

Analytical Methods

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Development of selective sorbent based on magnetic ion imprinted polymer for preconcentration and FAAS determination of urinary cadmium

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Abstract

In this study an ion imprinted polymer (IIP), based on coating of polymer on Fe_3O_4 nanoparticles (NPs) core, was used as a magnetic sorbent for preconcentration and determination of Cd(II) ion level in human urine samples. Synthesized polymer was characterized by IR spectroscopy, scanning electron microscopy (SEM), X-Ray Diffraction analysis (XRD) and thermal analysis (TG/DTA). The optimum conditions, such as pH of the sample, eluent volume, concentration and duration of sorption and elution were investigated. In addition, the effect of some potentially interfering cations on the sorption of Cd(II) was studied. In all samples, Cd(II) was determined by flame atomic absorption spectrophotometer. Under optimal conditions, the detection limit (LOD) of cadmium was $0.6 \mu\text{g L}^{-1}$. The optimized method has been validated using day-to-day and within-day reproducibly experiments and achieved appropriate accuracy and precision. The accuracy of the method was confirmed by analyzing a certified reference material (Seronorm LOT NO2525) and spiked real samples. Ultimately, this method was applied successfully for the preconcentration and determination of Cd(II) ions in urine samples obtained from exposed individuals.

Key words: Cd(II); Magnetic ion imprinted polymer; Selective extraction; Urine samples.

1. Introduction

One of the major aims of analytical toxicology whether in the workplace or environment is determination of chemical substances in environmental or biological samples. Therefore, international or national societies like ACGIH, OSHA, NIOSH and ISO annually introduce and publish new TLVs (Threshold limit value) or PELs (Permissible Exposure Limits) [1-2]. Among various harmful materials, occupational toxicologists put special attention on determination of heavy metals. There is global concern due to rapid industrial development and releasing harmful materials to the environment. Today, most of industries use heavy metals directly or indirectly, so that, large numbers of people are exposed to such material, particularly heavy metals via water, food, air, workplace, and even smoking. Cadmium is one of these heavy metals mostly is used in plating, battery, welding, semi-conductive and alloys industries [2-3]. Also, cadmium is a nonessential material element for human body [4-5]. On the other hand, according to IARC ranking, cadmium has sufficient evidence of being human carcinogen and ranked as Group 1 [6]. Annually, 30000 tons of cadmium are released into the environment because of natural or human activities. Nevertheless, about 4000 to 13000 tons of releasing cadmium is because of human activities which mainly caused by fossil fuels combustion [<https://www.osha.gov/doc/outreachtraining/htmlfiles/cadmium.html>]. Also, OSHA reports that 70000 individuals are employed in their environment with potential risk of cadmium exposures [7]. It is proved that, cadmium has a harmful effect on heart, bones, lungs, and chiefly on the kidney. Cadmium could be accumulated on liver and kidney. After accumulation, it needs about 20 to 30 years to be removed completely from body tissues via urine [8].

According to these facts, evaluation of cadmium is essential. Because of low concentration and intervention of several factors, various separation and preconcentration methods have been developed to evaluate heavy metals such as cadmium [9-11]. Different separation and

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preconcentration methods such as liquid-liquid extraction [12], solid-phase microextraction [13-14], liquid-phase micro-extraction [15], solid-phase extraction [16-18], cloud point extraction [19], co-precipitation [20], molecular imprinted polymer phase extraction [21-25] as well as different instrumental analysis like flame atomic absorption spectrometry (FAAS) [26], graphite furnace atomic absorption spectrometry (GFAAS) [27], inductively coupled plasma mass spectrometry (ICP-MS) [28] have been used for evaluation of metal ions in the spiked or real samples. From these methods, solid-phase extraction (SPE) has been popularly used for detecting pollutants because of simplicity, rapidity, cost-benefit, and minimal reagent consumption [29]. Ion imprinting is a process in which co-polymerization has been performed between functional and cross-linking monomers in the presence of the target ion (the imprint ion). In order to enhance the sensitivity and selectivity of SPE procedure, new sorbent based on ion imprinted polymer which had synthetic specific sites with high selectivity and affinity for the metal ions was developed [30].

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Compared to the conventional imprinted polymers, magnetic imprinted polymers have large surface area and short diffusion route and consequently high extraction efficiency. Rapid extraction dynamics have been achieved by using such sorbent [31-33].

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 Fe_3O_4 nano-particles are one of these magnetic particles which have attracted global attention due to their unique physical and chemical properties like biocompatibility, high mass transfer rates, good fluid–solid contact and extraordinary magnetic properties [34-35]. Recently, a procedure in which combining Fe_3O_4 particles with ion imprinted polymer ($\text{Fe}_3\text{O}_4\text{@IIP}$) for extracting metal ions from samples with complex matrix has been developed [36-38].

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In this study, magnetic ion imprinted polymer has been synthesized and applied as a new sorbent for the separation and preconcentration of trace amount of cadmium from urine followed by its determination with FAAS.

2. Experimental

2.1. Instruments and apparatus

An AA-680 Shimadzu (Kyoto, Japan) flame atomic absorption spectrometer equipped with an air-acetylene flame and a deuterium lamp background correction was used for the determination of cadmium ion (Kyoto, Japan). Conventional hollow cathode lamp with wavelength 228.8 nm was used as the radiation source (Kyoto, Japan). All pH measurements were carried out by a digital WTW Metrohm 827 Ion analyzer (Herisau, Switzerland). Fourier transform infrared spectra ($4000\text{-}400\text{ cm}^{-1}$) were recorded on a Bruker IFS-66 FT-IR Spectrophotometer (Massachusetts, USA). Morphology and size of the magnetic IIP were observed with a Philips Scanning electron microscope (XL-30, Almelo, The Netherlands). The thermal gravimetric and differential thermal analysis (TG-DTA) was carried out on a BHR-Thermo analysis GmbH (Hüllhorst, Germany) under air atmosphere at the heating rate of $10\text{ }^{\circ}\text{C}/\text{min}$. The X-ray diffraction (XRD) patterns was achieved on a Philips-PW12C diffractometer (Amsterdam, The Netherlands) with $\text{Cu K}\alpha$ radiation.

2.2. Chemicals and reagents

All reagents were of analytical grade were purchased from Merck (Darmstadt, Germany). A 1000 mg L^{-1} standard solution of Cd(II), as well as HCl, HNO_3 , N $(\text{CH}_2\text{CH}_3)_3$, toluene, acetonitrile, 2-aminobanzothiazol, FeCl_3 , FeSO_4 , NaOH, NH_4OH (32 % solution), azobisisobutyronitrile (AIBN), ethylene glycol dimethacrylate (EDGMA), 3-Choloro1-propane and trimethoxy vinyl silane were purchased from Merck. The working solutions of Cd (II) were obtained by diluting the standard solution with deionized water, and diluted HCl and NaOH was used to adjust pH values to the various ranges. All the required solutions were prepared using deionized water provided by a Milli-Q (Millipore, Bedford, MA, USA) purification system.

2.3. Procedures

2.3.1. Preparation of Fe₃O₄ nanoparticles

Fe₃O₄ NPs were synthesized according to the previously reported procedure [33]. Briefly, 10.4 g of FeCl₃·6H₂O and 4.0 g of FeCl₂·4H₂O were dissolved in 100 mL of deionized water, degassed with nitrogen gas for 15 min and heated to 80 °C. Then, 15 mL of NH₄OH (32% solution) was added dropwise to the solution. After 15 min the solid was separated by a magnet and washed three times with 0.1 mol L⁻¹ NaCl solution. The formation of NPs was confirmed by IR spectroscopy and X-ray powder diffraction.

2.3.2. Preparation of vinyl functionalized Fe₃O₄

Vinyl functionalization of Fe₃O₄ NPs was performed by reaction of silane agents with Fe₃O₄ NPs according to the recent report [39]. To prepare vinyl functionalized Fe₃O₄ NPs, 1.0 g of prepared Fe₃O₄ NPs was suspended in 50 mL of toluene, following this, 1.0 g of 3-vinyltriethoxysilane was added to the solution and the mixture was stirred for 24 hours. The solid phase was separated from the solvent by a magnet and washed three times with 50 mL of acetone in order to remove any impurities, and then dried at room temperature.

2.3.3. Preparation of vinylated ligand

2-aminobenzothiazole has S and N atoms which show a high tendency to coordinate with cadmium ions, selectively. Furthermore, in the following references thiazole ligands are used as a complexing agent for cadmium/heavy metals determination [40]. On this step vinylated ligand was prepared by the reaction of 2-aminobanzotiazol as a ligand (1 mmol) and of 3-choloro1-propen (1 mmol) in 50 mL solution of triethylamine and methanol (1:4 v/v) at room temperature. After 4 hours, the solvent was removed under the reduced pressure.

2.3.4. Preparation of Fe₃O₄@IIP nanosorbent

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3 For a synthesis of the Cd(II) nano IIP, by a typical polymerization reaction, a two-necked
4 glass reactor was equipped with a condenser, a magnetic stirrer and a gas inlet to maintain a
5 nitrogen atmosphere. One mmol of the complex and 1.0 g of vinyl functionalized Fe₃O₄ were
6 dispersed in 100 mL of methanol, then, the mixture temperature was raised to 70 °C and
7 afterwards 0.08 g of AIBN and 1.1 mL of EGDMA were added. After 48 hours the nano IIP
8 was separated by a magnet, and template (cadmium ions) was removed by a solution
9 containing 1 mol L⁻¹ HCl.
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21 In order to confirm the removal of Cd(II) ions, the small amount of magnetic polymer was
22 treated by piranha solution, then Cd(II) ions was determined by FAAS. Piranha solution,
23 containing concentrated H₂SO₄ and 30% solution of H₂O₂ (3:1 v/v), dissolves the polymer
24 network as well as Fe₃O₄ NPs and releases metallic ions in solution, which can be
25 determined by FAAS [39]. It's important to remark that this process was performed just on
26 20 mg of polymer as a trial experiment to confirm that an efficient removing of Cd (II) ions
27 template by acidic elution was accomplished. Therefore, the results showed that eluting by
28 HCl solution is sufficient for Cd(II) removal. The formation of this IIP was confirmed by IR
29 spectroscopy, TG/DTA analysis, XRD and SEM. A schematic diagram of magnetic IIP is
30 shown in Fig. 1.
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45 **2.4. Extraction procedure**

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48 The extraction procedure consists of two steps: in the sorption step, pH of sample solution
49 was adjusted at 7.0, then, 20 mg of magnetic IIP was suspended in the aqueous solution
50 containing 0.5 mg L⁻¹ of Cd(II) and stirred for 5.0 min. After washing with 5 mL distilled
51 water, in the next step elution of adsorbed Cd(II) ions was performed with 4.0 mL of 2 mol
52 L⁻¹ HCl solution for five minutes. Concentration of cadmium ions in the eluent were
53 evaluated by FAAS.
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2.5. Real samples

The urine samples were obtained from workers employed in one of automobile industries (Tehran, Iran). The samples were collected in sterile sample containers and were kept in the refrigerator (4°C). It is worth mentioning that, all experiments were performed in compliance with the relevant laws and institutional guidelines, and also the institutional committee has approved the experiments. Digestion of urine certified reference material (Seronorm LOT NO 2525), powder was performed in a reaction vessel containing 2 mL of HNO₃ (1% v/v) and 2 mL of H₂O₂ (30% v/v) and was placed in an ultrasonic bath for 30 min. Then the solution was diluted to 5 mL using deionized water [41]. To determine Cd(II) content in urine samples, 20 mg of prepared sorbent was weighted and suspended in 20 mL of urine. Then, by using a standard addition method (SAM), Cd(II) concentrations were determined in each sample under the optimized condition by FAAS.

3. Results and discussion

3.1. Sorbent characterization

Formation of Fe₃O₄ NPs has been assessed by IR spectroscopy, SEM and XRD analysis. Modification of Fe₃O₄ NPs surface with vinyl groups was carried out through the direct method for functionalization of Fe₃O₄ with triethoxysilan agents reported before [8]. The reaction of vinyl functionalized Fe₃O₄ with vinylated ligand complex as another monomer in the presence of Cd(II) as a template, AIBN as an initiator and EGDMA as the cross linker cause the formation of this magnetic IIP (Fig. 1).

The synthesized imprinted polymer was characterized by IR spectroscopy, SEM, and TG/DTA analysis. The FT-IR spectra of IIP showed following bands: C=S (1145 to 1149 cm⁻¹), C=N (1629 to 1632 cm⁻¹) and N-H (3415 to 3437 cm⁻¹) as shown in Fig. 1S. The thermal

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3 stability of this nano IIP was evaluated by TG/DTA analysis (Fig. 2S). The results showed
4 that, Fe₃O₄@Cd-IIP may be stable at high temperatures, as it was stable up to 300 °C.
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6 Nevertheless, according to the loss of 20 % of the composite weight, it can be concluded that
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8 the composite was burned causing 20% loss of its weight. This reduction was belonging to
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10 the polymer and residue was contributed to Fe₃O₄. In addition, the size and morphology
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12 assessment of synthesized sorbent were confirmed by SEM (Fig. 2-a,b). By investigating
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14 these micrographs, it was becoming apparent that the sorbent consists of spherical NPs with
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16 approximately 40 nm in diameter. It is worth mentioning that, since the magnetic NPs are
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18 coated with polymer, they will not be destroyed in acidic solutions [9].
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25 According to the XRD pattern, the Fe₃O₄ structure remained unchanged after polymerization;
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27 it means that, the Fe₃O₄ did not decompose or convert to Fe₂O₃. Also, according to the Fig. 2-
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31 nanosized scale. The average crystallite size of magnetic IIP was estimated from the XRD
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33 pattern using Scherrer formula:
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$$D = \frac{k\lambda}{\beta \cos \theta}$$

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38 Where D is the average crystallite size, λ is the X-ray wavelength, β is the full-width at half
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40 maximum (FWHM) and θ is the diffraction angle. Here K = 0.9 is for spherical shape. So the
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42 crystallite size of magnetic IIP was computed from XRD pattern and found to be about 28
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44 nm. The XRD pattern of Fe₃O₄@IIP showed that, length of peaks reduced compared Fe₃O₄
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46 XRD pattern. It can be concluded that, Fe₃O₄ nano particles have been coated with polymer
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58 3.2. Optimization of the preconcentration method

59 3.2.1. Effect of sample pH

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3 The pH factor has ability to change the coordination sites of the ligand, so, it will be one of
4 the most controlling parameters for sorption of Cd(II) on synthesized polymer. In order to
5 explore the effect of this parameter the pH values were adjusted in the range of 3.5-8.0. After
6 stirring for 5.0 min, the adsorbed Cd(II) ions were eluted by 4.0 mL of 2 mol L⁻¹ HCl,
7 subsequently, the amount of Cd(II) ions in the eluent was determined by FAAS (Fig. 3S).
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9 Optimized sample pH for adsorption of Cd(II) onto magnetic IIP was 7.0. At low pH values
10 there is an excessive protonation of the lone pair of electrons on nitrogen and sulfur, resulting
11 in a decrease in the Cd(II) ions sorption. Moreover, in the alkaline pHs, the Cd(II) precipitate
12 as Cd(OH)₂ causing a decrease in sorption efficiency.
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25 **3.2.2. Effect of eluent type, volume and its concentration**

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27 In order to elute Cd(II) from IIP, HCl was chosen among H₂SO₄, HNO₃ and HCl. The
28 inspection of the effect of eluent volume on the recovery of Cd(II) showed that, 4.0 mL of the
29 2 mol L⁻¹ HCl was the optimum volume for elution of Cd(II) ions. Interestingly, at larger
30 volumes, the recoveries were not significantly raised by increasing the eluent volume.
31 Therefore, to obtain higher preconcentration factors, 4.0 mL of 2 mol L⁻¹ HCl was the
32 optimum eluent for extraction of cadmium.
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44 **3.2.3. Optimization of sorption and elution time**

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46 Effect of time is one of the important factors in the analytical process while the inadequate
47 time reduces the procedure efficiently and extra times cause lowering throughput. In order to
48 investigate the effect of time, 25 mL of solutions containing 0.5 mg L⁻¹ of Cd(II) was
49 adjusted to pH = 7.0 and 20 mg of magnetic IIP was suspended to the solutions and stirred for
50 various durations and at the end of this process was washed with 5.0 mL distilled water.
51 Then, the sorbent was separated by placing a magnet and the pre-concentrated analyte was
52 determined by FAAS after eluting by 2 mol L⁻¹ HCl. According to the Fig. 3, both optimized
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sorption and elution time were selected as 5.0 min. This fast extraction and elution duration could be attributed to the high surface area dealing with these IIP NPs. Compare to column technique, the magnetic method takes shorter sorption and desorption times [14, 17-18].

3.2.4. Influence of potentially interfering ions

In various matrices even biological ones, there are unknown potentially interfering agents. In order to extract Cd(II), several cations may cause decreasing of recovery. So, the method of extraction must be selective enough to measure an accurate value of Cd(II) even in complex matrices. In order to investigate the selectivity of the sorbent, the effect of some cations was assessed. For this purpose, 0.5 mg L⁻¹ of Cd(II) ion solutions contained, individually the interfering ions, Na⁺, Pb⁺², Zn²⁺, and Cu²⁺ were analyzed under the optimum condition. Also the X ions concentration was 2000 times of the Cd(II) ions (Table 1S). The tolerable amount was defined as the maximum concentration that could cause a change of less than 5% in signal compared to the signal of Cd(II) ions without any interference [9]. As shown in Table 1S, none of these ions do not interfere with the concentrations encountered in biological samples, and the method is selective toward Cd(II) extraction at pH = 7.0. This high selectivity could be attributed to very selective sites, the size and shape of which are fitted to the Cd(II) ions specifically.

It is worth mentioning that; Although IIP can be considered as an SPE procedure, based on the results obtained in this study, IIP is promising to be more selective. The phase can be synthesized in a short time followed by a simple protocol for extraction stage, making method to be more convenience compare to the usual silica based SPE technique.

3.3.1. Analytical figures of merit

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In order to prove the precision and accuracy of the present technique, reproducibility of the optimized method was validated for day-to-day and within-day reproducibility using spiked aqueous samples. 25 mL samples were used at low, medium, and high concentrations of 2, 10, 50 $\mu\text{g L}^{-1}$ respectively. A linear standard curve was obtained for extracting samples over the range concentrations every day for 6 consecutive days ($n = 6$) with the correlation coefficient of 0.997 or greater. From within-day experiments, evaluation, six experiments were performed each day for three consecutive days. The extraction procedure was reliable and reproducible from day-to-day and within-day. Coefficient of variations (CV%) of 0.092, 0.064, and 0.017 were obtained for 2, 10, and 50 $\mu\text{g L}^{-1}$ respectively, for day-to-day and 0.146, 0.058, and 0.014 at the same concentrations, respectively for within-day, showing appropriate accuracy and precision for the optimized method (Table 2S).

It is worth mentioning that cadmium in samples was determined using standard addition method and a linear curve was plotted ($R^2 > 0.98$).

3.3.2. Method validation

In order to establish the accuracy of the current method the concentration of Cd(II) ions was compared to the exact concentration of this ion in the certified reference material (Seronorm LOT NO2525) and the result was in good agreement with the certified value (Table 1).

3.3.3. Determination of urinary cadmium

The optimized method was applied for the determination of Cd(II) ion concentration in urine samples collected from the male workers from one of Iranian automobile industries whom supposed to be exposed to Cd(II) through welding and painting process. In this regard, after sample collection and shipping to the analytical laboratory, each sample divided into 3 parts. Then Cd(II) values were determined in each sample under optimized condition by using a standard addition technique (Table 2).

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3 According to the available researches, analytical performance of those methods which were
4 used specific instrument like graphite furnace atomic absorption spectrometry (GFAAS) or
5 electrothermal atomic absorption spectrometry (ETAAS) are better than FAAS; 0.09, 0.045
6 and 5.75 $\mu\text{g L}^{-1}$ respectively. Utilization of preconcentration techniques can cause increasing
7 the performance of analytical method even has done by FAAS.

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10 This method already had not been used for extraction of metallic ions through urine samples.
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12 Furthermore, according to the ATSDR profile of Cd(II), the lowest LOD for determination of
13 Cd(II) by FAAS method was 5.67 $\mu\text{g L}^{-1}$ [42], in which, obtained LOD of the present study
14 was 0.6 $\mu\text{g L}^{-1}$ (Table 3). Excellent sensitivity of magnetic IIP extraction allows researchers to
15 determine trace concentration of Cd(II) in urine samples. Also, there is no need to use
16 instruments like Graphite furnace atomic absorption Spectrometry (GFAAS) or
17 Electrothermal Atomic Absorption Spectrometry (ETAAS) in similar studies. In the other
18 hand measuring of Cd(II) by $\text{Fe}_3\text{O}_4\text{@IIP}$ needs no pretreatment like acidic digestion,
19 filtration, or centrifuge.

20 21 22 23 24 25 26 27 28 29 30 31 32 33 34 35 36 37 38 **4. Conclusion**

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41 A new magnetic ion imprinted (IIP) polymer has been synthesized by the coating an IIP on
42 the surface of Fe_3O_4 nanoparticles to achieve highest surface area for extraction aims. The
43 selectivity of this ion imprinted polymer has made it to be a suitable and selective sorbent for
44 the extraction and preconcentration of cadmium ions in biological samples. In order to
45 achieve the best performance by least expense optimum parameters achieved, i.e. pH = 7.0,
46 sorption and elution time 5.0 min, eluent volume 4.0 mL (2 mol L^{-1} HCl). The high
47 selectivity of this magnetic IIP can determine Cd(II) in biological samples without the need
48 of unintelligible instrument like GFAAS or ETAAS, causing reduced expenses as well as
49 saving time. Other advantages of this method are: low time-consuming due to the
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magnetically-assisted separation of the sorbent and high surface area; therefore, satisfactory results can be achieved using fewer amounts of the sorbent. On the other hand, according to day-to-day and whiten-day reproducible experiments, this technique gain an acceptable reliability degree for analyzing biological samples.

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Table 1

Determination of Cd(II) ions in the certified reference material.

<u>Sample</u>	<u>Concentration ($\mu\text{g L}^{-1}$)</u>		<u>Relative error</u>
	<u>Certified</u>	<u>Found</u>	<u>%</u>
<u>Seronorm LOT NO2525</u>	<u>5.06</u>	<u>5.23</u>	<u>3.4</u>

Table 2

Measurement of Cd(II) in urine samples by using magnetic IIP.

Sample	Age	Smoking habit	Job	Concentration of urine ($\mu\text{g L}^{-1}$)
1	32	no	painting	84.0
2	30	no	painting	54.0
3	31	no	painting	29.2
4	32	yes	painting	72.3
5	30	yes	painting	15.0
6	31	no	painting	20.0
7	31	no	Welding	147
8	27	no	Welding	21.3
9	33	yes	Welding	142
10	42	no	Welding	52.0
11	26	no	Welding	28.0

Table 3

Comparative LOD of various methods for determination of Cd(II) in urine samples.

Type of samples	Preparation method	Analytical method	LOD ($\mu\text{g L}^{-1}$)	Ref.
Urine	Modification of matrix with diammonium hydrogen phosphate/nitric acid	GFAAS	0.09	[42]
Urine	Digestion with nitric acid	AAS	5.67	[42]
Urine	Dilution with nitric acid	ETAAS	0.045	[42]
Biological	$\text{Fe}_3\text{O}_4@\text{SiO}_2$ Cd^{2+}	ETAAS	0.19	[17]
Urine	$\text{Fe}_3\text{O}_4@\text{IIP}$ Cd^{2+}	FAAS	0.6	This study

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3 **Figure captions:**
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5 **Fig. 1:** A schematic diagram for synthesis process of magnetic ion imprinted polymer of
6 cadmium.
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10 **Fig. 2:** 1 μm (a) and 10 μm (b) scaled SEM micrographs of synthesized magnetic IIP; (c) XRD
11 pattern of magnetic IIP.
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14 **Fig. 3:** Effect of sorption and elution times on the recovery of Cd(II), conditions: sample pH, 7.0;
15 eluent volume 4.0 mL (2 mol L^{-1} HCl).
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Fig. 1

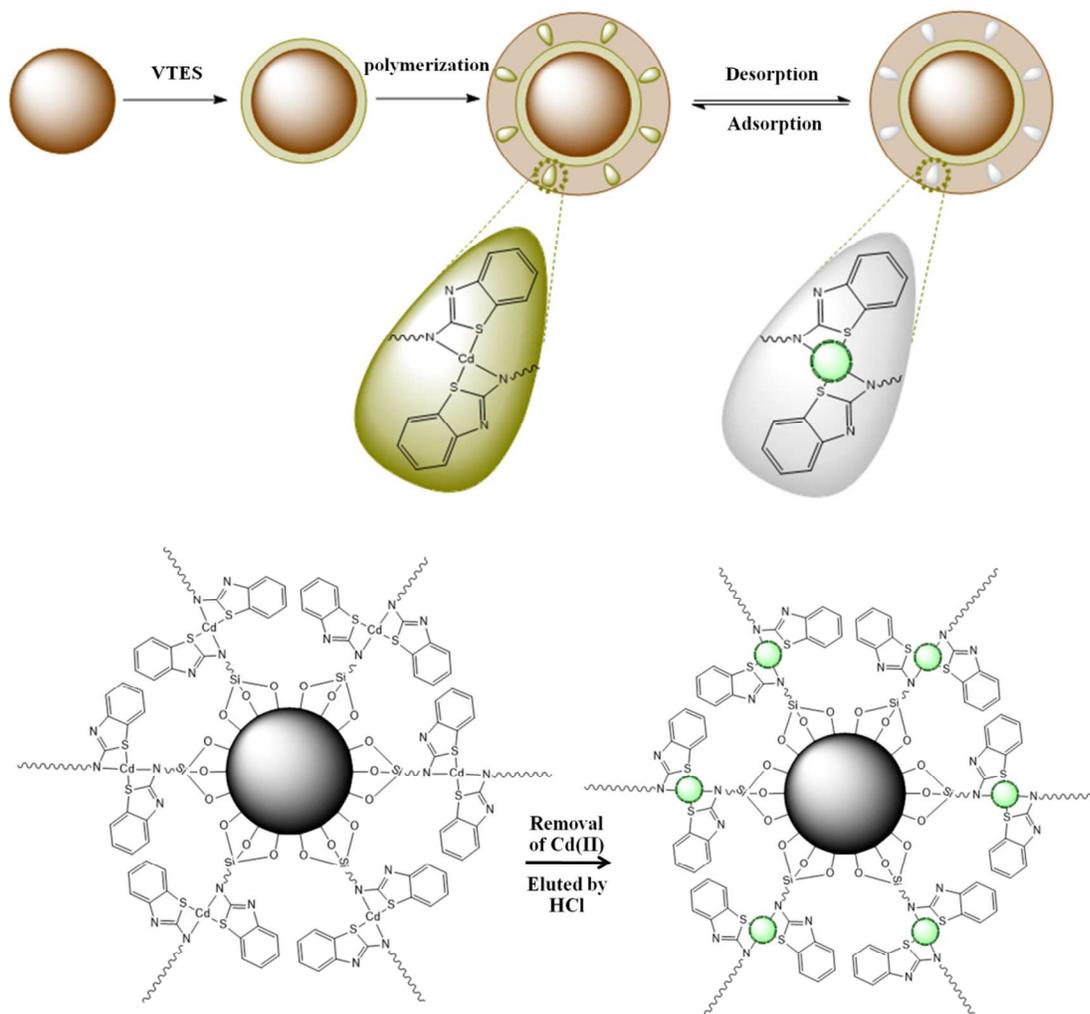


Fig. 2

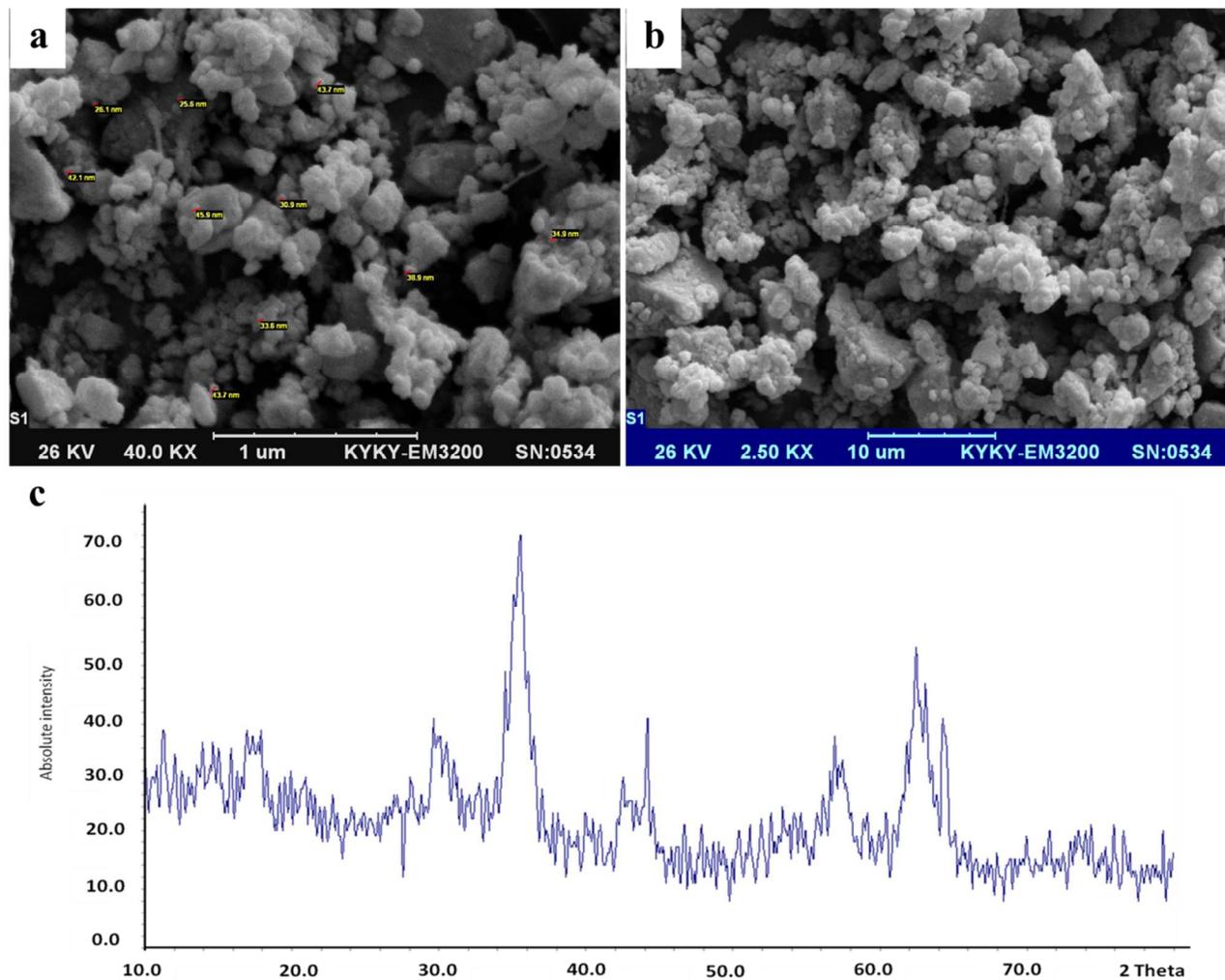


Fig. 3

