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COMMUNICATION

Non-covalent double functionalization of carbon nanotubes with a NADH oxidation Ru(II)-based molecular catalyst and a NAD-dependent glucose dehydrogenase

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We report the double functionalization of multiwalled carbon nanotube electrodes by two functional pyrene molecules. In combination, immobilized Ru(II)-based NADH oxidation catalyst and glucose dehydrogenase achieves highly efficient glucose oxidation with low overpotential of -0.10 V and high current densities of 6 mA cm⁻².

NAD-dependent dehydrogenases are versatile enzymes, having the great advantage of oxidizing a wide range of relevant substrates such as glucose, lactate, or alcohols. These enzymes were thus intensively investigated in applications ranging from biosensing to biosynthesis or biofuel cells. In particular, dehydrogenase-based biofuel cells are envisioned for harvesting energy from oxidation of glucose¹, alcohols² or L-lactate³. Furthermore, their wide biodiversity makes these enzymes also promising for enzyme cascade system in which a biofuel can be fully oxidized by successive multi-enzymatic oxidation performed by immobilized dehydrogenases, using NADH/NAD as ubiquitous cofactor.² However, such process requires, an efficient electrochemical catalyst for the NADH oxidation in order to maximize anodic catalytic currents and minimize the NADH oxidation overpotential. Although the redox potential of NADH is -0.56V vs SCE, its oxidation often requires high overpotentials. In this context, organic dyes and metal complexes have demonstrated low overpotential oxidation for NADH⁴⁻⁶. In particular, we have recently demonstrated that a Ru(II) complex, previously developed by H. Abruna et al⁷ for low-potential oxidation of NADH, could be combined to carbon nanotubes for efficient oxidation of NADH, via the electrogeneration of a metallopolymer at the surface of carbon nanotubes (CNTs) sidewalls.⁸ CNTs are widely investigated as an electrode material for both electrocatalytic and bioelectrocatalytic applications. Their ability to immobilize high concentration of redox catalysts and to efficiently transfer electrons have made this nanomaterial a key material in

designing electrodes for molecular electrocatalysis^{9,10}. In addition, CNTs can be functionalized by a large variety of covalent or non-covalent techniques. We and others have especially developed the non-covalent functionalization of CNTs for immobilization of molecular electrocatalysts based on modified transition-metal complexes¹¹⁻¹³. Non-covalent functionalization is based on the strong pi-pi interactions between pyrene and CNT sidewalls. By similar means, we and other have also developed the immobilization of redox enzymes by pyrene derivatives previously pi-stacked on CNTs¹⁴⁻¹⁷. In addition, it was recently demonstrated that covalent or non-covalent multiple functionalization of carbon nanotube sidewalls could have promising applications in drug delivery^{18,19} and biosensing²⁰ applications. Here we report the first example of a double functionalization of Multi-Walled CNTs (MWCNTs) with a molecular catalyst and an enzyme for a combined electroenzymatic process. Namely, a glucose dehydrogenase (GDH) was chemically grafted on MWCNTS via a N-hydrosuccinimide-modified pyrene to assume the enzymatic oxidation of glucose in presence of NAD⁺. In addition, an original Ru(II) complex, bearing phenanthrolinequinone (phendione) and pyrene-modified bipyridine ligands was used for the electrocatalytic oxidation of NADH cofactor. With the aim to design new glucose biofuel cells, these bi-functionalized MWCNT electrodes were electrochemically characterized and investigated for the improved glucose oxidation.

An original [(1,10-phenanthroline-5,6-dione)₂((4,4'-bis(4-pyrenyl-1-ylbutyloxy)-2,2'-bipyridine)Ru(II))] hexafluorophosphate complex (**RuQ-pyrene**) was synthesized by refluxing [Ru(1,10-phenanthroline-5,6-dione)₂Cl₂] with 4,4'-bis(4-pyrenyl-1-ylbutyloxy)-2,2'-bipyridine in ethylene glycol. **RuQ-pyrene** was fully characterized by ¹H NMR, mass spectroscopy and UV-visible spectrophotometry. Its redox behaviour was characterized using cyclic voltammetry in 0.1M

TBAP/MeCN (figure 1). The complex exhibits one reversible peak system at $E_{1/2}^{ox} = 0.98\text{V}$ corresponding to the $\text{Ru}^{III/II}$ reversible redox system and an overlapping irreversible oxidation peak around $E_p^{ox} = 1\text{V}$ corresponding to the oxidation of the two pyrene groups. Two quasi-reversible peak systems at $E_{1/2}(Q^{\cdot-}/Q) = -0.50\text{V}$ and $E_{1/2}(Q^{2-}/Q^{\cdot-}) = -1.30\text{V}$ were attributed to the two successive one-electron reduction of the phenidone ligand. The one-electron reduction of the bipyridine ligand (bpy/bpy) leads to a reversible couple at -2.34V .

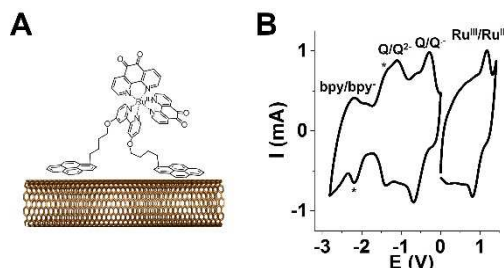


Figure 1. (A) Schematic representation of the preparation of MWCNT/RuQ-pyrene electrode. (B) Electrochemical characterisation of a RuQ-pyrene-functionalized MWCNT electrode by cyclic voltammetry in MeCN + 0.1M TBAP ($\nu = 100\text{ mV s}^{-1}$, starred peaks corresponds to charge trapping)

The establishment of pi-pi stacking interactions between pyrene moieties of the Ru complex and graphene planes of the MWCNT film affords its physisorption on MWCNT electrodes. A MWCNT electrode was simply incubated for 60 min in 0 to 8 mM RuQ-pyrene solution. After several washings, the modified MWCNT electrode was characterized by cyclic voltammetry in pure electrolyte. In MeCN, the RuQ-pyrene-functionalized MWCNT electrode displays the characteristic redox activity of immobilized RuQ-pyrene, accompanied with irreversible charge trapping peaks (starred peaks at -1.4 and -2.2V), often observed for immobilized redox species (figure 1B)²¹. In water, a reversible bielectronic peak system is observed at -0.02V , corresponding to the electroactivity of the phenidone redox couple QH_2/Q in water. Oxidation and reduction peak currents are highly stable upon scanning and are directly proportional to the scan rate. As expected, this electrochemical behaviour reflects the characteristics of firmly immobilized redox specie. Surface concentrations were estimated by integration of the charge under the anodic peak current. Figure 2A displays the variation of the surface coverage measured in water over the initial incubating RuQ-pyrene solution in DMF at 25°C .

The RuQ-pyrene surface coverage increases with the starting concentration in solution, following a simple Langmuir isotherm, according to equation 1.

$$\Gamma_{eq,Ru} = \frac{\Gamma_{max} \times K_{Ru} \times [\text{RuQ-pyrene}]}{1 + K_{Ru} \times [\text{RuQ-pyrene}]} \quad (1)$$

where $\Gamma_{eq,Ru}$ is the equilibrium surface coverage of RuQ-pyrene, Γ_{max} is the saturating surface coverage and K_{Ru} is the association constant of RuQ-pyrene with MWCNTs in DMF at 25°C . The best fit was achieved with a Γ_{max} of 17.8 nmol cm^{-2} and K_{Ru} of 1900 L mol^{-1} . It is noteworthy that we did not observe any influence of the incubation time on the RuQ-

pyrene surface coverage between one and 60 minutes, showing that the equilibrium of the pi-pi interactions is rapidly reached.

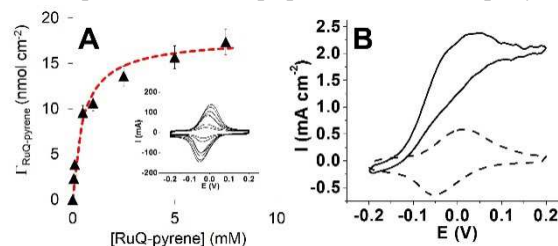


Figure 2. (A) Variation of the RuQ-pyrene surface coverage with incubating RuQ-pyrene concentrations in DMF accompanied with the fitting binding isotherm (red dashed line); (inset): Cyclic voltammetry of RuQ-pyrene-functionalized MWCNT electrodes performed in 0.2 M PBS, after incubation in different concentrations of Ru complex in DMF ($\nu = 10\text{ mV s}^{-1}$); (B) Cyclic voltammograms of MWCNT/ RuQ-pyrene electrode ($\Gamma_{\text{RuQ-pyrene}} = 15\text{ nmol cm}^{-2}$) in absence (dashed line) and presence (full line) of NADH (10mM) in PBS (0.2 M, pH 7) at RT ($\nu = 10\text{ mV s}^{-1}$).

The maximum surface coverage is equivalent to 220 layers of closely-packed RuQ-pyrene, underlining the high specific surface of MWCNT films. This is confirmed by scanning electron microscopy (SEM) image of the surface of the RuQ-pyrene-functionalized MWCNT electrode (figure S1).

Electrocatalytic properties of these functionalized electrodes were studied in water. We investigated the redox behaviour of phenidone ligand in the presence of NADH (figure 2B). In the presence of 10 mM NADH, an irreversible anodic peak current is observed, starting at -0.10V , at the foot of the QH_2/Q oxidation peak, confirming the electrocatalytic properties of RuQ-pyrene-functionalized MWCNTs. Maximum catalytic current of 2.5 mA cm^{-2} was achieved at 10 mM NADH.

Thanks to the easy self-assembly of pyrene molecules onto MWCNT sidewalls, we studied the double functionalization of MWCNT electrodes with both RuQ-pyrene and NHS-pyrene (figure 3A).

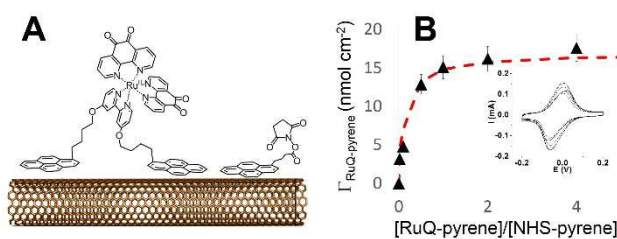


Figure 3. (A) Schematic representation of the double functionalization of MWCNTs with RuQ-pyrene and NHS-pyrene; (B) Variation of the RuQ-pyrene surface coverage with incubating RuQ-pyrene/NHS-pyrene concentration ratios in DMF accompanied with the fitting binding isotherm (red dashed line); (inset): Cyclic voltammetry of RuQ-pyrene-functionalized MWCNT electrodes performed in 0.2 M PBS ($\nu = 10\text{ mV s}^{-1}$), using different incubating concentrations of RuQ-pyrene and NHS-pyrene in DMF

MWCNT electrodes were simply incubated in solution containing different ratios of RuQ-pyrene and 1-pyrenebutyric acid N-hydroxysuccinimide ester (NHS-pyrene). As investigated for RuQ-pyrene incubating solutions, we measured the RuQ-pyrene surface coverage by electrochemistry in water.

In this case, the **RuQ-pyrene** surface coverage increases with increasing **RuQ-pyrene/NHS-pyrene** concentration ratios. The evolution of the **RuQ-pyrene** surface coverage follows a binary competitive Langmuir isotherm, according to equation 2:

$$\Gamma_{eq,Ru} = \frac{\Gamma_{max} \times K_{Ru} \times [RuQ-pyrene]}{1 + K_{Ru} \times [RuQ-pyrene] + K_{NHS} \times [NHS-pyrene]} \quad (2)$$

where K_{NHS} is the association constant of **NHS-pyrene** with MWCNTs in DMF at 25°C. This equation can be applied because **NHS-pyrene** and **RuQ-pyrene** do not interact in solution in the range of 0 to 10 mM and interact with same adsorption sites on CNT sidewalls. This model is corroborated by the fact that neither K_{Ru} (1900 L mol⁻¹) nor Γ_{max} (17.8 nmol cm⁻²) are influenced by the presence of **NHS-pyrene**. Furthermore, a K_{NHS} of 180 L mol⁻¹ was determined, which is more than ten times lower than K_{Ru} . This implies that **RuQ-pyrene** interacts stronger than **NHS-pyrene** with MWCNT sidewalls. This phenomenon likely arises from the presence of two pyrene groups in **RuQ-pyrene** that facilitate its adsorption and reinforce the mechanical stability of the immobilized form. Finally, these electrodes were incubated in a 2mg mL⁻¹ solution of GDH (figure 4). Figure 4B shows a typical CV in the presence of 160mM glucose and 10mM of NAD⁺. The addition of glucose triggers the appearance of an irreversible catalytic wave, starting at -0.10V and corresponding to the oxidation of NADH, enzymatically generated by oxidation of glucose by GDH active site. Kinetic measurements of immobilized GDH activity was performed by UV-Vis measurement, giving access to the real amount of grafted enzymes for each **[RuQ-pyrene]/[NHS-pyrene]** concentration ratio. As expected from the larger association constant K_{Ru} compared to K_{NHS} and from the bigger size of the enzyme compared to the polypyridine Ru^{II} complex, a similar amount of enzymes were immobilized on MWCNT whatever the ratio of the two pyrene derivatives. The latter, 7.1 x 10⁻¹⁰ mol cm⁻² of immobilized GDH, is approximately equivalent to 8 compact enzyme layers, illustrating the efficient chemical grafting of GDH within the whole 3D structure of the MWCNT coating.²² Moreover, this enzyme loading leads to a **RuQ-pyrene/enzyme** ratio of almost 260, assuming thus an efficient NADH oxidation.

The performances of bi-functionalized MWCNT electrodes were investigated by chronoamperometric measurements at 0.1V (figure 4C). Electro-catalytic currents increase with the increasing glucose concentration, according to a typical Michaelis-Menten dependence, reaching high catalytic currents of 1.95 mA cm⁻² at 5mM glucose and maximum catalytic currents of 6.0 mA cm⁻² at 70mM glucose. A calibration curve for glucose biosensing was obtained with a sensitivity of 0.27 mA mM⁻¹ cm⁻² with a detection limit of 10 μM (inset, figure 4C). This is among the best performances for glucose oxidation based on NAD-dependant GDH. High-performance previously-reported GDH-based bioanodes exhibited maximum currents of 0.4 mA cm⁻² for hydrogel/CNT composite²³, 1.5 mA cm⁻² for

microfluidic-based bioelectrodes²⁴ or 4.1 mA cm⁻² for NADH and poly-L-lysine-entrapped GDH on carbon fibers¹.

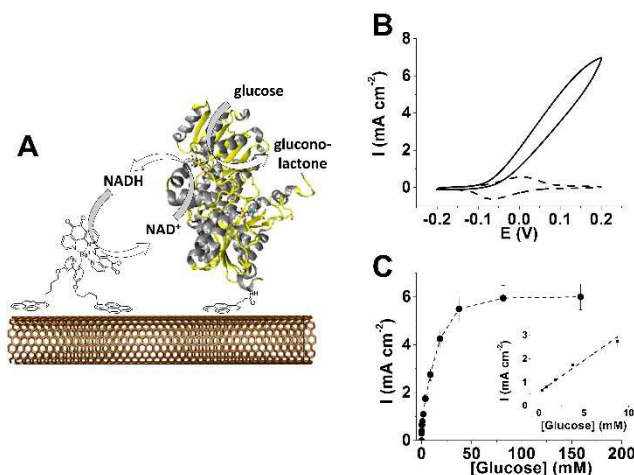


Figure 4. (A) Schematic representation of the preparation of the bi-functionalized MWCNT electrode with **RuQ-pyrene** catalyst and GDH. (B) Cyclic voltammograms of RuQ-pyrene/GDH-functionalized MWCNT electrode in absence (dashed line) and presence (full line) of glucose (160 mM) in phosphate buffer (0.2M, pH 7) containing 10 mM NAD⁺ at 37°C (v= 10 mV s⁻¹). (C) Calibration curve for glucose between 0 and 160 mM and in the linear range (inset). Applied potential 0.1V vs SCE in 0.2 M stirred phosphate buffer (pH7) containing 10mM NAD⁺ at 37°C.

The soft and efficient supramolecular functionalization with an original pyrene-modified Ru^{II} complex and GDH affords the easy double functionalization of MWCNT electrodes, while ensuring maximum surface coverage of both molecular catalysts and biocatalysts. Furthermore, fine control over functionalization was assessed by a competitive Langmuir isotherm model, underlining the different interactions of modified pyrene molecules. Thanks to low overpotentials and high current densities, these functionalized MWCNT films show great promise for the design of novel bioelectrodes, especially for biofuel cell applications.

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Notes and references

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