



Cite this: DOI: 10.1039/d5nr00052a

Hydrothermal microwave synthesis of water soluble NIR-II emitting Ag₂S quantum dots†

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Hydrothermal-based synthetic methods of quantum dots allow for the exploration of reaction parameters normally inaccessible to typical aqueous-based batch reactions, such as elevated reaction temperatures (>100 °C) and reaction pressures above atmospheric pressure. Coupled with microwave heating, new instantaneously bio-compatible quantum dots (QDs) with enhanced optical properties can be yielded. As of today, aqueous-based synthetic methods often lag behind their organic analogues in terms of the photophysical properties of the QDs obtained and the ease of modulation of both the emission wavelength and crystallite size. Using a novel microwave-assisted hydrothermal approach, the synthesis of silver sulphide (Ag₂S) QDs exhibiting NIR emission spanning the biological transparency windows *via* modulation of the reaction parameters has been developed. The intrinsic link between their optical and structural properties is explored *via* laboratory and synchrotron-based structural analysis techniques. Their toxicity towards a hepatic cell line was assessed, and related back to their structure and size. Overall this work aims to not only further develop the repertoire of synthetic methods for the synthesis of Ag₂S QDs, but also paves the way for the development of safer QDs suitable for future clinical applications.

Received 6th January 2025,
Accepted 26th April 2025

DOI: 10.1039/d5nr00052a

rsc.li/nanoscale

1. Introduction

Since their initial discovery in the early 1980s,^{1,2} quantum dots (QDs) have become a rapidly expanding ‘hot’ research topic, gaining vast attention across a broad range of sciences, culminating in the award of the Nobel Prize in Chemistry in 2023.³

Their unique optical and electronic properties including size-tuneable light emission, simultaneous excitation of multiple fluorescence colours, high signal-brightness and long term-photo stability^{4,5} make them ideal as candidates for use in a wide range of applications including optoelectronic devices,⁶ photo-catalysis,⁷ solar cells,⁸ biomedical,^{9–11} biosensing^{12,13} and bioimaging.^{14–16} Specifically within the field of bioimaging, their outstanding optical properties and potential for surface functionalisation with bio-molecules

(such as proteins,¹⁷ antibodies,¹⁸ drug molecules¹⁹ or DNA²⁰) make QDs prime candidates for theranostic agents, combining targeted diagnosis and therapy on a single nanoplatform. Additionally, manipulation of their size and shape can be controlled, allowing for modulation of the biodistribution and pharmacokinetics.²¹ As part of ongoing research into their potential application as theranostic agents, a large emphasis has been placed on QDs with strong emission specifically in the near infrared (NIR) spectral region in order to design attractive bio-imaging probes for diagnosis or targeted monitoring of biological functions or diseases from the organ to the molecular level.^{22,23}

Defined by IUPAC as spanning from 780 to 3000 nm,²⁴ the NIR spectral region offers significant advantages over the ultra-violet (UV) or visible range, for *in vivo* imaging at higher penetration depths. This region can be segmented further into three windows: the NIR-I, which stretches from 700–950 nm, the NIR-II, from 1000–1350, and the NIR-III, from 1550–1870 nm, referred to as the three biological windows, with each window exhibiting increased transparency towards biological matter, due to the decreasing attenuation coefficient of blood, skin and fatty tissue.²⁵ Additionally, NIR emission is spectrally different from auto-fluorescence,²⁶ with the NIR regions above 1000 nm exhibiting minimal auto-fluorescence, reducing background noise, considerably increasing the detection sensitivity and optical contrast as well as higher spatial and temporal resolution.²⁷ Compared to UV or visible light,

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†Electronic supplementary information (ESI) available. See DOI: <https://doi.org/10.1039/d5nr00052a>



the lower energy NIR is less damaging towards cells²⁵ and also exhibits lower levels of light scattering by tissue and water.²⁸ Therefore, within the biomedical field, the long-term application of QDs relies on the development of safe, non-toxic, biocompatible QDs.

Consequently, recent work has focused on the development of safer and greener QD compositions. Silver sulphide (Ag₂S) QDs have recently emerged as a promising alternative to traditional toxic binary heavy metal-containing QDs for biomedical applications due to their reportedly low cytotoxicity.^{27,29–31} Bulk Ag₂S is intrinsically non-toxic, exhibits an ultralow solubility product constant (K_{sp}) ($Ag_2S = 6.3 \times 10^{-50}$),³² minimising the release of Ag⁺ ions in solution and potentially in biological media and cells. Owing to its narrow band gap Ag₂S QDs (Ag₂S 0.9–1.1 eV)³³ possess tuneable emission ranging from the Vis-Red spectral region to the NIR-II biological window.

Ag₂S QDs were first synthesised in 1999 by Brelle *et al.*,³⁴ with subsequent research in the 2000s demonstrating their capabilities as a bio-imaging agent.³⁵ Research has rapidly expanded in the last ten years with a concentrated effort on the synthesis of QDs *via* diverse synthetic approaches, focused on precisely tuning their optical properties and size distribution,^{36–38} often in conjunction with toxicology studies³⁹ and *in vivo/in vitro* bio-sensing,³⁹ bio-imaging⁴⁰ or theranostic studies.⁴¹

Traditionally, the synthesis of Ag₂S QDs has been conducted in organic media.⁴² However, due to the toxic chemicals and harsh conditions exploited in organic synthesis, there has been a focus on shifting towards greener, aqueous synthetic methods allowing for the synthesis of instantly biocompatible QDs at lower temperatures without the need for phase transfer and the associated diminution of their fluorescence intensity.⁴³ Yet as of today, the bio-applications of Ag₂S QDs synthesised *via* aqueous approaches are limited by their poor optical properties, with observed photoluminescence quantum yields (PLQY) of less than 2%^{27,44} compared to their analogues synthesised in organic solvents, exhibiting PLQYs up to 20%.⁴⁵ Additionally, in order to reach larger particle sizes and thus NIR emission above 1000 nm, large bulky and expensive aptamers⁴⁶ or proteins,⁴⁷ are frequently used, compared to the cheaper more readily available organic ligands.

More recently, the development of microwave-assisted synthesis as an alternative heating source provides a rapid and often facile synthetic approach, opening new possibilities for synthesising nano-structured materials. Compared to conventional convection heating, microwave radiation provides faster, more uniform heating rates at a lower energy cost with highly reproducible results. Yet as of today, examples of Ag₂S QDs synthesised *via* microwave-assisted synthetic approaches fail to exhibit tuneable emission in the NIR-II window,^{48–50} limiting their application in bio-imaging applications.

From here on in, a novel microwave-assisted hydrothermal synthetic approach that yields a panel of directly water-soluble Ag₂S QDs with tuneable emission in the NIR-II window is presented. Utilising the low-cost reagent L-glutathione (GSH) as

both a capping agent and as a sulphur source, yields directly water-soluble quantum dots without the need for phase transfer or the use of bulky and expensive aptamers. Their structure, photoluminescence (PL) and the intrinsic link between both are probed, in addition to toxicity assays in order to analyse their biocompatibility.

2. Materials and methods

2.1. Chemicals

All chemicals were used as received without further purifications. L-Glutathione reduced (GSH, 97%), sodium sulphide nonahydrate (Na₂S·9H₂O, ACS, ≥98%) and silver nitrate (ACS, ≥99%, metal basis) were purchased from Thermo Scientific. Ammonium hydroxide (NH₄OH, 28% NH₃ basis) and propan-2-ol (HPLC isocratic grade) were purchased from Carlo Erba. Milli-Q ultrapure water (18.2 MΩ cm) was used in all experimental procedures and analyses.

2.2. Synthesis of GSH-capped Ag₂S QDs

0.5 mmol of AgNO₃ and either 1 mmol or 2 mmol of GSH were dissolved in 20 mL of de-ionised water under magnetic stirring, resulting in the formation of a white precipitate, suspended in solution under stirring. The pH of the solution was raised to pH 8 using 2.5 M NH₄OH, resulting in the dissolution of the precipitate (at approximately pH 5) and the formation of a colourless solution. The solution was then transferred to a 40 mL microwave vessel, which was subsequently sealed and heated to 140 °C under microwave irradiation (CEM Discover 2.0) and magnetic stirring for a pre-determined time (ranging from 1 to 20 minutes). The resulting colloidal stable solutions ranged in colour from light brown to black. The temperature of the reaction solution under microwave irradiation was monitored *via* an in-built IR probe, and a constant temperature was maintained by the automated regulation of the microwave power supplied. Ag₂S QDs were purified *via* the addition of propan-2-ol to the QD dispersion in a ratio of 3/1 (propan-2-ol/QD dispersion), followed by centrifugation at 8500 rpm for 10 minutes. The supernatant was disposed and the precipitate redispersed in de-ionised water.

2.3. Synthesis of bulk Ag₂S

0.25 mmol of AgNO₃ and 1 mmol of Na₂S were independently dissolved in 2.5 mL of deionised water to create two stock solutions. The stock solutions were mixed in the dark, resulting in the immediate formation of a black precipitate. The precipitate was collected *via* filtration, washed and dried *in vacuo* at 40 °C overnight before being stored in an amber glass vial. The X-ray diffractogram of the as-synthesised bulk Ag₂S can be found in the ESI (Fig. S1†).

2.4. Photophysical characterization

For all optical spectroscopy unless stated, the aqueous colloidal solutions were placed in 4 × 10 mm quartz cuvettes. Absorption spectra were recorded using a Jasco V-770 Visible/



NIR spectrometer with a range of 400 to 1400 nm. PL spectra were recorded using a Flurolog FL1057 spectrophotometer equipped with a 450 W Xe lamp and a Hamamatsu R5509-73 photomultiplier tube NIR detector. An excitation wavelength of 400 nm was used if not stated otherwise, and the emission spectrum was recorded in the range of 800–1500 nm. Relative measurements of the PLQY ($\Phi_{f,x}$) were performed using the reference dye IR125 dissolved in ethanol ($\Phi_{f,st} = 0.13$). The relative PLQY was calculated according to the formula of Demas and Crosby, see the equation below.

$$\Phi_{f,x} = \Phi_{f,st} \cdot \frac{F_x}{F_{st}} \cdot \frac{f_{st}(\lambda_{ex,st})}{f_x(\lambda_{ex,x})} \cdot \frac{n_x^2(\lambda_{ex,x})}{n_{st}^2(\lambda_{ex,st})}$$

The subscripts x, st and ex denote the sample, standard and excitation, respectively. $f(\lambda_{ex})$ is the absorption factor, F is the integrated spectral fluorescence photon flux and n is the refractive index of the solvents used (Ethanol for IR125; water for Ag₂S QDs). An excitation wavelength of 700 nm was used for PLQY measurements. Time-resolved measurements were performed in the wavelength region of 940 ± 10 nm using a FLS 920 (Edinburgh Instruments, Edinburgh, United Kingdom) lifetime spectrofluorometer equipped with an EPL-405 (Edinburgh Instruments, Edinburgh, United Kingdom) picosecond pulsed diode laser (excitation wavelength of 405 ± 10 nm; power of 5 mW) and a fast PMT R2658P from Hamamatsu, respectively. For lifetime and PLQY measurements 10×10 mm quartz cuvettes were used.

2.5. Transmission electron microscopy (TEM)

Morphological characterisation was achieved *via* transmission electron microscopy (TEM) analysis, using a FEI Tecnai F20 Microscope and an accelerating voltage of 200 kV. TEM grids were prepared by drop-casting dilute nanocrystal suspensions onto a 400-mesh copper TEM grid covered with an ultra-thin carbon film which was subsequently dried under an ambient atmosphere. The average diameter and size distribution (the standard deviation) of the QDs were calculated based on the measurement of at least 100 nanoparticles selected by eye and ImageJ processing software.

2.6. Powder X-ray diffraction (XRD)

Powder X-ray diffraction (XRD) patterns were recorded using a Bruker D8 powder diffractometer equipped with a copper anode ($K\alpha_1 = 1.5406$ Å, $K\alpha_2 = 1.5444$ Å) and an X' cebrator 1D detector. XRD samples were prepared by drop-casting concentrated Ag₂S dispersed in de-ionised water onto a clean silicon substrate (disorientated Si). The substrates supporting the samples for XRD were dried under an ambient atmosphere.

2.7. X-ray absorption spectroscopy (XAS) sample preparation

Purified QD solutions were deposited on the sample holder and frozen in liquid N₂ (LN₂). Reference solutions for XAS analysis of Ag–GSH at a ratio of 1/2 or 1/4 of Ag : GSH at pH 8 were prepared by mixing 0.25 mmol of AgNO₃ and either 0.5 mmol

or 1 mmol of GSH in 10 mL of de-ionised water and adjusting the pH to 8 using 1.25 M NH₄OH. A reference compound of bulk Ag₂S powder (synthesised as above) was prepared as a pellet diluted with cellulose. All liquid samples were deposited in the 3-position sample holders, equipped with Kapton windows, immediately frozen and stored in LN₂ until the experiment. The size of the frozen droplets was approximately 5 mm in diameter and 2 mm thick.

2.8. X-ray absorption spectroscopy (XAS) data acquisition and analysis

Ag K-Edge XAS acquisitions were conducted in cryogenic conditions at the CRG-FAME-UHD (BM16)⁵¹ beamline at the European Synchrotron Radiation Facility (ESRF, France). The samples were transferred into the liquid He cryostat and measured at 10 K. Ag K absorption edge was continuously scanned in the energy range 25.3–26.5 keV with a nitrogen-cooled Si(220) double-crystal monochromator.⁵² Beamsite on the sample was $240 \times 750 \mu\text{m}^2$ (HxV, Full-Width Half Maximum values). The incoming photon energy was calibrated with an Ag metallic foil, by defining the first inflection point of its XAS spectrum at 25.514 keV. Spectra were recorded in fluorescence mode with a 16-element Ge solid state detector (Mirion, Lingolsheim, France). The Ag₂S reference pellet was measured in transmission mode. The I_0 and I_1 signals are measured using Si diodes collecting a fraction of the transmitted beams scattered by 7 μm Kapton foils. The data were normalised using a linear function for the pre-edge and a 4th-order polynomial function for the after-edge and analysed as linear combinations fitting (LCF) of reference compounds by using the Larix application provided with the Larch package (version 0.9.81).⁵³

2.9. MTT cell toxicity assays

Cytotoxicity was evaluated using the 3-(4,5-dimethyl-2-thiazolyl)-2,5-diphenyltetrazolium bromide (MTT) assay. HepG2/C3A cells were provided by the ATCC. 7×10^4 HepG2/C3A cells per well were seeded in a transparent flat-bottom 96-well plate and were incubated for 24 hours at 37 °C with 5% CO₂ before the addition of QDs. To determine the LD₅₀ of the QDs, a concentration series between 0 and 300 $\mu\text{g mL}^{-1}$ of QDs was used. Five batches of Ag₂S QDs were added to the medium: GSH-coated Ag₂S QDs, synthesised from an AgNO₃ : GSH precursor ratio of 1/4 at pH 8 for either 1, 5, 10 or 15 minutes and GSH-coated Ag₂S QDs, synthesised from an AgNO₃ : GSH precursor ratio of 1/2 at pH 8 for either 15 or 20 minutes. In all experiments, each condition was performed in triplicate on the plate. Each experiment was conducted three times independently. After 24 hours of incubation in the presence of the QDs, the medium was discarded, the cells rinsed with PBS and 100 μL of medium containing 0.5 mg mL^{-1} of MTT was added to each well. After 1 hour of incubation at 37 °C, the MTT solution was discarded and formazan crystals formed were dissolved in 150 μL of 4 mM HCl, 0.1% NP40 in isopropanol. The absorbance in the wells was read in a microplate reader (BioTek Synergy H1) at 570 nm.



3. Results and discussion

Reported aqueous synthetic methods for Ag₂S QDs emitting in the second biological window typically use Na₂S as the sulphur source and mercaptopropionic acid or human/bovine serum albumin (HSA/BSA) as stabilising ligands.^{30,45,46,54,55} The utilisation of bulkier ligands, such as HSA or BSA, allowed for the regulation of the fast reaction kinetics, albeit relatively large size distributions were obtained. Following the experience gained in the aqueous synthesis of highly luminescent AgInS₂ QDs,⁵⁶ we utilised L-glutathione (GSH) in the present study. Brelle *et al.* initially used GSH in combination with Na₂S, yielding Ag₂S NCs of approximately 9 nm, however, with no mention of any photoluminescent properties.³⁴ Preliminary experiments showed Na₂S was too reactive of a sulphur source to allow for good size control, and it was found raising the temperature to 140 °C in hydrothermal conditions enabled the use of GSH as both the surface ligand and sulphur source. As in the reported works, the Ag/S precursor ratio is of prime importance for governing the size range of the yielded QDs.

3.1. Photophysical properties

Two panels of Ag₂S QDs were synthesised from differing Ag/S precursor ratios of 1/2 and 1/4, at varying reaction times of 15 to 20 minutes and 1 to 15 minutes, respectively. From here on, individual samples will be referred to as Ag₂S-X-Y, where X is the Ag/S precursor ratio and Y is the reaction time, for

example, the sample Ag₂S-14-1 corresponds to the sample synthesised from the Ag/S precursor ratio of 1/4 after a reaction time of 1 minute. Fig. 1 exhibits the absorption and emission spectra of the QDs yielded from an Ag/S precursor ratio of 1/2 and 1/4, respectively, at differing reaction times. As commonly seen for Ag₂S QDs, all absorption spectra are characterised by broad bands void of any marked features, which has previously been associated with the presence of sub-bandgap transitions involving defect states, where lattice defects have good spatial overlap with the delocalised valence or conduction band wave functions increasing the transition probability involving the valence/conduction band and a localised defect state.⁴⁵ For QDs synthesised with a reaction time between 15 and 18 minutes, from an Ag/GSH ratio of 1/2, the absorbance profiles are similar with no red-shift exhibited, however, the PL peak at 930 nm shows a continued decrease in intensity with reaction time (Fig. 1(b)). A red shift in the absorbance onset can be observed using a reaction time between 18, 19 and 20 minutes, accompanied by the loss of all emission after 19 minutes, followed by the re-emergence of a PL band at 1220 nm after a reaction time of 20 minutes. After 20 minutes, the PL band remained at 1220 nm and exhibited a significant decrease in intensity; therefore, the reaction was not explored at longer reaction times. Consequently, due to the absence of a clear red shift in the emission wavelength with reaction time, it can be postulated that the change in the emission wavelength is not a result of quantum confinement but a change in the origin of the emission.

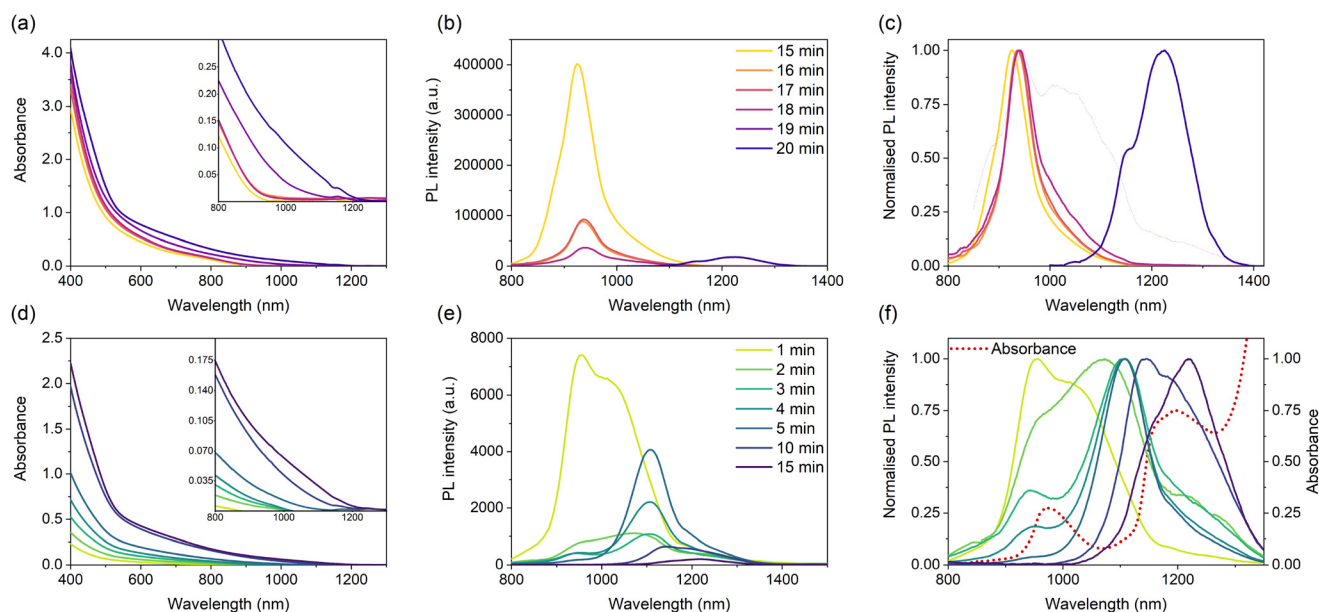


Fig. 1 Optical properties of Ag₂S QDs obtained via the microwave-assisted hydrothermal approach. (a)–(c) concern the optical properties of the Ag₂S QDs synthesised from an Ag/GSH precursor ratio of 1/2. (d)–(f) Concern the optical properties of the QDs synthesised from an Ag/GSH precursor ratio of 1/4. (a) and (d) Absorbance spectra of Ag₂S QDs synthesised from an Ag/GSH precursor ratio of 1/2 and 1/4 respectively at differing reaction times, with an inset of a zoom of the absorbance on-set; (b) and (e) absorption-corrected PL spectra of Ag₂S QDs of QDs synthesised from an Ag/GSH precursor ratio of 1/2 and 1/4 respectively obtained after different reaction times; (c) and (f) [0–1] Normalised PL spectra of Ag₂S QDs of QDs synthesised from an Ag/GSH precursor ratio of 1/2 and 1/4 respectively obtained after different reaction times, where (f) highlights the effects of absorption due to water (red dotted line) on the shape and FWHM of the PL emission peaks in the NIR domain.



Fig. 1(d–f) exhibits the absorbance and PL spectra of QDs synthesised from the 1/4 precursor ratio using reaction times between 1 and 15 minutes. In contrast to QDs synthesised from the 1/2 Ag/S precursor ratio, those synthesised from a 1/4 precursor ratio exhibit a red shift in absorbance on-set with increasing reaction time. Interestingly, between 2 and 4 minutes, the absorbance profiles (Fig. 1(d)) exhibit a minute redshift, far smaller than that exhibited between other reaction times, specifically between identical time intervals of 1 to 2 minutes and 4 to 5 minutes. This potentially may be a result of limited growth in this time domain. Fig. 1(f) exhibits a red-shift of the PL maxima with increasing reaction time from approximately 950–1220 nm, spanning the entire NIR-II spectral region. However, the absorption of NIR light by water in this spectral region (indicated by the red dotted line in Fig. 1(f)) corresponding to the bending and stretching vibrations of the hydroxyl group (–OH),⁵⁷ results in the deformation of the ‘true’ shape of the emission peak. Therefore, it is difficult to determine the exact peak positions (and hence the precise value of the redshift) along with the full-width half maxima (FWHM) of the individual emission peaks. Unexpectedly, as opposed to what is commonly seen in the literature, the emission intensity does not decrease uniformly with reaction time (typically a result of an increased number of surface traps with increasing crystallite size). However, instead the emission intensity initially decreases before increasing significantly between 3 to 5 minutes before subsequently decreasing once again (Fig. 1(e)). Despite the PL spectra for the QDs synthesised at 3, 4 and 5 minutes appearing to have a similar peak position and shape, specifically in the region of 1050–1200 nm, the difference in the area of the emission peak below 1000 nm (in the spectral region containing an absorption feature of water) indicates the true nature of the emission peaks between 3 and 5 minutes vary significantly. This is further corroborated by the samples’ absorbance spectra, which indicate a clear red shift, a feature that would be expected to be repeated in the PL plot. The photoluminescence quantum yield (PLQY) values of chosen samples (Table S1†) were obtained *via* the comparative method utilising the NIR dye IR125. For all samples tested, the PLQY was recorded as <1%, likely due to their poorly passivated surfaces, potentially arising from using GSH as both a capping ligand and sulphur source resulting in a high concentration of non-radiative surface traps.

The luminescence lifetime reflects the possible recombination paths involved in the emission of the QDs. The resulting decay signal can be fitted by a sum of exponential functions, each corresponding to a distinct recombination path. Time-resolved photoluminescence (TRPL) spectroscopy was conducted for the QD formulations with the highest PLQY for each precursor ratio, Ag₂S-12-15 and Ag₂S-14-1 synthesised from the 1/2 and 1/4 Ag/GSH precursor ratios, respectively. Commonly, the decay behaviour of Ag₂S QDs can be fitted with a two-exponential function. However, it remains important to note that due to the presence of many different de-excitation pathways resulting from differing surface trap combinations

Table 1 Tabulated fitting parameters obtained from a three-exponential fit of the time-resolved photoluminescence spectra of QD formulations Ag₂S-12-15 and Ag₂S-14-1

QD formulation	τ_1 (ns), A_1	τ_2 (ns), A_2	τ_3 (ns), A_3	τ_{int} (ns)	τ_{amp} (ns)
Ag ₂ S-12-15	5.2, 84%	22.6, 6%	79.6, 10%	50.3	13.6
Ag ₂ S-14-1	8.4, 53%	33.4, 40%	116.1, 7%	55.0	25.9

between different Ag₂S QDs formulations, a bi-exponential fit does not indicate the presence of only two relaxation pathways.⁵⁸ For TRPL spectra recorded for Ag₂S-14-1 and Ag₂S-12-15 (λ_{Em} : 940 ± 10 nm, Fig. S2†), a three-exponential function was found to best fit the experimental data. Both the intensity weighted and amplitude weighted average lifetimes, along with the individual lifetimes and relative weightings of the three decay components, are presented in Table 1. The short decay rate for both QD formulations has been previously exhibited for less efficient QDs,⁵⁹ where the τ_1 is commonly associated with decay due to non-radiative relaxation *via* the trapping of the exciton in surface quenching pathways. Despite exhibiting similar intensity weighted average lifetimes, the decrease in the weighting of τ_1 for Ag₂S-14-1 (53%) in comparison to Ag₂S-12-15 (84%) can be attributed to a decrease in the concentration of surface trap states, presumably due to higher ligand coverage and better surface passivation resulting from the higher Ag/GSH precursor ratio. Consequently, the increased passivation of non-radiative surface traps is also reflected in the higher PLQY of Ag₂S-14-1 (0.62%) in comparison to Ag₂S-12-15 (0.34%). Additionally, the shorter reaction time implies less GSH has decomposed, which also contributes to a higher ligand coverage. From thermogravimetric analysis (TGA), it can be shown for QDs of a similar average size of approximately 5 nm, namely Ag₂S-12-15 and Ag₂S-14-5, that QDs synthesised from the 1/4 Ag/GSH precursor ratio possess a higher percentage (by mass) of organics (25% and 50%, respectively), confirming the increased ligand density on the surface.

3.2. Structural properties

In order to determine the average particle size of selected QDs synthesised, TEM and the relevant image analysis were performed. The TEM images and size distribution histograms of Ag₂S-14-5, Ag₂S-14-10 and Ag₂S-14-15 are shown in Fig. 2; the TEM images and size distributions of Ag₂S-14-1, Ag₂S-12-15 and Ag₂S-12-20 can be found in the ESI (Fig. S3 and S4†). The particle diameters and their standard deviations are reported in Table 2. An increase in reaction time regardless of the precursor ratio is accompanied by an increase in the average particle size. Despite the QDs synthesised from the 1/4 Ag/GSH precursor ratio exhibiting a shift in the PL emission wavelength with reaction time consistent with that expected due to quantum confinement, the corresponding average particle sizes fall outside the quantum confinement regime. Due to its small Bohr radius of approximately 1 nm, it has been previously reported that the strong quantum confinement regime



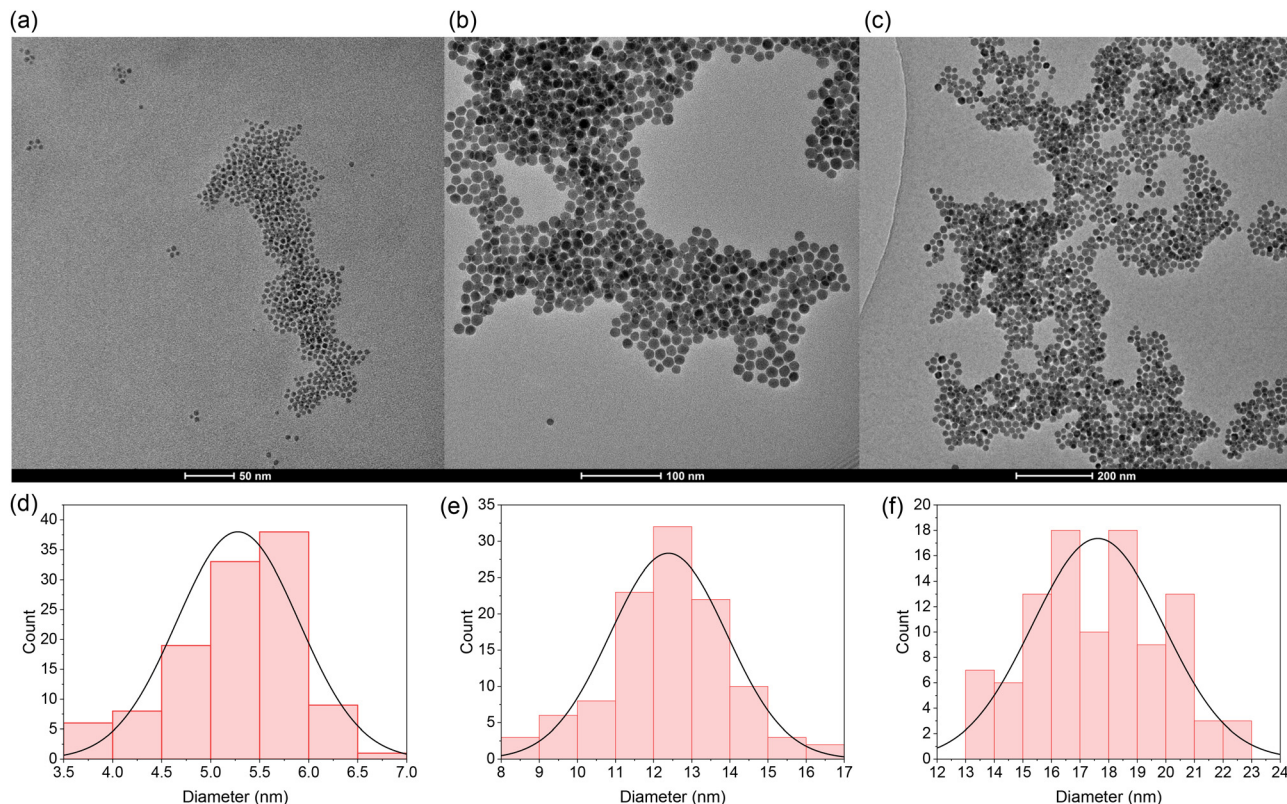


Fig. 2 Transmission electron microscopy (TEM) images of Ag₂S-14-5 (a), Ag₂S-14-10 (b) and Ag₂S-14-15 (c). Histograms of particle size overlaid with the distribution curve (black) yielded from manual particle counting image analysis ($n > 100$) for Ag₂S-14-5 (d), Ag₂S-14-10 (e) and Ag₂S-14-15 (f).

Table 2 Crystallite diameter, yielded from TEM image analysis, and LD₅₀ values (in $\mu\text{g mL}^{-1}$), yielded via MTT toxicity assays, of Ag₂S QDs synthesised from a precursor ratio of either 1/2 or 1/4 Ag/GSH with reaction times of 15 and 20 minutes or 1, 5, 10 or 15 minutes and the corresponding size of the QDs

QD formulation	Diameter (nm)	LD ₅₀ ($\mu\text{g mL}^{-1}$)
Ag ₂ S-12-15	4.6 ± 1.4	29 ± 8
Ag ₂ S-12-20	9.4 ± 1.4	>300
Ag ₂ S-14-1	3.9 ± 0.65	9 ± 1
Ag ₂ S-14-5	5.3 ± 0.65	11 ± 3
Ag ₂ S-14-10	12.5 ± 1.5	130 ± 42
Ag ₂ S-14-15	16.9 ± 2.0	>300

exists for Ag₂S QDs up to approximately 4 nm in diameter.^{33,37} Therefore, in this case, from the currently available literature, the shift in emission results from alternative origins.

X-ray diffraction patterns were collected to probe the relationship between the structure and the PL. Fig. 3(a) exhibits the diffractograms of QDs synthesised from the 1/2 Ag/S precursor ratio. In the case of the QDs synthesised after 15, 16, 17, 18 and 19 minutes a wide diffuse feature in the 2θ of $25\text{--}50^\circ$ is observed. Diffuse blurring is characteristic of the low-order Ag₂S monoclinic lattice, commonly seen for smaller Ag₂S crystallite sizes or materials with high structural disorder and defect concentrations. Contrastingly, the XRD diffracto-

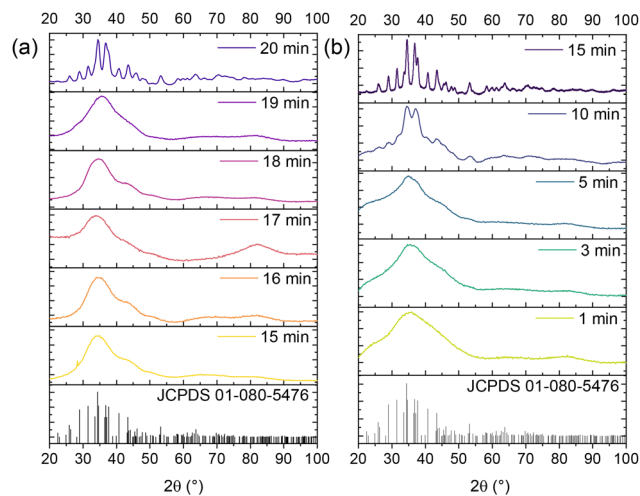


Fig. 3 X-ray diffractograms of Ag₂S QDs synthesised from an Ag/GSH precursor ratio of 1/2 (a) and 1/4 (b) at differing reaction times. The reference pattern of bulk monoclinic Ag₂S is given for comparison (JCPDS 01-080-5476).

gram of the QDs after 20 minutes exhibits a clearer more structured diffraction pattern with distinctive narrower peaks which correspond to the Ag₂S monoclinic lattice. The sudden formation of crystalline Ag₂S after 20 minutes from a highly amorphous, low crystallinity material (after 19 minutes),



results in a decrease in vacancies present in the lattice and hence trap-state energy levels potentially revealing an intrinsic link between the origin of the emission and the structure of the QDs. X-ray diffractograms (Fig. 3(b)) of the QDs synthesised from an Ag/S precursor ratio of 1/4 at shorter reaction times exhibit a wide diffuse feature in the 2θ range of 25–50°. Contrastingly, the diffractogram of the QDs synthesised after 10 minutes begins to exhibit diffraction peaks which can be assigned to the characteristic peaks of monoclinic Ag₂S. Despite this, many of the peaks at higher and lower angles compared to the two prominent peaks remain convoluted. Finally, the QDs synthesised after 15 minutes exhibit a clearer, more structured diffraction pattern with distinctive and narrower peaks which distinctly correspond to the Ag₂S monoclinic lattice. The increased definition of the peaks between the diffractograms of the QDs synthesised after 10 and 15 minutes, respectively, is likely a result of the increase in size from 12 to 16.9 nm and an increase in crystallinity of the core. In summary, XRD analysis indicates that a rapid crystallisation occurs for the samples synthesised with 1/2 Ag/GSH ratio between 19 and 20 min, while the 1/4 sample exhibits a slower crystallisation taking place between 10 and 15 min of reaction. The sharp transition in crystallinity observed for QDs synthesised with a 1/2 Ag/GSH ratio is reflected in the pronounced shift in the emission wavelength from 970 nm to 1220 nm at 20 minutes. In contrast, QDs synthesised with a 1/4 Ag/GSH ratio exhibit a more gradual increase in crystallinity, hence, this vast change in the emission wavelength is not observed. In both cases, the increase in reaction time leads to an increase of the particle size, yielding spherical nanocrystals of around 10 nm (1/2) and 17 nm (1/4) for the longest reaction time investigated.

3.3. X-Ray absorption near edge structure

The experimental X-ray Absorption Near-Edge Structure (XANES) spectra, normalised to an edge jump of 1, are reported in Fig. 4: the reference compounds are represented as dashed black lines and the QDs as symbols. The reference compounds were chosen to represent the different stages of the synthesis: from the precursor Ag–GSH solutions in 1/2 and 1/4 ratio, respectively, to the crystalline Ag₂S structure. Additionally, the spectrum of a silver foil was used as a reference for Ag(0) species, which could arise from the reduction of some metal centres during the reaction.⁶⁰ In order to quantify the composition of the internal structure of the QDs, a Linear Combination Fitting (LCF) analysis was performed; normalised XANES spectra were fitted with a least-squares minimisation algorithm as linear combinations of the reference compounds, in the energy range [25.450; 25.630] keV. It is important to notice that the main spectral difference between crystalline Ag₂S and Ag–GSH solutions, is a shift to higher energy of the second post-edge oscillation, that extends till 25.600 keV (Fig. 4). Therefore, in order to discriminate between crystalline and amorphous Ag–S complexes, it is crucial to include this oscillation in the XANES LCF range. Each experimental spectrum was fitted with all possible combinations of three stan-

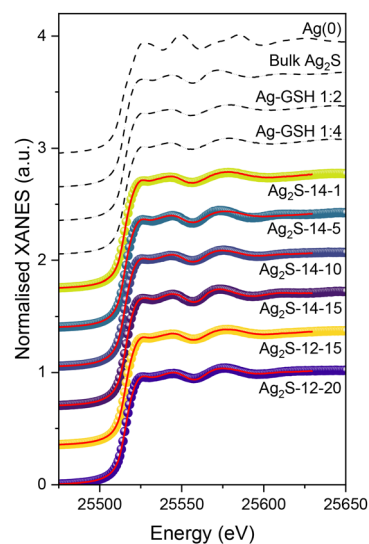


Fig. 4 Ag K-edge XANES spectra of the reference compounds (Ag(0), bulk Ag₂S, and the Ag–GSH complexes synthesised from a 1/2 and 1/4 Ag/GSH ratio) and the best-fitting curves (red) obtained as linear combinations of the reference spectra, overlapped to the experimental spectra (dots) of six different Ag₂S QD formulations.

dard compounds (Ag(0), bulk Ag₂S, and the Ag–GSH complex with stoichiometry corresponding to the QD feed ratio), where the free parameters are the weights of all the components. The results are reported in Table 3, where weights of the LCF indicate the fraction of the Ag sites in the QD structure that show a local coordination equal to the corresponding reference compound; the best-fitting curves are represented in Fig. 4 as solid red lines overlapped with the corresponding experimental curve. The fits are based on the combination of the Ag–GSH complex, with the corresponding ratio of Ag/GSH to the feed ratio for the QD, bulk Ag₂S and Ag(0) provide excellent agreement with the experimental data. For all QDs, the presence of Ag(0) reported as a fraction of the core structure falls close to or below the detection limit of the technique (5%). Furthermore, performing an LCF of the XANES region as a combination of only the corresponding Ag–GSH complex and bulk Ag₂S yielded fits with marginally inferior R_{fit} values, exhi-

Table 3 Tabulated results of the LCF XANES analysis of Ag₂S QDs synthesised from a precursor ratio of either 1/2 or 1/4 Ag/GSH with reaction times of 15 and 20 minutes or 1, 5, 10 or 15 minutes, respectively. The standard deviation error relative to the last digit is reported in brackets

QD formulation	Bulk Ag ₂ S fraction (%)	Ag–GSH fraction (%)	Ag(0) fraction (%)	R_{fit} (10^{-5})	χ^2_{reduced} (10^{-6})
Ag ₂ S-12-15	25(2)	70(2)	5(1)	1.0	6.4
Ag ₂ S-12-20	53(3)	41(3)	6(1)	3.1	19.2
Ag ₂ S-14-1	8(2)	91(2)	2(1)	1.8	11.2
Ag ₂ S-14-5	29(3)	69(3)	2(1)	3.4	20.9
Ag ₂ S-14-10	53(3)	44(3)	4(1)	2.5	15.3
Ag ₂ S-14-15	97(2)	3(2)	—	1.5	9.9



biting an increase on the order of 10^{-6} . From this, the presence of Ag(0) in the QDs can be excluded and the composition of the QDs considered as a combination of bulk Ag₂S and the corresponding Ag–GSH complex. For all QDs, regardless of the precursor ratio, the fraction of Bulk Ag₂S in the core structure increases with reaction time, from 25% to 53% for Ag₂S-12-15 and Ag₂S-12-20 and from 8% to 97% for Ag₂S-14-1 and Ag₂S-14-15, respectively. This is a reflection of not only the crystallisation of the particles from amorphous Ag₂S_y-based clusters to QDs with a bulk Ag₂S crystal structure (Fig. 3), but is also a reflection of the decreasing surface-to-volume ratio with increased particle size. For instance, core Ag atoms are more likely to occupy a bulk Ag₂S-like structure than surface atoms, which probably bind also the thiolate moieties of the ligand shell in an Ag–GSH-like coordination environment. For the QDs synthesised from an Ag/GSH precursor ratio of 1/2, however, the transformation of the core structure occurs over a far shorter period than that of the QDs synthesised from a 1/4 Ag/GSH precursor ratio. Furthermore, despite both Ag₂S-14-15 and Ag₂S-12-20 exhibiting optical properties consistent with the bulk band gap, the proportion of the QD core structure attributed to bulk Ag₂S for Ag₂S-14-15 is nearly twice that for Ag₂S-12-20 (97% versus 53%). This data is simultaneously reflected in the respective XRD of the two QDs, where the diffractogram of Ag₂S-14-15 exhibits more defined diffraction peaks than seen for Ag₂S-12-20. Furthermore, despite their differing optical properties, the LCF fit of the QDs Ag₂S-14-10 and Ag₂S-12-20 yield a similar fraction, within error, of the core corresponding to bulk Ag₂S and the initial Ag–GSH complex (53% and 41–43%, respectively), again in line with their similar X-ray diffractograms.

3.4. Toxicity assays

To further assess the feasibility of using these novel NIR-II emissive QDs as bioimaging probes, their cytotoxicity was evaluated. The viability of HepG2/C3A cells was quantified after exposure to different formulations of Ag₂S QDs for 24 hours in the concentration range of 0–300 $\mu\text{g mL}^{-1}$ using a MTT assay. For the following cytotoxicity assays, the samples Ag₂S-14-1, Ag₂S-14-5, Ag₂S-14-10, Ag₂S-14-15, Ag₂S-12-15 and Ag₂S-12-20 were selected. Conventionally, the toxicology of a species towards cells is quantified by the median lethal dose (LD₅₀), also commonly referred to as the median lethal concentration – LC₅₀, defined as the dose (or concentration) of a substance required to kill 50% of the members of a population after incubation. The plots yielded from the cell viability assay are presented in the ESI (Fig. S5†).

Interestingly, the aforementioned QDs exhibit a clear correlation between the crystallite size, crystallinity and their cytotoxicity with larger more crystalline QDs synthesised from the same precursor ratio exhibiting lower or no cytotoxicity (Table 2). For the series of QDs synthesised from the Ag/S precursor ratio of 1/4, Ag₂S-14-1 and Ag₂S-14-5 exhibited a high cytotoxicity with LD₅₀ values of less than 15 $\mu\text{g mL}^{-1}$. Contrastingly, the Ag₂S-14-10 exhibits an LD₅₀ of 130 ± 42 , whereas Ag₂S-14-15 exhibits no cytotoxicity within the concen-

tration range tested. QDs synthesised from the 1/2 Ag/S precursor ratio, show a similar pattern, where the smaller, less crystalline Ag₂S-12-15 QDs display cytotoxicity, with an LD₅₀ of $29 \pm 8 \mu\text{g mL}^{-1}$, whereas the larger, more crystalline Ag₂S-12-20 QDs exhibit no cytotoxicity in the concentration range tested. Despite Ag₂S-12-20 and Ag₂S-14-10, exhibiting both a similar crystallite size and crystallinity (Fig. 3) only Ag₂S-14-10 exhibits cytotoxic effects within the concentration range tested. However, as previously reported the cytotoxicity of QDs can rarely be associated with a single factor^{61,62} and can arise from a variety of origins including their surface structure, environmental factors, ligand coverage or leakage of ions from the core.⁶³

4. Conclusions

A novel synthesis *via* a hydrothermal-based microwave-assisted synthetic method has been presented, yielding NIR emitting QDs. *Via* modulation of the Ag/GSH precursor ratio in addition to the reaction time, the emission can be tuned across the NIR-I and NIR-II domains and biological transparency windows. Contrary to previous work in the literature, TEM analysis revealed that the particles (ranging from 4–17 nm) fall outside of the strong confinement regime despite the modulation of the emission wavelength. From XRD studies, it was determined that the PL maxima were strongly dependent on the structure, with a shift, where crystallisation of the particles resulted in a change in the origin of the emission. Despite the low PLQY of the QDs synthesised, the adaptation of shelling or doping approaches with Se,⁶⁴ Zn³⁶ or Mn,⁶⁵ currently utilised for the organic phase reactions, for aqueous phase synthetic approaches, may permit the enhancement of the QDs presented, especially those that emit at longer wavelengths. Analysis of the XANES *via* LCF of the synthesised QDs reported an increase in the structural fraction of the core corresponding to the bulk crystalline Ag₂S with increasing reaction time for all precursor ratios examined, corresponding to the X-ray diffractograms yielded for each sample. Finally, MTT assays were utilised to assess the biocompatibility of the QDs in HepG2/C3A cells, where an increase in crystallite size and crystallinity of the QDs was found to correspond to a decrease in toxicity. The presented results highlight, to the best of our knowledge, the first example of Ag₂S QDs synthesised *via* a microwave approach whose emission can be modulated to span the entire NIR-II window, paving the way for the development of new instantly water-soluble NIR emitting QDs for further bio-applications. Furthermore, more generally, the results from the toxicity assays contribute to the understanding of the size-related toxicity of these materials and lay the basis for the development of safe bio-compatible nano-probes for clinical applications.

Author contributions

Omar El-Dahshan – Investigation, writing – original draft, writing – review & editing; Aurélien Deniaud – Investigation,



writing – review & editing; Wai Li Ling – Investigation; Karl David Wegner – Investigation; writing – review & editing; Olivier Proux – Methodology, resources; Giulia Veronesi – Supervision, writing – review & editing; Peter Reiss – Supervision, writing – review & editing.

Data availability

The data supporting this article have been included as part of the ESI.† The dataset acquired during the synchrotron measurements has been deposited as follows: Deniaud, A., El-Dahshan, O., and Veronesi, G. (2027). Structure and in-cellulo stability of NIREmitting Ag₂S-based quantum dots [Dataset]. European Synchrotron Radiation Facility. <https://dx.doi.org/10.1515/ESRF-ES-1931737108>.

Conflicts of interest

There are no conflicts of interest to declare.

Acknowledgements

The authors thank the Labex ARCANÉ (ANR-11-LABX-0003-01) for the PhD funding of O. E. D and ANR for financial support (grant ANR-24-CE09-0786-01 PIQUANT). Stéphanie Pouget is thanked for assistance with powder X-ray diffraction. This work used the platforms of the Grenoble Instruct-ERIC centre (ISBG; UAR 3518 CNRS-CEA-UGA-EMBL) within the Grenoble Partnership for Structural Biology (PSB), supported by FRISBI (ANR-10-INBS-0005-02) and GRAL, financed within the University Grenoble Alpes graduate school (Ecoles Universitaires de Recherche) CBH-EUR-GS (ANR-17-EURE-0003). The IBS Electron Microscope facility is supported by the Auvergne Rhône-Alpes Region, the Fonds Feder, the Fondation pour la Recherche Médicale and GIS-IBiSA. The FAME-UHD project is financially supported by the French “grand emprunt” EquipEx (MAGNIFIX, ANR-21-ESRE-0011), the French “Programmes et équipements prioritaires de recherche” (DIADEM ESRF – ANR-22-PEXD-0011), the CEA-CNRS CRG consortium and the INSU CNRS institute. The authors would like to acknowledge SOLEIL-CRG program committee for the beamtime allocation (proposal 20240636/A16-1-839). Finally, the authors would like to acknowledge Isabelle Maurin for her help during the beamtime at FAME-UHD (BM16).

References

- R. Rossetti, S. Nakahara and L. E. Brus, *J. Chem. Phys.*, 1983, **79**, 1086–1088.
- A. I. Ekimov, A. L. Efros and A. A. Onushchenko, *Solid State Commun.*, 1985, **56**, 921–924.
- K. D. Wegner and U. Resch-Genger, *Anal. Bioanal. Chem.*, 2024, **416**, 3283–3293.
- Q. Yuan, T. Wang, P. Yu, H. Zhang, H. Zhang and W. Ji, *Org. Electron.*, 2021, **90**, 106086.
- H. Moon, C. Lee, W. Lee, J. Kim and H. Chae, *Adv. Mater.*, 2019, **31**, 1804294.
- A. P. Litvin, I. V. Martynenko, F. Purcell-Milton, A. V. Baranov, A. V. Fedorov and Y. K. Gun'ko, *J. Mater. Chem. A*, 2017, **5**, 13252–13275.
- M. Jouyandeh, S. S. M. Khadem, S. Habibzadeh, A. Esmaeili, O. Abida, V. Vatanpour, N. Rabiee, M. Bagherzadeh, S. Irvani, M. R. Saeb and R. S. Varma, *Green Chem.*, 2021, **23**, 4931–4954.
- G. H. Carey, A. L. Abdelhady, Z. Ning, S. M. Thon, O. M. Bakr and E. H. Sargent, *Chem. Rev.*, 2015, **115**, 12732–12763.
- A. A. H. Abdellatif, M. A. Younis, M. Alsharidah, O. Al Rugaie and H. M. Tawfeek, *Int. J. Nanomed.*, 2022, **17**, 1951–1970.
- A. Mukherjee and N. Sarkar, *Application of Quantum Dots in Biology and Medicine: Recent Advances*, Springer Nature, Singapore, 2022, pp. 103–123.
- S. Kargozar, S. J. Hoseini, P. B. Milan, S. Hooshmand, H.-W. Kim and M. Mozafari, *Biotechnol. J.*, 2020, **15**, 2000117.
- M. Cardoso Dos Santos, W. R. Algar, I. L. Medintz and N. Hildebrandt, *TrAC, Trends Anal. Chem.*, 2020, **125**, 115819.
- N. Hildebrandt, C. M. Spillmann, W. R. Algar, T. Pons, M. H. Stewart, E. Oh, K. Susumu, S. A. Díaz, J. B. Delehanty and I. L. Medintz, *Chem. Rev.*, 2017, **117**, 536–711.
- Y. Wang, R. Hu, G. Lin, I. Roy and K.-T. Yong, *ACS Appl. Mater. Interfaces*, 2013, **5**, 2786–2799.
- K. D. Wegner and N. Hildebrandt, *Chem. Soc. Rev.*, 2015, **44**, 4792–4834.
- W. R. Algar, M. Massey, K. Rees, R. Higgins, K. D. Krause, G. H. Darwish, W. J. Peveler, Z. Xiao, H.-Y. Tsai, R. Gupta, K. Lix, M. V. Tran and H. Kim, *Chem. Rev.*, 2021, **121**, 9243–9358.
- W. Liu, M. Howarth, A. B. Greytak, Y. Zheng, D. G. Nocera, A. Y. Ting and M. G. Bawendi, *J. Am. Chem. Soc.*, 2008, **130**, 1274–1284.
- S. Pathak, M. C. Davidson and G. A. Silva, *Nano Lett.*, 2007, **7**, 1839–1845.
- C. T. Matea, T. Mocan, F. Tabaran, T. Pop, O. Mosteanu, C. Puia, C. Iancu and L. Mocan, *Int. J. Nanomed.*, 2017, **12**, 5421–5431.
- D. Sun and O. Gang, *Langmuir*, 2013, **29**, 7038–7046.
- A. M. Smith, M. C. Mancini and S. Nie, *Nat. Nanotechnol.*, 2009, **4**, 710–711.
- J. Zhao, D. Zhong and S. Zhou, *J. Mater. Chem. B*, 2018, **6**, 349–365.
- G. Hong, A. L. Antaris and H. Dai, *Nat. Biomed. Eng.*, 2017, **1**, 1–22.
- T. I. U. o. P. a. A. Chemistry (IUPAC), *IUPAC - infrared (IT07399)*, <https://goldbook.iupac.org/terms/view/IT07399>.



- 25 E. Hemmer, A. Benayas, F. Légaré and F. Vetrone, *Nanoscale Horiz.*, 2016, **1**, 168–184.
- 26 A. Croce and G. Bottioli, *Eur. J. Histochem.*, 2014, **58**, 2461.
- 27 C. Ding, Y. Huang, Z. Shen and X. Chen, *Adv. Mater.*, 2021, **33**, 2007768.
- 28 A. N. Bashkatov, E. A. Genina, V. I. Kochubey and V. V. Tuchin, *J. Phys. D: Appl. Phys.*, 2005, **38**, 2543–2555.
- 29 B. Purushothaman and J. M. Song, *Biomater. Sci.*, 2021, **9**, 51–69.
- 30 K. D. Wegner and N. Hildebrandt, *TrAC, Trends Anal. Chem.*, 2024, **180**, 117922.
- 31 Z. Zhang, H. Yang, M. Wang, Y. Zhang, J. Jiang and Q. Wang, *Nano Res.*, 2024, **17**, 10620–10643.
- 32 R. Gui, H. Jin, Z. Wang and L. Tan, *Coord. Chem. Rev.*, 2015, **296**, 91–124.
- 33 S. Lin, Y. Feng, X. Wen, P. Zhang, S. Woo, S. Shrestha, G. Conibeer and S. Huang, *J. Phys. Chem. C*, 2015, **119**, 867–872.
- 34 M. C. Brelle, J. Z. Zhang, L. Nguyen and R. K. Mehra, *J. Phys. Chem. A*, 1999, **103**, 10194–10201.
- 35 R. D. Robinson, B. Sadtler, D. O. Demchenko, C. K. Erdonmez, L.-W. Wang and A. P. Alivisatos, *Science*, 2007, **317**, 355–358.
- 36 Z. Tang, H. Yang, Z. Sun, Y. Zhang, G. Chen and Q. Wang, *Nano Res.*, 2023, **16**, 12315–12322.
- 37 Y. Zhang, Y. Liu, C. Li, X. Chen and Q. Wang, *J. Phys. Chem. C*, 2014, **118**, 4918–4923.
- 38 H. Yang, R. Li, Z. Sun, Y. Zhang, H. Zhan, X. Chen and Q. Wang, *J. Chem. Phys.*, 2023, **159**, 014709.
- 39 P. Jiang, S. Li, M. Han, Y. Liu and Z. Chen, *Analyst*, 2019, **144**, 2604–2610.
- 40 P. Jiang, C.-N. Zhu, Z.-L. Zhang, Z.-Q. Tian and D.-W. Pang, *Biomaterials*, 2012, **33**, 5130–5135.
- 41 P. T. Buz, F. D. Duman, M. Erkisa, G. Demirci, F. Ari, E. Ulukaya and H. Y. Acar, *Nanomedicine*, 2019, **14**, 969–987.
- 42 P. Wang, R. Morales-Márquez, G. Cervás, A. H. Medel, M. P. Ogayar, D. J. de Aberasturi, A. I. de Isidro-Gomez, A. Torres-Pardo, F. J. Palomares, S. Garcia-Orrit, C. T. Sousa, A. Espinosa, H. H. Telle, D. H. Ortgies, V. Vega-Mayoral, J. Cabanillas-González, E. M. Rodríguez, U. Resch-Genger, K. D. Wegner and B. H. Juárez, *Mater. Horiz.*, 2024, **11**, 6158–6168.
- 43 G. Rotko, J. Cichos, E. Wysokińska, M. Karbowiak and W. Kałas, *Colloids Surf., B*, 2019, **181**, 119–124.
- 44 M. Jiao, A. S. Portniagin, X. Luo, L. Jing, B. Han and A. L. Rogach, *Adv. Opt. Mater.*, 2022, **10**, 2200226.
- 45 W. J. Mir, A. Swarnkar, R. Sharma, A. Katti, K. V. Adarsh and A. Nag, *J. Phys. Chem. Lett.*, 2015, **6**, 3915–3922.
- 46 J. Gao, C. Wu, D. Deng, P. Wu and C. Cai, *Adv. Healthcare Mater.*, 2016, **5**, 2437–2449.
- 47 H.-Y. Yang, Y.-W. Zhao, Z.-Y. Zhang, H.-M. Xiong and S.-N. Yu, *Nanotechnology*, 2013, **24**, 055706.
- 48 W. Bao, L. Ga, R. Zhao and J. Ai, *Biosens. Bioelectron.*, 2022, **10**, 100112.
- 49 D. Magalhães Sousa, W. Chiappim, J. P. Leitão, J. C. Lima and I. Ferreira, *ACS Omega*, 2020, **5**, 12877–12881.
- 50 Q. Ren, Y. Ma, S. Zhang, L. Ga and J. Ai, *ACS Omega*, 2021, **6**, 6361–6367.
- 51 O. Proux, X. Biquard, E. Lahera, J.-J. Menthonnex, A. Prat, O. Ulrich, Y. Soldo, P. Trévisson, G. Kapoujyan, G. Perroux, P. Taunier, D. Grand, P. Jeantet, M. Deleglise, J.-P. Roux and J.-L. Hazemann, *Phys. Scr.*, 2005, **2005**, 970.
- 52 O. Proux, V. Nassif, A. Prat, O. Ulrich, E. Lahera, X. Biquard, J.-J. Menthonnex and J.-L. Hazemann, *J. Synchrotron Radiat.*, 2006, **13**, 59–68.
- 53 M. Newville, *J. Phys.: Conf. Ser.*, 2013, **430**, 012007.
- 54 P. Jiang, Z.-Q. Tian, C.-N. Zhu, Z.-L. Zhang and D.-W. Pang, *Chem. Mater.*, 2012, **24**, 3–5.
- 55 T. Yang, Y. Tang, L. Liu, X. Lv, Q. Wang, H. Ke, Y. Deng, H. Yang, X. Yang, G. Liu, Y. Zhao and H. Chen, *ACS Nano*, 2017, **11**, 1848–1857.
- 56 C. Rivaux, T. Akdas, R. Yadav, O. El-Dahshan, D. Moodelly, W. L. Ling, D. Aldakov and P. Reiss, *J. Phys. Chem. C*, 2022, **126**, 20524–20534.
- 57 V. S. Langford, A. J. McKinley and T. I. Quickenden, *J. Phys. Chem. A*, 2001, **105**, 8916–8921.
- 58 J. W. de Wit, I. Zabala-Gutierrez, R. Marin, A. Zhakeyev, S. Melle, O. G. Calderon, J. Marques-Hueso, D. Jaque, J. Rubio-Retama and A. Meijerink, *J. Phys. Chem. Lett.*, 2024, **15**, 8420–8426.
- 59 H. D. A. Santos, I. Zabala Gutiérrez, Y. Shen, J. Lifante, E. Ximendes, M. Laurenti, D. Méndez-González, S. Melle, O. G. Calderón, E. López Cabarcos, N. Fernández, I. Chaves-Coira, D. Lucena-Agell, L. Monge, M. D. Mackenzie, J. Marqués-Hueso, C. M. S. Jones, C. Jacinto, B. del Rosal, A. K. Kar, J. Rubio-Retama and D. Jaque, *Nat. Commun.*, 2020, **11**, 2933.
- 60 A. Ortega-Rodríguez, Y. Shen, I. Zabala Gutierrez, H. D. A. Santos, V. Torres Vera, E. Ximendes, G. Villaverde, J. Lifante, C. Gerke, N. Fernández, O. G. Calderón, S. Melle, J. Marques-Hueso, D. Mendez-Gonzalez, M. Laurenti, C. M. S. Jones, J. M. López-Romero, R. Contreras-Cáceres, D. Jaque and J. Rubio-Retama, *ACS Appl. Mater. Interfaces*, 2020, **12**, 12500–12509.
- 61 A. Sukhanova, S. Bozrova, E. Gerasimovich, M. Baryshnikova, Z. Sokolova, P. Samokhvalov, C. Guhrenz, N. Gaponik, A. Karaulov and I. Nabiev, *Nanomaterials*, 2022, **12**, 2734.
- 62 A. Nagy, A. Steinbrück, J. Gao, N. Doggett, J. A. Hollingsworth and R. Iyer, *ACS Nano*, 2012, **6**, 4748–4762.
- 63 R. Hardman, *Environ. Health Perspect.*, 2006, **114**, 165–172.
- 64 D. Ruiz, M. Mizrahi, H. D. A. Santos, D. Jaque, C. M. S. Jones, J. Marqués-Hueso, C. Jacinto, F. G. Requejo, A. Torres-Pardo, J. M. González-Calbet and B. H. Juárez, *Nanoscale*, 2019, **11**, 9194–9200.
- 65 W. Ge, S. Huang, X. Huang, B. Gao, Z. Shen, X. Zhuang and F. Wang, *ACS Nano*, 2025, **19**, 6601–6612.

