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STEM-in-SEM high resolution imaging of gold nanoparticles and bivalve tissues in bioaccumulation experiments

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The methodology termed scanning transmission electron microscopy in scanning electron microscopy (STEM-in-SEM) has been used in this work to study the uptake of citrate stabilized gold nanoparticles (AuNPs), average particle sizes of 23.5±4.0 nm) into tissue samples upon *in vitro* exposure of the dissected gills of the Ruditapes philippinarum marine bivalve to the nanoparticle suspensions. The STEM-in-SEM methodology has been optimized for achieving optimum resolution at SEM low voltage operating conditions (20-30 kV). Based on scanning microscope assessments and resolution testing (SMART), resolutions well below 10 nm were properly achieved working at magnifications over 100 kx, with experimental sample thickness between 300 and 200 nm. These relative thick slices appear stable under the beam and help avoid NPs displacement during cutting. We show herein that both localizing of the internalized nanoparticles and imaging of ultrastructural disturbances in gill tissues are strongly accessible thanks to the improved resolution, even at sample thicknesses higher than those normally employed in standard TEM techniques at higher voltages. Ultrastructural imaging of Bio-Nano features in bioaccumulation experiments have been demonstrated in this study

Introduction

The characterization of *in vitro* nanoparticle's uptake and localization is an issue of increasing attention[1.] Transmission electron microscopy (TEM) is currently used to provide detailed information regarding nanoparticles uptake and localization by allowing both visualization of nanoparticles location within target cells and/or tissues and, in conjunction with spectroscopic methods, characterization of the composition of internalized nanoparticles[1, 2]. However, characterization of soft materials by TEM has been limited due to their susceptibility to high-voltage electron beams[3]. A hybrid characterization technique such as STEM-in-SEM may be a convenient characterization approach for these materials offering advantages such as lower accelerating voltages, larger field of view, and exclusion of a post-specimen projection lens[3, 4]. A STEM system

added to standard SEM is often designated as STEM-in-SEM or "low voltage STEM" (term referring to the 20-30 kV regime, i.e. low relative to typical TEM operating energies)[5]. The STEM-in-SEM technique is based on two principles. First, as in SEM, the beam focuses on a small spot that scans over the sample and imaging is performed by mapping some signal intensity synchronously with the scan[5]. Second, as in TEM, image information is extracted from electrons that have passed through a thin sample[5].

Bivalve molluscs are recognized pollution indicators in ecological monitoring programs[6-8]. Among several bivalves, *Ruditapes philippinarum* is a model frequently used for investigating the effects and mechanisms of action underlying the potential toxicity of NPs in marine invertebrates[9, 10]. Recently, an *in vitro* approach based on haemocytes of the clam *Ruditapes philippinarum* was used to investigate effects of TiO_2 nanoparticles on phagocytic activity[10] while efforts to establish primary cell cultures from the mantle of the clam *P. malabarica* were also performed[11]. Regarding the toxicity exerted by nanoparticles in a variety of taxa, details were also described[12] of how nanoparticles may interact with organisms through different ways such as adsorption to the surface (cell, organ or body), cellular internalization, or dissolution of ions from the NPs. The STEM-in-SEM technique was applied by the authors in a previous paper after *in vivo* exposure of *Ruditapes philippinarum* to high and low relevant concentrations of Au NPs and to ionic gold (Au^{3+}) ranging from ppm to ppb levels[13]. This technique successfully allowed the ultrastructural localization of AuNPs in target tissues and cells when delivered to artificial SW media.

In the present work, STEM-in-SEM has been carefully optimized and used to study the uptake and subcellular distribution of citrate stabilized gold nanoparticles in gill samples from *Ruditapes philippinarum* upon *in vitro* exposure of gill explants (*ex vivo*) to the nanoparticles. The development of *in vitro* models for assessing the toxicity of NPs, via direct exposure of isolated cells or with organ explants (*ex vivo*), allow experimentation on an organism cells or tissue under more controlled conditions than *in vivo* and can contribute to develop new models as test systems for NP toxicity in the future. Location studies were performed together with bioaccumulation analytical measurements in the same experiments at ecotoxicological relevant conditions, such as sea water media and low (sublethal) concentrations. We show how gold nanoparticles accumulate in the gills, as well as their subcellular location with nano-scale resolution. In addition, the paper presents the optimization of the STEM-in-SEM methodology for achieving optimum resolution at the SEM low voltage operating conditions.

Experimental section

AuNP suspensions: Preparation and characterization

AuNPs suspensions with a concentration of 60 $mg \cdot L^{-1}$ (as Au) were prepared following a procedure based on the synthesis method of Frens[14]. Specifically, 0.006 g of $HAuCl_4 \cdot 3H_2O$ (Aldrich, 99.9%) was dissolved in 50 mL of MilliQ® water in a 100 mL round bottomed flask and heated to boiling under reflux conditions. Then, 0.01 g of sodium citrate (Sigma-Aldrich, 99%) dissolved in 1 mL of MilliQ® water was added, and the solution was allowed to react until a red colour remained in the final suspension. From this starting solution, another sample was prepared by dilution to 750 $\mu g \cdot L^{-1}$ in standard sea water (SW) media.

For basic TEM characterization, colloidal solutions of 60 $mg L^{-1}$ of Au in milliQ® water and 750 $\mu g \cdot L^{-1}$ in SW were used, while for UV-Visible spectroscopy measurements, representative samples with 5 $mg \cdot L^{-1}$ of Au in either milliQ® or SW were tested to ensure sufficient signal and to allow a comparison. For *in vitro* testing, the as prepared 60 $mg \cdot L^{-1}$ in milliQ® water as well as the 750 $\mu g \cdot L^{-1}$ in SW solutions were used.

Primary particle size and morphology were characterized using a TEM microscope (Philips CM200) operating at 200 kV. For sample preparation, 5 µL of a particular AuNP suspension was pipetted onto a carbon-coated copper TEM grid and left to dry in air. A Perkin Elmer Lambda 12 spectrometer was used for UV-Vis spectra acquisition.

In vitro exposure experiments

Adult clams (*Ruditapes philippinarum*) of the same age with a shell length (maximum axis) between 4.4 and 3.2 cm were supplied by a commercial clam aquaculture facility (Amalthea S. L., Chiclana, Spain). The clams were dissected over ice and the gill explants were immersed into colloidal solutions of AuNPs at concentrations of 60 $mg.L^{-1}$ in milliQ® water and 750 $\mu g \cdot L^{-1}$ in SW. A control series was run in parallel by immersing gills in pure milliQ® and standard SW without AuNPs. After immersing the gill explants for 1 and 6 hours, they were removed from the solution and washed at least three times with milliQ® water or SW. Portions of gill explants were then processed for gold accumulation analysis and for STEM-in-SEM imaging and analysis as described below.

Analysis of gold accumulation in tissue samples

Gold accumulation was determined in the gill tissues after *in vitro* exposure to AuNPs suspensions of 750 $\mu g \cdot L^{-1}$ and after control exposures (both in seawater media). After washing as described above, the tissues were first washed with an AEDT solution 0.1 mM in SW and then with SW solution alone to remove Au traces that were adsorbed on the surface of the gills. The gills were then digested with 2 mL of "aqua regia" at 95ºC for 60 min employing a digestion system (DigiPrep MS, SCP Science, France). The digested samples were filled to 10 mL. All of the reagents were of Suprapur quality (Merck). The results are expressed as $\mu g \cdot g^{-1}$ wet weight.

The STEM-in-SEM methodology and the imaging of gold NPs in tissue samples

After the *in vitro* exposure experiment (described in section above), the gill tissues were cut into portions of 3-4 mm^2 and then fixed in cold 0.1 M sodium cacodylate trihydrate buffer solution (pH=7.4) containing 2.5% of glutaraldehyde for 2.5 h at 4 °C. Then, the samples were washed three times in a sodium trihydrate cacodylate buffer solution (pH=7.4) for 5 min each and stored at 4 °C.

Samples were post-fixed in 1% osmium tetroxide for 1 h at 4 °C and rinsed in a buffer solution (3 times) containing 7.5% sucrose for 20 min. The samples were then dehydrated at 4 °C in acetone (30, 0 and 70% acetone baths for 15 min each). Afterwards, the samples were rinsed in a 70% acetone bath containing 2% uranyl acetate for 4 h for mild pre-staining and dehydrated once in 90% acetone (30 min) and twice in 100% acetone (15 min each). Embedding in Spurr's resin was performed at the same temperature (4 °C) with acetone/resin mixtures of 3:1 for 1 h, 1:1 for the second hour, and 1:3 for the third hour, followed by embedding in pure resin for 12 h. Samples were polymerized at 70 °C for a period of 7 h. Lastly, semi-thin sections (range: 100-400 nm) were cut with an ultramicrotome (Leica EM UC7) using a diamond knife and placed on a carbon-coated copper TEM grid.

Studies by FE-SEM (field emission gun SEM microscope) were performed in a Hitachi S4800 microscope also coupled to an Energy Dispersive X-ray (EDX) detector (Bruker, XFlash 410) and equipped with a transmission mode detector. Details of these measurements will be thoroughly reported in the Results and Discussion section.

Results and discussion

AuNPs characterization

First, colloidal solutions of AuNPs in both milliQ® water and SW were characterized. Representative TEM images are shown in Figs. 1a-b. Nearly spherical particles with average diameters of 22.5±4.6 nm and 23.5±4.0 nm were obtained for samples containing 60 mgL^{-1} of Au in milliQ® water and 750 $\mu g \cdot L^{-1}$ of Au in SW. Although the Au concentration used in the milliQ® water was much higher than in SW, the agglomeration in both cases was comparatively similar because of the greater ionic strength of the SW media which favours agglomeration. Nevertheless the TEM study showed that for Au concentrations close to 750 $\mu g \cdot L^{-1}$ in SW, NPs assemblies can still dissociate easily[13] and only limited coalescence was observed (Fig 1b). Particles were therefore available for uptake during the tissue exposure experiments as shown in the next sections.

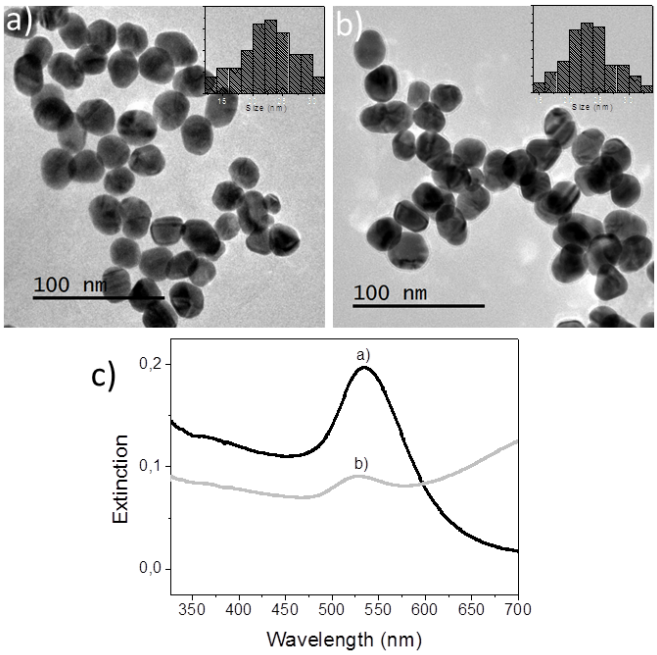

Fig. 1 TEM images of (a) 60 $mg \cdot L^{-1}$ AuNP suspension in milliQ® water and (b) 750 $\mu g \cdot L^{-1}$ AuNP suspension in standard SW. Insets in TEM images correspond to particle size distributions. c) Extinction spectra for 5 $mg \cdot L^{-1}$ AuNP suspensions, both in milliQ® water and standard SW.

The characterization by UV-VIS absorption spectroscopy could not be performed at very low concentrations due to a lack of signal. For this study, nanoparticles suspensions were prepared with mass-normalized concentrations of 5 $mg \cdot L^{-1}$ (as Au) in either milliQ® water or SW. As shown in Figure 1c, the signals due to the localized surface plasmon resonance (SPR) of Au were clearly observed. The SPR signal of Au in SW had an expected intensity loss that is attributable to higher inter-particle interactions[13] at these concentrations. A concentration of 60 ppm in milliQ® water,

although it is not ecotoxicologically relevant, was used in this work to optimize some of the parameters in the STEM-in-SEM methodology. As shown by TEM in this section, these suspensions showed limited coalescence and therefore were available for uptake during the tissue exposure experiments.

A more detailed analysis of the physico-chemical evolution of citrate-stabilized AuNPs in SW media is available in Ref. 13, where attempts to approach concentration values predicted[15, 16] in the environment were also performed.

The STEM-in-SEM methodology for imaging of gold NPs in tissue samples

The first main objective in this work was to explore the use of a SEM-FEG microscope operated in transmission mode at 20-30 kV for imaging AuNPs in tissue samples after our *in vitro* exposure experiments. The microscope configuration for STEM in a FE-SEM (see Fig. 2) includes a dedicated sample holder for conventional copper TEM grids and a dedicated bright field STEM detector (BF-STEM). This detector is located below the dedicated sample holder, which also includes a fixed STEM aperture. The upper detector, a through-the-lens (TTL) detector, permits to record the conventional secondary electrons SEM images. In addition, the EDX detector is also available for chemical analysis.

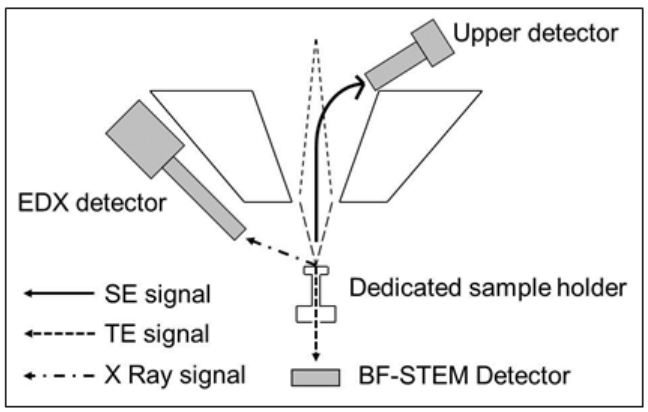

Fig. 2 Microscope configuration for STEM analysis in our FE-SEM Hitachi S4800 microscope. SE: secondary electrons; TE: transmitted electrons.

Despite the lower resolution compared to conventional TEM at typical voltages of 80-100 kV, the STEM-in-SEM offers contrast enhancement over TEM due to lower (20-30 kV) electron energy in the SEM[17]. The increased electron scattering cross-section enables better insight into the morphology of low Z (atomic number) materials, such as polymers or carbon nanotubes[17, 18]. Less damage in soft condensed matter is also expected. In addition, the use of a STEM detector in a standard SEM has the advantage of avoiding chromatic aberration. As there is no projection lens, no image deterioration occurs due to chromatic aberrations, even in the case of inelastic interactions at low voltages[5].

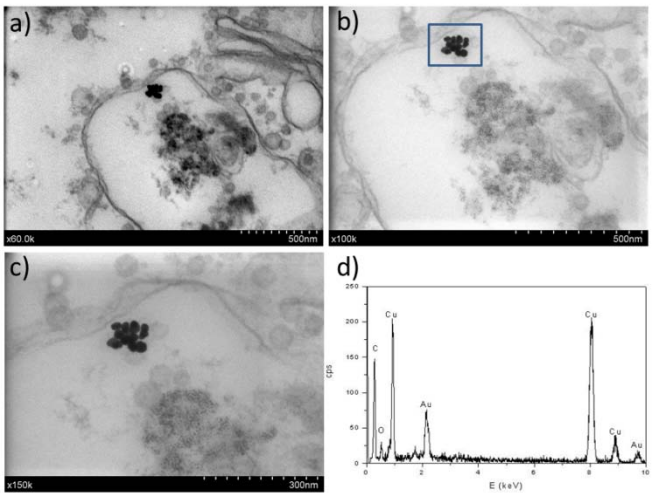

Fig. 3 STEM images (a to c) at different magnifications showing gills tissue after 1 hour exposure with a 60 mg·L^{-1} of AuNPs in milliQ water. d) Corresponding EDX spectra of the electron-dense contrasts region (marked area in panel b).

In the particular case of imaging gold NPs of ca. 23 nm in diameter in tissue bioaccumulation experiments, several specific features have been pursued with this methodology. Measurements were optimized with the goal of working with thicker slices (ca. 200-300 nm) to stabilize the samples under the electron beam (also improved by the use of lower voltage compared to TEM). The work with thick samples likely also avoids the NPs displacement during cutting and increases the possibility of finding NPs when working with low NPs doses (e.g., environmentally-relevant concentrations). In addition, the good contrast for low Z elements may allow for reducing (or even eliminating) heavy metal staining, which is of high interest when analysing the location of NPs. In all cases, EDX analysis were undertaken to confirm the chemical nature of the high contrast features.

The first results are presented in Fig. 3 for slices obtained after *in vitro* exposures for 1 hour into AuNPs colloidal solutions containing 60 mg·L^{-1} of gold in milliQ® water. The localization of high Z nanoparticles in low Z tissue matrices was clearly shown using the STEM-in-SEM coupled to EDX analysis (Fig. 3 a-d). As mentioned above, the exposure conditions were not relevant for ecotoxicological studies because the concentration of AuNPs was too high, and they were not supplied in sea water media. The experimental conditions were however chosen to ensure the presence

of AuNPs in the gill tissue with the purpose of using these slices to optimize some parameters of the methodological approach before going to more dilute solutions in the SW media. With actual FE-SEM microscopes, probe sizes fall into values smaller than 10 nm. To determine the actual resolution, a sputtered gold is commonly used, measuring the smallest gap between gold islands that are visible on a micrograph[19]. However, this method is not totally representative of all specimens of interest. In addition, Demers et al.[20, 21] studied the broadening of a scanning electron probe inside a micrometre thick film using Monte Carlo simulations. They found that the lateral resolution was noise limited for film thicknesses < 0.2 µm and by the probe broadening for film thicknesses >1 µm; the intermediate thicknesses represent a transition regime. Considering this background, we have estimated the resolution in our FE-SEM microscope directly from the images of tissue slices by Fast Fourier Transform (FFT) algorithms[19] on specific images. We have used the SMART macro running inside the "SCION Image" program under windows[19]. Using different set of images, such as those shown in Fig. 3, the microscope resolution for these particular samples has been determined under different measure conditions. Fig. 4 shows the magnification dependence of the estimated resolution. It is clearly seen that the resolution improves with the increase of magnification. A prominent feature of working at magnifications over 100 kx is that resolutions below 10 nm can be obtained for slices of 200-300 nm. Another interesting fact that can be seen from Fig. 4 is that operating voltages of either 20 or 30 kV are adequate to achieve resolutions below 10 nm.

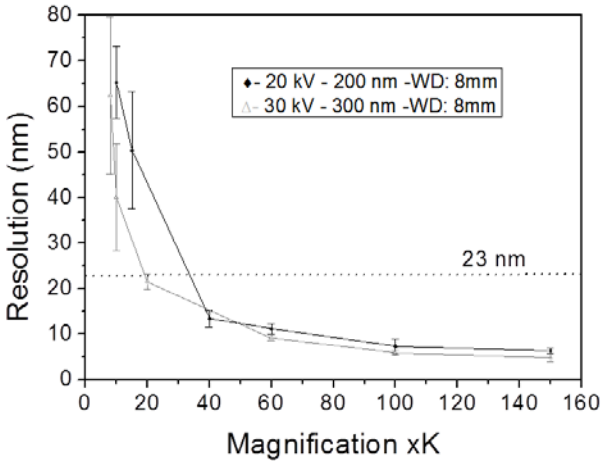

Fig. 4 Magnification dependence of the estimated resolution for two different series of images. Data given as mean ± standard deviation for n = 4 replicates. Measure conditions (acceleration voltage, sample thickness and working distance WD) are indicated in the symbols description within the graph.

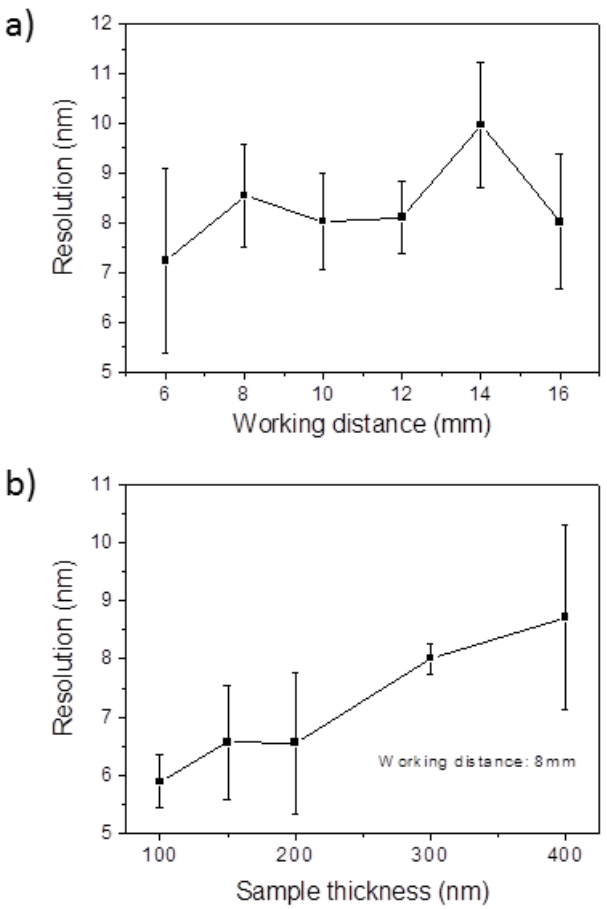

Fig. 5 a) Estimated resolution as a function of the working distance for a 300 nm slice; b) sample thickness dependence of the estimated resolution at a WD of 8 mm. Data given as mean ± standard deviation for n = 4 replicates. The magnification used was 100kx.

Further optimization of the observation conditions were then performed by taking into account the working distance dependence (Fig. 5a) for 300 nm slices, as well as the sample thickness dependence (Fig. 5b) for an optimum working distance of 8 mm at the reference magnification of 100 kx. In this optimized condition, the tissue slices used to acquire all images came from the *in vitro* experiment. As shown in Fig. 5a-b, the conditions for improved resolutions feasibly gave values below 10 nm, which is adequate for imaging relatively small objects, such as the AuNPs with average diameters of 23 nm.

The *in vitro* experiments: Gold accumulation in gill explants

In vitro exposure of gill explants into AuNPs colloidal solutions containing 750 µg.L^{-1} of gold in standard SW for 1 and 6 hours was the following step in our study. Assays of bivalve tissue exposures to AuNPs (750 µgL^{-1}) were carried out, together with control experiments. Analyses of tissue samples by ICP-MS are summarized

in Fig. 6. It was evidenced that the exposure time had an effect on accumulation, which was detected in all NP treatments, even in those cases where AEDT-mediated washing was used. Trace amounts of gold for the control experiments are within the experimental errors and are considered as zero values. Gold concentrations found after one hour of exposure were significantly less than those observed after six hours of exposure, showing that gold accumulation increases with the exposure time.

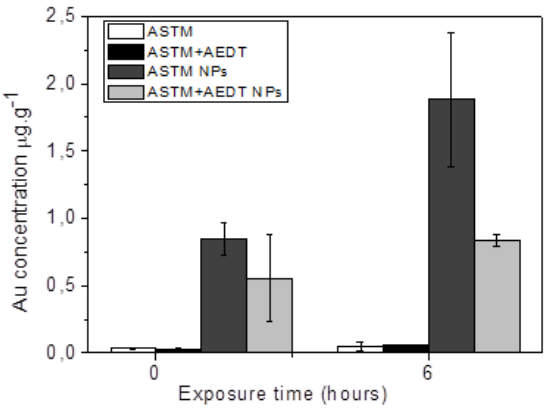

Fig. 6 Au concentrations ($\mu g.g^{-1}$ wet weight) in gills tissues after *in vitro* exposures of 1 and 6 hours exposure with a 750 $\mu g \cdot L^{-1}$ of AuNPs in standard SW. Control experiments are also included. Tissues were washed with SW or alternatively with a SW EDTA solution and SW.

Uptake found in these accumulation analyses, as well as location by electron microscopy in the section below, demonstrates cell viability for the exposure time in our *in vitro* experiments. In particular, histopathological alterations in the gills after exposure to Au NPs, compared to control experiments, have been concluded by optical microscopy from semi-thin sections after toluidine blue staining (ESI, Fig. S1†). These alterations will be discussed below with regard to ultrastructural characterization using the STEM-in-SEM methodology.

Subcellular localization of AuNPs

The feasibility of the optimized STEM-in-SEM methodology for ultrastructural characterization at the Bio-Nano interface is corroborated below. The main results are summarized in Fig. 7. Unlike in control gill samples (Figs. 7A-B), electron-dense particles were observed in the gill samples exposed to AuNPs (Figs. 7C-H). These electron-dense particles were located in secretory granules placed in the apex of ciliary cells (Figs. 7C-D), outside the gill epithelium attached to microvilli and cilia, or associated to excreted secretory granules and cell debris (Figs. 7E-G). For all of these electron-dense particles, the presence of Au was corroborated by X-ray microanalysis (ESI, Fig. S2†), in contrast to control samples in which Au was not found. As we discussed in the previous section, analysis of tissue samples by ICP-MS can be performed in order to obtain representative quantitative data of bioaccumulation. The joint approach between results obtained by ICP-MS analysis and those corresponding to X-ray microanalysis coupled to STEM-in-SEM is ideal for a most pertinent chemical element analysis of the sample. The detection limit in EDX analysis reaches values as low as 0.1 wt% under optimum conditions [22]. This should be borne in mind because as the Au concentrations found in this study by ICP-MS are below the detection limit, then the technique does not meet the requirement of representativeness in quantification capability. However, this cannot rule out the possibility to analyze and to differentiate the chemical composition of distinct particles through EDX analysis. In fact, Fe and Ti containing particles (likely from natural bivalve´s feeding) were also detected as shown in ESI (Fig. S3 and S4†).

In a previous study with the marine bivalve *Scrobicularia plana*, 40 nm AuNPs were detected close to the basal site of microvilli, suggesting the ability of AuNPs to penetrate this epithelium[23]. Although accumulation pathways between *in vivo* and *in vitro* experiments cannot be compared directly, the improved STEM-in-SEM methodology that is shown here allowed for the precise location of the accumulation of AuNPs in distinct cell organelles. Interestingly, the AuNPs attachment to microvilli (between other organelles) is a point of similarity between the bivalve *in vivo* experiment and our *in vitro* assay. The improved STEM-in-SEM technique has also been demonstrated to be highly useful to assess histopathological effects. Histopathological alterations in the gills of clams have been reported to be very sensitive after exposure to a variety of xenobiotics and to the presence of different kinds of parasites since they play a crucial role in respiration, food collection and absorption/digestion[24-28]. In the present work, a well-known series of histopathological lesions in the gill tissues have been observed in gill explants after short exposures (6 h) to the AuNPs solution. These alterations included cell and tissue level alterations. Specifically, epithelial cells (both ciliary and cuboidal cells) were highly vacuolated (Fig. 7C-H), with a hydropic cytoplasm, thickened basal lamina, and a loss of cilia and microvilli in the apex of both cell types. Considering the general picture of histopathological alterations, it can be concluded that this may cause an impairment of several physiological functions, such as respiration and feeding,

which could affect the health status of the gill samples as a result of short-term exposure to AuNPs.

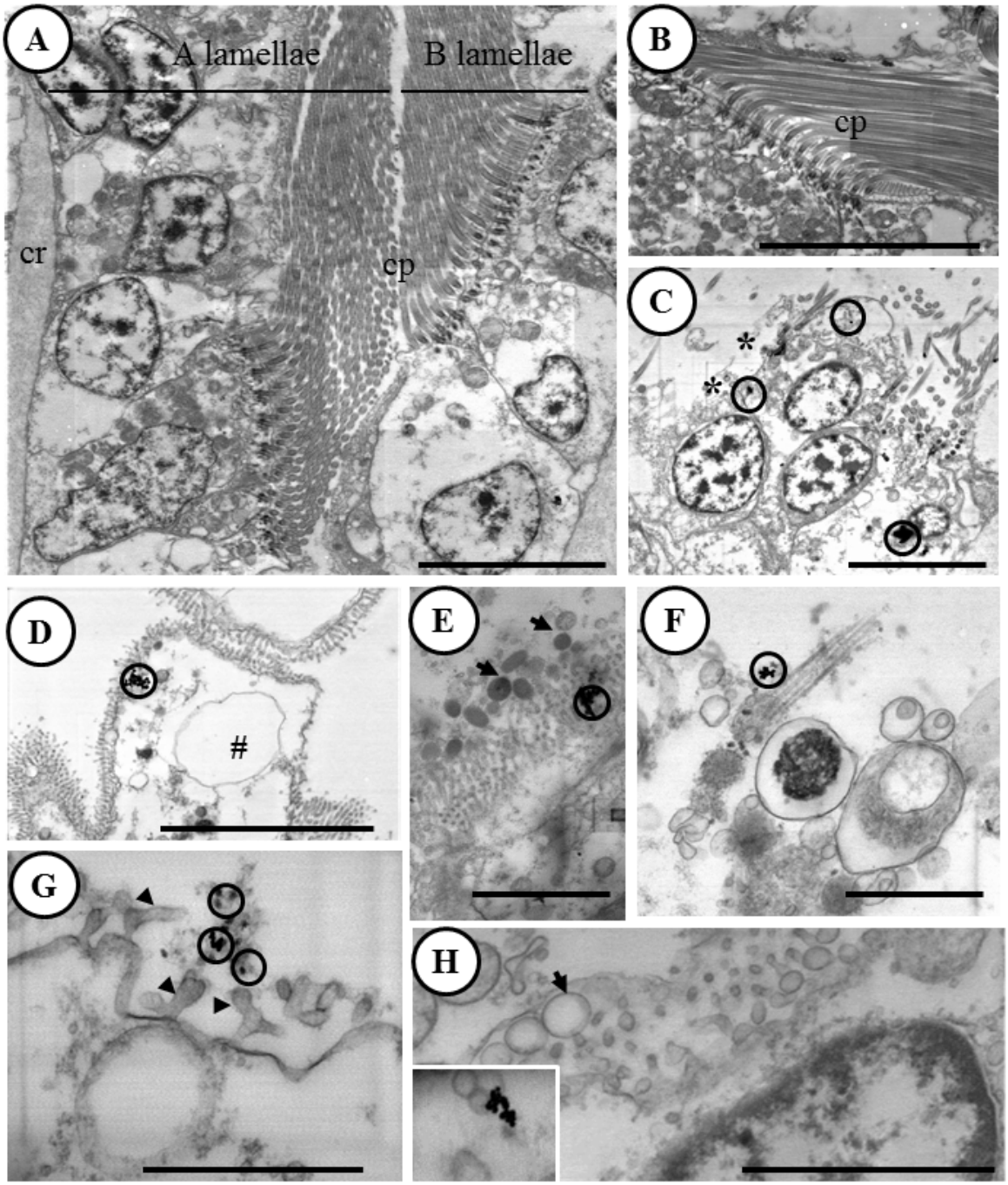

Fig. 7 Electron micrographs (STEM) of the gills of control clams (A, B) and clams exposed to Au NPs (C-H). (A) Regular arrangement of two adjacent lamellae with the ciliary plate (cp) between them. (B) Detail of the apex of ciliated cells. (C) Ciliary cells nearly devoid of cilia (*). Note Au NPs (circles) in the apex and in the extracted cytoplasm of damaged cells. (D) AuNPs internalized in the apex of disrupted epithelial cells. (E) Secretion granules (arrows) and AuNPs among cilia. (F) Detail of secretory granules and AuNPs attached to the basement of cilia. (G) AuNPs among modified microvilli (arrowheads). (H) Secretory granules among microvilli and detail (inset) of AuNPs attached to the granules. cr, cartilage rods; cp, ciliary plates; circles, AuNPs; #, haemolymphatic sinuses. Scale bars: A: 10 µm; B, C, D, E: 5 µm; F, G: 1 µm; H: 3 µm.

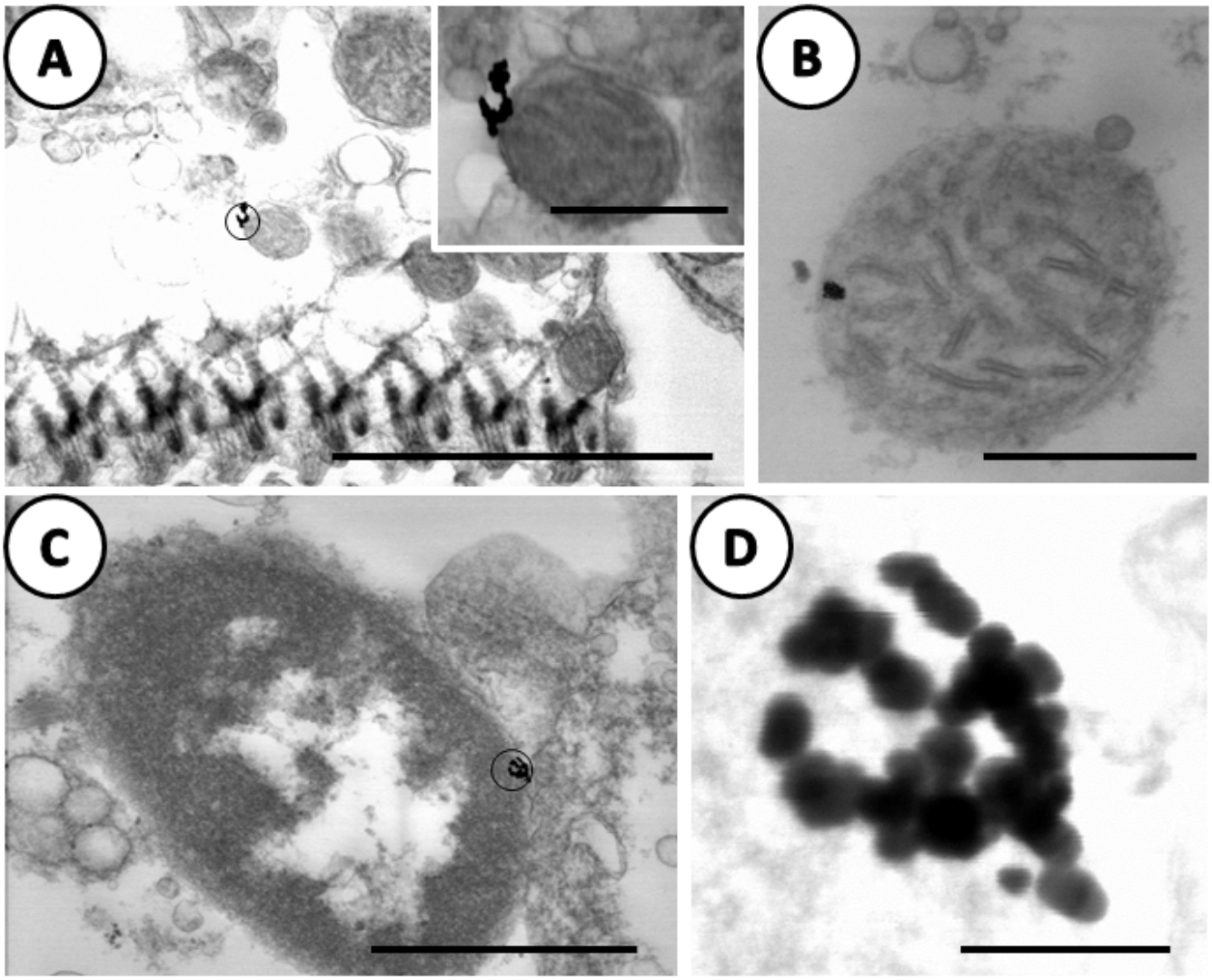

Fig. 8 High-resolution electron micrographs (STEM) of the gills of clams exposed to Au NPs (A-D). (A) Electron-dense particles in the apical part of epithelial cells associated to the outer membrane of the mitochondria (inset). (B) Detail of electron-dense particles in a mitochondrion. (C) Electron-dense particles attached to the nuclear envelop. (D) Detail of electron-dense nanoparticles encircled in C. Scale bars: A: 3 µm (inset: 500 nm), B: 500 nm, C: 200 nm, D: 100 nm.

In Fig. 8, images have been selected at higher magnifications. Electron-dense particles in the gills of clams exposed to AuNPs were clearly visible in the apex of epithelial cells attached to the outer membrane of the mitochondria (Figure 8A) and to the nuclear envelop of the same cell type (Fig. 8C). In Figure 8B, the electron-dense particles appeared to be inside the mitochondria; the interpretation is not clear as this image was obtained from an area with excreted secretory granules and cell debris around the mitochondria. What is important to emphasize is the high resolution achieved both at the tissue and NPs features. Another important result is that in the case of gold NPs, which are highly resistant to solubilization or oxidation, the electron-dense particles, clearly visible in Fig. 8D, maintain the original shape and size of the dosed AuNPs even after internalization.

Conclusions

Well characterized AuNPs suspensions have been prepared by the citrate-reduction method and used for *in vitro* exposure experiments of tissue samples to determine the location of accumulated NPs as well as to evaluate the tissue damage. The imaging of relatively thick specimens (~200-300 nm slices) by STEM-in-SEM is presented here as a powerful electron microscopy technique that makes the localization of high Z contrast engineered NPs in low Z contrast tissue matrices relatively easy.

In accordance with the worldwide use of marine bivalve molluscs as pollutant indicators in ecological monitoring programs, application of the technique to ecotoxicity research has been demonstrated i*n vitro* using gill explants of the bivalve *Ruditapes philippinarum.* The optimized STEM-in-SEM methodology showed great feasibility for ultrastructural characterization at the Bio-Nano interface achieving resolutions well below 10 nm at magnifications over 100 kx for experimental sample thickness between 300 and 200 nm. AuNPs that accumulated in the gill tissues after *in vitro* exposure in sea water media were localized not only in secretory granules and cell debris but also attached to organelles such as mitochondria and, most interestingly, to microvilli.

It can be concluded that the improved STEM-in-SEM technique presented herein provides a simple and useful tool to be applied in ecotoxicological research to assess the subcellular location of nanomaterials and their possible toxic effects. Its extensive use is encouraged due to its time efficiency and sample thickness versatility.

Acknowledgements

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Notes and references

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† Electronic Supplementary Information (ESI) available: Micrographs of semithin sections of the gills as well as the typical identification of electron-dense contrasts by EDX analysis.

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