

View Article Online
View Journal

Journal of Materials Chemistry B

Materials for biology and medicine

Accepted Manuscript

This article can be cited before page numbers have been issued, to do this please use: A. Mansour, M. Hossam Eldin and I. El-Sherbiny, *J. Mater. Chem. B*, 2025, DOI: 10.1039/D4TB02428A.



This is an Accepted Manuscript, which has been through the Royal Society of Chemistry peer review process and has been accepted for publication.

Accepted Manuscripts are published online shortly after acceptance, before technical editing, formatting and proof reading. Using this free service, authors can make their results available to the community, in citable form, before we publish the edited article. We will replace this Accepted Manuscript with the edited and formatted Advance Article as soon as it is available.

You can find more information about Accepted Manuscripts in the <u>Information for Authors</u>.

Please note that technical editing may introduce minor changes to the text and/or graphics, which may alter content. The journal's standard <u>Terms & Conditions</u> and the <u>Ethical guidelines</u> still apply. In no event shall the Royal Society of Chemistry be held responsible for any errors or omissions in this Accepted Manuscript or any consequences arising from the use of any information it contains.



Open Access Article. Published on 17 February 2025. Downloaded on 2/21/2025 12:49:03 PM.

Metallic Nanomaterials in Parkinson's Disease: A Transformative Approach for Early Detection and Targeted Therapy

Amira Mansour[#], Mariam Hossam Eldin[#], Ibrahim M. El-Sherbiny^{*}

Nanomedicine Laboratories, Center for Materials Science, Zewail City of Science and Technology, 6th October City, 12578 Giza, Egypt

^{*}These authors contributed equally to this work

^{*}Corresponding author email: <u>ielsherbiny@zewailcity.edu.eg</u>

Abstract

Parkinson's disease (PD) is a progressive neurodegeneration where there is substantial dopaminergic neurons loss in the substantia nigra, causing both motor and non-motor symptoms that significantly impact the quality of life. The prevalence of PD is expected to increase with the aging population as it affects millions globally. Current detection techniques, including clinical assays and neuroimaging, lack the sensitivity and specificity to sense PD at its earliest stages. Despite extensive research, there is no cure for PD, and the available treatments primarily focus on symptomatic relief rather than halting disease progression. Conventional treatments, such as levodopa and dopamine agonists provide limited and often temporary relief, while long-term use is associated with significant side effects and diminished efficacy. Nanotechnology, particularly the use of metallic-based nanomaterials (MNMs), offers a promising approach to overcome these limitations. MNMs, due to their unique physicochemical properties and can be engineered to target specific cellular and molecular mechanisms involved in PD. These MNMs can improve drug delivery, enhance imaging and biosensing techniques, and provide neuroprotective effects. For example, gold and silver nanoparticles have shown potential in crossing the blood-brain barrier, providing real-time imaging for early diagnosis and delivering therapeutic agents directly to the affected neurons. This review aims to reveal the current advancements in the use of MNMs for the detection and treatment of PD. It will provide a comprehensive overview of the limitations of conventional detection techniques and therapies, followed by a detailed discussion on how nanotechnology can address these challenges. The review will also highlight recent pre-clinical research and examine the potential toxicity of MNMs. By emphasizing the potential of MNMs, this review article aims to underscore the transformative impact of nanotechnology in revolutionizing the detection and treatment of PD.

Open Access Article. Published on 17 February 2025. Downloaded on 2/21/2025 12:49:03 PM

1. Introduction

Parkinson's disease (PD) is a public debilitating progressive neurodegenerative disease affecting the elderly. It is predominant in the second place after Alzheimer disease. PD prevalence increased in the last 40 years compared to the previous period reaching about 9% among the elderly population over 60 years with higher incidence in males. This gender-related occurrence is thought to be due to the doubtful neuroprotection of estrogen. ¹, ² According to the WHO, it is expected that the PD affected population will rise up to about 2 million at 2030. ³, ⁴ Although the etiology of PD is precocious in terms of clarity, PD is attributed to either non-genetic factors as dietary imbalance and pesticides or genetic mutations related to α-syn and dopamine which is more prevalent in patients less than 40 years. ³, ⁵ To achieve definitive diagnosis and efficient therapy, the pathophysiology of PD should be elucidated. PD etiology is not fully understood. Hitherto, deciphering research is recruited to provide coherent and substantial mechanistic explanation.

The dopaminergic neurons of the substantia nigra (SN) provides the dopamine for the striatum. In PD, the substantia nigra pars compacta (SNpc) neurons, that are responsible for the motor functions, are disrupted, and privation of the extracellular dopamine in the synapse culminates the striatal malfunction and induces motor disability. ⁶, ⁷, ⁸, ⁹ Other coherently pertinent factors that abrogate the PD ailment are the altered mitochondrial functions, triggered cerebral inflammatory response, and exacerbated ROS generation and activation of microglia. ¹⁰, ¹¹, ¹²

Remarkably, the aggregation of the α -synuclein (α -syn) and formation of Lewy bodies within the dopaminergic neurons of the substantia nigra is one of the main hallmarks. α -syn protein is formed from 140 amino acids divided into 3-regions; N-terminal, amyloid-binding central domain and the C-terminal. The first region is responsible for the protein's aggregation where the phosphorylation of serine-129 within the last region abrogates this event. ¹³, ^{14,15} The aggregation of the α -syn contributes in the dissipated dopaminergic neurons within the SNpc leading to dopamine deficiency. ¹⁶, ¹⁷

The PD development and progression is mostly explained by the Braak hypothesis that divides the disease into six stages where the symptoms starts with olfactory disorders and ends by the cognitive impairment. ¹⁸, ¹⁹ PD symptoms can be classified as motor and non-motor symptoms as

well (**Fig. 1**). Neuromotor disability is mainly observed as tremors at rest and bradykinesia. The latter class is heterogenous and includes cognitive disorders in addition to fast movement of the eye and some non-PD specific symptoms. All these symptoms culminate the declined life quality and can occasionally embars the patients. ²⁰, ²¹, ²², ²³ The focus of this review will be the inciting development of metallic nanomaterials (MNMs)-based detection techniques, therapeutics and theranostics. Furthermore, toxicity, biocompatibility, challenges, and future outlook would be delineated.

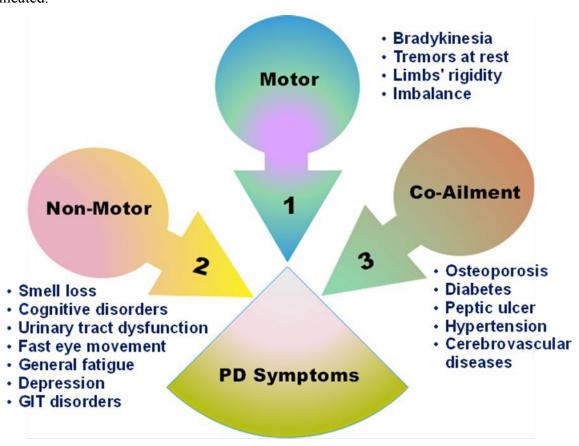


Fig. 1. A summary of the PD symptoms ²⁰, ²⁴, ²⁵

2. Detection of PD

PD clinical diagnosis remains challenging and misleading in most cases due to the overlap of its symptoms with other neurodegenerative disorders. Intriguingly, accumulation of misfolded α -syn protein clumps and development of lewy bodies in various brain parts is evident to be the primary pathogenic hallmark of PD. These α -syn aggregates, as well as DA, levodopa (LD), and miRNAs, form essential biomarkers for the early identification and control of this disease. ^{26–29} Current

Open Access Article. Published on 17 February 2025. Downloaded on 2/21/2025 12:49:03 PM

diagnostic platforms involve enzyme linked immunosorbent assay (ELISA), fluorescent and electrochemical immunoassays, high-performance liquid chromatography (HPLC), real-time polymerase chain reaction (rt-PCR) and neurobiosensors. ^{26,30,31} ELISA is the most commonly used method for the detection of markers like PS65-Ub indicating any mitochondrial dysfunction and α -syn especially in CSF because its levels are higher than in biofluids. ³² Other methods, like electrochemiluminescence and immunomagnetic reduction, are becoming more common and undergo more developments to detect extremely low levels of biomarkers. ³³These conventional techniques are used to selectively detect and quantify the levels of DA and α -syn in body fluids like blood and urine ³⁴. However, they suffer from several pitfalls; they can be complex, expensive, time-consuming, insufficient accuracy and false results, and require result interpretation. 34,35

Molecular imaging techniques are also implemented in PD diagnosis; they include magnetic resonance imaging (MRI), single-photon emission computed tomography (SPECT), positron emission tomography (PET), transcranial ultrasound imaging and their advancements. They provide quantitative information about structural changes in the brain according to their modalities and computer algorithms, and utilize tracers and contrast agents to bind to specific targets and markers. ^{36–38} Nevertheless, they are still expensive and labor-intensive techniques and this limits their accessibility for PD patients.

2.1. Artificial intelligence (AI)-based PD detection

With the outgrowing need for development and advancements of PD differential diagnosis, scientists have implemented and integrated artificial intelligence (AI) and machine learning (ML) in this process. They have employed data analytics to accurately diagnose PD in the early stages and it is showing promising results. AI holds a great potential for the automation of diagnosis and its accessibility in remote areas with high demands ^{39,40}. They mainly depend on using different models and algorithms to process inputs according to well-established libraries. These inputs can be medical history, genetic information, symptoms or PET images that need expertise to be analyzed. Such inputs are provided to AI systems possibly through wearable devices (WDs) so that they can detect patterns and anomalies indicating PD incidences with remarkable precision. Even breath patterns, as shown in Fig.2, were proved to have a link with PD, where a study was conducted on nocturnal breath signals to predict the disease. 41 The developed model achieved specificity and sensitivity of 82.83% and 86.23%, respectively using wireless signals. While ML uses algorithms to enable computers to process data without being programmed, deep learning (DL) exploits more sophisticated multi-layered neural networks for more complex data analysis and accurate predictions. ³⁹ This allowed expansion of sample size, reaching remote places, early decision making, and optimizing more algorithms to outperform human expertise. Moreover, WDs emerged as a promising tool that allows the non-invasive detection and continuous monitoring of PD. They have advanced sensors that capture motion patterns and physiological signals real-time and collect data to analyze motor functions indicative of PD patient. The design of these devices was integrated with AI to aid in data interpretation and highlight any deviation from normal motor functions.⁴² AI also facilitates the continuous improvement of WDs and adaptation through learning from any new inputs for further enhancement of the accuracy. 43 Besides, different types of sensors were exploited in WDs such as inertial, acoustic, optical, electrical, and force sensors. Once the patient uses the device, it begins retrieving and collecting specific data to be processed and analyzed using different AI and DL or ML models and algorithms. 44 However, this technology suffers from several drawbacks; ML models implemented in the studies lack full description of findings or techniques. Moreover, number and kind of subjects and their medical history are not accurately reported and assessed; while, DL models are limited due to difficulty in handling of their datasets. 45,46

Open Access Article. Published on 17 February 2025. Downloaded on 2/21/2025 12:49:03 PM

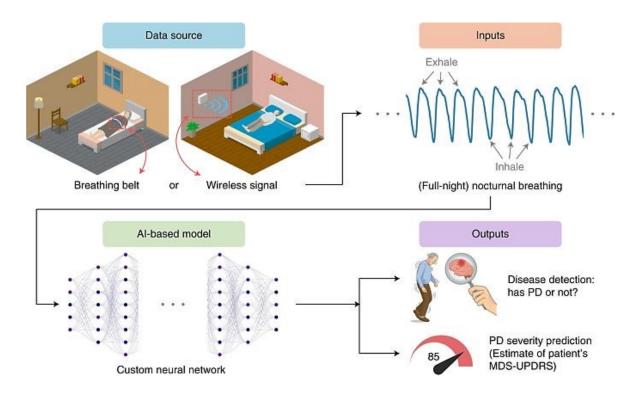


Fig. 2. A schematic illustration showing the process of AI-based diagnosis of PD. Breathing patterns is collected through signals or wearable device and processed using AI models. Based on the analysis, PD is predicted if present and its severity is determined according to Movement Disorder Society Unified Parkinson's Disease Rating Scale (MDS-UPDRS) questionnaire. Reproduced from reference ⁴¹.

2.2. MNMs-based biosensors for the detection of PD

Nanotechnology holds great potential to overcome the aforementioned challenges and confer revolutionary advances in developing diagnostic assays to enable the early detection of this incurable neurological disorder. Even though the mechanism of nanomaterials in biosensing remains unclear, they are currently experimented with offering high sensitivity, specificity and low limit of detection (LOD) ⁴⁷. Metallic nanoparticles (MNPs), like gold and silver nanoparticles, exhibit unique tunable electronic and optical properties enabling them to be used for imaging of PD. They can interact with incident light and form characteristic absorption signals that shift depending on their sizes and shapes. ⁴⁸ Moreover, they can be functionalized with specific chemical moieties to be able to cross the blood brain barrier (BBB) and act as contrast agents or bind to any desired biomarker and quantify it with high sensitivity ^{49–51}. Different metals can be

used to develop diagnostic assays for the detection of PD depending on their properties and the type of sensors. 52,53 For instance, a study was conducted to develop an immunosensor based on electrochemical impedance spectroscopy. They used an electrode system modified with Palladium nanoparticles (PdNPs) to enhance the signals and the conductivity. This PdNPs-plated electrode was functionalized with α -syn antibodies to quantify the levels of α -syn and epinephrine in clinical serum samples. It showed linear response in phosphate buffer and serum samples with LOD of 0.13 µg/ml and 1.3 µg/ml, respectively. ⁵⁴ Based on that, research has recently focused mainly on exploiting MNMs to enhance the sensitivity and performance of different types of biosensors. Biosensors can be categorized into immunosensors, DNA- and enzyme-based biosensors, and piezoelectric and thermal biosensors. 35,55 This review will focus mainly on different MNMs like zinc, platinum, gold, silver, cerium, and iron in the fabrication of biosensors for the detection and imaging of PD.

2.2.1. Gold nanoparticles (AuNPs)-based biosensors for the detection of PD

The nanoparticles of the noble metal gold (AuNPs) are extensively used for the detection of PD owing to the ease of fabrication, their inert nature, large surface area, and high affinity to biomolecules. ⁵⁶ In addition, they are biocompatible and good conductors of electricity, and they have unique optical properties manifested in their surface plasmon resonance (SPR). SPR is related to the oscillation of conducting electrons resulting in a distinguished absorption profile that depends on the shape and size of AuNPs. On account of this phenomenon, they are considered as signal transducers as shown in Fig. 3A.^{57,58} Moreover, they have been used in different structures to construct simple and sensitive biosensing platforms to enable the rapid and early detection of biomarker of PD as shown in Fig 3B. For instance, biotinylated antibody-conjugated AuNPs combined with graphene were used to design an electrochemical immunosensor to linearly detect the concentration of α -syn antigen in plasma. This nanocomposite enabled immobilization of the antibody, enhanced the conductivity, sensitivity, and specificity of the immunosensor, and produced higher current peak of 5.5 µA. Moreover, its electrochemical performance was investigated using square wave voltammetry (SWV) and showed linear detection of the antigen from 4 – 128 ng/ml with LOD of 4 ng/ml. However, the immunosensor stability and reproducibility were still under investigation before it can be used as a point-of-care device. ³¹

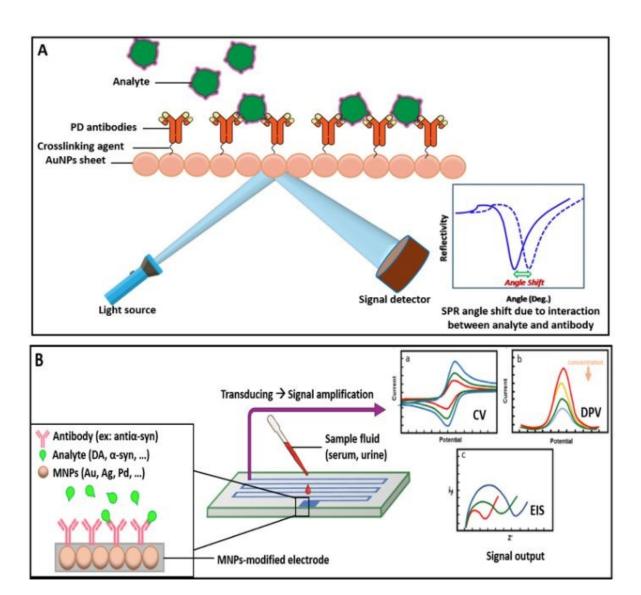


Fig. 3. (**A**) Schematic illustration of SPR phenomenon of AuNPs and its implementation in a diagnostic platform, where the interaction between the antibody and the analyte (target) causes SPR signal to shift indicating the presence of the target. (**B**) Schematic illustration of an electrochemical biosensor exploited in PD diagnosis. The antibodies are linked to an electrode, and the presence of the analyte alters the electrical signal output whether in the form of (**a**) cyclic voltammetry (CV) signal, (**b**) differential pulse voltammetry (DPV) signal, and (**c**) electrochemical impedance spectroscopy (EIS) signal.

Another research work was performed using activated charcoal modified with AuNPs to detect the levels of LD in synthetic serum, urine, and river water. This nanocomposite was deposited on a

glassy carbon to fabricate an electrode that measures the electrochemical response of LD using square-wave adsorptive anodic stripping voltammetry. Various factors were also studied according to Doehlert experimental matrix to assess their impact on the electrode performance. The results showed linear detection of LD and LOD of 50 nmol/L - 10 μ mol/L and 8.2 nmol/L, respectively.³⁴

Several studies were also conducted using the same integrated nanocomposite with different carbon allotropes to construct an electrochemical immunosensor. The reason is that it proved to have high surface area and conductivity, unique electronic properties, and biocompatibility. For example, recent work was done using multi-walled carbon nanotubes with AuNPs-doped modified with indium tin oxide electrode. This electrode was developed to detect levels of DJ-1 protein as a biomarker of PD in cerebrospinal fluids and saliva. Analysis of the biosensor efficiency was conducted using electrochemical impedance spectroscopy (EIS), single frequency impedance, and cyclic voltammetry (CV) and it showed detection range and LOD of 4.7 – 4700 fg/ml and 0.5 fg/ml, respectively. ^{59,60}

Carbon nanotubes were also used in another work as single-walled (SWCNTs) to fabricate interdigitated electrode functionalized chemically with anti- α -syn-conjugated Au nanourchin.⁵⁶ This electrode was designed to detect the levels of α -syn and the results were confirmed using ELISA technique. Due to integrating Au nanomaterial to the electrode, higher sensitivity and selectivity were achieved because it can detect low currents. Therefore, LOD was enhanced from 1 pM using the bare SWCNTs electrode to be 1 fM after applying conjugated gold nanourchin.⁵⁶

A different study was conducted using labelled gold nanobipyramids (GNBPs) to construct labon-a-chip diagnostic system based on surface-enhanced Raman scattering (SERS). This nanostructure is anisotropic which offered sharp tips and edges and consequently, enriched the plasmonic hot points. In addition, it was functionalized with hairpin DNA and Raman reporter moieties which caused GNBPs to aggregate if the analyte was present. This platform enabled the detection of altered expression of miR-221 and miR-214 as biomarkers of PD through amplified SERS signal due to aggregation. ⁶¹

SERS technique was also implemented in another study conducted on AuNPs. However, this study aimed at imaging DA in retinal tissues and live cells. AuNPs were labelled with modified thiol molecules due to their high affinity to Au surface where DA-positive samples result in the aggregation of functionalized AuNPs. The positive response was detected by the Raman scattering signals due to formed plasmonic hot spots, and it is speculated that this technique can be further applied in the detection of DA in live cells in PD patients.⁶² Furthermore, the biodistribution of administered gold nanoclusters (AuNCls) was studied as a function of the route of administration in mice as animal models. Intravenous, intraperitoneal, intranasal, intragastric routes were assessed, and it was found that AuNCs were mostly located in the brain in the case of intraperitoneal administration. This implies that they had the ability to cross the BBB, therefore, can be further investigated to use such nanoparticles in the imaging of PD upon irradiation along with treatment.⁶³ Besides, Adam et el. focused on developing AuNPs-based interdigitated tetraelectrode to detect α-syn in the fibril formation process. This electrochemical biosensor was investigated using cyclic and differential pulse voltammeter, and the results showed linear range of 1 aM – 1 pM and LOD of 100 aM. ⁶⁴ All these biosensing models will enable the early detection and monitoring of PD once they are well-developed and pass the stages of the clinical trials.

2.2.2. Silver nanoparticles (AgNPs)-based biosensors for the detection of PD

Silver nanoparticles (AgNPs) were investigated in literature for the detection of PD by biosensing of different PD biomarkers.⁶⁵ Like Au, Ag has unique SPR properties, good chemical stability and biocompatibility, and enables Raman spectroscopy. For example, Ma et al. exploited silver nanoclusters (AgNCls) functionalized with 5-mercapto-2-nitrobenzoic acid (MNBA) to fabricate photoelectrochemical biosensor to detect the levels of DA and glutathione (GSH), simultaneously in PD-bearing animal models with different stages. The cell structure was designed using Ag electrodes as multi-channels for the detection of multiple analytes through changes of photocurrent. AgNCls were further integrated with hybrid nanomaterials like graphene oxide (GO), carbon dots (CDs), and AgNPs to enhance the photoelectric activity, sensing performance, and the selectivity. The results showed that the biosensor hybrid with AgNPs and GO achieved LOD and linear range of DA of 53 nM and 0.16 – 6 μM, respectively. Whereas, the biosensor hybrid with CDs reported LOD and linear range of GSH of 34.3 nM and 0.1 – 1 μM, respectively. Another work employed AgNPs to fabricate label-free SERS platform as a sensitive and robust

diagnostic assay to detect 5-S-Cysteinyl-DA which is an important metabolite in PD. The results showed that this technique allowed the detection of the metabolite in synthetic cerebrospinal fluid with low concentrations of about 10 nM while 100 nM in simulated urine. ⁶⁶

The same technique was implemented in another work to detect the levels of DA in-situ using AgNPs where the label-free SERS measurements were conducted with laser of wavelength 488 nm. The results displayed reproducibility of SERS spectrum when using citrate-reduced AgNPs upon binding to DA through surface adsorption. ⁶⁷ Besides, Rouhani et al. worked on developing a biosensing electrode made of AgNPs with GO as a nanocomposite to detect LD in serum and urine samples. The performance of the modified electrode was measured through CV, and it showed enhanced oxidation-reduction peaks. Different parameters, such as concentration of GO, pH, and accumulation time were optimized, and the results showed that this accurate assay achieved LOD of 0.76 nM and linear range of 0.003 – 10 μM of LD. ⁶⁸

2.2.3. Iron oxides nanoparticles (IONs)-based biosensors for the detection of PD

Avowedly, iron oxides nanoparticles (IONs) are extensively used in the bioimaging and biosensing of neurodegenerative diseases due to their inherent biocompatibility, biodegradability, and small tunable sizes. ⁶⁹ They also exhibit unique SPR properties and high refractive index, therefore, they can cross BBB and serve as excellent MRI contrast agents in clinical bioimaging. Moreover, they can be functionalized with specific chemical moieties for enhancing the active targeting and bypassing BBB. Superparamagnetic IONs (SPIONs) align and become magnetic in the presence of external magnetic field, and this property can be employed for sustained drug release, control the targeting and avoid the clearance. ⁵⁷ Furthermore, SPIONs have also been used to tag, track, and monitor stem cells and their spatial distribution via MRI. 70 An et al. managed to synthesize ultra-small IONs functionalized with polyacrylic acid (PAA) and polyethylene glycol (PEG) to prevent protein corona formation and reduce the immunogenic response *in-vivo*. These particles acted as effective MRI agents with relaxation rate of 84.65/s.mM and they enabled labeling and tracing of primary human adipose-derived stem cells for up to 3 weeks. 71 This can be further implemented as a diagnostic technique to trace potentially therapeutic stem cells in PD patients. Another study employed PEGylated SPIONs conjugated to W20 antibody as MRI probe to detect amyloid oligomers in PD transgenic mice. The results proved the BBB penetration of these small SPIONs and showed high magnetic resonance relaxation and significant contrast in T2 weighted image. Therefore, this system was able to detect amyloids at the early stages because of its high sensitivity and biocompatibility. 72 Additionally, magnetic IONs were exploited in the biosensing of different biomarkers of PD due to the ease of their functionalization and their high sensitivity. For instance, Yang et al. developed a sensitive immunosensor depending on immunomagnetic reduction (IMR) of magnetic IONs labelled with anti-α-syn. The IMR signals in plasma samples were detected with magnetosusceptometer and the results showed that they achieved LOD and range of α -syn concentration of 0.3 fg/ml and 0.1 – 100 pg/ml, respectively. ^{73,74} Another work was done using anti-α-syn-functionalized IONs hybrid with GO to construct Mg-based micromotors for the detection of α -syn in whole blood samples. This sensitive electrochemical biosensor enabled the effective capturing of the biomarker in 100 s and its concentration was inversely proportional to the signal amplification capability of the developed biosensor. ⁷⁵ Besides, Zhang et al. investigated IONs-based interdigitated electrode for the identification of α -syn through using both aptamer and antibody to sandwich the analyte. They enhanced the performance of this biosensor by modifying it with AuNPs where the current changed as a response to binding to the target in a linear manner. The results recorded LOD of 10 aM and a linear range of $10-10^7$ aM with $R^2 = 0.9729$, thus, enabled the sensitive and accurate detection of α -syn (Fig. 4). ⁷⁶ Table 1 summarizes the main studies of MNMs-based biosensors of PD.

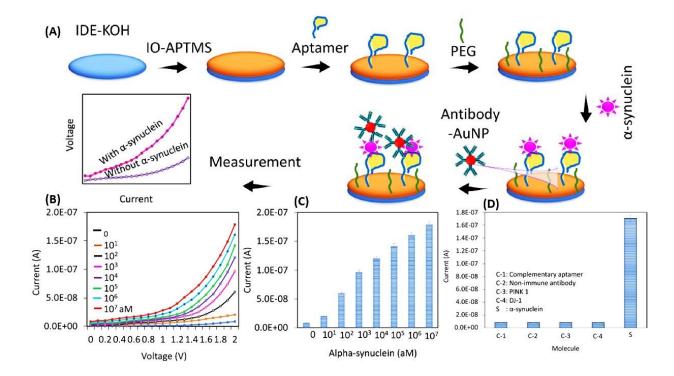


Fig. 4. (**A**) Scheme of the biosensor building. Iron oxide (IO)-modified aptamer and polyethylene glycol (PEG) were immobilized on the surface of interdigitated di-electrode (IDE) where the α-syn is sandwiched by the anti-α-syn aptamer and antibody-AuNPs, (**B**) and (**C**) quantitative detection of α-syn several concentrations and (**D**) selective detection of α-syn in the presence of other analytes. Reproduced from reference 76

Table 1. Summary of the main studies reported the development of MNMs-based PD biosensors.

2.43.45 omgencia 2.43.46 omgencia	5 of 54			Journal of Materials	Chemistry B			
Article. Published on 1 / February 2025. Downloaded on 2/21/2025 12:49:45 Ph This article is licensed under a Creative Commons Attribution-NonCompercia of Physics and Physics of Physics of Physics of Physics of Physics article is licensed under a Creative Commons Attribution-NonCompens	Table 1	1. Summary of the	main studies repor	rted the development of MN	NMs-based PD biosensors.			
ownlo	Target	Targeting Ligand	Sample Type	Detection Method	Nanomaterial	Linear Range	LOD	Ref.
2025. Do ative Co	α-syn	rabbit Ab138501 mAb	Serum	Label-free SPR	Iron oxide (Fe ₃ O ₄) NPs	0.01 – 100 pg/ml	5.6 fg/ml	30
ruary í	α-syn	Specific antibodies	Synthetic	Electrochemical impedance		1.5 – 15 μg/ml	0.13 μg/ml	54
ı 17 Feb ed unde:	Epinephrine		cerebrospinal fluid	spectroscopy	Palladium (Pd) NPs	0.75 – 100 μmol/L	0.051 μmol/L	_ 54
shed or s licens	DA		Live human nerve cells	Enzyme-less electrochemical biosensor (CV)	NiAl layered double hydroxides nanosheets integrated with GO	$0.1 - 97 \mu M$	2 nM	77
e. Publi ırticle is		antiα-syn	Diluted human sera	Label-free electrochemical immunoassay	Cysteamine-functionalized fluorine-doped tin oxide NPs	10 – 1000 ng/ml	1.13 ng/ml	78
s Article This a		antiα-syn	Human plasma	Electrochemical immunoassay	Gold Nanostars-decorated zinc oxide (ZnO) nanowires	0.5 – 10 pg/ml	0.08 pg/ml	79
Open Access /	α-syn	Monoclonal antibody	Plasma and serum	Immunomagnetic reduction assay	Magnetic Fe ₃ O ₄ NPs	ND	In plasma: $3.60 \pm 2.53 \text{ pg/ml}$ In serum: $0.03 \pm 0.04 \text{ pg/ml}$	80
(3)		Thiolated aptamer	Diluted serum	Electrochemiluminescence	AuNPs@Metal organic	2.43 fM – 0.486 pM	0.42 fM	- 81
		Carboxylated aptamer	Diffuted Setulii	(ECL)	frameworks (MOFs) composite	1.39 fM – 0.243 pM	0.38 fM	

These MNPs-based electroanalytical techniques have advantages over conventional ones as they require small volume of sample and exhibit high sensitivity and specificity. They also allow multiplexing where one electrochemical biosensor can be functionalized with various ligands for the detection of multiple analytes simultaneously, and this, as a return, enhances the accuracy of the result. As for the imaging techniques, contrast agents, like IONs, enabled higher resolution in MRI and easy penetrability due to their ultra-small sizes, and they can detect any minor changes in the brain structure. However, using MNPs may encounter some limitations where they can be expensive to manufacture, complex according to the design, and difficult in standardization because of the low reproducibility and high reactivity of the nanoparticles. These hurdles require more advanced research and optimization in implementing nanotechnology in diagnostic techniques to overcome such limitations and enhance the nanoparticles applicability.

Treatment of PD 3.

BBB is a semi-permeable membrane that surrounds the central nervous system including the brain. It confers selective permeability that constrains the passage of pathogens and toxins but passively transport the essential nutrients for survival and homeostasis. 82 On the other hand, this semipermeability constrains the treatment as only 1% of the therapeutic doses reaches the brain. However, increasing the doses is stumbled by the severity of the adverse effects. ¹⁶, ⁸³ PD is irremediable and the available therapy is mainly palliative for both the motor and non-motor symptoms. Therefore, the patients' plight is ameliorated through the easiness of the symptoms by increasing the intraneuronal dopamine (DA) or enhances DA receptors. ²⁰ Due to the noted DA deficiency in the PD patient's brains, the recruited efforts lead to the development of DA precursor; levodopa (L-3,4-dihydroxyphenylalanine, known as L-dopa). L-dopa is characterized by its ability to penetrate the BBB and converted into DA when absorbed by the nerve cells. Hitherto, the DA precursor; L-dopa, is the indispensable choice for the treatment and it exerts its effect through replenishing the deficiency. L-dopa can be undesirably converted to DA in the peripheral nerves, therefore, carbidopa (peripheral dopa-carboxylase inhibitor) is co-administered with L-dopa.⁸⁴ However, DA does not exert efficient remedial effect on the non-motor symptoms as hallucination and can culminate dopaminergic "type A" adverse effects. 85 Also, there are some drugs classes that were proven to have anti-PD effect as monoamine oxidase-B (MAO-B) inhibitors and catechol-O-methyltransferase (COMT) inhibitors. However, they may also induce the

Open Access Article. Published on 17 February 2025. Downloaded on 2/21/2025 12:49:03 PM.

dopaminergic "type A" adverse effects. ⁸⁵ Different treatment mechanisms are illustrated in **Fig. 5.**

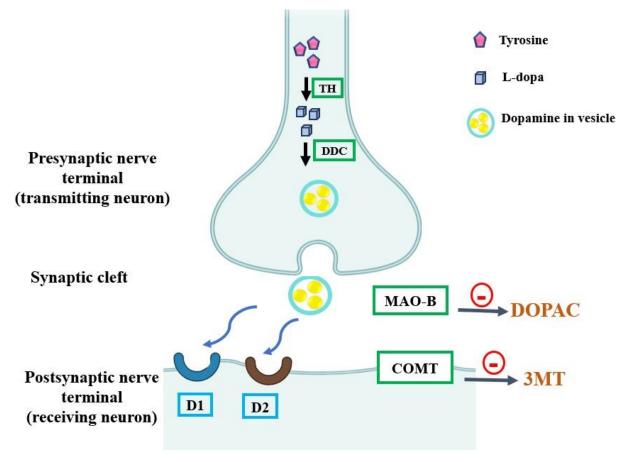


Fig. 5. Classes of anti-PD drugs affecting the DA (dopamine) synapse. In the presynaptic nerve terminal, tyrosine is converted to L-dopa by tyrosine hydroxylase (TH) and then to DA by L-dopadecarboxylase (DDC). Monoamine oxidase-B (MAO-B) inhibitors prevents DA break down and catechol-O-methyltransferase (COMT) inhibitors prevent L-dopa peripheral degradation. D1 and D2 are DA agonists as the non-ergot agonists. DOPAC, dihydroxyphenylacetic acid; 3MT, 3 methoxy-tyramine. Recreated from reference ⁸⁶.

The resistance-dependent debilitating ability of the neurons to maintain the DA in addition to the limited plasma half-life of L-Dopa aggravates the PD motor stability. For instance, the DA level fluctuates causing on/off cycles of akinesia and dyskinesia. So, non-ergot DA agonists as ropinirole and pramipexole are currently used as long-acting treatments available for PD. ⁸⁶, ²¹ In order to sequester the limitations of PD, the application of nanotechnology and development of

nano-based therapeutics emerged as efficient and probable definitive treatment. Nanomaterials can penetrate the BBB, interact with the cells, encapsulate and deliver drugs to obtain desirable drug release rate while dissipating off-targeting of the drugs. ⁸⁷,⁸⁸ Intriguingly, many studies have shown the ability of the MNMs to overcome this permeability obstacle and effortlessly penetrate the BBB ⁸⁹. Consequently, the MNMs effect is potentiated to have a crucial role in the developing of central nervous system (CNS) novel treatments (**Fig. 6**). Other classes of nanostructures are described in literature elsewhere as liposomes ⁹⁰, nanoemulsions ⁹¹ and polymeric nanoparticles.

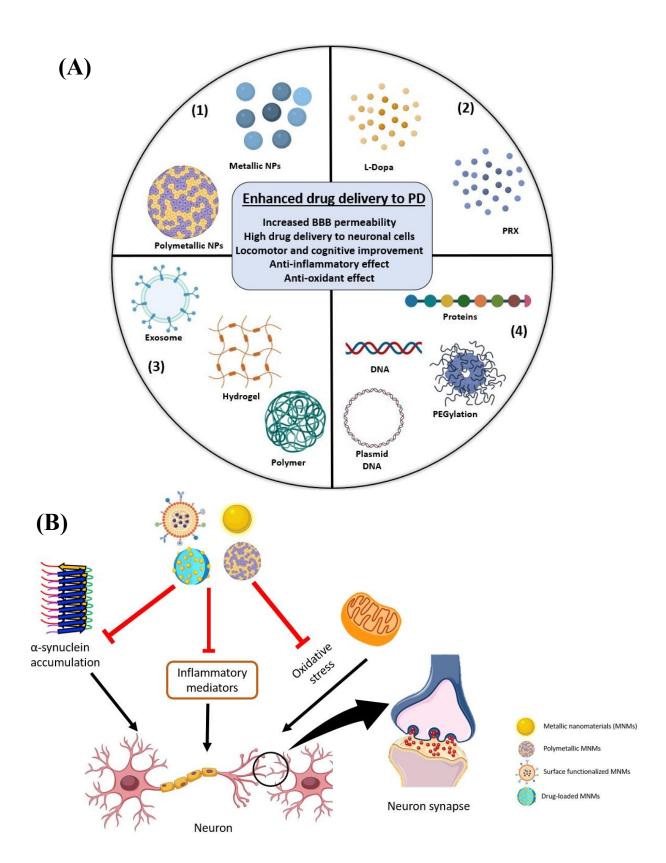


Fig. 6. (A) Schematic representation of several protocols for the implementation of MNMs to develop PD-based drug delivery platform. MNMs can be prepared from a single metal or multiple

metals (1), loaded with drugs (2), impeded into nanoformulations (3) and different classes of surface-attached targeting ligands (4), (B) Scheme of how the MNMs exert their anti-PD effect through decreasing α -syn level and blocking inflammation and oxidative stress to normalize the neurons' function.

3.1. MNMs-based treatment of PD

3.1.1. Gold nanoparticles (AuNPs)-based systems for the treatment of PD

Gold-based nanostructures have been extensively exploited in biomedical applications due to their biocompatibility, inertness and the presence of versatile well-established preparation approaches. Several factors govern the use of AuNPs such as the size, shape, charge, incorporation within nanoformulations and the use of specific surface-modified ligands, DNA or proteins. Concerning the CNS specifically, AuNPs intrigued specific interest due to their capability to cross the BBB and their antioxidant and anti-inflammatory effect without significant toxicity. 89 To consolidate the importance of size, Gao and coworkers reported the inversely proportion relation between the size of the AuNPs and the intensity of Aβ fibrillation. For instance, relatively bigger AuNPs (36 and 18 nm) enhanced the fibrillation while small AuNCs (6 nm) constrained this process. Intriguingly, even smaller AuNCs (1.9 nm) showed utter fibrillation. 93 Ultrasmall nanoclusters (AuNC, <3 nm) were used to develop remedial nanoformulations for PD ⁹⁴, ⁹⁵, ⁹⁶ due to their remarkable penetration of the BBB. 97 For example, Ma et al, formulated AuNC coated with peptides that are either positive or negative. Both types bind to 20S proteosome, Nonetheless, the AuNC(-) preferentially conferred cellular neuroprotective effect simultaneously with decreasing of α-syn. 95 This study highlighted the impact of different surface charges on PD manifestations and the shape also represented a controversial aspect in terms of its effect. Wang et al, compared Au nanospheres (AuNSs) and Au nanocubes (AuNCs) of the same size (20 nm) and found that AuNSs induce more fibrillation of amyloid- β peptide (A β (1-40)) more than {100}-faceted AuNCs owing to the greater chemical activity of the surface atoms. 98 Additionally, the stabilizers of the AuNPs were found to have a potentiation effect. Citrate-stabilized AuNPs were reported to decrease the membrane potential by binding to glia neurotransmitters receptors and thus, neutralize the astrocytes activation. 99 Their release causes diverse neurological disorders as PD through the activation of astrocytes by surface-attached receptors. 100

Open Access Article. Published on 17 February 2025. Downloaded on 2/21/2025 12:49:03 PM

Naturally derived AuNPs were found to be effective treatment for PD ¹⁰¹, ¹⁰² and can ameliorate the induced neurotoxicity through exerting protective effect from destructive oxidation, inflammation and cell death. 103 For instance, AuNPs from Hibiscus sobdariffa and from have been reported to have remedial effect on PD by abolishing the aggregation of α -lactalbumin. ¹⁰¹ AuNPs (30-50 nm) derived from *Cinnamomum verum* neutralized the oxidative activity and neuromotor dysfunction in the MPTP-induced PD model in addition to mitigated tumor necrosis factor-α (TNF-α), Interleukin-1β (IL-β) and Interleukin-6 (IL-6) levels and normalized TLR/NF-κB signal. 104 Hu, et al., adopted gene drug delivery for the treatment of PD using a nanocomposite based on chitosan and AuNPs loaded with plasmid DNA and nerve growth factor (CTS@GNPpDNA-NGF) (Fig. 7). The NGF facilitated the cellular endocytosis into the PC12 cells *in-vitro* model. Also, the MPP⁺-based apoptosis was inhibited by the nanocomposite as proven by confocal microscope due to the suppressed overexpression of α -syn. These results resembled those obtained from the western blot analysis that proved that CTS@GNP-pDNA-NGF deteriorated the expression level of α -syn. Additionally, in the PD *in-vivo* model induced in C57bl/6 mice by MPTP, CTS@GNP-pDNA-NGF nanocomposite improved the body weight, healed the substantianigra density, crossed the BBB and cleared from the body through the spleen after exerting its therapeutic effect. 105

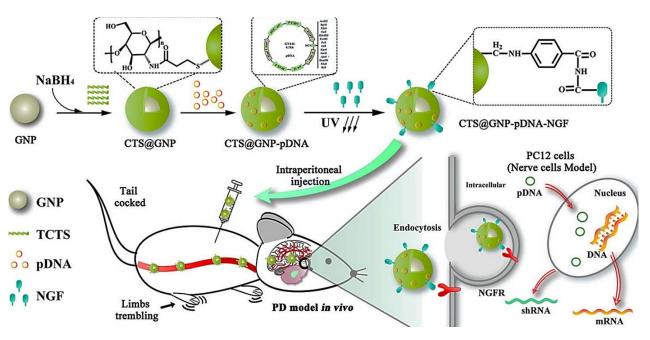


Fig. 7. Schematic illustration for the preparation of the nanocomposite, coating with the pDNA (plasmid DNA) and nerve growth factor (NGF). The NFG-driven endocytosis into PC12 cells was

followed by inhibited enhanced cellular and dopaminergic neurons proliferation. *In-vivo* model of PD showed BBB permeability and blocking of α -syn overexpression. Adapted from reference ¹⁰⁵ with permission from Elsevier, copyright 2018.

AuNPs were exploited as neuronal drug delivery cargo for the most efficient therapies of PD. Nanoflowers of multibranched AuNPs were prepared and acted as transposing carrier of L-DOPA to penetrate the BBB. ¹⁰⁶ A novel AuNPs-based platform for the cerebral drug delivery of L-DOPA and DA was developed. The AuNPs surface was functionalized by three different amantadine derivatives due to their ability to bypass the BBB and their biocompatibility. 87, 107, 108 The surfacefunctionalized AuNPs were compared to polyethylene glycol-coated AuNPs (PEG-AuNPs). L-DOPA or DA were attracted to the AuNPs by several potential surface interactions. The developed systems were investigated in the presence of bovine serum albumin (BSA) as it is the most abundant protein in the body that may form a protein corona hindering the drug's release. Peptidoglycan monomer-AuNPs (PGM-AuNPs) was proved to be the most successful drug delivery tool as it had the highest drug cargo especially from the DA. Additionally, the impact of the BSA on pharmacokinetics and pharmacodynamics was almost diminished. 87 However, neither *in-vitro* assessments nor *in-vivo* studies were performed to further reveal the anti-PD activity and efficiency. AuNPs were reported to be incorporated with other therapeutic agents to exploit the beneficial characters of the AuNPs. For example, it acted as the conductive component in selfhealing hydrogel that exerted anti-inflammatory effect. 109 Another hydrogel was also prepared using AuNPs conductive properties to design an injectable implant in the brain for PD treatment. ¹¹⁰ Also, AuNPs acted as a drug carrier for through electrostatic interactions, ⁸⁷ NIR-responsive, ¹¹¹ gene carrier and ¹¹² allowing CT imaging. ¹¹³ Diverse applications of AuNPs for the treatment of PD are listed in **Table 2**.

This article is licensed under a Creative Commons Attribution-NonCommercial 3.0 Unported Licence

Open Access Article. Published on 17 February 2025. Downloaded on 2/21/2025 12:49:03 PM

Table 2. AuNPs-based PD drug delivery systems, performed biological assessment, outcome and the mechanism of action

of 54			Journal of Materials Chemis	try B				
Table 2. AuNPs-based PD drug delivery systems, performed biological assessment, outcome and the mechanism of action								
Nanoformulation	Size (nm)	Zeta potential (mV)	Biological assessment	Efficacy outcome	Mechanism of action	Re		
Table 2. AuNPs-ba Nanoformulation Electroconductive hydrogel using dialdehyde polyurethane as nano-crosslinker, AuNPs (CDAH)	15	-26.2 ± 2.7	In-vitro cytotoxicity by neural stem cells and in-vivo by oxidopamine (6-OHDA)-induced rat model	CDAH2 hydrogel was found to be biocompatible by quantifying M2/M1 macrophages. The ratio of the CDAH2 group was double the control one. In the <i>in-vivo</i> model, there was significant functional recovery	The hydrogel neutralized the ROS production, decrease inflammation of neurons and delay dopaminergic neurons degradation in SNpc, striatum glial fibrillary acidic protein (GFAP+) astrocytes were limited	1		
AuNPs-pDNA loaded into liposomes, NGF and docosahexaenoic acid linked to the liposomes surface as targeting ligands to develop AuNPs(pDNA)-Lipo-NGF-DHA	20-40 nm of the AuNPs, increased to 200 nm	-15	Cell viability test of SH-SY5Y cells and 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP) <i>in-vivo</i> model	AuNPs-(pDNA)-Lipo-NGF-DHA significantly enhanced the cell viability relative to the control but not a significant difference compared to AuNPs(pDNA), in PD disease model, TH was increased, improved bradykinesia, improvement in the cognitive impairment and motor dysfunction	AuNPs (pDNA)-Lipo- NGF-DHA conferred protection of the dopaminergic neurons by elevating the TH and constrain the overexpression of α-syn, opposed long-term potentiation (LTP) inhibition	1		
AuNCs functionalized with dihydrolipoic acid (DHLA-AuNCs)	1.87 ±0.14	-	Cytotoxicity and microglial protein-inflammatory response on BV2 cells, cytotoxicity, neuronal differentiation and axonal growth assay on N2a cells, ex-vivo organotypic brain slice model	Inhibited proinflammation in BV2 cell, improved neurogenesis in N2a cells and in <i>ex-vivo</i> brain slice stroke model	Stimulated polarization near to the M2-like phenotype in addition to enhanced anti-oxidant effect and decreased NF-kB signal, prohibited apoptosis that led to enhanced cell viability, limited astrogliosis	ç		
Hybrid nanoassembly of AuNPs decorated with thiolated amphiphilic β-	150–300 nm, size of AuNPs was 20–	-	Estimation the rate of DA release after the additions of stimulants by CV	Release of 70% of the DA amount through half an hour as the AuNPs@SC16SH/DA	Thiolated-compounds dependent redox responsive drug delivery system for the	1		

Accepted Manuscript

Journal of Materials Chemistry B

cyclodextrin incorporated with DA; AuNPs@SC16SH/DA	30 nm			acted as stimuli responsive nanocarrier	DA, β-cyclodextrin acted as carrier for the DA by forming a complex with it, AuNPs allowed the monitoring of the DA release intracellularly followed the penetration of the BBB	
AuNPs coated with brain-targeting exosomes functionalized with rabies viral glycoprotein (RVG)	AuNPs: 48 ±1.5 and AuNPs/exosomes : 105 ±10.1	About -7	In-vitro binding efficiency using bEnd.3 mouse brain endothelial cell line, C6 rat glioma cells, or Hela cervical cancer cells, in-vitro BBB model using Astrocyte cells and bEnd.3 cells and murine model and in-vivo assay to determine the accumulation of the AuNPs/exosomes in the brain	bEnd.3 and C6 displayed the greatest binding affinity towards the AuNPs/exosomes than the Hela cell so the nanoparticles have brainspecific affinity, the AuNPs/exosome crossed the BBB with a significant increase relative to the AuNPs coated with unmodified exosomes, the <i>in-vivo</i> model demonstrated intense fluorescence from the brains treated with RVG-exosomes-coated AuNPs	Enhanced BBB permeability of the AuNPs after the coating with brain targeting RVG-protein ligated to exosomes	115
L-DOPA-functionalized multi-branched nanoflower-like gold nanoparticles (L- DOPA-AuNF)	90 nm	-34.6 ±1	In-vitro test of the L-DOPA-AuNF accumulation in the hCMEC/D3 brain endothelial cells, penetration of L-DOPA-AuNF across the brain endothelial monolayers, internalization of L-DOPA-AuNF into brain macrophages	High intracellular accumulation in the brain endothelial cells in the cytoplasm and the perinuclear space, penetration and internalization of L-DOPA-AuNF into the brain microglia without inflammation	Energy-dependent cellular internalization of L-DOPA-AuNF into brain endothelial monolayers and brain microglia due to L-DOPA functionalization and may be due to receptor-independent mechanism	116, 106

25 c	of 54			Journal of Materials Chemis	try B		
	NIR-responsive PLGA microspheres loaded with pramipexole (PRX) and hollow gold nanospheres (HGNS) (PRX/HGNS MS)	HGNS was 40 nm, PRX/HGNS MS was 24 μm	-	Cytotoxicity assessment using C2C12 myotubes, RAW macrophages, PC12 model neurons and 3T3 fibroblasts, <i>in-vivo</i> model including pharmacokinetics, pharmacodynamics and immunohistochemistry	HGNS and PRX/HGNS MS did not exert significant cytotoxicity were found biocompatible, the release rate of PRX from PRX/HGNS MS significantly increased compared to PXR from PXR MS, NIR-exposed mice treated with PRX/HGNS MS exhibited faster improvement relative to non- NIR exposed mice, significant decrease of TH in the striatum	Significant post-treatment elevation if DA, homovanilic acid (HVA) and 3,4-dihydroxyphenylacetic acid (DOPAC), protective effect of the neurons conferred by PRX/HGNS MS and NIR-treated mice	111
	Self-healing composite hydrogel of O- carboxymethyl chitosan (CMC)OTA@Au (oxidized tannic acid modified gold nanocrosslinker) (COA)	27.79 ± 2.89 nm	-45.6 ±2.7	In-vitro antioxidant test by neural stem cells (NSCs) and anti-inflammatory test by J774A.1 murine macrophages, in-vivo PD model followed by behavioral and electrophysiological assessment, immunofluorescent and immunohistochemical assays	NSCs proliferation was enhanced in the presence of COA, upregulation of GFAP, β-tubulin and MAP2, about 90% of the inflamed cells were healed after 12 h of COA treatment, inflammatory proteins expression was not expressed in COA-treated macrophages, significant increase in the counterclockwise rotation period, improved forelimbs contact, low spikes count in the projection neurons discharge behavior	The developed COA has efficient self-healing ability and injectability, can enhance the cellular proliferation and subsequent differentiation, can be developed as implant injected to the brain	110

3.1.2. Silver nanoparticles (AgNPs)-based systems for the treatment of PD

AgNPs are widely distributed to multiple organs. However, it is cleared from all the organs after 8 weeks of the initial exposure except from the testis and remarkably, the brain, as they have the ability to cross the BBB. ¹¹⁷, ¹¹⁸, ¹¹⁹ They were reported to stimulate inflammation, especially in the first 24 h of exposure. ¹¹⁷, ¹²⁰ However, based on a comparative study, several *in-vitro* assays were comparing biogenically prepared AgNPs and AuNPs by *Acacia auriculiformis* leaves. AgNPs decreased the fluorescence and absorption magnitude in the *in-vitro* assessments in addition to the enhanced anti-oxidant effect. Thus, AgNPs were speculated to have superior inhibitory effect in comparison to the Au counterparts. ¹²¹ Gonzalez-Carter et al, proved that the citrate-capped AgNPs-induced immune response enhanced the microglia ability to upregulate anti-inflammatory enzymes and reduce ROS. Following AgNPs microglial internalization, AgNPs induce the expression of cystathionine-γ-lyase (CSE) that detains the release of Ag⁺ ions. Also, Ag₂S is formed as a covering layer on the surface of the AgNPs. Consequently, ROS, nitric oxide and TNF-α production are mitigated. From all these findings, it can be concluded that AgNPs suppresses the inflammation and neurotoxicity of the microglia cells. ¹²²

The green synthesis of AgNPs for the treatment of PD ^{123–126} and exerting anti-oxidant effect have been previously reported. ^{127, 128} The plant extract of *Mucuna pruriens* was found to have reduction capability that was exploited to produce AgNPs and simultaneously, containing high L-dopa amounts. ¹²⁹ Sardjono et al, reported the preparation of AgNPs (36.5 nm) by the seeds extract and performed catalepsy assessment to investigate the efficiency of the prepared NPs in the *in-vivo* model. Gradually increasing doses starting from 5 to 25 mg/kg were tested on 3 months male mice. Groups of mice treated with 5, 10 and 15 mg/kg of AgNPs significantly improved the catalepsy symptoms relative to the control and pure extract-treated groups where the dose of 5 mg/kg-treated group was remarkably the most efficient. ¹²⁴ Therefore, in order to circumvent the potential toxicity of AgNPs, green synthesis, coating with biocompatible polymers or decorating the surface with neural-cells specific targeting ligands are all potential pathways that should be adopted. ¹³⁰

3.1.3. Iron oxide NPs (IONPs)-based systems for the treatment of PD

The application of iron oxide nanoparticles (IONPs) can be adopted through either of two approaches. The first one is through actively targeted therapy. For example, IONPs were coated

with oleic acid which acted as a cargo and then layered with short hairpin RNA (shRNA). The molecule was endocytosed through NGF-receptors. The developed nanoparticles constrained the expression of α-syn as concluded from the adopted *in-vitro* and *in-vivo* models by PC12 cells and MPTP-bearing mice, respectively. ¹³¹ Cheng et al, exploited the NIR-assisted penetration of the BBB to deliver the anti-PD drug; minocycline through loading it into Fe₃O₄ NPs. ¹³² A theoretical simulation study was performed to design PEGylated Fe₃O₄-based nanocargo as carrier for DA. PEGylated Fe₃O₄ NPs were coated with albumin to enhance the BBB penetrability and were loaded with DA thus, the developed NPs were speculated to provide sustained DA release after the neuronal uptake. 133 Iron oxide NPs coated with curcumin were developed and the results showed rescued DA and norepinephrine levels in addition to the anti-oxidant effect ¹³⁴. Ferric ions were coordinated with curcumin and poly(vinylpyrrolidone) (PVP) to develop PVP ultrasmall coordination polymer NPs (Fe-Cur NCPs, 10 nm) that was proven through the *in-vivo* model to alleviate the PD symptoms by improving the mitochondrial stress and having anti-inflammatory effect. ¹³⁵ The purpose of the study conducted by Moayeri et al., was the replacement of the missing dopaminergic neurons with stem cell therapies that are guided towards specific injury. So, external magnetic field was applied to PD *in-vivo* rat model and the therapeutic effect of superparamagnetic iron oxide nanoparticles (SPION)/poly-L-lysine hydrobromide (PLL) was found to be about 96%. Consequently, the transfection with SPIONs/PLL was found to be successful strategy for the translocation of the stem cells in the target tissue. 136

The second approach is underpinned on the application of external magnetic field. SPIONs can ameliorate the remedial effect of human mesenchymal stem cells (hMSCs). Dextran-coated IONPs. Both rotational and motor behaviors were significantly improved relative to the control. This improvement was attributed to the enhanced migration of hMSCs towards the damaged DA neurons and their subsequent differentiation to resemble the DA-neurons. To enhance the selectivity of the SPIONs towards PD, the binding affinity of streptavidin (SA) and biotin was exploited. The developed SA/PEI-SPIONs were tested using biotin-treated PC12 cell as an *in-vitro* model. The surface modified-SPIONs demonstrated greater binding the PC12 cells' membrane as observed by TEM. 138

3.1.4. Other MNMs-based systems for the treatment of PD

The experiment 6-OHDA-bearing hemiparkinson's mice showed that the release of DA from the implanted DA-loaded TiO₂ lattices improved the motor symptoms for one month and significant reduction of the symptoms was still observed after two months.¹³⁹ Interestingly, TiO₂-based implant with nanopores provided sustained release up to 30 days post-implant incorporation.¹⁴⁰ Punitha and coworkers adopted green synthesis pathway to prepare TiO₂ NPs (34 nm) and the cytotoxicity using PC12 cells was in the range of 31.2-62.5 µg/mL.¹⁴¹ Yet, dose-dependent dopaminergic neural toxicity of TiO₂ was corroborated after accumulation in the midbrain substantia nigra of Balb/c mice. ⁶

Selenium NPs (Se NPs) were proven to have neuroprotective effect so were reviewed as potential therapy for CNS diseases as PD. 142 For instance, effect of the Se NPs is attributed to the crucial role of Se to allow the normal function of several peroxidase enzymes 143 and inhibiting inflammatory mediators as TNF- α , Nf- $_k$ B and PEG2 144 to exert its anti-oxidant and anti-inflammatory effects, respectively. Derivatives of aminothiazole were synthesized and used for the development of Se NPs pursuing fortified enhancement of the neurological functions. The molecular docking studies revealed blocking of the hMAO isoforms (A and B). Simultaneously, the IC50 of the NPs was only 0.033 μ M, preceding the normal-sized counterparts' potency by about 70%. In the haloperidol-induced PD in-vivo model, the behavioral test results of the NPs group showed improvements and enhanced exploration. Therefore, the Se-based nanoformulation is elicited as effective anti-PD. 145

Gao et al, developed combined genetic and antioxidant therapy through NIR-stimulated magnesium oxide (MgO)-based nanocomposite; MgOp@PPLP and both *in-vitro* and *in-vivo* assessments results demonstrated enhanced permeability to the BBB in addition to the exerted anti-inflammatory and antioxidant effects. ¹⁴⁶

PD-lesioned cell model by 1-methyl-4-phenylpyridinium (MPP⁺) was adopted to reveal the remedial effects of hexagonal boron nitride NPs (hBNs). MTT and LDH showed boosted cells' viability and neural cells protection. Also, hBNs had aggravated defense against destructive oxidants, opposed the MPP⁺-induced cellular apoptosis and exerted neural cryoprotection. ¹⁴⁷

Cerium oxide nanoparticles (CeO₂ NPs) are well-known anti-oxidant materials that resembles the effect of superoxide dismutase and catalase. $^{148-150}$ So, the exploiting of the CeO₂ NPs antioxidant properties was pertinent with PD treatment. *Saccharomyces cerevisiae* yeast model of PD was adopted to investigate the effect of CeO₂ NPs. The α -syn expressing yeast cells showed enhanced viability with gradually increasing concentrations of CeO₂ NPs up to 50 ng. μ L⁻¹ due to the accumulation of α -syn in the plasma membrane instead of the cytoplasm as shown by the cell lysate analysis. Also, there was observable diminishing of the mitochondrial impairment and ROS generation. This improvement was thought to be through the surface absorption of the α -syn on CeO₂ NPs which constrained the α -syn fibrillation. 151 Also, several shapes of ceria NPs were investigated and the flower shape had the most powerful anti-oxidant activity. 152

Although *in-vivo* manganese (Mn) accumulation causes idiopathic PD-resembling symptoms 153 , controversial results were obtained from literature. Based on Sharma and coworkers' study, Mn NPs (30-40 nm) showed impairment of the cognition and motor ability of the rats after 8 days of administration. Additionally, definitive brain injuries were detected in multiple regions accompanied by BBB distortion, dissipated blood flow to the brain and cerebral edema. 154 On the other hand, citrate-capped Mn₂O₃ NPs (C- Mn₂O₃ NPs) ameliorated the PD consequences and chelate the excess Mn so, prevents its subsequent neuro-damage. 155 Wang et al, prepared chiral Mn₂O₃ NPs. The *D*-NPs showed enhanced α -syn fibrillation inhibition and anti-oxidant effect driven by electromagnetic field. 156 Additionally, using dopaminergic neural cells; MN9D, PD lesion was developed then treated with NPs of different facets. It was observable the Mn₃O₄ nanorods with (103) facet had high anti-oxidant capacity and diminishing of α -syn in the cerebrospinal fluid as elaborated by the biological assessments. 157

3.1.5. Polymetallic nanoparticles (nanozymes)-based systems for the treatment of PD

Nanozymes are recently generated nanomaterials class that gained special interest due to its antioxidant capacity by scavenging the harmful hydrogen peroxide into oxygen and water. 158 , 159 For
example, PtCu nanoalloys (NAs) were found to have catalase-like and superoxide dismutase-like
activity. Therefore, they were applied to develop anti-oxidant platform for the amelioration of PD. 158 , 159 Liu et al., investigated the effect of PtCu NAs and found that they are effective antioxidants.
They dissipated the PD through significantly mitigating the level of ROS and α -syn preformed
fibrils (PFF) in the adopted *in-vitro* and *in-vivo* models using primary cortical neurons and in PFF-

treated mice, respectively. ¹⁵⁸ The tri-element nanozyme PtCuSe was developed and found to have anti-oxidant effect. PtCuSe nanozymes caused the degradation of hydrogen peroxide and inhibited xanthine oxidase therefore, they have catalytic activity for hydrogen peroxide reduction (CAT) and resembles the action of super oxide dismutase (SOD). The cell viability of SH-SY5Y cells was investigated and the cytotoxic effect of PtCuSe was minute at concentrations below 120 µg/ml and the cellular uptake investigation by laser confocal microscopy showed a great endocytosis of PtCuSe. MPTP-induced PD *in-vivo* model was adopted, and the nanozyme treated groups showed improvement in the locomotor and cognitive functions. ¹⁵⁹ (**Fig. 8**).

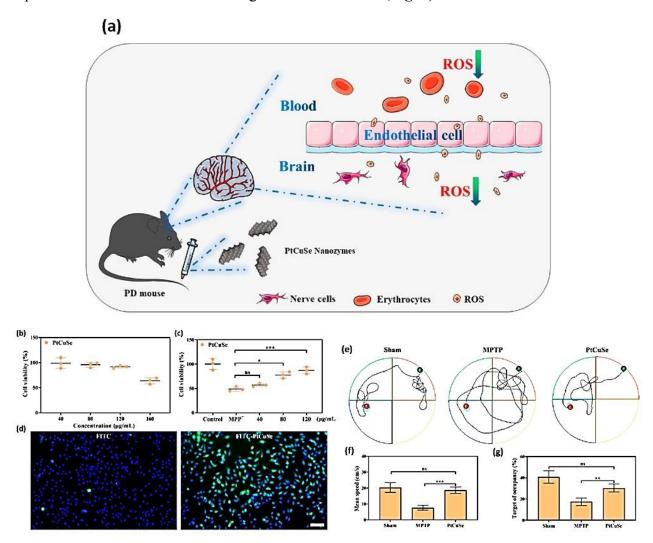


Fig. 8 (a) Scheme of exploiting the antioxidant capability of the nanozyme PtCuSe, **(b)** *in-vitro* cell viability of SH-SY5Y cells after addition of PtCuSe, **(c)** *in-vitro* cell viability of MPP-treated SH-SY5Y cells after addition of PtCuSe, **(d)** cellular uptake of PtCuSe, **(e)** behavioral assessment

Open Access Article. Published on 17 February 2025. Downloaded on 2/21/2025 12:49:03 PM

by Morris water maze test showing the path of mice, (**f** and **g**) the mean time and the relative mean time of the target quadrant. Reproduced from reference ¹⁵⁹

The experiment of chiral $_{L/D}$ -Cu $_{x}$ Co $_{y}$ S SPs comparison between the two counterparts indicated that the $_{D}$ -SPs had preceding ability to prevent the formation of α -syn and disintegrate the already-existing fibrils. This effect was attributed to the capability of $_{D}$ -SPs to generate ROS that exert its effect on α -syn. However, the study did not elucidate any off-target harmful effect of the generated ROS or any subsequent neutralizing cascades.

Li et al, formulated nano-bioconjugate/nanozyme of lactoferrin (Lf)-modified Au-Bi₂Se₃ nanodot (ND) (Lf-Au-Bi₂-Se₃). The nanozyme Lf-Au-Bi₂-Se₃ had characteristic protective antioxidant activity resembles that's of SOD, CAT, POD and GPx, a group of enzymes that scavenges the ROS and normalize its level in the normal cells. The catalytic activity of the Au-Bi₂-Se₃ is attributed to two reasons. First, the Au atom can alter the Se atoms within the lattice and replace it with Bi atoms representing defect points. The Bi defect points would enhance the electron transport and consequently, the catalytic effect. 161 Second, the Au atoms would act synergistically by allowing electron transport. 162 Additionally, the LF-surface modification did not affect the catalytic activity of Au-Bi₂-Se₃ To assess the cellular internalization, bEnd.3 cells were used and Lf-Au-Bi₂-Se₃ was found to have great ability to penetrate the BBB. The Morris water maze invivo model was adopted to assess the treatable effect of Lf-Au-Bi₂-Se₃ and the treated mice exhibited the best intellectual and physical improvement and confirms the pivotal role of Lf in enhancing of the transcytosis in the BBB. Additionally, high levels of tyrosine hydroxylase (TH), healed mitochondria, protected Nissel-positive cells and normalized ex-vivo lipid peroxidation were confirmed in the Lf-Au-Bi₂-Se₃-treated group. The biosafety of the Lf-Au-Bi₂-Se₃ was corroborated by the normally functioning main organs. The uptake of the Lf-Au-Bi₂-Se₃ into the brain cells was found to be 2.67 times more than Au-Bi₂-Se₃ which further confirms the ability of the nanorods to penetrate the BBB. Finally, the clearance of Lf-Au-Bi₂-Se₃ was found to be mainly through the urine which is attributed to the enhanced renal infiltration of the tiny-sized nanorods. 88

4. MNMs-based systems as theranostics for PD

Due to the continuous urge of early accurate detection and effective treatment of PD, researchers invested their efforts to develop theranostic platforms that achieve both goals. Novel

multifunctional nanoparticles were designed to real-time monitor the disease as well as act as a targeted therapeutic agent like SPIONs. As mentioned earlier, SPIONs are biocompatible and biodegradable contrast agents exploited in MRI and can be manipulated to cross BBB with their tunable sizes and shapes and can be green-synthesized pursing great biocompatibility. Moreover, they can remain circulating in the body allowing for better therapy and can be controlled using external magnetic field. 57,163 For instance, SPIONs-loaded liposomes were used as theranostic agent for image-guided drug delivery in glioma condition, therefore, this formulation can be employed in PD. 164

Another study was conducted using resveratrol-Fe₃O₄-loaded liposomes for sustained drug release at the target site using external magnetic field. This formulation was guided using MRI to ensure crossing BBB and reaching the target site in PD rat models. Niu et al. investigated the use of magnetic nanoparticles in gene delivery, where they synthesized N-isopropylacrylamide-acrylic acid-functionalized Fe₃O₄ NPs to confer pH and temperature responsiveness. Then, they photo-immobilized nerve growth factor to the NPs along with short hairpin RNA for gene delivery in PD models, however, they did not assess the MRI properties of such NPs. 166 Thus, this multifunctional system can be further implemented in MRI-guided drug and gene delivery in PD cases. Garcia-Pardo et al. also formulated DA-encapsulated iron nanoparticles made of iron metal nodes polymerized using bidentate ligand as bioinspired nanotheranostic agent. It displayed MRI properties, allowed for efficient DA delivery to PD animal models, and proved to be biosafe *invivo*. 167

Switchable nanoparticles for simultaneous drug/gene delivery and CT imaging were developed. The programmed drug delivery system (MBPCS) was composed of levodopa-quinone gold nanoparticles (GNP) integrated with 2 derivatives of Zwitterionic poly-(carboxybetaine)-based curcumin where the last was releasable via a cleavable link in the DA neurons of PD. B6 peptide was linked on the surface of the nanoparticles facilitating their penetration of the BBB. The intracellular internalization was adopted by using mazindol (MA) that binds preferentially to the DA neurons. Following the endosomal/lysosomal escape, the gene delivery phase starts by releasing α -syn gene (SNCA). The GNP and curcumin are then released because of MBPC degradation. GNP interact with Fe³⁺ and quinone groups allowing the CT. 106

Open Access Article. Published on 17 February 2025. Downloaded on 2/21/2025 12:49:03 PM

5. MNMs and BBB penetration Generally several BBB penetration

Generally, several BBB penetration mechanisms of the different classes of the nanomaterials were described and reviewed elsewhere. 168 Concerning the MNMs, they can penetrate the BBB either by active or passive pathways. Receptor-mediated endo/transcytosis, adsorption-mediated endocytosis and carrier mediated transport are all examples of the active pathways. On the other hand, the passive pathway involves the passage of the hydrophilic small-sized NPs across the endothelial cells. ¹⁶⁸ (**Fig. 9**). Proteins as nerve growth factor (NGF) ¹⁰⁵, ¹¹² poly unsaturated fatty acids as docosahexaenoic acid (DHA)¹¹² and PGM⁸⁷ are all examples of surface functionalizing moieties allowing receptor-mediated endocytosis. ¹⁶⁸ Li et al, prepared Au-Bi₂Se₃ nanodots for PD diseases and functionalized the surface with lactoferrin (LF) to enable the active targeting of the BBB through receptor-driven transcytosis. 88 The B6 peptide (CGHKAKGPRK) was used to design switchable AuNPs-based theranostic formulation where the penetration of the BBB was confirmed and the cellular internalization of SH-SY5Y cells was described to be through caveolae and clathrin-driven endocytosis. 113 AuNCs (2.5 \pm 1 nm) were investigated, and their penetration of the BBB was proven. The AuNCs were injected intraperitoneally in mice and after 6 h of 20 mg/kg injection, the AuNCs were detected by TEM in the SN sectors in addition to the neurons. ⁹⁶ Cheng et al, investigated the penetration of an iron-based nanoformulation through an *in-vitro* model and found that the cellular penetration can be through 3 different pathways. 135 Consequently, the MNMs were reported to penetrate the BBB through different mechanisms depending on the sizes and surface properties.

Open Access Article. Published on 17 February 2025. Downloaded on 2/21/2025 12:49:03 PM.

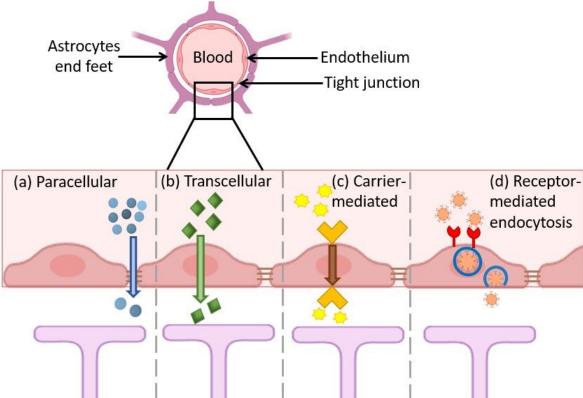


Fig. 9. Mechanisms of BBB penetration by the nanomaterials. Passive pathways include (a) paracellular and transcellular penetration while the active transport pathways include (c) carriermediated and (d) receptor-mediated BBB penetration.

Toxicity and biocompatibility of MNMs

Majorly, the axiomatic transportation of nanomaterials into clinical application is constrained by the inscrutable toxicity effects on the CNS and specifically, the brain. Heavy metals were previously reported to induces neurotoxicity that may resemble the PD-associated manifestation. ^{169,170} Therefore, the toxicity of the nano-constructed counterparts would be of great importance and pertinent as many of these nanoparticles release metallic ions after *in-vivo* administration. Additionally, the tiny size of the nanostructures permeates their smooth flow across the BBB and distribution within the brain parts remarkably, to the SP which has great importance due to the existence of dopaminergic neurons. ^{6,171} PD is associated with other physiological alterations such as inflammation, α-syn aggregation and impaired antioxidant response. All these impairments were observable from the exposure to several MNMs. 172,173, 174 For instance, TiO₂ NPs affect the hippocampus, cerebellum and substantia nigra. ¹⁷⁵, ⁶ Certain specific pathways of the TiO₂-induced toxicity have been illustrated in literature proving detrimental effect. Wu and Xie conducted a study to investigate the *in-vitro* and *in-vivo* toxicity of TiO₂ adopting PC12 cells and zebrafish embryos, respectively. The results corroborated the presence of neurotoxic effect due to the accumulation of TiO₂ NPs and the generation of ROS. Also, the degradation of the dopaminergic neurons was observable. ¹⁷⁶ Also, TiO₂ administration caused electrolyte imbalance and activated inflammatory response by stimulating IL-6 and NF-KB. ¹⁷⁷, ¹⁷⁸ Moreover, TiO₂ promoted cerebral stimulating the oxidative stress-driven cellular damage by constraining the anti-oxidants factors ¹⁷⁹ and elevating the intra-mitochondrial ROS alone or with simultaneous exposure to AgNPs. ¹⁸⁰ From other studies, TiO₂ caused the accumulation of α-syn that mitigates the DA in the brain which is a substantial mechanism causing PD. ¹⁸¹, ¹⁷², ¹⁸²

Despite the remedial effect of green-synthesized zinc oxide NPs (ZnO NPs) ¹⁸³, other studies reported their cerebral cytotoxicity. ¹⁸⁴, ¹⁷¹ A study performed by Jin and coworkers compared the induced cytotoxic effect by several ZnO-based nanostructures. ZnO NPs, long ZnO nanorods (1-ZnO NRs) and short ZnO nanorods (s-ZnO NRs) were prepared and the *in-vitro* assessment was performed using human neuroblastoma cells SH-SY5Y and revealed that 1-ZnO NPs had the least toxicity (LD50 = 17 µg/ml). Several concentrations of 1-ZnO NPs were tested using zebrafish larvae in-vivo model. From the investigation of the brain, 1-ZnO NRs high doses stimulated the ROD production that consequently, sequestered the motor ability and neurons development, disruption of dopaminergic neurons and cerebral apoptosis coherently leading to PD-resembling manifestations. ¹⁸⁴ The toxicity of the Zn²⁺ ions from ZnO NPs was investigated through *in-vitro* and *in-vivo* assessments. The cell viability of the glial cells A172 was greatly decreased after 1 day of exposure and DNA damage was observable after only 3h as confirmed by comet assay. The zebrafish embryos were exposed to gradient concentrations of ZnO NPs. After 96 hours postfertilization (hpf), locomotor impairment was resulted suggesting the possible role of the released Zn²⁺. The cytotoxicity and genotoxicity were believed to be caused by enhanced ROS production. Additionally, the locomotor disorder was not elucidated explicitly but attributed to dopaminergic cells destruction and therefore, affecting the muscles and nerves (Fig. 10). ¹⁸⁵ Table 3 summarizes some PD-related toxicities associated with MNMs exposure.

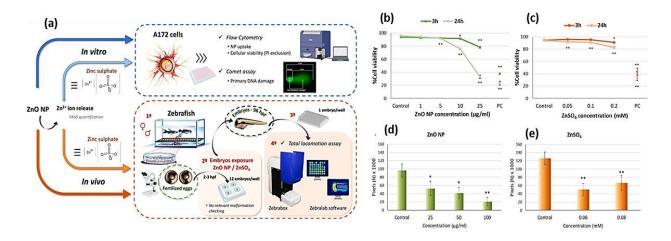


Fig. 10. (a) Schematic illustration of the prepared ZnO NPs and the effect of the released Zn²⁺ ions on A172 cells and the adopted *in-vivo* model using zebrafish embryos, **(b and c)** *in-vitro* cell viability assessment of A172 compared to ZnSO₄, **(d and e)** locomotor impairment of zebrafish embryos after exposure to ZnO NPs and ZnSO₄. Adapted from reference ¹⁸⁵ with permission from Elsevier, copyright 2024.

Table 3. Several MNMs' exposure duration and the resulting neurotoxicity

Type & size of NPs	Exposure duration	Biocompatibility & Toxicity	Ref.
Fe ₃ O ₄ , about 10 nm	24 h	Neuro-destructive effect through oxidative stress. Interaction between the iron and DA may induce neurons toxicity by generating toxic products. The aggregated α -syn can cause accretion of the iron level that leads to generation of OH destructive radicals.	186 187 188 189
Ag NPs, less than 100 nm	28 days	Induced neuronal apoptosis and cellular degeneration known as dark neurons due to ROS production and exaggerated inflammatory response.	190
Co NPs, less than 100 nm	24 h and 6 days	Destructive oxidative stress evidenced by the depletion of GSH, neurotoxicity due to calcium homeostasis and lipid peroxidation. Also, mitochondrial swelling and structures that resembles fibrosis were notable, leading to ferroptosis-like cell death.	191
CuO NPs, 50 nm	60 and 120 h	α-syn aggregates, promote pertinent cellular cytotoxicity	192
CuS NPs, 77.89 nm	24 h	Cytotoxic effect at low concentration, caused developmental neurotoxicity	193 194
NiO NPs, 50 nm	24 h	Altered the fibrillation kinetics of α-syn, enhanced ROS production due to the alleviated defensive pathways and the enhanced inflammatory mediators' expression	195

Long ZnO nanorods (1-ZnO NRs), diamter: 159.3 ± 17.9 nm and length: 1.1 ± 0.15 μ m	24-144 h post fertilization (hpf)	High doses stimulated the ROD production, sequestered the motor ability and neurons development, disruption of dopaminergic neurons and cerebral apoptosis leading to PD-resembling manifestations	184	
--	-----------------------------------	--	-----	--

7. Challenges and future perspectives

The bridling of clinical improvement in the journey of PD starting from the detection to treatment is attributed to the lacunae that can be recapitulated as long-time of asymptomatic period, absence of sharply definitive biomarker, heterogenous nature of the disease, dearth of effective treatment and resistance development. The use of MNMs represents a promising but complex avenue for addressing these issues. While NPs have shown potential in enhancing detection through sensitive biosensors and improving therapeutic delivery, challenges persist. These include ensuring biocompatibility and stability in biological systems, achieving precise targeting to diseased cells, and avoiding off-target effects.⁴⁸ The encapsulation of the MNMs into emulsions would provide prolonged drug release. Also, the design of the carrier medium to be NIR-responsive would confer more specific release in the target cells/sites. 111 The incorporation of the MNMs into biocompatible nanoshells 196 or porous nanocarriers would add a protective shield from pre-mature in-vivo degradation. Moreover, the surface modification of the MNMs with targeting ligands would enhance the on-site neurons-specific drug release 88, 197. The exploiting of specific BBBtransporters would also enhance the cerebral drug delivery surface modification of the MNMs by ligands specific to these receptor, reviewed by Mhaske et al. 198 Furthermore, scaling up from laboratory success to clinical application involves overcoming significant hurdles in manufacturing and regulatory approval. Concerning the MNMs, the design of a formulation for PD therapy or *in-vivo* detection as imaging would require prolonged testing to ensure the safety during the use and subsequent complete clearance from the body. Additionally, challenging of animal models reciprocation, ethical issues about human samples and the ambiguity to provide clear mechanistic molecular pathways of aging and neurodegenerative diseases, particularly PD, are all insistent issues that need to be addressed. 199,200 Future research should focus on refining nanoparticle technologies to enhance their specificity, reduce potential toxicity, and improve targeting mechanisms. Advancements in AI and machine learning could further aid in analyzing data from nanoparticle-based detection and therapies, leading to more personalized and effective

treatment strategies. Addressing these challenges through innovative approaches in biomarker discovery, model development, and therapeutic applications will be crucial for advancing clinical outcomes and improving the quality of life for individuals with PD.

Acknowledgement

The authors extend their appreciation to the Science, Technology and Innovation Funding Authority (STDF), Egypt, for funding and supporting research related to this review through the funds (Capacity Building Fund, CB-22808 and STDF FLUG Call 1-Project ID 46715).

Biographies



Amira Mansour is a pharmacist, graduated from Misr international university (MIU). She obtained her PhD in Nanoscience – Nanomedicine from Zewail City of Science and Technology, Egypt in 2023. Her research focused on the biomedical applications of diverse nanomaterials especially, in diseases' treatment as breast and hepatic cancer. Currently, she is an Adjunct Assistant Professor at the Nanoscience Program and a Senior Researcher at the CMS in Zewail City.



Mariam HossamEldin is a Nanoscience/Nanomedicine Master's student and a teaching assistant in Chemistry and Nanoscience Program in Zewail City of Science and Technology. She holds a bachelor's degree in Nanoscience (Nanomedicine Concentration) along with a minor in drug design and development. Her research work is specialized in nanoformulations exploited for drug delivery in pulmonary route. She has also investigated the use of metallic nanoparticles in colorimetric biosensors for the detection of hepatitis C virus (HCV) and developed a prototype.

Open Access Article. Published on 17 February 2025. Downloaded on 2/21/2025 12:49:03 PM



Ibrahim M. El-Sherbiny (BSc-Hons, MSc, PhD, DSc, FAAS) is a Tenured Professor of Nanotechnology and Nanomedicine, and the Founding Chairman of Nanoscience Program, and Director of the Center of Materials Science (CMS) at Zewail City of Science and Technology. El-Sherbiny's research focuses on the design and development of new classes of smart nano- and nano-in-micro matrices using advanced nanotechnology techniques and evaluating these new structures as potential candidates in various biotechnological, pharmaceutical and biomedical applications including targeted and controlled drug delivery, tissue engineering, regenerative medicine, and biosensing. El-Sherbiny's achievements have garnered widespread global recognition, with over 70 national and international awards, including Order of Egyptian Republic in Science and Arts, State Award for Excellence in Advanced Sciences. He has been honored by several esteemed organizations worldwide, showcasing his global impact and influence in the fields of nanotechnology and Nanomedicine.

References

- 1. Jinqiao Zhu, Yusha Cui, Junjiao Zhang, Rui Yan, Dongning Su, Dong Zhao, Anxin Wang TF. Prevalence of Parkinson's disease and its trend from 1980 to 2023: A systematic review and meta-analysis. *Lancet*. 2023;(127):1-26.
- 2. Lisa M Shulman. Is there a connection between estrogen and Parkinson's disease? *Park Relat disorers*. 2002;8(5):289-295.
- 3. Tysnes OB, Storstein A. Epidemiology of Parkinson's disease. *J Neural Transm*. 2017;124(8):901-905. doi:10.1007/s00702-017-1686-y
- 4. Organization WH. Neurological disorders: public health challenges. *World Heal Organ*. Published online 2006.
- 5. Chen SY, Tsai ST. Epidemiology of Parkinson's disease. *lancet Neurol*. 2006;5(6):525-535. doi:10.1016/S1016-3190(10)60044-4
- 6. Heidari Z, Mohammadipour A, Haeri P, Ebrahimzadeh-bideskan A. The effect of titanium dioxide nanoparticles on mice midbrain substantia nigra. *Iran J Basic Med Sci*. 2019;22(7):745. doi:10.22038/ijbms.2019.33611.8018
- 7. Haeri P, Mohammadipour A, Heidari Z, Ebrahimzadeh-bideskan A. Neuroprotective effect of crocin on substantia nigra in MPTP-induced Parkinson's disease model of mice. *Anat Sci Int.* 2019;94(1):119-127. doi:10.1007/s12565-018-0457-7
- 8. Cragg, S. J., & Rice ME. DAncing past the DAT at a DA synapse. *Trends Neurosci*. 2004;27(5):270-277.
- 9. Bartels AL, Leenders KL. Parkinson's disease: The syndrome, the pathogenesis and pathophysiology. *cortex*. 2009;45:915-921. doi:10.1016/j.cortex.2008.11.010
- 10. Mcgeer PL, Itagaki S, Boyes BE, Mcgeer EG. Reactive microglia are positive for HLA-DR in the substantia nigra of Parkinson's and Alzheimer's disease brains. *Neurology*.

- 1988;38(8):1285.
- 11. Ahmadinejad F, Møller SG, Hashemzadeh-Chaleshtori M, Bidkhori G, Jami MS. Molecular mechanisms behind free radical scavengers function against oxidative stress. *Antioxidants*. 2017;6(3):1-15. doi:10.3390/antiox6030051
- 12. Franco-Iborra S, Cuadros T, Parent A, Romero-Gimenez J, Vila M, Perier C. Defective mitochondrial protein import contributes to complex I-induced mitochondrial dysfunction and neurodegeneration in Parkinson's disease. *Cell Death Dis.* 2018;9(11):1122. doi:10.1038/s41419-018-1154-0
- 13. Chawla S, Kalyane D, Tambe V, Deb PK, Kalia K, Tekade RK. Evolving nanoformulation strategies for diagnosis and clinical interventions for Parkinson's disease. *Drug Discov Today*. 2020;25(2):392-405. doi:10.1016/j.drudis.2019.12.005
- 14. Kim TD, Choi E, Rhim H, Paik SR, Yang C hak. -Synuclein has structural and functional similarities to small heat shock proteins. *Biochem Biophys Res Commun*. 2004;4(324):1352-1359. doi:10.1016/j.bbrc.2004.09.208
- 15. Sode K, Ochiai S, Kobayashi N, Usuzaka E. Effect of Reparation of Repeat Sequences in the Human α -Synuclein on Fibrillation Ability. *Int J Biol Sci.* 2007;3(1):1.
- 16. Teixeira MI, Lopes CM, Amaral MH, Costa PC. Current insights on lipid nanocarrier-assisted drug delivery in the treatment of neurodegenerative diseases. *Eur J Pharm Biopharm*. 2020;149:192-217. doi:10.1016/j.ejpb.2020.01.005
- 17. Zambon F, Cherubini M, Fernandes HJR, et al. Cellular α-synuclein pathology is associated with bioenergetic dysfunction in Parkinson's iPSC-derived dopamine neurons. *Hum Mol Genet*. 2019;28(12):2001-2013. doi:10.1093/hmg/ddz038
- 18. Braak H, Del Tredici K, Rüb U, De Vos RAI, Jansen Steur ENH, Braak E. Staging of brain pathology related to sporadic Parkinson's disease. *Neurobiol Aging*. 2003;24(2):197-211. doi:10.1016/S0197-4580(02)00065-9
- 19. Dubois, Bruno and BP. Cognitive deficits in Parkinson's disease. *J Neurol*. 1997;244:2-8.
- 20. Lorraine V Kalia and Anthony E Lang. Parkinson's disease. *Lancet*. 2015;29(389):896-912.
- 21. Armstrong MJ, Okun MS. Diagnosis and Treatment of Parkinson Disease: A Review. *JAMA J Am Med Assoc.* 2020;323(6):548-560. doi:10.1001/jama.2019.22360
- 22. Chaudhuri KR, Prieto-Jurcynska C, Naidu Y, et al. The nondeclaration of nonmotor symptoms of Parkinson's disease to health care professionals: An international study using the nonmotor symptoms questionnaire. *Mov Disord*. 2010;25(6):704-709. doi:10.1002/mds.22868
- 23. Berg D, Postuma RB, Adler CH, et al. MDS research criteria for prodromal Parkinson's disease. *Mov Disord*. 2015;30(12):1600-1611. doi:10.1002/mds.26431
- 24. Hospital H. Parkinson's disease and osteoporosis. *Age Ageing*. 2013;42(2):156-162. doi:10.1093/ageing/afs161
- 25. Hong, Chien Tai, Han-Hwa Hu, Lung Chan and CHB. Prevalent cerebrovascular and cardiovascular disease in people with Parkinson's disease: a meta-analysis. *Clin Epidemiol*. Published online 2018:1147-1154. doi:10.2147/CLEP.S163493
- 26. Ma S, Yang Q, Zhang W, et al. Silver nanoclusters and carbon dots based light-addressable sensors for multichannel detections of dopamine and glutathione and its applications in probing of parkinson's diseases. *Talanta*. 2020;219:121290. doi:https://doi.org/10.1016/j.talanta.2020.121290

- 27. Meade RM, Fairlie DP, Mason JM. Alpha-synuclein structure and Parkinson's disease lessons and emerging principles. *Mol Neurodegener*. 2019;14(1):29. doi:10.1186/s13024-019-0329-1
- 28. Gómez-Benito M, Granado N, García-Sanz P, Michel A, Dumoulin M, Moratalla R. Modeling Parkinson's Disease With the Alpha-Synuclein Protein. *Front Pharmacol*. 2020;11:356. doi:10.3389/fphar.2020.00356
- 29. Srinivasan E, Chandrasekhar G, Chandrasekar P, et al. Alpha-Synuclein Aggregation in Parkinson's Disease. *Front Med.* 2021;8. doi:10.3389/fmed.2021.736978
- 30. Mandala SH, Liu TJ, Chen CM, et al. Enhanced Plasmonic Biosensor Utilizing Paired Antibody and Label-Free Fe3O4 Nanoparticles for Highly Sensitive and Selective Detection of Parkinson's α-Synuclein in Serum. *Biosensors*. 2021;11(10). doi:10.3390/bios11100402
- 31. Aminabad ED, Mobed A, Hasanzadeh M, Hosseinpour Feizi MA, Safaralizadeh R, Seidi F. Correction: Sensitive immunosensing of α-synuclein protein in human plasma samples using gold nanoparticles conjugated with graphene: an innovative immuno-platform towards early stage identification of Parkinson's disease using point of care (POC) analys. *RSC Adv.* 2022;12(10):5765. doi:10.1039/d2ra90011d
- 32. Jens O. Watzlawik Xu Hou DFCRSKBTFGMGHMDJSZKWCRSOARGBDWDMSGFCF, Springer W. Sensitive ELISA-based detection method for the mitophagy marker p-S65-Ub in human cells, autopsy brain, and blood samples. *Autophagy*. 2021;17(9):2613-2628. doi:10.1080/15548627.2020.1834712
- 33. Tsao HH, Huang CG, Wu YR. Detection and assessment of alpha-synuclein in Parkinson disease. *Neurochem Int.* 2022;158:105358. doi:https://doi.org/10.1016/j.neuint.2022.105358
- 34. Santos AM, Wong A, Ferreira LMC, et al. Multivariate optimization of a novel electrode film architecture containing gold nanoparticle-decorated activated charcoal for voltammetric determination of levodopa levels in pre-therapeutic phase of Parkinson's disease. *Electrochim Acta*. 2021;390:138851. doi:https://doi.org/10.1016/j.electacta.2021.138851
- 35. Mobed A, Razavi S, Ahmadalipour A, Shakouri SK, Koohkan G. Biosensors in Parkinson's disease. *Clin Chim Acta*. 2021;518:51-58. doi:https://doi.org/10.1016/j.cca.2021.03.009
- 36. Samson E, Noseworthy MD. A review of diagnostic imaging approaches to assessing Parkinson's disease. *Brain Disord*. 2022;6:100037. doi:https://doi.org/10.1016/j.dscb.2022.100037
- 37. Tolosa E, Garrido A, Scholz SW, Poewe W. Challenges in the diagnosis of Parkinson's disease. *Lancet Neurol.* 2021;20(5):385-397. doi:10.1016/S1474-4422(21)00030-2
- 38. Ghosh N, Sinha K, Sil PC. A review on the new age methodologies for early detection of Alzheimer's and Parkinson's disease. *Basic Clin Pharmacol Toxicol*. 2024;134(5):602-613. doi:https://doi.org/10.1111/bcpt.14003
- 39. Keserwani PK, Das S, Sarkar N. A comparative study: prediction of parkinson's disease using machine learning, deep learning and nature inspired algorithm. *Multimed Tools Appl*. Published online 2024. doi:10.1007/s11042-024-18186-z
- 40. Wang J, Xue L, Jiang J, et al. Diagnostic performance of artificial intelligence-assisted PET imaging for Parkinson's disease: a systematic review and meta-analysis. *npj Digit*

Med. 2024;7(1):17. doi:10.1038/s41746-024-01012-z

This article is licensed under a Creative Commons Attribution-NonCommercial 3.0 Unported Licence

- 41. Yang Y, Yuan Y, Zhang G, et al. Artificial intelligence-enabled detection and assessment of Parkinson's disease using nocturnal breathing signals. *Nat Med*. 2022;28(10):2207-2215. doi:10.1038/s41591-022-01932-x
- 42. Sigcha L, Borzì L, Amato F, et al. Deep learning and wearable sensors for the diagnosis and monitoring of Parkinson's disease: A systematic review. *Expert Syst Appl*. 2023;229:120541. doi:https://doi.org/10.1016/j.eswa.2023.120541
- 43. Sotirakis C, Su Z, Brzezicki MA, et al. Identification of motor progression in Parkinson's disease using wearable sensors and machine learning. *npj Park Dis*. 2023;9(1):142. doi:10.1038/s41531-023-00581-2
- 44. Moreau C, Rouaud T, Grabli D, et al. Overview on wearable sensors for the management of Parkinson's disease. *npj Park Dis.* 2023;9(1):153. doi:10.1038/s41531-023-00585-y
- 45. Dixit S, Bohre K, Singh Y, et al. A Comprehensive Review on AI-Enabled Models for Parkinson's Disease Diagnosis. *Electronics*. 2023;12(4). doi:10.3390/electronics12040783
- 46. Perju-Dumbrava L, Barsan M, Leucuta DC, et al. Artificial intelligence applications and robotic systems in Parkinson's disease (Review). *Exp Ther Med*. 2022;23(2):153. doi:10.3892/etm.2021.11076
- 47. Qin C, Xia J, Wen Y, Wang J, Zhong C. A new immunofluorescence determination of Parkinson's disease biomarkers using silver nanoparticles. *Alexandria Eng J*. 2025;111:404-414. doi:https://doi.org/10.1016/j.aej.2024.10.069
- 48. Scarpa E, Cascione M, Griego A, Pellegrino P, Moschetti G, De Matteis V. Gold and silver nanoparticles in Alzheimer's and Parkinson's diagnostics and treatments. *Ibrain*. 2023;9(3):298-315. doi:https://doi.org/10.1002/ibra.12126
- 49. Du Q, Liu Y, Fan M, Wei S, Ismail M, Zheng M. PEG length effect of peptide-functional liposome for blood brain barrier (BBB) penetration and brain targeting. *J Control Release*. 2024;372:85-94. doi:https://doi.org/10.1016/j.jconrel.2024.06.005
- 50. Abujamai J, Satar R, Ansari SA. Designing and Formulation of Nanocarriers for "Alzheimer's and Parkinson's" Early Detection and Therapy. *CNS & amp; Neurol Disord Drug Targets (Formerly Curr Drug Targets CNS & amp; Neurol Disord*. 2024;23(10):1251-1262. doi:https://doi.org/10.2174/0118715273297024240201055550
- 51. Parihar A, Gaur K, Khan R. Chapter 10 Rapid diagnostic assays for the detection of Alzheimer's and Parkinson's diseases. In: Parihar A, Khan R, Srivastava AK, eds. *Smart Diagnostics for Neurodegenerative Disorders*. Academic Press; 2024:221-250. doi:https://doi.org/10.1016/B978-0-323-95539-3.00008-9
- 52. Chawla S, Kalyane D, Tambe V, Deb PK, Kalia K, Tekade RK. Evolving nanoformulation strategies for diagnosis and clinical interventions for Parkinson's disease. *Drug Discov Today*. 2020;25(2):392-405. doi:https://doi.org/10.1016/j.drudis.2019.12.005
- 53. Thakur R, Saini AK, Taliyan R, Chaturvedi N. Neurodegenerative diseases early detection and monitoring system for point-of-care applications. *Microchem J.* 2025;208:112280. doi:https://doi.org/10.1016/j.microc.2024.112280
- 54. Orzari LO, Silva LRG e., de Freitas RC, Brazaca LC, Janegitz BC. Lab-made disposable screen-printed electrochemical sensors and immunosensors modified with Pd nanoparticles for Parkinson's disease diagnostics. *Microchim Acta*. 2024;191(1):76. doi:10.1007/s00604-023-06158-3
- 55. Kim JH, Suh YJ, Park D, et al. Technological advances in electrochemical biosensors for the detection of disease biomarkers. *Biomed Eng Lett.* 2021;11(4):309-334.

- doi:10.1007/s13534-021-00204-w
- 56. Zhang R, Wang S, Huang X, et al. Gold-nanourchin seeded single-walled carbon nanotube on voltammetry sensor for diagnosing neurogenerative Parkinson's disease. *Anal Chim Acta*. 2020;1094:142-150. doi:https://doi.org/10.1016/j.aca.2019.10.012
- 57. Zoey FL, Palanivel M, Padmanabhan P, Gulyás B. Parkinson's Disease: A Nanotheranostic Approach Targeting Alpha-Synuclein Aggregation. *Front Cell Dev Biol*. 2021;9. doi:10.3389/fcell.2021.707441
- 58. Natarajan B, Kannan P, Guo L. Metallic nanoparticles for visual sensing: Design, mechanism, and application. *Chinese J Struct Chem*. 2024;43(9):100349. doi:https://doi.org/10.1016/j.cjsc.2024.100349
- 59. Sonuç Karaboğa MN, Sezgintürk MK. A nano-composite based regenerative neuro biosensor sensitive to Parkinsonism-associated protein DJ-1/Park7 in cerebrospinal fluid and saliva. *Bioelectrochemistry*. 2021;138:107734. doi:https://doi.org/10.1016/j.bioelechem.2020.107734
- 60. Zhu H, Fohlerová Z, Pekárek J, Basova E, Neužil P. Recent advances in lab-on-a-chip technologies for viral diagnosis. *Biosens Bioelectron*. 2020;153(January). doi:10.1016/j.bios.2020.112041
- 61. Ge S, Chen G, Deng J, et al. Multiplex signal amplification strategy-based early-stage diagnosis of Parkinson's disease on a SERS-enabled LoC system. *Anal Chim Acta*. 2023;1247:340890. doi:https://doi.org/10.1016/j.aca.2023.340890
- 62. Ren X, Zhang Q, Yang J, et al. Dopamine Imaging in Living Cells and Retina by Surface-Enhanced Raman Scattering Based on Functionalized Gold Nanoparticles. *Anal Chem*. 2021;93(31):10841-10849. doi:10.1021/acs.analchem.1c01108
- 63. Hu J, Gao G, He M, et al. Optimal route of gold nanoclusters administration in mice targeting Parkinson's disease. *Nanomedicine*. 2020;15(6):563-580. doi:10.2217/nnm-2019-0268
- 64. Adam H, Gopinath SCB, Krishnan H, et al. Cyclic and differential pulse voltammetric measurements on fibrils formation of alpha synuclein in Parkinson's disease by a gold interdigitated tetraelectrodes. *Process Biochem*. 2024;136:212-220. doi:https://doi.org/10.1016/j.procbio.2023.11.019
- 65. Nagaraj K, Thangamuniyandi P, Velmurugan G, Alotaibi KM, Raja K, Sharma BK. Green Synthesis of Eosin-Y Coated Silver Nanoparticles for Sensitive and Selective Fluorometric Detection of L-Dopa. *J Fluoresc*. Published online 2025. doi:10.1007/s10895-024-04116-7
- 66. Badillo-Ramírez I, Landeros-Rivera B, Saniger JM, Popp J, Cialla-May D. SERS-based detection of 5-S-cysteinyl-dopamine as a novel biomarker of Parkinson {'}s disease in artificial biofluids. *Analyst*. 2023;148(8):1848-1857. doi:10.1039/D3AN00027C
- 67. Badillo-Ramírez I, Saniger JM, Popp J, Cialla-May D. SERS characterization of dopamine and in situ dopamine polymerization on silver nanoparticles. *Phys Chem Chem Phys.* 2021;23(21):12158-12170. doi:10.1039/D1CP00966D
- 68. Rouhani M, Soleymanpour A. Preparation of Dawson heteropolyacid-embedded silver nanoparticles/graphene oxide nanocomposite thin film used to modify pencil graphite electrode as a sensor for trace electrochemical sensing of levodopa. *Mater Sci Eng C*. 2020;117:111287. doi:https://doi.org/10.1016/j.msec.2020.111287
- 69. Huang Z, Lu H, Dong H, et al. Fe3O4/Ni nanoparticles anchored nitrogen-doped porous carbon derived from core-shell MOF for simultaneous electrochemical detection of

- dopamine and 5-hydroxytryptamine. *Talanta*. 2025;286:127522. doi:https://doi.org/10.1016/j.talanta.2025.127522
- 70. Abdelmonem M, Albert EL, Alhadad MA, Abdullah CA. Plant-Polyphenol-Mediated Synthesis of Magnetic Biocompatible Iron Oxide Nanoparticles for Diagnostic Imaging and Management of Neurodegenerative Diseases. *Precis Nanomedicine*. 2024;7(1):1233-1251. doi:10.33218/001c.92424
- 71. An L, Tao Q, Wu Y, et al. Synthesis of SPIO Nanoparticles and the Subsequent Applications in Stem Cell Labeling for Parkinson's Disease. *Nanoscale Res Lett*. 2021;16(1):107. doi:10.1186/s11671-021-03540-z
- 72. Liu X ge, Lu S, Liu D qun, et al. ScFv-conjugated superparamagnetic iron oxide nanoparticles for MRI-based diagnosis in transgenic mouse models of Parkinson's and Huntington's diseases. *Brain Res.* 2019;1707:141-153. doi:https://doi.org/10.1016/j.brainres.2018.11.034
- 73. Yang SY, Chiu MJ, Lin CH, et al. Development of an ultra-high sensitive immunoassay with plasma biomarker for differentiating Parkinson disease dementia from Parkinson disease using antibody functionalized magnetic nanoparticles. *J Nanobiotechnology*. 2016;14(1):41. doi:10.1186/s12951-016-0198-5
- 74. Luo S, Ma C, Zhu MQ, Ju WN, Yang Y, Wang X. Application of Iron Oxide Nanoparticles in the Diagnosis and Treatment of Neurodegenerative Diseases With Emphasis on Alzheimer's Disease. *Front Cell Neurosci*. 2020;14. doi:10.3389/fncel.2020.00021
- 75. Chen Q, Xue Y, Huang Y, Guo W, Wan M, Shen J. Mg-based micromotors for electrochemical detection of Parkinson's disease blood biomarkers. *Sensors Actuators B Chem.* 2024;402:135035. doi:https://doi.org/10.1016/j.snb.2023.135035
- 76. Zhang X, Wu M, Gopinath SCB, Chen Y. Dual-probe sandwich for Lewy body detection on nano-composite modified dielectric surface to determine Parkinson's disease. *Sens Bio-Sensing Res.* 2023;42:100599. doi:https://doi.org/10.1016/j.sbsr.2023.100599
- 77. Aziz A, Asif M, Azeem M, et al. Self-stacking of exfoliated charged nanosheets of LDHs and graphene as biosensor with real-time tracking of dopamine from live cells. *Anal Chim Acta*. 2019;1047:197-207. doi:10.1016/j.aca.2018.10.008
- 78. Ge CY, Rahman MM, Zhang W, et al. An Electrochemical Immunosensor Based on a Self-Assembled Monolayer Modified Electrode for Label-Free Detection of α-Synuclein. *Sensors*. 2020;20(3). doi:10.3390/s20030617
- 79. Di Mari GM, Scuderi M, Lanza G, et al. Pain-Free Alpha-Synuclein Detection by Low-Cost Hierarchical Nanowire Based Electrode. *Nanomaterials*. 2024;14(2). doi:10.3390/nano14020170
- 80. Chang CW, Yang SY, Yang CC, Chang CW, Wu YR. Plasma and Serum Alpha-Synuclein as a Biomarker of Diagnosis in Patients With Parkinson's Disease. *Front Neurol*. 2020;10. doi:10.3389/fneur.2019.01388
- 81. Wu Q, Tan R, Mi X, Tu Y. Electrochemiluminescent aptamer-sensor for alpha synuclein oligomer based on a metal–organic framework. *Analyst*. 2020;145(6):2159-2167. doi:10.1039/D0AN00169D
- 82. Obermeier B, Daneman R, Ransohoff RM. review Development, maintenance and disruption of the blood-brain barrier. *Nat Med.* 2013;19(12):1584-1596. doi:10.1038/nm.3407
- 83. Zhang TT, Li W, Meng G, Wang P, Liao W. Strategies for transporting nanoparticles

- across the blood-brain barrier. *Biomater Sci.* 2016;4(2):219-229. doi:10.1039/c5bm00383k
- 84. Merajoth AL, Pillai PS, Iype T. Clinical Response of Levodopa Carbidopa Combination in Patients with Idiopathic Parkinsonism. *J Clin diagnostic Res JCDR*. 2016;10(5):FC07. doi:10.7860/JCDR/2016/16043.7886
- 85. Rascol O, Payoux P, Ory F, Ferreira JJ, Brefel-courbon C, Montastruc J louis. Limitations of Current Parkinson's Disease Therapy. *Ann Neurol Off J Am Neurol Assoc Child Neurol Soc.* 2003;53(S3):S3-15. doi:10.1002/ana.10513.Address
- 86. Oertel WH. Recent advances in treating Parkinson's disease. *F1000Research*. 2017;6:1-14. doi:10.12688/f1000research.10100.1
- 87. Peranic N, Barbir R, Hall CR, Smith TA, Sani MA. Spectroscopic study of L-DOPA and dopamine binding on novel gold nanoparticles towards more efficient drug-delivery system for Parkinson's disease. *Spectrochim Acta Part A Mol Biomol Spectrosc*. 2022;5(268):120707. doi:10.1016/j.saa.2021.120707
- 88. Lihua Li, Yao Lu, Xingyi Xu, Xianfeng Yang, Lingling Chen, Chengming Jiang YW, Wenyan Hu, Xiaoming Wei and ZY. Catalytic-Enhanced Lactoferrin-Functionalized Au-Bi2Se3 Nanodots for Parkinson's Disease Therapy via Reactive Oxygen Attenuation and Mitochondrial Protection. *Adv Heal care Mater*. 2021;10(13):2100316.
- 89. Eduarda M, Emilio P, Andrez R, Avila M de, Avila M de. Effects of gold nanoparticles administration through behavioral and oxidative parameters in animal model of parkinson's disease. *Colloids Surfaces B Biointerfaces*. 2020;196:111302.
- 90. Liu J, Gao D, Hu D, Lan S, Liu Y, Zheng H. Delivery of Biomimetic Liposomes via Meningeal Lymphatic Vessels Route for Targeted Therapy of Parkinson's Disease. *research*. 2023;6:0030. doi:10.34133/research.0030
- 91. Sharma, G., Wadhwa, K., Kumar, S., Singh, G. and Pahwa R. Revolutionizing Parkinson's treatment: Harnessing the potential of intranasal nanoemulsions for targeted therapy. *Drug Deliv Transl Res*. Published online 2025:1-19.
- 92. Danz K, Fleddermann J, Koch M, et al. Evaluation of the Transport and Binding of Dopamine-Loaded PLGA Nanoparticles for the Treatment of Parkinson's Disease Using In Vitro Model Systems. *Pharmaceutics*. 2024;16(5571).
- 93. Gao G, Zhang M, Gong D, Chen R, Hu X, Sun T. The size-effect of gold nanoparticles and nanoclusters in the inhibition of amyloid-β fibrillation. *Nanoscale*. 2017;9(12):4107-4113. doi:10.1039/c7nr00699c
- 94. Xiao L, Wei F, Zhou Y, Anderson GJ, Frazer DM, Lim YC. Dihydrolipoic Acid Gold Nanoclusters Regulate Microglial Polarization and Have the Potential To Alter Neurogenesis. *nano Lett.* 2020;20(1):478-495. doi:10.1021/acs.nanolett.9b04216
- 95. Ma X, Lee S, Fei X, et al. Proteasome activity regulated by charged gold nanoclusters: Implications for neurodegenerative diseases. *Nano Today*. 2020;35:100933. doi:10.1016/j.nantod.2020.100933
- 96. Gao G, Chen R, He M, Li J, Wang L, Sun T. Gold nanoclusters for Parkinson's disease treatment. *Biomaterials*. 2019;194:36-46. doi:10.1016/j.biomaterials.2018.12.013
- 97. Nair L V., Nair R V., Shenoy SJ, Thekkuveettil A, Jayasree RS. Blood brain barrier permeable gold nanocluster for targeted brain imaging and therapy: An: in vitro and in vivo study. *J Mater Chem B*. 2017;5(42):8314-8321. doi:10.1039/c7tb02247f
- 98. Wang W, Han Y, Fan Y, Wang Y. Effects of Gold Nanospheres and Nanocubes on Amyloid-β Peptide Fibrillation. *Langmuir*. 2019;35(6):2334-2342.

- doi:10.1021/acs.langmuir.8b04006
- 99. Barthakur M, Kalita P, Mondal S. Modulation of astrocytic membrane potential using citrate stabilized gold nanoparticle to control brain hyper-excitability. *AIP Conf Proc*. 2020;2259(September). doi:10.1063/5.0015941
- 100. González-Reyes RE, Nava-Mesa MO, Vargas-Sánchez K, Ariza-Salamanca D, Mora-Muñoz L. Involvement of astrocytes in Alzheimer's disease from a neuroinflammatory and oxidative stress perspective. *Front Mol Neurosci*. 2017;10(December):1-20. doi:10.3389/fnmol.2017.00427
- 101. Talebpour F, Ghahghaei A. Effect of Green Synthesis of Gold Nanoparticles (AuNPs) from Hibiscus sabdariffa on the Aggregation of α Lactalbumin. *Int J Pept Res Ther*. 2020;26(4):2297-2306. doi:10.1007/s10989-020-10023-9
- 102. Xue J, Liu T, Liu Y, et al. Neuroprotective effect of biosynthesised gold nanoparticles synthesised from root extract of Paeonia moutan against Parkinson disease In vitro & In vivo model. *J Photochem Photobiol B Biol*. 2019;200:111635. doi:10.1016/j.jphotobiol.2019.111635
- 103. Cicek, B., Hacimuftuoglu, A., Yeni, Y., Kuzucu, M., Genc, S., Cetin, A., Yavuz, E., Danisman, B., Levent, A., Ozdokur, K.V. and Kantarcı, M.Cicek, B., Hacimuftuoglu, A., Yeni, Y., Kuzucu, M., Genc, S., Cetin, A., Yavuz, E., Danisman, B., Levent, A., Ozdok M. AuNPs with Cynara scolymus leaf extracts rescue arsenic-induced neurobehavioral deficits and hippocampal tissue toxicity in Balb/c mice through D1R and D2R activation. *Environ Toxicol Pharmacol*. 2024;107:104417.
- 104. Ling L, Jiang Y, Liu Y, et al. Role of gold nanoparticle from Cinnamomum verum against 1-methyl-4-phenyl-1, 2, 3, 6-tetrahydropyridine (MPTP) induced mice model. *J Photochem Photobiol B Biol*. 2019;201:111657. doi:10.1016/j.jphotobiol.2019.111657
- 105. Hu K, Chen X, Chen W, et al. Neuroprotective effect of gold nanoparticles composites in Parkinson's disease model. *Nanomedicine Nanotechnology, Biol Med.* 2018;14(4):1123-1136. doi:10.1016/j.nano.2018.01.020
- Daniel Gonzalez-Carter, Zhan Yuin Ongc, Catriona M. McGilverya IED, David T. Dexterb AEP. L-DOPA functionalized, multi-branched gold nanoparticles as braintargeted nano-vehicles. *nanotechnology*, *Biol Med*. 2019;1(15):1-11.
- 107. Frkanec L, Frkanec R, Štimac A, et al. Adamantane in Drug Delivery Systems and Surface Recognition. *molecules*. 2017;16(22):297. doi:10.3390/molecules22020297
- 108. Tomašić, Jelka and IH. Peptidoglycan monomer originating from Brevibacterium divaricatum—its metabolism and biological activities in the host. In: *Surface Structures of Microorganisms and Their Interaction with the Mammalian Host.*; 1987:113-121.
- 109. Xu J, Tai CH, Chen TY, Hsu S hui. An anti-inflammatory electroconductive hydrogel with self-healing property for the treatment of Parkinson's disease. *Chem Eng J*. 2022;446(1). doi:10.1016/j.cej.2022.137180
- 110. Xu J, Chen TY, Tai CH, Hsu S hui. Bioactive self-healing hydrogel based on tannic acid modified gold nano-crosslinker as an injectable brain implant for treating Parkinson's disease. *Biomater Res.* 2023;27(1):1-24. doi:10.1186/s40824-023-00347-0
- 111. Li S, Liu J, Li G, et al. Near-infrared light-responsive, pramipexole-loaded biodegradable PLGA microspheres for therapeutic use in Parkinson's disease. *Eur J Pharm Biopharm*. 2019;141(May):1-11. doi:10.1016/j.ejpb.2019.05.013
- 112. Liu L, Li M, Xu M, et al. Actively targeted gold nanoparticle composites improve behavior and cognitive impairment in Parkinson's disease mice. *Mater Sci Eng C*.

- 2020;114(January):111028. doi:10.1016/j.msec.2020.111028
- 113. Liu L, Li Y, Liu R, et al. Switchable nanoparticle for programmed gene-chem delivery with enhanced neuronal recovery and CT imaging for neurodegenerative disease treatment. *Mater Horizons*. 2019;6(9):1923-1929. doi:10.1039/c9mh00482c
- 114. Trapani M, Scala A, Mineo PG, et al. Thiolated amphiphilic b -cyclodextrin-decorated gold colloids: Synthesis, supramolecular nanoassemblies and controlled release of dopamine. *J Mol Liq.* 2021;336:116880. doi:10.1016/j.molliq.2021.116880
- 115. Khongkow M, Yata T, Boonrungsiman S. Surface modification of gold nanoparticles with neuron-targeted exosome for enhanced blood brain barrier penetration. 2019;(January):1-9. doi:10.1038/s41598-019-44569-6
- 116. Ong ZY, Chen S, Nabavi E, et al. Multi-Branched Gold Nanoparticles with Intrinsic LAT-1 Targeting Capabilities for Selective Photothermal Therapy of Breast Cancer. *ACS Appl Mater Interfaces*. 2017;15(9):39259-39270. doi:10.1021/acsami.7b14851
- 117. Khan AM, Korzeniowska B, Gorshkov V, et al. Silver nanoparticle-induced expression of proteins related to oxidative stress and neurodegeneration in an in vitro human blood-brain barrier model. *Nanotoxicology*. 2019;13(2):221-239. doi:10.1080/17435390.2018.1540728
- 118. Zande M Van Der, Vandebriel RJ, Doren E Van, et al. Distribution, Elimination, and Toxicity of Silver Nanoparticles and Silver Ions in Rats after 28-Day Oral Exposure. *ACS Nano*. 2012;6(8):7427-7442.
- 119. Skalska J, Strużyńska L. Toxic effects of silver nanoparticles in mammals does a risk of neurotoxicity exist? *folia Neuropathol*. 2015;53(4):281-300. doi:10.5114/fn.2015.56543
- 120. Huang CL, Hsiao IL, Lin HC, Wang CF, Huang YJ, Chuang CY. Silver nanoparticles affect on gene expression of inflammatory and neurodegenerative responses in mouse brain neural cells. *Environ Res.* 2015;136:253-263. doi:10.1016/j.envres.2014.11.006
- 121. Parveen M, Kumar A, Khan MS, et al. Comparative study of biogenically synthesized silver and gold nanoparticles of Acacia auriculiformis leaves and their efficacy against Alzheimer's and Parkinson's disease. *Int J Biol Macromol*. 2022;203(September 2021):292-301. doi:10.1016/j.ijbiomac.2022.01.116
- 122. Gonzalez-carter DA, Leo BF, Ruenraroengsak P, et al. Silver nanoparticles reduce brain inflammation and related neurotoxicity through induction of H 2 S-synthesizing enzymes. *Sci Total Environ*. 2017;7(1):42871. doi:10.1038/srep42871
- 123. Shivanna DK. Datura Stramonium leaves extract Silver Nanoparticles regulates PINK1 gene in Parkinson's disease model of Drosophila melanogaster. *Res Sq.* 2022;(Dm):1-16.
- 124. R E Sardjono, F Khoerunnisa, I Musthopa NSMMA and RR. Synthesize, characterization, and anti-Parkinson activity of silver-Indonesian velvet beans (Mucuna pruriens) seed extract nanoparticles (AgMPn). *J Phys Conf Ser*. 2018;1013(1):012195.
- 125. Silva CS, Tonelli FMP, Delgado VMS, et al. Nanoremediation and Antioxidant Potential of Biogenic Silver Nanoparticles Synthesized Using Leucena's Leaves, Stem, and Fruits. *Int J Mol Sci.* 2024;25(7):3993. doi:10.3390/ijms25073993
- 126. Mohamed Salam S, K.N., PV, R., NA, C.J. and MA A. Effect of Tualang Honey-Mediated Silver Nanoparticles on TNF-a level, Caspase-3 Activity and Hippocampal Morphology in Kainic Acid-Induced Neurodegeneration in Male Rats. *IIUM Med J Malaysia*. 2024;23(4).
- 127. Devi, N.A., Ravikumar, P., Devendran, P., Mohan, R., Ravichandran, K., Veeralakshmi, M., Yuvaloshini, J. and Sundari MM. Efficacy of Catharanthus roseus leaf and flower

- extracts mediated Ag incorporated ZnO nanoparticles for enhanced antimicrobial and antioxidant abilities: a comparative analysis. *Appl Phys A*. 2025;131(2):97.
- 128. Kaliappan, K., Nagarajan, P., Jayabalan, J., Pushparaj, H., Elumalai, S., Paramanathan, B., Manickam, V., Jang, H.T. and Mani, G., Kaliappan, K., Nagarajan, P., Jayabalan, J., Pushparaj, H., Elumalai, S., Paramanathan, B., Manickam, V., Jang, H.T. and Mani G. Systematic antimicrobial, biofilm, free radical inhibition and tyrosinase inhibition assessments of efficient green silver nanoparticles from the aqueous root extract of Cyphostemma adenocaule (CA). *RSC Pharm*. Published online 2025.
- 129. Arulkumar S, Sabesan M. Rapid preparation process of antiparkinsonian drug Mucuna pruriens silver nanoparticle by bioreduction and their characterization. *Pharmacognosy Res.* 2010;2(4):233-236. doi:10.4103/0974-8490.69112
- 130. Vissers C, Ming G li, Song H. Nanoparticle technology and stem cell therapy team up against neurodegenerative disorders. *Adv Drug Deliv Rev.* 2019;1(148):239-251. doi:10.1016/j.addr.2019.02.007
- 131. Niu S, Zhang L kun, Zhang L, et al. Inhibition by Multifunctional Magnetic Nanoparticles Loaded with Alpha-Synuclein RNAi Plasmid in a Parkinson's Disease Model. *Theranostics*. 2017;7(2):344. doi:10.7150/thno.16562
- 132. Cheng, G., Liu, Z., Yan, Z., Wu, J., Li, Z., Gao, S., Zheng, C., Guo, S., Pan, Y., Chen, X. and Lin G. Minocycline nanoplatform penetrates the BBB and enables the targeted treatment of Parkinson's disease with cognitive impairment. *J Control Release*. 2025;377:591-605.
- 133. Harris RA. Simulation study on the physicochemical properties of Fe 3 O 4 nanoparticles as drug delivery vehicles for dopamine replacement therapy of Parkinson's disease. *Mater Today Commun.* 2022;31(March):103829. doi:10.1016/j.mtcomm.2022.103829
- 134. Khadrawy YA, Hosny EN, Mohamed HSE. Assessment of the neuroprotective effect of green synthesized iron oxide nanoparticles capped with curcumin against a rat model of Parkinson's disease. *Iran J Basic Med Sci.* 2024;27(1):81-89. doi:10.22038/IJBMS.2023.73124.15892
- 135. Cheng G, Liu X, Liu Y, et al. Ultrasmall Coordination Polymers for Alleviating ROS-Mediated Inflammatory and Realizing Neuroprotection against Parkinson's Disease. *research*. 2022;jul 18.
- 136. Moayeri A, Darvishi M, Amraei M. Homing of Super Paramagnetic Iron Oxide Stem Cells by Magnetic Attraction in a Rat Model of Parkinson's Disease Homing of Super Paramagnetic Iron Oxide Nanoparticles (SPIONs) Labeled Adipose-Derived Stem Cells by Magnetic Attraction in a Rat Model of. *Int J Nanomedicine*. Published online 2020:1297-1308. doi:10.2147/IJN.S238266
- 137. Tsai-Hua Chung, Szu-Chun Hsu, Shu-Hui Wu, Jong-Kai Hsiao, Chih-Peng Lin MY and, Huang DM. Dextran-coated iron oxide nanoparticle-improved therapeutic effects of human mesenchymal stem cells in a mouse model of Parkinson's disease. *Nanoscale*. 2018;10(6):2998-3007. doi:10.1039/C7NR06976F
- 138. Dong Hana, Baolin Zhanga, Lichao Sua BY. Attachment of streptavidin-modified superparamagnetic iron oxide nanoparticles to the PC-12 cell membrane. *Biomed Mater*. 2020;12(15):045014.
- 139. Prado-prone G, Padilla P, Garc JA, Gutie G. Dopamine Released from TiO 2 Semicrystalline Lattice Implants Attenuates Motor Symptoms in Rats Treated with 6 Hydroxydopamine. *ACS Omega*. 2019;4:7953-7962. doi:10.1021/acsomega.8b00626

- 140. M. Velázquez-Paniaguaa, A.M. Vázquez-Álvarezb, G. Valverde-Aguilarc, P. Vergara-Aragóna. Current treatments in Parkinson's including the proposal of an innovative dopamine microimplant. Rev Médica del Hosp Gen México. 2016;79(2):79-87. doi:10.1016/j.hgmx.2015.10.006
- 141. Punitha VN, Vijayakumar S, Sakthivel B, Praseetha PK. Protection of neuronal cell lines, antimicrobial and photocatalytic behaviours of eco-friendly TiO 2 nanoparticles. *J Environ Chem Eng.* 2020;8(5):104343. doi:10.1016/j.jece.2020.104343
- 142. Balaraju PC, Manchegowda JB, Kumar C, Ahmed SS. Exploring the Therapeutic Potential of Selenium Nanoparticles in Central Nervous System Disorders: A Nanomedicine Approach. *Int J Pharm Investig.* 2024;14(4):1025-1034. doi:10.5530/ijpi.14.4.112
- 143. Shayganfard M. Are Essential Trace Elements Effective in Modulation of Mental Disorders? Update and Perspectives. *Biol Trace Elem Res.* 2022;200(3):1032-1059. doi:10.1007/s12011-021-02733-y
- 144. El-Ghazaly, M.A., Fadel, N., Rashed, E., El-Batal, A. and Kenawy S. Anti-inflammatory effect of selenium nanoparticles on the inflammation induced in irradiated rats. *Can J Physiol Pharmacol*. 2017;95(2):101-110.
- 145. El-Halaby, L.O., El-Magd, N.F.A., Almehmadi, S.J., El-Sayed, A.A., Khattab, R.R., El-Kalyoubi, S. and Elfeky SM. Synthesis, In-Vitro, In-Vivo screening, and molecular docking of disubstituted aminothiazole derivatives and their selenium nanoparticles as potential antiparkinson agents. *J Mol Struct*. 2024;1315:138951.
- 146. Yifei Gao, Yuxue Cheng, Jiapeng Chen, Danmin Lin, Chao Liu, Ling-Kun Zhang LY, Runcai Yang and YQG. NIR-Assisted MgO-Based Polydopamine Nanoparticles for Targeted Treatment of Parkinson's Disease through the Blood–Brain Barrier. *Adv Heal care Mater*. 2022;11:2201655.
- 147. Küçükdo R, Türkez H, Arslan ME, Tozlu ÖÖ, Sönmez E. Neuroprotective effects of boron nitride nanoparticles in the experimental Parkinson's disease model against MPP + induced apoptosis. *Metab Brain Dis.* 2020;35:947-957.
- 148. Colon J, Hsieh N, Ferguson A, et al. Cerium oxide nanoparticles protect gastrointestinal epithelium from radiation-induced damage by reduction of reactive oxygen species and upregulation of superoxide dismutase 2. *Nanomedicine Nanotechnology, Biol Med.* 2010;6(5):698-705. doi:10.1016/j.nano.2010.01.010
- 149. Eric Heckert, Ajay Karakoti, Sudipta Seal and WTS. The role of cerium redox state in the SOD mimetic activity of nanoceria. *Biomaterials*. 2008;29(18):2705-2709.
- 150. Pirmohamed T, Dowding JM, Singh S, et al. Nanoceria exhibit redox state-dependent catalase mimetic activity. *Chem Commun.* 2010;46(16):2736-2738. doi:10.1039/b922024k
- 151. Ruotolo R, De Giorgio G, Minato I, Bianchi MG, Bussolati O, Marmiroli N. Cerium oxide nanoparticles rescue α-synuclein-induced toxicity in a yeast model of parkinson's disease. *Nanomaterials*. 2020;10(2):235. doi:10.3390/nano10020235
- 152. Tameh, F.A., Jahani, Z., Sedghiniya, S., Aghtaei, M.A., Abtahi, M., Xiang, W., Akbari, M., Soleimannejad, J. and Janczak J. Morphology-dependent multienzyme activity of nanoceria in antioxidant protection of MnCl2-treated PC-12 Cells, and the potential application for Parkinson's disease treatment. *Inorg Chem Commun*. 2024;169:113117.
- 153. Dieter HH, Bayer TA and GM. Environmental Copper and Manganese in the Pathophysiology of Neurologic Diseases (Alzheimer's Disease and Manganism)*. *acta Hydrochim Hydrobiol*. 2005;33(1):72-78. doi:10.1002/aheh.200400556

- 154. Sharma, A., Feng, L., Muresanu, D.F., Sahib, S., Tian, Z.R., Lafuente, J.V., Buzoianu, A.D., Nozari, A., Wiklund, L. and Sharma HS. Manganese nanoparticles induce blood-brain barrier disruption, cerebral blood flow reduction, edema formation and brain pathology associated with cognitive and motor dysfunctions. In: *Nanomedicine and Neuroprotection in Brain Diseases*. Elsevier; 2021:385-406.
- 155. Adhikari A, Das M, Mondal S, et al. Manganese neurotoxicity: nano-oxide compensates for ion damage in mammals. *Biomater Sci.* 2019;7:4491-4502. doi:10.1039/c9bm01039d
- 156. Wang X, Zhao J, Wang W, et al. Electromagnetic field-enhanced chiral dimanganese trioxide nanoparticles mitigate Parkinson's disease. *Sci China Chem.* 2022;65(10):1911-1920.
- 157. Zhuojia Xu, Aihua Qu, Weiwei Wang, Meiru Lu, Baimei Shi, Chen Chen CH, Liguang Xu, Maozhong Sun,* Chuanlai Xu and HK. Facet-Dependent Biodegradable Mn3O4 Nanoparticles for Ameliorating Parkinson's Disease. *Adv Heal care Mater*. 2021;10(3):2101316.
- 158. Liu Y qing, Mao Y, Xu E, Jia H, Zhang S. Nanozyme scavenging ROS for prevention of pathologic __-synuclein transmission in Parkinson 's disease. *Nano Today*. 2021;36:101027. doi:10.1016/j.nantod.2020.101027
- 159. Xu H, Ding X, Li L, Li Q, Li Z. Tri-element nanozyme PtCuSe as an ingenious cascade catalytic machine for the amelioration of Parkinson's disease-like symptoms. *Front Bioeng Biotechnol.* 2023;30(11):1208693. doi:10.3389/fbioe.2023.1208693
- 160. Shi B, Qu A, Wang W, et al. Chiral Cu x Co y S Supraparticles Ameliorate Parkinson's Disease. *CCS Chem.* 2022;14(4):2440-2451. doi:10.31635/ccschem.021.202101107
- 161. Cheng Y, Chang Y, Feng Y, Jian H, Tang Z, Zhang H. Deep-Level Defect Enhanced Photothermal Performance of Bismuth Sulfide–Gold Heterojunction Nanorods for Photothermal Therapy of Cancer Guided by Computed Tomography Imaging. *Angew Chemie Int Ed.* 2018;57(1):246-251. doi:10.1002/anie.201710399
- 162. Xiao, L., Zhu, A., Xu, Q., Chen, Y., Xu, J. and Weng J. Colorimetric biosensor for detection of cancer biomarker by Au nanoparticle-decorated Bi2Se3 nanosheets. *ACS Appl Mater Interfaces*. 2017;9(8):6931-6940.
- 163. Ansari SR, Mahajan J, Teleki A. Iron oxide nanoparticles for treatment and diagnosis of chronic inflammatory diseases: A systematic review. *WIREs Nanomedicine and Nanobiotechnology*. 2024;16(3):e1963. doi:https://doi.org/10.1002/wnan.1963
- 164. Calle D, Negri V, Ballesteros P, Cerdán S. Magnetoliposomes Loaded with Poly-Unsaturated Fatty Acids as Novel Theranostic Anti-Inflammatory Formulations. *Theranostics*. 2015;5(5):489-503. doi:10.7150/thno.10069
- 165. Wang M, Li L, Zhang X, et al. Magnetic Resveratrol Liposomes as a New Theranostic Platform for Magnetic Resonance Imaging Guided Parkinson's Disease Targeting Therapy. *ACS Sustain Chem Eng.* 2018;6(12):17124-17133. doi:10.1021/acssuschemeng.8b04507
- 166. Niu S, Zhang LK, Zhang L, et al. Inhibition by Multifunctional Magnetic Nanoparticles Loaded with Alpha-Synuclein RNAi Plasmid in a Parkinson's Disease Model. *Theranostics*. 2017;7(2):344-356. doi:10.7150/thno.16562
- 167. García-Pardo J, Novio F, Nador F, et al. Bioinspired Theranostic Coordination Polymer Nanoparticles for Intranasal Dopamine Replacement in Parkinson's Disease. *ACS Nano*. 2021;15(5):8592-8609. doi:10.1021/acsnano.1c00453
- 168. Zhou Y, Peng Z, Seven ES, Leblanc RM. Crossing the blood-brain barrier with

- nanoparticles. J Control Release. 2018;270:290-303. doi:10.1016/j.jconrel.2017.12.015
- Wei X, Cai M, Jin L. The Function of the Metals in Regulating Epigenetics During Parkinson's Disease. Front Genet. 2021;11:616083. doi:10.3389/fgene.2020.616083
- Pyatha S, Kim H, Lee D, Kim K. Association between Heavy Metal Exposure and 170. Parkinson's Disease: A Review of the Mechanisms Related to Oxidative Stress. Antioxidants. 2022;11(12):2467. doi:10.3390/antiox11122467
- Xiaoli F, Junrong W, Xuan L, Zhang Yanli, Wei Limin LJS and, Longquan. Prenatal 171. exposure to nanosized zinc oxide in rats: neurotoxicity and postnatal impaired learning and memory ability. Nanomedicine. 2017;12(7):777-795.
- 172. Oliver T. Phillipson. Alpha-synuclein, epigenetics, mitochondria, metabolism, calcium traffic, & circadian dysfunction in Parkinson's disease. An integrated strategy for management. Ageing Res Rev. 2017;1(40):149-167. doi:10.1016/j.arr.2017.09.006
- David N Hauser and Teresa G Hastings. Mitochondrial dysfunction and oxidative stress in Parkinson's disease and monogenic parkinsonism. *Neurobiol Dis.* 2013;51:35-42. doi:10.1016/j.nbd.2012.10.011.Mitochondrial
- 174. Mohammadipour A, Haghir H, Bideskan AE. A link between nanoparticles and Parkinson's disease. Which nanoparticles are most harmful? Rev Environ Health. 2020;35(4):545-556. doi:10.1515/reveh-2020-0043
- 175. Valentini X, Deneufbourg P, Paci P, et al. Morphological alterations induced by the exposure to TiO 2 nanoparticles in primary cortical neuron cultures and in the brain of rats. Toxicol Reports. 2018;5:878-889. doi:10.1016/j.toxrep.2018.08.006
- Hu O, Guo F, Zhao F, Fu Z. Effects of titanium dioxide nanoparticles exposure on Parkinsonism in zebrafish larvae and PC12. *Chmeosphere*. 2017;173:373-379. doi:10.1016/j.chemosphere.2017.01.063
- Hu R, Gong X, Duan Y, et al. Neurotoxicological effects and the impairment of spatial 177. recognition memory in mice caused by exposure to TiO 2 nanoparticles. *Biomaterials*. 2010;31:8043-8050. doi:10.1016/j.biomaterials.2010.07.011
- 178. Grissa I, Guezguez S, Ezzi L, Chakroun S. The effect of titanium dioxide nanoparticles on neuroinflammation response in rat brain. Environ Sci Pollut Res. 2016;23:20205-20213. doi:10.1007/s11356-016-7234-8
- 179. Wang J, Li N, Zheng L, Wang S, Wang Y. P38-Nrf-2 Signaling Pathway of Oxidative Stress in Mice Caused by Nanoparticulate TiO 2. Biol Trace Elem Res. 2011;140:186-197. doi:10.1007/s12011-010-8687-0
- 180. Cristina L, Pazin M, Franco-bernardes MF, et al. A perspective of mitochondrial dysfunction in rats treated with silver and titanium nanoparticles (AgNPs and TiNPs). JTrace Elem Med Biol. 2018;47:63-69. doi:10.1016/j.jtemb.2018.01.007
- 181. Journal AI, Wu J, Xie H, Wu J, Xie H. Effects of titanium dioxide nanoparticles on α synuclein aggregation and the ubiquitin- proteasome system in dopaminergic neurons Effects of titanium dioxide nanoparticles on a -synuclein aggregation and the ubiquitinproteasome system in dopaminergic ne. artifical cells, nanomedicine, Biotechnol. 2016;44:690-694. doi:10.3109/21691401.2014.980507
- 182. Mohammadi S, Nikkhah M. TiO2 Nanoparticles as Potential Promoting Agents of Fibrillation of α-Synuclein, a Parkinson's Disease-Related Protein. Natl Inst Genet Eng Biotechnol. 2017;15(2):87-94. doi:10.15171/ijb.1519
- 183. Akintunde JK, Farai TI, Arogundade MR, Adeleke JT. Biogenic zinc-oxide nanoparticles of Moringa oleifera leaves abrogates rotenone induced neuroendocrine toxicity by

- regulation of oxidative stress and acetylcholinesterase activity. *Biochem Biophys Reports*. 2021;26(April):100999. doi:10.1016/j.bbrep.2021.100999
- 184. Jin M, Li N, Sheng W, Ji X, Liang X, Kong B. Toxicity of different zinc oxide nanomaterials and dose-dependent onset and development of Parkinson's disease-like symptoms induced by zinc oxide nanorods. *Environ Int.* 2021;146:106179. doi:10.1016/j.envint.2020.106179
- 185. Fernández-Bertólez N, Alba-González A, Touzani A, et al. Toxicity of zinc oxide nanoparticles: Cellular and behavioural effects. *Chemosphere*. 2024;363:142993. doi:10.1016/j.chemosphere.2024.142993
- 186. Ayton S, Lei P. Nigral Iron Elevation Is an Invariable Feature of Parkinson's Disease and Is a Sufficient Cause of Neurodegeneration. *biomed reseach*. 2014;2014.
- 187. Hare DJ, Double KL. Iron and dopamine: a toxic couple. *brain*. 2016;1(139):1026-1035. doi:10.1093/brain/aww022
- 188. Yarjanli Z, Ghaedi K, Esmaeili A, Rahgozar S, Zarrabi A. Iron oxide nanoparticles may damage to the neural tissue through iron accumulation, oxidative stress, and protein aggregation. *BMC Neurosci*. 2017;18:1-12. doi:10.1186/s12868-017-0369-9
- 189. Imam SZ, Lantz-mcpeak SM, Cuevas E, et al. Iron Oxide Nanoparticles Induce Dopaminergic Damage: In vitro Pathways and In Vivo Imaging Reveals Mechanism of Neuronal Damage. Published online 2015:913-926. doi:10.1007/s12035-015-9259-2
- 190. Bagheri-Abassi F, Alavi H, Mohammadipour A, Motejaded F, Ebrahimzadeh-Bideskan A. The effect of silver nanoparticles on apoptosis and dark neuron production in rat hippocampus. *Iran J Basic Med Sci.* 2015;18(7):644-648.
- 191. Gupta G, Gliga A, Hedberg J, Odnevall I, Bengt W. Cobalt nanoparticles trigger ferroptosis-like cell death (oxytosis) in neuronal cells: Potential implications for neurodegenerative disease. *Fed Am Soc Exp Biol.* 2020;(August 2019):5262-5281. doi:10.1096/fj.201902191RR
- 192. Lyu J, Long X, Xie T, et al. Copper oxide nanoparticles promote α -synuclein oligomerization and underlying neurotoxicity as a model of Parkinson's disease. *J Mol Liq.* 2021;323(323):115051. doi:10.1016/j.molliq.2020.115051
- 193. Botha NL, Cloete KJ, Šmit Ž, et al. Ionome mapping and amino acid metabolome profiling of Phaseolus vulgaris L. seeds imbibed with computationally informed phytoengineered copper sulphide nanoparticles. *Discov Nano*. 2024;19(1):8. doi:10.1186/s11671-023-03953-y
- 194. Stern M, Botha N, Cloete KJ, Maaza M, Tan S, Bicker G. Neurotoxicity and Developmental Neurotoxicity of Copper Sulfide Nanoparticles on a Human Neuronal In-Vitro Test System. *Int J Mol Sci.* 2024;25(11):5650. doi:10.3390/ijms25115650
- 195. Li X, Li Q, Zhang Y, Bai Y, Cao Y, Yang Y. Nickel oxide nanoparticles increase a synuclein amyloid formation and relevant overexpression of inflammatory mediators in microglia as a marker of Parkinson's disease. *Arab J Chem.* 2021;14(10):103380. doi:10.1016/j.arabjc.2021.103380
- 196. Gao, C., Lyu, F. and Yin Y. Encapsulated metal nanoparticles for catalysis. *Chem Rev.* 2020;121(2):834-881.
- 197. Boltman T, Sibuyi NRS, Ekpo O, Meyer M. Synthesis of chlorotoxin functionalized metallic nanoparticles and their in vitro evaluation of cytotoxic effects in nervous system cancer cell lines. *Nano Express*. 2024;5(4):045002. doi:10.1088/2632-959X/ad80b0
- 198. Mhaske A, Shukla S, Ahirwar K, Singh KK, Shukla R. Receptor-Assisted

- Nanotherapeutics for Overcoming the Blood–Brain Barrier. *Mol Neurobiol*. 2024;61(11):8702-8738. doi:10.1007/s12035-024-04015-9
- 199. Lang AE. C O M M E N TA RY Clinical trials of disease-modifying therapies for neurodegenerative diseases: the challenges and the future. *Nat Publ Gr*. 2010;16(11):1223-1226. doi:10.1038/nm.2220
- 200. Dawson TM, Golde TE, Lagier-tourenne C. Animal models of neurodegenerative diseases. *Nat Neurosci.* 2018;21(10):1370-1379. doi:10.1038/s41593-018-0236-8

Data Availability Statement:

This review article does not contain original data. All data referenced in this article are from published studies and sources cited in the reference list. Additional information can be obtained from these sources as noted.