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Examination of mercaptobenzyl sulfonates as catalysts for native chemical ligation: Application to the assembly of a glycosylated Glucagon-Like Peptide 1 (GLP-1) analogue

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3/4-mercaptobenzyl sulfonates were investigated as aryl thiol catalysts for native chemical ligation (NCL). Whilst catalysing NCL processes at a similar rate to 4mercaptophenyl acetic acid (MPAA), the increased polarity and solubility of 3-mercaptobenzyl sulfonate in particular may favour its selection as NCL catalyst in many instances.

Native chemical ligation (NCL), where peptide thioesters and cysteinyl peptides combine to form native peptide linkages,^{1, 2} has benefited significantly from the use of 4-mercaptophenyl acetic acid (MPAA) as catalyst.³ MPAA (1, scheme 1) is relatively non-malodorous, has good water solubility under most commonly employed NCL reaction conditions, and catalyses NCL through formation of a reactive aryl thioester intermediate (3).^{4, 5}



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Scheme 1. *In-situ* thiol-thioester exchange during NCL converts the 2-mercaptoethanesulfonate (MESNa) thioester 2 to the highly reactive MPAA thioester 3.



Scheme 2. MPAA analogues investigated as NCL catalysts.⁷

5 and **6** were readily prepared on gram scale from the corresponding nitrobenzyl chlorides (7/8) in four steps (Scheme 3)^{8, 9} Notably, throughout the synthesis no flash column chromatography was required.



Scheme 3. Synthesis of mercaptobenzyl sulfonates 5 and 6.

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With **5** and **6** in hand we were pleased to observe that they both eluted 10 minutes earlier than MPAA on a C₁₈ analytical reverse-phase column¹⁰ and we next examined application in NCL. To compare their performance as catalysts model ligations were conducted using H-LYRAG-SCH₂CH₂SO₃Na (**15**) and H-CRAFS-OH (**16**) over a pH6-pH8 range (Figure 1).¹¹ The thioester component was conveniently prepared from H-LYRAGC-OH (**17**) in near quantitative yield via $N \rightarrow S$ acyl shift.¹²⁻¹⁷ NCL progress was then monitored by analytical reverse-phase HPLC and confirmed by LC-MS.



Figure 1. a) Preparation of LYRAG thioester **15** and reaction with **16** in NCL reactions in the presence of thiol catalyst. b) **5** and **6** are compared with MPAA as NCL catalysts at pH 6.¹⁸

At this stage the anticipated instability of 4-MBSA (5) became evident.¹⁹ Whilst preparing 0.25 M aqueous stock solutions of 4-MBSA for immediate use at 50 mM final concentration in NCL reactions allowed us to evaluate it as a catalyst, at higher concentrations 4-MBSA rapidly precipitated and/or polymerised. Consequently no reliable kinetic data could be obtained for the 5-mediated ligation between 15 and 16 at typical working concentrations (0.1-0.2 M) at pH 6 or above.

Interestingly 5 also precipitated from all solutions prepared in 6 M guanidine hydrochloride, a common NCL component yet did not precipitate when identical solutions were prepared in 8 M urea, allowing 4-MBSA mediated ligation reactions between peptides of low solubility to take place. We took advantage of this to assemble a glycosylated Glucagon-Like Peptide-1 (GLP-1) analogue (Scheme 4). GLP-1 (residues 7-36) has gained attention as a therapeutic for the treatment of type 2 diabetes and affects glucose control by stimulating Although GLP-1 is not naturally insulin secretion. glycosylated, a recent study showed that the addition of carbohydrate moieties at N34 (18, Scheme 4a) prolonged the half-life of the peptide in vivo, thus increasing its potential for therapeutic applications.²⁰ We envisaged that GLP-1 analogues of 18 could be reached via simply glycosylated 19 (Scheme 4b). Following straightforward production of NCL components 20 and 21 we successfully assembled 19 in 8 M Urea using 50 mM 4-MBSA as catalyst (Scheme 4c). In this case we

observed similar catalytic performance to 50 mM MPAA.²¹ However, the poor stability of **5** at neutral pH led us to focus on 3-MBSA (**6**) since 1 M solutions were fully soluble at pH7 and compatible with 6 M guanidine hydrochloride (Figure 2).



Scheme 4 a) Sequence of glycosylated GLP-1(7-36) 18. b) Simply glycosylated analogue 19. c) Preparation of 19 from thioester 20 and cysteinyl glycopeptide 21 employing 50 mM 5 or 0.1 M 6.



Figure 2. Analytical HPLC analysis of a) 3-MBSA and b) MPAA catalysed ligation between 20 and excess 21 (0.82 mM 20, 1.4 mM 21, 0.1 M catalyst, 6 M guanidine hydrochloride, 0.1 M Na phosphate buffer; pH 7, 35 mM TCEP). c-d) Comparison of semipreparative HPLC purification for c) the 3-MBSA catalysed reaction and d) the MPAA catalysed reaction.

The 0.1 M 3-MBSA catalysed reaction was successful but appeared to proceed significantly more slowly than the corresponding MPAA catalysed process (Figure 2a and 2b), requiring an extra 2 h reaction time to reach a similar level of conversion. It is clear from Figure 2a that a more significant **Journal Name**

be accelerated further by employing the higher concentrations (0.2 M) that are optimal for MPAA catalysis, and are more comparable with employing the pre-formed thioester.³ Despite the slightly slower reaction rate when using 3-MBSA, its enhanced polarity enabled straightforward purification of the ligation product whereas MPAA co-eluted with it.

After isolation of 19 the Gln \rightarrow Cys mutation at the Gly-Cys ligation site was simply carboxamidomethylated to restore a pseudo-glutamine ("Q") residue at this position (scheme 5). The N-acetyl glucosamine unit was finally extended to the native N-glycoprotein pentasaccharide core structure upon exposure to oligosaccharyl oxazoline (22) in the presence of Endoglycosidase A.^{22, 23}



Ac-HAEGTFTSDVSSYLEGCAAKEFIAWLVNGR-NH2



24 Ac-HAEGTFTSDVSSYLEG"Q"AAKEFIAWLVNGR-NH2

Scheme 5. Thiol capping and Endoglycosidase A mediated elaboration of 19.

Conclusions

Overall, the results demonstrate that compounds based on the mercaptobenzyl sulfonate scaffold can accelerate NCL reactions at a comparable rate to MPAA. 3-MBSA (6) served as the more stable catalyst and, owing to the increased polarity conferred by the sulfonate group, facilitated straightforward ligation of two GLP-1 fragments, and isolation of the product. MPAA can be significantly removed from ligation reaction upon acidification and repeated extraction of the reaction mixture,^{24,25} or by using MPAA hydrazide analogues which can be captured on suitably functionalised solid supports.^{26, 27} However it is hoped that the additional flexibility provided by 3-MBSA may allow purification of peptide products without these additional handling steps. Furthermore the high solubility of these aryl thiols, and further analogues such as

mercaptobenzyl phosphonates, at low pH may additionally find application in processes relevant to $N \rightarrow S$ acyl transfer and peptide transamidation reactions where MPAA has already been employed advantageously, despite its low solubility under these conditions.17,28

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References

- 1. S. B. H. Kent, Chem Soc Rev, 2009, 38, 338-351.
- 2. P. E. Dawson, T. W. Muir, I. Clark-Lewis and S. B. Kent, Science, 1994, 266, 776-779.
- 3. E. C. B. Johnson and S. B. H. Kent, J. Am. Chem. Soc., 2006, 128, 6640-6646
- 4. P. E. Dawson, M. J. Churchill, M. R. Ghadiri and S. B. H. Kent, J Am. Chem. Soc. 1997, 119, 4325-4329.
- 5. K. Mandal, B. L. Pentelute, D. Bang, Z. P. Gates, V. Y. Torbeev and S. B. H. Kent, Angew. Chem. Int. Ed. 2012, 51, 1481-1486.
- 6. T. K. Tiefenbrunn, J. Blanco-Canosa and P. E. Dawson, Peptide Science, 2010, 94, 405-413
- 7. We suspected 5 may have limited stability as a consequence of its predisposition to p-quinone methide formation, and 6 was prepared as a potentially stabilised analogue.
- 8. F. M. Beringer and R. A. Falk, J. Am. Chem. Soc. 1959, 81, 2997-3000
- 9. H. Kawai, F. Sakamoto, M. Taguchi, M. Kitamura, M. Sotomura and G. Tsukamoto, Chem. Pharm. Bull. 1991, 39, 1422-1425.
- 10. See supporting information for HPLC traces.
- 11. Juan B. Blanco-Canosa and Philip E. Dawson, Angew. Chem. Int. Ed. 2008, 47, 6851-6855.
- 12. D. Macmillan, A. Adams and B. Premdjee, Isr. J. Chem. 2011, 51, 885-899
- 13. J. Kang and D. Macmillan, Org. Biomol. Chem. 2010, 8, 1993-2002.
- 14. J. Kang, J. P. Richardson and D. Macmillan, Chem. Commun. 2009, 407-409.
- 15. J. Kang, N. L. Reynolds, C. Tyrrell, J. R. Dorin and D. Macmillan, Org. Biomol. Chem. 2009, 7, 4918-4923.
- 16. J.-S. Zheng, S. Tang, Y.-C. Huang and L. Liu, Acc. Chem. Res. 2013.
- 17. F. Burlina, G. Papageorgiou, C. Morris, P. D. White and J. Offer, Chem. Sci. 2014, 5, 766-770.
- 18. For clarity results are shown at pH 6. See supporting information for NCL catalysis at pH 7 and pH 8.
- 19. M. M. Toteva and J. P. Richard, in Advances in Physical Organic Chemistry, ed. P. R. John, Academic Press, 2011, pp. 39-91.
- 20. T. Ueda, K. Tomita, Y. Notsu, T. Ito, M. Fumoto, T. Takakura, H. Nagatome, A. Takimoto, S.-I. Mihara, H. Togame, K. Kawamoto, T.

ChemComm

Iwasaki, K. Asakura, T. Oshima, K. Hanasaki, S.-I. Nishimura and H. Kondo, *J. Am. Chem. Soc.* 2009, **131**, 6237-6245.

- See supporting information for the analytical HPLC traces for the 4-MBSA and MPAA catalysed reactions.
- W. Huang, Q. Yang, M. Umekawa, K. Yamamoto and L.-X. Wang, Chembiochem, 2010, 11, 1350-1355.
- T. B. Parsons, J. W. B. Moir and A. J. Fairbanks, Org. Biomol. Chem. 2009, 7, 3128-3140.
- 24 H. van de Langemheen, A. J. Brouwer, J. Kemmink, J. A. W. Kruijtzer and R. M. J. Liskamp, J. Org. Chem. 2012, 77, 10058-10064.
- N. Ollivier, J. Dheur, R. Mhidia, A. Blanpain and O. Melnyk, *Org Lett*, 2010, **12**, 5238-5241.
- 26. T. Moyal, H. P. Hemantha, P. Siman, M. Refua and A. Brik, *Chem. Sci.* 2013, **4**, 2496-2501.
- 27. It has also been shown that MPAA can be replaced with non-aryl thiol 2,2,2-trifluoroethane thiol, which catalyses NCL reactions at a rate comparable with MPAA: R. E. Thompson, X. Liu, N. Alonso-García, P. J. B. Pereira, K. A. Jolliffe and R. J. Payne, *J. Am. Chem. Soc.* 2014, **136**, 8161-8164.
- N. Ollivier, A. Blanpain, E. Boll, L. Raibaut, H. Drobecq and O. Melnyk, Org. *Lett.* 2014, 16, 4032-4035.