

Cite this: *Chem. Sci.*, 2024, 15, 11444

All publication charges for this article have been paid for by the Royal Society of Chemistry

Received 9th May 2024  
Accepted 17th June 2024

DOI: 10.1039/d4sc03054k

rsc.li/chemical-science

# Unlocking flavin photoacid catalysis through electrophotochemistry†

Samuel Gary,<sup>a</sup> Jack Woolley,<sup>b</sup> Sofia Goia<sup>c</sup> and Steven Bloom<sup>id</sup>\*<sup>a</sup>

Molecular flavins are one of the most versatile photocatalysts. They can coordinate single and multiple electron transfer processes, gift hydrogen atoms, form reversible covalent linkages that support group transfer mechanisms, and impart photonic energy to ground state molecules, priming them for downstream reactions. But one mechanism that has not featured extensively is the ability of flavins to act as photoacids. Herein, we disclose our proof-of-concept studies showing that electrophotochemistry can transform fully oxidized flavin quinones to super-oxidized flavinium photoacids that successfully guide proton-transfer and deliver acid-catalyzed products. We also show that these species can adopt a second mechanism wherein they react with water to release hydroxyl radicals that facilitate hydrogen-atom abstraction and  $\text{sp}^3\text{C-H}$  functionalization protocols. Together, this unprecedented bimodal reactivity enables electro-generated flavinium salts to affect synthetic chemistries previously unknown to flavins, greatly expanding their versatility as catalysts.

## Introduction

Flavins are Nature's multi-tools, choreographing a myriad of chemical reactions. Flavins catalyze single-<sup>1–3</sup> and multiple-electron transfer processes,<sup>4,5</sup> group transfer chemistries,<sup>6</sup> and other redox fluctuations<sup>7</sup> that orchestrate complex bimolecular synthesis and cellular crosstalk. Flavins also have rich photochemistry.<sup>1</sup> Depending on their redox state—quinone, semiquinone, or hydroquinone—flavins serve as powerful photo-oxidants and -reductants and can even confer their newly acquired photonic energy to ground state molecules, activating them for high-barrier transformations.<sup>8,9</sup> Many of these pathways have been successfully leveraged for modern organic synthesis with visible light.<sup>10–12</sup> A less explored pathway involves the use of flavins as photoacids. Protonation of the flavin quinone affords a flavinium salt,  $\text{FL}^+\text{ox}$ . In the excited state ( $\lambda_{\text{max}} = 394 \text{ nm}$ ), these species become 'strong' acids. Rarely do organic photoacids exceed  $\text{p}K_{\text{a}} < 0$ .<sup>13–18</sup> Schulman determined a  $\text{p}K_{\text{a}}^*$  for protonated riboflavin ( $\text{RF}^+\text{ox}$ ) of  $-6.2$ .<sup>19</sup> Liu *et al.* theorized that bis-protonation of the flavin quinone gives rise to an intermediate,  $\text{FLH}^+\text{ox}_{\text{N1/N5}}$ , with even greater acid-like character in the

ground state,  $\text{p}K_{\text{a}}$  values of 0 and  $-10.1$  at N1 and N5, respectively.<sup>20</sup> Photoexcitation increases the acidity of N1 to  $-4.9$  and shifts the basicity of N5 to  $+1.7$ , Fig. 1. Unlocking the unique photoacid reactivity of  $\text{FL}^+\text{ox}$  has been a significant challenge. In the excited state,  $\text{FL}^+\text{ox}$  is a strong oxidant, and promotes competitive single-electron transfer (SET) and hydrogen-atom transfer (HAT) mechanisms.<sup>21–26</sup> Strong acids are typically used to generate  $\text{FL}^+\text{ox}$  from their precursor quinones but cause competing background reactions with many acid-sensitive substrates.<sup>27</sup> Mager *et al.* showed that flavin quinones are rapidly converted to  $\text{FL}^+\text{ox}$  by anodic oxidation at  $+1.9 \text{ V}$  vs. SCE in MeCN solvent, Fig. 1.<sup>28</sup> The advantage of this electrochemical approach is that many unproductive free radicals are eliminated by reaction at the cathode or anode, suppressing background SET and HAT processes. It also precludes the use of strong acids as additives. Hence, the combination of electrochemistry and photochemistry, a.k.a., electrophotochemistry (EPC), could offer a superior route to access  $\text{FL}^+\text{ox}$ , enabling organic chemists to probe its photoacid reactivity for the first time, Fig. 1. These studies would further complement previous work showing that the combination of electricity and flavins can entice new reaction discovery<sup>29</sup> and that intermediates generated through EPC can facilitate synthetic disconnections and mechanisms not possible with prior art.<sup>30–34</sup>

## Results and discussion

Sclareolide (**1**) is an ideal substrate to probe the competency of  $\text{FL}^+\text{ox}$  as a photoacid. Functionalization of sclareolide has been demonstrated with a swathe of chemical platforms that include HAT,<sup>35–37</sup> SET,<sup>38</sup> enzymatic,<sup>39–41</sup> base-mediated,<sup>42,43</sup> and

<sup>a</sup>Department of Medicinal Chemistry, University of Kansas, Lawrence, 66045, USA. E-mail: [spbloom@ku.edu](mailto:spbloom@ku.edu)

<sup>b</sup>Department of Physics, University of Warwick, Coventry, CV4 7AL, UK

<sup>c</sup>Forensic Centre for Digital Scanning and 3D Printing, WMG, University of Warwick, Coventry, CV4 7AL, UK

† Electronic supplementary information (ESI) available: Detailed experimental procedures, computational files, product characterization data and NMR spectra, and CV, UV/Vis, fluorescence, and transient absorption spectra. CCDC 2255574. For ESI and crystallographic data in CIF or other electronic format see DOI: <https://doi.org/10.1039/d4sc03054k>

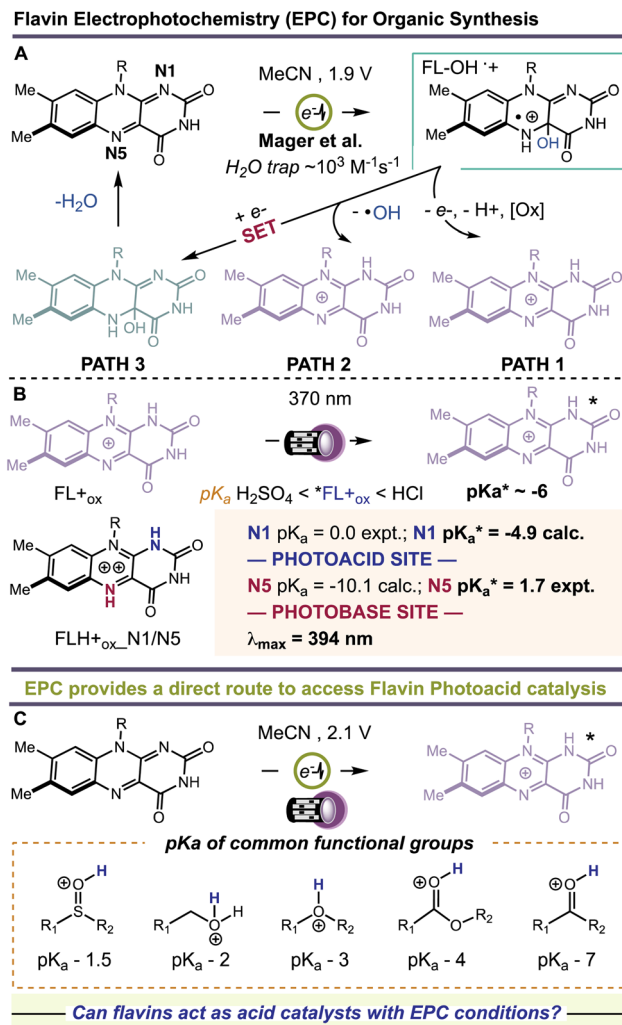


Fig. 1 (A) Electrochemical generation of 4a-hydroxy flavins and decomposition to flavinium salts. (B) Photoacid and photobase behavior of flaviniums. (C) Electrophotocatalysis route to flavin photoacid catalysis.

transition metal-catalyzed processes,<sup>44,45</sup> Fig. 2. Most of these technologies replace a single C–H bond with a new functional group. No extent method directly replaces the key skeletal atoms of sclareolide, *i.e.*, carbon and oxygen, with new atomic structures; although some methods do facilitate O → N ‘swapping’ by first opening the ‘C’ ring lactone to a hydroxamate or dioxazolone, and then recycling the ring.<sup>46,47</sup> A more straightforward path could be achieved through photoacid catalysis. The carbonyl oxygen of the ‘C’ ring lactone can be protonated with a suitable acid,  $\text{pK}_a \text{ CH}_3(\text{C}=\text{OH}^+)\text{OMe} = -3.9$ .<sup>48</sup> Protonation induces C–O bond heterolysis and 3° carbocation formation. The carbocation can be trapped with a suitable nucleophile, such as solvent MeCN, to establish a new C–N bond. Closure of the ‘C’-ring through a Mumm rearrangement affords an imide product, excising the ‘O’ atom from the former lactone and introducing a ‘N’ atom in its place.<sup>49,50</sup> If  $* \text{FL}^+_{\text{ox}}$  is a sufficient photoacid, we should obtain the imide product *in lieu* of C–H functionalized products. For this pilot

reaction we selected riboflavin tetrabutyrate (RFTB) as a catalyst (10 mol%). RFTB ( $\lambda_{\text{Abs}} = 440 \text{ nm}$ ,  $\lambda_{\text{Em}} = 519 \text{ nm}$ ) undergoes irreversible electrooxidation at +1.9 V vs. SCE to form the flavinium RFTB $^+_{\text{ox}}$  ( $\lambda_{\text{Abs}} = 396 \text{ nm}$ ,  $\lambda_{\text{Em}} = 512 \text{ nm}$ ), Fig. 1A. The four large butyrate groups give RFTB exceptional solubility in MeCN and shield the photoexcited state from nucleophilic attack, extending its lifetime in solution. The  $\text{pK}_a^*$  of RFTB $^+_{\text{ox}}$  is expected to be in line with other protonated flavins,  $-5$  to  $-8$ .<sup>19</sup> We elected to use an IKA ElectraSyn 2.0 Pro system with an undivided cell, a pair of graphite electrodes, and MeCN solvent. LiClO<sub>4</sub> was included as an electrolyte and AcOH was added as a cathodic oxidant. An alternating current of +2.1 V vs. SCE was applied for 16 h and irradiation was provided by two 370 nm Kessil lights (40 W each). From this reaction, we observed a trace amount of sclareolide-imide product (2). Increasing the loading of RFTB to 50 mol% improved the isolated yield to 17%. The imide product retained the original ‘C’ ring stereochemistry, yielding a single stereoisomer by X-ray crystallography (see ESI, page S105†). No C–H functionalization products were evident. A large fraction of the remaining mass balance is unreacted sclareolide, which was not consumed even with longer reaction times (>4 days). The imide product could not be formed in MeCN through bulk electrolysis at +4.0 V vs. SCE, direct photolysis at 370 nm, or treatment with stoichiometric acids such as H<sub>2</sub>SO<sub>4</sub> and HClO<sub>4</sub>. All conditions led to epimerization and/or carbon skeleton rearrangements, as determined by crude <sup>1</sup>H NMR of the reaction mixtures.<sup>51,52</sup> Epimerization with acids like H<sub>2</sub>SO<sub>4</sub> and HClO<sub>4</sub> is consistent with reversible proton transfer to the lactone. Proton transfer from  $* \text{FL}^+_{\text{ox}}$  is irreversible ( $\text{pK}_a$  of ground state  $\text{FL}^+_{\text{ox}} < 0$ ), obviating epimerization, and allowing the intermediate carbocation that results from C–O bond heterolysis to be trapped by solvent acetonitrile. Ammonolysis with NH<sub>3</sub> in MeOH also failed to produce the imide product, affording only the ‘C’ ring-opened amide alcohol.<sup>53</sup> Alternative photoacid catalysts, *i.e.* eosin Y, pyranine, Schreiner’s thiourea, and 7-bromonaphthalen-2-ol, were likewise unable to furnish imide 2. One-step formation of an imide 2 was only observed with flavin photoacid catalysis.

We attempted to optimize the reaction by testing alternative organic acids, electrodes, and electrolytes, but none of these changes offered any improvement. We also surveyed different flavin photocatalysts, *e.g.* lumiflavin (R = Me), riboflavin, and riboflavin 5′-monophosphate. Only lumiflavin afforded imide 2 (29% yield), but its cost is prohibitive at the high catalyst loadings needed in our reaction. Based on these results, we performed four sets of mechanistic experiments to gain deeper insight into our reaction. First, we carried out transient spectroelectrochemistry experiments to determine the excited state lifetime of RFTB $^+_{\text{ox}}$ . Passing a current of +1.7 V vs. SCE through an MeCN solution of RFTB with irradiation at 390 nm produced  $* \text{RFTB}^+_{\text{ox}}$ , which decayed over  $\sim 3 \text{ ns}$  by fluorescence at  $\sim 512 \text{ nm}$  (see ESI, page S44†). A 3 ns lifetime is comparable to riboflavin tetraacetate in 10 mM HClO<sub>4</sub>,  $\tau = 2.4 \text{ ns}$ , which has a relatively low fluorescence quantum yield,  $\Phi_F \sim 0.03$ .<sup>24</sup> The fact that  $* \text{RFTB}^+_{\text{ox}}$  exhibits a nanosecond lifetime means that it is sufficiently long-lived to engage in bimolecular photocatalysis. One important observation made during these

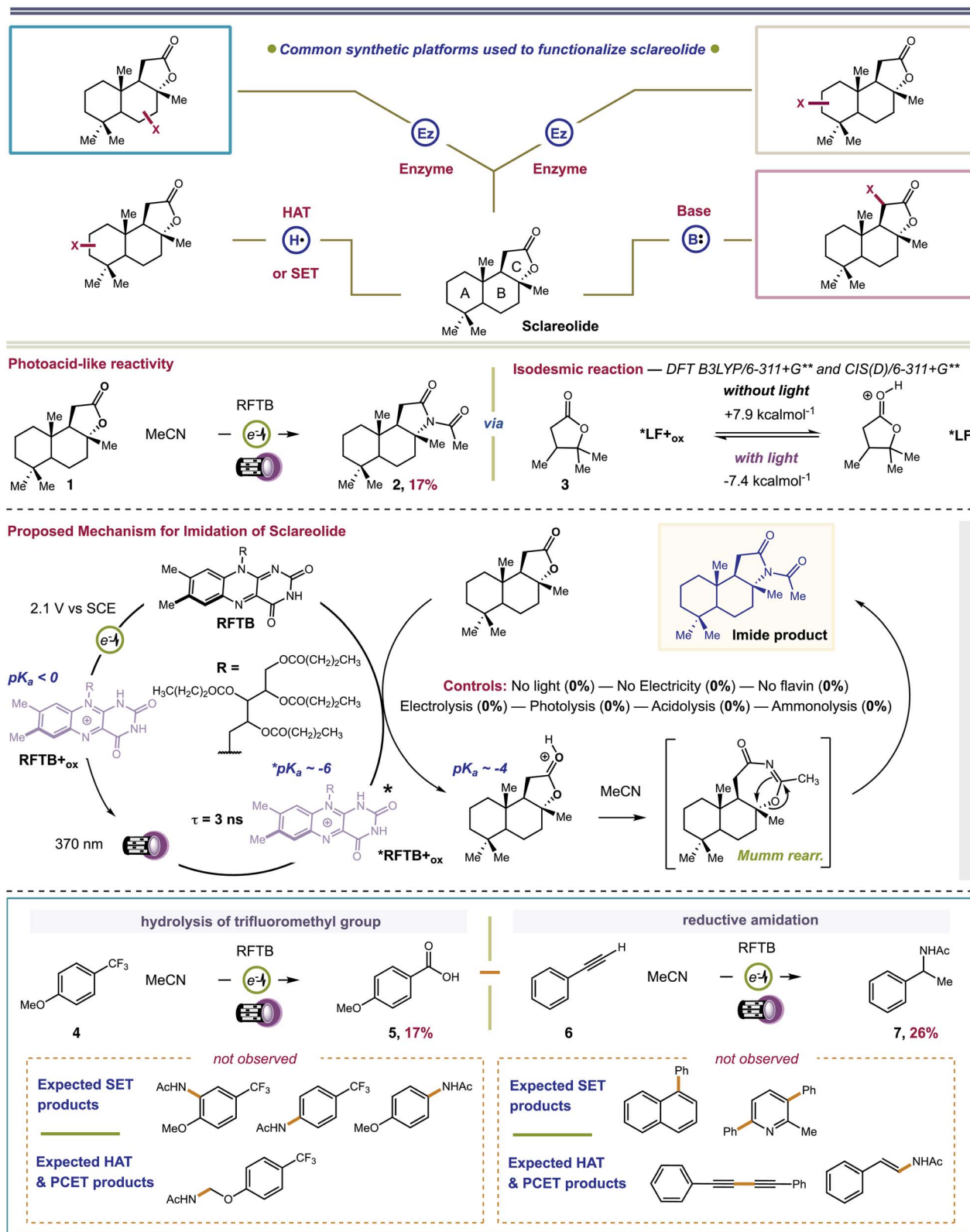


Fig. 2 Top panel: Established platforms and regioselectivities observed for functionalization of sclareolide. Second panel: Observed imidation of sclareolide under flavin photoelectrocatalysis conditions and isodesmic calculations for excited-state proton transfer using a computational surrogate for sclareolide. Third panel: Proposed mechanism for formation of imido-sclareolide 2. Bottom panel: Substrates used to distinguish photoacid reactivity from alternate mechanisms.

experiments was that extended exposure to light and anodic current converted small amounts of RFTB to several side products. By LC-MS, we tentatively assigned these products as oxygenated and amidated materials, and flavin derivatives having one less butyrate group attached (ESI, page S39†). These same byproducts were detected in our crude reaction mixture with sclareolide. While these products may only form in low amounts, their appearance suggests that at least some of our catalyst degrades over time and this could hinder the efficiency of our reaction. It also provides experimental evidence that some reactive intermediates, such as free radicals derived from MeCN (amidated catalyst) or water (oxygenated catalyst), may still exist in our reaction, and their open-shell chemistries could become more apparent when substrates with non-basic functional groups are employed; a case that we will address later in this manuscript. Second, we performed computations to study the thermochemistry of proton transfer. We used lactone **3** as a computational surrogate for the 'C' ring of sclareolide. Ground state proton transfer from a simplified flavin, lumiflavin<sup>+</sup><sub>ox</sub> (LF<sup>+</sup><sub>ox</sub>; R = CH<sub>3</sub>), to **3** is endergonic by +7.9 kcal mol<sup>-1</sup>. It is conceivable that this barrier can be overcome by visible light excitation of LF<sup>+</sup><sub>ox</sub>. Absorption of a single photon of 370 nm light provides +77 kcal mol<sup>-1</sup> driving force. In the first singlet excited state, proton transfer becomes exergonic by -7.4 kcal mol<sup>-1</sup>. Our computational analysis shows that proton transfer is only favorable in the excited state, in agreement with the proposed role of RFTB<sup>+</sup><sub>ox</sub> as a photoacid. It is also consistent with the fact that no imide product is formed in the absence of light. Third, we conducted Stern-Volmer quenching studies. We found that sclareolide quenched the excited state of RFTB<sup>+</sup><sub>ox</sub>, generated by treatment of RFTB with HClO<sub>4</sub>. Sclareolide failed to quench neutral RFTB. However, the Stern-Volmer plot for RFTB<sup>+</sup><sub>ox</sub> displayed non-linear quenching (ESI, page S15†). At high concentrations of sclareolide, we observed a negative curvature. This occurs when a decrease in the quenching constant is evident and can manifest with a change in the fluorescence spectra of the fluorophore.<sup>54</sup> It was clear from our experiment that RFTB<sup>+</sup><sub>ox</sub> fluorescence shifted from 510 to 505 nm. The absorption spectrum of RFTB<sup>+</sup><sub>ox</sub> remained constant when subjected to increasing concentrations of sclareolide, ruling out a Stokes' shift due to a change in solvent polarity. A deviation in the fluorescence spectra is also associated with the existence of multiple fluorescent states, with at least two subpopulations of the fluorophore being available. In our case, RFTB<sup>+</sup><sub>ox</sub> can exist in two tautomeric forms where the proton resides on either N1 or N5.<sup>19</sup> These can interconvert upon absorption and fluorescence (phototropism), accompanied by a dramatic change in charge density, accounting for the observed negative curvature. But this finding does not distinguish between individual mechanisms, *i.e.*, proton transfer, HAT, SET or PCET. To delineate these pathways, we proceeded to our fourth set of mechanistic experiments, that is, the extension of our method to other substrates. We chose two substrates where a photoacid product would be mechanistically distinct from PCET, HAT, SET, base-catalyzed or transition metal catalyzed processes. We initially examined 4-trifluoromethylanisole (**4**). Trifluoromethylenes are hydrolyzed

under strongly acidic conditions, producing benzoic acids.<sup>55-63</sup> SET, HAT, or PCET mechanisms lead to S<sub>N</sub>Ar of the methoxy-<sup>64-66</sup> or trifluoromethyl-group,<sup>67</sup> electrophilic aromatic substitution,<sup>68</sup> or  $\alpha$ -functionalization of the aryl ether,<sup>69-71</sup> Fig. 2. Benzoic acid **5** was isolated as the *exclusive* product from our reaction. We next examined phenylacetylene (**6**). Phenylacetylene undergoes aromatic dimerization and N-insertion chemistries when irradiated at 400 nm in the presence of strongly oxidizing (SET) photocatalysts in MeCN solvent.<sup>72</sup> It can also react with electrophilic free radicals (HAT species) to give  $\beta$ -substituted styrenes or alkyne dimers.<sup>73,74</sup> We observed none of these products. The *only* product we found was benzylic acetamide **7**, which forms from styrene produced by partial reduction of the alkyne at the cathode. Indeed, the use of styrene as reactant also afforded **7**, albeit with competitive styrene polymerization. Addition of the amide group to the benzylic carbon is consistent with formation of a carbocation intermediate, expected from a photoacid mechanism. Neither of the two products reported above could be formed by electrolysis, photolysis, acidolysis or ammonolysis. Collectively, our experiments provide evidence for the involvement of RFTB<sup>+</sup><sub>ox</sub> as a photoacid and show that its catalytic efficiency may be compromised by low levels of radical intermediates that survive past the electrodes and covalently modify the catalyst.

We examined substrates that are known to react through acid-catalyzed mechanisms to give defined products. Our first efforts focused on the Pinacol rearrangement. Our approach converted pinacol (**8**) to the classical pinacolone product (**9**) with transposition of a methyl group. Next, we examined a Ritter reaction with *tert*-butyl acetate (**10**). Proton transfer from RFTB<sup>+</sup><sub>ox</sub> causes heterolytic C-O bond fragmentation and the release of *t*Bu<sup>+</sup>, which can be trapped by MeCN solvent to give acetamide **11** after hydrolysis. This reaction worked well, yielding *tert*-butyl acetamide **11** in 59% isolated yield. Finally, we examined a Meyer-Schuster reaction with propargyl alcohol **12** and carbonyl reduction with phthalic anhydride **14**. For our Meyer-Schuster reaction, we obtained acetophenone (**13**) as the *exclusive* product. This species can be explained by acid-catalyzed [3,3]-sigmatropic rearrangement of the tertiary alcohol to give the  $\alpha,\beta$ -unsaturated phenone followed by acid-catalyzed conjugate addition of water and a retro aldol. The acid-catalyzed reduction of a single carbonyl group in phthalic anhydride also proceeded smoothly to provide phthalide (**15**). Here, the flavin photoacid activates the carbonyl group through proton transfer and reducing equivalents of electrons are supplied by the cathode. This type of electrochemical reduction was demonstrated by Baran for a related phthalimide system.<sup>75</sup> In all cases, we fully acknowledge that many of our isolated yields are 'low to moderate'. We should mention that all isolations were complicated by co-elution of the desired products with the flavin catalyst. Multiple columns were required to fully remove the catalyst, lowering our isolated yields. Still, despite the lower yields obtained, this is the first time that a flavin has ever been shown to promote acid catalyzed reactions with any success. Moreover, control reactions show that none of the products in Fig. 3 can be made in the absence of either light or current.





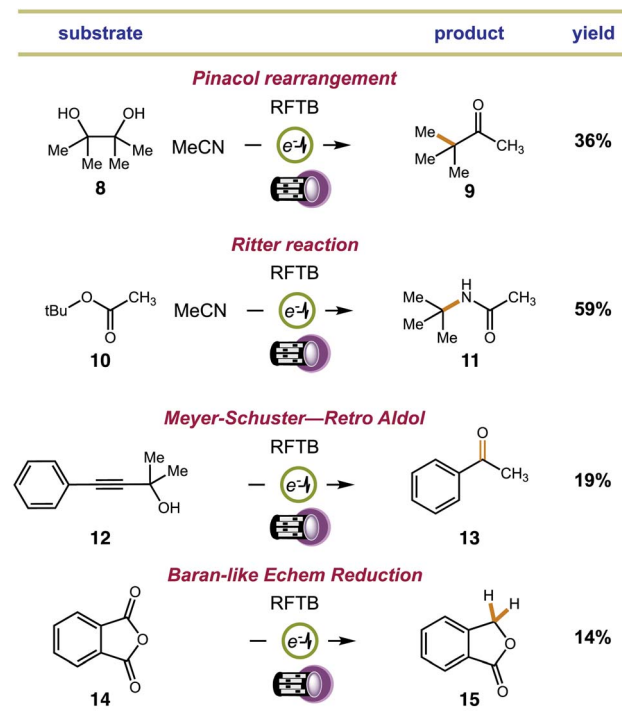


Fig. 3 Extension of electrophotochemical flavin photoacid generation to alternative acid-catalyzed reactions.

Earlier, we showed that in the absence of a suitably basic substrate, that our catalyst is covalently modified. This likely occurs by reaction with background free radical species. Mager *et al.* showed that electrophilic  $\cdot\text{OH}$  can form during flavin electrooxidation, Fig. 1 – path 2. The  $\cdot\text{OH}$  can abstract strong C–H bonds producing open shell  $\text{C}^\cdot$  intermediates. If this pathway becomes viable in the absence of non-basic substrates, then we should be able to observe C–H functionalized products in such cases. To test this, we examined cyclohexane as substrate. Cyclohexane contains no functional groups for our flavin photoacid to activate, and it is very unlikely that  $\text{RFTB}^+_{\text{ox}}$  is acidic enough to perform hydride abstraction or carbonium ion formation with alkanes, which requires acids with  $\text{pK}_a < -20$ .<sup>76</sup> Applying identical conditions as before, we isolated a small amount of *N*-cyclohexyl acetamide as the sole product from our reaction. A plausible mechanism involves C–H abstraction by  $\cdot\text{OH}$ , electrooxidation of the cyclohexyl radical to a carbocation, trapping by MeCN, and hydrolysis of the transient nitrilium ion to give the final acetamide product. The only byproduct of the reaction is  $\text{H}_2\text{O}$ , consistent with prior art by Lambert using electrophotochemical conditions.<sup>77,78</sup> Mager showed that addition of  $\text{H}_2\text{O}$  to  $\text{FL}^+_{\text{ox}}$  affords the 4a-hydroxy flavin, Fig. 1. This species is quite stable but can inevitably dehydrate to return the ground state flavin, which is then anodically re-oxidized. Because the final dehydration event dictates catalyst turnover, we wondered if we could accelerate this step. Glusac *et al.* showed that Lewis-acidic  $\text{Ce}^{\text{IV}}$  can encourage dehydration of 4a-hydroxy flavins by forming metal-hydroxides.<sup>79</sup> We surveyed a number of Lewis acids for this task,

finding a significant improvement with  $\text{Sc}(\text{OTf})_3$ . The cooperative effect between FL and  $\text{Sc}(\text{OTf})_3$  is well established, and these two species are often combined to achieve new photocatalytic reactions.<sup>80–82</sup> Before proceeding, we wished to validate the proposed free radical pathway. We ran a competitive Kinetic Isotope Effect (KIE) experiment with  $\text{d}_{12}$ -cyclohexane and performed Stern–Volmer quenching studies with cyclohexane,  $^*2\text{ScRFTB}$  and  $^*\text{RFTB}^+_{\text{ox}}$  (ESI, pages S11–S14†). We obtained a  $\text{KIE} = 1.2$ , which is not typically consistent with  $\text{Csp}^3\text{–H}$  HAT in the rate-determining step, especially by highly reactive radicals like  $\cdot\text{OH}$  ( $k \sim 10^{12} \text{ cm}^3 \text{ per molecule per s}$ )<sup>83</sup> or others that can be generated by electrophotochemical methods to activate C–H bonds.<sup>84–87</sup> In our case, generation of hydroxyl radicals, which was experimentally validated using methylene blue quenching of our crude reaction mixture—see ESI, pages S40–S41†—from 4a-hydroxyflavin radical cation is expected to be a much slower process,<sup>88</sup> and likely rate-determining, which better reflects our KIE value. We documented no quenching of any flavin species with cyclohexane, occluding the possibility of SET, proton transfer, or energy transfer.

We examined the scope and chemoselectivity of alkane amidation. For most alkanes, poor solubility was observed in MeCN solvent. Therefore, we included trichloroacetonitrile (TCA) as a cosolvent. TCA greatly improves solubility without hindering reaction efficiency or competing with MeCN in the solvent trapping step. We are aware that TCA can expel  $\text{Cl}^\cdot$  when irradiated with UV light and that  $\text{Cl}^\cdot$  can abstract hydrogen atoms from alkanes.<sup>89</sup> In a control reaction with 370 nm or 254 nm lights and 14 : 5 MeCN/TCA solvent, no product was formed when RFTB was excluded, showing that  $\text{Cl}^\cdot$  does not engender product formation when TCA is used as a cosolvent. We found that cycloalkanes  $\text{C}_5\text{–C}_{12}$  (compounds **16a–e**,  $E^0_{\text{ox}} = 2.3\text{–}2.6 \text{ V vs. SCE}$ )<sup>90</sup> were converted to new acetamide products, isolated yields 18–37%. For alkanes with both  $2^\circ$  and  $3^\circ$  C–H bonds (compounds **17–20**), we observed site-selectivities consistent with Ritter-type amidation procedures mediated by an electrophilic HAT species, like  $\cdot\text{OH}$ .<sup>91–101</sup> For instance, in methylcyclohexane, amidation at more sterically congested C2 is preferred over less crowded C4. The difference is that C2 has 2-fold more hydrogen atoms to choose from, mirroring C3. The fact that C3 has four hydrogen atoms and is sterically accessible makes it the most preferred site for kinetic C–H amidation. Our observed selectivity,  $\text{C3} > \text{C2} > \text{C4}$  is reflective of some evolved P450 nitrene-transferases<sup>102</sup> and metal-laphotoredox platforms.<sup>103</sup> This same effect can be seen for *trans*-decalin **20** and *n*-hexane **21**. When the number of hydrogen atoms is equal, the methylene site that is most accessible is amidated. Thus, a general set of rules for predicting site-selectivity in alkane substrates is established. These regiochemical preferences should hold true regardless of the selected nitrile (nucleophile) or the experimental setup used to perform our reaction. To test this, we selected three commercial nitriles that replace acetonitrile as solvent and nucleophile, and a three-neck round bottom flask equipped with a DC power supply and graphite electrodes as a substitute for our standard IKA ElectroSyn 2.0 Pro electrochemical setup. For all experiments, norbornane was used as the test alkane and TCA was omitted due to improved solubility, Fig. 4B and D. All three nitriles fashioned



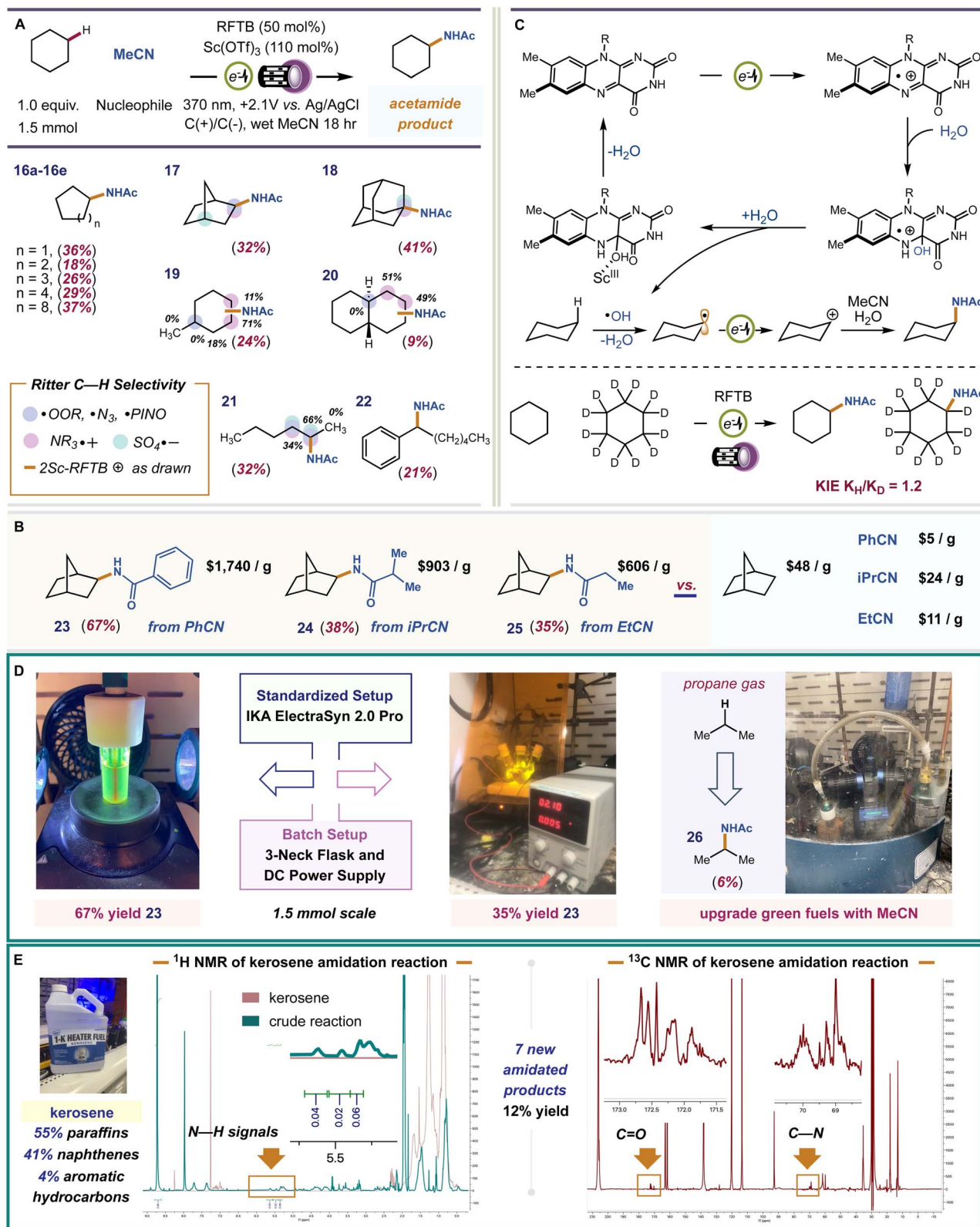


Fig. 4 Scope of the reaction and application to hydrocarbon fuels. (A) Survey of alkane substrates and a comparison of observed site-selectivities to established amidation protocols (Ritter-type) that use different catalysts/mediators. % Regiochemical distribution for substrates with multiple products is indicated beside each site. (B) Scope of alternative nitrile nucleophiles with average prices of products and starting materials from commercial vendors. (C) Proposed mechanism of C—H amidation. (D) Results with gaseous propane and a comparison of reaction efficiencies between our standard IKA ElectroSyn 2.0 Pro setup and a three-neck flask electrochemical setup. (E) Experiments performed using commercial kerosene as C—H substrate.



a single amide product (compounds **23–25**, yields 35–67%) that would be very expensive to procure otherwise. The site- and stereo-selectivities of the new amide products were identical to reactions performed with acetonitrile. In a three-neck flask, **23** was formed in 35% isolated yield. Despite its lower efficiency, this proof-of-principle experiment is evidence that our reaction can be performed in alternative setups, including those that are more conducive to large-scale batch reactions, without loss in product selectivity. Finally, we tested the capacity of our catalyst to upgrade off-the-shelf samples of common hydrocarbon fuels. The hydroxy radical formed by flavin electrooxidation is highly reactive and, therefore, can abstract a wide variety of C–H bonds (1°, 2°, and 3°) in different alkanes, improving feedstock throughput. This is harder to achieve with more selective HAT reagents that may only activate a select few hydrogen atoms, 2° and 3°. Our preliminary studies focused on gaseous propane ( $E_{\text{ox}}^0 = 2.90$  V vs. SCE) and kerosene (jet fuel), a primary distillate from crude oil that contains a heterogeneous mixture of C9–C16 alkanes and alkylarenes (Fig. 4D and E). Functionalization of C–H bonds in light alkanes (C<sub>1</sub>–C<sub>4</sub>) is intrinsically more difficult because of their higher bond-dissociation energies (BDE) and low polarities. Under our electrophotochemical conditions, *N*-isopropylacetamide (**26**) was forged in 6% LC-MS yield (Fig. 4D). For kerosene, our conditions produced a ~12% NMR yield of amidated products, which includes ~7 unique amides by <sup>1</sup>H–<sup>13</sup>C HMBC. Thus, our system can harness petroleum side streams as veritable bench deposits to pan out value-added amide products.

## Conclusions

In summary, we show that electrophotochemistry provides a direct route to access flavin photoacids, allowing them to be harnessed in organic synthesis for the first time. We show that when substrates lacking a basic functional group are employed that <sup>•</sup>OH formation becomes more apparent, enabling C–H amidation protocols. Together, our results demonstrate the unique ability of flavins (FL<sub>ox</sub>) to exhibit bimodal reactivity under electrophotochemical conditions, unveiling new mechanistic opportunities for using flavins as dynamic electrophotocatalysts.

## Data availability

Crystallographic data for compound **2** has been deposited at the Cambridge Crystallographic Data Centre (CCDC) under accession number 2255574. All other data is present in the manuscript or ESI.†

## Author contributions

S. Ga. performed all chemical reactions. J. W. and S. Go. acquired all spectroelectrochemical spectra. S. B. supervised this work. The manuscript was written with input and approval from all authors.

## Conflicts of interest

There are no conflicts to declare.

## Acknowledgements

This work was performed in the Department of Medicinal Chemistry at the University of Kansas, and supported by the School of Pharmacy, University of Kansas Startup 2506031-099, and graduate teaching assistant support for S. Ga. This work was also supported by the National Institutes of Health Graduate Training at the Biology Chemistry Interface Grant T32 GM132061 from the National Institutes of General Medical Sciences (NIGMS) and NIGMS P20GM113117. Minor salary support for S. Ga. was provided by NIGMS R35GM147169. The content is solely the responsibility of the authors and does not necessarily represent the official views of the National Institutes of Health. We thank Alonso Rodriguez Ugalde, Pei-Hsuan Chen, and Farjana Afroj for their help in reviewing the manuscript. We also thank Dr Travis Witte for assistance with collecting absorbance and fluorescence spectra, Dr Justin Douglas for help with the acquisition of 2D NMR spectra, Dr Scott Lovell and Dr Mohammad Rasel Mian for crystallography, and Dr Christopher Elles for helpful discussions. J. W. and S. Go. thank the Warwick Electrochemistry & Interfaces Group for their help with the non-aqueous spectroelectrochemical set-up. We thank Anjali John and Julie Macpherson (University of Warwick) for use of their electrochemical cell and for proofreading the manuscript.

## Notes and references

- V. Srivastava, P. K. Singh, A. Srivastava and P. P. Singh, Synthetic applications of flavin photocatalysis: a review, *RSC Adv.*, 2021, **11**, 14251–14259.
- R. Lechner, S. Kümmel and B. König, Visible light flavin photo-oxidation of methylbenzenes, styrenes, and phenylacetic acid, *Photochem. Photobiol. Sci.*, 2010, **10**, 1367–1377.
- T. Hering, B. Mühldorf, R. Wolf and B. König, Halogenase-inspired oxidative chlorination using flavin photocatalysis, *Angew. Chem., Int. Ed.*, 2016, **55**, 5342–5345.
- C. Smit, M. W. Fraaije and A. J. Minnaard, Reduction of carbon-carbon double bonds using organocatalytically generated diimide, *J. Org. Chem.*, 2008, **73**, 9482–9485.
- J. Svoboda, H. Schmaderer and B. König, Thiourea-enhanced flavin photooxidation of benzyl alcohol, *Chem.–Eur. J.*, 2008, **14**, 1854–1865.
- A. Rehpenn, A. Walter and G. Storch, Molecular flavin catalysts for C–H functionalisation and derivatisation of dehydroamino acids, *Chem. Sci.*, 2022, **47**, 14151–14156.
- Y. Imada and T. Naota, Flavins as organocatalysts for environmentally benign molecular transformations, *Chem. Rec.*, 2007, **7**, 354–361.
- J. B. Metternich and R. Gilmour, A bio-inspired, catalytic *E*→*Z* isomerization of activated olefins, *J. Am. Chem. Soc.*, 2015, **137**, 11254–11257.
- V. Mojir, E. Svobodová, K. Straková, T. Nevesely, J. Chudoba, H. Dvořáková and R. Cibulka, Tailoring flavins for visible light photocatalysis: organocatalytic [2+2] cycloadditions mediated by a flavin derivative and visible light, *Chem. Commun.*, 2015, **51**, 12036–12039.





- 10 B. König, S. Kümmel, E. Svobodová and R. Cibulka, Flavin photocatalysis, *Phys. Sci. Rev.*, 2018, **3**, 20170168.
- 11 M. Jirásek, K. Straková, T. Nevesely, E. Svobodová, Z. Rottnerová and R. Cibulka, Flavin-mediated visible-light [2+2] photocycloaddition of nitrogen- and sulfur-containing dienes, *Eur. J. Org. Chem.*, 2016, **15**, 2139–2146.
- 12 R. Cibulka and M. Fraaije, *Flavin-Based Catalysis: Principles and Applications*, Wiley-VCH Verlag GmbH & Co. KGaA, 2021.
- 13 R. Simkovitch, S. Shomer, R. Gepshtein, M. E. Roth, D. Shabat and D. Huppert, Comparison of the rate of excited-state proton transfer from photoacids to alcohols and water, *J. Photochem. Photobiol., A*, 2014, **277**, 90–101.
- 14 N. Sülzner, B. Geissler, A. Grandjean, G. Jung and P. Nuernberger, Excited-state proton transfer dynamics of a super-photoacid in acetone-water mixtures, *ChemPhotoChem*, 2022, **6**, e202200041.
- 15 I. Carmeli, D. Huppert, L. M. Tolbert and J. E. Haubrich, Ultrafast excited-state proton transfer from dicyanonaphthol, *Chem. Phys. Lett.*, 1996, **260**, 109–114.
- 16 Y.-J. Choi, H. Kim and O.-H. Kwon, Acid-base reaction of a super-photoacid with a cooperative amide hydrogen-bonded chain, *Bull. Korean Chem. Soc.*, 2022, **43**, 501–507.
- 17 J. Saway, Z. M. Salem and J. J. Badillo, Recent advances in photoacid catalysis for organic synthesis, *Synthesis*, 2021, **53**, 489–497.
- 18 B. Finkler, C. Spies, M. Vester, F. Walte, K. Omlor, I. Riemann, M. Zimmer, F. Stracke, M. Gerhards and G. Jung, Highly photostable “super”-photoacids for ultrasensitive fluorescence spectroscopy, *Photochem. Photobiol. Sci.*, 2014, **13**, 548–562.
- 19 S. G. Schulman, pH dependence of fluorescence of riboflavin and related isoalloxazine derivatives, *J. Pharm. Sci.*, 1971, **60**, 628–631.
- 20 J. Wang and Y. Liu, Systematic theoretical study on the pH-dependent absorption and fluorescence spectra of flavins, *Molecules*, 2023, **28**, 3315.
- 21 T. Hartman, M. Reisnerová, J. Chudoba, E. Svobodová, N. Archipowa, R. J. Kutta and R. Cibulka, Photocatalytic oxidative [2+2] cycloelimination reactions with flavinium salts: mechanistic study and influence of the catalyst structure, *ChemPlusChem*, 2021, **86**, 373–386.
- 22 A. Pokluda, Z. Anwar, V. Boguschová, I. Anusiewicz, P. Skurski, M. Sikorski and R. Cibulka, Robust photocatalytic method using ethylene-bridged flavinium salts for the aerobic oxidation of unactivated benzylic substrates, *Adv. Synth. Catal.*, 2021, **363**, 4371–4379.
- 23 T. Hartman and R. Cibulka, Photocatalytic systems with flavinium salts: from photolyase models to synthetic tool for cyclobutane ring opening, *Org. Lett.*, 2016, **18**, 3710–3713.
- 24 C. Pac, K. Miyake, Y. Masaki, S. Yanagida, T. Ohno and A. Yoshimura, Flavin-photosensitized monomerization of dimethylthymine cyclobutane dimer: remarkable effects of perchloric acid and participation of excited-singlet, triplet, and chain-reaction pathways, *J. Am. Chem. Soc.*, 1992, **114**, 10756–10762.
- 25 S. Fukuzumi and S. Kuroda, Photooxidation of benzyl alcohol derivatives by oxygen, catalyzed by protonated flavin analogues, *Res. Chem. Intermed.*, 1999, **25**, 789–811.
- 26 A. H. Tolba, F. Vávra, J. Chudoba and R. Cibulka, Tuning flavin-based photocatalytic systems for application in the mild chemoselective aerobic oxidation of benzylic substrates, *Eur. J. Org. Chem.*, 2019, **2020**, 1579–1585.
- 27 K. Miyake, Y. Masaki, I. Miyamoto, S. Yanagida, T. Ohno, A. Yoshimura and C. Pac, Flavin-photosensitized monomerization of dimethylthymine cyclobutane dimer in the presence of magnesium perchlorate, *J. Photochem. Photobiol.*, 1993, **58**, 631–636.
- 28 H. I. X. Mager, S.-C. Tu, Y.-H. Liu, Y. Deng and K. M. Kadish, Electrochemical superoxidation of flavins: generation of active precursors in luminescent model systems, *Photochem. Photobiol.*, 1990, **52**, 1049–1056.
- 29 W. Zhang, K. L. Carpenter and S. Lin, Electrochemistry broadens the scope of flavin photocatalysis: photoelectrocatalytic oxidation of unactivated alcohols, *Angew. Chem., Int. Ed.*, 2020, **59**, 409–417.
- 30 H. Huang, K. A. Steiniger and T. H. Lambert, Electrophotocatalysis: Combining light and electricity to catalyze reactions, *J. Am. Chem. Soc.*, 2022, **144**, 12567–12583.
- 31 L. Qian and M. Shi, Contemporary photoelectrochemical strategies and reactions in organic synthesis, *Chem. Commun.*, 2023, **24**, 3487–3506.
- 32 J. P. Barham and B. König, Synthetic photoelectrochemistry, *Angew. Chem., Int. Ed.*, 2019, **59**, 11732–11747.
- 33 J. Liu, L. Lu, D. Wood and S. Lin, New redox strategies in organic synthesis by means of electrochemistry and photochemistry, *ACS Cent. Sci.*, 2020, **6**, 1317–1340.
- 34 F. Medici, V. Chiroli, L. Raimondi and M. Benaglia, Latest updates in electrophotochemical reactions, *Tetrahedron Chem*, 2024, **9**, 100061.
- 35 G. Choi, G. S. Lee, B. Park, D. Kim and S. H. Hong, Direct C(sp<sup>3</sup>)-H trifluoromethylation of unactivated alkanes enable by multifunctional trifluoromethyl copper complexes, *Angew. Chem., Int. Ed.*, 2020, **60**, 5467–5474.
- 36 Y. Kawamata, M. Yan, Z. Liu, D.-H. Bao, J. Chen, J. T. Starr and P. S. Baran, Scalable, electrochemical oxidation of unactivated C-H bonds, *J. Am. Chem. Soc.*, 2017, **139**, 7448–7451.
- 37 R. K. Quinn, Z. A. Konst, S. E. Michalak, Y. Schmidt, A. R. Szklarski, A. R. Flores, S. Nam, D. A. Horne, C. D. Vanderwal and E. J. Alexanian, Site-selective aliphatic C-H chlorination using N-chloroamides enables a synthesis of chlorolissoclimide, *J. Am. Chem. Soc.*, 2016, **138**, 696–702.
- 38 S. Bloom, J. L. Knippel and T. Lectka, A photocatalyzed aliphatic fluorination, *Chem. Sci.*, 2014, **5**, 1175–1178.
- 39 F. Li, H. Deng and H. Renata, Remote B-ring oxidation of sclareol with an engineered P450 facilitates divergent access to complex terpenoids, *J. Am. Chem. Soc.*, 2022, **144**, 7616–7621.





- 40 J. Li, F. Li, E. King-Smith and H. Renata, Merging chemoenzymatic and radical-based retrosynthetic logic for rapid and modular synthesis of oxidized meroterpenoids, *Nat. Chem.*, 2020, **12**, 173–179.
- 41 A. Ata, L. J. Conci, J. Betteridge, I. Orhan and B. Sener, Novel microbial transformations of sclareolide, *Chem. Pharm. Bull.*, 2007, **55**, 118–123.
- 42 Z. Li, X. Yao, X. Zhang, H. Mei and J. Han, Carboxylic acid O-H insertion reaction of  $\beta$ -ester diazos enabling synthesis of  $\beta$ -acyloxy esters, *J. Org. Chem.*, 2022, **87**, 15483–15491.
- 43 J. Guo, X. Xu, Q. Xing, Z. Gao, J. Gou and B. Yu, Furfuryl cation induced cascade formal [3+2] cycloaddition/double ring-opening/chlorination: an approach to chlorine-containing complex triazoles, *Org. Lett.*, 2018, **20**, 7410–7414.
- 44 N. D. Chiappini, J. B. C. Mack and J. Du Bois, Intermolecular C(sp<sup>3</sup>)-H amination of complex molecules, *Angew. Chem., Int. Ed.*, 2018, **57**, 4956–4959.
- 45 K. Chen, A. Eschenmoser and P. S. Baran, Strain release in C-H bond activation?, *Angew. Chem., Int. Ed.*, 2009, **48**, 9705–9708.
- 46 H. Jung, H. Keum, J. Kweon and S. Chang, Tuning triplet energy transfer of hydroxamates as the nitrene precursor for intramolecular C(sp<sup>3</sup>)-H amidation, *J. Am. Chem. Soc.*, 2020, **142**, 5811–5818.
- 47 H.-Y. Jung, S. Chang and S. Hong, Strategic approach to the metamorphosis of  $\gamma$ -lactones to NH  $\gamma$ -lactams *via* reductive cleavage and C-H amidation, *Org. Lett.*, 2019, **21**, 7099–7103.
- 48 A. Bagno and G. Scorrano, Acid-base properties of organic solvents, *J. Am. Chem. Soc.*, 1988, **110**, 4577–4582.
- 49 R. Giri, I. Mosiagin, I. Franzoni, N. Y. Notel, S. Patra and D. Katayev, Photoredox activation of anhydrides for the solvent-controlled switchable synthesis of gem-difluoro compounds, *Angew. Chem., Int. Ed.*, 2022, **61**, e202209143.
- 50 J. Bolsakova and A. Jirgensons, The ritter reaction for the synthesis of heterocycles, *Chem. Heterocycl. Compd.*, 2017, **53**, 1167–1177.
- 51 A. Si, A. D. Landgraf, S. Geden, S. J. Sucheck and K. H. Rohde, Synthesis and evaluation of marine natural product-inspired meroterpenoids with selective activity toward dormant mycobacterium tuberculosis, *ACS Omega*, 2022, **7**, 23487–23496.
- 52 H.-S. Wang, H.-J. Li, X. Nan, Y.-Y. Luo and Y.-C. Wu, Enantiospecific semisynthesis of puupehedione-type marine natural products, *J. Org. Chem.*, 2017, **82**, 12914–12919.
- 53 D. Li, S. Xiang, Z. Song, W. Li, F. Zhu, J. Zhang and S. Li, Synthesis and bio-inspired optimization of drimenal: discovery of chiral drimane fused oxazinones as promising antifungal and antibacterial candidates, *Eur. J. Med. Chem.*, 2018, **143**, 558–567.
- 54 J. Keizer, Nonlinear fluorescence quenching and the origin of positive curvature in Stern-Volmer plots, *J. Am. Chem. Soc.*, 1983, **105**, 1494–1498.
- 55 D. Herrera, D. Peral and C. J. Bayon, Preparation of carboxylic arylphosphines by hydrolysis of the trifluoromethyl group, *RSC Adv.*, 2022, **12**, 7103–7114.
- 56 J. H. Simons and E. O. Ramler, Preparation and properties of some new trifluoromethyl compounds, *J. Am. Chem. Soc.*, 1943, **65**, 389–392.
- 57 H. Gilman and D. Blume, 5- and 7-trifluoromethylquinolines, *J. Am. Chem. Soc.*, 1943, **65**, 2467–2468.
- 58 G. M. Le Fave, Some reactions of the trifluoromethyl group in the benzotrifluoride series. I. Hydrolysis, *J. Am. Chem. Soc.*, 1949, **71**, 4148–4149.
- 59 G. M. Le Fave and P. G. Scheurer, Some reactions of the trifluoromethyl group in the benzotrifluoride series. II. Alcoholysis, *J. Am. Chem. Soc.*, 1950, **72**, 2464–2465.
- 60 P. G. Scheurer and G. M. Le Fave, Isophthalic and terephthalic acids, *J. Am. Chem. Soc.*, 1950, **72**, 3308–3309.
- 61 K. Wang, H. Li and J. Wen, Synthesis and mesomorphic properties of cholesteryl p-polyfluoroalkoxy-m-nitrobenzoate, *J. Fluorine Chem.*, 2001, **109**, 205–208.
- 62 Y. V. Zonov, V. M. Karpov and V. E. Platonov, Transformation of perfluorinated benzocycloalkenes and alkylbenzenes to their carbonyl derivatives under the action of CF<sub>3</sub>COOH/SbF<sub>5</sub>, *J. Fluorine Chem.*, 2007, **128**, 1058–1064.
- 63 A. Kethe, A. F. Tracy and D. A. Klumpp, Protolytic defluorination of trifluoromethyl-substituted arenes, *Org. Biomol. Chem.*, 2011, **9**, 4545–4549.
- 64 X. Wu, W. Chen, N. Holmberg-Douglas, G. T. Bida, X. Tu, X. Ma, Z. Wu, D. A. Nicewicz and Z. Li, 11C-, 12C-, and 13C-cyanation of electron-rich arenes *via* organic photoredox catalysis, *Chem*, 2023, **9**, 343–362.
- 65 V. A. Pistritto, S. Liu and D. A. Nicewicz, Mechanistic investigations into amination of unactivated arenes *via* cation radical accelerated nucleophilic aromatic substitution, *J. Am. Chem. Soc.*, 2022, **144**, 15118–15131.
- 66 N. J. Venditto and D. A. Nicewicz, Cation radical-accelerated nucleophilic aromatic substitution for amination of alkoxyarenes, *Org. Lett.*, 2020, **22**, 4817–4822.
- 67 S. Wu, J. Žurauskas, M. Domański, P. S. Hitzfeld, V. Butera, D. J. Scott, J. Rehbein, A. Kumar, E. Thyraug, J. Hauer and J. P. Barham, Hole-mediated photoredox catalysis: tris(p-substituted)biarylaminium radical cations as tunable, precomplexing and potent photooxidants, *Org. Chem. Front.*, 2021, **8**, 1132–1142.
- 68 T. Morofuji, G. Ikarashi and N. Kano, Photocatalytic C-H amination of aromatics overcoming redox potential limitations, *Org. Lett.*, 2020, **22**, 2822–2827.
- 69 Z.-Y. Dai, Z.-S. Nong, S. Song and P.-S. Wang, Asymmetric photocatalytic C(sp<sup>3</sup>)-H bond addition to  $\alpha$ -substituted acrylates, *Org. Lett.*, 2021, **23**, 3157–3161.
- 70 S. M. Treacy and T. Rovis, Copper catalyzed C(sp<sup>3</sup>)-H bond alkylation *via* photoinduced ligand-to-metal charge transfer, *J. Am. Chem. Soc.*, 2021, **143**, 2729–2735.
- 71 J. D. Bell, I. Robb and J. A. Murphy, Highly selection  $\alpha$ -aryloxyalkyl C-H functionalization of aryl alkyl ethers, *Chem. Sci.*, 2022, **13**, 12921–12926.
- 72 S. L. Mattes and S. Farid, Photosensitized electron-transfer reactions of phenylacetylene, *J. Chem. Soc., Chem. Commun.*, 1980, **3**, 126–128.



- 73 A. Souibgui, M. ben Mosbah, R. ben Salem, Y. Moussaoui and A. Tlili, Hydrotrifluoromethylation of styrene and phenylacetylene derivatives under visible-light photoredox conditions, *Chemistry*, 2022, **4**, 1010–1015.
- 74 S. S. Zaleskiy, N. S. Shlapakov and V. P. Ananikov, Visible light mediated metal-free thiol-yne click reaction, *Chem. Sci.*, 2016, **7**, 6740–6745.
- 75 Y. Kawamata, K. Hayashi, E. Carlson, S. Shaji, D. Waldmann, B. J. Simmons, J. T. Edwards, C. W. Zapf, M. Saito and P. S. Baran, Chemoselective electrosynthesis using rapid alternating polarity, *J. Am. Chem. Soc.*, 2021, **143**, 16580–16588.
- 76 G. A. Olah and R. H. Schlosberg, Chemistry in super acids. I. Hydrogen exchange and polycondensation of methane and alkanes in  $\text{FSO}_3\text{H-SbF}_5$  (“magic-acid”) solution. Protonation of alkanes and the intermediacy of  $\text{CH}_5^+$  and related hydrocarbon ions. The high chemical reactivity of “paraffins” in ionic solution reactions, *J. Am. Chem. Soc.*, 1968, **90**, 2726–2727.
- 77 T. Shen and T. H. Lambert, Electrophotocatalytic diamination of vicinal C–H bonds, *Science*, 2021, **371**, 620–626.
- 78 T. Shen and T. H. Lambert, C–H Amination via Electrophotocatalytic Ritter-type Reaction, *J. Am. Chem. Soc.*, 2021, **143**, 8597–8602.
- 79 V. Sichula, Y. Hu, E. Mirzakulova, S. F. Manzer, S. Vyas, C. M. Hadad and K. D. Glusac, Mechanism of N(5)-ethyl-flavinium cation formation upon electrochemical oxidation of N(5)-ethyl-4a-hydroxyflavin pseudobase, *J. Phys. Chem. B*, 2010, **114**, 9452–9461.
- 80 S. Fukuzumi, S. Kuroda and T. Tanaka, Flavin analog-metal ion complexes acting as efficient photocatalysts in the oxidation of p-methylbenzyl alcohol by oxygen under irradiation with visible light, *J. Am. Chem. Soc.*, 1985, **107**, 3020–3027.
- 81 D. Shen, F. Zhong, L. Li, H. Zhang, T. Ren, C. Sun, B. Wang, M. Guo, M. Chao and S. Fukuzumi, Aerobic oxyfunctionalization of alkynes by a bioinspired flavin-metal ion photocatalytic system, *Org. Chem. Front.*, 2023, **10**, 2653–2662.
- 82 B. Mühldorf and R. Wolf, Photocatalytic benzylic C–H bond oxidation with a flavin scandium complex, *Chem. Commun.*, 2015, **51**, 8425–8428.
- 83 E. S. C. Kwok and R. Atkinson, Estimation of hydroxyl radical reaction rate constants for gas-phase organic compounds using a structure-reactivity relationship: An update, *Atmos. Environ.*, 1995, **29**, 1685–1695.
- 84 P. Xu, P.-Y. Chen and H.-C. Xu, Scalable photoelectrochemical dehydrogenative cross-coupling of heteroarenes with aliphatic C–H bonds, *Angew. Chem., Int. Ed.*, 2020, **59**, 14275–14280.
- 85 L. Niu, C. Jiang, Y. Liang, D. Liu, F. Bu, R. Shi, H. Chen, A. D. Chowdhury and A. Lei, Manganese-catalyzed oxidative azidation of  $\text{C}(\text{sp}^3\text{-H})$  bonds under electrophotocatalytic conditions, *J. Am. Chem. Soc.*, 2020, **142**, 17693–17702.
- 86 L. Capaldo, L. L. Quadri, D. Merli and D. Ravelli, Photoelectrochemical cross-dehydrogenative coupling of benzothiazoles with strong aliphatic C–H bonds, *Chem. Commun.*, 2021, **57**, 4424–4427.
- 87 H. Huang, Z. M. Strater and T. H. Lambert, Electrophotocatalytic C–H functionalization of ethers with high regioselectivity, *J. Am. Chem. Soc.*, 2020, **142**, 1698–1703.
- 88 H. I. X. Mager, D. Sazou, Y. H. Liu, S.-C. Tu and K. M. Kadish, Reversible one-electron generation of 4a,5-substituted flavin radical cations: Models for a postulated key intermediate in bacterial bioluminescence, *J. Am. Chem. Soc.*, 1988, **110**, 3759–3762.
- 89 Y. Toda, R. Matsuda, S. Gomyou and H. Suga, Use of trichloroacetonitrile as a hydrogen chloride generator for ring-opening reactions of aziridines, *Org. Biomol. Chem.*, 2019, **17**, 3825–3829.
- 90 See ESI page S17† for a complete list of substrate oxidation potentials and S18–S30† for substrate cyclic voltammograms.
- 91 Q. Michaudel, D. Thevenet and P. S. Baran, Intermolecular ritter-type C–H amination of unactivated  $\text{sp}^3$  carbons, *J. Am. Chem. Soc.*, 2014, **134**, 2547–2550.
- 92 Y.-W. Zheng, R. Narobe, K. Danabauer, S. Yakubov and B. König, Copper(II)-photocatalyzed N–H alkylation with alkanes, *ACS Catal.*, 2020, **10**, 8582–8589.
- 93 V. Nair, T. D. Suja and K. A. Mohanan, A convenient protocol for C–H oxidation mediated by an azido radical culminating in ritter-type amidation, *Tetrahedron Lett.*, 2005, **46**, 3217–3219.
- 94 B. Tran, B. Li, M. Driess and J. F. Hartwig, Copper-catalyzed intermolecular amidation and imidation of unactivated alkanes, *J. Am. Chem. Soc.*, 2014, **136**, 2555–2563.
- 95 H. Lai, J. Xu, J. Lin, B. Su and D. Zha, Chemo-selective control of ritter-type reaction by coordinately unsaturated inorganic salt hydrates, *Org. Chem. Front.*, 2022, **9**, 1541–1549.
- 96 L. Zhang, Y. Fu, Y. Shen, C. Liu, M. Sun, R. Cheng, W. Zhu, X. Qian, Y. Ma and J. Ye, Ritter-type amination of  $\text{C}(\text{sp}^3)\text{-H}$  bonds enabled by electrochemistry with  $\text{SO}_4^{2-}$ , *Nat. Commun.*, 2022, **13**, 4138.
- 97 K. Kiyokawa, K. Takemoto and S. Minakata, Ritter-type amination of C–H bonds at tertiary carbon centers using iodic acid as an oxidant, *Chem. Commun.*, 2016, **52**, 13082–13085.
- 98 E. Kotani, S. Kobayashi, Y. Ishii and S. Tobinaga, A new combined oxidizing reagent system,  $\text{Fe}(\text{CH}_3\text{CN})_6^{3+}\text{-IO}_4^-$ : oxidation of paraffin hydrocarbons, *Chem. Pharm. Bull.*, 1985, **33**, 4680–4684.
- 99 J. Lee, S. Jin, D. Kim, S. H. Hong and S. Chang, Cobalt-catalyzed intermolecular C–H amidation of unactivated alkanes, *J. Am. Chem. Soc.*, 2021, **143**, 5191–5200.
- 100 A. M. Fuentes, R. Gava, N. I. Saper, E. A. Romero, A. Cabellero, J. F. Hartwig and P. J. Pérez, Copper-catalyzed dehydrogenative amidation of light alkanes, *Angew. Chem., Int. Ed.*, 2021, **60**, 18467–18471.



- 101 A. Bakhoda, Q. Jiang, Y. M. Badiei, J. A. Bertke, T. R. Cundari and T. H. Warren, Copper-catalyzed C(sp<sup>3</sup>)-H amidation: sterically driven primary and secondary C-H site selectivity, *Angew. Chem., Int. Ed.*, 2019, **58**, 3421–3425.
- 102 S. V. Athavale, S. Gao, A. Das, S. C. Mallojjala, E. Alfonz, Y. Long, J. S. Hirschi and F. H. Arnold, Enzymatic nitrogen insertion into unactivated C-H bonds, *J. Am. Chem. Soc.*, 2022, **144**, 19097–19105.
- 103 Q. Wang, S. Ni, L. Yu, Y. Pan and Y. Wang, Photoexcited direct amination/amidation of inert Csp<sup>3</sup>-H bonds *via* tungsten-nickel catalytic relay, *ACS Catal.*, 2022, **12**, 11071–11077.

