View Article Online View Journal

Soft Matter

Accepted Manuscript

This article can be cited before page numbers have been issued, to do this please use: L. G. Vaniyan, P. Kumar Borah, G. E. Pavlovskaya, N. J. Terrill, J. E. S. J. Reid, M. W. Boehm, P. Prochasson, R. A. Nicholson, S. K. Baier and G. E. Yakubov, *Soft Matter*, 2025, DOI: 10.1039/D4SM00705K.



This is an Accepted Manuscript, which has been through the Royal Society of Chemistry peer review process and has been accepted for publication.

Accepted Manuscripts are published online shortly after acceptance, before technical editing, formatting and proof reading. Using this free service, authors can make their results available to the community, in citable form, before we publish the edited article. We will replace this Accepted Manuscript with the edited and formatted Advance Article as soon as it is available.

You can find more information about Accepted Manuscripts in the Information for Authors.

Please note that technical editing may introduce minor changes to the text and/or graphics, which may alter content. The journal's standard <u>Terms & Conditions</u> and the <u>Ethical guidelines</u> still apply. In no event shall the Royal Society of Chemistry be held responsible for any errors or omissions in this Accepted Manuscript or any consequences arising from the use of any information it contains.



rsc.li/soft-matter-journal

Wet Spinning of Sodium Carboxymethyl Cellulose - Sodium Mo0705K Caseinate Hydrogel Fibres: Relationship between Rheology and Spinnability

4

Lathika Vaniyan^a, Pallab Kumar Borah^{a,f}, Galina E. Pavlovskaya^b, Nick Terrill^c, Joshua E.S.J.
Reid^a, Michael Boehm^d, Philippe Prochasson^d, Reed A. Nicholson^d, Stefan Baier^{d,e}, Gleb E.
Yakubov*^{a,g}

8

This article is licensed under a Creative Commons Attribution 3.0 Unported Licence.

Article. Published on 24 February 2025. Downloaded on 2/24/2025 10:45:03 PM.

Open Access

- 9 ^aFood Materials Research Group, University of Nottingham, Sutton Bonington, LE12 5RD,
 10 United Kingdom
- ¹¹ ^bSir Peter Mansfield Imaging Centre, University of Nottingham, Nottingham, NG7 2RD,
 ¹² United Kingdom
- ¹³ ^cDiamond Light Source, Harwell Science and Innovation Campus, Didcot, OX11 0DE, United
 ¹⁴ Kingdom
- 15 ^dMotif FoodWorks Inc, 27 Drydock Avenue, Boston, MA 02210, USA.
- ⁶School of Chemical Engineering, University of Queensland, Brisbane, QLD 4072, Australia.
 ^fHeinz Maier-Leibnitz Zentrum, Technical University of Munich, Lichtenbergstraβe 1, 85748,
 Germany
- 19 gFood Biopolymers Laboratory, School of Food Science and Nutrition, University of Leeds,
- 20 Leeds, LS2 9JT, United Kingdom
- 21
- 22 *Corresponding author: Gleb E. Yakubov, Professor of Food Biopolymers, School of Food
- 23 Science and Nutrition, University of Leeds, Leeds, LS2 9JT, United Kingdom.

View Article Online DOI: 10.1039/D4SM00705K

This article is licensed under a Creative Commons Attribution 3.0 Unported Licence.

39

Open Access Article. Published on 24 February 2025. Downloaded on 2/24/2025 10:45:03 PM.

25 Abstract

26 Mimicking the fibrous structures of meat is a significant challenge as natural plant protein 27 assemblies lack the fibrous organisation ubiquitous in mammalian muscle tissues. In this work, 28 wet-spun hydrogel fibres resembling the anisotropic fibrous microstructure of meat are 29 fabricated using carboxymethyl cellulose as a model polysaccharide and sodium caseinate as a 30 model protein which are crosslinked using 1-ethyl-3-(3-dimethylaminopropyl) carbodiimide 31 (EDC). Hydrogels and spun fibres were characterised using a combination of rheology (shear, 32 oscillatory, and extensional), microscopy (light, polarised, and fluorescence), rheo-NMR, and 33 x-ray diffraction. Examination of structuring behaviour under shear uncovered a relationship 34 between enhanced biopolymer orientation along the fibre axis and a viscoelastic time-35 dependent ageing window for optimal hydrogel spinnability. This study provides novel rheological and structural insights into mechanisms of protein-polysaccharide assembly that 36 may prove instrumental for development of tuneable fibres for applications in plant-based 37 38 foods, tissue engineering, and biomaterials.

40 Keywords: fibre spinning; carboxymethyl cellulose; hydrogel; plant-based meat analogue;
41 anisotropy; rheo-NMR

1. Introduction.

Soft Matter

The growing demand for sustainable, protein-rich foods requires the development of 43 innovative technologies to mimic the texture of meat-based products using plant-based 44 45 ingredients^{1, 2}. The distinctive fibrous texture of meat stems from the arrangement of collagen 46 fibres and myofibrils in the muscle tissue. However, creating analogous fibrous structures using 47 plant-based proteins poses a marked challenge, as most commercial plant proteins, such as those derived from soy or pea, lack the inherent fibrous organisation in their native state^{3, 4}. 48 49 The use of hydrogel fibres is rapidly expanding across food and biomaterial applications, 50 including the alternative meat products, where fibres have been used for imparting meat-like texture attributes to the plant-based analogue products⁵. Equally, fibres are routinely used to 51 52 design scaffolds for cellular-agriculture applications, also known as lab-grown meat⁶. The 53 fibre-based structures are favoured due to high versatility, tuneability, and a wide spectrum of mechanical behaviours that fibres can impart to the final products^{7, 8}. In addition, the inherent 54 ability to hold large amounts of water provides high structuring efficiency using small amounts 55 56 of fibre material, thus providing cost-effective solutions for food reformulation and material 57 design⁹.

Despite these advantages, progress in the spinning of fibres remains limited, 58 59 particularly with respect to spinnability and scalability for industrial applications, hampering 60 its widespread adoption, with the food industry being in a particularly challenging position due 61 to stringent requirements of cost, performance, scalability, and safety. Recently, both wet and dry spinning techniques for hydrogel fibres have gained attention. For instance, Bordignon and 62 63 coworkers explored the spinning of low molecular weight gels and demonstrated that altering 64 the molecular structure of carbohydrates allows fragile hydrogels to be wet-spun, making them suitable for 3D printing applications¹⁰. Lundahl and coworkers investigated the impact of shear 65 66 and extensional viscosities on carbon nanofibrils and found that improved spinnability was Soft Matter Accepted Manuscript

Soft Matter Accepted Manuscript

Open Access Article. Published on 24 February 2025. Downloaded on 2/24/2025 10:45:03 PM.

associated with increased shear viscosity, storage modulus, and extensional viscosity 67 Rheological studies of polyacrylonitrile (PAN)-carbon nanotube (CNT) dispersions carried out 68 by Lu and coworkers revealed that increasing CNT concentration enhances elastic-like 69 70 behaviour and shear thinning, alongside fibre spinning performance improving for lower molecular weight PAN and similar rheological properties observed in PAN/CNT dispersions 71 at high filler loading¹². Tan and coworkers¹³ studied the effects of temperature, coagulation 72 conditions, and non-solvent on the spinnability of polyacrylonitrile-dimethyl sulfoxide 73 74 solutions, showing that wet spinning is strongly influenced by the temperature and 75 concentration of the coagulating bath, while dry spinning is primarily affected by the air gap. It was also found that addition of non-solvent such as water deteriorated the quality of wet spun 76 77 fibre. Sharma and coworkers studied the extensional rheology of weakly elastic, polymeric 78 complex fluids, by characterising their extensional relaxation time and extensional viscosity¹⁴⁻ ¹⁶. By analysing the elastocapillary self-thinning, they have established the relationship 79 between extensional relaxation time and polymer concentration¹⁷. 80

Despite these advancements, optimising fibre formation remains challenging¹⁸. A 81 particularly challenging aspect is achieving a balance between extensibility and structural 82 integrity during the sol-gel transition¹⁹. This balance should be considered across the length 83 scales, starting from the molecular level and extending to micro- and macrostructures. 84 85 Understanding the fundamental rheological and crosslinking properties that govern this 86 transition is essential, as they ultimately determine the final structure and mechanical 87 characteristics of fibrous hydrogels. These properties are key for final applications across 88 foods, pharmaceuticals, and (bio)materials.

In this work, we aim to address some of these challenges. One of the key targets is to identify hydrogel formulations and crosslinking conditions where gel-setting properties are optimised to allow the formation of consistent and uniform protein-polysaccharide fibres. For

this purpose, we utilise a model binary, weakly associating biopolymer system containing/hereaticle Online 92 sodium salt of carboxymethyl cellulose (NaCMC) and sodium caseinate (NaCas) which is 93 crosslinked into a hydrogel using 1-ethyl-3-[3-dimethylaminopropyl] carbodiimide 94 95 hydrochloride (EDC). Sodium carboxymethyl cellulose is a water-soluble polysaccharide, a cellulose derivative that is widely used in industrial applications across foods, hygiene 96 97 products, pharmaceuticals and materials. This is due to tuneable and well-defined viscosifying 98 and, more broadly, rheological properties, biocompatibility, biodegradability, and crosslinking 99 abilities²⁰⁻²³. NaCas is a protein derived from milk^{24, 25}. Its secondary protein structure features a high content of random coil configurations (~ 20%)^{25, 26}, which makes it effective at tethering 100 NaCMC chains due to higher conformational flexibility²⁷ (i.e., as compared to tightly folded 101 102 globular proteins such as, for example, lysozyme²⁸. Previous studies on carboxymethyl 103 cellulose and sodium caseinate hydrogels have primarily focussed on the bulk gels and their viscoelastic properties^{26, 29-31}. Although a wide range of cross-linking and complexation 104 mechanisms have been explored^{20, 32} which includes physical interactions³³, irradiation³⁴, use 105 of multi-valent metal ions³⁵⁻³⁷ and low-molecular weight crosslinkers^{23, 38-40}, little is known 106 about the mechanisms of fibre formation when fibre spinning is performed under transient 107 108 crosslinking conditions i.e., when the cross-linking reaction is not fully completed. To 109 modulate the conditions of the crosslinking reaction, we systematically vary the concentrations of the tethering molecule (NaCas) and the crosslinker (EDC). By decoupling the two key 110 111 factors of cross-linking, i.e., tether density and the speed of crosslinking, we attain the 112 possibility of adjusting and probing the dynamic balance between extension of the polymer 113 network in a sol state during spinning and its relaxation during the transition into a hydrogel.

We hypothesise that the emergence of anisotropic characteristics of biologic fibrous materials is associated with the alignment of polysaccharide chains in extensional flow⁴¹, which is subsequently stabilised by covalent bonds between the polysaccharide and protein (*i.e.*, Soft Matter Accepted Manuscript

Soft Matter Accepted Manuscript

supressing chain relaxation upon shear cessation). To probe and scrutinise this hypothesis were added and a scrutinise this hypothesis were added and a scrutinise this hypothesis were added as the second added and a scrutinise the second added and a scrutinise the second added adde 117 employed a range of rheological methods, including steady shear rotational rheometry, small 118 amplitude oscillatory shear rheometry, and capillary breakup and extensional rheometry 119 120 (CABER). These techniques were used to probe viscoelastic properties of the hydrogels. 121 Polarised light microscopy has been used to reveal the effect of crosslinking conditions on the 122 microstructure that formed during fibre spinning process. The structural characteristics on the 123 molecular level have been probed using X-ray diffraction (XRD) and rheology coupled to sodium nuclear magnetic resonance (rheo-²³Na-NMR) spectroscopy^{42, 43}. Our results highlight 124 125 the importance of rheological characteristics in the fibre spinning process and provide a deeper understanding of the factors that govern spinnability which remains underexplored, especially 126 127 in the context of the controlled formation of fibrous structures with tuneable mechanical 128 properties. As such, our work aims to provide foundational insights into designing plant-based 129 fibrous structures for food applications, which could extend to fields like biomedical 130 scaffolding and sustainable packaging.

132 **2. Experimental**

133 **2.1. Materials**

134 1-ethyl-3-(3-dimethylaminopropyl) carbodiimide (EDC, 99%) and calcofluor white stain were purchased from Sigma-Aldrich, UK. Sodium caseinate (NaCas, > 92% protein) was purchased 135 136 from Thermo Scientific Chemicals. Sodium carboxymethyl cellulose (NaCMC) was supplied 137 by CP Kelco, Norway (CEKOL 4000: M_w, 450 kDa; DS, 0.8). Molecular weight was verified using intrinsic viscosity $[\eta]$ measurements (details of experimental procedure is in 138 Supplementary methods S1 and S2). The $[\eta]$ was found to be ~18 dL g⁻¹ in 100 mM NaCl 139 140 leading to an estimated M_w of 340 - 470 kDa and is consistent with the information provided by the manufacturer (Supplementary Information S1 and Supplementary Figure S1). Milli-O 141 water (Millipore Corp., USA) was used throughout the experiments (18.2 M Ω .cm ionic purity 142 at 25 °C). All experiments were carried out at 25 °C. 143

144 2.2. Preparation of NaCMC-NaCas hydrogels

145 Hydrogels were prepared at different concentrations of NaCas (0.1, 0.3, 0.5, 1, and 2 wt.%) in 0.5 wt.% NaCMC, using the 'zero-length' crosslinker, EDC (0, 1, 5, 10, 20 and 50 mM) (Table 146 147 1). Briefly, the mechanism of the crosslinking reaction comprises a step of: (i) EDC forming an unstable reactive ester upon interaction with a carboxyl group of the polysaccharide, and 148 149 (ii) the intermediate ester then interacts with a primary amine of protein to form a peptide 150 bond⁴⁴. For the formation of hydrogels, NaCas and NaCMC solutions were prepared separately in 50 mL deionized water. EDC was added to the NaCMC solution at pH 4.5 with continuous 151 stirring for 60 sec, followed by addition of the NaCas solution. The pH was lowered to 4.5 to 152 153 ensure optimal EDC activity and stability of the intermediates formed during the crosslinking 154 process.

Soft Matter Accepted Manuscript

	View	Article	Online
DOI:	10.1039/D	4SM0()705K

Variables	Concentrations
NaCMC	0.5%
EDC	0 mM
	1 mM
	5 mM
	10 mM
	20 mM
	50 mM
NaCas	0.1%
	0.3%
	0.5%
	1%
	2%

156 Table 1. Concentrations of EDC and NaCas used with 1 wt.% NaCMC.

NaCas, Sodium caseinate; EDC, 1-ethyl-3-(3-dimethylaminopropyl) carbodiimide; NaCMC, Sodium carboxymethyl cellulose. Note, percentages are wt.%.

160 **2.3. Steady-state shear and oscillatory rheology**

This article is licensed under a Creative Commons Attribution 3.0 Unported Licence.

157

158

159

Open Access Article. Published on 24 February 2025. Downloaded on 2/24/2025 10:45:03 PM.

161 The flow behaviour and viscoelastic properties of the hydrogel were measured on a MCR 301 rheometer (Anton Paar GmbH, Austria) equipped with a Peltier temperature control system. A 162 163 concentric cylinder geometry (CC27; outer diameter, 28.9 mm; inner diameter, 26.66 mm; gap size, 1.13 mm; cone angle, 120°; effective cylinder height, 39.997 mm) was used for testing 164 bulk solutions and gel samples. Amplitude sweeps were performed with a constant angular 165 166 frequency, $\omega = 10$ rad s⁻¹, to identify the linear viscoelastic regime (Figure S2). Optimum angular frequency, ω and shear strain, γ were determined by performing frequency sweeps for 167 sample containing NaCMC and NaCas in 1:1 ratio, in the frequency range, $\omega = 1 - 100$ rad s⁻ 168 169 ¹, at constant strains of 1%. Time-dependent oscillatory shear experiments were carried out 170 with varying concentrations of NaCas and EDC (Table 1) to identify the optimum 171 concentrations for crosslinking and fibre spinnability. Measurements were carried out within 172 the linear viscoelastic range to ensure that sample properties were not affected by the imposed strain or stress. Experiments were performed at a constant angular frequency, $\omega = 6.28$ rad s⁻ 173 174 ¹, and constant strain, 1% for 120 min. For constructing a crosslinking diagram for biopolymer

175 mixtures, the G' values at t = 100 min of the reaction were plotted as a function of concentration $f_{M00705K}$ 176 of EDC and NaCas.

177 2.4. Capillary breakup and extensional rheology

Extensional capillary breakup tests were performed using a CaBER-1 extensional rheometer (Thermo Scientific Haake, Germany) equipped with an enclosed measuring unit to minimise evaporation, as described in our earlier study⁴⁵. For all measurements (n = 5), 76 µL of sample was utilised in the parallel geometry (diameter, 6 mm; initial gap, 3.01mm, and final gap, 9.92 mm). The initial and final aspect ratio were 1 and 3.31, respectively, corresponding to a Hencky strain of 1.19. Hydrogels were tested at different time intervals to determine the effects of timedependent ageing on crosslinking *via* analysis of capillary thinning and breakup^{15, 17, 18}.



186 Figure 1. Changes in the extensional behaviour of a liquid bridge filament as a function of crosslinking time. During extensional flow measurements, the liquid sample is placed between 187 two plates of the CaBER apparatus (plate diameter, 6 mm, initial height, ca. 3 mm); then the 188 189 upper movable plate rapidly opens the gap, allowing the formation of a filament (final height, ca. 10 mm). Initially (t = 2 min), breakup is rapid as expected for low viscosity, Newtonian-190 191 like fluids. With the increase in crosslinking time, the filament breakup slows down, indicating the growing contribution of fluid elasticity. At t = 30 min of the crosslinking reaction, the 192 193 filament remains sustained between the plates without breaking. As the crosslinking reaction 194 continues, the formation of the filament represents the uniaxial extension of a viscoelastic gel. 195 Ultimately, the strength of the gel exceeds the adhesion force between the plate and the sample, 196 making the formation of a filament unfeasible.

197

198 **2.5. Sodium rheo-NMR**

View Article Online DOI: 10.1039/D4SM00705K

199 A non-magnetic cone and plate geometry with 8° cone angle and 12 mm diameter (Bruker, 200 Germany) was placed inside the 25 mm dual channel ²³Na/¹H resonator (Bruker, Germany) 201 tuned to 105.68 MHz and 400.18 MHz corresponding to sodium and proton resonance 202 frequencies, respectively. Using a specialised shaft, the assembly was positioned in the centre 203 of the 9.4T superconducting magnet and connected to the shear control unit (RheoSpin, Bruker, 204 Germany). Shear rate in all rheo-NMR experiments was controlled using automated TopSpin software and maintained within +/- 0.01 s⁻¹. The employed shear rates were 10.89, 21.00, and 205 206 30.00 s¹. Sodium detection in rheo-NMR experiments was performed using two-dimensional 207 triple quantum time proportional phase increment method (TQ-TPPI) with 1024 complex 208 points sampled in the direct dimension and 512 points in the indirect dimension with 50 µs increment steps. The width of sodium $\frac{\pi}{2}$ was 50 µs, recycle delay was 100 ms, and a total 209 210 time for a TQTPPI scan was under 5 min. Further details of this method are provided elsewhere^{42, 46}. 211

212 **2.6. X-ray diffraction**

This article is licensed under a Creative Commons Attribution 3.0 Unported Licence.

Open Access Article. Published on 24 February 2025. Downloaded on 2/24/2025 10:45:03 PM.

NaCMC-NaCas crosslinked fibres under different time-dependent ageing and crosslinking conditions were studied using an Echo D8 Advance Bruker AXS powder diffractometer (Bruker, UK) equipped with a copper tube operating at a tube voltage of 40 kV and an accelerating current of 25 mA, using Cu-K α radiation at a wavelength of 0.1541 nm and controlled by DIFFRAC EVA software. The diffraction range (2 θ) was 4 to 45°, with a step size of 0.02° (n = 3).

219 2.7. Wet Spinning of hydrogel fibres

NaCMC-NaCas fibres were spun using a syringe pump (KD Scientific, USA), as described
 previously by Li and coworkers⁴⁷, with some modifications. The experimental setup for

229

234

spinning hydrogel fibres is illustrated in Figure 2. Briefly, fibres were spun from hydrocolligitate online
mixtures containing varying concentration of sodium caseinate (0.5, 0.75, 1 and 1.5 wt.%),
EDC (10, 15, 20 and 30 mM), and 0.5 wt.% NaCMC. The flow rate was fixed at 0.4 mL min⁻¹
for all experiments. Acetone was used as coagulant. After spinning, the fibres were air-dried.
Additionally, the time-dependent ageing of hydrogels was studied using a representative
hydrocolloid mixture containing 0.5 wt.% of NaCMC, 0.5 wt.% of NaCas (*i.e.*, 1:1 ratio) and
20 mM EDC).



Figure 2. (a) Experimental set up for spinning hydrogel fibres using a syringe Pump (KD
Scientific, USA): (b) Visual representation of hydrogel filament obtained after spinning 0.5
wt.% NaCMC-NaCas solution, 30 minutes after onset of crosslinking (0.4 mL min⁻¹, 21G
needle, ID 800 µm).

235 **2.8. Microscopy**

Bright field and polarised light micrographs of the fibre samples were obtained using an Eclipse Ci-POL microscope (Nikon, Japan). Fluorescence micrographs were obtained on a EVOS FL microscope (Life technologies, USA) using the DAPI light cube ($\lambda_{ex} = 350$ nm; $\lambda_{em} = 440$). Briefly, calcofluor white stain was prepared in water at a concentration of 0.1 mg mL⁻¹. Fibre samples were stained for 1 min before observation under the microscope. Mean diameter of fibres were estimated (n = 5) using ImageJ software (NIH, USA). Microscope images have

- been adjusted to improve contrast and brightness to accurately represent the data or to high in the bright of the
- specific features of interest. The original images are provided in the Figure S3 and S4.

This article is licensed under a Creative Commons Attribution 3.0 Unported Licence.

()

Open Access Article. Published on 24 February 2025. Downloaded on 2/24/2025 10:45:03 PM.

3. Results and Discussion

246 The ability to successfully wet-spin hydrogel fibres is highly dependent on optimising the formulation parameters, particularly the concentration of biopolymer components and the 247 248 rheological properties of the precursor solution⁴⁸. Previous studies have demonstrated that an optimal balance of viscoelasticity, surface tension, and shear-thinning behaviour is required to 249 ensure continuous fibre formation and structural integrity during extrusion^{12, 49, 50}. Building 250 251 upon these insights, we explored the effect of hydrogel composition on spinnability by varying 252 systematically the concentrations of a tether (NaCas) and a crosslinker (EDC). The choice of a 253 model system has been instrumental in studying the relationship between viscoelastic properties of hydrogels and fibre spinnability. Using carboxymethyl cellulose (NaCMC) as a 254 255 model polysaccharide and sodium caseinate (NaCas) as a model protein, we have formulated 256 a crosslinked hydrogel system with tuneable gel-setting properties. This is achieved by 257 independently varying the concentration of tethers that determines cross-linking density and 258 the concentration of a crosslinker, which influences the kinetics of the crosslinking reaction 259 (Due to a size difference of approximately two orders of magnitude between EDC (\sim 5Å) and NaCas (~ 500Å), it is assumed that EDC has much higher diffusivity compared to NaCas). 260

Our hypothesis suggests that under specific spinning conditions, the extension of polysaccharide molecules will occur, resulting in stable fibres with an anisotropic molecular and microstructure which resembles collagen and myofibril. The concept of anisotropic fibre formation is particularly important in this context, as it replicates the structural organisation observed in natural fibres like collagen, which achieve high tensile strength through their aligned molecular arrangement as reported by Li and coworkers⁷.

267 **3.1. Rheological properties of the hydrogel**

268 The rheological properties of NaCMC-NaCas crosslinked hydrogels were assessed in 269 the concentrations ranging from semi-dilute state to concentrated solution state⁵¹, covering

relevant regions for technical applications. To determine the relationship between biopolymetrice online concentration and hydrogel formation, the crosslinking process was monitored using small amplitude oscillatory shear (SAOS) rheology as a function of time. This is in agreement with the work of Lopez and coworkers $^{20, 51, 52}$. SAOS measurements yielded values of *G*' and *G*'' as a function of time and formulation. Monitoring the evolution of *G*' enables assessment of the changes in crosslinking density (ν) of the hydrogel as^{53, 54}:

$$v_c = \frac{G'_C}{RT} \tag{1}$$

where *R* is the gas constant, *T* is the absolute temperature and G'_C is the storage modulus of the gel. Using the above equation, it is possible to understand the relative change in crosslinking density by normalising the storage modulus as:

This article is licensed under a Creative Commons Attribution 3.0 Unported Licence.

276

280

Open Access Article. Published on 24 February 2025. Downloaded on 2/24/2025 10:45:03 PM.

$$v = (G' - G'_0) / G'_0 \tag{2}$$

where, *G'* is the storage modulus of the gel at time = *t* and *G'*₀ is the storage modulus of the gel at time *t* = 0. The changes in crosslinking densities over time for various concentrations of NaCas and EDC are presented in Figure S5, provide important insights into the dynamics of hydrogel formation. The storage modulus of the gel can be correlated with the strength S_{ω_0} of the gel at a given angular frequency, ω_0 as⁵⁵:

$$S_{\omega_0} = G'_{\tan\delta = 1} \left(\frac{1}{2}\pi\omega_0\right)^{-1/2}$$
(3)

where $G'_{\tan\delta=1}$ is the value of G' at the time point of its intersection with intersects G''. The gel strength, S_{ω_0} is also proportional to the degree of crosslinking that enables monitoring the crosslinking reaction by measuring the corresponding time evolution of G' (data not shown). This article is licensed under a Creative Commons Attribution 3.0 Unported Licence.

289

291

293

295

296

298

301 302

303

3

Open Access Article. Published on 24 February 2025. Downloaded on 2/24/2025 10:45:03 PM.



290 Figure 3. (a) Evolution of the small amplitude oscillatory shear (SAOS) rheological behaviour of NaCMC-NaCas crosslinked hydrogel (1:1 ratio of protein and cellulose gum) with 20 mM 292 EDC showing changes in storage modulus (G'), loss modulus (G'') and tan δ values during gelation as a function of time. The sol-gel transition time is achieved at tan $\delta = 1$ when G' 294 = G''. This transition time can vary depending on the concentration of crosslinking agent or concentration of biopolymers. Black dashed line is a visual guide to show tan $\delta = 1$. (b) 'Protorheological'⁵⁶ inference of (a) as a function of crosslinking time. Here, 'zones' 297 correspond to the following crosslinking times: 0 - 20 min (Zone 1), 20 - 40 min (Zone 2), 40 - 60 min (Zone 3), 60 - 80 min (Zone 4), and 80 - 100 min (Zone 5). (c) Variation in storage 299 modulus (G') with concentration of EDC and NaCas (G' at $\omega = 6.28$ rad sec⁻¹) for NaCMC-300 NaCas hydrogel at cross-linking time t = 100 mins. (d) Variation in tan δ as a function of concentration of EDC and NaCas. Solid black line is a visual guide to denote the tan $\delta = 1$ boundary.

304 Figure 3a shows the corresponding values of the storage, G' and loss moduli, G'' as a 305 function of reaction time for NaCMC crosslinked with NaCas mixed in 1:1 ratio using 20 mM EDC. Prior to the crosslinking reaction, G'' is greater than G', indicating that the mixture is in 306 307 the free-flowing state. The crossover point of the loss and storage moduli after 30 min of 308 reaction, represents the sol-to-gel transition and it can be used to define the onset point of

Soft Matter Accepted Manuscript

crosslinking/gelation (G' = G'' or $G''/G' = \tan \delta = 1$). At the end of the reaction the $G'_{10.1}$ we sticle Online 309 were found to be significantly greater as compared to G'' indicating gel-like behaviour. At this 310 311 point in time, the mixture sets to form a self-supporting gel. Figure 3a shows the tan δ values with respect to time and depicts that the gelation point is obtained at 35 min where tan $\delta = 1$. 312 'Protorheological'⁵⁶ inference of the sample as a function of crosslinking time are shown in 313 314 Figure 3b, and clearly shows the emergence of the storage, G' moduli in the gels. Here, 'zones' 315 refer to the ranges of crosslinking times and compositions that represent specific phases in the hydrogel's rheo-mechanical evolution: **Zone 1**: $G' \ll G''$, rapid filament breakup; **Zone 2**: G'316 < G'', slowly thinning filament; **Zone 3**: G' > G'' stable filament; **Zone 4**: G' >> G'', filament 317 dominated by elastic extension; **Zone 5**: G' >> G'', elastic extension. 318

319 To map the crosslinking reaction as a function of composition, the G'values at t = 100min of crosslinking were plotted against the concentration of NaCas and EDC. The variation 320 321 in the G' values over time with increasing EDC concentrations is plotted in the Figure S6. Figure 3c shows the comparison of storage modulus at t = 100 min with increasing 322 concentrations of both NaCas and EDC. The variation in G' and G'' over time for different of 323 324 NaCas concentrations, specific to each EDC concentration are plotted separately in the Figure S7. In un-crosslinked systems the loss modulus (G'') value was greater than the storage 325 modulus (G'). As the gel crosslinks the storage modulus increases (G'' < G'). It was observed 326 327 that crosslinking occurred at a minimum concentration of 10 mM EDC with 1 wt.% NaCas and 328 50 mM EDC with 0.3 wt.% NaCas. The storage modulus and crosslinking density increase in 329 tandem with increasing concentration of EDC and NaCas. Figure 3d shows that the tan δ value decreased with increasing concentration of EDC and NaCas indicating the formation of 330 crosslinked gel. At low concentrations of NaCas and EDC, there was a delay in the onset of 331 332 crosslinking, while at higher concentrations, the gel sets rapidly, leaving very little window for

This article is licensed under a Creative Commons Attribution 3.0 Unported Licence.

3

336

337

338

339

340

341

342 343

Open Access Article. Published on 24 February 2025. Downloaded on 2/24/2025 10:45:03 PM.

Soft Matter



Figure 4. Normalised filament diameter as a function of time for crosslinking hydrogel with a total polymer concentration of 1 wt.% and 20 mM EDC. Early filament thinning and breakage was observed in weakly crosslinked polymer while completely crosslinked polymers exhibited no filament formation. (b) Characteristic relaxation time (λ_E) from CaBER experiments as a function of crosslinking time obtained by fitting the exponential phase of CaBER data. Red dashed box is a visual guide to indicate the evidence of percolation threshold behaviour. Error bars represent n = 5. Fitted curves for extensional relaxation time, λ_E are shown in Figure S8.

We also studied the extensional flow behaviour of the hydrogels using capillary 344 breakup and extensional rheology (CaBER)⁵⁷. Capillary breakup is widely understood as a 345 346 surface tension induced breakup of filaments at low concentrations of the crosslinked solution 347 which determines the lower limit of spinnability^{58, 59}. The time t = 0 is defined as the time at 348 which the upper plate has reached its final position (Hencky strain, 1.19). It was observed that 349 the hydrogel starts crosslinking immediately after sample preparation and forms a highly 350 viscous gel within two hours of preparation. The gel showed marked extensional properties for 351 a long period of time at a concentration of NaCMC and NaCas at 1:1 ratio with 20 mM of EDC. 352 The gel formed is highly flexible within the range of 30 min to 60 min after which it sets completely. Figure 4a shows the evolution of fibre diameter on the CaBER with respect to 353

Soft Matter Accepted Manuscript

Open Access Article. Published on 24 February 2025. Downloaded on 2/24/2025 10:45:03 PM. (cc) **BY** This article is licensed under a Creative Commons Attribution 3.0 Unported Licence.

time. The extensional relaxation time, λ_E were determined from the exponential function λ_E were determined from the exponential λ_E

356

$$D(t) = D_o exp\left(-\frac{t}{3\lambda}\right) \tag{4}$$

where, D is the diameter of the thinning capillary and D_0 is the diameter of the thinning 357 capillary at time, t = 0. One can clearly observe the evolution of the hydrogel from completely 358 359 un-crosslinked to completely crosslinked as a function of time. Changes in relaxation time of 360 the gel in response to crosslinking time provides further evidence of percolation and threedimensional network formation as the sol-to-gel transformation takes place⁶¹, and is shown in 361 Figure 4b. When G'' > G' in the system, the gel network is weakly formed, and the relaxation 362 time is lower than the percolation threshold value $(r < r_c)$. When $r > r_c$; G' > G'' and a 363 364 plateau region is observed, providing indications that the polymer network has completely 365 developed. We have provided a visual depiction of the network formation as a function of the 366 crosslinking time within insets in Figure 4b.

367 3.2. Microstructural analysis of hydrogel fibres

Hydrogel fibres were spun at a constant flow rate of 0.4 mL min⁻¹ at different 368 369 crosslinking times using a syringe pump. Microscopic images of EDC crosslinked NaCMC-370 NaCas fibres were analysed to determine the average diameter of the fibres spun at different time intervals. Figure 5a displays the variation in diameters of fibres spun at various time 371 372 intervals following the initiation of crosslinking. The results show that the fibre diameter 373 increased with the increase in polymer concentration and crosslinking time. The variation in 374 fibre diameter at different concentration regimes is displayed in Figure 5b. Smooth fibres were 375 obtained by wet spinning NaCMC-NaCas hydrogel within the interval of optimum spinnability 376 as discussed in section 3.1. These changes can be attributed to the increase in the extensional viscosity of the biopolymer mixture. The wet spun hydrogel fibres were coagulated in an 377



This article is licensed under a Creative Commons Attribution 3.0 Unported Licence.

380

381

382 383

384

385

386

387

388

389

390

ÅЗ

3

Open Access Article. Published on 24 February 2025. Downloaded on 2/24/2025 10:45:03 PM.





Figure 5. (a) Images of hydrogel fibres formed at different crosslinking times viewed using an EVOS fluorescent microscope. Fibre diameters were calculated using ImageJ (NIH, USA). *Star marks within the plot are relative to the corresponding *marks in the associated fluorescent micrographs. (b) Fibre diameter in different concentration regimes. The weak hydrogels formed at lower concentrations resulted in thin filaments. By contrast, rapid gelation at higher polymer concentrations led to the formation of fibres with irregular thickness. Inset is a rendition of Figure 3d, but with added demarcations for different regimes. Regimes I-IV indicates transition from viscosity-dominated to elasticity-dominated as a function of NaCas and EDC concentrations. Fluorescent micrographs show spun fibres at regimes I and IV.

391 The polarized images of NaCMC-NaCas crosslinked fibres alongside comparative 392 optical micrographs at different rheo-mechanical zones during the crosslinking process are 393 shown in Figure 6. Refractive index of the spun fibres was greater than air, evidenced by the 394 presence of Becke lines with positive relief (Figure 6, highlighted by arrows). Increased crosslinking time results in thicker fibres and the surface of the fibre transitioned from smooth 395 396 to irregular with scalloped edges. As can be seen, within the zones of optimum spinnability 397 (particularly Zone 3), fibres demonstrated strong birefringence, *i.e.*, anisotropy in the 398 transmission of light. The fibre appears to have a principle linear optical axis along the major 399 axis, where the polarised light is disintegrated into slow and fast components which were not

Soft Matter Accepted Manuscript

409

411

412

413 414

in phase with each other. Note, the extinction events upon rotation of the sample, stage/levents/le 400 401 shown in Figure S9. Here, the transmission intensity is a function of the angle, θ that the fibre principal linear axis makes with the axis of polarisation; extinction is observed when $\theta \approx$ zero 402 or $\pi/2$, whereas transmission is high when $\theta \approx \pi/4$ (transmitted light intensity $\approx \sin^2(2\theta)$). 403 404 Fibres within Zones 1, 2, and 5 did not demonstrate extinction as a function of θ , whereas a strong effect of θ was observed for fibres within Zones 3 and 4; the latter is indicative of fibre 405 406 anisotropy plausibly with nematic ordering. Previous studies have shown similar results for 407 fibril alignment in collagen fibres^{62, 63}.



Figure 6. Polarised optical micrographs ($\theta \approx \pi/4$) of crosslinked NaCMC-NaCas fibres spun at different rheo-mechanical zones during crosslinking. Zones correspond to crosslinking time 410 as: 0 - 20 min (Zone 1), 20 - 40 min (Zone 2), 40 - 60 min (Zone 3), 60 - 80 min (Zone 4), and 80 - 100 min (Zone 5). White arrows are a visual guide indicating Becke lines. Scale bar is 100µm. $\theta \approx \text{zero or } \pi/2$ are shown in Figure S9.

3.3. Short-range molecular order of hydrogel fibres 415

416 Powder x-ray diffraction (XRD) enables ascertaining molecular orientation in biopolymer fibres and was utilised to characterise such systems as spider silk⁶⁴ and binary 417 polysaccharide gels based on xanthan gum⁶⁵. In this work XRD is used to investigate the 418 419 emergence of molecular ordering in the NaCMC-NaCas crosslinked fibres as a function of crosslinking time. Figure 7a shows XRD diffraction patterns of pure NaCMC and hydrogel 420

This article is licensed under a Creative Commons Attribution 3.0 Unported Licence.

Open Access Article. Published on 24 February 2025. Downloaded on 2/24/2025 10:45:03 PM.

Soft Matter

fibres spun at different zones of the sol-gel transition. The data show the presence $_{DOf} a_0 sm = 0.07$ and $a_0 sm = 0.0$

For the hydrogel fibres, this peak becomes broader, indicating wider distribution of 424 intermolecular distances. Notably, a marked increase in peak intensity at $2\theta \approx 9^\circ$ was observed 425 426 for the hydrogel fibres compared to NaCMC alone, as well as compared to fibres spun in Zone 427 1. Our findings align with previous research conducted by Souza and coworkers⁵³. The latter 428 are equivalent to quiescently cross-linked gels, because NaCMC chains in these fibres have 429 sufficient time to relax before cross-linking takes place. Thus, we suggest that the peak at $2\theta \approx$ 9° is associated with longer range structure, which gets accentuated by the spinning process 430 431 during fibre manufacture. The ratio of peak areas was used to estimate the degree of long-range 432 structuring as shown in Table 2. The fibres spun within Zone 3 showed the highest degree of long-range structuring, followed closely by Zone 4 with the peak intensity ratio of ~0.1 433 compared to 0 for pure NaCMC. Although specific details of long-range structuring require 434 435 further analysis, it is evident that XRD findings appear to be consistent with the results obtained 436 from polarised microscopy.

437 Table 2. Peak areas and the ratio of area of peaks at $2\theta \approx 9^\circ$ to $2\theta \approx 20^\circ$.

Zone	Peak 1	Peak 2	Ratio
NaCMC	0	14857.73	0
1	270.48	31116.51	0.0087
2	1279.19	38632.77	0.0331
3	2404.99	23910.67	0.1006
4	2911.11	29348.14	0.0992
5	1856.37	32880.64	0.0565

⁴³⁸

To probe the hydrogel structure further, we used sodium rheo-NMR - a versatile and sensitive technique which provides insights into molecular alignment within sodiumcontaining hydrogel networks under shear⁴⁶. Since both our biopolymers, NaCas and NaCMC, contain Na ions, ²³Na rheo-NMR would provide a means of distinguishing between free and

Soft Matter Accepted Manuscript

Open Access Article. Published on 24 February 2025. Downloaded on 2/24/2025 10:45:03 PM.

bound sodium ions⁴². The triple quantum (TQ) coherence, contribution to sodium NMR symptometers with a soliton of the solito 443 arises from slow-motion states of sodium ions as compared to the freely diffusing ones. Such 444 slow-motion states can arise, for example, from electrostatic binding of sodium ions to 445 446 hydrogel macromolecules. This interaction can be detected using a two-dimensional (2D) scan 447 induced through specific ²³Na TQ-TPPI MR protocol, employed in this work. Figure 7b shows 448 the 2D²³Na TQ-TPPI spectrum, where sodium chemical shift is shown in the direct (horizontal) 449 dimension, while sodium multiple quantum spectra are shown in the indirect (vertical) 450 dimension that separates the single quantum and triple quantum coherences. Analysis of 451 sodium multiple quantum spectra taken at the sodium chemical shift frequency (dotted blue 452 line in Figure 7b) enables monitoring the degree of association of sodium cations with hydrogel 453 biomolecules by analysing the intensity of TQ population under shear. Figure 7c shows sodium 454 multiple quantum (MQ) spectra produced in this manner in the gel under pre-shear, sheared at 11 s⁻¹, 20 s⁻¹, 30 s⁻¹ and post shear conditions. No TQ sodium peaks were detected in the absence 455 of shear. This indicates an isotropic environment of the static crosslinked gel as probed by 456 457 sodium cations. The introduction of shear, however, results in the emergence of distinct TO peaks in the sheared gel system as seen in Figure 7c (highlighted with black arrow). 458



Figure 7. X-ray diffraction curves for NaCMC and hydrogel fibres spun at different
crosslinking times and rheo-mechanical zones: 0 - 20 min (Zone 1), 20 - 40 min (Zone 2), 40 60 min (Zone 3), 60 - 80 min (Zone 4) and 80 - 100 min (Zone 5). Dashed lines are a visual
guide to denote the diffraction peaks. NaCMC, sodium carboxymethyl cellulose. (b) ²³Na TQ-

Page 23 of 33

This article is licensed under a Creative Commons Attribution 3.0 Unported Licence.

Open Access Article. Published on 24 February 2025. Downloaded on 2/24/2025 10:45:03 PM.

Soft Matter

TPPI spectra of 1 wt.% NaCMC-NaCas crosslinked gels: 2D TQ-TPPI map measured wired article Online 30 s⁻¹ with all relative terminology. Dotted blue line is a visual guide denoting the sodium chemical shift frequency. (c) ²³Na multiple quantum (MQ) spectra taken from 2D maps as shown by dashed blue line in (b) at 0 s⁻¹, 11s⁻¹, 20 s⁻¹, 30 s⁻¹, and post shear. MQ spectrum of mix solution of biopolymers without crosslinking agent added and sheared at 30s⁻¹ is shown in red. Please note the absence of TQ peak at the highest shear rate as shown by arrows.

We postulate the emergence of the ²³Na TQ signals to the molecular order or molecular 471 472 alignment formed in the gel at the onset of shear. Tests were also performed on a mixed solution 473 of NaCMC and NaCas, without the addition of the crosslinking agent. As can be seen from 474 Figure 7c (MQ spectrum shown in red), no TQ sodium signal was detected in the mixed 475 biopolymer system under shear. This indicates that molecular alignment must be emerging at the interface of shear gel particles, most likely in a form of an electrical double layer. In the 476 477 mixed, un-crosslinked biopolymer system the ionic interactions lack a distinct interface, 478 making their relaxation more random than in the case of an interfacial layer of aligned counter-479 ions. During shear, the requirement of electroneutrality generates a streaming potential in the 480 direction of flow that separates sodium ions between those in a slow-motion state and freely 481 diffusing ones. Similar effects have been observed before in other biopolymeric fluids⁴⁶ where the formation of molecular order was confirmed by detection of sodium residual quadrupolar 482 483 coupling constant. Although the likely location of the sodium ions in slow-motion states is at 484 the interface between shear-gel particles, it is possible to propose an alternative localisation of 485 electrokinetically trapped states. Considering small amplitudes of TQ signals, the effects of gel 486 friction against the walls of the measuring geometry cannot be excluded⁴⁶.

In summary, the results of this study reveal a clear relationship between the rheological properties of covalently crosslinked NaCMC-NaCas hydrogels and their spinnability. The observed trends suggest that specific rheological parameters - such as viscoelasticity, precursor concentrations, sol-gel transition time, and polymer relaxation behaviour - play critical roles in determining molecular alignment and orientation during the fibre spinning process. In this

- 492 context, a suite of quantifiable parameters provides a valuable toolbox for optimising spinster and a valuable tool
- 493 conditions and enabling the rational design of biopolymer fibres.

4. Conclusions

Soft Matter

495 In this study, a model protein-polysaccharide hydrogel system was designed to probe 496 the effects of crosslinking on fibre spinnability and structural alignment during extension. The 497 EDC-mediated crosslinking of NaCMC and NaCas resulted in the formation of homogenous 498 gel. Hydrogels were prepared in different ratios by varying the concentrations of EDC and 499 NaCas at a constant concentration of NaCMC, 1.0 wt.%. By exploring capillary breakup and 500 rheological behaviour using small amplitude oscillatory shear rheology, a direct connection between the rheological properties of hydrogels and fibre spinnability was uncovered. This 501 502 study demonstrates that fibre spinnability is intricately linked to the evolution of viscoelastic 503 properties and is influenced by the sol-gel transition time. During the sol-to-gel transition, the 504 dynamic properties of the transient molecular networks facilitate the spinning of highly regular and homogeneous fibres. Specifically, for the 1 wt.% NaCMC-NaCas hydrogel, optimal 505 506 spinnability was observed between 40 and 60 min into the crosslinking reaction, where tan $\delta \approx$ 507 1. Notably, evidence of shear-induced anisotropic alignment of polysaccharide chains was 508 identified. This alignment was characterized using polarized light microscopy, XRD, and rheo-509 NMR.

510 Overall, our findings provide a foundational platform for future studies. By exploring 511 the fundamental principles governing spinnability and through careful tailoring of the 512 crosslinking conditions, we have uncovered a dynamic balance between extension and relaxation of weakly associated polymer networks to form aligned fibres. These insights will 513 514 help guiding the development and optimisation of materials tailored to specific applications in 515 fibre spinnability across diverse fields, including applications in foods, drug delivery, wound care scaffolding, tissue engineering⁶⁷, and sustainable packaging. From our perspective, the 516 517 insights generated using a model hydrocolloid system will enable the development of edible 518 fibres designed with food-grade hydrocolloids and food-compatible crosslinking agents.

Soft Matter Accepted Manuscript

View Article Online

DOI: 10.1039/D4SM00705K

519

520

This article is licensed under a Creative Commons Attribution 3.0 Unported Licence.

3

Open Access Article. Published on 24 February 2025. Downloaded on 2/24/2025 10:45:03 PM.

Author Contribution. Lathika Vaniyan, Conceptualisation, Data curation, Formal analysis, 521 522 Investigation, Validation, Visualisation, Writing - original draft, Writing - reviewing & 523 editing; Pallab Kumar Borah, Formal analysis, Investigation, Methodology, Visualisation, 524 Writing - reviewing & editing; Galina E. Pavlovskava, Data curation, Formal analysis, 525 Investigation, Validation, Resources, Writing - reviewing & editing; Nick Terrill, Formal analysis, Investigation, Validation, Writing - reviewing & editing; Joshua E.S.J. Reid, 526 527 Investigation, Writing - reviewing & editing; Michael Boehm, Conceptualisation, Investigation, Writing - reviewing & editing; Philippe Prochasson, Investigation, Writing -528 529 reviewing & editing; Reed A. Nicholson, Investigation, Formal analysis, Writing - reviewing 530 & editing; Stefan Baier, Conceptualisation, Investigation, Formal analysis, Funding 531 acquisition, Writing - reviewing & editing; Gleb E. Yakubov, Conceptualisation, Methodology, 532 Formal analysis, Investigation, Funding acquisition, Resources, Supervision, Project 533 administration, Writing – reviewing & editing. All authors reviewed the manuscript.

534 Conflict of interest. The authors declare that they have no competing financial interests or535 personal relationships that could have appeared to influence the work reported in this paper.

536 Data Availability. The data supporting this article has been included as part of the
537 Supplementary Information.

Acknowledgements. The authors wish to acknowledge funding from Motif FoodWorks Inc.
and Biotechnology and Biological Sciences Research Council (BBSRC) grant BB/T008369/1
(Nottingham Doctoral Training Partnership, Ref. 2604202) and BB/T006404/1. PKB
acknowledges funding received from the European Union's Horizon 2020 research and
innovation programme under the Marie Skłodowska-Curie grant agreement No. 101034266.

- 543 LV and GEY would like to acknowledge Dr Anca Pordea (University of Nottingham) effective Online
- 544 valuable discussions and suggestions.

- 547 DS, degree of substitution
- 548 EDC, 1-ethyl-3-(3-dimethylaminopropyl) carbodiimide
- 549 G', storage Modulus
- 550 G'', loss Modulus
- 551 NaCMC, sodium carboxymethyl cellulose
- 552 NaCas, sodium caseinate
- 553 Rheo-NMR, rheology-nuclear magnetic resonance
- 554 tan δ , loss Factor
- 555 TQF, triple Quantum Filter
- 556 TPPI, time-proportional phase incrementation
- 557 XRD, x-ray diffraction
- 558 λ_E , extensional relaxation time
- 559 ω , angular frequency

View Article Online DOI: 10.1039/D4SM00705K

	561	View Article Online DOI: 10.1039/D4SM00705K
l on 24 February 2025. Downloaded on 2/24/2025 10:45:03 PM. ensed under a Creative Commons Attribution 3.0 Unported Licence.	562	• SPSS interactive graphics 8.0 (1998). Chicago, Ill: SPSS Inc.
	563	• Mateos Pérez, J.M. and Pascau, J. (2013) 'Image processing with imageJ'.
	564	Birmingham, UK: Packt Publishing.
	565	• Software library for Bruker TopSpin NMR Data files' (2016). Washington, D.C: United
	566	States. Dept. of Energy.
	567	• Origin(Pro), Version Number (Version 2022). OriginLab Corporation, Northampton,
	568	MA, USA.
	569	Lectures, meetings & conferences:
	570	• Lathika Vaniyan presented in part at the Food Physics Conference, Institute of Physics
	571	(IOP), London, January 2024.
Publishec icle is lic	572	• Lathika Vaniyan presented in part at the Annual European Rheology Conference
Article. This art	573	(AERC), European Society of Rheology, Leeds, April 2024.
Open Access	574	

576	1.	B. L. Dekkers, R. M. Boom and A. J. van der Goot, Trends in Food Science & Technology, 2018,
577		81 , 25-36.
578	2.	H. Schösler, J. d. Boer and J. J. Boersema, Appetite, 2012, 58 , 39-47.
579	3.	M. Imran and Z. Liyan, European Food Research and Technology, 2023, 249 , 2189-2213.
580	4.	S. Lin, H. E. Huff and F. Hsieh, <i>Journal of Food Science</i> , 2002, 67 , 1066-1072.
581	5.	P. C. Nath, S. Debnath, K. Sridhar, B. S. Inbaraj, P. K. Nayak and M. Sharma, Journal, 2023, 9.
582	6.	A. Singh, V. Kumar, S. K. Singh, J. Gupta, M. Kumar, D. K. Sarma and V. Verma, Cell and Tissue
583		Research, 2023, 391 , 235-247.
584	7.	W. Li, J. Liu, J. Wei, Z. Yang, C. Ren and B. Li, Advanced Functional Materials, 2023, 33.
585		2213485.
586	8	7. Yu. D. Wang and 7. Lu. AIMS Materials Science, 2023. 10 , 1004-1033
587	9	H Khalesi W Lu K Nishinari and Y Fang Advances in Colloid and Interface Science 2020
588	5.	285 102278
580	10	D. Bordignon B. Lonetti C. Coudret D. Poblin P. Joseph I. Malaguin A. Chalard and J.
500	10.	Eitromann, Journal of Colloid and Interface Science, 2021, 602, 222, 242
501	11	M L Lundahl M Porta M Ago M Stading and O L Poias European Polymor Journal 2019
502	11.	100 267 279
502	10	107, 507-576.
595	12.	Wi. Lu, J. Lido, P. V. Guigunje, H. Chang, P. J. Anas-Wonje, J. Ramachandran, V. Breedveid and
594	10	S. Kumar, <i>Polymer</i> , 2021, 215 , 123369.
595	13.	L. Tan, H. Chen, D. Pan and N. Pan, Journal of Applied Polymer Science, 2008, 110 , 1997-
596		
597	14.	J. Dinic, L. N. Jimenez and V. Sharma, <i>Lab on a Chip</i> , 2017, 17 , 460-473.
598	15.	J. Dinic and V. Sharma, Proceedings of the National Academy of Sciences, 2019, 116 , 8766-
599		8774.
600	16.	J. Dinic, Y. Zhang, L. N. Jimenez and V. Sharma, ACS Macro Letters, 2015, 4, 804-808.
601	17.	K. Al Zahabi, L. Hassan, R. Maldonado, M. W. Boehm, S. K. Baier and V. Sharma, Soft Matter,
602		2024, 20 , 2547-2561.
603	18.	L. N. Jimenez, C. D. V. Martínez Narváez and V. Sharma, Physics of Fluids, 2020, 32.
604	19.	L. N. Jimenez, C. D. V. Martínez Narváez and V. Sharma, Macromolecules, 2022, 55, 8117-
605		8132.
606	20.	C. G. Lopez and W. Richtering, Carbohydr Polym, 2021, 267, 118117.
607	21.	M. Hashmi, S. Ullah, A. Ullah, Y. Saito, M. K. Haider, X. Bie, K. Wada and I. S. Kim, Polymers
608		(Basel), 2021, 13 .
609	22.	C. Hou, T. Watanabe, C. G. Lopez and W. Richtering, <i>Carbohydr Polym</i> , 2025, 347 , 122287.
610	23.	H. Kono, <i>Carbohydr Polym</i> , 2014, 106 , 84-93.
611	24.	R. Deschenes Gagnon, L. Bazinet and S. Mikhaylin, <i>Membranes (Basel)</i> , 2022, 12 .
612	25.	Q. Hu. L. Huang, J. Wang, J. Huangfu, Y. Cai, T. Liu, M. Zhao and Q. Zhao, Food Bioscience.
613		2024. 61 . 104570.
614	26.	M. Pereda, G. Amica, I. Rácz and N. F. Marcovich, <i>Carbohydrate Polymers</i> , 2011, 86 , 1014-
615		1021
616	27	Wusigale I Liang and Y Luo Trends in Food Science & Technology 2020 97 391-403
617	27.	S A Rodrigues C Pradal I. VII K I Steadman I. R. Stokes and G. F. Vakubov Food
618	20.	Hydrocolloids 2021 119 106826
619	20	IS Behra I Mattsson O I Cavre E S I Bohles H Tang and T N Hunter ACS Applied
620	25.	Dolumer Materials 2010 1 200.259
621	20	r orginici marchais, 2013, 1, 344-330. D. Wagner S. Bétańska E. Warmbier A. Frankjowicz and I. Bétański, Journal 2022, 16
622	50. 21	C. G. Lonoz and W. Bichtoring, Calluloca, 2010, 26 , 1517, 1524
672	51. 27	A. C. Alayarca E. C. G. Erachini P. da Silva V. H. Lima A. Chavandi and D. E. C. Datri Jat J. Bial
624	52.	A. C. Alavarse, E. C. G. Frachini, K. da Silva, V. H. Lima, A. Shavandi and D. F. S. Petri, <i>Int J Biol</i>
024		IVIULTUTIUT, 2022, 202 , 338-340.

575 **References**

This article is licensed under a Creative Commons Attribution 3.0 Unported Licence.

(cc) BY

Open Access Article. Published on 24 February 2025. Downloaded on 2/24/2025 10:45:03 PM.

View Article Online DOI: 10.1039/D4SM00705K This article is licensed under a Creative Commons Attribution 3.0 Unported Licence.

Open Access Article. Published on 24 February 2025. Downloaded on 2/24/2025 10:45:03 PM.

Soft Matter

View Article Online 625 G. Dürig and A. Banderet, Helvetica Chimica Acta, 1950, 33, 1106-1118. 33. DOI: 10.1039/D4SM00705K 626 34. M. Z. Pengfei Liu, Jiuqiang Li, Jing Peng, Jilan Wu, Radiation Physics and Chemistry 2002, 63, 627 525-528. 628 35. S. C. Lin, Y. Minamisawa, K. Furusawa, Y. Maki, H. Takeno, T. Yamamoto and T. Dobashi, 629 *Colloid and Polymer Science*, 2010, **288**, 695-701. 630 36. L. Fan, K. Peng, M. Li, L. Wang and T. Wang, J Biomater Sci Polym Ed, 2013, 24, 1099-1111. 631 37. Y. Xue, H. Zhang, F. Su, L. Zhang, G. Lang, Y. Zhu, C. Gu, P. Zhou, X. Zhan and D. Liu, 632 Biomacromolecules, 2024, 25, 4867-4878. 633 38. R. Barbucci, A. Magnani and M. Consumi, *Macromolecules*, 2000, 33, 7475-7480. 634 39. P. Toledo, D. Limeira, N. Siqueira and D. Petri, Cellulose, 2019, 26. 635 40. V. S. Ghorpade, A. V. Yadav, R. J. Dias, K. K. Mali, S. S. Pargaonkar, P. V. Shinde and N. S. 636 Dhane, International Journal of Biological Macromolecules, 2018, 118, 783-791. 637 V. Calabrese, T. Porto Santos, C. G. Lopez, M. P. Lettinga, S. J. Haward and A. Q. Shen, 41. 638 Physical Review Research, 2024, 6. 639 42. P. Hanson, C. J. Philp, H. S. Randeva, S. James, J. P. O'Hare, T. Meersmann, G. E. Pavlovskaya 640 and T. M. Barber, JCI insight, 2021, 6. 641 43. X. w. Jia, Z. y. Qin, J. x. Xu, B. h. Kong, Q. Liu and H. Wang, International Journal of Biological 642 Macromolecules, 2020, 157, 641-647. 643 44. R. F. Rivera-Santiago, S. Sriswasdi, S. L. Harper and D. W. Speicher, Methods, 2015, 89, 99-644 111. 645 45. X. Li, Manickavasagan, A., Lim, LT., Ali, A. In: Manickavasagan, A., Lim, LT., Ali, A., Plant 646 Protein Foods. Springer, Cham, 2022, 171-196. 647 46. G. E. Pavlovskaya and T. Meersmann, Soft Matter, 2023, 19, 3228-3237. 648 47. S. Li, M. Chandra Biswas and E. Ford, Carbohydrate Polymers, 2022, 297, 120001. 649 W. Fang, E. Y. Lim, K. L. Nieminen and H. Sixta, ACS Omega, 2023, 8, 34103-34110. 48. 650 S. Xu, Y. Yan, Y. Zhao, X. Qiu, D. Zhuang, H. Liu, X. Cui, J. Huang, X. Wu and C. Huang, Journal 49. 651 of Materials Chemistry C, 2021, 9, 5554-5564. 652 50. F. Javad, M. Azadeh and W. Holly, in Hydrogels, eds. H. Sajjad and H. Adnan, IntechOpen, 653 Rijeka, 2018, DOI: 10.5772/intechopen.74188, p. Ch. 6. 654 51. C. G. Lopez, Journal of Rheology, 2020, 64, 191-204. 655 52. C. G. Lopez, S. E. Rogers, R. H. Colby, P. Graham and J. T. Cabral, J Polym Sci B Polym Phys, 656 2015, **53**, 492-501. 657 53. D. S. d. S. d. Souza, V. A. P. Tartare, B. d. S. Bega, G. C. Zambuzi, T. S. Ribeiro, C. Ribeiro, O. d. 658 Freitas and K. R. Francisco, Colloids and Surfaces A: Physicochemical and Engineering 659 Aspects, 2024, 682. 660 54. J. Huang, P. Fu, W. Li, L. Xiao, J. Chen and X. Nie, RSC Advances, 2022, 12, 23048-23056. 661 55. H. H. W. a. F. Chambon, Journal of Rheology, 1986, 30, 367-382. 662 56. M. T. Hossain and R. H. Ewoldt, Journal of Rheology, 2024, 68, 113-144. 663 57. S. Różańska, K. Verbeke, J. Różański, C. Clasen and P. Wagner, Journal of Polymer Science 664 Part B: Polymer Physics, 2019, 57, 1537-1547. 665 58. J. Dinic, L. N. Jimenez and V. Sharma, *Lab Chip*, 2017, **17**, 460-473. 666 59. J. Dinic, Y. Zhang, L. N. Jimenez and V. Sharma, ACS Macro Lett, 2015, 4, 804-808. 667 M. Stelter, G. Brenn, A. L. Yarin, R. P. Singh and F. Durst, Journal of Rheology, 2002, 46, 507-60. 668 527. 669 61. T. Sakai, T. Katashima, T. Matsushita and U.-i. Chung, Polymer Journal, 2016, 48, 629-634. 670 A. Bublikova, F. Schütte and S. G. Mayr, Materials Advances, 2024, 5, 4807-4817. 62. 671 63. B. Yang, B. Brazile, N. J. Jan, Y. Hua, J. Wei and I. A. Sigal, J Biomed Opt, 2018, 23, 1-10. 672 D. T. Grubb and G. Ji, International Journal of Biological Macromolecules, 1999, 24, 203-210. 64. 673 65. P. Cairns, M. J. Miles, V. J. Morris and G. J. Brownsey, Carbohydrate Research, 1987, 160, 674 411-423.

Soft Matter Accepted Manuscript

- M. A. Saadiah, D. Zhang, Y. Nagao, S. K. Muzakir and A. S. Samsudin, Journal of Non-View Article Online View Article Online Vi 675 66. 676 *Crystalline Solids*, 2019, **511**, 201-211.
- 677 K. T. Shalumon, N. S. Binulal, N. Selvamurugan, S. V. Nair, D. Menon, T. Furuike, H. Tamura 67. 678 and R. Jayakumar, Carbohydrate Polymers, 2009, 77, 863-869.

679

Wet Spinning of Sodium Carboxymethyl Cellulose - DSodium M00705K Caseinate Hydrogel Fibres: Relationship between Rheology and Spinnability

Lathika Vaniyan^a, Pallab Kumar Borah^{a,f}, Galina E. Pavlovskaya^b, Nick Terrill^c, Joshua E.S.J.
Reid^a, Michael Boehm^d, Philippe Prochasson^d, Reed A. Nicholson^d, Stefan Baier^{d,e}, Gleb E.
Yakubov*^{a,g}

7

8

9

Data Availability. The data supporting this article have been included as part of the Supplementary Information.

Soft Matter Accepted Manuscript