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Photoinitiated thiol–ene mediated functionalisation of 4,5-enoses†

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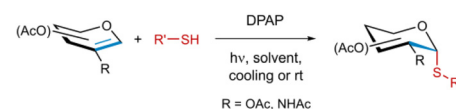
The photoinitiated thiol–ene reaction is emerging as a highly efficient methodology for thioglycoside synthesis. Herein, the radical-mediated hydrothiolation reaction of 4,5-unsaturated saccharides was extended, offering efficient access to C4-position, S-linked glycosides. A diverse range of 4,5-unsaturated saccharides were investigated with high-yields achieved for the thioether products with complete regioselectivity and good diastereoselectivity. 1,2-Ethanedithiol products furnished a thiol-residue suitable for tagging and fluorescent labelling of a disaccharide.

Introduction

Thio-linked oligosaccharides are valuable glycomimetics with inherent resistance to glycosidase activity, offering significant potential both as probes for glycobiology and as therapeutics.^{1–4} Consequently, methods for the chemical synthesis of thioglycosides *via* the regio- and stereoselective introduction of thioether groups at specific sites on the carbohydrate backbone are of considerable interest.^{5–14} Thiol–ene ligation, between a thiol and an unsaturated sugar, offers an efficient and selective approach for thioglycoside synthesis (Fig. 1A). Borbás, Somsák and co-workers have extensively investigated the thiol–ene reaction of both *endo*- and *exo*-enoses to furnish anomeric S-glycans in high yields, often with high stereoselectivity.^{9,10} Furthermore, the thiol–ene reaction of 2,3-unsaturated glycosides was applied to the introduction of a thio-linkage at either C2- or C3-position of pyranoses, with

regioselectivity highly influenced by the ring substituents. The methodology was further extended to furanose exomethylene derivatives to furnish thiol-analogues of nucleosides.⁹ The radical-mediated approach enabled access to novel thiosugars that would be challenging to synthesise through traditional ionic methods.¹⁵ 4,5-Unsaturated sugars are of particular interest in glycobiology owing to their formation as byproducts of polysaccharide lyase activity.^{16,17} These enzymes operate through an elimination mechanism resulting in oligosaccharide products displaying a terminal 4,5-unsaturated uronic acid residue. Lyase enzymes that process human glycosaminoglycans (GAGs) include heparin lyase and chondroitin lyase, critical carbohydrate processing enzymes, often overexpressed in disease states.^{18–22} The thia-Michael addition and thiol–ene reaction have been used to investigate labelling or modification of the non-reducing end of low molecular weight heparin^{23–25} or electron-deficient enoses.^{26,27}

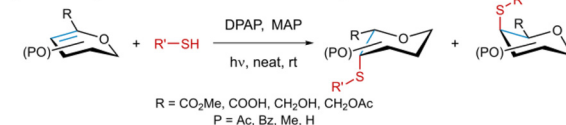
A) Photoinitiated thiol–ene reaction in 1,2-enoses



B) Base-mediated thia-Michael reaction in 4,5-enoses



C) This work: photoinitiated thiol–ene reaction in 4,5-enoses



* DPAP = 2,2-dimethoxy-2-phenylacetophenone; MAP = 4'-methoxyacetophenone

Fig. 1 (A) Photoinitiated thiol–ene in 1,2-enoses. (B) Prior thia-Michael addition study. (C) Photoinitiated thiol–ene in 4,5-enoses.

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Covalent modification of 4,5-unsaturated sugars with thiols remains relatively unexplored, with only one prior study reported to our knowledge. In this study, Liu and coworkers described a base-mediated thia-Michael addition in an electron-deficient 4,5-enose that required heating and prolonged reaction times, using thiophenol and benzyl mercaptan as model thiol reagents (Fig. 1B), and attempted the radical-mediated hydrothiolation reaction, concluding its inefficiency.²⁸ We set out to explore if the radical-mediated thiol-ene reaction could be expanded to other classes of thiols and sugar derivatives, being a milder and more efficient reaction compared to the thia-Michael process (Fig. 1C).

Results and discussion

Reaction conditions and thiol scope

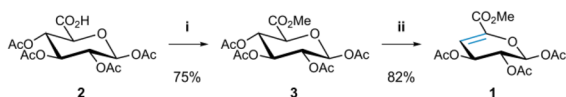
Given the unique steric challenges associated with 4,5-unsaturated systems, in particular protected uronic acid derivatives, we first sought to identify optimised reaction conditions. Prior to investigating 4,5-unsaturated sugars, we conducted our own extensive studies of commercially available 1,2-enoses to investigate if quantitative conversion could be achieved under photochemical thiol-ene conditions (see ESI†). A key finding emerging from these preliminary studies was that solvent-free reaction conditions could be employed to furnish thioether products with full conversion and simple work-up. These con-

ditions were subsequently employed as the starting point for the 4,5-unsaturated systems.

To commence our study, we first prepared 4,5-unsaturated saccharide **1**,²⁹ derived from D-glucuronic acid, using a modified protocol based on previously reported procedures (Scheme 1).^{29,30} The synthesis began with the methylation of the acetylated β-D-glucuronic acid **2**³¹ using methyl iodide and K₂CO₃ to furnish intermediate **3**,³² followed by β-elimination of the acetyl group at C4 under basic conditions yielding the desired 4,5-enose **1**.

Using **1** as a model substrate we carried out the photoinitiated reaction with 1,2-ethanedithiol **4a**, employing the latter as the solvent, under irradiation at λ_{max} 352 nm for 1 hour, at room temperature in the presence of 2,2-dimethoxy-2-phenylacetophenone (DPAP) and 4'-methoxyacetophenone (MAP), as a photoinitiator and a photosensitizer respectively. The 'neat' reaction conditions afforded the diastereomers **5a** and **6a** in a 26 : 74 ratio and a combined isolated yield of 97% (Table 1, entry 1).

NMR characterization and X-ray crystallography revealed the structures of the isolated diastereomers (Fig. 2A and ESI†). The observed coupling constants for **5a** exhibited small values (³J ~ 2–4 Hz), consistent with a *trans*-diequatorial arrangement of the coupled protons, in addition to long-range coupling constants (⁴J ~ 0.8 Hz) between diequatorial protons in a W disposition (H¹–H³ or H²–H⁴), hence supporting an α-L-idouronate in a ¹C₄ chair conformation.³³ In contrast, **6a** showed high values (³J ~ 8–10 Hz) for the *trans*-diaxial arranged protons (H¹, H², and H³) and small values for the couplings with one of the pyranose ring protons (H⁴, J ~ 2–4 Hz) in an equatorial disposition, characteristic of the ⁴C₁ chair of a β-D-galacturonate. The assigned conformations of the uronates were confirmed by 2D-NOESY spectra. Thus, NOE contacts were observed between the equatorial-oriented H⁴ and the axial-oriented protons (H¹, H³ and H⁵), justifying the ⁴C₁ conformation of **6a**. These axial NOE contacts were absent in **5a**,



Scheme 1 Synthesis of 4,5-unsaturated saccharide **1**. Reaction conditions: (i) CH₃I, K₂CO₃, DMF, rt, 5 h; (ii) DBU, CH₂Cl₂, 0 °C to rt, 19 h.

Table 1 Photoinitiated thiol-ene conditions using 4,5-unsaturated saccharide **1** as the model substrate

Entry	Photo-initiator	Light source (λ _{max})	T (°C)	Time (h)	Solvent	Yield ^a (%) (5a / 6a) ^b
1	DPAP/MAP	UV-A (352 nm)	25	1	Neat	97 (26 : 74)
2	DPAP/MAP	UV-A (352 nm)	25	2	EtOAc ^c	26 (29 : 71)
3	Eosin Y	Blue LED (450 nm)	25	2	EtOAc : THF (1 : 1, v : v) ^d	48 (22 : 78)
4	Thioxanthene-9-one	Blue LED (450 nm)	25	2	EtOAc : THF (1 : 1, v : v) ^d	10 (n/a)
5	DPAP/MAP	In dark	25	2	Neat	0 ^e (n/a)
6	DPAP/MAP	UV-A (352 nm)	–18 ^f	1	Neat	97 (37 : 63)
7	No catalyst ^g	In dark	50	1	CHCl ₃	— (n/a)

Scale: 0.24 mmol in 0.06 mL of thiol, neat. ^a Combined yield. ^b Isolated compounds. ^c 0.5 mL solution. ^d 0.5 mL added for photoinitiator solubility. ^e Recovered 96% of unreacted **1**. ^f MeOH/crushed ice cooling bath. ^g Thia-Michael addition using *N,N*-diethylamine (2 equiv.) as base led to a complex mixture of degradation products.



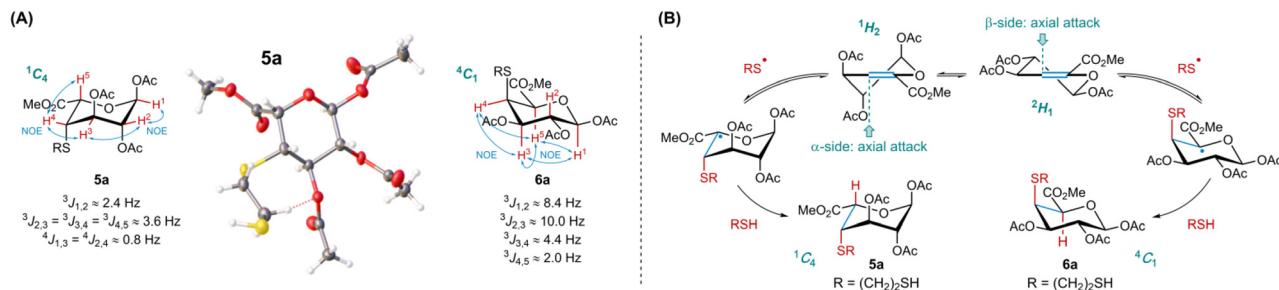


Fig. 2 (A) J values and NOE signals for **5a/6a** structural determination and X-ray crystal structure of **5a**. (B) Proposed mechanism and diastereoselectivity considerations for thiol-ene reaction on **1**.

as expected for the 1C_4 chair, showing instead the spatial interactions of each proton with its immediate vicinal protons in an all-equatorial arrangement, except for the axial-oriented H^5 . The structure of α -L-idouronate **5a**, was confirmed by single-crystal X-ray diffraction analysis, showing the absolute configuration of the substituents along the pyranose ring, albeit, the sugar crystallized in its 4C_1 conformation. Suitable crystals of **6a** could not be obtained, despite extensive crystallization attempts (see ESI†). However, suitable crystals of structurally related product **6b** were successfully obtained, and the X-ray single-crystal diffraction analysis showed the absolute configuration of the substituents, confirming a β -D-galacturonate structure (see ESI†).

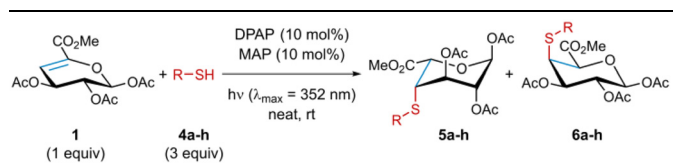
The solved crystal structures and initial diastereoselectivity outcomes led us to propose a reaction mechanism, illustrated in Fig. 2B. The minor product **5a** may arise from an α -side axial attack of the thiyl radical onto the 1H_2 conformation of **1**, followed by ring flip to the 1C_4 conformation and axial hydrogen abstraction from the thiol, affecting an overall *trans*-diaxial addition across the enose. Comparatively, for the formation of the major product **6a**, a β -side axial attack can occur on the ring flipped 2H_1 conformation of **1**, leading to a similar overall *trans*-diaxial addition but affording a 4C_1 galacturonate-configured product.^{34–36} Furthermore, only C4 addition products were observed and this complete regioselectivity may be explained by enhanced captodative stability of the intermediate tertiary radical, formed upon addition to the less substituted olefinic carbon.

Since clearly not all thiol substrates are amenable to solvent-free reaction conditions, further optimisation studies were carried out in organic solvents, varying reaction parameters including photoinitiator, light source and temperature. Optimisation studies quickly revealed however that use of neat conditions furnished the highest yields (Table 1, entries 1 and 6), when paired with the optimised photoinitiation system of DPAP/MAP under UV light irradiation. A significant drop in yield to 26% was observed when using EtOAc as a solvent (Table 1, entry 2). However this improved to 48% upon using a mixture of EtOAc : THF (1 : 1) and using Eosin Y as an initiator under Blue LED (450 nm) conditions (Table 1, entry 3). A drop in yield to 10% was observed when a less efficient photoinitiator, thioxanthene-9-one, was used highlighting the importance of an efficient photoinitiator system for achieving high

yields (Table 1, entry 4). Indeed, the dark reaction furnished no thioether products (as expected), demonstrating the radical nature of the process and the requirement for a photochemical initiation of the thiol-ene radical chain process (Table 1, entry 5). Reduced temperature furnished a 97% conversion, showing a slight variation in the diastereomeric ratio from 26 : 74 at room temperature to 37 : 63 at -18 °C (Table 1, entry 6), providing further evidence that the major product originates from thermodynamically favoured axial addition of the thiyl radical onto the alkene. The addition of a base to promote the competing thia-Michael addition was also investigated (Table 1, entry 7), however, this failed to furnish any thioether products. Finally, to demonstrate the ability to scale up, this solvent-free method was successfully completed on 1 mmol scale to produce **5a** and **6a** in 87% yield and 16 : 84 ratio (see ESI†). Interestingly, upon heating of compound **6a** during crystallization studies, a novel bicyclic, dithioether derivative was formed and its X-ray structure was solved (**6a'**, see ESI†).

With optimised conditions in hand and some insight into the expected regio- and stereochemical outcomes of the thiol-ene reaction, we next sought to investigate the scope of the reaction using a variety of thiols, including alkyl, aromatic and thioacid derivatives (Table 2). Since all of the thiols screened were either oils or liquids, solvent-free conditions were used in each case. Thus, the reaction between **1** and 1,3-propanedithiol (**4b**) furnished a 92% yield of the corresponding diastereomers (**5b/6b**) in a 2 : 8 ratio (Table 2, entry 2). Similarly, the reaction with 3-chloropropanethiol (**4c**), 3-mercapto-1,2-propanediol (**4d**), 3-mercaptopropionate (**4e**) and 2-mercaptoacetate (**4f**) each afforded the corresponding diastereomers **5c–f** and **6c–f** in good to excellent yields (50–99%, Table 2, entries 3–6), with slightly lower yields observed for the more bulky thiol substrates (Table 2, entries 4 and 6). The ratios of the isolated isomers were *ca.* 2 : 8, in favour of the axial addition products, with a slightly reduced ratio of 3 : 7 observed for **4f** (Table 2, entry 6), potentially arising from the proximity of the bulky carbonyl group to the thiol residue which may interfere with axial attack of the corresponding thiyl radical. Interestingly, reaction of **1** with thioacetic acid (**4g**), a well-known and highly utilised substrate for thiol-ene reactions with enoses, furnished a poor yield of only 21% and a **5g/6g** ratio of *ca.* 1 : 1 even after 6 hours (Table 2, entry 7), suggesting that steric bulk around the thiyl radical is detrimental to efficient thiol-ene reaction



Table 2 Scope of the photoinitiated thiol–ene reaction using uronate **1** with different thiols (**4a–h**)


Entry	Thiol	Time (h)	Yield ^a (%)	5/6 ratio ^b
1	4a	1.0	97	5a/6a (26 : 74)
2	4b	1.0	92	5b/6b (23 : 77)
3	4c	1.0	87	5c/6c (18 : 82)
4	4d	3.5	50	5d/6d (20 : 80)
5	4e	1.0	99	5e/6e (20 : 80)
6	4f	2.5	61	5f/6f (31 : 69)
7	4g	6.0	21	5g/6g (43 : 57)
8	4h	2.5	26	5h/6h (0 : 100)

^a Combined yield. ^b Isolated compounds.

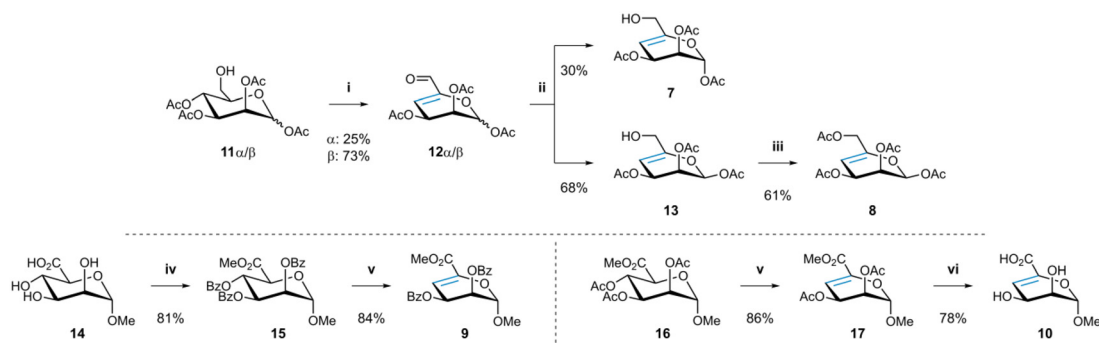
with **1**. Recent studies by Kelemen and co-workers suggest that degradation of the thioacid may also be a factor in the low yield for this substrate.³⁷ The reaction with benzyl mercaptan (**4h**) only furnished diastereomer **6h**, albeit in a low yield of 26% (Table 2, entry 8), again due to steric interference arising from the bulky benzyl moiety. Overall, these findings demonstrate that while the reaction does appear to be tolerant of a diverse range of thiols, having a bulky residue in the proximity of the reactive heteroatom radical hampers product formation, increases reaction time, lowers the reaction yields and/or varies the diastereomeric product ratios. Furthermore, in the series evaluated here the product ratio appears to follow a pattern whereby the ²H₁ enose conformer reacts preferentially, perhaps avoiding unwanted steric (axial substituent) penalties *via* the competing ¹H₂ pathway (see Fig. 2B).

Synthesis and thiol–ene scope on 4,5-enoses

To further investigate the scope of the thiol–ene reaction, we next prepared a range of 4,5-unsaturated enoses (**7–10**, Scheme 2) including both protected and fully unprotected sugars. We also included non-uronate derivatives to investigate the impact of having an alkyl group at C5 since this should lessen the steric constraints for the thiol–ene addition.

The synthesis of compounds **7** and **8** began with the preparation of intermediate **11** from D-mannose (see ESI†).^{38,39} Next, oxidation of the primary alcohol *via* Dess–Martin periodinane and subsequent β-elimination of the acetyl group at C4 under basic conditions furnished the 4,5-enose **12** as a separable mixture of α/β anomers. The partially protected mannosyl 4,5-unsaturated derivative **7** was obtained upon reduction of the aldehyde **12α** in the presence of sodium borohydride, whilst the fully protected mannosyl 4,5-enose **8** was isolated following reduction of **12β** to the respective primary alcohol **13**, followed by acetylation. Synthesis of benzoyl-protected 4,5-unsaturated methyl uronate **9** involved conversion of methyl α-D-mannosiduronic acid **14**⁴⁰ into the fully protected derivative **15**⁴¹ *via* a two-step process involving methyl iodide mediated esterification under basic conditions, followed by perbenzoylation upon treatment with BzCl in pyridine. DBU-mediated β-elimination of the benzoyl group at C4 furnished the desired 4,5-enose **9**. The fully unprotected 4,5-enose **10** was synthesised from the acetyl-protected methyl uronate **16**,⁴² a derivative of α-D-mannosiduronic acid (see ESI†), whose acetyl group at C4 was subjected to base-promoted β-elimination to furnish 4,5-enose **17**, which was subsequently deprotected upon treatment with NaOH in EtOH/H₂O (1 : 1).

Next, we proceeded to investigate the thiol–ene reaction of these alternate 4,5-enoses using the previously optimised conditions with 1,2-ethanedithiol **4a** employed as the thiol in each case (Table 3). For compound **7**, a C1,2,3-protected mannosyl 4,5-enose, and compound **8**, a fully acetylated form of **7**, only a single diastereomer was isolated in each case, identified as ⁴C₁ mannosyl derivatives **18** or **19**, respectively. Indicatively, a combination of changing the C6 uronate ox-



Scheme 2 Synthesis of 4,5-unsaturated monosaccharides **7–10**. Reaction conditions: (i) (a) DMP, CH₂Cl₂, rt, 6 h; (b) Et₃N, CH₂Cl₂, rt, 1–2 h; (ii) NaBH₄, THF, 0 °C to rt, 1–2 h; (iii) Ac₂O, DMAP, pyridine, CH₂Cl₂, rt, 3 h; (iv) (a) CH₃I, K₂CO₃, DMF, rt, 14 h; (b) BzCl, pyridine, 0 °C to rt, 12 h; (v) DBU, CH₂Cl₂, 0 °C to rt, 2 h; (vi) NaOH, EtOH/H₂O (1 : 1), rt, 6 h.



Table 3 Scope of the photoinitiated thiol–ene reaction in 4,5-unsaturated glycans **7–10** using 1,2-ethanedithiol (**4a**)

Entry	4,5-enose	Product	Yield ^a (%)
1			49
2			37
3			87 ^b
4			45 ^c

^a Yield of isolated compounds after 1 h irradiation using thiol **4a** (3 equiv.), DPAP (10 mol%) and MAP (10 mol%). ^b Combined yield, isolated compound ratio 32:68. ^c MeOH (0.1 mL) was added for solubility.

dation level to hexose, together with a C2-axially-oriented substituent, promoted an alternate overall *cis* addition across the enose (Table 3, entries 1 and 2), although the yields were low in both cases. By contrast, return to a benzoyl protected 4,5-unsaturated mannose uronate **9** formed diastereomers **20a** and **20b** in 87% yield and in a 32 : 68 ratio (Table 3, entry 3). This pair were identified as derivatives of α -D-taluronate (C2 and C4 epimer of glucose) and β -L-guluronate respectively and supports the finding that the presence of an ester at C5 is optimum for a *trans*-diaxial thiol–ene addition onto 4,5-enoses (here using thiol **4a** and considering D-manno and D-gluco configurations only). Finally, for a fully unprotected uronic acid enose **10**, only guluronate-configured addition product **21** was isolated in 45% yield (Table 3, entry 4)

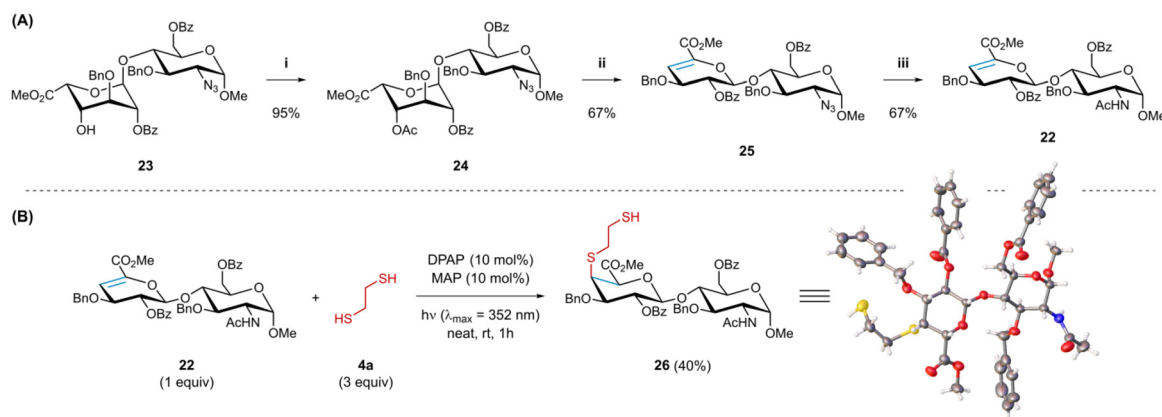
demonstrating potential for compatibility of the methodology in functionalising free sugars.

Thiol–ene reaction on 4,5-unsaturated disaccharide

As outlined in the introduction, a key motivation for this research was the development of a methodology that would enable facile tagging of the 4,5-unsaturated terminal residues of di- and oligosaccharide products from lyase activity in biological samples. As further proof of concept towards this objective, we selected the disaccharide **22**, a synthetic heparan sulfate building block, to test with the thiol–ene conditions (Scheme 3).

The 4,5-unsaturated disaccharide **22** was synthesised from the commercially available building block **23** (Scheme 3A). Acetylation of the free hydroxyl group at the non-reducing terminus under standard conditions (Ac_2O , pyridine) afforded intermediate **24**, which was subsequently converted into the 4,5-enose derivative **25** via β -elimination of the acetyl group at C4. To prevent unwanted side reactions, such as thiol-mediated azide reduction during the photoinitiated hydrothiolation, the azide group at C2 of the reducing end of **25** was modified into an acetamide group using thioacetic acid in the presence of 2,6-lutidine, following a previously reported method.⁴³

Gratifyingly, hydrothiolation of **22** using the optimised thiol–ene conditions furnished the thioether product **26** in 40% yield (Scheme 3B). A trace quantity of a second product was detected by TLC but the quantity proved insufficient for isolation and analysis. The colourless, plate-shaped crystals, obtained during the isolation step (column chromatography, hexane/acetone 60 : 40 solvent system), allowed the determination of the absolute configuration of the monosaccharide units in disaccharide **26** by X-ray analysis. Thus, the modification of the 4,5-unsaturated terminal residue furnished the β -D-galacturonate ($^4\text{C}_1$ conformation), as observed previously with monosaccharides, and the methyl α -D-glucopyranoside unit remained unaltered, highlighting the potential for derivatisation of complex oligosaccharides.



Scheme 3 (A) Synthesis of 4,5-unsaturated disaccharide **22**. (B) Application of the photoinitiated thiol–ene methodology. Reaction conditions: (i) Ac_2O , pyridine, rt, 2 h; (ii) DBU, CH_2Cl_2 , 0 °C to rt, 2 h; (iii) thioacetic acid, 2,6-lutidine, CHCl_3 , 60 °C, 36 h.



Conclusions

The thiol–ene reaction has been demonstrated on sterically challenging 4,5-unsaturated derivatives of uronic acids together with complete regioselectivity and good diastereoselectivity. The thermodynamically favoured axial attack of the photoinitiated thiyl radical appears to be the dominant process and is tolerant of a wide range of conditions and substrates. Of particular relevance is the finding that solvent-free conditions are optimal for thiol–ene reactions on 4,5-unsaturated sugars, since these conditions are scalable and have proven to be operationally simple in terms of work-up. The removal of solvents from organic reactions is important in the context of sustainable synthesis. The approach is compatible with fully protected, partially protected and fully unprotected monosaccharides. Finally, we have demonstrated how the methodology can be applied to a relevant disaccharide, which may allow fluorescent labelling, utilising a tag and modify approach, to ensue.⁴⁴ Further work on modification of oligosaccharide structures *via* thiol–ene ligation is ongoing in our laboratories.

Author contributions

Conceptualisation: G. J. M. and E. M. S.; methodology and investigation: A. P.-C., H. M., T. M., L. B., J. T. M., I. P. and I. R. G.; data curation: A. P.-C., H. M., T. M., I. P., I. R. G. and B. T.; visualization: A. P.-C. and E. M. S.; writing – original draft: A. P.-C., H. M. and E. M. S.; writing – review and editing: A. P.-C., H. M., T. M., I. P., G. J. M. and E. M. S.; supervision: G. J. M. and E. M. S.; funding acquisition: G. J. M. and E. M. S.

All authors have given approval to the final version of the manuscript.

Data availability

The data supporting this article have been included as part of the ESI:†

General methods, synthetic procedures and characterisation data for all compounds.

NMR spectra for all new compounds and X-ray crystallographic data for **5a**, **6a'**, **6b**, and **26**.

Crystallographic data for **5a**, **6a'**, **6b**, and **26** has been deposited at The Cambridge Crystallographic Data Centre under CCDC accession codes 2389449 (**5a**), 2389448 (**6a'**), 2389447 (**6b**) and 2389446 (**26**).†

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

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References

- 1 Z. J. Witzak and J. M. Culhane, *Appl. Microbiol. Biotechnol.*, 2005, **69**, 237–244.
- 2 A. Czubatka-Bienkowska, J. Sarnik and T. Poplawski, *Carbohydr. Res.*, 2023, **533**, 108934.
- 3 T. Morikawa, K. Nimomiya, G. Tanabe, G. Matsuda, M. Yoshikawa and O. A. Muraoka, *J. Nat. Med.*, 2021, **75**, 449–466.
- 4 J. Porter, A. R. Noble, N. Signoret, M. A. Fascione and G. J. Miller, *ACS Omega*, 2024, **9**, 31703–31713.
- 5 M. Fiore, A. Marra and A. Dondoni, *J. Org. Chem.*, 2009, **74**, 4422–4425.
- 6 S. Staderini, A. Chambery, A. Marra and A. Dondoni, *Tetrahedron Lett.*, 2012, **53**, 702–704.
- 7 A. Dondoni and A. Marra, *Chem. Soc. Rev.*, 2012, **41**, 573–586 and references cited therein.
- 8 T. Xiong, R. Xie, C. Huang, X. Lan, N. Huang and H. Yao, *J. Carbohydr. Chem.*, 2021, **40**, 401–439.
- 9 A. Borbás, *Chem. – Eur. J.*, 2020, **26**, 6090–6101 and references cited therein.
- 10 M. Császár, D. Eszenyi, E. Mező, L. Lázár, N. Debreczeni, M. Tóth, L. Somsák and A. Borbás, *Int. J. Mol. Sci.*, 2020, **21**, 573.
- 11 A. Malone and E. M. Scanlan, *Org. Lett.*, 2013, **15**, 504–507.
- 12 L. McSweeney, F. Dénès and E. M. Scanlan, *Eur. J. Org. Chem.*, 2016, 2080–2095.
- 13 J. Porter, D. Parisi, T. Miller, A. N. Cheallagh and G. J. Miller, *Carbohydr. Res.*, 2023, **530**, 108854.
- 14 J. Porter, M. A. Lima, I. Pongener and G. J. Miller, *Carbohydr. Res.*, 2023, **524**, 108759.
- 15 G. Goti, *ChemCatChem*, 2022, **14**, e202200290.
- 16 D. W. Abbott, H. J. Gilbert and A. B. Boraston, *J. Biol. Chem.*, 2010, **285**, 39029–39038.
- 17 D. Ndeh, A. Rogowski, A. Cartmell, A. S. Luis, A. Baslé, J. Gray, I. Venditto, J. Briggs, X. Zhang, A. Labourel, N. Terrapon, F. Buffetto, S. Nepogodiev, Y. Xiao, R. A. Field, Y. Zhu, M. A. O'Neill, B. R. Urbanowicz, W. S. York, G. J. Davies, D. W. Abbott, M.-C. Ralet, E. C. Martens, B. Henrissat and H. J. Gilbert, *Nature*, 2017, **544**, 65–70.
- 18 N. Afratis, C. Gialeli, D. Nikitovic, T. Tseggenidis, E. Karousou, A. D. Theocharis, M. S. Pavão, G. N. Tzanakakis and N. K. Karamanos, *FEBS J.*, 2012, **279**, 1177–1197.
- 19 A. Joshi, P. Chopra, A. Venot and G.-J. Boons, *Org. Lett.*, 2024, **26**, 2462–2466.
- 20 S. Zhu, J. Li, R. S. Loka, Z. Song, I. Vlodavsky, K. Zhang and H. M. Nguyen, *J. Med. Chem.*, 2020, **63**, 4227–4255.



- 21 P. Chopra, T. Yadavalli, F. Palmieri, S. A. K. Jongkees, L. Unione, D. Shukla and G.-J. Boons, *Angew. Chem., Int. Ed.*, 2023, **62**, e202309838.
- 22 T. K. Kandola, C. J. Mycroft-West, M. L. Andrade De Lima, A. Turner, C. Gardini, E. Urso, G. J. Miller, A. Bisio, E. A. Yates, M. Guerrini and L. Wu, *bioRxiv*, 2024, preprint, DOI: [10.1101/2024.09.26.615151](https://doi.org/10.1101/2024.09.26.615151).
- 23 Z. Wang, C. Shi, X. Wu and Y. Chen, *Chem. Commun.*, 2014, **50**, 7004–7006.
- 24 C. Przybylski, V. Bonnet and R. R. A. Vivès, *Chem. Commun.*, 2019, **55**, 4182–4185.
- 25 R. T. Taylor and D. P. Poudel, *Reactions*, 2022, **3**, 442–450.
- 26 L. Lázár, M. Csávás, Á. Hadházi, M. Herczeg, M. Tóth, L. Somsák, T. Barna, P. Herczegh and A. Borbás, *Org. Biomol. Chem.*, 2013, **11**, 5339–5350.
- 27 L. Lázár, L. Juhász, G. Batta, A. Borbás and L. Somsák, *New J. Chem.*, 2017, **41**, 1284–1292.
- 28 S.-T. Ma, C.-W. Lee and W.-M. Liu, *RSC Adv.*, 2021, **11**, 18409.
- 29 D. Combaud, M. Thomas, S. Papot and J.-P. Gesson, *Lett. Drug Des. Discovery*, 2005, **2**, 631–637.
- 30 L. Beswick, S. Ahmadipour, J. P. Dolan, M. Rejzek, R. A. Field and G. J. Miller, *Carbohydr. Res.*, 2019, **485**, 107819.
- 31 J. P. Malkinson, R. A. Falconer and I. Toth, *J. Org. Chem.*, 2000, **65**, 5249–5252.
- 32 W. Pilgrim and P. V. Murphy, *J. Org. Chem.*, 2010, **75**, 6747–6755.
- 33 B. M. Sattelle, B. Bose-Basu, M. Tessier, R. J. Woods, A. S. Serianni and A. Almond, *J. Phys. Chem. B*, 2012, **116**, 6380–6386.
- 34 F. Dénès, M. Pichowicz, G. Povie and P. Renaud, *Chem. Rev.*, 2014, **114**, 2587–2693.
- 35 Y. Araki, K. Matsuura, Y. Ishido and K. Kushida, *Chem. Lett.*, 1973, **2**, 383–386.
- 36 B. H. Northrop and R. N. Coffey, *J. Am. Chem. Soc.*, 2012, **134**, 13804–13817.
- 37 V. Kelemen, M. Bege, N. Debreczeni, I. Bereczki, A. C. Bényei, P. Herczegh and A. Borbás, *Chem. – Eur. J.*, 2025, **31**, e202500104.
- 38 H. Yu and X. Chen, *Org. Lett.*, 2006, **8**, 2393–2396.
- 39 M. Lopez, J. Trajkovic, L. F. Bornaghi, A. Innocenti, D. Vullo, C. T. Supuran and S.-A. Poulsen, *J. Med. Chem.*, 2011, **54**, 1481–1489.
- 40 M. M. Muthana, J. Qu, M. Xue, T. Klyuchnik, A. Siu, Y. Li, L. Zhang, H. Yu, L. Li, P. G. Wang and X. Chen, *Chem. Commun.*, 2015, **51**, 4595–4598.
- 41 R. A. Edington, E. L. Hirst and E. E. Percival, *J. Chem. Soc.*, 1955, 2281–2288.
- 42 R. N. Rej, J. N. Glushka, W. Chewand and A. S. Perlin, *Carbohydr. Res.*, 1989, **189**, 135–148.
- 43 N. Shangguan, S. Katukojvala, R. Greenberg and L. J. Williams, *J. Am. Chem. Soc.*, 2003, **125**, 7754–7755.
- 44 M. Singh, M. Watkinson, E. M. Scanlan and G. J. Miller, *RSC Chem. Biol.*, 2020, **1**, 352–368.

