ChemComm

Accepted Manuscript



This is an *Accepted Manuscript*, which has been through the Royal Society of Chemistry peer review process and has been accepted for publication.

Accepted Manuscripts are published online shortly after acceptance, before technical editing, formatting and proof reading. Using this free service, authors can make their results available to the community, in citable form, before we publish the edited article. We will replace this Accepted Manuscript with the edited and formatted Advance Article as soon as it is available.

You can find more information about *Accepted Manuscripts* in the **Information for Authors**.

Please note that technical editing may introduce minor changes to the text and/or graphics, which may alter content. The journal's standard <u>Terms & Conditions</u> and the <u>Ethical guidelines</u> still apply. In no event shall the Royal Society of Chemistry be held responsible for any errors or omissions in this *Accepted Manuscript* or any consequences arising from the use of any information it contains.



Cite this: DOI: 10.1039/c0xx00000x

www.rsc.org/xxxxxx

ARTICLE TYPE

Novel and selective detection of Tabun mimics

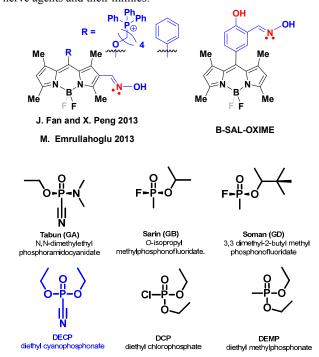
Yoon Jeong Jang, Olga Tsay, Dhiraj P. Murale, Jeong A Jeong, Aviv Segev, and David G. Churchill*a

Received (in XXX, XXX) Xth XXXXXXXXX 20XX, Accepted Xth XXXXXXXX 20XX 5 OI: 10.1039/b000000x

Detection of nerve agent-related molecules based on BODIPY-salicylaldehyde oxime congugation was studied. Fluorescence intensity of the B-SAL-OXIME species 10 increases in the presence of DECP, whereas it decreases in the presence of DCP and DEMP (Limit of detection = 997 nM). Benzonitrile formation in the novel Fluorescent B-SAL-**OXIME** system was elucidated using model substrates.

Organophosphorus chemicals include nerve agents and pesticides and are considered as being among the most toxic chemicals to living things.1 Nerve agents were developed to effectively harm people and decrease military power of an opposing force.² Historically, nerve agents have been used during 20 certain conflicts and/or have been stockpiled. The Chemical Weapons Convention (CWC) has placed a ban on synthesizing, stockpiling and deploying chemical warfare agents (CWAs) to prevent further casualties and reduce global dangers; however, recent news of stockpiles and utilization of nerve agents in the 25 Syrian Arab Republic (Ghouta) showed there remains a clear and present danger in the World regarding nerve agents.³ The toxic mechanism is phosphorylation and phosphonylation of the active serine-OH residue in synapses. Nerve agents bind covalently into the acetylcholinesterase active site and block the breakdown of 30 acetylcholine in synapses. It causes an increase in acetylcholine concentration and results in a neurological imbalance in the synapse.4 Tabun is a unique agent because of its nitrile group. Tabun includes a P-CN group, while other nerve agents of the Gseries possess a P-F group. It has been suggested that this 35 chemical group may reveal unique chemistry in chemosensing, but approaches for detection of Tabun (GA) selectively over other nerve agents are still scarce or non-existent. Recently, many methods of detecting nerve agents continue to be developed and optimized, which include ion mobility spectrometry, mass 40 spectrometry, NMR spectroscopy, and enzyme sensors. 5 These methods provide good sensitivity, but do not afford convenient access to appropriate selectivity and/or are not convenient and simplified real-time methods for the "field." Fluorescent and

45 widely used for detecting nerve agents because they are very sensitive, convenient, allow for detection in real-time and are easy to assess by the unassisted eye. Oximes (R1R2C=NOH) include aldoximes and ketoximes in which R1 is a hydrogen or another organic group.⁷ Aldoximes have been used for the 50 treatment of nerve agents. 8 An oxime is a "super-nucleophile" and is capable of attacking the internal phosphorus at the phosphorylated serine-OH in AChE to restore the serine-OH, and thus the function of AChE. 9 BODIPY species bearing oximes have recently been explored in chemosensing (reactive oxygen ss species). 10 Conjugates of (a) BODIPY(4,4-difluoro-4-bora-3a,4a-diaza-s-indacene), a well-known fluorophore class, and (b) the salicylaldehyde core, very widely used in transition metal ligand synthetic designs, have recently been prepared. Bodipy derivatives enable high quantum yield reporting and possess 60 robust chemical properties and photostability and tuanable solubility for use in bioimaging and chemosensing, 11,12 Herein, we extend efforts using this conjugation for selective detection of nerve agents and their mimics.



chromogenic sensing methods are becoming convenient and

Figure 1. Structures of Probes and G-series chemical warfare nerve agents and simulants (blue color–DECP–Tabun mimic). ¹

In the present study, a new oxime-based Bodipy system (B-SAL-OXIME) was synthesized and used as a fluorescent 5 sensor for the sensing and detection of nerve agent simulants, (diethyl **DCP** chlorophosphate), **DEMP** (diethyl methylphosphonate), and DECP (diethyl cyanophosphonate) (Fig. 1). Fluorescence emission changes for B-SAL-OXIME (compound 6) with 0.1 M of DCP, DEMP, and DECP were ₁₀ studied; solutions were made by 1×10^{-6} M in 0.1 mM, pH 7.4 HEPES buffer and 100 to 1200×10^{-6} M of DCP, DEMP and 2 × 10^{-7} M in 0.1 mM, pH 7.4 HEPES buffer and 100 to 1200×10^{-6} M of DECP. Fluorescence emission for B-SAL-OXIME with DCP and DEMP decreased (Fig. 2 and S15a, b), but with DECP, 15 intensity increased at $\lambda_{exic} = 499$ nm, $\lambda_{emis} = 508$ nm (Fig. 2a). Possible mechanisms of B-SAL-OXIME with DCP, DEMP and DECP were proposed and relate to standard nucleophilic attack pathways. Cl⁻ was a leaving group, departing from DCP, which together with H⁺ from R=N-OH, helps form R=N-O-P(O)(OEt)₂ 20 (Scheme 1) causing the decrease in fluorescence intensity (Scheme 1).¹³ The ethoxy group played the role of the leaving group from DEMP to give R=N-O-P(O)(OEt)Me (Scheme 1) which causes a decrease in fluorescence intensity.14 The fluorescence intensity of B-SAL-OXIME with DECP increases 25 when also in the presence of appreciable concentrations of DCP and DEMP. Here, cyanide (CN-) was the leaving group to give R=N-O-P(O)(Et)₂. Compound 7 for this reaction could not be found by HRMS or NMR spectra (¹H and ³¹P).

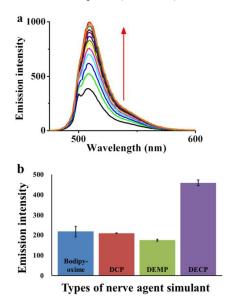


Figure 2. (a) Emission titration spectra of B-SAL-OXIME (2 \times 10^{-7} M in 0.1 mM, pH 7.4 HEPES buffer) with DECP (100 to 1200 μ M in acetonitrile) at λ_{exic} = 499 nm, λ_{emis} = 508 nm (b) 35 Fluorescence intensity comparing bar graph among B-SAL-OXIME (1 \times 10⁻⁶ M in 0.1 mM, pH 7.4 HEPES buffer) with 1200 μ M DCP, DEMP and DECP in acetonitrile at $\lambda_{exic} = 499$ nm, $\lambda_{emis} = 508 \text{ nm}.$

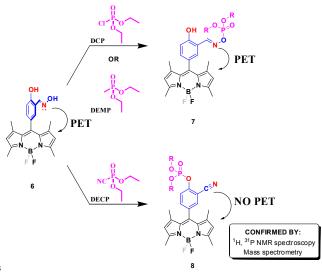
40 The detection limit of B-SAL-OXIME for DECP was determined to be 92.2 μM for a linear fit; but, in the case of nonlinear methods, the fitting that involving a lower LOD value also possesses a higher R^2 value (997 nM and $R^2 = 0.99$) was obtained (see Fig. S16 for comparative fittings). 15 This LOD (linear fit) 45 value is not extraordinarily low. Therefore, as part of our future aims we will continue to develop new systems that possess novel modalities, or enhance newly discovered methods to tune the sensitivity: e.g., conjugates with GNPs (Gold Nanoparticles). 15-16 B-SAL-OXIME however does have enough sensitivity in the 50 detection of nerve agent stimulants, considering the respective, relevant LD₅₀ values (Table. S1).

Chemicals	Structure	³¹ P NMR Shift (ppm)	
		DECP	DCP
Solvent (CDCl ₃)		- 20.5	5.0
Phenol	OH	- 5.7	- 6.0
Benzaldehyde oxime	N OH←	- 0.1	- 0.5
Salicylaldehyde oxime	OH OH←	0.02	0.7
Bodipy-oxime	OH OH	- 7.1	0.3

Table 1. ³¹P NMR spectra results for B-SAL-OXIME, phenol, 55 benzaldehyde oxime and salicyladehyde oxime with nerve agent simulants DECP and DCP.

To support the proposed pathway(s) (Scheme 1), a mechanistic study was conducted through the use of model 60 benzene derivatives bearing the same substituents found in salicylaldehyde-oxime. Phenol, benzaldehyde oxime and salicylaldehyde oxime, underwent respective reactions with DCP and DECP under basic conditions (triethylamine) in acetonitrile (Table 1, Fig. S2); ¹H NMR and ³¹P NMR spectra were studied 65 after purification. While kinetic differences exist between the model system and the actual probe, the model systems importantly help clarify the reactivity. ³¹P NMR spectra results of DECP and DCP gave a singlet (δ -20.5 and 5.0, resp.) with shifting to $\delta - 5.7$ and -6.0, respectively, in the presence of 70 phenol (Fig. S12). This data supports the phenol, with DECP and DCP gives Ph-O-P(O)(OEt)₂ functionality through nucleophilic substitution. A singlet in the ³¹P NMR spectrum was found to be -0.1 and -0.5 in the presence of benzaldehyde oxime (Fig. S13), and at 0.02 and 0.7 in the presence of salicylaldehyde oxime (Fig. 75 S14) upon treatment with DECP and DCP, respectively (Table 1). The ³¹P NMR spectrum of B-SAL-OXIME with DECP reveals a singlet (δ –7.1), supporting that DECP is pheonolate-bound in B– SAL-OXIME, and not bound to the R=NOH group. The model study with salicylaldehyde oxime revealed dehydration occurs to 80 give the 2-hydroxyl-benzonitrile with DECP and DCP; ³¹P NMR

and ¹H NMR spectroscopy confirm this reaction (Fig. S14). ¹⁷ The dehydration of the oxime to the nitrile occurs with DECP, in B-SAL-OXIME; the OH group in B-SAL-OXIME was also attacked by DECP wherein mass spectroscopy helps confirm this 5 mechanism. The mass spectrum of B-SAL-OXIME with DECP was observed at M/Z 524.1621; compound 8 formulated as $[C_{28}H_{30}O_{11}P_2Na]^+$ gives a calculated value of 524.1693 (Fig. S7). We believe that the nitrogen of the oxime is the electron donating group and PET mechanism gives no strong signalling 10 for compound 6 and 7. However, the cyano group in compound 8 works as an electron-withdrawing group and allows for strong fluorescence by inhibiting PET between the donor-acceptor units of the dyad (Scheme 1).



Scheme 1. Proposed mechanism of B-SAL-OXIME with DCP, DEMP, and DECP.

To assist in recognition, a logic gating treatment¹⁸ was invoked 20 where data was interpreted in blocks of emission intensity; these help form exclusive logic gate tiers of increasing intensity (Figure 3). Intensity for **A** is 0 to 50 nm, for **B** 50 to 100 nm, for **C** 100 to 150 nm, for **D** 150 to 200, for **E** 200 to 250 nm, and for **F** 250 to 500 nm. Each emission intensity block can be identified by a 25 combination of levels of B-SAL-OXIME from DCP, DEMP, and DECP according to fluorescence intensity, using "AND," "OR," and "NOT" logic gates. For the 250-500 nm region, the concentration, DECP is 700 µM or greater with two different levels of fluorescence intensity. In the regions of B, C and D, the 30 concentration of DCP may be zero; for the E region, none of the three agents may be zero. The high intensity gate for the 250-500 nm region (F) is a three-input gate based on levels of DECP and no DCP and DEMP; the 100-250 nm zone is a four-input gate; and the 0-50 nm gate is a six-input gate with DCP, DEMP, and 35 DECP, each with two different options of fluorescence intensity for 900 µM or lower.

Fluorescence change monitoring for compound 8 with metal ions (Ag⁺, Ca²⁺, Cd⁺, Co²⁺, Cu²⁺, Fe²⁺, Fe³⁺, K⁺, Hg²⁺,

40 Mg²⁺, Mn²⁺, Na⁺, Pb²⁺, Zn²⁺) shows no interference exists except for Ag⁺ at concentrations equimolar to that of the organophosphonate species. A strong quenching event was found (99.7%, probe: 1×10^{-6} M in 0.1 mM, pH 7.4 HEPES buffer λ_{exic} = 499 nm, λ_{emis} = 508 nm, acetonitrile). Other trials including B-45 SAL-OXIME and Ag⁺ or probe with DCP and Ag⁺ show no change (Figures S16, S17).

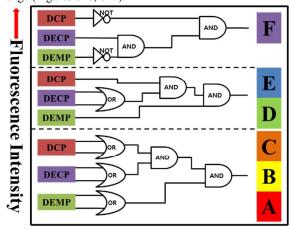


Figure 3. Logic gate construct for B-SAL-OXIME with DCP, 50 DEMP and DECP according to fluorescence intensity.

In conclusion, herein we introduce a novel B-SAL-OXIME probe for detecting chemical warfare nerve agent simulants. In the most straight-forward manifestation, it can be 55 implemented as a fluorescent detection medium for the detection of DECP over DCP and DEMP. Fluorescence intensity of B-SAL-OXIME increased with DECP selectively, and decreased with DCP and DEMP concentration. Models were treated with DECP and DCP and monitored by ¹H NMR and ³¹P NMR 60 spectroscopy to help interpret spectra obtained after the reaction of the B-SAL-OXIME probe with simulants. Through these model studies, B-SAL-OXIME was found to be dehydrated to the nitrile and the OH bonds to DECP giving loss of HCl.

65 Acknowledgement: D.G.C. acknowledges research support from the National Research Foundation (NRF) (Grant # 2011-0017280). Mr. Hack Soo and Ms. Sung A Kim are acknowledged respectively for facilitating the acquisition of NMR spectroscopic and MS data.

70 Notes and references

- ^a Molecular Logic Gate Laboratory, Department of Chemistry, Korea Advanced Institute of Science and Technology (KAIST), 291 Daehak-ro, Yuseong-gu, Daejeon, 305-701, Republic of Korea. Fax: (+82)42-350-2810; Tel: (+82)42-350-2845; E-mail: dchurchill@kaist.ac.kr
- 75 Department of Knowledge Service Engineering Korea Advanced Institute of Science and Technology (KAIST). 291 Daehak-ro, Yuseonggu, Daejeon, 305-701, Republic of Korea. Fax: (+82)42-350-1610; Tel: (+82)42-350-1614; E-mail: aviv@kaist.edu
- 80 1. K. Kim, O. G. Tsay, D. A. Atwood and D. G. Churchill, Chem. Rev., 2011, 111, 5345.

- (a) T. Okumura, N. Takasu, S. Ishimatsu, S. Miyanoki, A. Mitsuhashi, K. Kumada, K. Tanaka and S. Hinohara, *Ann. Emergency Med.*, 1996, 28, 129; (b) H. Okudera, H. Morita, T. Iwashita, T. Shibata, T. Otagiri, S. Kobayashi and N. Yanagisawa, *Am. J. Emerg. Med.*, 1997, 15, 527.
- 3 (a) S. W. W. a. R. S. Hoffman, *J. Intensive Care Med.*, 2004, 19; (b) I. R. Kenyon, K. Gutschmidt and O. Cosivi, *Toxicology*, 2005, 214, 249; (c) C. E. H. Christopher M. Blanchard, and Mary Beth D. Nikitin, Congressional Research Service, 2014.
- W.-h. Wu, J.-j. Dong, X. Wang, J. Li, S.-h. Sui, G.-y. Chen, J.-w. Liu and M. Zhang, *Analyst*, 2012, 137, 3224.
- 5. (a) G. R. Asbury, C. Wu, W. F. Siems and H. H. Hill, *Anal. Chim. Acta*, 2000, **404**, 273; (b) T. J. Henderson, *Anal. Chem.*, 2002, **74**, 191; (c) R. M. Black, R. J. Clarke, R. W. Read and M. T. J. Reid, *J. Chromatogr. A*, 1994, **662**, 301; (d) A. Mulchandani, I. Kaneva and W. Chen, *Biotechnol. Bioeng.*, 1999, **63**, 216; (e) S. Royo, R. Martinez-Manez, F. Sancenon, A. M. Costero, M. Parra and S. Gil, *Chem. Commun.*, 2007,
- (a) I. Walton, M. Davis, L. Munro, V. J. Catalano, P. J. Cragg, M. T. Huggins and K. J. Wallace, *Org. Lett.*, 2012, 14, 2686;
 (b) X. Wu, Z. Wu and S. Han, *Chem. Commun.*, 2011, 47, 11468;
 (c) B. Díaz de Greñu, D. Moreno, T. Torroba, A. Berg, J. Gunnars, T. Nilsson, R. Nyman, M. Persson, J. Pettersson, I.
- Eklind and P. Wästerby, *J. Am. Chem. Soc.*, 2014, **136**, 4125; (d) M. Burnworth, S. J. Rowan and C. Weder, *Chem. Eur. J.*, 2007, **13**, 7828; (e) A. Wild, A. Winter, M. D. Hager and U. S. Schubert, *Chem. Commun.*, 2012, **48**, 964; (f) D. Ajami and J. Rebek, Jr., *Org. Biomol. Chem.*, 2013, **11**, 3936.
- J. Kassa, J. Z. Karasova, V. Sepsova, G. Kunesova, F. Caisberger, M. Pohanka and J. Bajgar, *Toxicol. Lett.*, 2011, 205, S128.
- 8. (a) F. Worek, H. Thiermann, L. Szinicz and P. Eyer, *Biochem. Pharmacol.*, 2004, **68**, 2237; (b) J. Kassa, *J. Toxicol. Clin. Toxicol.*, 2002, **40**, 803.
- 9. (a) F. Worek, P. Eyer, N. Aurbek, L. Szinicz and H. Thiermann, *Toxicol. Appl. Pharmacol.*, 2007, **219**, 226; (b) O. G. Tsay, S. T. Manjare, H. Kim, K. M. Lee, Y. S. Lee and D. G. Churchill, *Inorg. Chem.*, 2013, **52**, 10052.
- (a) M. Emrullahoglu, M. Ucuncu and E. Karakus, *Chem. Commun.*, 2013, 49, 7836;
 (b) G. Cheng, J. Fan, W. Sun, K. Sui, X. Jin, J. Wang and X. Peng, *Analyst*, 2013, 138, 6091.
- 11. G. Ulrich, R. Ziessel and A. Harriman, *Angew. Chem. Int. Ed.*, 2008, **47**, 1184.
- C. Thivierge, J. Han, R. M. Jenkins and K. Burgess, J. Org. Chem., 2011, 76, 5219.
- I. Damljanovic, M. Vukicevic and R. D. Vukicevic, Monatsh. Chem., 2006, 137, 301.
- 50 14. J. J. Topczewski and D. M. Quinn, Org. Lett., 2013, 15, 1084.
 - 15. A. Hakonen and N. Stromberg, Analyst, 2012, 137, 315.
 - (a) Y. J. Jang, D. P. Murale and D. G. Churchill, *Analyst*,
 2014, 139, 1614; (b) A. Hakonen, *Anal. Chem.*, 2009, 81,
- 4555; (c) A. Hakonen and N. Stromberg, *Chem. Commun.*, 2011, 47, 3433; (d) N. Stromberg and A. Hakonen, *Anal. Chim. Acta*, 2011, 704, 139.
- 17. A. R. Sardarian, Z. Shahsavari-Fard, H. R. Shahsavari and Z. Ebrahimi, *Tetrahedron Lett.*, 2007, **48**, 2639.
- 60 18. A. P. de Silva, Chemistry-an Asian Journal, 2011, 6, 750.