

Analytical Methods

Accepted Manuscript



This is an *Accepted Manuscript*, which has been through the Royal Society of Chemistry peer review process and has been accepted for publication.

Accepted Manuscripts are published online shortly after acceptance, before technical editing, formatting and proof reading. Using this free service, authors can make their results available to the community, in citable form, before we publish the edited article. We will replace this *Accepted Manuscript* with the edited and formatted *Advance Article* as soon as it is available.

You can find more information about *Accepted Manuscripts* in the [Information for Authors](#).

Please note that technical editing may introduce minor changes to the text and/or graphics, which may alter content. The journal's standard [Terms & Conditions](#) and the [Ethical guidelines](#) still apply. In no event shall the Royal Society of Chemistry be held responsible for any errors or omissions in this *Accepted Manuscript* or any consequences arising from the use of any information it contains.

1
2
3
4
5
6
7 **Laser ionization/time-of-flight mass spectrometry**
8
9
10
11 **for the direct analysis of emulsions**
12
13
14
15
16
17
18
19

20 **Hidaka Ishigami, Yukihiro Tsuda and Tomohiro Uchimura***
21
22
23
24
25

26
27 Department of Materials Science and Engineering, Graduate School of Engineering, University
28 of Fukui, 3-9-1 Bunkyo, Fukui 910-8507, Japan
29
30
31
32
33
34
35

36 * To whom correspondence should be addressed. E-mail: uchimura@matse.u-fukui.ac.jp
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

Abstract

A direct method for the monitoring of emulsions was developed using laser ionization/time-of-flight mass spectrometry. A pair of concentric capillaries stably introduced an oil-in-water microemulsion into a mass spectrometer. The system was applied to the monitoring of a milky white emulsion. With this system, the average of the peak areas was calculated to monitor the local concentration of dispersed toluene, which was related to a collapse of the emulsion. Strong spikes appeared, particularly when the emulsion was measured during high turbidity, which suggested the existence of highly concentrated toluene in the emulsion. The mass of the constituents was measurable in both micro- and macroemulsions. Therefore, this method could be widely applied to emulsion studies that are needed for stability evaluation and quality control.

Introduction

Laser ionization/time-of-flight mass spectrometry (LI/TOFMS) has several advantages such as a superior optical selectivity and robustness against contamination.¹⁻⁵ Here, it was applied to the trace analysis and/or real-time monitoring of components in mixture samples.⁶⁻¹¹ This method is mainly used to measure gas samples. Solid or liquid samples normally are vaporized by heating or by laser ablation/desorption.¹²⁻¹⁸ Though electrospray ionization (ESI)-MS is generally employed for the mass analysis of a liquid sample, LI/TOFMS may become a superior means for the detection of a liquid sample due to great advantages in spectroscopic selectivity. We recently reported the introduction of an aqueous solution directly into TOFMS to measure an aromatic compound.¹⁹

The target sample in the present study was an emulsion, which is a heterogeneous system wherein one liquid is dispersed as droplets into another liquid but is not dissolved, such as with oil and water. Emulsions exist in a diverse range of applications such as materials, foods, and pharmaceuticals.²⁰ Studies on the stability of emulsions are very important, because the condition of an emulsion can easily be changed, e.g., creaming, flocculation, coalescence, and Ostwald ripening.²⁰⁻²² The collapse process is quite complicated, and depends on several factors that includes the concentrations of the dispersoid and the surfactant. To date, several techniques have been used to evaluate emulsions: spectrometry, light scattering, and small X-ray scattering.²³⁻²⁹

To the best of our knowledge, an online mass spectrometric and stability evaluation approach of emulsions has not yet been accomplished. Normally, appropriate pretreatments such as drying or matrix addition are performed before the mass analysis.^{30,31} Recently, the mass

1
2
3 spectra of surfactant-stabilized droplets have been reported using ESI.³² If real-time monitoring
4
5 of an intact emulsion in mass spectrometry could be achieved without pretreatment, the method
6
7 would be quite useful for a detailed evaluation of the collapse process and for the quality control
8
9 of an emulsion. Further applications of the method also would be possible. Study on the
10
11 kinetics of emulsion polymerization would be one example.³³
12
13
14
15

16 The use of LI/TOFMS for the real-time monitoring of an emulsion was first developed in
17
18 the present study. Using an oil-in-water (O/W) emulsion, a stable sample introduction method
19
20 was investigated, and the application of this method for the measurement of an unstable milky
21
22 emulsion is discussed here.
23
24
25
26
27
28
29

30 **Experimental**

31 32 33 **Reagents and sample preparation**

34
35
36
37 Toluene, sodium dodecyl sulfate (SDS), and 1-pentanol were all purchased from Wako Pure
38
39 Chemical Industries (Osaka, Japan) and used as a dispersoid, a surfactant, and a co-surfactant,
40
41 respectively. In the sample preparation, SDS and 1-pentanol were added to distilled water, and it
42
43 was shaken manually. Then toluene was slowly added, and the mixture was shaken manually
44
45 and/or sonicated. The mixture was sealed and left at room temperature for at least a few days
46
47 until use. The final concentrations are mentioned in each figure caption. In the present study,
48
49 two types of emulsions, a transparent microemulsion and a milky emulsion (macroemulsion),
50
51 were prepared. The transparent microemulsion remained clear for an extended period of time,
52
53
54
55
56
57
58
59
60

1
2
3 but a collapse phenomenon, likely to be creaming, was observed in the case of the milky
4
5 emulsion.
6
7
8
9

10 11 12 **Apparatus** 13

14
15
16 The LI/TOFMS instrument used in the present study is shown in Figure 1, as described in the
17
18 literature.^{19,34}
19
20

21
22 Two types of the sample introduction technique were examined in the present study. In
23
24 the first, an emulsion was introduced using a single deactivated fused-silica capillary (GL
25
26 Sciences), and the second used a pair of concentric capillaries, as illustrated in Figure 1. In the
27
28 latter case, the emulsion was passed through the inner capillary (outer diameter; 150 μm), while
29
30 ambient air or nitrogen gas was introduced from the outer column. The flow rate of the gas was
31
32 ca. 2 mL/min in the present study; in some cases, that was adjusted by a flow meter (Kofloc, RK-
33
34 1250, Kyoto, Japan). The pressure in the chamber was ca. 2×10^{-3} Pa when using a single
35
36 capillary and ca. 1×10^{-2} Pa or below when using a pair of concentric capillaries. The length of
37
38 the capillary column for the passage of the emulsion was ca. 450 mm. The position of the tip of
39
40 the inner column was set 2-3 mm inward from that of the outer column. The inlet side of the
41
42 capillary was directly inserted into a sample container with rims that were sealed with a piece of
43
44 parafilm, while a lid with a small hole for passage through the capillary was used in the
45
46 experiment shown in Figure 5 in order to suppress the sample volatilization. The inlet side of the
47
48 capillary was set a few centimeters from the bottom of the container. The capillary column was
49
50 set almost even with the ground.
51
52
53
54
55
56
57
58
59
60

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

The fourth harmonic emission of a Nd:YAG laser (Rayture Systems, GAIA II, 10 Hz, 266 nm, 20-400 μ J, 4 ns, Tokyo) was used for ionization. The laser was focused with a plano-convex lens with a focal length of 200 mm. The ionization position was adjusted to 2 mm from the capillary tip. A linear-type TOFMS, which is now commercially available (Hikari-GK, HGK-1, drift length 60 cm, Fukuoka, Japan), was used in the present study. The capillary nozzle was indirectly heated by raising the temperature of the flange to 40 °C. The ion signals were detected by a microchannel plate assembly (Hamamatsu Photonics, F4655-11, Shizuoka, Japan) and were recorded using either a digitizer (Acqiris/Agilent Technologies, AP240) or a digital oscilloscope (Tektronix, TDS5104). All data were recorded after a certain time following the insertion of the capillary into the sample. In the present study, the ionization laser was introduced 10-30 seconds after starting the recording in order to determine the baseline level. In the measurement of Figures 2 and 3, the data were recorded without averaging, i.e., the signal intensity was acquired every 0.1 seconds. On the other hand, an average of 5 laser shots was applied in the experiment shown in Figure 5 in order to reduce the data volume arising from an extended period of recording. The data were processed and analyzed with a program constructed in-house using LabVIEW programming software (National Instruments).

Results and discussion

Introduction of a microemulsion using a single capillary

The sample introduction was first applied using a single capillary column. Figure 2 shows a two-dimensional display of the data for a microemulsion using a capillary with an inner diameter

1
2
3 of 50 μm . In this figure, two lines arising from a toluene ion ($m/z = 92$) and a fragment ion (m/z
4 = 91, a tropylium ion was one of the candidates) were primarily observed. It seemed odd that
5 these lines seemed to be in waves, or in other words, that the flight times were frequently
6 shortened. The result of the selected ion monitoring of the corresponding species, which was
7 constructed by extracting the region of ca. 10 μs with regards to flight time, is also shown in
8 Figure 2. Several spikes, which had simultaneously arisen when the flight times were shortened,
9 can be observed in the figure. These results can be explained by assuming that the initial
10 velocity and the concentration of toluene molecules both were often increased at the ionization
11 point, probably due to the explosive introduction of an emulsion that had accumulated at the tip
12 of the capillary. A fluctuation in the flight time is inappropriate for calculating the mass of an
13 analyte, and, therefore, the introduction method using a single capillary was considered to be
14 unsuitable for emulsion analysis.
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35

36 **Introduction of a microemulsion using a pair of concentric capillaries**

37
38
39 In order to correct the flight time fluctuation, as shown in Figure 2, several experimental
40 conditions were modified. First, a pair of concentric capillaries, rather than a single capillary,
41 was employed as a sample introduction technique. The result is shown in Figure S1
42 (Supplemental information), where the inner diameter of the inner capillary was 50 μm . In this
43 case, though spikes still appeared, the fluctuation of the flight time almost disappeared. These
44 results suggest that, despite the fact that the explosive sample introduction still existed, the initial
45 velocity of the analyte molecules became constant by introducing the gas from the outer capillary.
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

In order to suppress the explosive introduction, an inner capillary with a smaller inner diameter was employed to reduce the amount of the analyte molecule per unit of time. Schepler et al. have reported that they employed a thinner capillary (inner diameter; 25 μm) to prevent the formation of droplets at the tip, although their configuration for liquid sample introduction was considerably different from that of the present study.³⁵ Figure 3 shows an example of the result using an inner capillary with an inner diameter of 20 μm . Neither the spikes nor the fluctuation of the flight time was observed. As a result, the modification of the capillary configuration for applying the outer gas and the use of an inner capillary with a smaller inner diameter reduced the size of the species undergoing laser ionization from small droplets to clusters to possibly even isolated molecules. Note that, though the deviation of the signal intensity of the selected ion monitoring seems large at first glance, the figure is depicted using raw data, and of course it can be altered with effects such as smoothing in order to reduce the deviation when requested. The mass spectrum obtained using this experimental condition is shown in Figure 4. A selective measurement was definitely achieved, i.e., only the peaks of toluene and the fragment were observed, while the peaks of SDS ($m/z = 288$) or 1-pentanol ($m/z = 88$), the concentrations of which were about 1 order of magnitude higher than that of toluene, were not detected.

The appropriate experimental conditions to stably introduce an emulsion are as follows:

- (1) a smaller inner diameter of the inner capillary is preferable, e.g., 20 μm is preferred and 50 μm is too large;
- (2) the position of the tip of the inner capillary should be set 2-3 mm inward from the tip of the outer diameter, and when the tips of both capillaries are true, the spikes are substantially observed.

There was an additional problem with this sample introduction method. Once the inner capillary was removed from the bottle containing the emulsion and attempts were made to measure the next sample, the flow of the emulsion was often stopped due to the clogging

1
2
3 of SDS. The cause of this problem remains unknown, but the capillary can easily be exchanged
4
5 with a new one if it occurs.
6
7
8
9

10 11 12 **Monitoring of a milky emulsion** 13

14
15
16 The present method was applied to the measurement of a milky white emulsion. Figure 5 shows
17
18 the time course of the peak area for a milky emulsion (black line), and that of the peak area
19
20 averaged every 30 seconds is also shown (red line) in order to simply judge the obtained results.
21
22 For this measurement, the emulsion container was handled gently. After that, the capillary was
23
24 inserted into the lower part of the container and left for at least 30 minutes, and then the
25
26 container was shaken by hand to give the emulsion a white turbidity. As shown in this figure,
27
28 the red line indicates that the average peak area was increased a few minutes after shaking, and
29
30 then decreased with elapsed time. The time course of the peak area was considered to be the
31
32 concentration of toluene at the sampling point, and the decay was correlated to the collapse of an
33
34 emulsion such as creaming. The peak area approached a nearly constant value, which meant that
35
36 a certain concentration of toluene was present, e.g., as a microemulsion, after the visible
37
38 creaming.
39
40
41
42
43
44

45
46 It is noteworthy that many strong spikes appeared by the peak area depicted with a black
47
48 line after the emulsion had gained turbidity, then subsequently, the number, as well as the height,
49
50 of the spikes gradually decreased. It is true that the average signal area showed the same
51
52 tendency, but this does not mean that both forms of data provide the same information about the
53
54 local environment in an emulsion. That is to say, that although the possibility of an unstable
55
56 sample introduction cannot be denied, as discussed previously, it is reasonable to assume that the
57
58
59
60

1
2
3 arising spikes were caused by the introduction of highly concentrated toluene, e.g., toluene
4 droplets, into the inlet of the capillary, and/or the occurrence of a collapse, such as flocculation,
5 during passage through the capillary column. The decrease in the spikes could be explained by a
6 decrease in the domain of highly concentrated toluene around the inlet. Good reproducibility of
7 the appearance of strong spikes could have been obtained by measuring the emulsions with high
8 turbidity.
9

10
11
12
13
14
15
16
17
18
19 In the present study, the real-time monitoring of emulsions, even where the local
20 concentrations and conditions were dramatically changed, was achieved using LI/TOFMS. The
21 average peak area, as well as the spikes, can provide interesting information about the stability of
22 an emulsion. The tendencies towards decreases that were demonstrated were considered to be a
23 function of the sampling point; they may be increased in other sampling points. Other methods
24 for evaluating the stability of an emulsion should be used and compared against the results
25 obtained in the present study. Fortunately, the present system can easily be combined with
26 several other conventional methods,^{26,27} since the capillary was simply inserted into the container
27 of an emulsion with no other interference.
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43

44 **Conclusions**

45
46
47
48 An on-line mass analytical method of emulsion was developed using LI/TOFMS. The local
49 concentration of dispersed toluene in both the micro- and macroemulsions was measurable by
50 the average peak area. In addition, the existence of high concentrations of toluene, such as
51 droplets, was observable by the characteristic spikes. All the results could be obtained with no
52 pretreatment as demonstrated. From the point of view of the advantages such as selectivity and
53
54
55
56
57
58
59
60

1
2
3 robustness, LI/TOFMS can be suitable for the evaluation of an emulsion with inherently high
4 concentrations of each component. Therefore, this method could be a powerful tool to evaluate
5
6 the stability of emulsions and could be useful for many applications, such as the quality control
7
8 of emulsions and the monitoring of intermediates/products generated from emulsion
9
10 polymerization.
11
12
13
14
15
16
17
18
19

20 **Acknowledgments**

21
22
23 This research was supported by a Grant-in-Aid for Scientific Research from the Japan Society
24 for the Promotion of Science (JSPS). This work was also supported by a research grant provided
25
26 by The Iwatani Naoji Foundation.
27
28
29
30
31
32
33
34

35 **Supplemental information**

36
37
38 Supplementary data associated with this article are available on the Web of *Analytical Methods*.
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

References

- 1 R. Tembreull, C. H. Sin, H. M. Pang and D. M. Lubman, *Anal. Chem.*, 1985, **57**, 2911.
- 2 R. Zimmermann, U. Boesl, D. Lenoir, A. Kettrup, Th. L. Grebner and H. J. Neusser, *Int. J. Mass Spectrom. Ion Processes*, 1995, **145**, 97.
- 3 O. P. Haefliger and R. Zenobi, *Anal. Chem.*, 1998, **70**, 2660.
- 4 H. J. Heger, R. Zimmermann, R. Dorfner, M. Beckmann, H. Griebel, A. Kettrup and U. Boesl, *Anal. Chem.*, 1999, **71**, 46.
- 5 Y. Sakoda, T. Uchimura and T. Imasaka, *Anal. Chem.*, 2010, **82**, 1283.
- 6 G. Rhodes, R. B. Opsal, J. T. Meek and J. P. Reilly, *Anal. Chem.*, 1983, **55**, 280.
- 7 A. Marshall, A. Clark, K. W. D. Ledingham, J. Sander, R. P. Singhal, C. Kosmidis and R. M. Deas, *Rapid Commun. Mass Spectrom.*, 1994, **8**, 521.
- 8 K. Misawa, K. Tanaka, H. Yamada, Y. Goto, J. Matsumoto, Y. Yamato, S. Ishiuchi, M. Fujii, K. Tanaka, K. Endo and S. Hayashi, *Int. J. Engine Res.*, 2009, **10**, 409.
- 9 T. Matsui, K. Fukazawa, M. Fujimoto and T. Imasaka, *Anal. Sci.*, 2012, **28**, 445.
- 10 Y.-C. Chang and T. Imasaka, *Anal. Chem.*, 2013, **85**, 349.
- 11 T. Kuraishi and T. Uchimura, *Anal. Chem.*, 2013, **85**, 3493.
- 12 S. G. Hansen, *J. Appl. Phys.*, 1989, **66**, 3329.
- 13 M. Tsunekawa, S. Nishio and H. Sato, *Jpn. J. Appl. Phys.*, 1995, **34**, 218.

- 1
2
3 14 C.-H. Lin, Y. Murata and T. Imasaka, *Anal. Chem.*, 1996, **68**, 1153.
4
5
6
7 15 M. S. de Vries and H. E. Hunziker, *Appl. Surf. Sci.*, 1996, **106**, 466.
8
9
10 16 T. Uchimura and T. Imasaka, *Anal. Chem.*, 2000, **72**, 2648.
11
12
13 17 J. Matsumoto, K. Kai and T. Imasaka, *Anal. Chem.*, 2003, **75**, 346.
14
15
16
17 18 J. Matsumoto, K. Nishimura, T. Uchimura and T. Imasaka, *Anal. Chim. Acta*, 2003, **484**,
18 163.
19
20
21
22 19 G. Tokumoto, H. Saburi, S. Miyagawa and T. Uchimura, *Bunseki Kagaku*, 2013, **62**, 595.
23
24
25
26 20 K. Holmberg, B. Jönsson, B. Kronberg and B. Lindman, *Surfactants and Polymers in*
27 *Aqueous Solution*, 2nd ed.; John Wiley & Sons: England, 2003.
28
29
30
31 21 M. M. Robin, *Curr. Opin. Colloid Interface Sci.*, 2000, **5**, 265.
32
33
34
35 22 I. Capek, *Adv. Colloid Interface Sci.*, 2004, **107**, 125.
36
37
38
39 23 L. Auvray, J.-P. Cotton, R. Ober and C. Taupin, *J. Physique*, 1984, **45**, 913.
40
41
42 24 M. J. Monteiro and J. de Barbeyrac, *Macromolecules*, 2001, **34**, 4416.
43
44
45 25 F. Kitagawa, M. Murase and N. Kitamura, *J. Org. Chem.*, 2002, **67**, 2524.
46
47
48
49 26 K. Kageshima, T. Takei and Y. Sugitani, *Anal. Sci.*, 2003, **19**, 757.
50
51
52 27 O. Mengual, G. Meunier, I. Cayré, K. Puech and P. Snabre, *Talanta*, 1999, **50**, 445.
53
54
55 28 S. Tsukahara, Y. Shishino and T. Fujiwara, *Langmuir*, 2011, **27**, 7392.
56
57
58
59
60

- 1
2
3
4 29 D. S. Miller, X. Wang, J. Buchen, O. D. Lavrentovich and N. L. Abbott, *Anal. Chem.*,
5 2013, **85**, 10296.
6
7
8
9 30 B. Thomson, Z. Wang, A. Paine, G. Lajoie and A. Rudin, *J. Polym. Sci. Pol. Chem.*, 1995,
10 **33**, 2297.
11
12
13
14 31 F. G. Hoogland and J. J. Boon, *Int. J. Mass Spectrom.*, 2009, **284**, 72.
15
16
17
18 32 C. A. Smith, X. Li, T. H. Mize, T. D. Sharpe, E. I. Graziani, C. Abell, and W. T. S. Huck,
19 *Anal. Chem.*, 2013, **85**, 3812.
20
21
22
23 33 M. Nomura, H. Tobita and K. Suzuki, *Adv. Polym. Sci.*, 2005, **175**, 1.
24
25
26
27 34 H. Okudaira, T. Uchimura and T. Imasaka, *Anal. Sci.*, 2012, **28**, 638.
28
29
30
31 35 C. Schepler, M. Sklorz, J. Passig, G. Famiglini, A. Cappiello and R. Zimmermann, *Anal.*
32 *Bioanal. Chem.*, 2013, **405**, 6953.
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

Figure Captions

Figure 1. Experimental apparatus of LI/TOFMS. An enlarged view of a pair of concentric capillaries for sample introduction is also shown.

Figure 2. Two-dimensional display for a microemulsion obtained using a single capillary for sample introduction. The inner diameter of the capillary was 50 μm . The time course of the peak area of observed ions, as indicated in the display, is also shown. Concentration: 500 $\text{ng}/\mu\text{L}$ for toluene, 10,200 $\text{ng}/\mu\text{L}$ for SDS, and 6,300 $\text{ng}/\mu\text{L}$ for 1-pentanol.

Figure 3. Two-dimensional display for a microemulsion obtained using a pair of concentric capillaries where the inner diameters of the inner and outer capillaries were 20 and 250 μm , respectively. The time course of the peak area of toluene ion is also shown. Concentration: 400 $\text{ng}/\mu\text{L}$ for toluene, 6,600 $\text{ng}/\mu\text{L}$ for SDS, and 3,000 $\text{ng}/\mu\text{L}$ for 1-pentanol.

Figure 4. Mass spectrum for a microemulsion averaged from 200 single transients. The experimental conditions were the same as those in Figure 3.

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

Figure 5. Time course of the peak area of toluene in an unstable emulsion (black line). The red circle/line exhibits the results of the black line averaged every 30 seconds. The photos of an emulsion taken at different times are indicated by gray arrows. The inner diameters of the inner and outer capillaries were 25 and 320 μm , respectively. Concentration: 5,000 $\text{ng}/\mu\text{L}$ for toluene, 5,700 $\text{ng}/\mu\text{L}$ for SDS, and 3,000 $\text{ng}/\mu\text{L}$ for 1-pentanol.

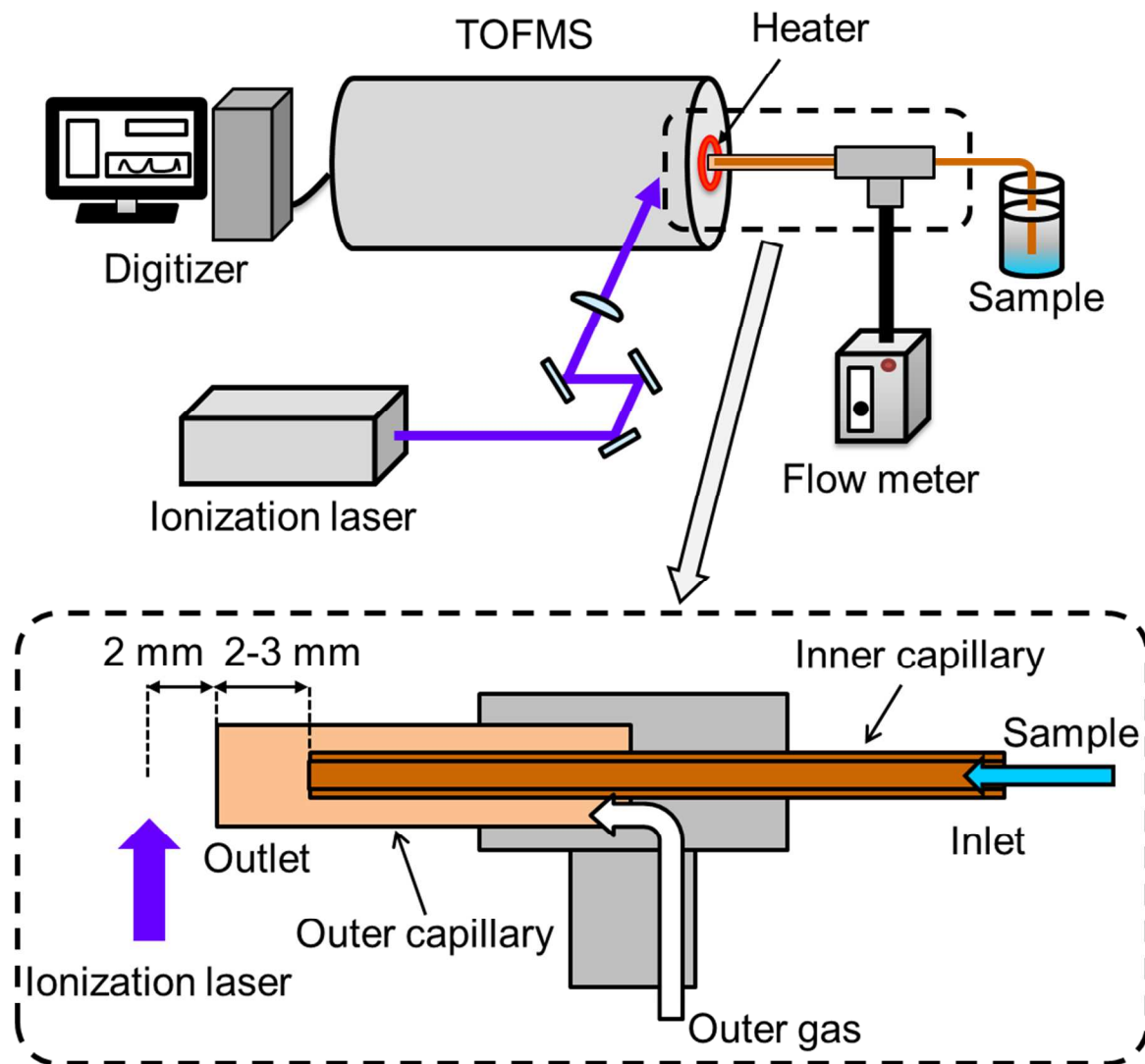


Figure 1. Experimental apparatus of LI/TOFMS. An enlarged view of a pair of concentric capillaries for sample introduction is also shown.

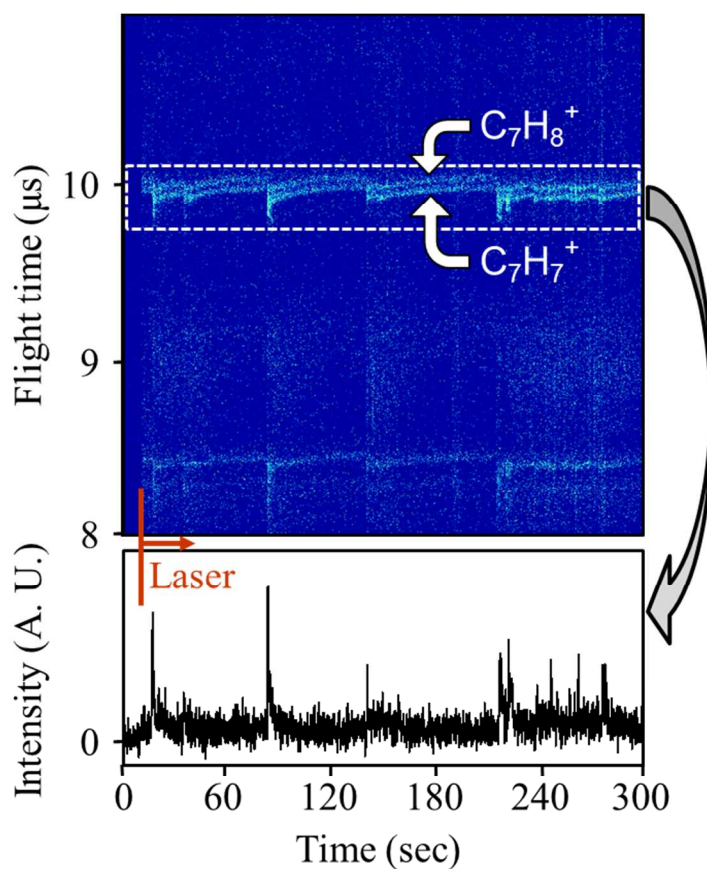


Figure 2. Two-dimensional display for a microemulsion obtained using a single capillary for sample introduction. The inner diameter of the capillary was 50 μm . The time course of the peak area of observed ions, as indicated in the display, is also shown. Concentration: 500 $\text{ng}/\mu\text{L}$ for toluene, 10,200 $\text{ng}/\mu\text{L}$ for SDS, and 6,300 $\text{ng}/\mu\text{L}$ for 1-pentanol.

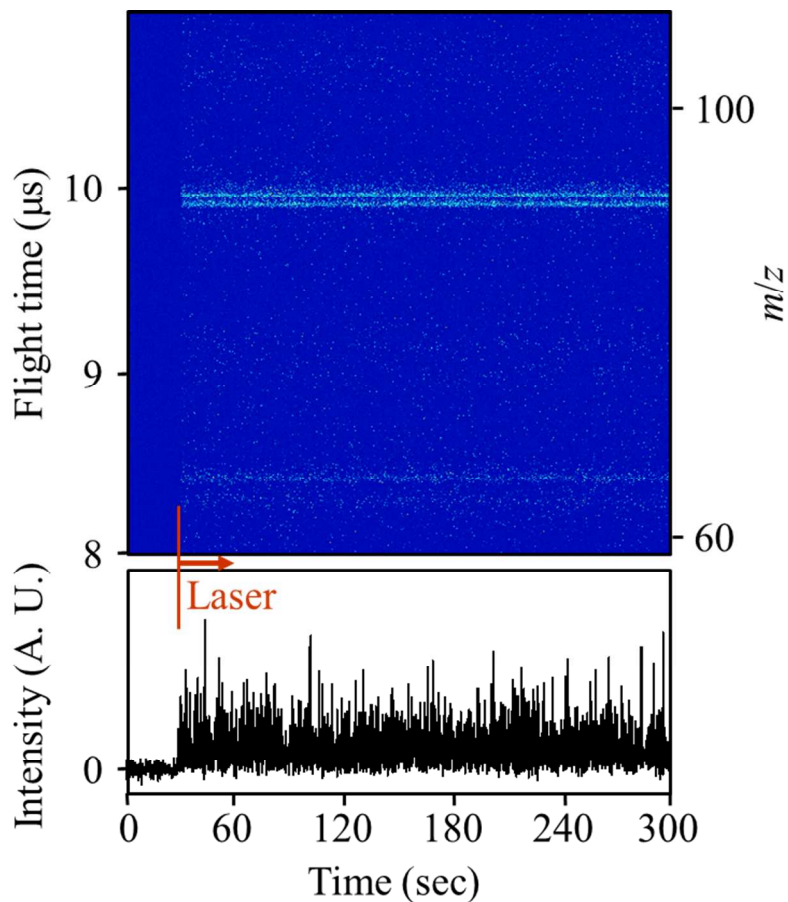


Figure 3. Two-dimensional display for a microemulsion obtained using a pair of concentric capillaries where the inner diameters of the inner and outer capillaries were 20 and 250 μm , respectively. The time course of the peak area of toluene ion is also shown. Concentration: 400 $\text{ng}/\mu\text{L}$ for toluene, for 6,600 $\text{ng}/\mu\text{L}$ SDS, and 3,000 $\text{ng}/\mu\text{L}$ for 1-pentanol.

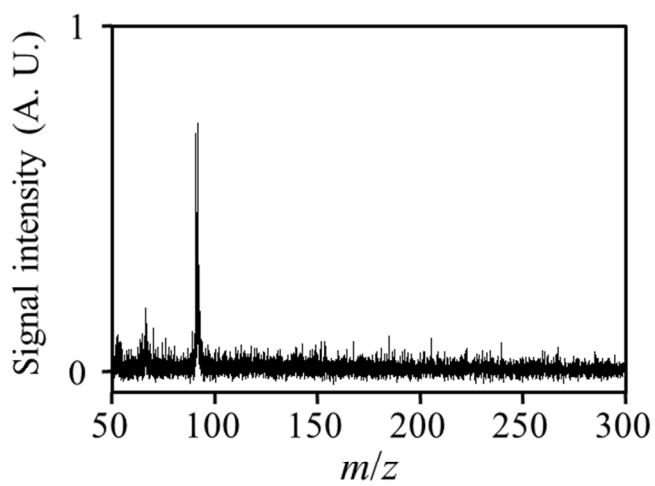


Figure 4. Mass spectrum for a microemulsion averaged from 200 single transients. The experimental conditions were the same as those in Figure 3.

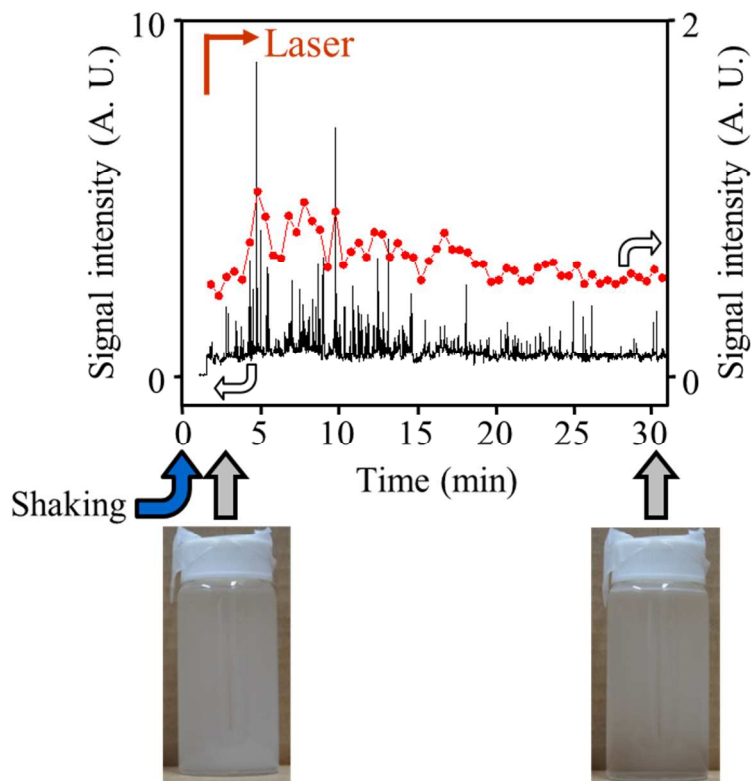
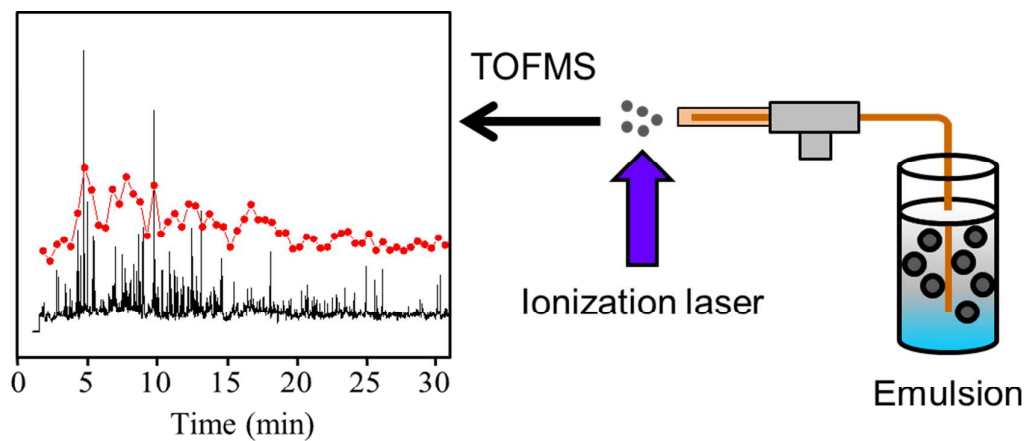


Figure 5. Time course of the peak area of toluene in an unstable emulsion (black line). The red circle/line exhibits the results of the black line averaged every 30 seconds. The photos of an emulsion taken at different times are indicated by gray arrows. The inner diameters of the inner and outer capillaries were 25 and 320 μm , respectively. Concentration: 5,000 $\text{ng}/\mu\text{L}$ for toluene, 5,700 $\text{ng}/\mu\text{L}$ for SDS, and 3,000 $\text{ng}/\mu\text{L}$ for 1-pentanol.

<For Table of Contents>



A direct method for the monitoring of emulsions was developed using laser ionization/time-of-flight mass spectrometry.