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In order to test our hypothesis, we designed the molecule of Nap-FFYGK-CA (Scheme 1). Many peptide derivatives based on FF or FFY are molecules with excellent self-assembling properties.¹⁸ We envisioned that our designed molecule might self-assemble into nanofibers with good water dispersity. The CA moiety was used to form a complex with melamine to increase the inter-fiber interactions for hydrogelation. The synthetic route was described in Scheme 1, we firstly prepared CA-acid in four steps with a total yield of about 70%. The Nap-FFYGK was produced by standard solid phase peptide synthesis (SPPS). We then obtained Nap-FFYGK-CA by coupling CA-acid with the peptide. The pure compound was achieved by reverse phase high performance liquid chromatography (HPLC).

After obtaining the Nap-FFYGK-CA, we tested its self-assembling property by the heating-cooling process. We firstly prepared a phosphate buffer saline (PBS, pH = 7.4) solution of it at a concentration of 0.5 wt%. After a heating-cooling process, we observed the formation of a clear solution (Fig. 1A), suggesting that the compound itself could not form a hydrogel at this concentration. We observed a light beam when we used a laser pointer to shine the solution, indicating the presence of nanoscale materials in the solution. We then tested whether the addition of melamine would lead to hydrogelations or not. We mixed the solution of Nap-FFYGK-CA at a final concentration of 0.5 wt% and the solution of melamine with different concentrations. As shown in Fig. 2B, the solution changed to a hydrogel within 10 minutes when the concentration of melamine was higher than 35 ppm. We also observed the hydrogel formation when the concentration of melamine was 20 ppm, but the gelation took a longer time of about 8 hours. These observations clearly indicated the success of our design. As the sol-gel phase transition can be easily identified by naked eyes, this system might be suitable for the detection of melamine without instruments.

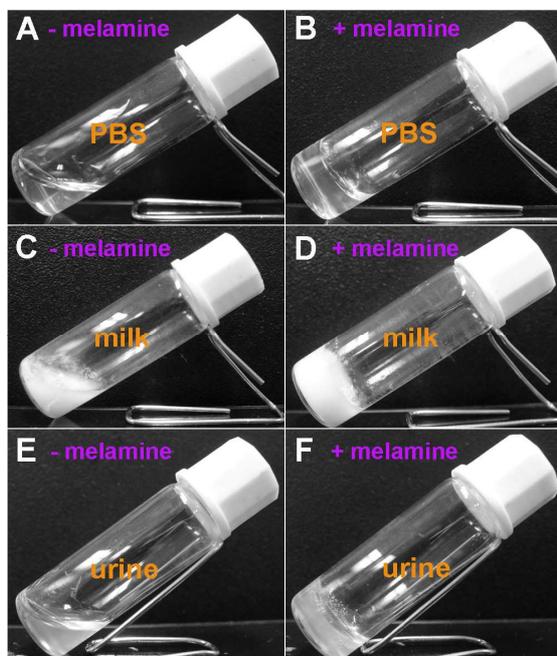


Fig. 1. Optical images of the solutions of Nap-FFYGK-CA without (left images) or with (right images) 35 ppm, 20 ppm, 20 ppm melamine in PBS (A and B), milk (C and D), and urine (E and F), respectively.

We also tested whether our method could be used to detect melamine in foods and biological fluids. We choose melamine tainted milk and urine because of the importance of detection of melamine in these two samples. There are proteins and urea in milk and urine, respectively, which may interfere with the hydrogelation. As shown in Fig. 1C, milk without melamine can not form a hydrogel at the concentration of 0.5 wt%. With the addition of melamine above the concentration of 20 ppm, a hydrogel would form within 30 min (Fig. 1D). Similar observations were achieved for melamine tainted urine, and the minimum detection concentration of melamine in urine was also about 20 ppm (Fig. 1E and 1F). Compared with other melamine detecting systems needing to extract melamine from the samples, our method was much simpler that the analytes could be detected directly by naked eyes. Such convenient method is very promising for the detection of melamine where no instrument is accessible.

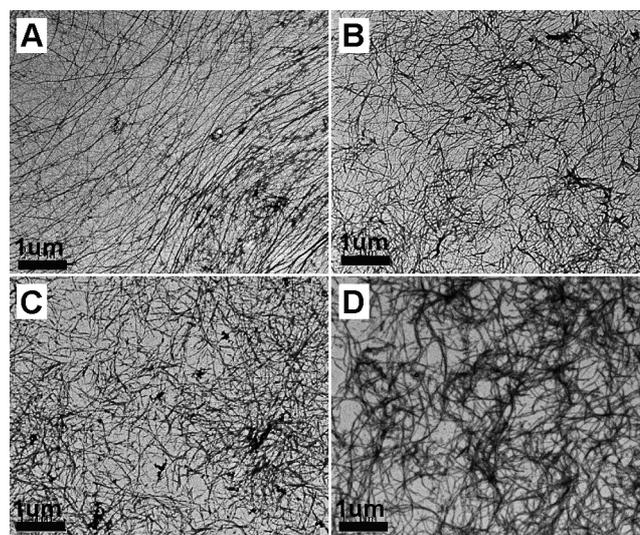


Fig. 2. TEM images of A) solution of Nap-FFYGK and gels with different equiv. of melamine: B) 2 equiv., C) 4 equiv. and D) 16 equiv.

We then used transmission electron microscopy (TEM) to characterize the morphology of self-assembled structures in the solution and hydrogels with different amounts of melamine. As shown in Fig. 2A, the compound Nap-FFYGK-CA self-assembled into filamentous structures with the diameter of about 25 nm. The density of cross-linking point was low probably due to the lack of strong interactions between fibers. In the presence of melamine, we observed higher densities of cross-linking point in the gel samples (Fig. 2B-2D, S-5, S-6, S-7, S-8). Compared with the fibers in the solution without melamine, the diameter of those in gels was bigger. Many reports have demonstrated that the hydrogen bonding and π - π stacking between melamine could assist the formation and tune the morphology of self-assembled nanostructures.¹⁹ In our study, we also observed that the diameter of fibers became bigger in the presence of more melamine and it was about 33, 40, 48, 53, 58, 68, and 77 nm in the presence of 0.25, 0.5, 1, 2, 4, 8, 16 equivalent of melamine, respectively (Fig. S-9). It is well-known that in order to form supramolecular

hydrogels, there should be strong or at least medium interactions between self-assembled nanostructures.²⁰ Our previous study had also showed that using a recombinant protein with multiple binding sites, we could enhance the inter-fiber interaction by the specific protein-peptide interaction for hydrogelations.²¹ In this study, with the assistance of CA-melamine complexation, we could enhance the interaction between supramolecular nanofibers, resulting in the formation of stable 3D networks and hydrogels.

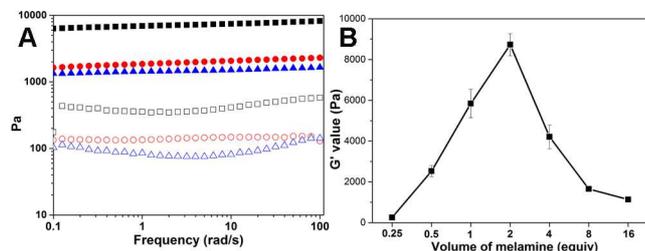


Fig. 3. A) Dynamic frequency sweep at the strain of 0.1% of the hydrogels with addition of 0.5 (circles), 2 (squares) and 8 (triangles) equiv of melamine (filled symbols: G' and open symbols: G''), and B) G' value in the mode of dynamic frequency sweep at the strain of 0.1% of the gels with different equivalents of melamine

We further investigated the mechanical property of hydrogels by rheology. The hot aqueous solutions containing Nap-FFYGK-CA and different amounts of melamine were directly transferred to the rheometer. After 2 hours' incubation to achieve stable hydrogels, we performed a dynamic strain/frequency sweep. The value of the storage moduli (G' , elasticity) of all samples was bigger than that of their corresponding loss moduli (G'' , viscosity), suggesting that all samples behaved as viscoelastic materials. The hydrogels showed bigger G' values with the increased amounts of melamine when the equivalent of melamine was lower than 2 (Fig. 3, S-10, S-11, S-12). For instance, the G' value reached 8728 Pa at the frequency value of 0.1 rad/s for the gel with 2 equivalent of melamine, which was about 35 times bigger than that with 0.25 equivalent of melamine. However, when the amount of melamine was more than 2 equivalent, the G' value of resulting hydrogels dropped (Fig. 3). These results indicated that the mechanical property of the hydrogels might be regulated by changing the molar ratios of melamine. As the mechanical properties of hydrogels are critical for their biological applications, a hydrogel that can finely tune the mechanical properties is highly desired.²²

Based on the above observations, we proposed possible interactions between melamine and supramolecular nanofibers in the hydrogels. As shown in Fig. 4, Nap-FFYGK-CA itself could self-assemble into aligned nanofibers with CA moiety at their surfaces. With low density of cross-linking points, these nanofibers can only form solutions. Assisted by the hydrogen bonds between melamine and the CA derivative, the nanofibers entangled with each other and formed stable 3D networks for hydrogelations. When the concentration of melamine was low, it served as the cross-linker. With the increase of melamine, more cross-linking points formed, resulting in higher G' values of the gels. However, melamine could also stack with each other. When excess amount of melamine was present in the gel, melamine

assembly formed at the surface of nanofibers, leading to the increase of diameter of nanofibers. In such situation, melamine assembly served as the cross-linkers to form fiber networks. Since the interaction between melamine was not so strong as that between melamine and CA, the mechanical property of gels with excess amount of melamine decreased.

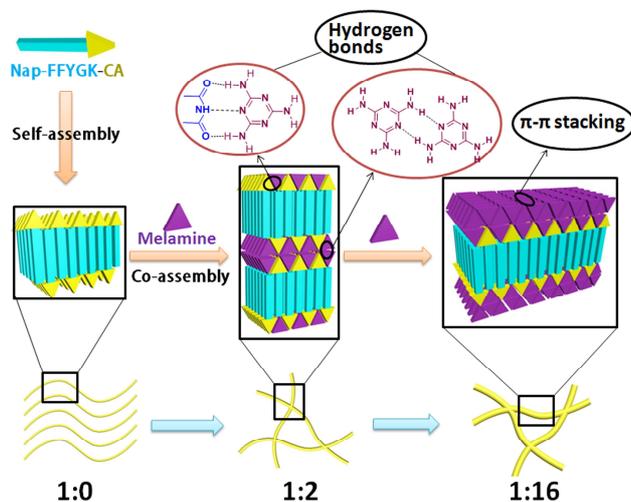


Fig. 4. Possible molecular arrangements in self-assembled nanofibers of Nap-FFYGK-CA and in gels with different amounts of melamine

In summary, we reported a supramolecular hydrogel-based detection system for melamine. Using this method, melamine could be directly visualized by naked eyes in milk and urine without pre-extraction process and the need of any instruments. Although the minimum detection concentration of melamine for our system is a little bit higher than the safety limits (2.5 ppm in the USA and EU and 1 ppm in China), we have provided a simple, real-time, and on-site way to detect melamine at the concentrations of higher than 20 ppm in foods and in biological samples. Our method is useful for the detection of high concentration of melamine in remote places without instruments.

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Notes and references

^aDepartment of Cardiology, Zhujiang Hospital of Southern Medical University, Guangzhou 510280, P. R. China; E-mail: gzminsheng@vip.163.com

^bCollege of Pharmacy and Tianjin Key Laboratory of Molecular Drug Design, Nankai University, Tianjin 300071, P. R. China

^cState Key Laboratory of Medicinal Chemical Biology, Key Laboratory of Bioactive Materials, Ministry of Education, and College of Life Sciences, Nankai University, Collaborative Innovation Center of Chemical Science and Engineering, Tianjin 300071, P. R. China; E-mail: yangzm@nankai.edu.cn

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