

COMMUNICATION

View Article Online
View Journal



Cite this: DOI: 10.1039/d5ob00683j

Received 26th April 2025,

Accepted 13th May 2025

DOI: 10.1039/d5ob00683j

rsc.li/obc

Superacid-catalysed α -deuteration of ketones with D_2O [†]

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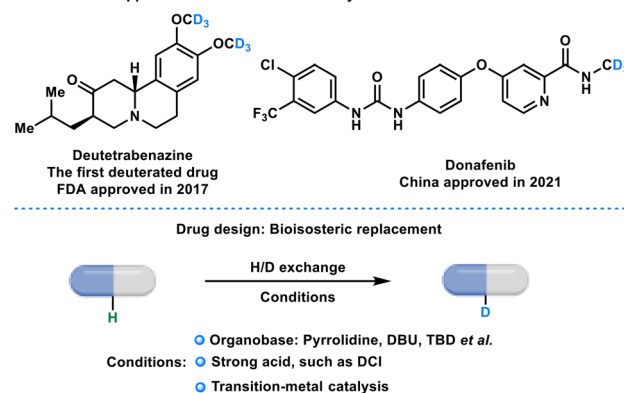
In this study, we present a superacid catalyzed protocol for the α -deuteration of ketones with D_2O using $[Ph_3C]^+[B(C_6F_5)_4]^-$ as a pre-catalyst to generate *in situ* the superacidic species $[D]^+[B(C_6F_5)_4]^-$. The features of this catalytic process include simple manipulation, high deuteration efficiency (up to 99%), excellent functional group compatibility and a broad substrate scope, including 30 substrates comprising common building blocks and bioactive molecules like pentoxifylline. Moreover, the avoidance of toxic reagents enables sustainable access to deuterated products, demonstrating the method's practical potential for use in mechanistic studies and pharmaceutical applications.

Deuterium-labeled compounds play a pivotal role in pharmaceutical research, mechanistic studies, and synthetic chemistry due to their unique kinetic isotope effects and broad applications.¹ In recent years, deuterated drugs have attracted significant attention in pharmaceutical science,² with several deuterated drugs being approved and introduced to the market,³ including deutetrabenazine, the first deuterated drug approved by the FDA in 2017, and donafenib, first approved in 2021. In addition, numerous deuterated drug candidates are currently in clinical development, such as d_1 -NJ38877605, an ATP-competitive inhibitor of c-Met.⁴

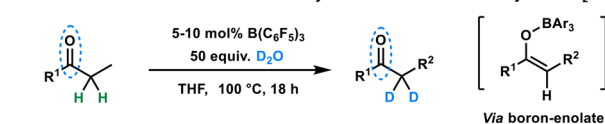
In modern drug discovery, bioisosteric replacement remains a fundamental strategy for lead compound optimization and bioactivity enhancement. Keto groups are ubiquitous functional motifs in natural products and drugs. Therefore, the hydrogen–deuterium exchange (H/D exchange) at the α -position of carbonyl groups is of particular significance in drug design. For instance, CTP-499, a phase II clinical

drug, which is a deuterated compound derived from the metabolite of pentoxifylline (HDX), demonstrated improved safety, enhanced absorption, and reduced interindividual variability compared to HDX.⁵ Recent advancements in selective H/D exchange have focused on organobase-catalyzed H/D exchange⁶ and strong acid mediated deuteration.⁷ Additionally, transition metal-catalyzed H/D exchange has also

A. Deuteration application in medicinal chemistry



B. Wasa's work: Frustrated Lewis Pair catalyzed α -deuteration of carbonyls with D_2O



C. This work: Metal-free catalytic α -deuteration of ketones with D_2O

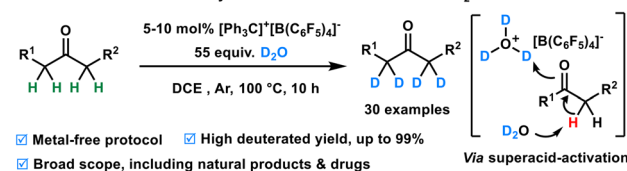


Fig. 1 (A) Deuteration application in medicinal chemistry. (B) Wasa's work: frustrated Lewis pair catalyzed α -deuteration of carbonyls with D_2O . (C) This work: metal-free catalytic α -deuteration of ketones with D_2O .

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[†]Electronic supplementary information (ESI) available. See DOI: <https://doi.org/10.1039/d5ob00683j>



been recognized as an alternative approach (Fig. 1A).⁸ However, these processes often suffer from several limitations, such as the need for harsh conditions, scalability challenges, poor functional group tolerance, and potential toxicity concerns associated with metal residues.

More recently, Wasa and co-workers developed an efficient α -deuteration of carbonyl compounds with D₂O under metal-free conditions, leveraging the catalytic ability of frustrated Lewis pairs (Fig. 1B).^{7a} Building upon our previous research on triaryl carbenium ion-pair mediated organic transformations,⁹ we herein describe a metal-free catalytic protocol for H/D exchange at the α -position of keto moieties, enabling the synthesis of a series of deuterated products with potential bioactivity. In addition, this method exhibits high deuteration efficiency, broad functional group compatibility and substrate versatility, accommodating both ketones and structurally complex molecules (Fig. 1C).

Based on our work on catalytic H/D exchange of arenes with D₂O,¹⁰ we started our investigation using pentoxifylline (**1**) as the model substrate, employing trityl tetrakis(pentafluorophenyl)borate ([Ph₃C]⁺[B(C₆F₅)₄][−]) as the catalyst with D₂O as the deuteration reagent to optimize the reaction conditions. Finally, we achieved the desired deuterated product **2** (pentoxifylline-*d*₅) in 93% yield with up to 96% deuterium incorporation at the α -positions. The optimal reaction conditions included the use of 5 mol% [Ph₃C]⁺[B(C₆F₅)₄][−], 55 equiv. D₂O (0.2 mL), at 100 °C in 10 hours under an argon atmosphere (Table 1, entry 1). Control experiments provided crucial insights, confirming the essential role of the ion-pair in facilitating this metal-free deuteration process efficiently (Table 1, entry 2). When [Ph₃C]⁺[BF₄][−] replaced [Ph₃C]⁺[B(C₆F₅)₄][−], the

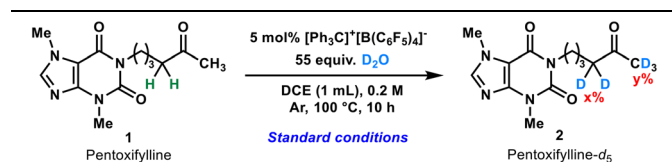
expected deuteration was not observed due to weaker anion basicity (Table 1, entry 3), underscoring the significant influence of the anion. Furthermore, the use of K⁺[B(C₆F₅)₄][−] resulted in substantially reduced deuteration efficiency, affording the desired product in 86% yield (Table 1, entry 4). When dichloromethane (DCM) was tested as an alternative solvent to dichloroethane (DCE), deuterium incorporation at the α -position of the keto group was observed at 91% and 96%, respectively (Table 1, entry 5). Consequently, DCE was chosen as the optimal solvent for the subsequent reaction optimization, owing to its high boiling point that helped drive efficient and robust hydrogen–deuterium exchange. Also, deuterated methanol (CD₃OD) was used as the deuterium source to conduct the hydrogen–deuterium exchange reaction under the standard conditions. Comparative analysis revealed only a marginal decrease in keto deuteration efficiency at the α -position (85% and 89%), while the isolated yield was maintained at consistently high levels of 95% (Table 1, entry 6).

A slight decrease in reaction temperature reduced the deuteration efficiency, affording **2** in 90% and 93% deuterated yields (Table 1, entry 7). When the reaction time was reduced to 8 hours, the deuterated product **2** was obtained in 89% yield (Table 1, entry 8). Notably, strict atmospheric control proved essential, as oxygen-rich conditions significantly compromised reaction efficiency, leading to 50% yield of the deuterated product with lower deuterium incorporation, likely due to competing oxidative side reactions (Table 1, entry 9).¹¹

With the optimal reaction conditions in hand, we next explored the substrate scope to demonstrate the versatility and functional group tolerance of this deuteration method across diverse molecular architectures (Scheme 1). First, natural products and drugs with keto moieties were successfully deuterated to yield the corresponding deuterated product at the α -position of keto groups (**2–11**). For instance, pentoxifylline (**2**), tonalide (**3**), nootkatone (**4**), geranylacetone (**5**), and dihydro- β -ionone (**6**) were converted into deuterated products with high levels of deuterium incorporation at expected positions, underscoring the feasibility of direct deuteration of bioactive molecules without the need for complex protection/deprotection strategies. Moreover, our strategy demonstrated exceptional performance in the regioselective deuteration of steroidal and polycyclic frameworks (**7–11**), achieving over 95% deuterium incorporation at diverse positions. Notably, deuterium labeling of these structurally complex scaffolds presents significant advantages for pharmaceutical development and drug discovery, highlighting the potential application of these labeled molecules while maintaining the integrity of functional groups, including hydroxyls, lactones, and alkenes.

Subsequently, we evaluated the catalytic performance of this protocol using acetophenone derivatives with a wide range of electronic properties, and noted that this method exhibited excellent functional group tolerance. Electron-donating groups (–Me, –*n*-Bu, and –OMe), halogen groups (–F, –Cl, and –Br) and electron-withdrawing (–CF₃, –NO₂, –CN, –COOMe, –SO₂Me, and –Bpin) substituents (**12–23**) were well tolerated, with 76%–99% deuterium incorporation in 70%–97% yields, highlighting

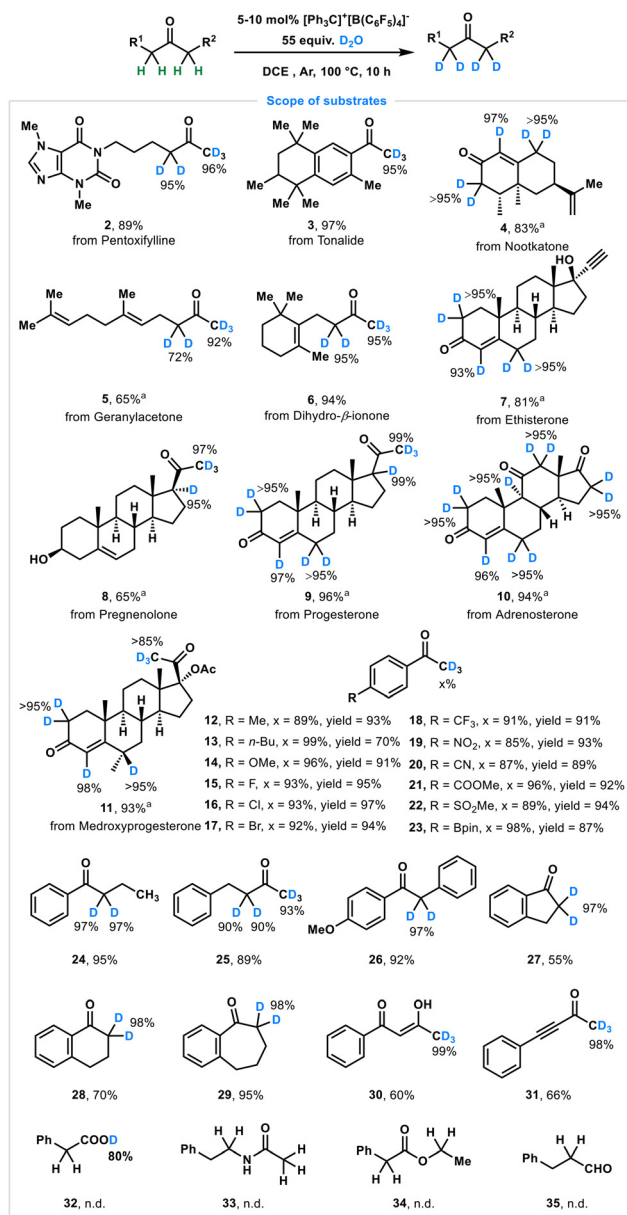
Table 1 Optimization of reaction conditions^a



Entry	Variation from 'standard conditions'	<i>x</i> ^b (%)	<i>y</i> ^b (%)	Yield ^c (%)
1	None	95	96	93 (89 ^d)
2	No [Ph ₃ C] ⁺ [B(C ₆ F ₅) ₄] [−]	n.d.	n.d.	99
3	[Ph ₃ C] ⁺ [BF ₄] [−] instead of [Ph ₃ C] ⁺ [B(C ₆ F ₅) ₄] [−]	n.d.	n.d.	90
4	K ⁺ [B(C ₆ F ₅) ₄] [−] instead of [Ph ₃ C] ⁺ [B(C ₆ F ₅) ₄] [−]	40	34	86
5	DCM instead of DCE	91	96	86
6	CD ₃ OD instead of D ₂ O	89	85	95
7	90 °C instead of 100 °C	90	93	94
8	8 h instead of 10 h	89	93	86
9	O ₂ instead of argon	50	50	82

^a Reaction conditions: pentoxifylline (0.1 mmol, 27.8 mg), [Ph₃C]⁺[B(C₆F₅)₄][−] (5 mol%, 0.005 mmol), D₂O (55 equiv., 0.2 mL), DCE (1 mL), 100 °C, argon, in 10 hours. ^b Yield of deuteration was based on hydrogen, indicated in red. ^c All yields are NMR yields using CH₃NO₂ as an internal standard. ^d Isolated yields. n.d.: no deuteration.





Scheme 1 Scope of substrates. Reaction conditions: substrates (0.1 mmol), $[\text{Ph}_3\text{C}]^+[\text{B}(\text{C}_6\text{F}_5)_4]^-$ (5 mol%), D_2O (55 equiv., 0.2 mL), DCE (1 mL), 100 °C, argon, in 10 hours. ^a DCE : DMSO (1 : 1) as co-solvent. All yields are isolated yields. n.d.: no deuteration.

the general compatibility of this method. Other ketones, including 1-phenylbutan-1-one (24), 4-phenylbutan-2-one (25), 1-(4-methoxyphenyl)-2-phenylethan-1-one (26), indanone (27), 1-tetralone (28) and 1-benzosuberone (29), underwent selective deuteration efficiently to produce the expected products in 55%–95% yields with up to 98% deuterium incorporation. Additionally, the enolate of ketone was also compatible with our metal-free deuteration to give 30 in 60% yield with 99% deuterium labeling. Alkynones, which were recently employed in bioconjugation for protein modification by our group and others,¹² underwent deuterium labeling at the methyl group to

generate 31 in 98% deuteration yield. Additionally, some typical carbonyl compounds, such as carboxylic acids, esters, and aldehydes, were evaluated for H/D exchange under standard conditions. Unfortunately, none of these underwent hydrogen–deuterium exchange (32–35).

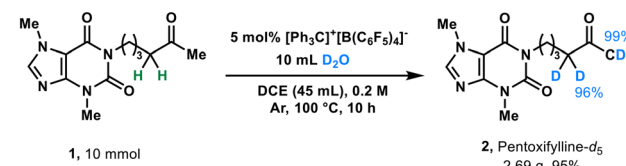
To demonstrate the practical utility and scalability of our metal-free deuteration protocol, we scaled up the synthesis of deuterated pentoxifylline- d_5 at the gram level, using 10 mL of D_2O as the deuterium source. The product was obtained in high yield without the need for column chromatographic purification, highlighting the potential application of our method (Scheme 2A).

Interestingly, the deuterium incorporation was reversible.

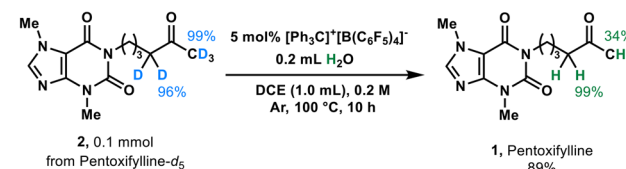
As shown in Scheme 2B, D/H exchange with deionized water was achieved when deuterated pentoxifylline- d_5 was used as the substrate and treated under slightly modified conditions for 24 hours. Moreover, when 1.0 equiv. Brønsted acids, such as AcOH, HCl, H_2SO_4 , TFA and TfOH, were employed instead of the catalytic amount of 5 mol% $[\text{Ph}_3\text{C}]^+[\text{B}(\text{C}_6\text{F}_5)_4]^-$ ion pair catalyst, only minimal amounts of the deuterated products were observed, showcasing the distinct activity of the ion pair in facilitating the desired deuteration process (Scheme 2C). Comparative experiments revealed that the superacid-catalyzed method reported here demonstrated clear advantages over stoichiometric base-catalyzed protocols (e.g., using 1.0 equivalent of K_2CO_3 as the catalyst). (For further details, please refer to the ESI.†)

As shown in Scheme 3, the proposed mechanism began with the hydrolysis of the triaryl carbenium ion-pair in an

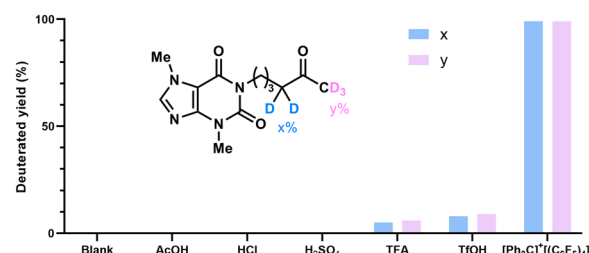
A. Scale-up to 10 mmol for deuteration of Pentoxifylline



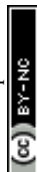
B. Reverse D/H exchange using d -Pentoxifylline

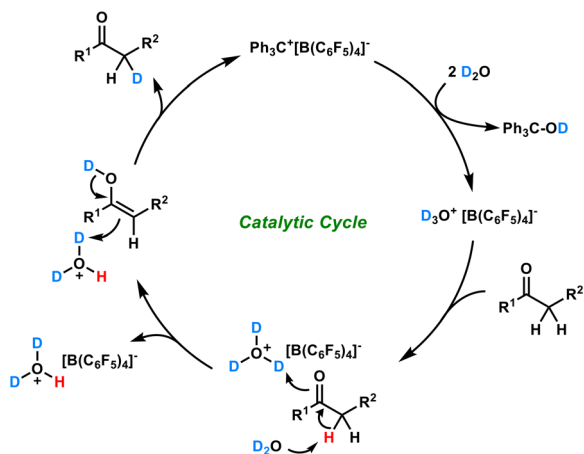


C. H/D exchange using 1.0 eq. Brønsted acids vs. 5 mol% $[\text{Ph}_3\text{C}]^+[\text{B}(\text{C}_6\text{F}_5)_4]^-$



Scheme 2 Practical application and mechanistic studies.





Scheme 3 Proposed mechanism.

aqueous medium, generating the hydrated proton complex $(D_2O)_nD^+[B(C_6F_5)_4]^-$ along with triphenylmethanol as a byproduct. In the presence of ketone substrates, the oxygen atom of the keto group underwent rapid protonation, leading to the formation of the corresponding enol tautomer. This reactive intermediate subsequently tautomerized back to the keto structure, simultaneously regenerating the superacid species. This catalytic cycle enabled single H/D exchange at the α -position, and through the cumulative effect of deuterium isotope exchange, ultimately achieved complete deuteration of the ketone compound at the α -position.

In summary, we have developed a highly efficient and selective metal-free catalytic H/D exchange strategy for precise deuterium incorporation across a broad range of keto-containing pharmaceuticals, ketones, sterically hindered frameworks, and bioactive steroids. This approach exhibits excellent functional group tolerance and scalability, making it a valuable tool for the synthesis of deuterium-labeled compounds with significant applications in pharmaceutical research, drug metabolism studies, and mechanistic investigations of organic reactions.

Data availability

The data underlying this study are available in the published article and its ESI.[†]

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

We gratefully acknowledge the financial support provided by the Distinguished University Professor Grant (Nanyang Technological University) and the Agency for Science,

Technology, and Research (A*STAR) under its MTC Individual Research Grant (M21K2c0114) and RIE2025 MTC Programmatic Fund (M22K9b0049) to T.-P.L.

References

- (a) E. M. Simmons and J. F. Hartwig, *Angew. Chem., Int. Ed.*, 2012, **51**, 3066–3072; (b) J. Atzrodt and V. Derdau, *J. Labelled Compd. Radiopharm.*, 2010, **53**, 674–685.
- (a) R. M. C. Di Martino, B. D. Maxwell and T. Pirali, *Nat. Rev. Drug Discovery*, 2023, **22**, 562–584; (b) S. L. Harbeson and R. D. Tung, *Med. Chem.*, 2014, **2**, 8–22; (c) T. G. Gant, *J. Med. Chem.*, 2014, **57**, 3595–3611; (d) T. Pirali, M. Serafini, S. Cargnin and A. A. Genazzani, *J. Med. Chem.*, 2019, **62**, 5276–5297; (e) A. Mullard, *Nat. Rev. Drug Discovery*, 2016, **15**, 219–221.
- (a) A. Mullard, *Nat. Rev. Drug Discovery*, 2017, **16**, 305; (b) S. Qin, F. Bi, S. Gu, Y. Bai, Z. Chen, Z. Wang, J. Ying, Y. Lu, Z. Meng, H. Pan, P. Yang, H. Zhang, X. Chen, A. Xu, C. Cui, B. Zhu, J. Wu, X. Xin, J. Wang, J. Shan, J. Chen, Z. Zheng, L. Xu, X. Wen, Z. You, Z. Ren, X. Liu, M. Qiu, L. Wu and F. Chen, *J. Clin. Oncol.*, 2021, **39**, 3002–3011; (c) A. Mullard, *Nat. Rev. Drug Discovery*, 2022, **21**, 623–625; (d) S. H. DeWitt and B. E. Maryanoff, *Biochemistry*, 2018, **57**, 472–473.
- (a) X. Li, E. K. Dowling, G. Yan, Z. Dereli, B. Bozorgui, P. Imanirad, J. H. Elnaggar, A. Luna, D. G. Menter, P. G. Pilié, T. A. Yap, S. Kopetz, C. Sander and A. Korkut, *Cancer Discovery*, 2022, **12**, 1542–1559; (b) K. Sanderson, *Nature*, 2009, **458**, 269.
- (a) V. Braman, P. Graham, C. Cheng, D. Turnquist, M. Harnett, L. Sabounjian and J. Shipley, *Clin. Pharmacol. Drug Dev.*, 2013, **2**, 53–66; (b) M. V. Perez-Gomez, M. D. Sanchez-Niño, A. B. Sanz, C. Martín-Cleary, M. Ruiz-Ortega, J. Egido, J. F. Navarro-González, A. Ortiz and B. Fernandez-Fernandez, *J. Clin. Med.*, 2015, **4**, 1325–1347.
- (a) C. Rape and W. H. Sachs, *Tetrahedron*, 1968, **24**, 6287–6290; (b) C. Sabot, K. A. Kumar, C. Antheaume and C. Mioskowski, *J. Org. Chem.*, 2007, **72**, 5001–5004; (c) M. Rempt and G. Pohnert, *Angew. Chem., Int. Ed.*, 2010, **49**, 4755–4758; (d) S. Peng, N. M. Okeley, A.-L. Tsai, G. Wu, R. J. Kulmacz and W. A. van der Donk, *J. Am. Chem. Soc.*, 2002, **124**, 10785–10796; (e) K. I. Galkin, E. G. Gordeev and V. P. Ananikov, *Adv. Synth. Catal.*, 2021, **363**, 1368–1378; (f) C. Berthelette and J. Scheigetz, *J. Labelled Compd. Radiopharm.*, 2004, **47**, 891–894; (g) M. Zhan, T. Zhang, H. Huang, Y. Xie and Y. Chen, *J. Labelled Compd. Radiopharm.*, 2014, **57**, 533–539; (h) T. Tanaka, Y. Koga, Y. Honda, A. Tsuruta, N. Matsunaga, S. Koyanagi, S. Ohdo, R. Yazaki and T. Ohshima, *Nat. Synth.*, 2022, **1**, 824–830.
- (a) Y. Chang, T. Myers and M. Wasa, *Adv. Synth. Catal.*, 2020, **362**, 360–364; (b) M. Shang, X. Wang, S. M. Koo, J. Youn, J. Z. Chan, W. Yao, B. T. Hastings and M. Wasa, *J. Am. Chem. Soc.*, 2017, **139**, 95–98.



- 8 (a) J. Krüger, B. Manmontri and G. Fels, *Eur. J. Org. Chem.*, 2005, 1402–1408; (b) B. M. Wheatley and B. A. Keay, *J. Org. Chem.*, 2007, 72, 7253–7259.
- 9 (a) Z. Zhang, X. Liu, L. Ji, T. Zhang, Z. Jia and T.-P. Loh, *ACS Catal.*, 2022, 12, 2052–2057; (b) Z. Zhang, L. Ji, X. Liu, Z. Li, Y. Lv, Z. Jia and T.-P. Loh, *Org. Chem. Front.*, 2022, 9, 5154–5159.
- 10 Z. Zhang, Y. Lv, W. Q. R. Ong, X. Zhao, Z. Jia and T.-P. Loh, *Angew. Chem., Int. Ed.*, 2024, 63, e202408509.
- 11 (a) Z. Zhang, J. Gu, Y. Lv, L. Ji, X. Liu, B. Wu, F. Liu, Z. Jia and T.-P. Loh, *Cell Rep. Phys. Sci.*, 2023, 4, 101246; (b) Z. Zhang, J. Gu, L. Ji, X. Liu, T. Zhang, Y. Lv, F. Liu, Z. Jia and T.-P. Loh, *ACS Catal.*, 2022, 12, 14123–14129; (c) Z. Zhang, Y. Lv, L. Ji, P. Chen, S. Han, Y. Zhu, L. Li, Z. Jia and T.-P. Loh, *Angew. Chem., Int. Ed.*, 2024, 63, e202406588.
- 12 (a) S. Teng, E. W. H. Ng, Z. Zhang, C. N. Soon, H. Xu, R. Li, H. Hirao and T.-P. Loh, *Sci. Adv.*, 2023, 9, eadg4924; (b) Z. Zhang, L. Li, H. Xu, C.-L. K. Lee, Z. Jia and T.-P. Loh, *J. Am. Chem. Soc.*, 2024, 146, 1776–1782.

