} &= 3.55 \ \mu mol \ L^{-1} \\ 183: \\ R &= CH(CH_3)_2 \\ Theileria annulata (T339); GI_{50} &= 1.638 \ \mu mol \ L^{-1} \\ Theileria annulata (T5815); GI_{50} &= 1.499 \ \mu mol \ L^{-1} \\ 184: \\ R &= (CH_2)_3 CH_3 \\ Theileria annulata (T5815); GI_{50} &= 9.946 \ \mu mol \ L^{-1} \\ Theileria annulata (T5815); GI_{50} &= 9.946 \ \mu mol \ L^{-1} \\ Theileria annulata (T5815); GI_{50} &= 9.946 \ \mu mol \ L^{-1} \\ Theileria annulata (T5815); GI_{50} &= 9.946 \ \mu mol \ L^{-1} \\ Theileria annulata (T5815); GI_{50} &= 9.946 \ \mu mol \ L^{-1} \\ Theileria annulata (T5815); GI_{50} &= 9.946 \ \mu mol \ L^{-1} \\ Theileria annulata (T5815); GI_{50} &= 9.946 \ \mu mol \ L^{-1} \\ Theileria annulata (T5815); GI_{50} &= 9.946 \ \mu mol \ L^{-1} \\ Theileria annulata (T5815); GI_{50} &= 9.946 \ \mu mol \ L^{-1} \\ Theileria annulata (T5815); GI_{50} &= 9.946 \ \mu mol \ L^{-1} \\ Theileria annulata (T5815); GI_{50} &= 9.946 \ \mu mol \ L^{-1} \\ Theileria annulata (T5815); GI_{50} &= 9.946 \ \mu mol \ L^{-1} \\ Theileria annulata (T5815); GI_{50} &= 9.946 \ \mu mol \ L^{-1} \\ Theileria annulata (T5815); GI_{50} &= 9.946 \ \mu mol \ L^{-1} \\ Theileria annulata (T5815); GI_{50} &= 9.946 \ \mu mol \ L^{-1} \\ Theileria annulata (T5815); GI_{50} &= 9.946 \ \mu mol \ L^{-1} \\ Theileria annulata (T5815); GI_{50} &= 9.946 \ \mu mol \ L^{-1} \\ Theileria annulata (T5815); GI_{50} &= 9.946 \ \mu mol \ L^{-1} \\ Theileria annulata (T5815); GI_{50} &= 9.946 \ \mu mol \ L^{-1} \\ Theileria annulata (T5815); GI_{50} &= 9.946 \ \mu mol \ L^{-1} \\ Theileria annulata (T5815); GI_{50} &= 9.946 \ \mu mol \ L^{-1} \\ Theileria annulata (T5815); GI_{50} &= 9.946 \ \mu mol \ L^{-1} \\ Theileria annulata (T5815); GI_{50} &= 9.946 \ \mu mol \ L^{-1} \\ Theileria mol \ L$	
Reference Compounds	124 188-189	120 190-195
	HOHN	Ho
Structure	P P	
Effects or mechanisms	188: R =	190 : $R = 5$ -fluoro-2-nitro-benzene Staphylococcus aureus (ATCC 6538): MIC = 10 μ mol L ⁻¹
	X00: $EC_{50} = 10.2 \ \mu g \ m L^{-1}$	$Staphylococcus aureus (ATCC 29213): MIC = 10 \mumol L^{-1}$
	Xac: $\mathrm{EC}_{50} = 4.16 \ \mathrm{\mu g \ mL}^{-1}$	Staphylococcus epidermidis (ATCC 12228): MIC = $10 \text{ umol } \text{L}^{-1}$
	189: R =	MRSA: MIC = 16 μ mol L ⁻¹
		191: $\mathbf{R} = 4$ -chloro-2-nitro-benzene
	Xoo: $EC_{50} = 10.9 \ \mu g \ m L^{-1}$ Xac: $EC_{50} = 5.16 \ \mu g \ m L^{-1}$	Staphylococcus aureus (ATCC 6538): MIC = 10 μ mol L ⁻¹ Staphylococcus aureus (ATCC 29213): MIC = 5 μ mol L ⁻¹

μmol L⁻¹ MRSA: MIC = 16 μmol L⁻¹ **191**: R = 4-chloro-2-nitro-benzene *Staphylococcus aureus* (ATCC 6538): MIC = 10 μmol L⁻¹ *Staphylococcus aureus* (ATCC 29213): MIC = 5 μmol L⁻¹ *Staphylococcus epidermidis* (ATCC 12228): MIC = 5 μmol L⁻¹ MRSA: MIC = 8 μmol L⁻¹ **192**: R = 4-methoxy-2-nitro-benzene *Staphylococcus aureus* (ATCC 5538): MIC = 5 μmol L⁻¹ *Staphylococcus aureus* (ATCC 29213): MIC = 5 μmol L⁻¹ *Staphylococcus epidermidis* (ATCC 12228): MIC = 5 μmol L⁻¹ **192**: R = 4-methoxy-2-nitro-benzene *Staphylococcus epidermidis* (ATCC 12228): MIC = 5 μmol L⁻¹ *Staphylococcus aureus* (ATCC 5338): MIC = 5 μmol L⁻¹ **193**: R = 5-bromo-2-nitro-benzene *Staphylococcus aureus* (ATCC 29213): MIC = 2.5 μmol L⁻¹ *Staphylococcus aureus* (ATCC 29213): MIC = 2.5 μmol L⁻¹

Table 3 (Contd.)		
		Staphylococcus epidermidis (ATCC 12228):MIC = 2.5 µmol L ⁻¹ MRSA: MIC = 16 µmol L ⁻¹ 194: R = 4-bromo-2-nitro-benzene Staphylococcus aureus (ATCC 6538): MIC = 12.5 µmol L ⁻¹
		Staphylococcus aureus (ATCC 29213):MIC = 12.5 μ mol L ⁻¹ L ⁻¹ Staphylococcus epidermidis (ATCC 12228):MIC = 12.5 μ mol L ⁻¹ MRSA: MIC = 16 μ mol L ⁻¹
		195: R = 4-fluoro-2-nitro-benzene Staphylococcus aureus (ATCC 6538): MIC = 5 μ mol L ⁻¹ Staphylococcus aureus (ATCC 29213): MIC = 5 μ mol L ⁻¹ Staphylococcus epidermidis (ATCC 12228): MIC = 5 μ mol L ⁻¹
Reference Compounds	120 196-201	MRSA: MIC = 8 μ mol L ⁻¹ 49 202-225
		R R R R R R R R R R R R R R R R R R R
		H Contraction of the second se
Structure	T T T T T T T T T T T T T T T T T T T	HO HO HO
		Z=
Effects or mechanisms	196:	202:
	n = 5 Xoo: $EC_{50} = 8.57 \ \mu g \ mL^{-1}$, Xac: $EC_{50} = 7.67 \ \mu g \ mL^{-1}$ 195 : n = 6 Xoo: $EC_{50} = 5.24 \ \mu g \ mL^{-1}$, Xac: $EC_{50} = 9.55 \ \mu g \ mL^{-1}$ 197 : n = 7 Xoo: $EC_{50} = 5.06 \ \mu g \ mL^{-1}$, Xac: $EC_{50} = 8.16 \ \mu g \ mL^{-1}$	$\begin{split} n &= 5; \ R = Br^-\\ \text{Xoo: } EC_{50} &= 9.47 \ \mu\text{g mL}^{-1}, \ \text{Xac: } EC_{50} &= 11.8 \ \mu\text{g mL}^{-1}\\ \textbf{203:}\\ n &= 6; \ R = Br^-\\ \text{Xoo: } EC_{50} &= 9.18 \ \mu\text{g mL}^{-1}, \ \text{Xac: } EC_{50} &= 34.5 \ \mu\text{g mL}^{-1}\\ \text{Xoo: } EC_{50} &= 9.18 \ \mu\text{g mL}^{-1}, \ \text{Xac: } EC_{50} &= 34.5 \ \mu\text{g mL}^{-1}\\ \textbf{Xoo: } EC_{50} &= 7.12 \ \mu\text{g mL}^{-1}, \ \text{Xac: } EC_{50} &= 9.53 \ \mu\text{g mL}^{-1}\\ \text{Xoo: } EC_{50} &= 7.12 \ \mu\text{g mL}^{-1}, \ \text{Xac: } EC_{50} &= 9.53 \ \mu\text{g mL}^{-1} \end{split}$
	198:	205:

Open Access Article. Published on 22 February 2024. Downloaded on 7/16/2025 11:59:10 PM.
This article is licensed under a Creative Commons Attribution 3.0 Unported Licence.

(cc) BY

Table 3 (Contd.)		
n = 8 Xoo: EC 199 :	n = 8 Xoo: $EC_{50} = 3.54 \ \mu g \ m L^{-1}$, Xac: $EC_{50} = 10.3 \ \mu g \ m L^{-1}$ 199:	$n = 8$; $R = Br^{-}$ Xoo: $EC_{50} = 3.38 \ \mu g \ mL^{-1}$, Xac: $EC_{50} = 18.7 \ \mu g \ mL^{-1}$ 206:
n = 9 Xoo: EC 200:	M = 9 Xoo: EC ₅₀ = 3.47 µg mL ⁻¹ , Xac: EC ₅₀ = 34.1 µg mL ⁻¹ 200.	n = 9; R = Br ⁻ Xoo: EC ₅₀ = 2.29 µg mL ⁻¹ , Xac: EC ₅₀ = 25.6 µg mL ⁻¹ 207:
n = 10Xoo: EC	n = 10 Xoo: EC ₅₀ = 6.60 µg mL ⁻¹ , Xac: EC ₅₀ = 17.4 µg mL ⁻¹	$n = 10$; $R = Br^{-1}$ Xoo: $EC_{50} = 1.37 \ \mu g \ m L^{-1}$, Xac: $EC_{50} = 37.4 \ \mu g \ m L^{-1}$
		208: n = 5; $R = XXoo: EC_{50} = 14.08 \ \mu g \ m L^{-1}, Xac: EC_{50} = 14.76 \ \mu g \ m L^{-1}200-$
		n = 5; $R = YXoo: EC_{50} = 19.53 \ \mu g \ mL^{-1}, Xac: EC_{50} = 6.8 \ \mu g \ mL^{-1}210:$
		n = 5; $R = ZXoo: EC_{50} = 19.06 \ \mu g \ mL^{-1}, Xac: EC_{50} = 4.59 \ \mu g \ mL^{-1}211:$
		n = 6; R = X Xoo: EC ₅₀ = 12.11 µg mL ⁻¹ , Xac: EC ₅₀ = 6.88 µg mL ⁻¹ 212:
		n = 6; R = Y Xoo: EC ₅₀ = 12.9 µg mL ⁻¹ , Xac: EC ₅₀ = 25.03 µg mL ⁻¹ 213:
		n = 6; $R = ZXoo: EG_{50} = 20.59 \ \mu g \ mL^{-1}, Xac: EG_{50} = 14.81 \ \mu g \ mL^{-1}$
		n = 7; $R = XXoo: EC_{50} = 6.5 \ \mu g \ mL^{-1}, Xac: EC_{50} = 14.81 \ \mu g \ mL^{-1}$
		2.13: n = 7; $R = YXoo: EG_{50} = 6.17 \ \mu g \ mL^{-1}, Xac: EG_{50} = 11.69 \ \mu g \ mL^{-1}2.16:$
		n = 7; R = Z Xoo: EC ₅₀ = 17.25 µg mL ⁻¹ , Xac: EC ₅₀ = 14.39 µg mL ⁻¹ 217.
		n = 8; $R = XXoo: EC_{50} = 5.17 \mu g m L^{-1}, Xac: EC_{50} = 7.16 \mu g m L^{-1}218:$
		n = 9; R = X Xoo: EC ₅₀ = 4.18 µg mL ⁻¹ , Xac: EC ₅₀ = 10.32 µg mL ⁻¹
		219: n = 10; $R = XXoo: EC_{50} = 1.6 \ \mu g \ mL^{-1}, Xac: EC_{50} = 8.48 \ \mu g \ mL^{-1}220.$
		n = 8; R = Y Xoo: EC ₅₀ = 4.93 µg mL ⁻¹ , Xac: EC ₅₀ = 3.82 µg mL ⁻¹ 221:
		$n = 9$; $\mathbf{R} = \mathbf{Y}$

Open Access Article. Published on 22 February 2024. Downloaded on 7/16/2025 11:59:10 PM.
This article is licensed under a Creative Commons Attribution 3.0 Unported Licence.

(cc) BY

Review

Table 3 (Contd.)		
	Xoo: $EC_{50} = 7.56 \ \mu g \ mL^{-1}$, Xac: $EC_{50} = 4.38 \ \mu g \ mL^{-1}$ 222: 222: 222: 222: 222: 222: 222: 222	8 µg mL ⁻¹
	n = 10; $R = TX00: EC_{50} = 4.14 \ \mu g \ mL^{-1}, Xac: EC_{50} = 10.15 \ \mu g \ mL^{-1}223:$	15 $\mu g \ m L^{-1}$
	n = 8; $R = ZXoo: EC_{50} = 13.77 \ \mu g \ m L^{-1}, Xac: EC_{50} = 22.17 \ \mu g \ m L^{-1}224:$	$17 \ \mu g \ m L^{-1}$
	n = 9; $R = ZXoo: EC_{50} = 12.46 \ \mu g \ mL^{-1}, Xac: EC_{50} = 2.07 \ \mu g \ mL^{-1}225:$	$07 \ \mu g \ m L^{-1}$
	n = 10; $R = ZXoo: ECen = 2.98 ug mL-1. Xac: ECen = 6.08 ug mL-1$	8 ug mL ⁻¹
Reference		0
Compounds	226	
	HO	
Structure	$\langle \rightarrow$	
	HOOC	
Effects or mechanisms	226: MRSA SA5002: MIC = 16 mg L^{-1} MRSA SA5033: MIC = 16 mg L^{-1}	
Reference Abbreviations	MSSA SA5028: MIC = 16 mg L^{-1} 122 Xac: <i>Xanthomonas citri</i> subsp. <i>citri</i> . Xoo: <i>Xanthomonas oryzae</i> pv. o <i>ryzae</i> . Psa: <i>Pseudomonas syringae</i> pv. actinidiae. MRSA: methicillin-resistant <i>Staphylococcus aureus</i>	s aureus

This article is licensed under a Creative Commons Attribution 3.0 Unported Licence.

(cc) BY

Open Access Article. Published on 22 February 2024. Downloaded on 7/16/2025 11:59:10 PM.

and biological activities. By studying and modifying these compounds, scientists hope to develop more potent and effective antibiotics. Among the natural compounds explored for their antibacterial properties, 18 β -GA and related compounds have shown promise. These compounds have exhibited antibacterial effects against various bacterial strains, suggesting their potential as therapeutic agents. Further investigations are underway to elucidate the mechanisms of action and optimize the activity of these compounds.¹⁰³

The antimicrobial properties of 18β-GA, a compound extracted from the licorice plant, have been extensively studied by various researchers. Kim et al. discovered that 18β-GA has the ability to disrupt bacterial cell membranes, leading to the eradication of these microorganisms. This finding has generated significant interest in the potential of 18β-GA as a novel antibacterial agent.¹⁰⁴ Salari et al. further supported the antibacterial activity of 18β-GA against periodontopathogenic and capnophilic bacteria, while another investigation found that this natural compound can inhibit the growth of Helicobacter pylori.^{105,106} In a comprehensive study, Schrader et al. explored the antibacterial properties of various natural plant compounds, including 18β-GA and 18α-GA, and evaluated their efficacy against common pathogens found in pond-cultured channel catfish.107 It has been demonstrated that 18β-GA can effectively combat antibiotic-resistant bacterial strains, such as methicillin-resistant Staphylococcus aureus (MRSA), by inhibiting their survival and virulence gene expression.¹⁰⁸ Furthermore, this compound has shown potential in preventing the growth and formation of supragingival plaque bacteria and treating H. pylori infections.109,110 In the fight against opportunistic nosocomial P. aeruginosa, 18β-GA has proven to be a valuable ally.¹¹¹ Additionally, 18β-GA has been investigated for its ability to enhance the activity of tobramycin and polymyxin B against MRSA.¹¹² In the quest to combat opportunistic nosocomial P. aeruginosa, 18β-GA has been found to be a valuable ally.113 Moreover, 18β-GA has been used in combination with nanoparticles and hydrogels to combat bacterial infections. Darvishi et al. developed and evaluated the antibacterial activity of 18β-GA-loaded PL18β-GA nanoparticles, which demonstrated significant antibacterial activity against both Gram-positive and Gram-negative bacteria.114 Similarly, Zhao et al. engineered an injectable moldable hydrogel assembled from natural glycyrrhizic acid, which exhibited remarkable antibacterial activity against both types of bacteria.115 Recently, the remarkable antibacterial capabilities of 18β-GA derivatives have come to light. These derivatives have shown promising inhibitory effects against various bacterial strains, making them potential candidates for combating bacterial infections.¹¹⁶ In this review, our objective is to classify and elucidate the antibacterial activities of different 18β-GA derivatives against specific bacterial species. 18β-GA and its derivatives, as shown in Table 3, have demonstrated significant potential in inhibiting pathogens.

Compounds **157–163** have emerged as potent inhibitors of *Streptomyces scabies*, a notorious plant pathogen. These derivatives have exhibited remarkable inhibitory activity, suggesting their potential application in managing plant bacterial

diseases.¹¹⁷ Compound **161** has demonstrated superior inhibitory activity against *Bacillus subtilis*, *Staphylococcus aureus*, and MRAS compared to conventional antibiotics such as ampicillin, streptomycin, and vancomycin. This finding highlights the potential of 18 β -GA derivatives as effective alternatives for combating drug-resistant bacterial strains.

Furthermore, compounds **164–166**, compounds **177–178**, compounds **183–187**, and compounds **196–225** have displayed robust inhibitory activity against *Xanthomonas oryzae* pv. *oryzae* (Xoo) and *X. axonopodis* pv. *citri* (Xac).^{118–121} Xiang *et al.* particularly emphasized the potency of compounds **164** and **165**. *In vivo* trials have further confirmed the potential of these compounds in managing rice bacterial blight disease, with control efficacy ranging between 50.57% and 53.70% at 200 μ g mL^{-1.118}

Moreover, Yang *et al.* discovered that derivatives of 18β-GA (compounds **167–176**, **190–195**, and **226**) exhibit potent antibacterial activity against *Staphylococcus aureus*, *Staphylococcus epidermidis*, and MRAS.^{43,122} Compound **172**, as identified by Guo *et al.*, has demonstrated robust antibacterial properties and has been used to prepare supramolecular self-assembly hydrogels with exceptional thermodynamic stability and high melting temperatures.¹²³ Additionally, compounds **173–176** have exhibited high activity against various bacteria, particularly showing enhanced antibacterial effects against *Micrococcus luteus* compared to gentamicins.⁷⁹

Tropical bovine theileriosis (TBT) is one of the progressive and lymphoproliferative tick-borne diseases caused by *Theileria annulata*. Buvanesvaragurunathan *et al.* investigated the effect of 18β-GA esters (compounds **179–184**) on the growth of *Theileria annulata* and found that they induced apoptosis in parasite cells. Among these esters, the isopropyl ester of 18β-GA (compound **183**) showed improved anti-theileriosis efficacy than other 18β-GA derivatives.¹²⁴

In conclusion, the rise of drug-resistant bacteria necessitates the discovery of novel antibiotics that can effectively combat these resilient strains mentioned above. Natural compounds, such as 18 β -GA and its derivatives, offer a promising avenue for antibiotic development. Future research efforts should focus on understanding the mode of action of these compounds and optimizing their efficacy against drug-resistant bacteria.

Antiviral activity

Over the past two decades, the potencies have been extensively investigated for pentacyclic triterpenoids, such as asiatic acid, betulinic acid, boswellic acid, glycyrrhizin, 18 β -GA, lupeol, oleanolic acid, and ursolic acid, and their analogs and derivatives, as potent antitumor and antiviral agents. These triterpenoids have displayed remarkable cytotoxic activity against various tumor cell lines and exhibit antiviral properties, in particular, anti-HIV activity.¹²⁶ The main active constituents of licorice are triterpenoids, which have shown inhibitory effects on several viruses, including SARS-CoV-2.¹²⁷ It has been revealed that these compounds achieve their antiviral effects through various mechanisms such as inhibiting virus replication, directly inactivating viruses, halting inflammation mediated by HMGB1/TLR4, preventing β -chemokines, reducing the binding



of HMGB1 to DNA to weaken virus activity, and inhibiting reactive oxygen species formation.^{128,129} While these natural products offer great potential as anti-viral and anti-microbial agents, they comprise complex mixtures of organic molecules, making it difficult to determine their exact effectiveness. Hence, further research is required to gain an intricate understanding of their mechanisms of action and their potential for use as food or herbal medicine. Additionally, it is vital to carefully consider the pleiotropic effects of these compounds to avoid potential negative consequences.

Several studies have shown that 18β -GA inhibit several viruses (Fig. 5), for example, Sato *et al.* reported that 18β -GA inhibits *hepatitis B virus* (HBV) by suppressing surface antigens,¹³⁰ while Hardy *et al.* showed that 18β -GA exhibits significant antiviral activity against rotavirus replication *in vitro.*¹³¹ Other investigations demonstrated that 18β -GA inhibited rotavirus SA11 *via* the Fas/FasL pathway, inhibits *Epstein–Barr virus* (EBV) in superinfected Raji cells, showed significant antiviral activity against human immunodeficiency virus (HIV), inhibits infection of *human respiratory syncytial virus* (HRSV), and significantly protects against *murine hepatitis virus* (MHV)-induced severe hepatic injury by suppressing HMGB1 release.^{35,132–135}

In recent years, researchers have also worked on the antiviral properties of 18β-GA derivatives (Table 4). Baltina *et al.* synthesized a series of 18β-GA derivatives. They found that compounds **227–230** exert the most significant antiviral activity ($IC_{50} = 0.13 \mu M$) against ZIKV, with compound **227** demonstrating promising potential as an antiviral agent against ZIKV infection.¹³⁶ Similarly, Zígolo *et al.* reported that compound **231** exhibited significant antiviral activity against TK+ and TK– strains of *herpes simplex* virus type 1 (HSV-1).¹³⁷ Liang *et al.* found that water-soluble β-cyclodextrin-18β-GA (compounds **232–237**) showed promising antiviral activity against the influenza A/WSN/33 (H1N1) virus.^{138,139} More recently, Ding *et al.*

suggested that 18β-GA and its derivatives (compounds 238–241) could alleviate the symptoms of COVID-19 patients.¹⁴⁰ Additionally, Wang *et al.* synthesized several compounds and observed that compounds 242–243 exhibited significant inhibitory activities against HBV DNA replication.⁷³ These findings highlight the potential of 18β-GA and its derivatives as potent antiviral agents with remarkable antiviral activity against numerous viral infections.

In summary, the research on pentacyclic triterpenoids, including 18 β -GA and its derivatives, suggests their immense potential as effective and safe antiviral agents. These compounds have demonstrated varying degrees of antiviral activity against numerous viral infections, making them a promising area of ongoing research. However, further studies are necessary to comprehensively investigate their mechanisms of action and how they can be effectively used as food or herbal medicine while considering the possible negative consequences of their pleiotropic effects.

Antioxidant activity

18β-GA has been found to exhibit significant antioxidant activity, which makes it of great interest in the research of antioxidants. Alanazi et al. found that the serum concentrations of final glucose, aspartate aminotransferase (AST), alanine aminotransferase (ALT), and alkaline phosphatase (ALP) in mice treated with 20 mg per kg acrylamide (Acr) increased to $131 \pm 12.2 \text{ mg dL}^{-1}$, 76.5 $\pm 12.0 \ \mu \ U^{-1}$, 47.7 $\pm 9.17 \ \mu \ L^{-1}$, and 82.5 \pm 10.3 μ L⁻¹, which is much higher than the normal concentrations (serum final glucose, AST, ALT, and alkaline ALP concentrations of 87.7 \pm 5.93 mg dL $^{-1}$, 21.1 \pm 2.60 μ U $^{-1}$, 10.7 \pm 1.16 μ L⁻¹, and 24.1 \pm 3.97 μ L⁻¹), respectively, compared to these serums in the 18β-GA-Acr (50 mg per kg 18β-GA) group. The biochemical variables of rats return to normal. The findings provide sufficient evidence to demonstrate that 18β-GA possesses the capability to suppress the production of oxygen species and reinstate the antioxidant mechanisms in diabetic rats afflicted with acrylamide-induced liver and kidney cytotoxicity.141 Similarly, Melekoglu et al. discovered that the antioxidant defense system parameters, encompassing malondialdehyde (MDA), reduced glutathione (GSH), superoxide dismutase (SOD), and catalase (CAT), were significantly higher in the ovarian tissues of rats treated with 18β-GA (100 mg kg⁻¹ day⁻¹) compared to those subjected to ischemiareperfusion (I/R) alone.¹⁴² These findings suggest that 18β-GA may have protective effects against oxidative stress in a variety of tissues and systems. In addition to its potential antioxidant properties, recent research has also explored the potential therapeutic applications of 18β-GA in the context of viral infections. For example, Rehman et al. found that 18β-GA exhibited a solid binding affinity for several SARS-CoV-2 protein targets, including main protease (binding energy mol^{-1}), -9.46helicase (binding kcal energy = -9.91 kcal mol⁻¹), spike glycoprotein (S) (binding energy = -8.08 kcal mol⁻¹), and E-channel proteins (binding energy = -9.72 kcal mol⁻¹), through ligand-protein interactions. This



This article is licensed under a Creative Commons Attribution 3.0 Unported Licence.

(cc)) BY

Open Access Article. Published on 22 February 2024. Downloaded on 7/16/2025 11:59:10 PM.



Linker

п

236:

Influenza A/WSN/33 (H1N1) virus:IC₅₀ = 20.7 μ M, CC₅₀ Influenza A/WSN/33 (H1N1) virus: IC₅₀ = 11.0 μ M, CC₅₀ > 100 $\mu M,$ SI > 4.8 > 100 $\mu M,$ SI > 9.1 $\mathbf{R} = \mathbf{A}\mathbf{c}$ $\mathbf{R} = \mathbf{H}$ 237:

238:

 \mathbb{R}

HBV: $CC_{50}>985.68~\mu M,~IC_{50}=5.71~\mu M,$ SI > 172.6 239:

 $\mathbf{R} =$

Linker

HBV: $CC_{50} > 1373.13 \ \mu\text{M}, \ IC_{50} = 5.36 \ \mu\text{M}, \ SI > 255.9 \ SI > 255$

240: $\mathbf{R} =$

> mechanisms Effects or

Ś

HBV: $CC_{50} > 1327.92 \ \mu M$, IC_{50} : 8.90 μM , SI > 149.2 241: R =

HBV: $CC_{50}=37.17~\mu M,\,IC_{50}=9.08~\mu M,\,SI$ = 4.1



finding suggests that 18 β -GA may have the potential as a therapeutic agent in the fight against COVID-19.¹⁴³

We have discovered that a significant number of studies on the antioxidant properties of 18β-GA focus on its hepatoprotective function. In the mouse model of carbon tetrachloride (CCl₄)-induced chronic liver fibrosis, it was observed that CCl₄ inhibited the expression of Nrf2 regulatory genes, including CAT, glutathione peroxidase 2 (GPX2), and superoxide dismutase 3 (SOD3). However, 18β-GA was found to protect the mouse liver from oxidative stress by potentially activating the nuclear trans of Nrf2, enhancing the expression of its target genes, and increasing the activity of antioxidant enzymes.37 Furthermore, 18β-GA was also found to have the ability to inhibit the activity of xanthine oxidase (XO) significantly. XO is responsible for reducing O_2 to superoxide anionic radical O2, leading to oxidative stress.144 In a mouse model of methotrexate (MTX)-induced liver injury, Mahmoud et al. discovered that 18β-GA was able to reverse the significant manifestations of Nrf2, hemooxygenase-1, and PPARg induced by MTX, thus restoring antioxidant defense.³⁸ Another study demonstrated that 18β-GA significantly reduced alphanaphthylisothiocyanate (ANIT)-induced liver damage primarily by increasing the expression of nuclear factors (such as Sirt1, FXR, and Nrf2) and their targeted excretion transporters in the liver, which play a crucial role in maintaining bile acidosis in hepatocytes. The plasma levels of ALT, AST, ALP, γ -glutamyl transpeptidase (GGT), and total bilirubin (TBIL) were significantly elevated by 31.2-, 33.4-, 5.1-, 5.0-, and 91.3-fold, respectively, in rats induced with ANIT (P < 0.0001). However, for 18 β -GA (60 mg kg⁻¹ for 7 days treatment), all of these levels showed a significant reduction of 62.0%, 38.5%, 45.7%, 51.6%, and 39.7%, respectively (P < 0.05).¹⁴⁵ Moreover, the study also revealed that 18β-GA exerts its hepatoprotective effects against RTS-induced liver damage through the phosphatidylinositol 3kinase (PI3K)/protein kinase B (AKT) pathway and enhanced glycogen synthase kinase 3 beta (GSK3B) pathway, which promotes the Nrf2-mediated antioxidant system.146 Fig. 6 briefly illustrates the hepatoprotective effect of 18β-GA based on antiinflammatory and antioxidant mechanisms. Additionally, other hepatoprotective mechanisms are also discussed, such as the inhibition of the TLR/NF-KB pathway and upregulation of hepatic FXR to facilitate bile acid synthesis, transport, and detoxification, competitive inhibition of cyto P450 (CYP) enzymes responsible for the activation of pyrrolizidine alkaloid (PA) metabolism, particularly C3A1, which protects against liver damage, activation of PXR to regulate autophagy and lysosomal biogenesis, thereby alleviating acute liver injury, inhibition of hepatic stellate cell activation, and direct transcriptional inhibition of $\alpha 2$ (I) collagen gene (COL1A2), as observed in transgenic reporter mice, and other mechanisms.147-150

18β-GA derivatives (Table 5) also demonstrated significant antioxidant activity. It was discovered that compounds **244–247** exhibited robust antioxidant activity and inhibited ROS activity by up to 41%.¹⁵¹ Maitraie *et al.* observed that compounds **249– 258** displayed both anti-inflammatory and antioxidant properties, with compound **254** specifically exerting inhibitory effects on NO and superoxide anions in RAW 246.7 cells.¹⁵² Moreover,



Fig. 6 Mechanism of hepatoprotective effect of glycyrrhetinic acid.

Zhang et al. found that compounds 259-263 hindered the proliferation of activated hepatic stellate cells (HSC)-T6 cells by inducing apoptosis and arresting them in the G0/G1 phase. They used rat hepatic stellate cell line T6 cells activated by transforming growth factor-\beta-1 as the cell model and as the 18\beta-GA control. The IC₅₀ value of the compound on cell proliferation was determined by tetrazolium salt colorimetry. It was found that the inhibitory effect of compounds 259-263 on activated HSC-T6 was stronger than that of GA (IC_{50} = 78.4 \pm 2.3 μM). 153 Numerous studies have demonstrated a strong association between COX-2 and the activation of hepatic stellate cells (HSCs), thereby facilitating the initiation and progression of hepatic fibrosis. Among them, compounds 262 and 265 strongly inhibit the activation of HSC-T6 cells by downregulating the expression of alpha-smooth muscle actin (a-SMA) and type I collagen (Col1) proteins, which are biomarkers of liver fibrosis. After treatment with compound activated HSC-T6, the expression levels of the two biomarkers were down-regulated. Second, both compounds downregulated the expression levels of COX-2 and transforming growth factor beta1 (TGF- β_1) and reduced ROS levels in a concentration-dependent manner. This suggests that they inhibit HSC-T6 activation and may also be due to downregulation of COX-2 levels, inhibition of the TGF-B1 signaling pathway, and reduction of ROS levels.

Overall, while the study of oxidative stress and its effects on the body is complex, recent research has shed light on the potential benefits of compounds like 18β -GA in combatting this process. By exploring the mechanisms of these compounds and their effects on various tissues and systems, we can better understand how to combat oxidative stress and its associated health risks.

Discussion

Experience has imparted the understanding that when a compound manifests a biological activity characterized by an IC_{50} value lower than 10 μ M, it may be classified as potential biological efficacy. Additionally, in the process of scrutinizing lead and candidate compounds, it is importance to consider both cost-effectiveness and the intricacy of synthetic routes. Keeping these pivotal factors in consideration, the investigation unveiled that compounds 16-21 exhibited noteworthy inhibitory activities against 11β-HSD2 within the sub-micromolar (nM) range. Particularly remarkable is compound 16, which boasts an exceptionally modest synthetic complexity, necessitating a single-step reaction initiated from 18β-GA. The incorporation of amide and hydroxyl groups at the C-30 position has substantially augmented the solubility of 18β-GA. Compounds of this kind exhibit tremendous promise for further in-depth exploration. Moreover, numerous studies have demonstrated that the majority of structural alterations to 18β-GA revolve around rigid five-ring skeleton structure, encompassing the



RSC Advances

This article is licensed under a Creative Commons Attribution 3.0 Unported Licence.

(cc) BY

6592 | RSC Adv., 2024, 14, 6557-6597

Open Access Article. Published on 22 February 2024. Downloaded on 7/16/2025 11:59:10 PM.





© 2024 The Author(s). Published by the Royal Society of Chemistry RSC Adv., 20

RSC Adv., 2024, 14, 6557-6597 | 6593

addition, removal, and replacement of functional groups. Comparatively, few studies explore the strategy, such as scaffold hopping and changes in the skeleton itself to the biological activity. Reports about compounds **38–41**, **116–122**, **227–230**, and **248–259** have discernible indicated that brought about a substantial augmentation in the anti-tumor, antiviral, and antioxidant properties of 18β-GA through the processes of ring opening and ring expansion. The modifications in 18β-GA from the complexity of the derivative structure is mainly due to addition rather than subtraction. It may be connected with that there are few reaction methods for removing carbon atoms in the rigid alkyl skeleton.

It is particularly noteworthy that compounds 227-230 demonstrate an inhibitory activity against the ZIKA virus within the sub-micromolar (nM) range. Perhaps designing modifications that involve adding or reducing rings could provide excellent solutions for enhancing the target binding strength, selectivity, bioavailability, selective tissue distribution, and metabolic stability of 18β-GA derivatives. However, further studies are necessary to comprehensively reveal their mechanisms or the target protein to further guide the modification of compounds. Moreover, 18β-GA derivatives that self-assemble, including gels, micelles, nanoparticles, and liposomes, hold potential for application in food additives and intelligent drug delivery due to availability, biocompatibility, and controllable degradability.¹⁵⁴ Additionally, while the mainstream research direction focuses on the aforementioned topics, shifting the focus to other biologically active research areas such as antidiabetes, anti-coagulation, and neuroprotection, could prove worthwhile, as the studies in these areas are still relatively scarce. This could further broaden the development prospects of 18β-GA derivatives and increase their role in various fields.

Conclusions

In conclusion, the past decade has yielded promising research on the therapeutic potential of 18β-GA and its derivatives for various diseases, including cancer, inflammation, bacterial infection, hepatic diseases, and viral infections. Pharmacological effects have been observed through a variety of pathways, including inflammation-related signaling, immune response modulation, and gene expression regulation. However, it is unfortunate that no derivatives have entered clinical trials (from https://www.clinicaltrials.gov) due to their poor pharmacological properties, low bioavailability, significant toxic side effects, and other factors.

The review of over 200 chemical structures and key activity data in this review article serves as a valuable data resource for pharmaceutical chemists and also provides future research directions. Future research, except self-assembling derivatives, as well as exploring other related fields should more focus on revealing the mechanisms of action or the target protein and the relationship with the SAR of derivates and to further guide the structural modifications. With further research and optimization, 18 β -GA derivatives will address the above crucial issues that hold great promise as potential therapeutic agents for various diseases.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

The author thanks Hainan Provincial Natural Science Foundation of China (No. 2019RC229 and No. 20152023) and Hainan Provincial Graduate Students Scientific Innovative Foundation (Qhys2021-368) for financial support. Sincerely thanks Professor Wang, Professor Li and Professor Xu for their selfless help and guidance in the process of writing.

Notes and references

- 1 J. Padmavathy and S. Devarajan, *Bangladesh J. Pharmacol.*, 2017, **12**, 151–161.
- 2 C. Sabbadin, L. Bordin, G. DonÃ, J. Manso, G. Avruscio and D. Armanini, *Front. Endocrinol.*, 2019, **10**, 484.
- 3 D. J. Newman and G. M. Cragg, *J. Nat. Prod.*, 2020, **83**, 770–803.
- 4 S. E. Osagie-Eweka, N. E. J. Orhue, E. K. I. Omogbai and F. C. Amaechina, *Toxicol Rep*, 2021, **8**, 239–247.
- 5 H. B. Wang, Y. He, M. L. Jian, X. G. Fu, Y. H. Cheng, Y. J. He, J. Fang, L. Li and D. Zhang, *Molecules*, 2022, **27**, 21.
- 6 M. Ahmad, M. Jalaluddin and B. P. Panda, *Ann. Microbiol.*, 2013, **64**, 683–688.
- 7 Q. Ni, Y. Gao, X. Yang, Q. Zhang, B. Guo, J. Han and S. Chen, *Front. Pharmacol*, 2022, **13**, 1001018.
- 8 C. S. Graebin, *The Pharmacological Activities of Glycyrrhizinic Acid ("Glycyrrhizin") and Glycyrrhetinic Acid*, Springer International Publishing, Cham, 2018.
- 9 A. Kowalska and U. Kalinowska-Lis, *Int. J. Cosmet. Sci.*, 2019, **41**(4), 325–331.
- 10 L. Jin, L. Dai, M. Ji and H. Wang, *Bioorg. Chem.*, 2019, **85**, 179–190.
- 11 H. Wang, R. W. Li, Y. Rao, S. X. Liu, C. H. Hu, Y. Zhang, L. C. Meng, Q. L. Wu, Q. H. Ouyang, H. Liang and M. Qin, *Pharmaceutics*, 2022, 14, 1797.
- 12 A. Mittal, M. Nagpal and V. K. Vashistha, *Rev. Bras. Farmacogn.*, 2023, **33**, 1154–1169.
- 13 R. Guo, Y. Liu, R. Sheng and J. Fan, *Mini-Rev. Med. Chem.*, 2022, **22**, 2024–2066.
- M. G. Netea, F. Balkwill, M. Chonchol, F. Cominelli, M. Y. Donath, E. J. Giamarellos-Bourboulis, D. Golenbock, M. S. Gresnigt, M. T. Heneka, H. M. Hoffman, R. Hotchkiss, L. A. B. Joosten, D. L. Kastner, M. Korte, E. Latz, P. Libby, T. Mandrup-Poulsen, A. Mantovani, K. H. G. Mills, K. L. Nowak, L. A. O'Neill, P. Pickkers, T. van der Poll, P. M. Ridker, J. Schalkwijk, D. A. Schwartz, B. Siegmund, C. J. Steer, H. Tilg, J. W. M. van der Meer, F. L. van de Veerdonk and C. A. Dinarello, *Nat. Immunol.*, 2017, 18, 826–831.
- 15 R. Medzhitov, Cell, 2010, 140, 771-776.
- 16 T. Lawrence, Cold Spring Harbor Perspect. Biol., 2009, 1, a001651.

- 17 M. Groslambert and B. F. Py, *J. Inflammation Res.*, 2018, **11**, 359–374.
- 18 A. Quintás-Cardama and S. Verstovsek, *Clin. Cancer Res.*, 2013, **19**, 1933–1940.
- 19 L. A. O'neill, D. Golenbock and A. G. Bowie, *Nat. Rev. Immunol.*, 2013, **13**, 453–460.
- 20 J.-M. Cavaillon, Clin. Rev. Allergy Immunol., 2023, 65, 183– 187.
- 21 V. Thiruchenthooran, E. Sánchez-López and A. Gliszczyńska, *Cancers*, 2023, **15**, 475.
- 22 S. A. Richard, Mediators Inflammation, 2021, 2021, 1-15.
- 23 J.-X. Zhou and M. Wink, Medicines, 2019, 6, 55.
- 24 C.-Y. Wang, T.-C. Kao, W.-H. Lo and G.-C. Yen, *J. Agric. Food Chem.*, 2011, **59**, 7726–7733.
- 25 L.-N. Peng, L. Li, Y.-F. Qiu, J.-H. Miao, X.-Q. Gao, Y. Zhou, Z.-X. Shi, Y.-L. Xu, D.-H. Shao, J.-C. Wei and Z.-Y. Ma, *J. Asian Nat. Prod. Res.*, 2011, **13**, 942–950.
- 26 J. Liu, Y. Xu, M. Yan, Y. Yu and Y. Guo, *Sci. Rep.*, 2022, **12**, 3121.
- 27 G. L. Gupta, L. Sharma and M. Sharma, *Neurochem. Res.*, 2023, 48, 551–569.
- 28 T. Ishida, I. Miki, T. Tanahashi, S. Yagi, Y. Kondo, J. Inoue, S. Kawauchi, S. Nishiumi, M. Yoshida, H. Maeda, C. Tode, A. Takeuchi, H. Nakayama, T. Azuma and S. Mizuno, *Eur. J. Pharmacol.*, 2013, **714**, 125–131.
- 29 W. Zheng, X. Huang, Y. Lai, X. Liu, Y. Jiang and S. Zhan, *Front. Pharmacol*, 2021, **12**, 631206.
- 30 R. Li, K. Wu, Y. Li, X. Liang, K. P. Lai and J. Chen, *Briefings Bioinf.*, 2021, **22**, 1161–1174.
- 31 Z. Zhao, Y. Xiao, L. Xu, Y. Liu, G. Jiang, W. Wang, B. Li, T. Zhu, Q. Tan and L. Tang, ACS Appl. Mater. Interfaces, 2021, 13, 20995–21006.
- 32 J.-X. Zhou and M. Wink, Medicines, 2019, 6, 55.
- A. V. Shetty, S. Thirugnanam, G. Dakshinamoorthy,
 A. Samykutty, G. Zheng, A. Chen, M. C. Bosland,
 A. Kajdacsy-Balla and M. Gnanasekar, *Int. J. Oncol.*, 2011,
 39, 635–640.
- 34 Y. Xiao, J. Xu, C. Mao, M. Jin, Q. Wu, J. Zou, Q. Gu, Y. Zhang and Y. Zhang, *J. Biol. Chem.*, 2010, **285**, 1128–1137.
- 35 X. Shi, L. Yu, Y. Zhang, Z. Liu, H. Zhang, Y. Zhang, P. Liu and P. Du, *Int. Immunopharmacol.*, 2020, **84**, 106578.
- 36 A. M. Mahmoud and H. S. Al Dera, *Genes Nutr.*, 2015, **10**, 1– 13.
- 37 S. Chen, L. Zou, L. Li and T. Wu, PLoS One, 2013, 8, e53662.
- 38 A. M. Mahmoud, O. E. Hussein, W. G. Hozayen and S. M. Abd El-Twab, *Chem.-Biol. Interact.*, 2017, 270, 59–72.
- 39 Z. Wang, J. Ma, Y. He, K. K. Miu, S. Yao, C. Tang, Y. Ye and G. Lin, *Phytomedicine*, 2022, **102**, 154162.
- 40 Y. Ma, J. M. Liu, R. D. Chen, X. Q. An and J. G. Dai, *Chin. Chem. Lett.*, 2017, 28, 1200–1204.
- 41 B. Y. Fan, B. C. Jiang, S. S. Yan, B. H. Xu, H. L. Huang and G. T. Chen, *Planta Med.*, 2019, **85**, 56–61.
- 42 B. Li, Y. Yang, L. Chen, S. Chen, J. Zhang and W. Tang, *MedChemComm*, 2017, 8, 1498–1504.
- 43 Y. Yang, Q. Zhu, Y. Zhong, X. Cui, Z. Jiang, P. Wu, X. Zheng,
 K. Zhang and S. Zhao, *Bioorg. Chem.*, 2020, **101**, 103985.

- 44 M. Bian, D. Zhen, Q.-K. Shen, H.-H. Du, Q.-Q. Ma and Z.-S. Quan, *Bioorg. Chem.*, 2021, **107**, 104598.
- 45 Q. P. Zhang, Y. N. Wang, Z. Y. Wang, E. A. H. Mohammed, Q. Y. Zhao, D. He and Z. Wang, *Bioorg. Chem.*, 2022, **119**, 105542.
- 46 B. Li, S. Cai, Y. A. Yang, S. C. Chen, R. Chen, J. B. Shi, X. H. Liu and W. J. Tang, *Eur. J. Med. Chem.*, 2017, 139, 337–348.
- 47 H. B. Wang, J. W. Zuo, L. Zha, X. Jiang, C. X. Wu, Y. A. Yang, W. J. Tang and T. L. Shi, *Bioorg. Chem.*, 2021, **110**, 104755.
- 48 A. V. Markov, A. V. Sen'kova, I. I. Popadyuk,
 O. V. Salomatina, E. B. Logashenko, N. I. Komarova,
 A. A. Ilyina, N. F. Salakhutdinov and M. A. Zenkova, *Int. J. Mol. Sci.*, 2020, 21, 3511.
- 49 B. Tu, J. Liang, Y. Ou, X. Zhang, W. Zheng, R. Wu, L. Gan, D. Li, Y. Lu, J. Wu, W. David Hong, K. Zhang, P. Wu, J. Jin and W.-L. Wong, *Bioorg. Chem.*, 2022, **122**, 105714.
- 50 X. Su, H. Lawrence, D. Ganeshapillai, A. Cruttenden, A. Purohit, M. J. Reed, N. Vicker and B. V. L. Potter, *Bioorg. Med. Chem.*, 2004, **12**, 4439–4457.
- 51 R. Gaware, R. Khunt, L. Czollner, C. Stanetty, T. D. Cunha, D. V. Kratschmar, A. Odermatt, P. Kosma, U. Jordis and D. Claßen-Houben, *Bioorg. Med. Chem.*, 2011, **19**, 1866– 1880.
- 52 D. V. Kratschmar, A. Vuorinen, T. D. Cunha, G. Wolber, D. Classen-Houben, O. Doblhoff, D. Schuster and A. Odermatt, *J. Steroid Biochem. Mol. Biol.*, 2011, 125, 129– 142.
- 53 R. You, W. Long, Z. Lai, L. Sha, K. Wu, X. Yu, Y. Lai, H. Ji, Z. Huang and Y. Zhang, J. Med. Chem., 2013, 56, 1984–1995.
- 54 B. Xu, G.-R. Wu, X.-Y. Zhang, M.-M. Yan, R. Zhao, N.-N. Xue, K. Fang, H. Wang, M. Chen, W.-B. Guo, P.-L. Wang and H.-M. Lei, *Molecules*, 2017, 22, 924.
- 55 H. Sung, J. Ferlay, R. L. Siegel, M. Laversanne, I. Soerjomataram, A. Jemal and F. Bray, *Ca-Cancer J. Clin.*, 2021, **71**, 209–249.
- 56 F. Bray, J. Ferlay, I. Soerjomataram, R. L. Siegel, L. A. Torre and A. Jemal, *Ca-Cancer J. Clin.*, 2018, **68**, 394–424.
- 57 S. Tauro, B. Dhokchawle, P. Mohite, D. Nahar, S. Nadar and E. Coutinho, *Curr. Med. Chem.*, 2023, 7, 848–870.
- 58 S. Wang, Y. Shen, R. Qiu, Z. Chen, Z. Chen and W. Chen, *Int. J. Oncol.*, 2017, **51**, 615–624.
- 59 Y.-H. Luo, C. Wang, W.-T. Xu, Y. Zhang, T. Zhang, H. Xue, Y.-N. Li, Z.-R. Fu, Y. Wang and C.-H. Jin, *OncoTargets Ther.*, 2021, **14**, 5131.
- 60 J. Shi, J. Li, J. Li, R. Li, X. Wu, F. Gao, L. Zou, W. W. S. Mak, C. Fu and J. Zhang, *Phytomedicine*, 2021, **81**, 153408.
- 61 A. Speciale, C. Muscarà, M. S. Molonia, M. Cristani, F. Cimino and A. Saija, *Molecules*, 2022, 27, 1775.
- 62 C. Liu, Q. Ma, G. Gong and F. Su, Molecules, 2023, 28, 5855.
- 63 H. Hussain, I. Ali, D. Wang, F. L. Hakkim, B. Westermann, I. Ahmed, A. M. Ashour, A. Khan, A. Hussain, I. R. Green and S. T. A. Shah, *Expert Opin. Drug Discovery*, 2021, 16, 1497–1516.
- 64 Y. Lai, L. Shen, Z. Zhang, W. Liu, Y. Zhang, H. Ji and J. Tian, *Bioorg. Med. Chem. Lett.*, 2010, **20**, 6416–6420.

- 65 J. Hu, Y. Wu, C. Zhao, Y. Ju and Y. Ju, *Chem. J. Chin. Univ.*, 2010, 31, 1762–1768.
- 66 D. P. Alho, J. A. Salvador, M. Cascante and S. Marin, *Molecules*, 2019, 24, 766.
- 67 D. Cai, Z. Zhang, Y. Chen, Y. Zhang, Y. Sun and Y. Gong, *Molecules*, 2019, 24, 3631.
- 68 F. Zhou, G.-R. Wu, D.-S. Cai, B. Xu, M.-M. Yan, T. Ma, W.-B. Guo, W.-X. Zhang, X.-M. Huang, X.-h. Jia, Y.-Q. Yang, F. Gao, P.-L. Wang and H.-M. Lei, *Eur. J. Med. Chem.*, 2019, **178**, 623–635.
- 69 L. Dai, J. Li, J. Yang, Y. Men, Y. Zeng, Y. Cai and Y. Sun, *Catalysts*, 2018, **8**, 615.
- 70 R. Wang, Y. Li, X. Huai, Q. Zheng, W. Wang, H.-J. Li and Q. Huai, *Drug Des., Dev. Ther.*, 2018, **12**, 1321–1336.
- 71 D. K. Yadav, A. Meena, A. Srivastava, D. Chanda, F. Khan and S. Chattopadhyay, *Drug Des., Dev. Ther.*, 2010, 4, 173– 186.
- 72 D. Cai, Z. hua Zhang, Y. Chen, C. Ruan, S. qiang Li, S. qin Chen and L. shan Chen, *RSC Adv.*, 2020, **10**, 11694–11706.
- 73 L.-J. Wang, C.-A. Geng, Y.-B. Ma, X.-Y. Huang, J. Luo, H. Chen, X.-M. Zhang and J.-J. Chen, *Bioorg. Med. Chem. Lett.*, 2012, 22, 3473–3479.
- 74 B. Lallemand, M. Gelbcke, J. Dubois, M. Prévost, I. Jabin and R. Kiss, *Mini-Rev. Med. Chem.*, 2011, **11**, 881–887.
- 75 Y. Li, L. Feng, Z.-F. Song, H.-B. Li and Q.-Y. Huai, *Molecules*, 2016, **21**, 199.
- 76 J. Tatsuzaki, M. Taniguchi, K. F. Bastow, K. Nakagawa-Goto, S. L. Morris-Natschke, H. Itokawa, K. Baba and K.-H. Lee, *Bioorg. Med. Chem.*, 2007, **15**, 6193–6199.
- 77 S.-C. Yu, Y.-T. Hou, C.-M. Hsu, F.-J. Tsai and Y. Tsai, J. Inclusion Phenom. Macrocyclic Chem., 2022, 102, 339–346.
- 78 R. Csuk, S. Schwarz, R. Kluge and D. Ströhl, *Phytomedicine*, 2010, 45, 5718–5723.
- 79 G. O. Moustafa, A. Shalaby, A. M. Naglah, M. M. Mounier, H. El-Sayed, M. M. Anwar and E. S. Nossier, *Molecules*, 2021, 26, 4573.
- 80 R. Csuk, S. Schwarz, R. Kluge and D. Ströhl, *Eur. J. Med. Chem.*, 2010, 45, 5718–5723.
- 81 I. Serbian, R. K. Wolfram, L. Fischer, A. Al-Harrasi and R. Csuk, *Mediterr. J. Chem.*, 2018, 7, 286–293.
- 82 G. Chadalapaka, I. Jutooru, A. McAlees, T. Stefanac and S. Safe, *Bioorg. Med. Chem. Lett.*, 2008, 18, 2633–2639.
- 83 Y. Gao, X. Guo, X. Li, D. Liu, D. Song, Y. Xu, M. Sun, Y. Jing and L. Zhao, *Molecules*, 2010, 15, 4439–4449.
- 84 E. B. Logashenko, O. V. Salomatina, A. V. Markov, D. V. Korchagina, N. F. Salakhutdinov, G. A. Tolstikov, V. V. Vlassov and M. A. Zenkova, *ChemBioChem*, 2011, 12, 784–794.
- 85 O. V. Salomatina, A. V. Markov, E. B. Logashenko, D. V. Korchagina, M. A. Zenkova, N. F. Salakhutdinov, V. V. Vlassov and G. A. Tolstikov, *Bioorg. Med. Chem.*, 2014, 22, 585–593.
- 86 L.-F. Yang, Y. Xing, J.-X. Xiao, J. Xie, W. Gao, J. Xie, L.-T. Wang, J. Wang, M. Liu and Z. Yi, ACS Med. Chem. Lett., 2018, 9, 1105–1110.
- 87 P. Alper, O. V. Salomatina, N. F. Salakhutdinov, E. Ulukaya and F. Ari, *Bioorg. Med. Chem.*, 2021, **30**, 115963.

- 88 S. Wang, Y. Shen, R. Qiu, Z. Chen, Z. Chen and W. Chen, *Int. J. Oncol.*, 2017, **51**, 615–624.
- 89 S. Schwarz and R. Csuk, *Bioorg. Med. Chem.*, 2010, **18**, 7458–7474.
- 90 R. Csuk, S. Schwarz, B. Siewert, R. Kluge and D. Ströhl, *Eur. J. Med. Chem.*, 2011, 46, 5356–5369.
- 91 R. Csuk, S. Schwarz, R. Kluge and D. Ströhl, Arch. Pharm., 2012, 345, 28–32.
- 92 R. Csuk, S. Schwarz, B. Siewert, R. Kluge and D. Ströhl, Z. Naturforsch. B Chem. Sci., 2012, 67, 731–746.
- 93 R. Csuk, S. Schwarz, B. Siewert, R. Kluge and D. Ströhl, *Arch. Pharm.*, 2012, **345**, 223–230.
- 94 W. Guo, M. Yan, B. Xu, F. Chu, W. Wang, C. Zhang, X. Jia,
 Y. Han, H. Xiang and Y. Zhang, *Chem. Cent. J.*, 2016, 10,
 1–11.
- 95 K. Li, T. Ma, J. Cai, M. Huang, H. Guo, D. Zhou, S. Luan, J. Yang, D. Liu and Y. Jing, *Bioorg. Med. Chem.*, 2017, 25, 5441–5451.
- 96 K.-W. Lin, A.-M. Huang, T.-C. Hour, S.-C. Yang, Y.-S. Pu and C.-N. Lin, *Bioorg. Med. Chem.*, 2011, **19**, 4274–4285.
- 97 M. Huang, P. Gong, Y. Wang, X. Xie, Z. Ma, Q. Xu, D. Liu, Y. Jing and L. Zhao, *Bioorg. Chem.*, 2020, **103**, 104187.
- 98 Q.-X. Zheng, R. Wang, Y. Xu, C.-X. He, C.-Y. Zhao, Z.-F. Wang, R. Zhang, W. Dehaen, H.-J. Li and Q.-Y. Huai, *Biol. Pharm. Bull.*, 2020, 43, 102–109.
- 99 J. Sun, H.-Y. Liu, C.-Z. Lv, J. Qin and Y.-F. Wu, J. Agric. Food Chem., 2019, 67, 9643–9651.
- 100 N. Dheman, N. Mahoney, E. M. Cox, J. J. Farley, T. Amini and M. L. Lanthier, *Clin. Infect. Dis.*, 2020, **73**, e4444–e4450.
- 101 J. Sun, H.-Y. Liu, C.-Z. Lv, J. Qin and Y.-F. Wu, J. Agric. Food Chem., 2019, 67, 9643–9651.
- 102 N. Dheman, N. Mahoney, E. M. Cox, J. J. Farley, T. Amini and M. L. Lanthier, *Clin. Infect. Dis.*, 2020, **73**, ciaa859.
- 103 D. Langer, B. Czarczynska-Goslinska and T. Goslinski, *Curr. Issues Pharm. Med. Sci.*, 2016, 29, 118–123.
- 104 H. K. Kim, Y. Park, H. N. Kim, B. H. Choi, H. G. Jeong, D. G. Lee and K.-S. Hahm, *Biotechnol. Lett.*, 2002, 24, 1899–1902.
- 105 M. H. Salari and Z. Kadkhoda, *Clin. Microbiol. Infect.*, 2003, 9, 987–988.
- 106 J. Bielenberg and R. Krausse, Phytother. Res., 2004, 2, 37–39.
- 107 K. K. Schrader, *Toxins*, 2010, 2, 1676–1689.
- 108 R. Long Danyelle, J. Mead, M. Hendricks Jay, E. Hardy Michele and M. Voyich Jovanka, *Antimicrob. Agents Chemother.*, 2013, 57, 241–247.
- 109 N. Dewake, X. Ma, K. Sato, S. Nakatsu, K. Yoshimura,
 Y. Eshita, H. Fujinaka, Y. Yano, N. Yoshinari and
 A. Yoshida, *Microbiol. Immunol.*, 2021, 65, 343–351.
- 110 M. M. Celik and N. Duran, *Revista Romana de Medicina de Laborator*, 2019, 27, 63–71.
- 111 S. Kannan, G. Sathasivam and M. Marudhamuthu, *Microb. Pathog.*, 2019, **126**, 332–342.
- 112 A. d. Breij, T. G. Karnaoukh, J. Schrumpf, P. S. Hiemstra, P. H. Nibbering, J. T. v. Dissel and P. C. d. Visser, *Eur. J. Clin. Microbiol. Infect. Dis.*, 2016, **35**, 555–562.
- 113 Y. Zhao and X. Su, BB Rep., 2023, 33, 101427.

- 114 B. Darvishi, S. Manoochehri, M. Esfandyari-Manesh, N. Samadi, M. Amini, F. Atyabi and R. Dinarvand, *Drug Res.*, 2015, **65**, 617–623.
- 115 X. Zhao, H. Zhang, Y. Gao, Y. Lin and J. Hu, *ACS Appl. Bio Mater.*, 2020, **3**, 648–653.
- 116 E. A. H. Mohammed, Y. Peng, Z. Wang, X. Qiang and Q. Zhao, *Russ. J. Bioorg. Chem.*, 2022, **48**, 906–918.
- 117 L.-R. Huang, X.-J. Hao, Q.-J. Li, D.-P. Wang, J.-X. Zhang, H. Luo and X.-S. Yang, *J. Nat. Prod.*, 2016, **79**, 721–731.
- 118 M. Xiang, Y.-L. Song, J. Ji, X. Zhou, L.-W. Liu, P.-Y. Wang, Z.-B. Wu, Z. Li and S. Yang, *Pest Manage. Sci.*, 2020, 76, 2959–2971.
- 119 L. Zhang, Y. Fu, Y. Ding, J. Meng, Z. Wang and P. Wang, Chem. Res. Chin. Univ., 2021, 37, 662–667.
- 120 Y.-l. Song, H.-w. Liu, Y.-h. Yang, J.-j. He, B.-x. Yang, L.-l. Yang, X. Zhou, L.-w. Liu, P.-y. Wang and S. Yang, J. Integr. Agric., 2022, 22, 2759–2771.
- 121 J.-J. He, T. Li, H.-W. Liu, L.-L. Yang, Y.-H. Yang, Q.-Q. Tao, X. Zhou, P.-Y. Wang and S. Yang, *Arabian J. Chem.*, 2023, 16, 104771.
- 122 K. Oyama, M. Kawada-Matsuo, Y. Oogai, T. Hayashi, N. Nakamura and H. Komatsuzawa, *PLoS One*, 2016, **11**, e0165831.
- 123 S. Guo, S. Chen, N. Cao, W. Zheng, D. Li, Z. Sheng, X. Xu, Q. Zhang, X. Zheng, K. Wu, P. Wu, K. Zhang and W. D. Hong, *J. Mater. Sci.*, 2021, 56, 17254–17267.
- 124 K. Buvanesvaragurunathan, J. Ganesh, S. N. Kumar,
 V. Porchezhiyan, A. Radha, P. Azhahianambi,
 P. Pandikumar and S. Ignacimuthu, *Exp. Parasitol.*, 2022,
 236, 108258.
- 125 N. Dewake, X. Ma, K. Sato, S. Nakatsu, K. Yoshimura, Y. Eshita, H. Fujinaka, Y. Yano, N. Yoshinari and A. Yoshida, *Med. Microbiol. Immunol.*, 2021, 65, 343–351.
- 126 R. Paduch and M. Kandefer-Szerszen, *Mini-Rev. Org. Chem.*, 2014, **11**, 262–268.
- 127 D. Elebeedy, W. F. Elkhatib, A. Kandeil, A. Ghanem, O. Kutkat, R. Alnajjar, M. A. Saleh, A. I. Abd El Maksoud, I. Badawy and A. A. Al-Karmalawy, *RSC Adv.*, 2021, 11, 29267–29286.
- 128 J.-Y. Pu, L. He, S.-Y. Wu, P. Zhang and X. Huang, *J. Virol.*, 2013, **29**, 673–679.
- 129 C. Huan, Y. Xu, W. Zhang, T. Guo, H. Pan and S. Gao, *Front. Pharmacol*, 2021, **12**, 680674.
- 130 H. Sato, W. Goto, J.-i. Yamamura, M. Kurokawa, S. Kageyama, T. Takahara, A. Watanabe and K. Shiraki, *Antiviral Res.*, 1996, 30, 171–177.
- 131 M. E. Hardy, J. M. Hendricks, J. M. Paulson and N. R. Faunce, *Virol. J.*, 2012, **9**, 96.
- 132 X. Wang, F. Xie, X. Zhou, T. Chen, Y. Xue and W. Wang, *Pharmaceut. Biol.*, 2021, **59**, 1096–1103.
- 133 J.-C. Lin, J.-M. Cherng, M.-S. Hung, L. A. Baltina, L. Baltina and R. Kondratenko, *Antiviral Res.*, 2008, **79**, 6–11.

- K. Fukuchi, N. Okudaira, K. Adachi, R. Odai-Ide, S. Watanabe, H. Ohno, M. Yamamoto, T. Kanamoto, S. Terakubo and H. Nakashima, *In Vivo*, 2016, 30, 777–785.
- 135 C. F. Yeh, K. C. Wang, L. C. Chiang, D. E. Shieh, M. H. Yen and J. S. Chang, *J. Ethnopharmacol.*, 2013, **148**, 466–473.
- 136 L. A. Baltina, H.-C. Lai, Y.-C. Liu, S.-H. Huang, M.-J. Hour, L. A. Baltina, T. R. Nugumanov, S. S. Borisevich, L. M. Khalilov and S. F. Petrova, *Bioorg. Med. Chem.*, 2021, 41, 116204.
- 137 M. A. Zígolo, M. Salinas, L. Alché, A. Baldessari and G. G. Liñares, *Bioorg. Chem.*, 2018, 78, 210–219.
- 138 S. Liang, M. Li, X. Yu, H. Jin, Y. Zhang, L. Zhang, D. Zhou and S. Xiao, *Eur. J. Med. Chem.*, 2019, **166**, 328–338.
- 139 S. Liang, X. Ma, M. Li, Y. Yi, Q. Gao, Y. Zhang, L. Zhang, D. Zhou and S. Xiao, *Front. Chem.*, 2022, **10**, 836955.
- 140 H. Ding, W. Deng, L. Ding, X. Ye, S. Yin and W. Huang, *J. Med. Virol.*, 2020, **92**, 2200–2204.
- 141 I. S. Alanazi, M. Emam, M. Elsabagh, S. Alkahtani and M. M. Abdel-Daim, *Environ. Sci. Pollut. Res.*, 2021, 28, 58322–58330.
- 142 R. Melekoglu, O. Ciftci, S. Eraslan, S. Alan and N. Basak, *BioMed Res. Int.*, 2018, **2018**, 5421308.
- 143 M. F. u. Rehman, S. Akhter, A. I. Batool, Z. Selamoglu,
 M. Sevindik, R. Eman, M. Mustaqeem, M. S. Akram,
 F. Kanwal and C. Lu, *Antibiotics*, 2021, 10, 1011.
- 144 S. K. Hasan, R. Khan, N. Ali, A. Q. Khan, M. U. Rehman, M. Tahir, A. Lateef, S. Nafees, S. J. Mehdi and S. Rashid, *Hum. Exp. Toxicol.*, 2015, 15, 104187.
- 145 S.-y. Wu, S.-c. Cui, L. Wang, Y.-t. Zhang, X.-x. Yan, H.-l. Lu, G.-z. Xing, J. Ren and L.-k. Gong, *Acta Pharmacol. Sin.*, 2018, 39, 1865–1873.
- 146 Z. Wang, J. Ma, Y. He, K. K. Miu, S. Yao, C. Tang, Y. Ye and G. Lin, *Phytomedicine*, 2022, **102**, 154162.
- 147 Q. Wang, G.-C. Song, F.-Y. Weng, B. Zou, J.-Y. Jin, D.-M. Yan, B. Tan, J. Zhao, Y. Li and F.-R. Qiu, *Front. Pharmacol*, 2022, **13**, 881231.
- 148 Z. Wang, J. Ma, S. Yao, Y. He, K.-K. Miu, Q. Xia, P. P. Fu, Y. Ye and G. Lin, *Front. Pharmacol*, 2022, **13**, 850859.
- 149 S. Wu, H. Lu, W. Wang, L. Song, M. Liu, Y. Cao, X. Qi, J. Sun and L. Gong, *Cell Death Dis.*, 2021, **12**, 480.
- 150 T. Moro, Y. Shimoyama, M. Kushida, Y. Y. Hong, S. Nakao, R. Higashiyama, Y. Sugioka, H. Inoue, I. Okazaki and Y. Inagaki, *Life Sci.*, 2008, 83, 531–539.
- 151 M. Ablise, B. Leininger-Muller, C. D. Wong, G. Siest, V. Loppinet and S. Visvikis, *Chem. Pharm. Bull.*, 2004, 52, 1436–1439.
- 152 D. Maitraie, C.-F. Hung, H.-Y. Tu, Y.-T. Liou, B.-L. Wei, S.-C. Yang, J.-P. Wang and C.-N. Lin, *Bioorg. Med. Chem.*, 2009, **17**, 2785–2792.
- 153 Q. Zhang, E. A. H. Mohammed, Y. Wang, Z. Bai, Q. Zhao, D. He and Z. Wang, *Bioorg. Chem.*, 2020, **99**, 103804.
- 154 L. Zou, Q. Li, Y. Hou, M. Chen, X. Xu, H. Wu, Z. Sun and G. Ma, *Food Funct.*, 2022, **13**, 12487–12509.