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Some pyrimidohexahydroquinoline candidates: synthesis, DFT, cytotoxic activity evaluation, molecular docking, and *in silico* studies†

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Some hexahydroquinoline candidates were prepared by reacting 2-amino-3-cyano-1-cyclohexylhexahydroquinoline with oxalyl chloride and triethyl orthoformate. The computational chemical approach agreed with the product-testing results. The produced substances were examined *in vitro* for their antiproliferative activity against liver carcinoma (HepG2), breast adenocarcinoma (MCF7), prostate cancer (PC3), and colon cancer (HCT116) cell lines. The highest potency against the four cell lines was exhibited by hydrazide, thiosemicarbazide, and thiazolidinone derivatives. The best docking score was presented by thiosemicarbazide and thiazolidinone derivatives as they showed the highest binding to the Mcl-1 enzyme with binding energies of -8.97 and -8.90 kcal mol⁻¹, respectively, which were higher than that of the co-crystallized ligand (LC3) with a binding energy of -8.74 kcal mol⁻¹. Besides, the modeling pharmacokinetics disclosed their desirable drug-likeness and oral bioavailability characteristics.

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Introduction

Pyrimidoquinolines have piqued the consequence of researchers due to their potential pharmaceutical applications.^{1–3} Pyrimidoquinolines and quinolines have demonstrated antimicrobial,⁴ antitumor,^{5,6} antiviral,⁷ and anti-inflammatory properties.⁸ Diverse quinoline structures are effective inhibitors of major microbial pathogen proteins. Some quinolines with diverse biological effects are depicted in Fig. 1.^{9–21} Therefore, this work describes the design and preparation of certain pyrimidohexahydroquinolines with potential antiproliferative effects, reinforced by computational chemical approach and molecular docking, which might serve as a drug development target.

Rationale and design

Quinolines have been recognized as fundamental scaffolds owing to their existence in various pharmacologically potent compounds. In our case, to develop some antiproliferative substances, inspiration was drawn from quinoline's pharmacophoric features, which has exhibited efficacy in treating many diseases (*cf.* Fig. 2). Initially, using the nitro substituent in the

synthesized compounds is attributed to the significant effect of the electron-withdrawing group, which improves the ability to inhibit cancer cell proliferation.^{22,23} This electron-withdrawing head plays a substantial role in the binding interactions with the receptor subsites *via* van der Waals interactions and hydrogen bonding.¹⁰ To design a new series of some hexahydroquinoline derivatives, fundamental pharmacophoric features of quinoline were retained while integrating heterocyclic and side chain moieties as follows: (i) a planar aromatic core (chromophore) associated with the active sites of enzymes was retained in their design, which displayed pi-hydrophobic or pi-pi interactions with amino acid residues; (ii) the linker was altered by adding heteroatoms like nitrogen (N), oxygen (O), and sulfur (S) to demonstrate more hydrogen bonding through interactions with tumor proteins; (iii) to enhance the anti-proliferative potential, cores of pyrimidine, pyridine, indoline, thiazolidine, and phenyl were included in the design. With these features, this work intended to create some new hexahydroquinoline derivatives with enhanced antiproliferative properties.

Results and discussion

Chemistry

A one-pot cyclo-condensation treating cyclohexanone **1** with 2-(3-nitrobenzylidene)-malononitrile **2** and cyclohexylamine **3** in boiling ethanol and triethylamine (Et₃N) yielded the hexahydroquinoline-enaminonitrile derivative **4** as yellow crystals (Scheme 1). The absorption bands for primary amino and

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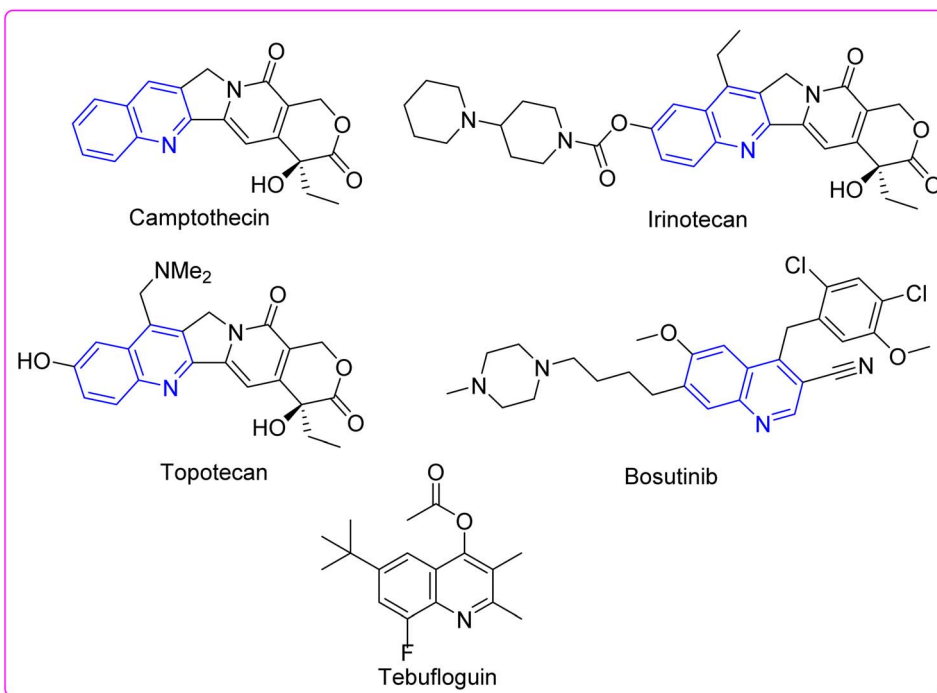



Fig. 1 Structures of some quinoline-containing drugs.

nitrile functionalities are illustrated in the IR spectra of **4**. Also, its ^1H NMR spectra showed exchangeable singlet signals for NH_2 protons. Furthermore, its mass spectrum displayed the molecular ion peak at m/z 378 (19%). A plausible mechanistic pathway for the formation of β -enaminonitrile **4** could be visualized to occur *via* Scheme 2. The initial step might be the formation of the carbanion of cyclohexanone **1** using triethylamine as a base, which subsequently attacked the β -carbon of the activated arylidene malononitrile **2** to give the Michael adduct, followed by condensation with cyclohexylamine **3** to eliminate a water molecule, which underwent 1,6-*exo*-dig cyclization, followed by imine-amine tautomerism, to be more stabilized by conjugation.

DFT simulation of the one-pot cyclocondensation reaction for the synthesis of the enaminonitrile **4** could support the plausible mechanistic pathway, as depicted in Fig. 3, which outlines the structure optimization of the intermediate that reacted to yield the final product. The computed parameters of the reacting species can be found in Table 1. The smaller energy gap between the HOMO and LUMO was found for arylidene **2**, which showed good chemical reactivity.

Also, the initial step of the reaction pathway could be interpreted from the charge density calculations. It was noted that charge density on the α -carbon of cyclohexanone **1** and the β -carbon of arylidene **2** was -0.101 and $+0.146$, respectively, which enhanced Michael addition under basic conditions (Et_3N). In turn, the charge density on the amino group of cyclohexylamine **3** was -0.294 , which facilitated the

nucleophilic attack on the carbonyl-carbon (with charge density of $+0.499$) of the adduct obtained from the first step rather than the nitrile-carbon (with a charge density of $+0.374$).

Boiling the β -enaminonitrile **4** with triethyl orthoformate yielded the condensation product **5**. The NH_2 absorption was absent in the IR spectrum of **5**. Further, its ^1H NMR spectrum was devoid of the NH_2 singlet signal and displayed quartet and triplet signals for the $-\text{CH}_2\text{CH}_3$ group. In turn, treating **4** with oxalyl chloride in benzene and Et_3N at ambient temperature yielded the pyrimidoquinoline derivative **6** (*cf.* Scheme 1). In the IR spectrum of **6**, the NH_2 and CN absorptions were absent while it displayed the amide and acid chloride carbonyl absorption bands at 1675 and 1779 cm^{-1} , respectively. Besides, its ^1H NMR spectrum demonstrated an exchangeable singlet signal for the NH proton.

The plausible mechanistic pathway for the condensation of enaminonitrile **4** with oxalyl chloride could be described in Scheme 3. The initial step was the nucleophilic attack of the primary amino group in compound **4** on the carbonyl-carbon of oxalyl chloride to eliminate hydrogen chloride molecule *via* the tetrahedral mechanism, which was tautomerized into the lactim form to undergo 1,6-*exo*-dig cyclization, giving the intermediate **[X]**, which subsequently rearranged into pyrimidine skeleton **6**. The preferential formation of the structure of compound **6** may be interpreted by considering the stability of products. By constructing space models for pyrimidine **6** and oxazine **[X]** isomers using ChemBio3D Ultra 2014, it was found that the pyrimidine skeleton **6** is thermodynamically more



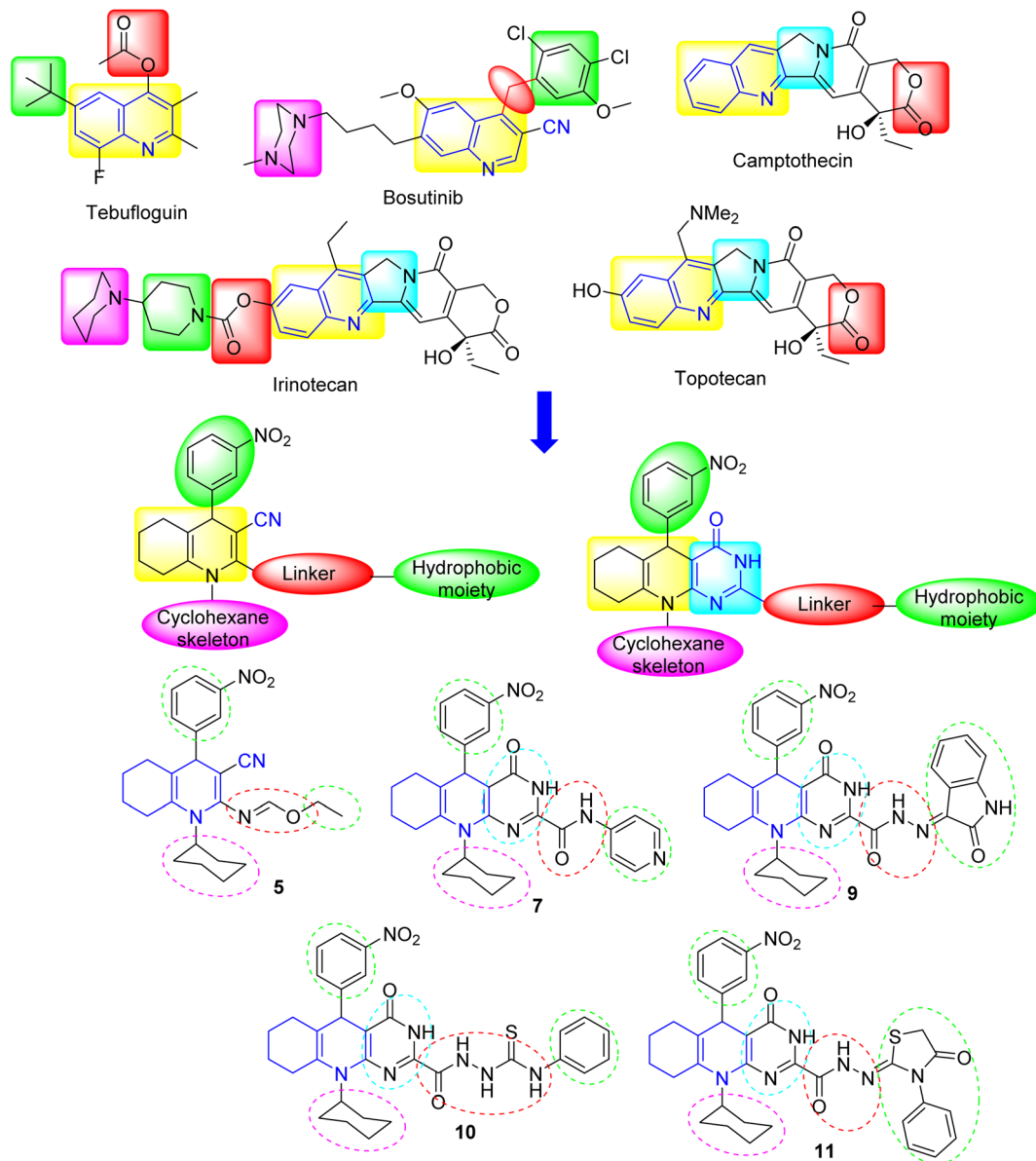


Fig. 2 Rationale and design of the structures of some anticancer agents (including quinoline core) and the target substances.

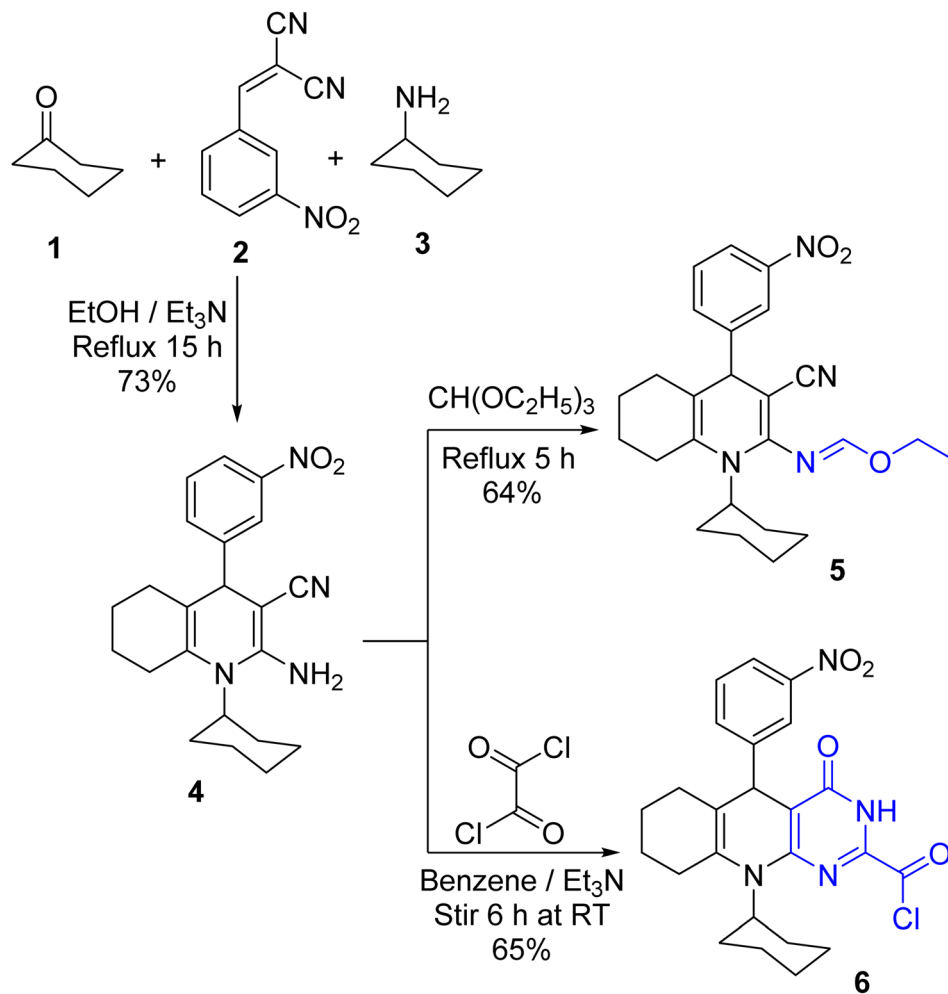
stable than the oxazine one [X] (cf. Fig. 4) according to MM2 calculations. The total energy of the isomer [X] is $52.8448 \text{ kcal mol}^{-1}$ with dipole/dipole of 10.1166, while that of pyrimidine 6 is $43.3355 \text{ kcal mol}^{-1}$ with dipole/dipole of 4.3151.

Reacting acid chloride 6 with 4-aminopyridine in benzene and Et_3N furnished the carboxamide derivative 7 (Scheme 4). The absorption of acid chloride carbonyl was absent in the IR spectrum of 7 but it unveiled the absorption of amide carbonyl. Further, its ^1H NMR spectrum established two exchangeable singlet signals for two NH protons. Analogously, hydrazide 8 was produced on treating 6 with hydrazine hydrate (80%) while

stirring in benzene at ambient temperature. The absorption bands for $\text{C}=\text{O}$, NH, and NH_2 groups are visible in the IR spectrum of hydrazide 8. Besides, its ^1H NMR spectrum indicated three exchangeable singlet signals for the NH of pyrimidine, NH of hydrazide, and NH_2 protons. Additionally, its mass spectrum compellingly proves structure 8 as it uncovered the molecular ion peak at m/z 464 with 23% intensity.

The condensation of hydrazide 8 with indolin-2,3-dione in boiling 1,4-dioxane gave hydrazone derivative 9 (Scheme 5). The IR chart of 9 unveiled the carbonyl absorption of indolinone core but no NH_2 absorption. Reacting phenyl isothiocyanate with hydrazide 8 in 1,4-dioxane achieved thiosemicarbazide





Scheme 1 Synthesis of β -enaminonitrile **4** and its reactions with triethyl orthoformate and oxalyl chloride.

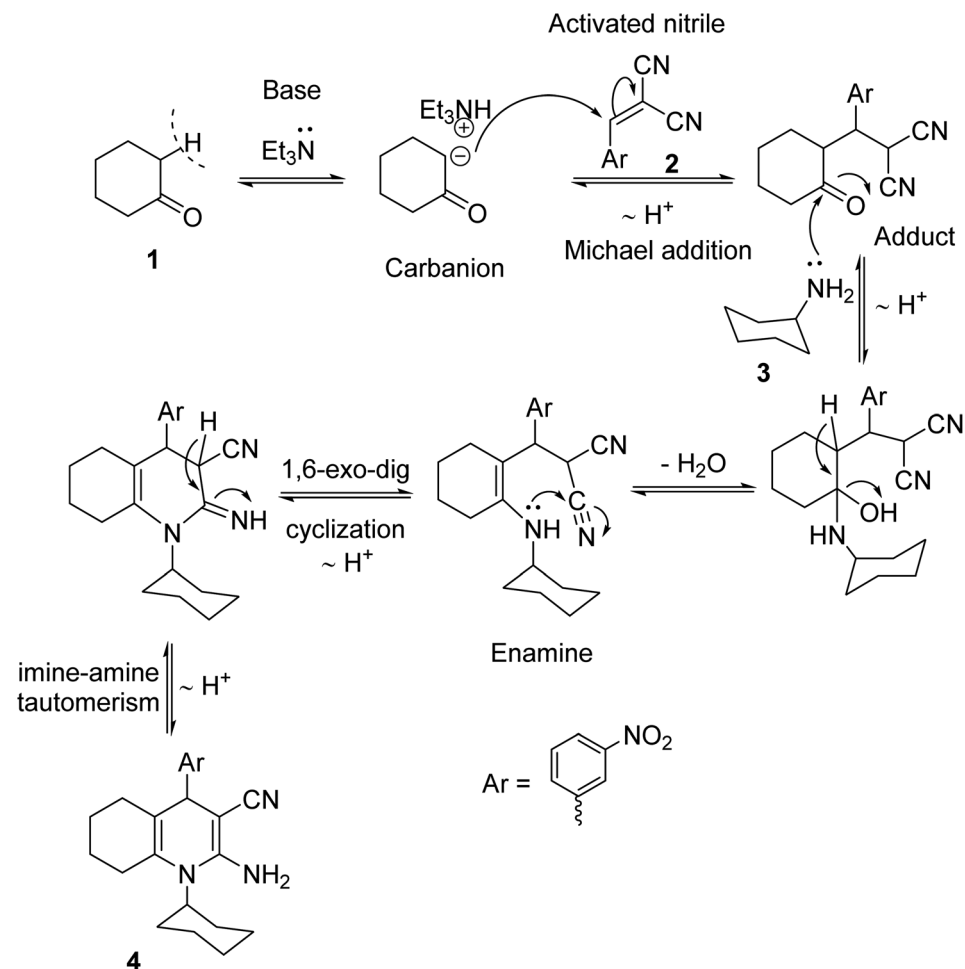
derivative **10**. Quite a lot of pyrimidoquinoline C-nucleosides⁴ and thiazolidinone C-nucleosides²⁴ demonstrate substantial antimicrobial and anticancer effects. These substances may impede the action of receptor-growth enzymes, like protein tyrosine kinase growth enzyme, in living cells.²⁵ As a result, combining the pyrimidoquinoline backbone with the thiazolidine-C-acyclic nucleoside core in a single molecule might increase the growth-inhibitory effect of these substances. Thiazolidinone derivative **11** was produced on treating **10** with chloroacetic acid in 1,4-dioxane (Scheme 5). Thiazolidinone carbonyl absorption appeared in IR chart **11**. Beyond that, a singlet signal for the CH₂ of the thiazolidine core was demonstrated in its ¹H NMR chart.

Density functional theory (DFT) study

The optimization of molecular structures for the prepared substances was run with the DFT approach applying the hybrid B3LYP functional with the 3-21G basis set. DFT based

on quantum chemical computation describes the structure optimization of the intermediate that reacted, furnishing the final product. The molecular structures of the substances are superior for forming a stable geometry, which is obtained through optimization. The structure's energy was reduced to a stationary point, the shape of tetrahydroquinoline derivatives was gradually optimized, and their energy was continuously reduced until the fluctuations in the molecule's energy were minimized.

The electrophilic-attacking locations are characterized by regions of highest electron density (HOMO), while the LUMO imitates the nucleophile-attacked locations. The HOMO and LUMO play strong roles in the electronic studies by quantum chemical calculations. Together, both are considered as vital tools in the computational investigations of the physicochemical, pharmacokinetic, and toxicological properties.^{26,27} The HOMO acts as an electron-donor, and the LUMO acts as an electron-acceptor.²⁸ A molecule with a high E_{HOMO} is likely to imply the strong tendency of a molecule to shift electrons.



Scheme 2 A plausible mechanism for the formation of enaminonitrile 4.

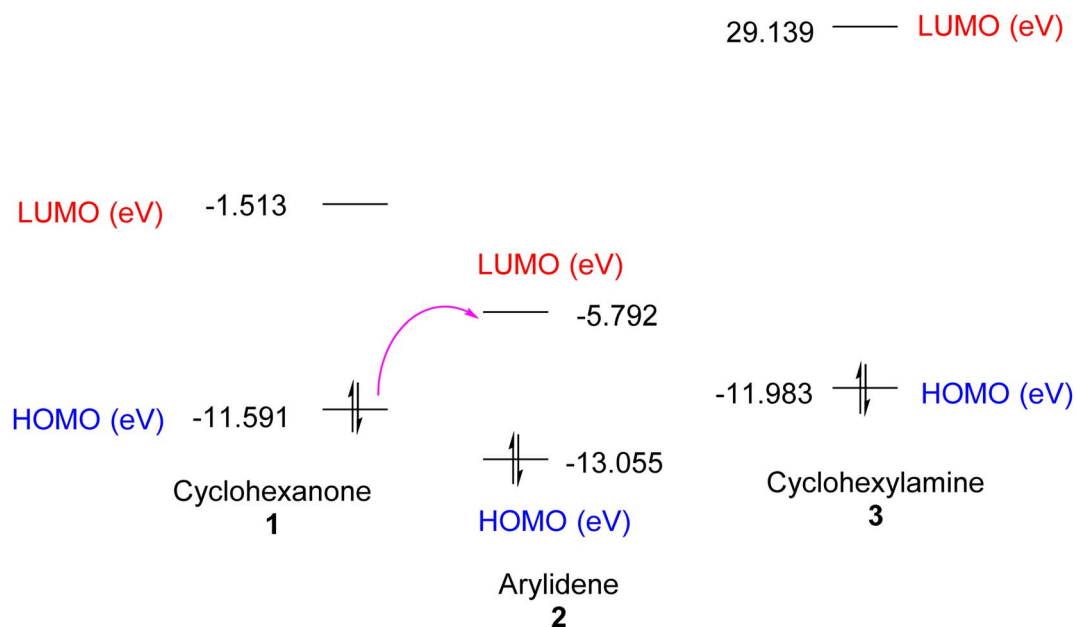
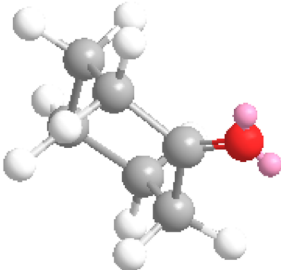
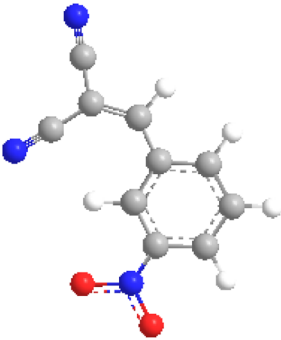
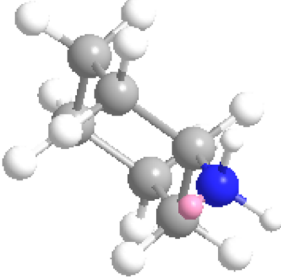
Fig. 3 E_{HOMO} and E_{LUMO} of reactant materials 1–3.

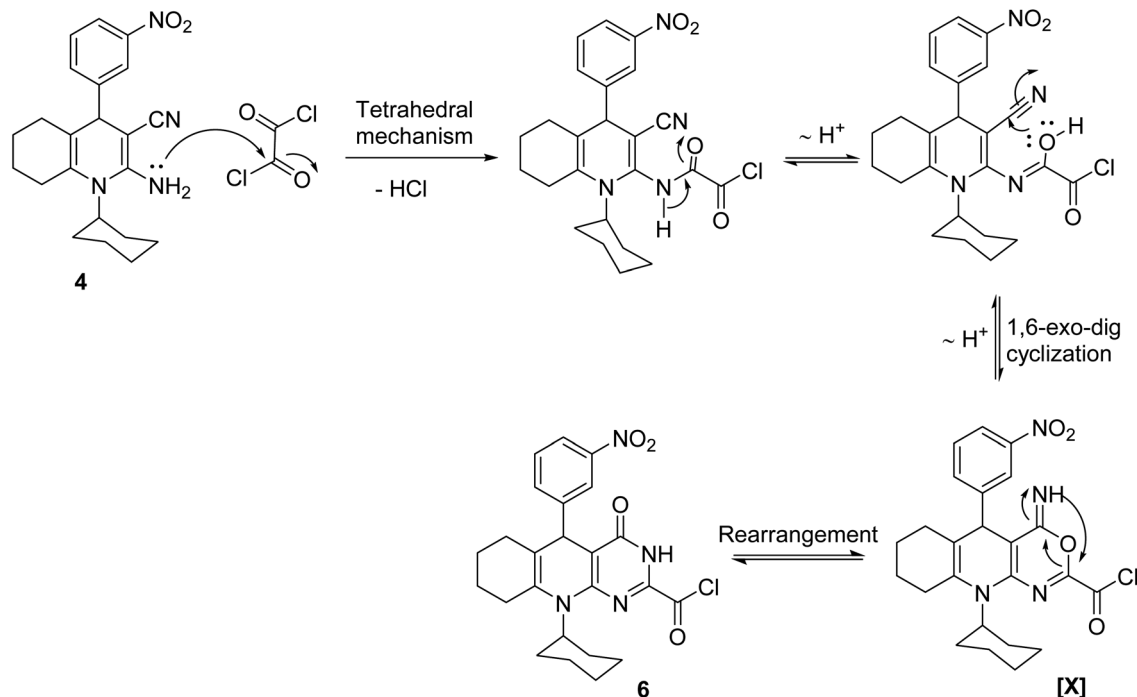
Table 1 Parameters of the reacting components to produce β -enaminonitrile 4

Reacting components	Stretch-bend	Torsion	Non-1,4 VDW	1,4 VDW	Dipole/dipole	Total energy (kcal mol ⁻¹)
Cyclohexanone 1 	0.059	3.839	-1.248	4.486	-1.711	6.414
Arylidene 2 	0.071	-9.690	2.602	11.225	7.561	10.648
Cyclohexylamine 3 	0.095	1.841	-1.490	4.709	0.0	5.930

A molecule with a smaller energy gap showed excellent chemical reactivity and such a molecule is designated as softer; meanwhile, a molecule with a larger energy gap exhibited good stability with high chemical hardness.^{29,30} Because the energy required to eliminate an electron from the last occupied orbital is small, the low values of the energy gap ($\Delta E = E_{\text{LUMO}} - E_{\text{HOMO}}$) will lead to good inhibition efficiency.^{18,21} Utilizing the E_{HOMO} and E_{LUMO} of the synthesized compounds, we can compute such global reactivity descriptors as energy gap, ionization energy, electron affinity, chemical hardness, chemical softness, chemical potential, electrophilicity, and electronegativity.³¹

Thus, DFT simulation assisted us in demonstrating the reactions of β -enaminonitrile derivative 4 with various reagents to generate substrates 5–11, and the computed parameters are shown in Table 2. The structures were endorsed by spectral and analytical data. The dipole moments for the tetrahydroquinoline derivatives were established with a decent explanation for the substrates. Fig. 5 depicts the optimized shapes and the HOMOs and LUMOs of compounds 4–11. In compounds 4 and 5, the HOMOs are distributed over the dihydropyridine unit. In compounds 6–10, the HOMOs are spread over the pyrimidinone unit. In compound 11, the HOMO is spread over the thiazolidinone unit. However, in all





Scheme 3 A probable mechanism for the condensation of **4** with oxalyl chloride.

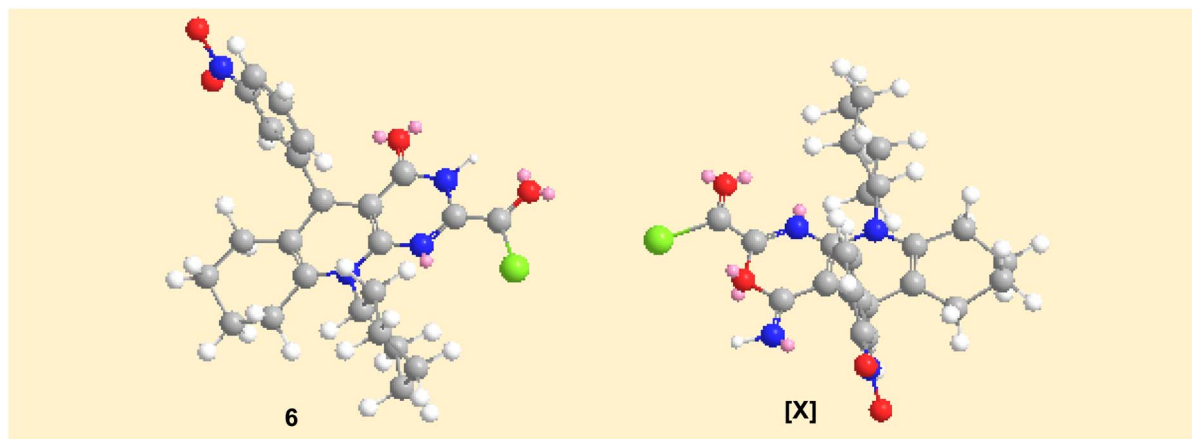


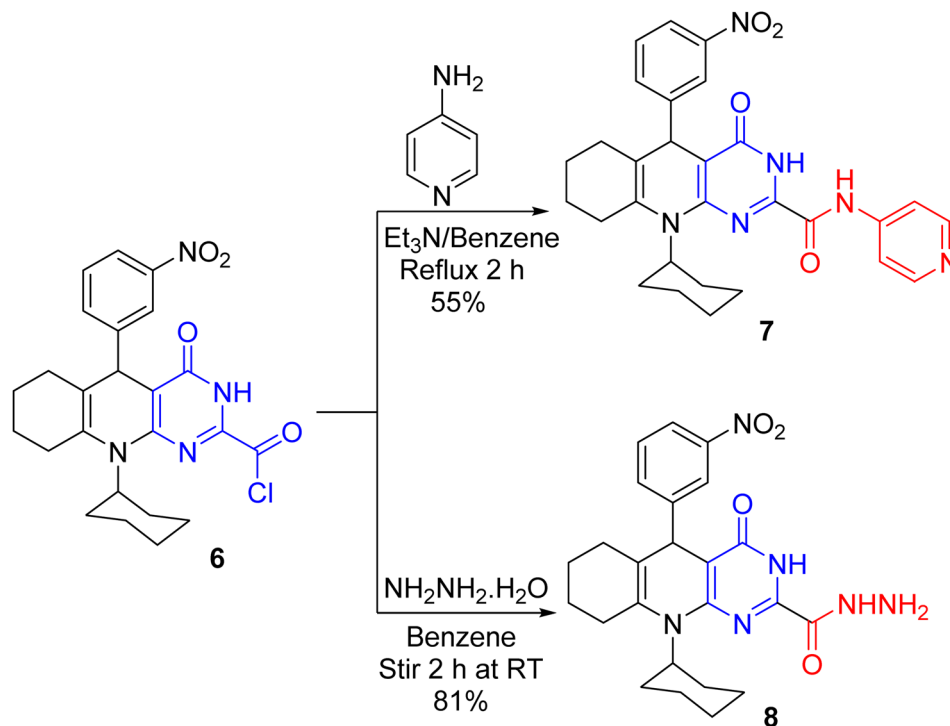
Fig. 4 Space models of both compounds obtained **6** and the intermediate **[X]**.

the compounds, the LUMOs are localized on the nitrophenyl moiety.

With ΔE as a criterion, three most typical and popular exchange-correlation functionals like PW91 were systematically compared in terms of the typical synthesis in the gas phase upon reactions of enaminonitrile **4** with triethyl orthoformate, oxalyl chloride, 4-aminopyridine, hydrazine hydrate, indolin-2,3-dione, and phenyl isothiocyanate. In addition, for the

further screening of proper functionalization, the verification of geometrical and electronic features of modeling anticancer products as well as the adsorption behaviors of typical probing reactants and solvents are suggested. This work provides general implications for how to choose a valid exchange-correlation functional in the computational solvents and catalyst on the reactant surface.





Scheme 4 Treating acid chloride **6** with 4-aminopyridine and hydrazine hydrate.

Quantum chemical parameters' calculations of the synthesized substrates are in good agreement with the anti-cancer efficiency (Table 2). The results indicated that the values of gap energy (ΔE) follow the sequence: heterocyclic derivatives **11** < **9** < doxorubicin. Substrates with small ΔE values are generally denoted as soft substrates, which are more reactive towards radical surface interactions, being capable of donating electrons simply to the hole surface. The scavenging ability toward positive hole, radical, tumor, and removable oxygen depend not only upon the E_{HOMO} values but, in addition, the heteroatoms number, surface area, electron distributions, and lipophilicity must be contemplated. The dipole moment (μ , debye) and softness (σ , eV^{-1}) for the most effective substances carrying hydrophobic groups showed decent correlation with the oxidation inhibition effectiveness. Also, the substrates with higher binding energy are of higher influence due to the strong interaction between these substrates and the receptor's active sites.

Cytotoxic activity

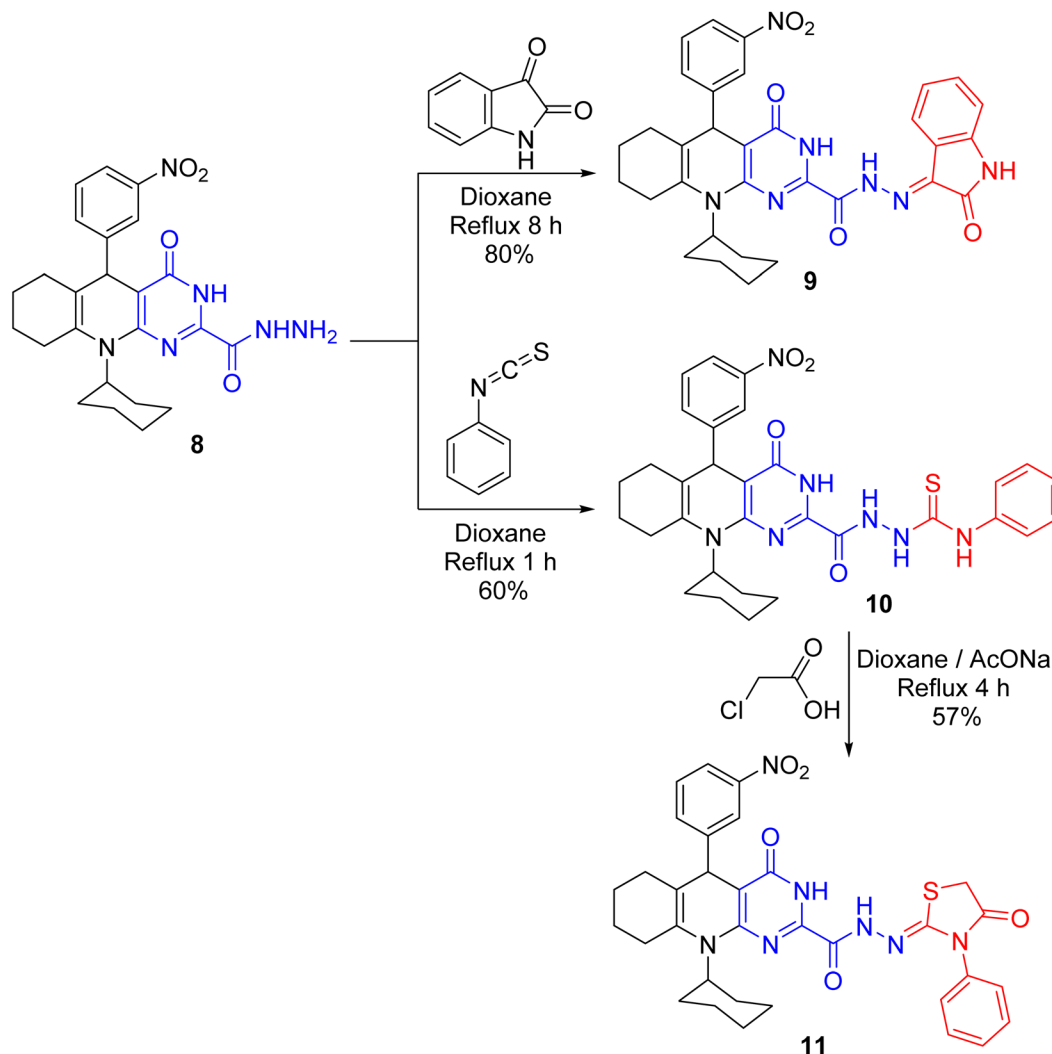
The cytotoxic activity of substrates was examined against four tumor cell lines, namely, liver carcinoma (HepG2), breast adenocarcinoma (MCF7), prostate cancer (PC3), and colon cancer (HCT116) cell lines, employing doxorubicin as

a standard agent. The synthesized compounds are water insoluble, and they were dissolved in a mixture of water and DMSO for the biological study, and they are quite stable as they do not have cleavable bonds. The cytotoxic activity was quantified as IC_{50} values, as described in Table 3, applying the MTT assay.³² The results illustrate that the tested substrates unveiled varying inhibitory potency levels. The highest potency against the four cell lines was presented by the following substances: hydrazide **8** (IC_{50} = 8.85, 11.44, 17.39, 16.21 μM), thiosemicarbazide derivative **10** (IC_{50} = 7.20, 5.67, 12.46, 7.34 μM), and thiazolidinone derivative **11** (IC_{50} = 9.67, 8.32, 10.22, 7.85 μM). Other compounds demonstrated moderate to weak cytotoxic effects.

Structure–activity relationship (SAR)

The inhibitory action of the tested substances could be correlated to structure variation and modifications compared with the doxorubicin reference. The structure of the parent substrate **4** is based on the presence of hexahydroquinoline core, which is an important scaffold of many anticancer drugs. The cytotoxic activity of the parent substrate **4** was moderate compared to the reference drug doxorubicin. The produced hexahydroquinolines with a sided electron-withdrawing head play a substantial role in binding





Scheme 5 Reaction of hydrazide 8 with indolin-2,3-dione and phenyl isothiocyanate.

Table 2 Energy level distribution and global reactivity indices of frontier orbitals^a

Compd	*E	E _{HOMO}	E _{LUMO}	ΔE	μ	η	ζ	μ _o	ω	n	I _p	EA	x
4	30.87	-7.669	-5.414	2.25	-2.90	1.12	0.89	-6.53	-23.87	-0.041	7.66	5.41	6.541
5	63.02	-7.705	-5.420	2.28	-1.07	1.14	0.87	-6.56	-24.52	-0.040	7.70	5.42	6.562
6	44.29	-7.391	-5.415	1.98	4.32	0.99	1.01	-6.40	-20.27	-0.049	7.39	5.41	6.403
7	29.88	-7.330	-5.415	1.92	-15.77	0.96	1.04	-6.37	-19.47	-0.051	7.33	5.41	6.372
8	24.19	-7.043	-5.413	1.63	-13.08	0.81	1.23	-6.22	-15.66	-0.063	7.04	5.41	6.228
9	29.25	-6.952	-5.411	1.54	-13.02	0.77	1.29	-6.18	-14.70	-0.068	6.95	5.41	6.181
10	22.04	-7.174	-5.417	1.76	-12.77	0.88	1.13	-6.29	-17.40	-0.057	7.17	5.41	6.295
11	47.52	-6.614	-5.408	1.21	-13.69	0.60	1.66	-6.00	-10.80	-0.092	6.61	5.40	6.011
Dox.	65.21	-6.69	-2.41	4.28	6.41	0.66	1.98	5.28	31.09	0.56	6.88	3.62	4.55

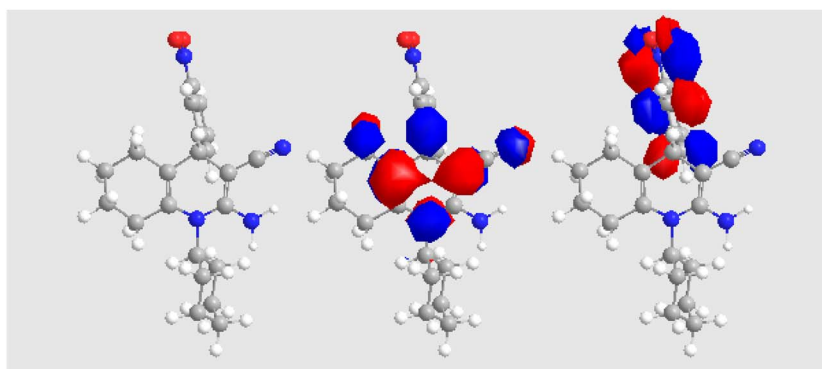
^a Dox.: doxorubicin, *E: minimized energy (kcal mol⁻¹), μ: dipole moment, η: global hardness, ζ: global softness, μ_o: chemical potential, ω: global electrophilicity index, n: nucleophilicity index, I_p: ionization potential, EA: electron affinity, x: electronegativity.

interactions with receptor subsites *via* van der Waals interactions and hydrogen bonding and promote pi-stacking interactions of the loop C aromatic residue with the side chain. The existence of an aromatic core in these compounds increased the hydrophobicity, which is essential for binding

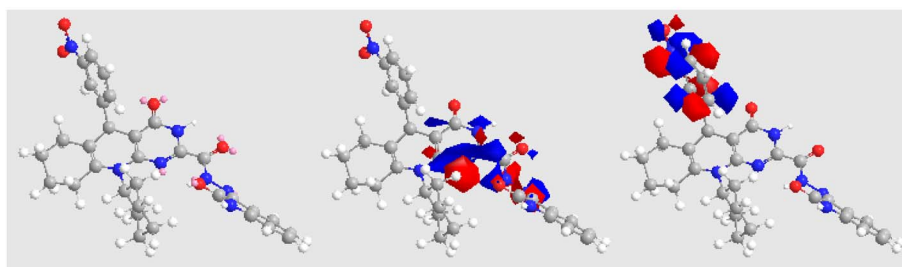
with the active site of many cellular enzymes and improves their permeability into the cell membrane, therefore developing the antiproliferative activity.

Besides, the analogs with extended conjugation had higher affinity to generate a face-to-edge aromatic interaction with the

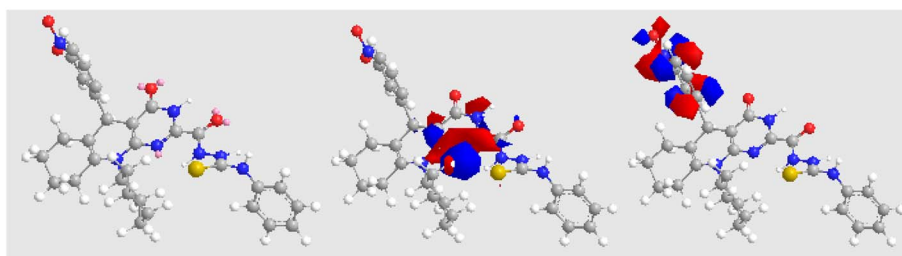




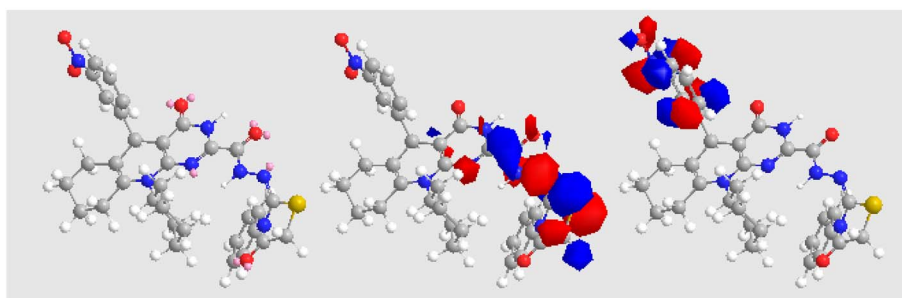
Compound 4



Compound 9



Compound 10



Compound 11

Fig. 5 Optimized configurations (left), HOMO (middle), and LUMO (right) for substrates 4–11. Atom color index: white H, grey C, blue N, red O, and green Cl (see all substrates in the ESI†).

receptor. Also, hydrazine and hydrazone moieties were important for bioactivity *via* azo-reduction into toxic amine derivatives. The hydrazone functionality is known for its antitumor properties.^{21,28} The introduction of another heterocyclic ring

like pyrimidine, pyridine, indole, and thiazolidine rings changed the cytotoxic properties towards the cell lines. These heterocycles show many essential features as anticancer agents. Consequently, it was revealed that substituting the chlorine



Table 3 Cytotoxic activity of the tested substances against the cancer cell lines

Compound	<i>In vitro</i> cytotoxicity IC ₅₀ ^a (μM) ± S.D.			
	HepG2	MCF7	PC3	HCT116
Doxorubicin	4.50 ± 0.3	4.17 ± 0.3	8.87 ± 0.5	5.23 ± 0.3
4	25.34 ± 3.9	28.19 ± 4.3	24.46 ± 4.2	35.10 ± 3.1
5	61.85 ± 3.2	73.15 ± 3.9	57.11 ± 3.4	48.20 ± 2.1
6	62.10 ± 4.5	75.24 ± 4.8	66.01 ± 4.2	58.07 ± 3.8
7	37.30 ± 1.8	49.57 ± 1.9	56.30 ± 2.9	27.99 ± 1.4
8	8.85 ± 3.5	11.44 ± 3.1	17.39 ± 3.7	16.21 ± 2.8
9	33.12 ± 2.1	45.44 ± 3.5	51.91 ± 3.9	37.63 ± 1.9
10	7.20 ± 0.5	5.67 ± 0.3	12.46 ± 1.0	7.34 ± 0.6
11	9.67 ± 2.9	8.32 ± 2.8	10.22 ± 2.4	7.85 ± 2.2

^a IC₅₀ (μM): 1–10 (very strong), 11–20 (strong), 21–50 (moderate), 51–100 (weak) and above 100 (non-cytotoxic). S.D.: standard deviation of *n* = 3.

atom in acid chloride **6** with hydrazinyl moiety (as in compound **8**) increased the inhibitory effect against the cell lines owing to hydrogen bonding with the receptor. The introduction of indolin-2-one (as in compound **9**) reduced its biological profile. The addition of a thiourea side chain (as in compound **10**) boosted the efficacy *via* extra hydrogen bonding with the receptor amino acids. The cyclization of the side chain to the thiazolidinone ring (as in compound **11**) is strongly superior to the cytotoxic potency (*cf.* Fig. 6).

Molecular docking studies

To grasp the pattern by which the examined compounds are bound to the active place, the most functioning candidates (**8**, **10**, and **11**) were subjected to the docking process into the Mcl-1 enzyme downloaded from the protein data bank (PDB code: 3WIX) at a resolution of 1.90 Å. Mcl-1 protein is a crucial

regulator of apoptosis; therefore, inhibitors of this protein could serve as promising new anticancer drugs. To design Mcl-1 inhibitors, we performed structure-guided analyses of the corresponding selective inhibitors. Firstly, water molecules were removed from the downloaded complex. The structures of the compounds and reference ligand were drawn using ChemBio3D Ultra 14.0.

The affinities of the most active synthesized ligands (**8**, **10**, and **11**) toward the target protein (Tables 4–6) were compared according to the docking score values calculated employing MOE 2019.0102.³³ The best docking score was presented by substances **10** and **11** as they unveiled the highest binding to the Mcl-1 enzyme with a binding energy of -8.97 kcal mol⁻¹ with an RMSD of 1.41 Å and -8.90 kcal mol⁻¹ with an RMSD of 1.99 Å, respectively, which were higher than the co-crystallized ligand (LC3) with a binding energy of -8.74 kcal mol⁻¹ with an RMSD of 1.19 Å. The leading interactions of the co-crystallized ligand with Mcl-1 pockets were almost like that of the most effective substrates (**8**, **10**, and **11**), for example, hydrogen bonding with MET 231 and ARG 263 receptors.

Modeling pharmacokinetics studies

To select substrates from a huge collection of prepared substances in the early stages of drug discovery, biological actions, and development for an effective drug, ADME profiles including physicochemical properties, lipophilicity, and drug likeness of the synthesized substances have been predicted.^{34–36} Substance **8**, satisfying Lipinski's rule of five, demonstrates a total polar surface area (TPSA) of 149.93 Å and respectable lipophilicity, expressed by the consensus Log *P*_{o/w}, which is 2.23. As seen in Table 7, it demonstrated a high GI absorption and was computed to acquire a good bioavailability score (0.55).

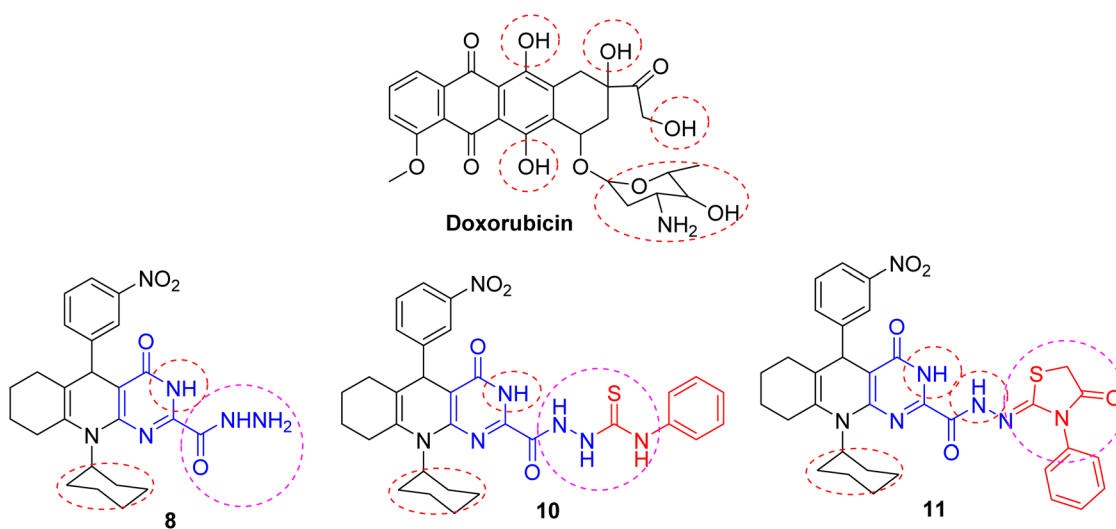


Fig. 6 SAR of the most potent substances.



Table 4 2D and 3D interactions of compounds (8, 10, and 11) with Mcl-1 binding pockets

Compound	2D-interaction	3D-interaction
8		
10		
11		
Co-crystallized ligand (LC3)		

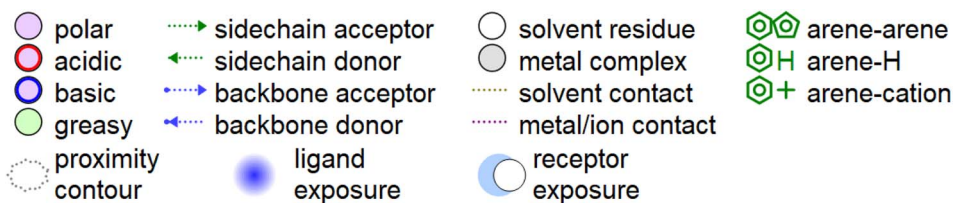


Table 5 Ligand interactions of compounds (8, 10, and 11) with the Mcl-1 enzyme

Compound	Ligand	Receptor	Type of interaction	Interaction distance (Å)	<i>E</i> (kcal mol ⁻¹)
8	N 7	SD MET 231	H-donor	3.90	-1.8
	O 23	NE2 HIS 224	H-acceptor	3.08	-2.5
10	O 1	NE ARG 263	H-acceptor	2.95	-5.6
	S 25	CA MET 250	H-acceptor	4.08	-1.1
	C 65	5-Ring HIS 224	H-pi	4.34	-0.6
11	O 1	NE ARG 263	H-acceptor	3.02	-5.6
	O 1	NH1 ARG 263	H-acceptor	3.34	-0.6
	C 68	5-Ring HIS 224	H-pi	4.40	-0.6
	6-Ring	CE MET 250	pi-H	3.79	-1.0
Co-crystallized ligand (LC3)	O 1	NE ARG 263	H-acceptor	3.02	-4.2
	C 23	6-Ring PHE 270	H-pi	3.50	-0.7
	6-Ring	CE MET 250	pi-H	3.93	-0.8

Table 6 Docking score and RMSD of compounds (8, 10, and 11) with the Mcl-1 enzyme

Compound	Docking score (kcal mol ⁻¹)	RMSD (Å)
8	-6.67	1.35
10	-8.97	1.41
11	-8.90	1.99
Co-crystallized (LC3)	-8.74	1.19

Table 7 ADME characteristics of compounds 8, 10, and 11

Entry	Compounds		
	8	10	11
Physicochemical properties/lipophilicity/druglikeness			
Molecular weight (g mol ⁻¹)	464.52	599.70	639.72
Num. heavy atoms	34	43	46
Num. arom. heavy atoms	12	18	18
Fraction Csp ³	0.46	0.35	0.36
Num. rotatable bonds	5	9	7
Num. H-bond acceptor	6	5	7
Num. H-bond donors	3	4	2
Molar refractivity	132.43	173.94	185.36
TPSA (Å ²)	149.93	180.06	181.88
Consensus Log <i>P</i> _{o/w}	2.23	3.98	4.29
Lipinski's rule	Yes	No	No
Bioavailability score	0.55	0.17	0.17
Pharmacokinetics			
GI absorption	Low	Low	Low
BBB permeant	No	No	No
P-gp substrate	Yes	No	Yes
CYP1A2 inhibitor	No	Yes	Yes
CYP2C19 inhibitor	No	No	No
CYP2C9 inhibitor	Yes	Yes	Yes
CYP2D6 inhibitor	No	No	No
CYP3A4 inhibitor	Yes	Yes	Yes
Log <i>K</i> _p (cm s ⁻¹)	-7.10	-6.34	-6.18

Its skin permeation (Log *K*_p) parameter was -7.10, thus facilitating the accessibility of the bioactive molecules through the skin. Its cyto-chrome P450 isoenzymes (CYP1A2/CYP2C19/

CYP2C9/CYP2D6/CYP3A4), playing a major part in the biotransformation of drugs through O-type oxidation reactions, have been likewise anticipated (*cf.* Table 7). The bioavailability of the produced substances was also estimated based on their pink area on the radar chart (*cf.* Fig. S7-S9, *cf.* the ESI†). The compounds were completely included in the pink area and justified their good predicted oral bioavailability, especially compounds 8, 10, and 11.

Conclusion

Some pyrimidohexahydroquinolines were achieved by reacting the hexahydroquinoline-enaminonitrile scaffold with some carbon-centered electrophiles. The computational chemical results were in good agreement with the experimental products. The substates were screened for their *in vitro* antitumor activity against liver carcinoma (HepG2), breast adenocarcinoma (MCF7), prostate cancer (PC3), and colon cancer (HCT116), and unveiled that hydrazide, thiosemicarbazide, and thiazolidinone derivatives displayed strong effectiveness. The molecular docking study upheld the biological effects. *In silico* findings with physicochemical characteristics, lipophilicity, and drug-likeness of the potent substances have been estimated.

Experimental

General

Solvents and chemicals were purified and dried by standard techniques. Melting points (uncorrected) were assessed on a MEL-TEMP II electric melting point apparatus. The IR spectra (ν , cm⁻¹) were recorded at the Faculty of Science, Ain Shams University, applying KBr disks on a FTIR Thermo Electron Nicolet iS10 infrared spectrometer. ¹H NMR spectra (δ , ppm) were run at 300 MHz on a GEMINI NMR spectrometer employing tetramethyl silane (TMS) as an internal standard in deuterated DMSO. Mass spectra (70 eV, *m/z*, %) were documented at the Faculty of Pharmacy, Al-Azhar University, Egypt. Elemental analyses were done at the Faculty of Science, Ain Shams University, applying a PerkinElmer 2400 CHN elemental analyzer. The reactions were monitored by TLC on silica gel-



precoated aluminum sheets 60 F₂₅₄-Merck, and the spots were perceived by exposure to a UV lamp.

2-Amino-1-cyclohexyl-4-(3-nitrophenyl)-1,4,5,6,7,8-hexahydroquinoline-3-carbonitrile (4). A mixture of cyclohexanone **1** (0.98 g, 0.01 mol), 2-(3-nitrobenzylidene)malononitrile (**2**) (1.99 g, 0.01 mol), and cyclohexylamine **3** (0.99 g, 0.01 mol) in ethanol (30 mL) including triethylamine (0.1 mL) was refluxed for 15 h and left overnight at ambient temperature. The obtained precipitate was filtered off, washed with cold water and recrystallized from ethanol to furnish yellow crystals, mp. 95–97 °C, yield: 2.76 g (73%). IR (ν , cm⁻¹): 2220 (CN), 3346, 3310 (NH₂). ¹H NMR (DMSO, δ , ppm): 1.42–1.44 (m, 2H, CH₂, cyclohexyl), 1.45–1.48 (m, 4H, 2CH₂, cyclohexyl), 1.64–1.67 (m, 4H, 2CH₂, tetrahydroquinoline), 1.75–1.78 (m, 4H, 2CH₂, cyclohexyl), 1.96–1.99 (m, 4H, 2CH₂, tetrahydroquinoline), 2.54–2.58 (m, 1H, N-CH, cyclohexyl), 5.47 (s, 1H, CH tetrahydroquinoline), 6.15 (br.s, 2H, NH₂, exchangeable), 6.49–6.82 (m, 4H, Ar-H). EIMS (m/z , %): 378 (M⁺, 19). Anal. calcd for C₂₂H₂₆N₄O₂ (378.48): C, 69.82; H, 6.92; N, 14.80; found: C, 69.81; H, 6.94; N, 14.82%.

Ethyl-N-(3-cyano-1-cyclohexyl-4-(3-nitrophenyl)-1,4,5,6,7,8-hexahydroquinolin-2-yl)formimidate (5). A suspension of compound **4** (0.76 g, 0.002 mol) in triethyl orthoformate (5 mL) was refluxed for 5 h; then, the reaction mixture was poured onto ice/cold water. The precipitate was filtered off, dried, and crystallized from ethanol to achieve yellow crystals, mp. 80–82 °C, yield: 0.55 g (64%). IR (ν , cm⁻¹): 2214 (C≡N), 1640 (C=N). ¹H NMR (DMSO, δ , ppm): 1.20 (t, 3H, CH₃CH₂, $J = 6.5$ Hz), 1.41–1.44 (m, 2H, CH₂ cyclohexyl), 1.45–1.48 (m, 4H, 2CH₂ cyclohexyl), 1.65 (m, 4H, 2CH₂, tetrahydroquinoline), 1.76 (m, 4H, 2CH₂ cyclohexyl), 1.98 (m, 4H, 2CH₂, tetrahydroquinoline), 2.55 (m, 1H, CH cyclohexyl), 3.57 (q, 2H, CH₂CH₂, $J = 6.5$ Hz), 5.42 (s, 1H, CH tetrahydroquinoline), 6.80–7.30 (m, 4H, Ar-H), 8.50 (s, 1H, CH = N). EIMS (m/z , %): 434 (M⁺, 15). Anal. calcd for C₂₅H₃₀N₄O₃ (434.54): C, 69.10; H, 6.96; N, 12.89; found: C, 69.01; H, 6.90; N, 12.91%.

10-Cyclohexyl-5-(3-nitrophenyl)-4-oxo-3,4,5,6,7,8,9,10-octahydropyrimido[4,5-*b*]quinoline-2-carbonyl chloride (6). To a solution of **4** (3.78 g, 0.01 mol) in benzene (20 mL) including triethylamine (0.1 mL), oxalyl chloride (1.89 g, 0.015 mol) was added dropwise at ambient temperature with stirring. The reaction mixture was stirred for 6 h. The precipitate obtained was collected to give a brown solid, yield: 3.04 g (65%). mp 151–153 °C. IR (ν , cm⁻¹): 3352 (NH), 1779 (C=O acid chloride), 1675 (C=O amide). ¹H NMR (DMSO, δ , ppm): 1.35–1.38 (m, 4H, 2CH₂, cyclohexyl), 1.40–1.43 (m, 2H, CH₂, cyclohexyl), 1.51–1.53 (m, 4H, 2CH₂, cyclohexyl), 1.62–1.65 (m, 4H, 2CH₂, tetrahydroquinoline), 1.94–1.98 (m, 4H, 2CH₂, tetrahydroquinoline), 2.59–2.62 (m, 1H, N-CH, cyclohexyl), 5.43 (s, 1H, CH tetrahydroquinoline), 6.54–6.99 (m, 4H, Ar-H), 10.03 (br.s, 1H, NH, exchangeable). EIMS (m/z , %): 470 (M²⁺, 10), 468 (M⁺, 30). Anal. calcd for C₂₄H₂₅ClN₄O₄ (468.94): C, 61.47; H, 5.37; N, 11.95; found: C, 61.39; H, 5.32; N, 11.94%.

10-Cyclohexyl-5-(3-nitrophenyl)-4-oxo-N-(pyridin-4-yl)-3,4,5,6,7,8,9,10-octahydropyrimido[4,5-*b*]quinoline-2-carboxamide (7). A solution of **6** (4.69 g, 0.01 mol) and 4-aminopyridine (0.94 g, 0.01 mol) in benzene (20 mL) containing triethylamine (0.1 mL) was refluxed for 2 h. The reaction mixture was concentrated and then cooled. The solid obtained

was filtered off and crystallized from ethanol to furnish yellow crystals, mp. 100–102 °C, yield: 2.89 g (55%). IR (ν , cm⁻¹): 3355 (NH), 1665, 1643 (C=O). ¹H NMR (DMSO, δ , ppm): 1.37–1.40 (m, 4H, 2CH₂, cyclohexyl), 1.43–1.46 (m, 2H, CH₂, cyclohexyl), 1.52–1.55 (m, 4H, 2CH₂, cyclohexyl), 1.65–1.67 (m, 4H, 2CH₂, tetrahydroquinoline), 1.95–1.99 (m, 4H, 2CH₂, tetrahydroquinoline), 2.59–2.62 (m, 1H, N-CH, cyclohexyl), 5.40 (s, 1H, CH tetrahydroquinoline), 6.40–6.64 (m, 8H, Ar-H), 8.89 (br.s, 1H, NH, exchangeable), 9.79 (br.s, 1H, NH, exchangeable). EIMS (m/z , %): 526 (M⁺, 21). Anal. calcd for C₂₉H₃₀N₆O₄ (526.60): C, 66.15; H, 5.74; N, 15.96; found: C, 66.07; H, 5.69; N, 15.98%.

10-Cyclohexyl-5-(3-nitrophenyl)-4-oxo-3,4,5,6,7,8,9,10-octahydropyrimido[4,5-*b*]quinoline-2-carbohydrazide (8). To a solution of **6** (4.69 g, 0.01 mol) in benzene (20 mL), hydrazine hydrate (0.55 mL, 0.011 mmol, 98%) was added dropwise at ambient temperature with stirring. The reaction mixture was stirred at room temperature for 2 h. The solid obtained was collected and recrystallized from ethanol to give pale-yellow crystals, mp. 130–132 °C, yield: 3.75 g (81%). IR (ν , cm⁻¹): 1668, 1655 (C=O), 3354, 3325, 3245 (NH, NH₂). ¹H NMR (DMSO, δ , ppm): 1.42–1.45 (m, 4H, 2CH₂, cyclohexyl), 1.46–1.48 (m, 2H, CH₂, cyclohexyl), 1.52–1.55 (m, 4H, 2CH₂, cyclohexyl), 1.65–1.69 (m, 4H, 2CH₂, tetrahydroquinoline), 2.15–2.22 (m, 4H, 2CH₂, tetrahydroquinoline), 2.58–2.61 (m, 1H, N-CH, cyclohexyl), 5.44 (s, 1H, CH tetrahydroquinoline), 5.70 (br.s, 2H, NH₂, exchangeable), 6.57–7.06 (m, 4H, Ar-H), 8.58 (br.s, 1H, NH, exchangeable), 8.85 (br.s, 1H, NH, exchangeable). EIMS (m/z , %): 464 (M⁺, 23). Anal. calcd for C₂₄H₂₈N₆O₄ (464.53): C, 62.06; H, 6.08; N, 18.09; found: C, 61.97; H, 6.00; N, 18.10%.

10-Cyclohexyl-5-(3-nitrophenyl)-4-oxo-N-(2-oxindolin-3-ylidene)-3,4,5,6,7,8,9,10-octahydropyrimido[4,5-*b*]quinoline-2-carbohydrazide (9). A mixture of hydrazide **8** (4.64 g, 0.01 mol) and isatin (1.47 g, 0.01 mol) in dioxane (20 mL) was refluxed for 8 h. The solid obtained was separated by filtration and crystallized from dioxane to give red crystals, mp 179–181 °C, yield: 4.74 g (80%). IR (ν , cm⁻¹): 1665 (C=O pyrimidinone), 1704 (C=O indolinone), 3350, 3240 (NH). ¹H NMR (DMSO, δ , ppm): 1.36–1.43 (m, 4H, 2CH₂, cyclohexyl), 1.42–1.46 (m, 2H, CH₂, cyclohexyl), 1.52–1.56 (m, 4H, 2CH₂, cyclohexyl), 1.63–1.66 (m, 4H, 2CH₂, tetrahydroquinoline), 1.99–2.33 (m, 4H, 2CH₂, tetrahydroquinoline), 2.58–2.87 (m, 1H, N-CH, cyclohexyl), 5.45 (s, 1H, CH tetrahydroquinoline), 6.45 (br.s, 1H, NH indolinone, exchangeable), 6.55–7.58 (m, 8H, Ar-H), 8.59 (br.s, 1H, NH, exchangeable), 9.01 (br.s, 1H, NH, exchangeable). EIMS (m/z , %): 593 (M⁺, 25). Anal. calcd for C₃₂H₃₁N₇O₅ (593.64): C, 64.74; H, 5.26; N, 16.52; found: C, 64.68; H, 5.20; N, 15.50%.

2-(10-Cyclohexyl-5-(3-nitrophenyl)-4-oxo-3,4,5,6,7,8,9,10-octahydropyrimido[4,5-*b*]quinoline-2-carbonyl)-N-phenylhydrazine-1-carbothioamide (10). A mixture of hydrazide **8** (4.64 g, 0.01 mol) and phenyl isothiocyanate (1.35 g, 0.01 mol) in dioxane (20 mL) was refluxed for 1 h. The reaction mixture was evaporated under reduced pressure and the residue was triturated with ethanol. The solid obtained was collected by filtration, washed with ethanol, and recrystallized from ethanol to achieve brown crystals, mp. 140–142 °C, yield: 3.59 g (60%). IR (ν , cm⁻¹): 1258 (C=S), 1669, 1649 (C=O), 3351, 3240 (NH), 3182 (NH). ¹H NMR (DMSO, δ , ppm): 1.43–1.46 (m, 4H, 2CH₂,



cyclohexyl), 1.47–1.50 (m, 2H, CH₂, cyclohexyl), 1.53–1.57 (m, 4H, 2CH₂, cyclohexyl), 1.68–1.73 (m, 4H, 2CH₂, tetrahydroquinoline), 2.02–2.05 (m, 4H, 2CH₂, tetrahydroquinoline), 2.61–2.64 (m, 1H, N-CH, cyclohexyl), 5.44 (s, 1H, CH tetrahydroquinoline), 6.59–7.49 (m, 9H, Ar-H), 8.97 (br.s, 1H, NH, exchangeable), 9.33 (br.s, 1H, NH, exchangeable), 9.95 (br.s, 1H, NH, exchangeable), 10.10 (br.s, 1H, NH, exchangeable). EIMS (*m/z*, %): 599 (M⁺, 21). Anal. calcd for C₃₁H₃₃N₇O₄S (599.71): C, 62.09; H, 5.55; N, 16.35; found: C, 61.97; H, 5.50; N, 16.37%.

10-Cyclohexyl-5-(3-nitrophenyl)-4-oxo-N'-(4-oxo-3-phenylthiazolidin-2-ylidene)-3,4,5,6,7,8,9,10-octahydropyrimido[4,5-*b*]quinoline-2-carbohydrazide (11). A mixture of **10** (3.00 g, 0.005 mol), chloroacetic acid (0.47 g, 0.005 mol) and anhydrous sodium acetate (0.01 mmol) in dioxane (20 mL) was refluxed for 4 h. The solid formed was separated by filtration and crystallized from dioxane to produce brown crystals, mp. 161–163 °C, yield: 1.82 g (57%). IR (ν , cm⁻¹): 1675, 1655 (C=O amide), 1705 (C=O thiazolidinone), 3360, 3275 (NH). ¹H NMR (DMSO, δ , ppm): 1.38–1.43 (m, 4H, 2CH₂, cyclohexyl), 1.44–1.47 (m, 2H, CH₂, cyclohexyl), 1.53–1.57 (m, 4H, 2CH₂, cyclohexyl), 1.66–1.69 (m, 4H, 2CH₂, tetrahydroquinoline), 1.97–2.01 (m, 4H, 2CH₂, tetrahydroquinoline), 2.59–2.62 (m, 1H, N-CH, cyclohexyl), 3.79 (s, 2H, CH₂, thiazolidinone), 5.41 (s, 1H, CH tetrahydroquinoline), 6.59–7.23 (m, 9H, Ar-H), 8.39 (br.s, 1H, NH, exchangeable), 9.37 (br.s, 1H, NH, exchangeable). EIMS (*m/z*, %): 639 (M⁺, 21). Anal. calcd for C₃₃H₃₃N₇O₅S (639.73): C, 61.96; H, 5.20; N, 15.33; found: C, 61.87; H, 5.14; N, 15.30%.

Molecular docking

Molecular docking studies are considered an influential method for the interpretation of molecular interactions between the produced substances and the main amino acid residues at the specific binding site of the target receptor.^{37,38} The affinities of the produced ligands toward the target protein Mcl-1 were compared according to the docking score values calculated using the MOE 2014.0901.³³ The most active derivatives (**8**, **10**, and **11**) were drawn employing ChemBio3D Ultra 14.0 and transferred to the MOE window, optimized for partial charges, and energy minimized, as previously reported.³⁹ Then, the target protein (Mcl-1) was extracted from the PDB database (PDB ID: 3WIX⁴⁰), corrected, 3D hydrogenated, and energy minimized. Finally, a docking process was run to evaluate the binding affinities of candidates toward the Mcl-1 receptor pockets. The co-crystallized inhibitor (LC3) was inserted as the reference standard and the program specifications were adjusted, as previously mentioned.⁴¹

Data availability

Full tables, figures, and all data generated or analyzed during this study are included in this article and its ESI† files.

Conflicts of interest

The authors declare no conflicts of interest.

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